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LETTER TO EDITOR

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A natural BACE1 and GSK3 β dual inhibitor Notopterol effectively ameliorates the cognitive deficits in APP/PS1 Alzheimer's mice by attenuating amyloid- β and tau pathology

Dear Editor,

BACE1 and GSK3 β are the key targets for A β production and tau phosphorylation, respectively. A β activated the phosphorylation of GSK3 β , resulting in the increase of tau phosphorylation.¹ In addition, tau protein stimulated A β toxicity, formed tau Fyn-A β pathway, and increased postsynaptic Fyn level and NMDA receptor sensitivity, thus making neuronal dendrites more sensitive to A β toxicity.² Given the cross-talk between BACE1 and GSK3 β , a welldesigned multitarget-directed ligand (MTDL) capable of targeting these two enzymes simultaneously may be a valuable and promising therapeutic strategy.³

So far, only two well-studied scaffold derivatives, triazinone scaffold and curcumin scaffold, have been reported as potential BACE1 and GSK3 β dual inhibitors.^{4,5} Getting benefit from virtual screening in silico, we identified a furan coumarin (Notopterol) with simultaneously inhibitory activity on BACE1 and GSK3 β from *Notopterygium incisum* (Table S1). This study is the first to systematically elucidate the benefits of BACE1 and GSK3 β dual inhibitors on the pathological mechanism of Alzheimer's disease (AD) in vitro and in vivo.

Notopterol was docked into the binding site of the proteins, and the predicted binding modes at the two targets are shown in Figure 1A and D. In BACE1, the hydroxyl group of the Notopterol fatty chain interacted with Asp32, Asp228, and Thr231, and formed a water bridge with Gly230 (Figure S1). For GSK3 β , Notopterol bound in the hinge region of the kinase and establishes two strong hydrogen-bonding interactions with the backbone of Asp133 and Tyr134. To evaluate the binding stability of protein ligand complexes, we have performed molecular dynamics (MD) simulations for 100 ns. The RMSD values of protein skeleton remained stable at about 2–3 Å during the simulation (ligand fit on protein). These

results showed that Notopterol can bind to protein in a stable state in the process of molecular dynamics simulation. We also tested the inhibitory effect of Notopterol on BACE1 and GSK3 β enzymes: Notopterol showed moderate inhibitory against BACE1 (IC₅₀: 26.01 μ M) and strong inhibitory against GSK3 β (IC₅₀: 1 μ M).

HEK APPswe293T cells stably overexpress human APP-695 harboring the Swedish double mutation (APPswe).⁶ The secretion of $A\beta$ in HEK APPswe293T cells is much higher than that of wild type cells, so it is an ideal cell model for screening drugs against AD.⁷ Figure 1H demonstrates that Notopterol could inhibit the production of $A\beta$. Okadaic acid induced tau hyperphosphorylation in SH-SY5Y cell is usually used as a cell model for tau pathology.⁸ Western blot results showed that Notopterol decreased the expression of ADAM17 and BACE1 in HEK APPswe293T cells, as well as the phosphorylation of tau in SH-SY5Y cell exposed in Okadaic acid (Figure 1I and J).

To assess whether Notopterol treatment had any beneficial effects on the cognition of APP/PS1 mice, we performed Morris water maze assay. Compared with APP/PS1 mice, the escape latency of Notopterol-treated mice was significantly decreased (Figures 2A and B). The levels of A β 40, A β 42, and A β in the hippocampus and cortex of were determined by enzyme linked immunosorbent assay (ELISA) assay. As shown in Figure 2C, Notopterol significantly reduced the levels of A β 40, A β 42, and A β in hippocampus and cortex compared to that of transgenic mice. In addition, we also verified that the A β 42 level in cortex was positively correlated with the escape latency of mice (Figure 2D), which is consistent with previous studies.⁹ Given Notopterol is a BACE1 and GSK3 β dual inhibitor, we first investigated the deposition of amyloid plaques and the phosphorylated tau in the hippocampus and cortex. The immunohistochemistry results showed that the

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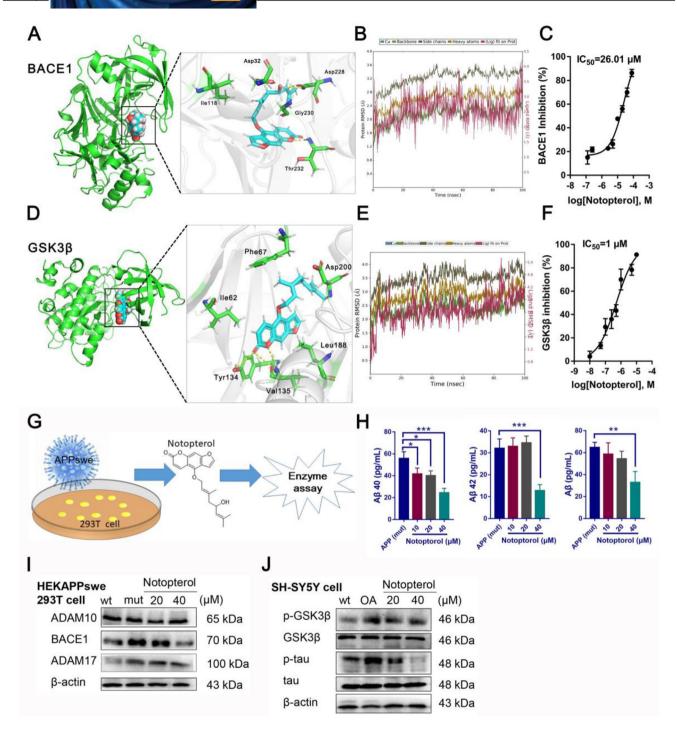


Figure 1 Binding modes of Notopterol with BCAE1 (5CLM) and GSK3 β (2OW3) and inhibiting the generation of A β and phosphorylated tau. A, Predicted conformation of Notopterol combined with BACE1. B, RMSD curve of Notopterol/BACE1 complex. C, Notopterol inhibited the activities of BACE1 enzyme. D, Predicted conformation of Notopterol combined with GSK3 β E, RMSD curve of Notopterol/GSK3 β complex. F, Notopterol inhibited the activities of GSK3 β enzyme. G, The experimental protocol for measuring A β levels by enzyme linked immunosorbent assay (ELISA). H, A β 40, A β 42, and total A β in HEKAPPswe239T cells with or without Notopterol. I, Western blot of ADAM10, BACE1, and ADAM17 in APPswe239T cells. J, Western blot of p-GSK3 β , GSK3 β , p-tau, and tau in Okadaic acid-treated SH-SY5Y cells. The error bars represent the SD. (*P < 0.05, **P < 0.01 and ***P < 0.001, other comparisons were not significant; One-way analysis of variance followed by Turkey's post hoc multiple-comparisons test)

3 of 5

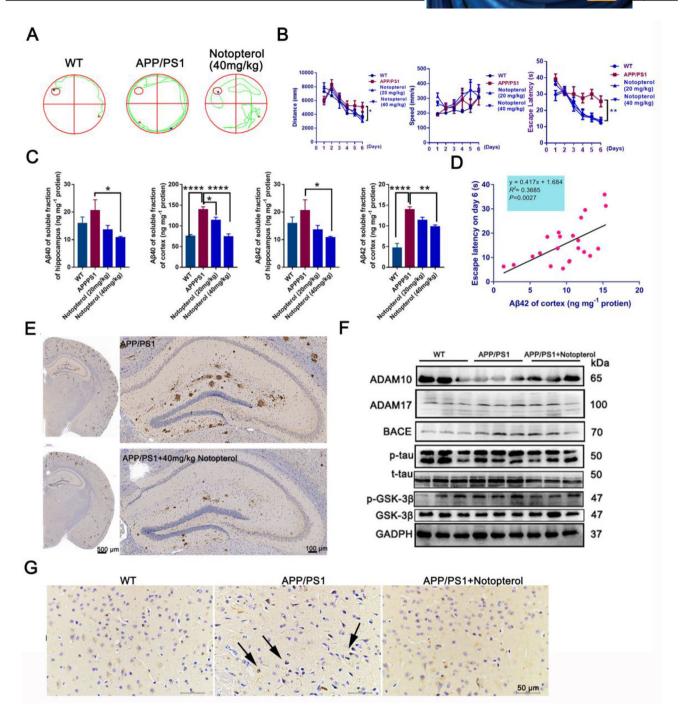


Figure 2 Notopterol improves the cognitive decline in APP/PS1 mice. A, Representative water maze traces of WT mice (left), APP/PS1 mice (middle), and Notopterol-treated (right) APP/PS1 mice on day 6. B, Analysis of the MWM test of WT, Notpterol-treated, or without treated APP/PS1 mice. C, The levels of A β 40 and A β 42 in hippocampal and cortes in APP/PS1 mice. D, The level of A β 42 in the cortex of APP/PS1 mice was positively correlated with memory impairment. E, Immunohistochemistry staining of deposited A β in APP/PS1 mice and Notopetrol (40 mg/kg) treated APP/PS1 mice; scale bars, 100 μ m and 500 μ m. F, Western blot of ADAM10, BACE1, ADAM17, p-GSK3 β , GSK3 β , p-tau, and tau in hippocampus of APP/PS1 mice and Notopetrol-treated (40 mg/kg) APP/PS1 mice. Quantitative data are shown in Figure S4. G, Immunohistochemistry staining of phosphorylated tau in cortex of APP/PS1 mice and Notopetrol-treated (40 mg/kg) APP/PS1 mice; scale bars, 50 μ m. The error bars represent the SD. (*P < 0.05, **P < 0.01 and ***P < 0.001, other comparisons were not significant; One-way analysis of variance followed by Turkey's post hoc multiple-comparisons test)

deposition of $A\beta$ plaques was significantly decreased in the hippocampus of Notopterol-treated APP/PS1 transgenic mice (Figure 2E and G), as well as the phosphorylated tau in cortex. Furthermore, we investigated the proteins expression of the $A\beta$ and tau pathways. We found that Notopterol can regulate the protein expression of ADAM10-BACE1-ADAM7 and GSK3 β -tau signaling pathway in hippocampus (Figure 2F). These results indicated that the advantage of BACE1 and GSK3 β dual inhibitor is reducing both the amyloid plaques and hyperphosphorylatic tau, which can improve the ability of AD mice to learn.

In summary, here we reported is the discovery of Notopterol as the natural small molecule capable of simultaneously inhibiting BACE1 and GSK3 β . We have demonstrated that inhibition of the activity of BACE1 and GSK3 β by Notopterol can effectively repair the pathophysiological and cognitive impairment of AD caused by abnormal A β accumulation and tau hyperphosphorylation. Given the complexity of AD pathology, BACE1 and GSK3 β dual inhibitor may be a promising treatment strategy.

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ETHICS APPROVAL AND CONSENT TO PARTICIPATE

All animal experiments were carried out in accordance with the National Institutes of Health guide for the care and use of laboratory animals (NIH Publications No. 8023, revised 1978). This study was approved by the Ethics Committee of the Institutional Animal Care and Use Committee of Shenyang Pharmaceutical University.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

AUTHOR CONTRIBUTIONS

Qingchun Zhao and Huiyuan Gao designed this study, Xiaowen Jiang performed this research and wrote the paper. Jingda Li and Qiong Wu assisted in isolating animal tissue. Jingda Li and Wenwu Liu performed Morris water maze test and analyzed data. Hongyuan Lu and Zihua Xu performed docking analysis. Qinglong Qiao and Haotian Zhang analyzed data. Xiaowen Jiang^{1,2,3} Hongyuan Lu^{3,4} Jingda Li^{1,2,3} Wenwu Liu^{1,2,3} Qiong Wu^{2,4} Zihua Xu^{2,3} Qinglong Qiao⁵ Haotian Zhang⁴ Huiyuan Gao^{1,3} Oingchun Zhao^{1,2,3,4}

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5 of 5

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