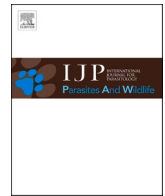


Contents lists available at [ScienceDirect](https://www.sciencedirect.com)

International Journal for Parasitology: Parasites and Wildlife

journal homepage: www.elsevier.com/locate/ijppaw

Gastrointestinal and cardiorespiratory endoparasites in the wild felid guigna (*Leopardus guigna*) in Chile: Richness increases with latitude and first records for the host species

Francisca Acuña-Olea^{a, **}, Irene Sacristán^b, Emilio Aguilar^a, Sebastián García^a,
María José López^a, Pablo Oyarzún-Ruiz^c, José Luis Brito^d, Fernando Fredes^e,
Constanza Napolitano^{f, g, *}

^a Facultad de Ciencias Veterinarias y Pecuarias, Universidad de Chile, La Pintana, Santiago, Chile

^b Doctorado de Medicina de La Conservación, Facultad de Ciencias de La Vida, Universidad Andrés Bello, Santiago, Chile

^c Facultad de Ciencias Veterinarias, Universidad de Concepción, Chillán, Chile

^d Museo de Historia Natural e Histórico de San Antonio, San Antonio, Chile

^e Unidad de Parasitología, Departamento Medicina Preventiva Animal, Facultad de Ciencias Veterinarias y Pecuarias, Universidad de Chile, La Pintana, Santiago, Chile

^f Departamento de Ciencias Biológicas y Biodiversidad, Universidad de Los Lagos, Osorno, Chile

^g Instituto de Ecología y Biodiversidad (IEB), Santiago, Chile

ARTICLE INFO

Keywords:

Leopardus guigna, endoparasite
Helminth
Necropsy

ABSTRACT

Guignas (*Leopardus guigna*) are small felids closely associated with native forest habitats. In fragmented landscapes, they use vegetation corridors and forest remnants to move across the landscape. In these contexts, guignas may increase contact probabilities with domestic animals, being therefore relevant to assess their pathogens and parasites. The aim of this study was to characterize the helminth fauna in the gastrointestinal tract and cardiorespiratory system of guignas from central and southern Chile. Between 2015 and 2018, 33 dead free-ranging guignas were found road-killed or were collected from wildlife rescue centers. Thirty-two gastrointestinal tracts and 32 cardiorespiratory organs were analyzed through direct analysis and artificial digestion. We found 81.8% (27/33) guignas were positive for helminth endoparasites (84.4% (27/32) positive for gastrointestinal parasites, 37.5% (12/32) positive for cardiorespiratory parasites). Fourteen parasites were identified (7 at genus level and 7 at species level), with *Angiostrongylus* sp., *Molineus* sp., *Oslerus* sp. and *Troglostrongylus* sp. as first records in guignas. The most prevalent parasites were the species *Toxascaris leonina*, *Toxocara cati* and *Uncinaria stenocephala*. *Uncinaria stenocephala* showed the highest intensity of infection. Multiparasitism was observed in 76% of the animals. Significant differences in richness of endoparasites and prevalence of cardiorespiratory parasites were found between geographic zones; higher values in the southern zone are possibly due to favorable environmental characteristics for endoparasite development. There were no statistically significant differences between sexes. All the parasites found in this study have been previously reported in domestic cats. These results are valuable to understand parasite transmission at the domestic-wildlife interface; the possibility of endoparasite transmission between domestic cats and guignas should be clarified with molecular analysis.

1. Introduction

Parasites are important for biodiversity; they belong to biological communities and have an impact on food chains (Hudson et al., 2006; Dobson et al., 2008). The diversity and abundance of parasites can reflect the diversity and abundance of the community of hosts, thus,

parasites may be indicative of the health of the ecosystems (Hudson et al., 2006), shedding light on disturbances in the hosts and their environments (Marcogliese, 2005; Thompson et al., 2010).

The current climate change and human perturbation of the environment could modify the dynamics of pathogen transmission (Seguel and Gottdenker, 2017). This would be more relevant for native species

* Corresponding author. Departamento de Ciencias Biológicas y Biodiversidad, Universidad de Los Lagos, Av. Fuchslocher 1305, Osorno, Chile.

** Corresponding author.

E-mail addresses: francisca.acuna@veterinaria.uchile.cl (F. Acuña-Olea), constanza.napolitano@ulagos.cl (C. Napolitano).

<https://doi.org/10.1016/j.ijppaw.2020.07.013>

Received 4 May 2020; Received in revised form 22 July 2020; Accepted 28 July 2020

Available online 31 July 2020

2213-2244/© 2020 The Authors. Published by Elsevier Ltd on behalf of Australian Society for Parasitology. This is an open access article under the CC BY-NC-ND

license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

in fragmented habitats, where they could increase their contact probability with domestic species and the infectious agents they may carry (Fiorello et al., 2006). The guigna (*Leopardus guigna*), a small wild felid, is currently affected by human perturbation, being classified as Vulnerable by the IUCN Red List (Napolitano et al., 2015). Guignas are closely associated with native forests of Chile and Argentina, but they can adapt to move across fragmented landscapes using vegetation corridors (Sanderson et al., 2002; Acosta-Jamett and Simonetti, 2004; Gálvez et al., 2013, 2018; Fleschutz et al., 2016). Thus, this felid could be an indicator species for the consequences of habitat fragmentation on forest-dwelling species, and it may be possible to study the contact between guignas and domestic carnivores through the pathogens and parasites they could share.

Previous studies on endoparasites of guignas were based on coprological analysis (Cortés, 2006; González-Acuña et al., 2010; Vallverdú, 2014; Acosta-Jamett et al., 2018), direct techniques with necropsy in a small number of animals (Wolffhügel, 1949; Álvarez, 1963; Álvarez et al., 1970; Fernández and Villalba, 1984; González-Acuña et al., 2010; Moleón et al., 2015), and mainly targeting gastrointestinal parasites. The aim of this study was to characterize the helminth fauna in the gastrointestinal tract and cardiorespiratory system of guignas from central and southern Chile. Our objective was to describe prevalence, richness and diversity of parasites in free-ranging guignas, assessed for the first time with a direct technique in a larger number of animals.

2. Material and methods

2.1. Sampling

From 2015 to 2018, 33 dead free-ranging guignas were collected, found road-killed or after death or being euthanized in wildlife rescue centers. All guignas came from anthropized areas. Seventeen (51.5%) guignas were from central Chile (hereinafter central zone, Valparaíso region to Biobío region, 32°S 71.6°W - 38°S 72.4°W) and 16 (48.5%) from southern Chile (hereinafter southern zone, Araucanía region to Los Lagos region, 38°S 72.4°W - 43°S 73.6°W) (Fig. 1, Supplementary Table S1). Eighteen females and 15 males were analyzed, 31 adults and 2 juveniles. Trachea, lungs and heart were collected. The gastrointestinal tracts were removed, knotting each portion separately (esophagus, stomach, small intestine and large intestine) with a cotton thread. The organs were frozen and stored at -20 °C until subsequent analysis.

In two different road-killed guignas, the cardiorespiratory organs (in one individual) and the gastrointestinal tract (in the other) were destroyed. Therefore 32 gastrointestinal tracts and 32 hearts and lungs were analyzed. Carcass collection was approved by Animal Ethics Committee of the Institute of Ecology and Biodiversity of the Universidad de Chile (resolution of 20 November 2015) and SAG (Agriculture and Livestock Service) (capture permits 7624/2015, 2288/2016, 2185/2017, 4072/2018).

2.2. Sample analysis

Carcasses, organs and helminths were analyzed at the Parasitology

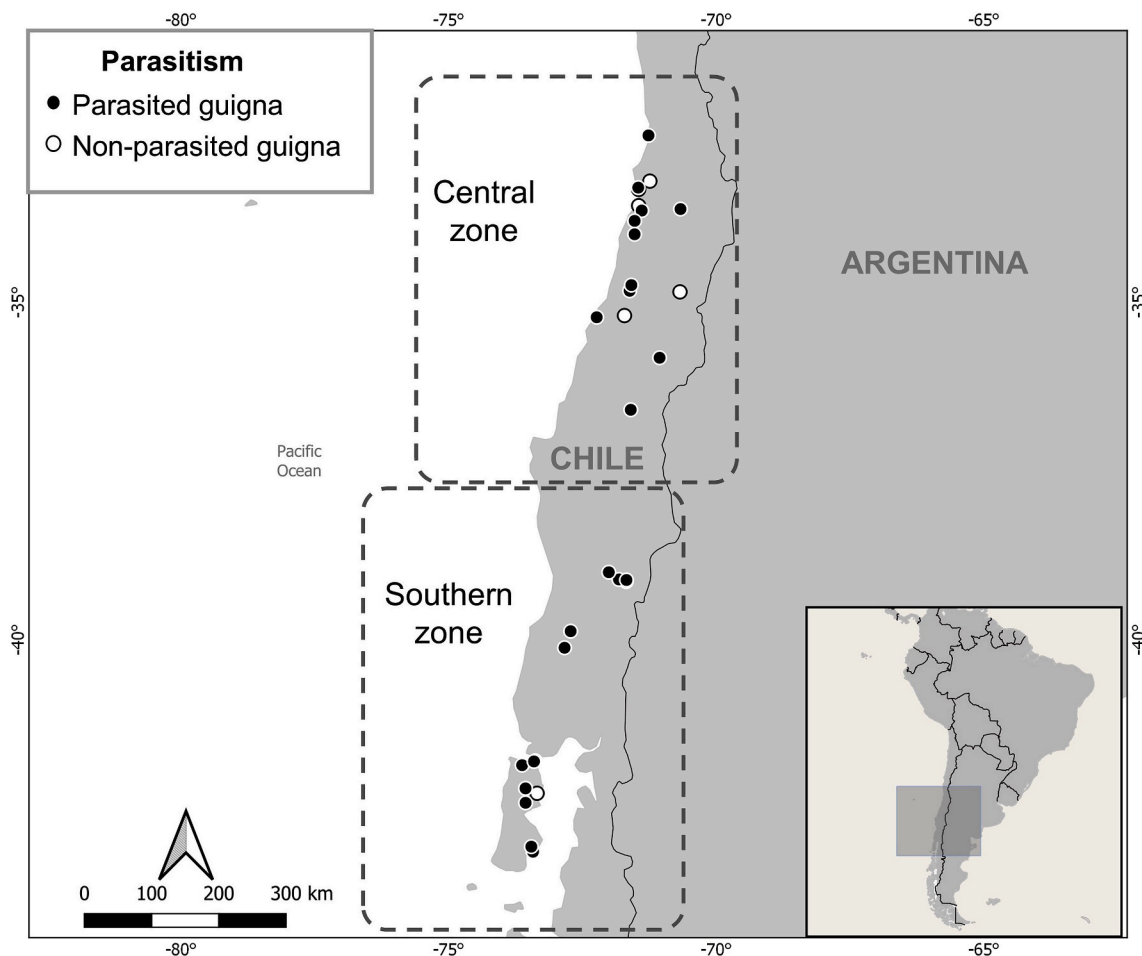


Fig. 1. Geographical locations and parasitism status of analyzed guigna samples in Chile. Dotted line boxes delimit the two geographic zones (Center, South) studied in Chile.

Laboratory of the Department of Animal Preventive Medicine at the Faculty of Veterinary and Animal Sciences of University of Chile.

Each portion of the gastrointestinal tract was dissected for macroscopic analysis. The content was washed into a 63 µm mesh stainless steel sieve, and the liquid obtained was observed under a stereo microscope (Zeiss Stemi 1000 Stereo Microscope), according to Tagle (1970). The heart and respiratory tract (trachea and bronchial tree) were dissected and observed under a stereo microscope. Then, for a detailed inspection for undetected helminths, we performed an enzymatic digestion with hydrochloric acid and pepsin to the lungs and hearts, according to Martínez-Rondán et al. (2017). All the retrieved helminths were stored in 70% ethanol at room temperature.

Nematodes were cleared with lactophenol, while cestodes were dehydrated in a series of alcohol concentrations (30°, 40°, 50°, 60°, 70° and 99.9°) and cleared with xylol. The helminths were observed under a light microscope (Miotic BA310 Led Binocular) at 10X and 40X. Parasites were identified according to morphological keys of Soulsby (1987), Khalil et al. (1994) and Anderson et al. (2009).

2.3. Statistical analysis

The observed prevalence (infected guignas/analyzed guignas), intensity of infection (number of parasites/guigna), abundance of infection and parasite species richness were obtained according to Bush et al. (1997), and compared between geographic zones (central/south) and sex of the guignas.

We conducted a Pearson's chi-squared test (statistical significance of $p \leq 0.05$) to compare the prevalence of parasite infection between sexes and geographic zones. Associations between intensity of infection and species richness, with sex or geographic zone, were analyzed with the Mann-Whitney U test. The Shannon-Wiener index was used to estimate diversity.

Statistical analyses were conducted with the software IBM SPSS Statistics 20 and PAST 3.2.

3. Results

Helminth endoparasites were recorded in 27 (81.8%) of 33 analyzed guignas. Twenty-seven (84.4%) of 32 gastrointestinal tracts were parasitized, and 12 (37.5%) of 32 guignas were positive for cardiorespiratory

helminths, with parasites in four hearts (12.5%) and 11 lungs (34.4%). Fourteen parasites were identified, 7 at genus level and 7 at species level, in addition to immature proglottids of Diphyllbothriidea cestodes that could not be identified (Table 1). This is the first record in guignas of *Angiostrongylus* sp. (Strongylida: Angiostrongylidae), *Molineus* sp. (Strongylida: Molineidae), *Oslerus* sp. (Strongylida: Filaroididae) and *Troglostrongylus* sp. (Strongylida: Crenosomatidae) (Fig. 2). *Angiostrongylus* sp. was present in the heart of four guignas from southern Chile (Pucón and Villarrica). *Molineus* sp. was found in the small intestine of a guigna from Pucón. *Oslerus* sp. was detected in the peribronchial tissue and lung parenchyma of guignas from Pucón and Paillaco (southern zone). *Troglostrongylus* sp. was retrieved in the lungs of four guignas from southern Chile (Pucón and Quellón) and one guigna from the central zone (Constitución).

The most prevalent parasites were the species *Toxascaris leonina* (Ascaridida: Ascarididae) (56.3%), *Toxocara cati* (Ascaridida: Toxocaridae) (37.5%) and *Uncinaria stenocephala* (Strongylida: Ancylostomidae) (37.5%). The highest intensity of infection was shown by *U. stenocephala* (105 helminths).

The most frequent parasite associations were *T. leonina* with *T. cati* and *T. leonina* with *U. stenocephala* in nine guignas, followed by *T. leonina* with *Hydatigera taeniaeformis* (Cyclophyllidea: Taeniidae) and *T. cati* with *U. stenocephala*, in six guignas. Nevertheless, these associations were not statistically significant. The parasites that showed positive associations and significant values were *Troglostrongylus* sp. with *Angiostrongylus* sp. (in three guignas, $p = 0.0005$) and *U. stenocephala* with *Oslerus* sp. (in two guignas, $p = 0.049$). A non-significant negative association was found between *T. cati* and *Troglostrongylus* sp. ($p = 0.052$), with no cases of coinfection.

Analyzing prevalence by geographic zone, we found a significant difference in the prevalence of cardiorespiratory parasites (center: 5.9% and south: 73.3%; $\chi^2 = 15.469$; $p = 0.00008$), with higher values in the south. No significant difference was found in parasite prevalence between sexes (Table 2, Supplementary Table S2). Mean abundance of parasites and general mean infection intensity showed significant differences between geographic zones, with higher values in guignas from the southern zone (Table 2).

Multiparasitism was present in 76% of the guignas, with a maximum of six parasite species/genera in four cases. There was a significant difference in parasite richness between geographic zones

Table 1

Observed prevalence, mean abundance (\pm SE), mean infection intensity (\pm SE) and intensity of infection range (min-max), of helminths retrieved in the sampled guignas.

	Organs ^a	Prevalence		Mean abundance	Mean infection intensity	Infection intensity range
		IG/AG ^b	% (CI) ^c			
Parasitized guignas		27/33	81.8 (67.9–95.7)			
Gastrointestinal parasites		27/32	84.4 (71.1–97.7)			
<i>Toxascaris leonina</i>	E, S, SI, LI	18/32	56.3 (38.1–74.4)	6.25 \pm 3.31	11.11 \pm 5.68	1–103
<i>Toxocara cati</i>	E, SI, LI	12/32	37.5 (19.8–55.2)	1.28 \pm 0.38	3.42 \pm 0.67	1–8
<i>Uncinaria stenocephala</i>	SI, LI	12/32	37.5 (19.8–55.2)	7.5 \pm 3.62	20 \pm 8.71	1–105
<i>Taenia</i> sp.	E, SI, LI	9/32	28.1 (11.7–44.6)	0.31 \pm 0.09	1.11 \pm 0.11	1–2
<i>Hydatigera taeniaeformis</i>	E SI	9/32	28.1 (11.7–44.6)	0.72 \pm 0.31	2.56 \pm 0.85	1–9
<i>Spirometra</i> sp.	SI	6/32	18.8 (4.5–33)	0.44 \pm 0.17	2.33 \pm 0.33	1–3
<i>Spirometra mansonioides</i>	E, SI, LI	4/32	12.5 (0.4–24.6)	0.78 \pm 0.49	6.25 \pm 2.84	2–14
Diphyllbothriidea	SI, LI	4/32	12.5 (0.4–24.6)	0.31 \pm 0.22	2.5 \pm 1.5	1–7
<i>Molineus</i> sp.	SI	1/32	3.1 (0–9.5)	0.03 \pm 0.03	1	1
Capillarinae gen. sp.	E	1/32	3.1 (0–9.5)	0.03 \pm 0.03	1	1
Cardiorespiratory parasites		12/32	37.5 (19.8–55.2)			
<i>Aelurostrongylus abstrusus</i>	L	6/32	18.8 (4.5–33)	1.53 \pm 1.16	8.17 \pm 5.8	1–37
<i>Troglostrongylus</i> sp.	T, L	5/32	15.6 (2.3–28.9)	1.16 \pm 0.79	7.4 \pm 4.41	1–24
<i>Angiostrongylus</i> sp.	H	4/32	12.5 (0.4–24.6)	0.13 \pm 0.06	1	1
<i>Eucoleus aerophilus</i>	T, L	3/32	9.4 (0–20.1)	0.25 \pm 0.19	2.67 \pm 1.67	1–6
<i>Oslerus</i> sp.	L	2/32	6.3 (0–15.1)	2.78 \pm 2.42	44.5 \pm 32.5	12–77

^a Anatomic location of the endoparasites: E, esophagus; S, stomach; SI, small intestine; LI, large intestine; T, trachea; L, lungs; H, heart.

^b Infected guignas/Analyzed guignas.

^c Prevalence of infection expressed in percentage (Confidence interval with 95% confidence level).



Fig. 2. Parasites recovered from analyzed guignas in Chile: **A**-anterior extremity of female *Molineus* sp. (40x); **B**- caudal extremity of female *Molineus* sp. with caudal spine (40x); **C**- oral extremity of female *Angiostrongylus* sp. (40x); **D**-lateral view of caudal extremity of female *Angiostrongylus* sp. (40x); **E**-caudal extremity of male *Angiostrongylus* sp., caudal bursa and spicules can be observed (40x); **F**- lateral view of the rounded caudal extremity of female *Oslerus* sp. (40x); **G**-lateral view of the caudal extremity of male *Oslerus* sp., with characteristic short and stout spicules (40x); **H**- oral extremity of female *Troglostrongylus* sp. (100x); **I**- lateral view of the caudal extremity of male *Troglostrongylus* sp., long spicules and caudal bursa can be observed (40x).

Table 2

Parasitism prevalence, mean abundance (\pm standard error), and mean infection intensity (\pm standard error) of helminths retrieved, by geographic zone and sex of the guignas.

	Geographic zone			Sex		
	Center	South	p value	Females	Males	p value
Parasitism prevalence	12 (70.6%)	15 (93.8%)	NS ^a	14 (77.8%)	13 (86.7%)	NS
GI prevalence ^b	12 (75.0%)	15 (93.8%)	NS	14 (77.8%)	13 (92.9%)	NS
CR prevalence ^c	1 (5.9%)	11 (73.3%)	0.00008	4 (23.5%)	8 (53.3%)	NS
Mean abundance	4.88 \pm 5.55	41.81 \pm 66.41	0.0004	23.89 \pm 13.74	21.47 \pm 9.78	NS
Mean infection intensity	6.92 \pm 1.57	44.6 \pm 67.77	0.001	30.71 \pm 17.36	24.77 \pm 11.05	NS

^a NS = non-significant p value ($p > 0.05$).

^b Guignas positive for gastrointestinal parasites.

^c Guignas positive for cardiorespiratory parasites.

(Mann–Whitney $U = 51.5$; $p = 0.002$), being higher in southern guignas (Fig. 3, Supplementary Table S3). Regarding parasite richness per system (gastrointestinal and cardiorespiratory organs) we found

significantly higher cardiorespiratory parasite richness in guignas from the southern zone (Mann–Whitney $U = 48.0$; $p = 0.0002$) and in males (Mann–Whitney $U = 88.0$; $p = 0.047$) (Fig. 4, Fig. 5).

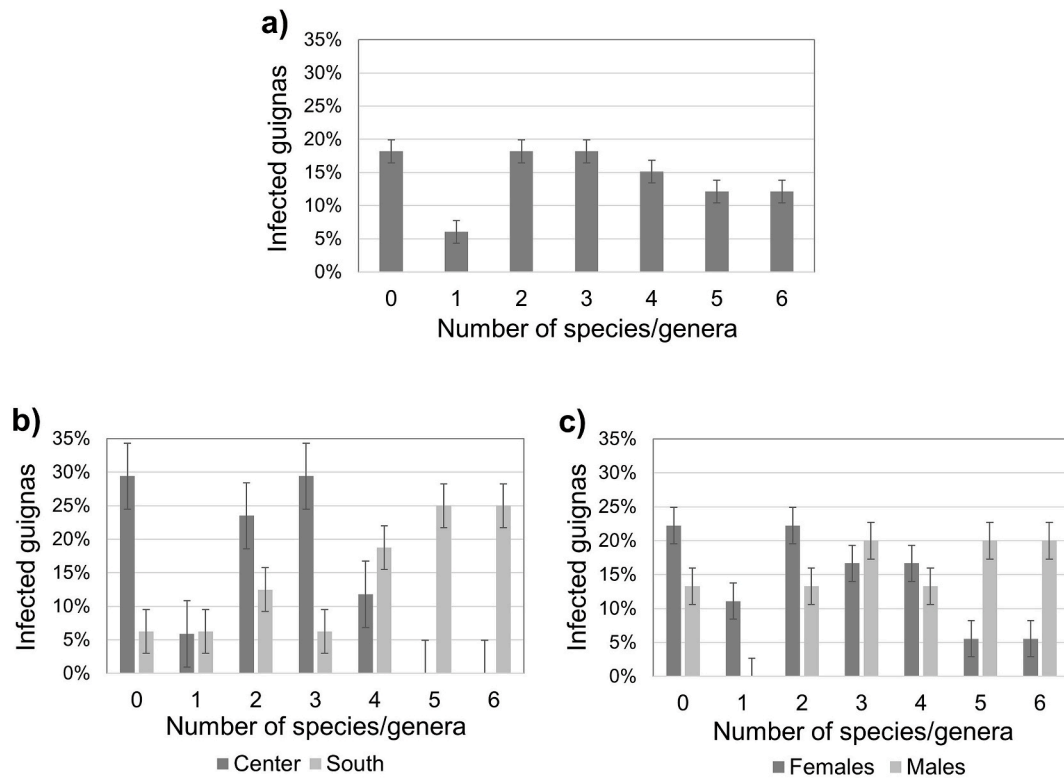


Fig. 3. Parasite richness (number of species/genera) in the analyzed guignas from Chile (a) overall, (b) by geographic zone and (c) by sex. Error bars represent standard error of 5%.

The Shannon-Wiener diversity index showed similar values between sexes and geographic zones. Nevertheless, the mean richness was different between them (Table 3).

4. Discussion

We report higher parasite richness compared to previous studies, with a maximum of 8 parasite species in Vallverdú (2014) and Aosta-Jamett et al. (2018). This could be due to the larger number of guignas sampled and the direct analysis applied to the organs. With a direct analysis to the organs, we could detect adult helminths, in contrast to previous coprological studies, where detection of parasites is affected by variations in egg and larvae excretion in the feces of the host (Houpin et al., 2016).

The Shannon-Wiener index considers species richness and abundance (Smith and Smith, 2001). In our study, the Shannon index showed similar values despite the difference in parasite richness between geographic zones. This could be due to greater variation in parasite abundance among guignas in the south, decreasing the diversity index. Guignas from the central zone have lower parasite richness, but abundance is similar among animals. The higher index in males could be due to the higher species/genera richness found.

Four parasite genera were detected for the first time in *L. guigna*. *Molineus* is a nematode genus with a direct transmission cycle; the definitive hosts (carnivores and primates) acquire the infective larvae from the environment (Bowman et al., 2002). Four species/genera have been previously reported in felids: *Molineus* sp. in jaguar (*Panthera onca*), cougar (*Puma concolor*) and ocelot (*Leopardus pardalis*) in Peru (Aranda et al., 2013); *M. barbatus* in bobcat (*Lynx rufus*) from USA (Hiestand et al., 2014); *M. felineus* in cougar from Argentina (Moleón et al., 2015) and *M. cati* in domestic cat (*Felis catus*) from South Africa (Durette-Desset et al., 2000). In this study the identification at species level could not be achieved because the only nematode found was an adult female.

The other three newly detected genera in guigna: *Angiostrongylus* sp., *Oslerus* sp. and *Troglostrongylus* sp. (Nematoda: Metastrongyloidea) are endoparasites with an indirect life cycle. The intermediate hosts are gastropods, but rodents, birds and reptiles can become paratenic hosts (Bowman et al., 2002). According to the dietary composition of guignas (Freer, 2004; Correa and Roa, 2005; Zúñiga et al., 2005; Astorga, 2013; Figueroa et al., 2018), paratenic hosts are possibly their main source of infection, as suggested in other felids (Bowman et al., 2002; Brianti et al., 2013). None of these three genera have been previously reported in the genus *Leopardus*. In the case of *Troglostrongylus* sp., our report is the first detection in a South American felid.

In our study, the four guignas positive for *Angiostrongylus* sp., had only one adult helminth in their hearts. These results differ from previous studies in felids (Vieira et al., 2013; Traversa et al., 2015; Diakou et al., 2016; Gherman et al., 2016), where the hosts carry several helminths; however, Varcasia et al. (2014) found only one female adult in the pulmonary artery of a domestic cat.

Metastrongyloid mixed infections were detected in five guignas from the southern zone; one of them, a male guigna from Pucón, was infected by the four metastrongyloids detected in this study (*Aelurostrongylus abstrusus* (Strongylida: Angiostrongylidae), *Angiostrongylus* sp., *Oslerus* sp. and *Troglostrongylus* sp). These co-infections have been previously reported in domestic cats (Traversa et al., 2015; Varcasia et al., 2015) and European wildcat (*Felis silvestris silvestris*) (Veronesi et al., 2016) in Europe. This could be due to the occurrence of shared intermediate and paratenic hosts for these parasites in the same area (Traversa et al., 2015).

The lack of previous reports of these parasites in guignas, may be due to difficulties in the diagnosis based on coprological analysis, where the larval stages of metastrongyloids are similar in size and morphology (Brianti et al., 2014b). Moreover, previous studies did not adopt the Baermann test, regarded as the gold standard technique for detection of metastrongyloid larvae in feces (Traversa and Di Cesare, 2013). The direct dissection and enzymatic digestion used in this study could reduce

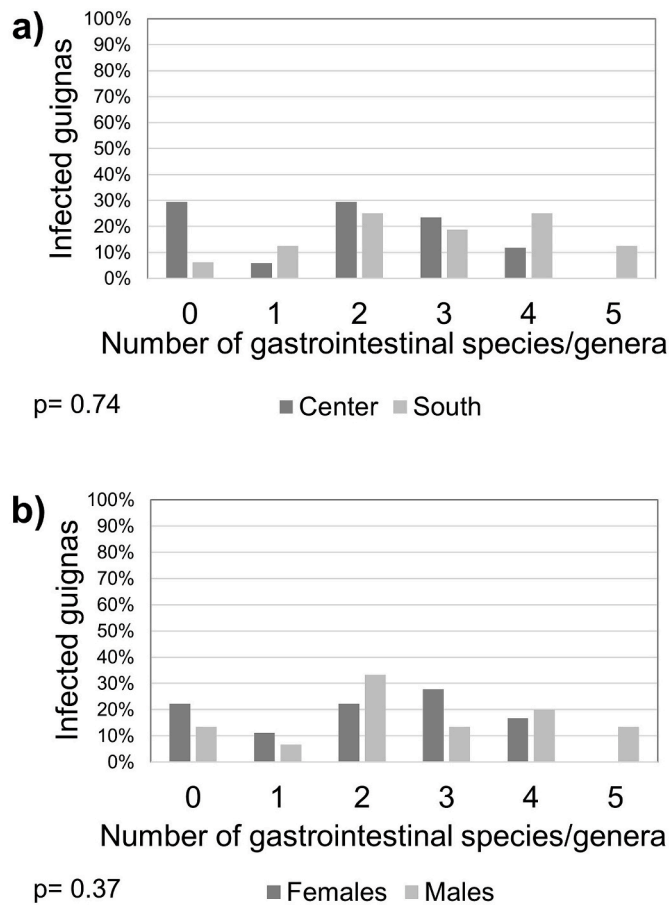


Fig. 4. Richness of gastrointestinal parasites in guignas from Chile, compared (a) by geographic zone and (b) by sex. p values show the statistical significance in the Mann-Whitney U test between (a) center-south and (b) females-males.

the false negative hosts compared to other parasitological analyses (Houpin et al., 2016; Martínez-Rondán et al., 2019). An underdiagnosis of Metastrongyloidea parasites has been proposed by other authors (Otranto et al., 2013; Traversa and Di Cesare, 2013; Penagos-Tabares et al., 2018), due to difficulties in identifying the larval stages in coprological studies, or by misdiagnosing them as *A. abstrusus* (Traversa and Di Cesare, 2013). Nevertheless, global warming, changes in host population dynamics and domestic animal contact with natural landscapes, among others, are also proposed as factors for the rise of reports of these parasites in various geographic areas (Traversa et al., 2010; Vieira et al., 2013; Brianti et al., 2014a; Veronesi et al., 2016; Lange et al., 2018). Awareness of these endoparasites is necessary because of their high pathogenic potential in felids and canids, especially in younger animals or in high worm burdens (Traversa et al., 2010).

Three statistically significant associations were found in this study. The presence of factors that favor or avoid coinfection with these parasites is unknown. Nevertheless, both positive associations are parasites with higher prevalence in southern zone, where intermediate hosts (for *Troglostrongylus* sp.-*Angiostrongylus* sp.) or paratenic hosts (for *U. stenocephala*-*Oslerus* sp.) could possibly be shared. Co-infection of cardiorespiratory parasites in intermediate hosts was reported by Lange et al. (2018) and Penagos-Tabares et al. (2019), finding larvae of two metastrongyloid species in the same slug. Thus, intermediate hosts with mixed infections could be eaten by paratenic hosts and then be transmitted to guignas. Parasite associations should be further explored in the future, to assess if this phenomenon is present in other carnivore species and its possible causes.

In our study, cardiorespiratory parasite prevalence, intensity of infection, abundance of infection and parasite richness showed

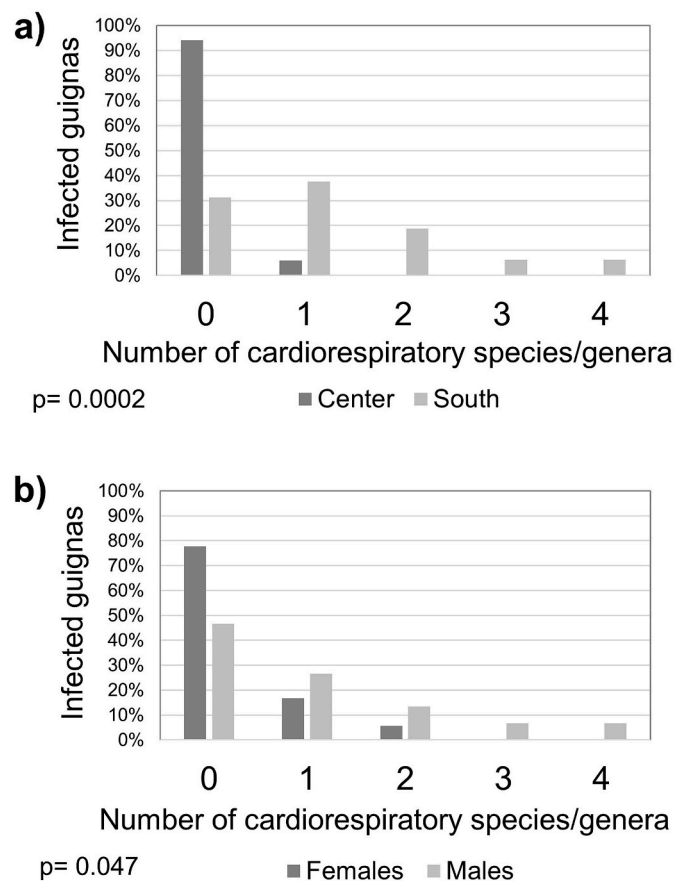


Fig. 5. Richness of cardiorespiratory parasites in guignas from Chile, compared by (a) geographic zone and (b) by sex. p values show the statistical significance in the Mann-Whitney U test between (a) center-south and (b) females-males.

Table 3

Specific richness (number of parasite species/genera), mean richness (\pm SD) (mean of parasite species/genera per sample) and Shannon-Wiener index of the helminths retrieved, by geographic zone and sex of the analyzed guignas.

Parameter	n	Specific richness	Mean richness	Shannon-Wiener (H')
Geographic zone				
Center	17	8	1.88 \pm 1.45	1.882
South	16	14	4.0 \pm 1.9	1.848
Sex				
Females	18	12	2.33 \pm 1.81	1.723
Males	15	13	3.6 \pm 1.99	1.881

statistically significant differences between geographic zones, with higher values in the south. High humidity and low temperature, such as those present in southern Chile (Chilean Meteorological Office, <http://www.meteochile.gob.cl>), are favorable conditions for the viability and abundance of gastropods (intermediate hosts of metastrongyloids) (Morgan et al., 2009; Brianti et al., 2014b). This could explain the higher prevalence of cardiorespiratory parasites found in southern guignas. Previous coprological studies in domestic cats have shown geographic differences in cardiorespiratory infection: in Santiago (central zone), Alcáño et al. (1992) found 10% prevalence of *A. abstrusus*, while López et al. (2006) and García (2014) did not find cardiorespiratory parasites, despite the large sample size they used (230 and 300 samples), while in Los Ríos Region (southern Chile), *A. abstrusus* showed prevalence of 10% (Oyarzún, 2013), 34.1% (Bonilla, 1980) and 38% (Escobar et al., 1984), whilst, *Eucoleus aerophilus* (Enoplida: Capillariidae) had a prevalence of 20% (Bonilla, 1980). This higher presence of cardiorespiratory helminths and the report of *Molineus* sp.

and capillarid worms only in guignas from the southern area, could explain the higher parasite richness we reported in southern Chile.

The higher intensity and abundance of infection we observed in guignas from the south, could be due to the fact that the species with higher intensity of infection, *U. stenocephala* and *T. leonina*, showed higher intensity in the southern zone. *Uncinaria stenocephala* had a total intensity of 8 helminths in the central zone and 232 in the south, while *T. leonina* showed a total intensity of 22 in the central zone and 178 helminths in the south. Both species can develop at low temperatures. The optimal temperature for *U. stenocephala* larval development is 20 °C, and their eggs can remain viable after a week at 0 °C (Bowman et al., 2002). *Toxascaris leonina* develops at higher temperatures than *U. stenocephala*, but it can also resist lower temperatures (Bowman et al., 2002). These characteristics and the auspicious humidity for the eggs and larvae, could make the presence and development of these species more feasible in southern Chile.

In this study there were no statistically significant differences in parasitism between sexes, but prevalence of infection and parasite richness were slightly higher in males. Larger home ranges and higher movement patterns of male guignas, who can move across the territories of different females (Dunstone et al., 2002; Sanderson et al., 2002; Schüttler et al., 2017), may increase male exposure to endoparasites.

Six guignas (20% of the guignas analyzed) carried 74% of the helminths found in this study (Fig. 6). These values resemble the 20/80 rule (Anderson and May 1985; Woolhouse et al., 1997) which postulates that less than 20% of the hosts harbor 80% of the helminth population, being responsible for most parasite transmission and persistence in the environment (Anderson and May 1985; Woolhouse et al., 1997). It would be interesting in the future, to analyze this rule in studies with a larger sample size and other carnivore species who could share endoparasites with guignas.

All the endoparasites that we found in guigna in this study have been previously reported in domestic cats (Bowman et al., 2002). The parasite species/genera in common between guignas and domestic cats may be shared between these two felids, and the domestic cat could represent a reservoir species for parasite transmission (Millán and Casanova, 2007; Otranto et al., 2015). This theory must be confirmed with molecular identification techniques, as has already been done for parasites shared by domestic and wild hosts (Epe et al., 1999; Millán and Blasco-Costa, 2012; Di Cesare et al., 2014; Hodžić et al., 2016). Cross-species transmission of feline immunodeficiency virus (FIV) and feline leukemia virus (FeLV) has been reported between free-roaming domestic cats and guignas in fragmented landscapes in Chile (Mora et al., 2015). This infectious agent transmission between domestic and wild species is probably facilitated by current habitat fragmentation and changes in land use (Napolitano et al., 2015), promoting proximity and contact between domestic cats and guignas, either by direct contact or by other species which could play the role of vectors or intermediate and paratenic hosts.

5. Conclusion

A large part of the analyzed guignas were parasitized, with higher parasite richness compared to previous studies, probably due to the analysis by necropsy that we used. Sex had no influence on parasitism, while geographic zone could be an important factor for infection, possibly due to favorable environmental conditions for the life cycles of the parasites.

For a more accurate identification at the species level of the parasites found in this study, we suggest conducting further analysis using molecular techniques, and also phylogenetic assessments of shared parasites between guigna and domestic cat, exploring the possibility of domestic-wildlife transmission.

The results of this study must be considered by veterinarians and researchers, to develop new research opportunities and detect if these helminths parasitize other wild and domestic carnivores in Chile,

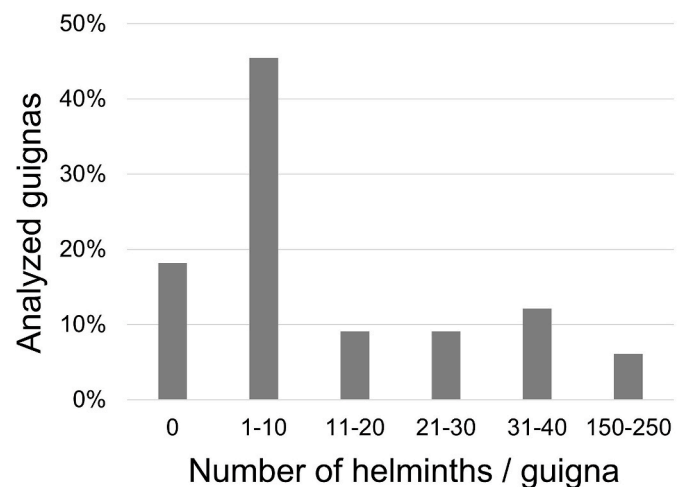


Fig. 6. Distribution of parasite infection intensity (total number of helminths per guigna) in the analyzed guignas from Chile.

clarifying their life cycle and epidemiology. These results will be valuable to inform conservation decisions for threatened carnivores like guignas and their habitat.

Declaration of competing interest

The authors declare no conflict of interest.

Acknowledgments

We are grateful to CONAF, especially Patricio Contreras, Patricia Barría; SAG, especially to Diego Ramírez, Rodrigo Villalobos and Luis Sepúlveda for logistic support and valuable samples. We thank Patricio Toro for technical support in laboratory work. Special thanks to Nicolás Gálvez, Jorge Valenzuela, Eduardo Silva, Brayan Zambrano, Javier Cabello, Gerardo Morales, Ricardo Pino, Daniel González, Nicole Salaberry, Angelo Espinoza, Diego Peñaloza, Mario Alvarado, Aitor Cevadanes, Frederick Toro, Paulette Abarca, Alfredo Catalán, Gabriella Svensson, Jaime Rau, Andrea Roa, Tomás Valdés and Manuel Valdés for their valuable support in sample collection. Our work was funded by CONICYT FONDECYT Iniciación 11150934 (CN), Morris Animal Foundation (MAF) D15ZO-413 (CN), National Geographic Society C309-15 (CN), Mohamed bin Zayed Species Conservation Fund 152510351 (CN), ANID PAI 77190064 (CN), the Wild Felid Association (IS).

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.ijppaw.2020.07.013>.

References

- Acosta-Jamett, G., Contreras, S., Muñoz, P., Briceño, C., Chirgwin, C., Hernández, F., 2018. Description of gastrointestinal parasitism through coprologic survey in Darwin's fox, *Lycalopex fulvipes* (Martin 1837), and kodkod, *Leopardus guigna* (Molina 1782), in Chiloé Island, Chile. *Gayana* 82, 160–165. <https://doi.org/10.4067/S0717-65382018000200160>.
- Acosta-Jamett, G., Simonetti, J.A., 2004. Habitat use by *Oncifelis guigna* and *Pseudalopex culpaeus* in a fragmented forest landscape in central Chile. *Biodivers. Conserv.* 13, 1135–1151. <https://doi.org/10.1023/B:BIOC.0000018297.93657.7d>.
- Alcaíno, H., Gorman, T., Larenas, I., 1992. Fauna endoparasitaria del gato doméstico en una zona urbana marginal de la Región Metropolitana de Chile. *Parasitol. al día* 16, 139–142.
- Álvarez, V., 1963. Echinococcosis silvestre en Chile. *Arch. Int. la Hídatid.* 21, 156–159.
- Álvarez, V., Rivera, G., Neghme, A., Schenone, H., 1970. Tiquinos en animales en Chile. *Bol. Chil. Parasitol.* 25, 83–86.
- Anderson, R.C., Chabaud, A.G., Willmott, S., 2009. *Keys to the Nematode Parasites of Vertebrates*. CAB International, London, UK.

- Anderson, R.M., May, R.M., 1985. Helminth infections of humans: mathematical models, population dynamics, and control. *Adv. Parasitol.* 24, 1–101. [https://doi.org/10.1016/S0065-308X\(08\)60561-8](https://doi.org/10.1016/S0065-308X(08)60561-8).
- Aranda, C., Serrano-Martínez, E., Tantaléan, M., Quispe, M., Casas, G., 2013. Identification and frequency of gastrointestinal parasites in captive wild cats in Peru. *Rev. investig. vet. Perú* 24 (3).
- Astorga, D., 2013. Comparación de la dieta de dos carnívoros silvestres, güiña (*Leopardus guigna*) y zorro chilla (*Pseudalopex griseus*), en el Parque Nacional Nahuelbuta, Región de la Araucanía. Universidad Austral de Chile, Chile.
- Bonilla, C., 1980. Estudio de la fauna helmintológica del gato en la ciudad de Valdivia. Universidad Austral de Chile, Chile.
- Bowman, D.D., Hendrix, C.M., Lindsay, D.S., Barr, S.C., 2002. *Feline Clinical Parasitology*, first ed. Iowa State University Press Blackwell Science Company, Iowa, EE. UU.
- Brianti, E., Gaglio, G., Napoli, E., Falsone, L., Giannelli, A., Annocchia, G., Varcasia, A., Giannetto, S., Mazzullo, G., Otranto, D., 2014a. Feline lungworm *Oeslerus rostratus* (Strongylida: filariidae) in Italy: first case report and histopathological findings. *Parasitol. Res.* 113, 3853–3857. <https://doi.org/10.1007/s00436-014-4053-z>.
- Brianti, E., Gaglio, G., Napoli, E., Falsone, L., Giannetto, S., Latrofa, M.S., Giannelli, A., Dantas-Torres, F., Otranto, D., 2013. Evidence for direct transmission of the cat lungworm *Troglostrongylus brevior* (Strongylida: Crenosomatidae). *Parasitology* 140, 821–824. <https://doi.org/10.1017/S0031182013000188>.
- Brianti, E., Giannetto, S., Dantas-Torres, F., Otranto, D., 2014b. Lungworms of the genus *Troglostrongylus* (strongylida: Crenosomatidae): neglected parasites for domestic cats. *Vet. Parasitol.* 202, 104–112. <https://doi.org/10.1016/j.vetpar.2014.01.019>.
- Bush, A.O., Lafferty, K.D., Lotz, J.M., Shostak, A.W., 1997. Parasitology Meets Ecology on Its Own Terms: margolis *et al.* Revisited. *J. Parasitol.* 83 <https://doi.org/10.2307/3284227>, 575.
- Correa, P., Roa, A., 2005. Relaciones tróficas entre *Oncifelis guigna*, *Lycalopex culpaeus*, *Lycalopex griseus* y *Tyto alba* en un ambiente fragmentado de la zona central de Chile. *Mastozoool. Neotrop.* 12, 57–60.
- Cortés, M., 2006. Identificación de formas reproductivas de parásitos gastrointestinales, en mamíferos nativos presentes en el Boin Zoo. Universidad de Concepción, Chile.
- Di Cesare, A., Otranto, D., Latrofa, M.S., Veronesi, F., Perrucci, S., Lalosevic, D., Gherman, C.M., Traversa, D., 2014. Genetic variability of *Eucoleus aerophilus* from domestic and wild hosts. *Res. Vet. Sci.* 96, 512–515. <https://doi.org/10.1016/j.rvsc.2014.03.018>.
- Diakou, A., Psalla, D., Migli, D., Di Cesare, A., Youlatos, D., Marcer, F., Traversa, D., 2016. First evidence of the European wildcat (*Felis silvestris silvestris*) as definitive host of *Angiostrongylus chabaudi*. *Parasitol. Res.* 115, 1235–1244. <https://doi.org/10.1007/s00436-015-4860-x>.
- Dobson, A., Lafferty, K.D., Kuris, A.M., Hechinger, R.F., Jetz, W., 2008. Homage to linnaeus: how many parasites? How many hosts? *Light Evol* 105, 11482–11489. <https://doi.org/10.1073/pnas.0803232105>.
- Dunstone, N., Durbin, L., Wyllie, I., Freer, R., Jamett, G.A., Mazzulli, M., Rose, S., 2002. Spatial organization, ranging behaviour and habitat use of the kodkod (*Oncifelis guigna*) in southern Chile. *J. Zool.* 257, 1–11. <https://doi.org/10.1017/S0952836902000602>.
- Durette-Desset, M.C., Boomker, J., Malan, F.S., 2000. *Molineus cati* n. sp. (Nematoda, Trichostrongylina, Molineoidea), a parasite of feral cats, *Felis catus* Linnaeus, 1758 in South Africa. *Onderstepoort J. Vet. Res.* 67, 173–177.
- Epe, C., Meuwissen, M., Stoye, M., Schnieder, T., 1999. Transmission trials, ITS2-PCR and RAPD-PCR show identity of *Toxocara canis* isolates from red fox and dog. *Vet. Parasitol.* 84, 101–112. [https://doi.org/10.1016/S0304-4017\(99\)00080-1](https://doi.org/10.1016/S0304-4017(99)00080-1).
- Escobar, R., Illanes, O., Fuentealba, I., Cubillos, V., 1984. Nematodiasis pulmonar en el gato doméstico. *Arch. Med. Vet.* 16, 47–49.
- Fernández, J., Villalba, C., 1984. Helminths parásitos de *Felis guigna* Molina, 1782. *Bol. Soc. Biol. Concepción, Chile* 55, 161–164.
- Figuroa, R.A., Corales, E.S., Rau, J.R., 2018. Prey of the güiña (*Leopardus guigna*) in an Andean mixed southern beech forest, southern Chile. *Stud. Neotrop. Fauna Environ.* 53, 211–218. <https://doi.org/10.1080/01650521.2018.1477032>.
- Fiorello, C.V., Robbins, R.G., Maffei, L., Wade, S.E., 2006. Parasites of free-ranging small canids and felids in the Bolivian chaco. *J. Zoo Wildl. Med.* 37, 130–134. <https://doi.org/10.1638/05-075.1>.
- Fleischütz, M.M., Gálvez, N., Pe'er, G., Davies, Z.G., Henle, K., Schüttler, E., 2016. Response of a small felid of conservation concern to habitat fragmentation. *Biodivers. Conserv.* 25, 1447–1463. <https://doi.org/10.1007/s10531-016-1118-6>.
- Freer, R.A., 2004. The Spatial Ecology of the Güiña (*Oncifelis Guigna*) in Southern. *Dep. Biol. Sci. University of Durham, Chile*.
- Gálvez, N., Guillera-Aroita, G., St John, F.A.V., Schüttler, E., Macdonald, D.W., Davies, Z.G., 2018. A spatially integrated framework for assessing socioecological drivers of carnivore decline. *J. Appl. Ecol.* 55, 1393–1405. <https://doi.org/10.1111/1365-2664.13072>.
- Gálvez, N., Hernández, F., Laker, J., Gilabert, H., Petitpas, R., Bonacic, C., Gimona, A., Hester, A., Macdonald, D.W., 2013. Forest cover outside protected areas plays an important role in the conservation of the Vulnerable güiña *Leopardus guigna*. *Oryx* 47, 251–258. <https://doi.org/10.1017/S0030605312000099>.
- García, M., 2014. Helminths y protozoos gastrointestinales de gatos (*Felis catus*) de la ciudad de Santiago. Universidad de Chile, Chile.
- Gherman, C.M., Ionică, A.M., D'Amico, G., Otranto, D., Mihalca, A.D., 2016. *Angiostrongylus chabaudi* (Biocca, 1957) in wildcat (*Felis silvestris silvestris*, S) from Romania. *Parasitol. Res.* 115, 2511–2517. <https://doi.org/10.1007/s00436-016-5032-3>.
- González-Acuña, D., Moreno, L., Ardiles, K., Flores, M., Duclos, M., Kinsella, M., 2010. Endoparasites of the kodkod, *Oncifelis guigna* (carnivora, felidae) in Chile. *Rev. Chil. Hist. Nat.* 83, 619–622. <https://doi.org/10.4067/S0716-078X2010000400015>.
- Hiestand, S.J., Nielsen, C.K., Jiménez, F.A., 2014. Epizootic and zoonotic helminths of the bobcat (*Lynx rufus*) in Illinois and a comparison of its helminth component communities across the American Midwest. *Parasite* 21. <https://doi.org/10.1051/parasite/2014005.4>.
- Hodžić, A., Alić, A., Klebić, I., Kadrić, M., Brianti, E., Duscher, G.G., 2016. Red fox (*Vulpes vulpes*) as a potential reservoir host of cardiorespiratory parasites in Bosnia and Herzegovina. *Vet. Parasitol.* 223, 63–70. <https://doi.org/10.1016/j.vetpar.2016.04.016>.
- Houpin, E., Mccarthy, G., Ferrand, M., Waal, T De, Neill, E.J.O., Zintl, A., 2016. Comparison of three methods for the detection of *Angiostrongylus vasorum* in the final host. *Vet. Parasitol.* 220, 54–58. <https://doi.org/10.1016/j.vetpar.2016.02.023>.
- Hudson, P.J., Dobson, A.P., Lafferty, K.D., 2006. Is a healthy ecosystem one that is rich in parasites? *Trends Ecol. Evol.* 21, 381–385. <https://doi.org/10.1016/j.tree.2006.04.007>.
- Khalil, L.F., Jones, A., Bray, R.A., 1994. *Keys to the Cestode Parasites of Vertebrates*. CAB International, Wallingford, Oxon, UK.
- Lange, M.K., Penagos-Tabares, F., Hirtzmann, J., Failing, K., Schaper, R., Van Bourgonie, Y.R., Backeljau, T., Hermosilla, C., Taubert, A., 2018. Prevalence of *Angiostrongylus vasorum*, *Aelurostrongylus abstrusus* and *Crenosoma vulpis* larvae in native slug populations in Germany. *Vet. Parasitol.* 254, 120–130. <https://doi.org/10.1016/j.vetpar.2018.03.011>.
- López, J., Abarca, K., Paredes, P., Inzunza, E., 2006. Parásitos intestinales en caninos y felinos con cuadros digestivos en Santiago, Chile. Consideraciones en Salud Pública. *Rev. Med. Chil.* 134, 193–200. <https://doi.org/10.4067/s0034-98872006000200009>.
- Marcogliese, D.J., 2005. Parasites of the superorganism: are they indicators of ecosystem health? In: *International Journal for Parasitology*, pp. 705–716. <https://doi.org/10.1016/j.ijpara.2005.01.015>.
- Martínez-Rondán, F.J., Ruiz de Ybáñez, M.R., López-Beceiro, A.M., Fidalgo, L.E., Berriatua, E., Lahat, L., Sacristán, I., Oleaga, A., Martínez-Carrasco, C., 2019. Cardiopulmonary nematode infections in wild canids: does the key lie on host-prey-parasite evolution? *Res. Vet. Sci.* 126, 51–58. <https://doi.org/10.1016/j.rvsc.2019.08.008>.
- Martínez-Rondán, F.J., Ruiz de Ybáñez, M.R., Tizzani, P., López-Beceiro, A.M., Fidalgo, L.E., Martínez-Carrasco, C., 2017. The American mink (*Neovison vison*) is a competent host for native European parasites. *Vet. Parasitol.* 247, 93–99. <https://doi.org/10.1016/j.vetpar.2017.10.004>.
- Millán, J., Blasco-Costa, I., 2012. Molecular evidence of shared hookworm *Ancylostoma tubaeformis* haplotypes between the critically endangered Iberian lynx and sympatric domestic cats. *Vet. Parasitol.* 186, 518–522. <https://doi.org/10.1016/j.vetpar.2011.11.027>.
- Millán, J., Casanova, J.C., 2007. Helminth parasites of the endangered Iberian lynx (*Lynx pardinus*) and sympatric carnivores. *J. Helminthol.* 81, 377–380. <https://doi.org/10.1017/S0022149X07869203>.
- Moleón, M.S., Kinsella, J.M., Moreno, P.G., Del Valle Ferreyra, H., Pereira, J., Pía, M., Beldomonte, P.M., 2015. New hosts and localities for helminths of carnivores in Argentina. *Zootaxa* 4057, 106–114. <https://doi.org/10.11646/zootaxa.4057.1.6>.
- Mora, M., Napolitano, C., Ortega, R., Poulin, E., Pizarro-Lucero, J., 2015. Feline immunodeficiency virus and feline leukemia virus infection in free-ranging guignas (*Leopardus guigna*) and sympatric domestic cats in human perturbed landscapes on Chiloé Island, Chile. *J. Wildl. Dis.* 51, 199–208. <https://doi.org/10.7589/2014-04-114>.
- Morgan, E.R., Jefferies, R., Krajewski, M., Ward, P., Shaw, S.E., 2009. Canine pulmonary angiostrongylosis: the influence of climate on parasite distribution. *Parasitol. Int.* 58, 406–410. <https://doi.org/10.1016/j.parint.2009.08.003>.
- Napolitano, C., Gálvez, N., Bennett, M., Acosta-Jamett, G., Sanderson, J., 2015. *Leopardus guigna*, güiña view on. www.iucnredlist.org/TheIUCNREDLISTOFTHREATENEDSPECIES™8235. <https://doi.org/10.2305/iucn.uk.2015-2.rlts.t15311a50657245.en>.
- Otranto, D., Brianti, E., Dantas-Torres, F., 2013. *Troglostrongylus brevior* and a nonexistent “dilemma”. *Trends Parasitol.* 29, 517–518. <https://doi.org/10.1016/j.pt.2013.09.001>.
- Otranto, D., Cantacessi, C., Dantas-Torres, F., Brianti, E., Pfeffer, M., Genchi, C., Guberti, V., Capelli, G., Deplazes, P., 2015. The role of wild canids and felids in spreading parasites to dogs and cats in Europe. Part II: helminths and arthropods. *Vet. Parasitol.* 213, 24–37. <https://doi.org/10.1016/j.vetpar.2015.04.020>.
- Oyarzún, J., 2013. *Pesquisa de nemátodos pulmonares en perros y gatos de las ciudades de Río Bueno y La Unión, Provincia del Ranco*. Universidad Austral de Chile, Chile.
- Penagos-Tabares, F., Lange, M.K., Chaparro-Gutiérrez, J.J., Taubert, A., Hermosilla, C., 2018. *Angiostrongylus vasorum* and *Aelurostrongylus abstrusus*: neglected and underestimated parasites in South America. *Parasites Vectors* 11, 1–13. <https://doi.org/10.1186/s13071-018-2765-0>.
- Penagos-Tabares, F., Lange, M.K., Vélez, J., Hirtzmann, J., Gutiérrez-Arboleda, J., Taubert, A., Hermosilla, C., Chaparro Gutiérrez, J.J., 2019. The invasive giant African snail *Lissachatina fulica* as natural intermediate host of *Aelurostrongylus abstrusus*, *Angiostrongylus vasorum*, *Troglostrongylus brevior*, and *Crenosoma vulpis* in Colombia. *PLoS Neglected Trop. Dis.* 13, 1–18. <https://doi.org/10.1371/journal.pntd.0007277>.
- Sanderson, J., Sunquist, M.E., Iriarte, A.W., 2002. Natural history and landscape-use of guignas (*Oncifelis guigna*) on isla grande de Chiloé, Chile. *J. Mammal.* 83, 608–613. [https://doi.org/10.1644/1545-1542\(2002\)083<0608.nhaluo>2.0.co;2](https://doi.org/10.1644/1545-1542(2002)083<0608.nhaluo>2.0.co;2).
- Schüttler, E., Klenke, R., Galuppo, S., Castro, R.A., Bonacic, C., Laker, J., Henle, K., 2017. Habitat use and sensitivity to fragmentation in America's smallest wildcat. *Mamm. Biol.* 86, 1–8. <https://doi.org/10.1016/j.mambio.2016.11.013>.

- Seguel, M., Gottdenker, N., 2017. The diversity and impact of hookworm infections in wildlife. *Int. J. Parasitol. Parasites Wildl.* 6, 177–194. <https://doi.org/10.1016/j.ijppaw.2017.03.007>.
- Smith, R.L., Smith, T.M., 2001. *Ecología*, fourth ed. Pearson Educacion, Madrid, España.
- Soulsby, E., 1987. *Parasitología y Enfermedades Parasitarias de los Animales Domésticos*, seventh ed. Nueva Editorial Interamericana, Mexico D.F., Mexico.
- Tagle, I., 1970. *Enfermedades parasitarias de los animales domésticos*, first ed. Editorial Andrés Bello, Santiago, Chile.
- Thompson, R.C.A., Lymbery, A.J., Smith, A., 2010. Parasites, emerging disease and wildlife conservation. *Int. J. Parasitol.* 40, 1163–1170. <https://doi.org/10.1016/j.ijpara.2010.04.009>.
- Traversa, D., Di Cesare, A., 2013. Feline lungworms: what a dilemma. *Trends Parasitol.* <https://doi.org/10.1016/j.pt.2013.07.004>.
- Traversa, D., Di Cesare, A., Conboy, G., 2010. Canine and feline cardiopulmonary parasitic nematodes in Europe: emerging and underestimated. *Parasites Vectors* 3, 1–22. <https://doi.org/10.1186/1756-3305-3-62>.
- Traversa, D., Lepri, E., Veronesi, F., Paoletti, B., Simonato, G., Diaferia, M., Di Cesare, A., 2015. Metastrongyloid infection by *Aelurostrongylus abstrusus*, *Troglostrongylus brevior* and *Angiostrongylus chabaudi* in a domestic cat. *Int. J. Parasitol.* 45, 685–690. <https://doi.org/10.1016/j.ijpara.2015.05.005>.
- Vallverdú, A., 2014. Descripción de parásitos gastrointestinales en güiña, zorro chilla, zorro culpeo y puma, mediante análisis coprológicos, en Parque Nacional Nahuelbuta, Región de la Araucanía. Universidad Austral de Chile, Chile.
- Varcasia, A., Brianti, E., Tamponi, C., Pipia, A.P., Cabras, P.A., Mereu, M., Dantas-Torres, F., Scala, A., Otranto, D., 2015. Simultaneous infection by four feline lungworm species and implications for the diagnosis. *Parasitol. Res.* 114, 317–321. <https://doi.org/10.1007/s00436-014-4207-z>.
- Varcasia, A., Tamponi, C., Brianti, E., Cabras, P.A., Boi, R., Pipia, A.P., Giannelli, A., Otranto, D., Scala, A., 2014. *Angiostrongylus chabaudi* Biocca, 1957: a new parasite for domestic cats? *Parasites Vectors* 7. <https://doi.org/10.1186/s13071-014-0588-1>, 588.
- Veronesi, F., Traversa, D., Lepri, E., Morganti, G., Vercillo, F., Grelli, D., Cassini, R., Marangi, M., Iorio, R., Ragni, B., Di Cesare, A., 2016. Occurrence of lungworms in European wildcats (*Felis silvestris silvestris*) of central Italy. *J. Wildl. Dis.* 52, 270–278. <https://doi.org/10.7589/2015-07-187>.
- Vieira, F.M., Muniz-Pereira, L.C., de Souza Lima, S., Neto, A.H.A.M., Guimarães, E.V., Luque, J.L., 2013. A new metastrongyloidean species (Nematoda) parasitizing pulmonary arteries of *Puma (Herpailurus) yagouaroundi* (É. Geoffroy, 1803) (Carnivora: felidae) from Brazil. *J. Parasitol.* 99, 327–331. <https://doi.org/10.1645/GE-3171.1>.
- Wolffhügel, K., 1949. ¿Es autóctono el *Diphyllobothrium* en Chile? *Bol. Soc. Biol. Concepc.* 24, 85–89.
- Woolhouse, M.E.J., Dye, C., Etard, J.-F., Smith, T., Charlwood, J.D., Garnett, G.P., Hagan, P., Hii, J.L.K., Ndhlovu, P.D., Quinnell, R.J., Watts, C.H., Chandiwana, S.K., Anderson, R.M., 1997. Heterogeneities in the transmission of infectious agents: implications for the design of control programs. *Proc. Natl. Acad. Sci. Unit. States Am.* 94, 338–342. <https://doi.org/10.1073/PNAS.94.1.338>.
- Zúñiga, A., Quintana, V., Fierro, A., 2005. Relaciones tróficas entre depredadores en un ambiente fragmentado del sur de Chile. *Gestión Ambient* 11, 31–42.