

Research Article

HOTTIP polymorphism may affect gastric cancer susceptibility by altering *HOTTIP* expression

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Background: Non-coding RNA polymorphisms can affect disease risk and prognosis by influencing gene expression. Here, we first investigated the association between single nucleotide polymorphisms (SNPs) of long non-coding RNA (lncRNA) *HOTTIP* and gastric cancer risk/prognosis. **Methods:** A total of five *HOTTIP* SNPs among 627 gastric cancer cases and 935 controls were tested by Kompetitive Allele Specific PCR (KASP) assay. The functional SNPs underwent eQTL analysis and the expression of *HOTTIP* was assessed by quantitative RT-PCR. **Results:** The rs2067087 and rs3807598 SNPs of *HOTTIP* increased susceptibility to gastric cancer (rs2067087: dominant model, $P=0.008$, odds ratio (OR) = 1.35; rs3807598: recessive model, $P=0.037$, OR = 1.29). Both *HOTTIP* rs2067087 and rs3807598 could affect the expression of mature lncRNA ($P=0.003$ and $P=0.032$, respectively). **Conclusion:** The rs2067087 and rs3807598 SNPs of *HOTTIP* are associated with gastric cancer risk, possibly by affecting the expression of mature *HOTTIP*.

Introduction

Gastric cancer is the second most fatal type of tumor [1]. Patients with gastric cancer often respond poorly to treatment because of the heterogeneity of gastric cancer and limited treatment methods [2,3]. Therefore, the study of factors involved in early detection of gastric cancer and elucidation of the underlying mechanisms of gastric cancer pathogenesis are of significant interest.

Long non-coding RNAs (lncRNAs) are defined as transcripts containing more than 200 nucleotides, which are research hotspots particularly in oncology because of their wide biological regulatory functions [4]. lncRNAs play important roles in cancer pathogenesis by affecting diverse biological processes, including transcription, post-transcriptional regulation and epigenome [5–8]. So far, three key lncRNAs (*HOTTIP*, *HOTAIR*, and *H19*) have been reported to be candidate genes involved in carcinogenesis and potential therapeutic targets [9–14]. Among them, *HOTTIP*, transcribed from the 5' tip of the *HOXA* cluster, is associated with various tumors including gastric cancer [15,16]. Recently, two studies have shown that *HOTTIP* is significantly overexpressed in gastric cancer cell lines and acts as a predictive factor for poor prognosis, suggesting that it may be a potential novel diagnostic and prognostic biomarker [17,18].

Genetic studies have shown that several single nucleotide polymorphisms (SNPs) are involved in increased susceptibility to gastric cancer [19–21]. Hu et al. showed that the functional *HOTTIP* rs1859168 A>C polymorphism might decrease the risk of pancreatic cancer [22]. However, the role of *HOTTIP* polymorphism in gastric cancer has not been investigated. With this in mind, we conducted the present study to identify functional SNPs in *HOTTIP* to determine any correlation of *HOTTIP* polymorphisms with gastric cancer susceptibility and prognosis, aiming to explore whether polymorphisms could affect the expression of mature *HOTTIP*.

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Materials and methods

Patients and study design

This research project was approved by the Ethical Committee of the First Hospital of China Medical University and written informed consent was obtained. The study consisted of risk and prognosis studies, followed by eQTL analysis by quantitative RT-PCR for a step-by-step screening to find SNPs functional for gastric cancer etiology. This case-control study enrolled 1562 participants, including 627 gastric cancer patients and 935 matched controls. The patients received surgery for gastric cancer at the First Hospital of China Medical University between 2002 and 2013. The participants who had surgery were diagnosed with gastric cancer by pathological confirmation based on WHO classification. Then, 183 patients were diagnosed with intestinal-type gastric cancer and 312 with diffused-type gastric cancer according to Lauren classification. A total of 935 frequency-matched controls were recruited from a health-screening program from Zhuanghe, Liaoning Province, China, between 2002 and 2012 [23]. A questionnaire survey was conducted to collect information of smoking and drinking. We performed a follow-up visit for gastric cancer patients whose medical record was completed thereafter. The median survival time (MST) was 36 months and the last follow-up day was 1 July 2017.

Selected SNP sites and genotyping

We selected polymorphic sites based on previous publication [24], which was shown in Supplementary Materials (Supplementary Figure S1 and Table S1). A total of five SNPs covering the *HOTTIP* gene were selected. Genomic DNA was extracted by a previously published method [25]. The genotyping assay was performed by Gene Company (Shanghai, China), using allele-specific PCR and Kompetitive Allele Specific PCR (KASP) reagents (LGC Genomics, Hoddesdon, U.K.) as previously described [24]. We repeated some samples for quality control, and the concordance rate reached more than 99% [24].

Quantitative RT-PCR by eQTL analysis for *HOTTIP* expression and functional SNP identification

Approximately 50 mg total RNA was isolated from 39 gastric cancer specimens and 27 related cancer-free tissue using TRIzol reagent (Life Technologies, Carlsbad, CA, U.S.A.) as described in previous reports [24,26] shown in Supplementary Materials. The *HOTTIP* primer sequences were F: 5'-CGACTGGGTCCCTCCTCAC-3' and R: 5'-GGCTCCTGCCGTCTTTTCT-3'. Analysis of eQTLs was performed by analyzing the effect of the polymorphisms on the lncRNA expression.

Statistical analysis

Inter-group differences in sex variability and the Hardy-Weinberg equilibrium were compared by the Chi-squared (χ^2) test, and the analysis of variance was performed for age variability. To evaluate the association between gene polymorphisms and gastric cancer risk, multivariate logistic regression adjusted for age and sex was used to calculate odds ratios (ORs) and their 95% confidence intervals (95% CIs). The haplotype of each gene was analyzed by SHEsis software [27]. The Student's *t* test was used to test the differences in relative mRNA levels between the two groups. All statistical tests were two-sided and a *P*-value <0.05 was considered to be statistically significant.

Results

The association of *HOTTIP* SNPs with gastric cancer risk

The demographic information was presented in Supplementary Table S2. No significant difference was observed in either age or sex in gastric cancer cases and controls ($P>0.05$). Four SNPs (rs3807598, rs17501292, rs2067087, and rs17427960) of *HOTTIP* were accorded with the Hardy-Weinberg test ($P>0.05$), while rs78248039 was excluded because only the AA genotype was detected. Therefore, four SNPs were involved in the subsequent analysis.

By logistic regression analysis adjusted for age and gender, we found that two SNPs in *HOTTIP*, rs2067087 and rs3807598, were associated with gastric cancer risk. The dominant model of *HOTTIP* rs3807598 showed a 1.29-fold increased gastric cancer risk ($P=0.037$, 95% CI = 1.02–1.63) while the recessive model of *HOTTIP* rs2067087 showed a 1.35-fold increased gastric cancer risk ($P=0.008$, 95% CI = 1.08–1.68, Table 1). In addition, stratified analysis indicated that the patients with rs17501292 TG genotype were more likely to develop gastric cancer compared with the patients with TT genotype in the *H. pylori*-positive subgroup ($P=0.022$, OR = 4.12, 95% CI = 1.23–13.77, Supplementary Table S3). Female patients with rs2067087 CC *HOTTIP* genotype were more likely to have gastric cancer compared with the rs2067087 GG genotype ($P=0.027$, OR = 1.85, 95% CI = 1.07–3.19, Supplementary Table S3).

Table 1 The association of *HOTTIP* polymorphisms and gastric cancer risk

Gene	Chr. Pos.	SNP ¹	Loc.	Genotype	Controls (%)	Cases (%)	P ²	OR (95% CI)	P _{HWE}	
HOTTIP	7p15.2	rs3807598	Exon 2	CC	257 (28.0)	143 (23.2)		1 (see footnote)	0.147	
				CG	454 (49.4)	326 (52.9)	0.043	1.29 (1.01–1.66)		
				GG	208 (22.6)	147 (23.9)	0.108	1.27 (0.95–1.71)		
				CG+GG vs. CC				0.037	1.29 (1.02–1.63)	
				G vs. C				0.105	1.13 (0.98–1.30)	
		rs17501292	Exon 2	TT	853 (91.2)	567 (90.4)		1 (see footnote)	0.676	
				TG	80 (8.6)	59 (9.4)	0.558	1.11 (0.78–1.58)		
				GG	2 (0.2)	1 (0.2)	0.846	0.79 (0.07–8.78)		
		rs2067087	Exon 2	GG	201 (21.6)	114 (18.4)		1 (see footnote)	0.390	
				CG	467 (50.2)	291 (47.0)	0.498	1.10 (0.84–1.44)		
				CC	262 (28.2)	214 (34.6)	0.014	1.44 (1.08–1.94)		
				CC vs. GC+GG				0.008	1.35 (1.08–1.68)	
				C vs. G				0.009	1.22 (1.05–1.40)	
		rs17427960	Intron 2	CC	195 (21.2)	120 (19.8)		1 (see footnote)	0.122	
				AC	446 (48.4)	278 (45.9)	0.926	1.01 (0.77–1.33)		
AA				280 (30.4)	208 (34.3)	0.191	1.21 (0.91–1.62)			
	rs78248039	Exon 3	AA	877 (100)	576 (100)		1 (see footnote)	NA		

Abbreviations: Chr. Pos., chromosomal position; Loc., localization; NA, not available; P_{HWE}, P-value for Hardy–Weinberg equilibrium.

¹, The sort order was according to the SNP location in its genes from 5' starting to 3' ends.

², P-value was calculated by adjusted age and sex.

Table 2 Association of *HOTTIP* polymorphisms with the risk of intestinal-type and diffuse-type gastric cancer

Gene	Variables	CON	Intestinal-type GC	Diffuse-type GC	Intestinal-type GC vs CON		Diffuse-type GC vs CON		
					P	OR (95% CI) ¹	P	OR (95% CI) ¹	
HOTTIP	rs3807598	CC	257 (28.0)	44 (24.3)	71 (23.3)		1 (see footnote)	1 (see footnote)	
		CG	454 (49.4)	95 (52.5)	156 (51.1)	0.301	1.23 (0.83–1.82)	0.189	1.24 (0.90–1.71)
		GG	208 (22.6)	42 (23.2)	78 (25.6)	0.512	1.17 (0.73–1.86)	0.097	1.37 (0.94–1.98)
	rs17501292	TT	853 (91.2)	160 (87.0)	284 (91.0)		1 (see footnote)	1 (see footnote)	
		TG	80 (8.6)	23 (12.5)	28 (9.0)	0.111	1.50 (0.91–2.48)	0.784	1.07 (0.68–1.67)
		GG	2 (0.2)	1 (0.5)	0 (0)	0.492	2.41 (0.20–29.64)	NA	NA
	rs2067087	GG	201 (21.6)	35 (19.1)	53 (17.3)		1 (see footnote)	1 (see footnote)	
		CG	467 (50.2)	76 (41.5)	153 (50)	0.708	0.92 (0.59–1.43)	0.219	1.25 (0.88–1.78)
		CC	262 (28.2)	72 (39.4)	100 (32.7)	0.056	1.55 (0.99–2.42)	0.046	1.48 (1.01–2.16)
	rs17427960	CC	195 (21.2)	34 (19.1)	59 (19.5)		1 (see footnote)	1 (see footnote)	
		AC	446 (48.4)	78 (43.8)	146 (48.4)	0.925	0.98 (0.63–1.52)	0.624	1.09 (0.77–1.54)
		AA	280 (30.4)	66 (37.1)	97 (32.1)	0.211	1.34 (0.85–2.12)	0.417	1.17 (0.80–1.70)
	rs78248039	AA	877 (100)	171 (100)	285 (100)	NA	NA	NA	NA

Abbreviations: CON, control; GC, gastric cancer; NA, not available. The significance values were showed as bold font.

¹, Using Logistic Regression adjusted by gender and age.

Regarding the Lauren classification, rs2067087 might be associated with the susceptibility to diffuse-type gastric cancer, as our analysis suggested that the CC genotype could increase the risk of diffuse-type gastric cancer compared with the GG genotype ($P=0.046$, OR = 1.48, 95% CI = 1.01–2.16, Table 2). However, in the further haplotype analysis,

Table 3 The association of haplotype of *HOTTIP* and gastric cancer risk

Haplotype	Case (%)	Control (%)	P	OR (95% CI)
HOTTIP				
CTGC	475.65 (0.441)	763.41 (0.472)	0.086	0.87 (0.74–1.02)
GTCA	501.79 (0.465)	698.72 (0.432)	0.086	1.15 (0.98–1.36)

Using SHEsis software to analyze (<http://analysis.bio-x.cn/>).

Table 4 Univariate and multivariate Cox proportional hazard analyses for the association of *HOTTIP* polymorphisms and gastric cancer

Variables	SNP	genotype	All GC, n (%)	Deaths, n	MST ¹ (M)	Univariate		Multivariate ³	
						P-value	Hazard ratio (95% CI)	P-value	Hazard ratio (95% CI)
HOTTIP	rs3807598		n=297	n=122					
		CC	63 (21.2)	26 (21.3)	55.3 ²		1 (see footnote)		1 (see footnote)
		CG	163 (54.9)	67 (54.9)	56.3 ²	0.802	1.06 (0.67–1.67)	0.747	1.08 (0.68–1.71)
		GG	71 (23.9)	29 (23.8)	68.0	0.990	1.00 (0.59–1.71)	0.703	1.11 (0.65–1.91)
			n=299	n=121					
		TT	273 (91.3)	110 (90.9)	57.5 ²		1 (see footnote)		1 (see footnote)
	rs17501292	TG	26 (8.7)	11 (9.1)	52.6 ²	0.869	1.05 (0.57–1.20)	0.429	0.78 (0.42–1.45)
		GG	NA	NA	NA	NA	NA	NA	NA
			n=297	n=122					
	rs2067087	CC	97 (32.7)	44 (36.1)	68.0		1 (see footnote)		1 (see footnote)
		CG	151 (50.8)	58 (47.5)	58.7 ²	0.321	0.82 (0.55–1.21)	0.213	0.78 (0.53–1.15)
		GG	49 (16.5)	20 (16.4)	54.9 ²	0.556	0.85 (0.50–1.45)	0.610	0.87 (0.51–1.48)
	rs17427960		n=295	n=120					
		AA	98 (33.2)	42 (35)	68.0		1 (see footnote)		1 (see footnote)
		AC	144 (48.8)	57 (47.5)	58.0 ²	0.678	0.92 (0.62–1.37)	0.644	0.91 (0.61–1.36)
		CC	53 (18.0)	21 (17.5)	55.2 ²	0.745	0.92 (0.54–1.55)	0.687	0.90 (0.53–1.52)

Abbreviations: HR, hazard rate; NA: not available.

¹, MST (months).

², Mean survival time was provided when MST could not be calculated.

³, Multivariate survival analysis was carried out by adding the polymorphisms variable to the clinicopathological parameters with $P < 0.05$.

no haplotype of *HOTTIP* was found to be correlated with gastric cancer risk (Table 3).

The association of *HOTTIP* SNPs with clinical parameters and prognosis of gastric cancer

We first analyzed the association of *HOTTIP* SNPs with clinical parameters of gastric cancer (Supplementary Table S4). In the univariate analysis of the clinical parameters and survival of gastric cancer patients, we found that the macroscopic type, TNM stage, depth of invasion, and lymphatic metastasis were associated with the survival time of gastric cancer patients ($P < 0.001$, Supplementary Table S5). All the four clinical parameters were adjusted in the multivariate analysis of *HOTTIP* SNPs and gastric cancer prognosis. However, no statistical correlation between any of these four *HOTTIP* SNPs and gastric cancer prognosis were observed (Table 4).

eQTL analysis

The impact of polymorphisms in *HOTTIP* on *HOTTIP* expression level was also analyzed. The *HOTTIP* expression level was significantly higher in the samples with heterozygous genotype than in wildtype samples for both *HOTTIP* rs3807598 and rs2067087 ($P = 0.032$ and $P = 0.003$, respectively, Table 5 and Figure 1).

Discussion

Previous studies have demonstrated that overexpression of lncRNA *HOTTIP* in gastric cancer promotes tumor invasion and results in poor prognosis [17]. However, no investigation has focused on *HOTTIP* polymorphisms, which

Table 5 Differences of HOTTIP mRNA levels in different genotypes in gastric cancer and non-cancer tissues

Variable	Non-cancer tissue				Cancer tissue			
	<i>n</i>	ΔC_t (Mean \pm SD)	Normalized $2^{-\Delta\Delta C_t}$	<i>P</i>	<i>n</i>	ΔC_t (Mean \pm SD)	Normalized $2^{-\Delta\Delta C_t}$	<i>P</i>
HOTTIP	27	10.73 \pm 2.28	1 (0.21, 4.86)	(see footnote)	39	11.59 \pm 2.65	0.55 (0.09, 3.46)	0.331
Effect of HOTTIP rs3807598 genotypes on HOTTIP								
CC	3	11.21 \pm 3.13	1 (0.11, 8.75)	(see footnote)	6	11.82 \pm 2.20	1 (0.22, 5.94)	(see footnote)
GC	12	9.29 \pm 1.12	3.78 (1.74, 8.22)	0.784	17	10.57 \pm 3.02	2.38 (0.29, 19.29)	0.032
GG	7	10.25 \pm 1.26	1.94 (3.24, 4.65)	0.370	7	13.85 \pm 2.21	0.24 (0.05, 1.13)	0.125
GC+GG vs. CC	19	10.86 \pm 2.59	1.27 (0.21, 7.67)	0.871	24	11.53 \pm 3.16	1.22 (0.14, 10.93)	0.097
Effect of HOTTIP rs2067087 genotypes on HOTTIP								
GG	2	9.47 \pm 1.59	1 (0.33, 3.01)		4	12.78 \pm 1.96	1 (0.26, 3.89)	(see footnote)
GC	10	10.95 \pm 2.67	0.36 (0.06, 2.28)	0.467	17	9.98 \pm 2.50	6.96 (1.23, 39.40)	0.003
CC	10	10.70 \pm 2.53	0.42 (0.07, 2.46)	0.788	11	10.51 \pm 2.61	4.82 (0.79, 29.45)	0.907
CC+GC vs. GG	10	10.57 \pm 2.56	0.47 (0.08, 2.75)	0.382	28	11.18 \pm 2.81	3.63 (0.52, 25.46)	0.247

Abbreviation: NA, not available. *P*, the statistical analysis for the effect of genotype to phenotype was used two-independent sample *t* test, and for the combination of genotype to phenotype was used ANOVA analysis. Significance values are shown in bold.

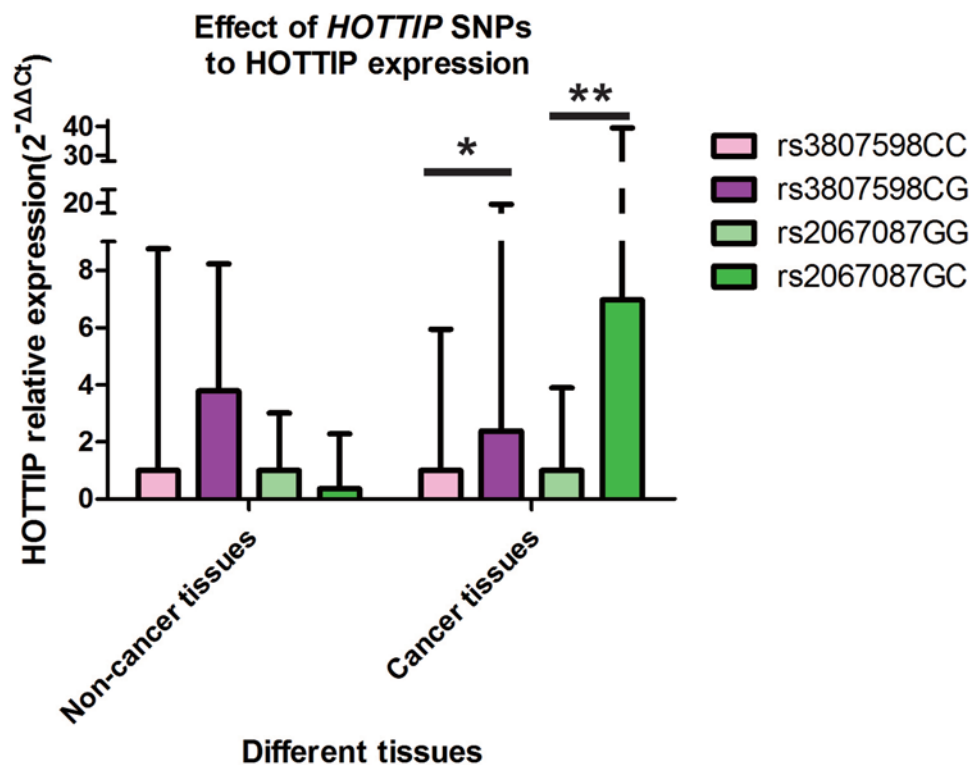


Figure 1. The effect of HOTTIPrs3807598 and rs2067087 polymorphisms on the HOTTIP corresponding mRNA expression
 P*=0.032, *P*=0.003.

can affect HOTTIP expression in gastric cancer. The current study is the first report on the association between HOTTIP polymorphisms and gastric cancer. Here, we identified two functional SNPs associated with gastric cancer susceptibility that affect the expression of mature HOTTIP.

One of the two functional HOTTIP SNPs was rs2067087 with the minor allele of C. We found that rs2067087 statistically increased the risk of gastric cancer, and that female individuals carrying the rs2067087 CC genotype

were more susceptible to gastric cancer compared with those with GG genotype. This suggested that determining the rs2067087 genotype might be a potentially meaningful test in gastric cancer screening, especially for females. In a previous study, we found that the variant genotype of rs2067087 could increase hepatocellular cancer (HCC) risk [24]. The rs2067087 polymorphism is located in an exon of the *HOTTIP* gene and can combine with some functional proteins. Through a CHIP-Seq experiment, this SNP was found to bind to SUZ12 protein [28], which promoted gastric cancer cell invasion [29]. Although the detailed mechanisms require further study, the rs2067087 polymorphism may be a functional SNP participating in gastric carcinogenesis and a candidate biomarker for the prediction of gastric cancer.

Another *HOTTIP* SNP detected to play a role in gastric carcinogenesis was rs3807598. It was suggested that the heterozygous genotype had a significantly increased risk of gastric cancer, and a significant association under the dominant model (GG+CG/CC) between rs3807598 and the risk of gastric cancer was observed. Although no statistical difference in the stratified analysis of rs3807598 was found considering gender, age, cigarette or alcohol consumption, and *Helicobacter pylori* infection status, this allele might serve as a risk marker of gastric cancer. Further study focused on the detailed molecular mechanism of rs3807598 in gastric cancer is required.

We also carried out genotype–phenotype analysis and observed that both the heterozygous genotypes of rs3807598 and rs2067087 associated with gastric cancer susceptibility contributing to higher *HOTTIP* expression than wildtype SNPs. A previous study showed that the rs1859168 A>C polymorphism regulated *HOTTIP* expression and reduced the risk of pancreatic cancer in a Chinese population [22]. In our study, we identified two SNPs (rs3807598 and rs2067087) in *HOTTIP* involved in gastric cancer susceptibility and the formation of mature *HOTTIP*. In recent years, Harrow et al. showed that the rs2067087 SNP could combine with SUZ12 protein [28], while Westra et al. found that another SNP, rs3807598, had an effect as trans-eQTL, acting as a putative driver in whole blood with a significant *P*-value ($P=0.00019$) [30]. Here, we found that the risk-associated heterozygous genotypes of these two SNPs showed higher *HOTTIP* expression. Because *HOTTIP* functions as an oncogenic lncRNA, we speculate that the risk-associated rs3807598 and rs2067087 SNPs could participate in gastric carcinogenesis by up-regulating the expression of mature *HOTTIP*. Further molecular experiments should be performed to verify our results.

In view of the impact of *HOTTIP* polymorphisms on the overall survival of patients with gastric cancer, we found that, in this case–control study, none of these four SNPs affected the prognosis of gastric cancer. We previously reported that *HOTTIP* overexpression was associated with poor gastric cancer prognosis [17], and Ye et al. identified *HOTTIP* expression levels as an independent factor for poor prognosis in gastric cancer patients [17]. Although both the heterozygous genotypes of rs3807598 and rs2067087 up-regulated *HOTTIP* expression, either rs3807598 or rs2067087 heterozygous genotype was not significantly correlated with poor survival. Further studies are needed to investigate the association between *HOTTIP* SNPs and the prognosis of gastric cancer.

It should be pointed that the study had some limitations. First, the sample size was limited, causing limited probability of the stratified and interaction analysis for variant genotypes. Second, the expression data of lncRNA-*HOTTIP* gene in the present study was only based on RNA level. Experiments *in vitro* are needed in future research. Third, it is a relatively peripheral association study and lacking functional evidence that links to the studied SNPs and *HOTTIP*. Therefore, further confirmation would be warranted.

Conclusion

The present study first identified two functional SNPs (rs3807598 and rs2067087) in *HOTTIP* with the potential to predict gastric cancer risk. These SNPs variants were associated with corresponding *HOTTIP* expression, providing clues for further studies focused on *HOTTIP* SNPs and gastric cancer pathogenesis. For the future perspective, lncRNA SNPs could have potential to be biomarkers for gastric cancer risk and help to elucidate the etiology of gastric cancer.

Competing Interests

The authors declare that there are no competing interests associated with the manuscript.

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Author Contribution

Yuan Yuan conceived and designed the present study. Ben-gang Wang, Zhi Lv and Han-xi Ding performed the experiment. Ben-gang Wang, Qian Xu and were responsible for the data analysis and performed data interpretation. Ben-gang Wang wrote the paper. Yi-zhi Li and Yuan Yuan revised the manuscript.

Ethics Approval

All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards.

Abbreviations

CI, confidence interval; eQTL, expression quantitative trait Loci; lncRNA, long non-coding RNA; MST, median survival time; OR, odds ratio; SNP, single nucleotide polymorphism; TNM, Tumor Node Metastasis.

References

- 1 Chen, W., Zheng, R., Baade, P.D., Zhang, S., Zeng, H., Bray, F. et al. (2016) Cancer statistics in China, 2015. *CA Cancer J. Clin.* **66**, 115–132, <https://doi.org/10.3322/caac.21338>
- 2 Zhang, X.Y. and Zhang, P.Y. (2017) Gastric cancer: somatic genetics as a guide to therapy. *J. Med. Genet.* **54**, 305–312, <https://doi.org/10.1136/jmedgenet-2016-104171>
- 3 Duraes, C., Almeida, G.M., Seruca, R., Oliveira, C. and Carneiro, F. (2014) Biomarkers for gastric cancer: prognostic, predictive or targets of therapy? *Virchows Arch.* **464**, 367–378, <https://doi.org/10.1007/s00428-013-1533-y>
- 4 Moran, V.A., Perera, R.J. and Khalil, A.M. (2012) Emerging functional and mechanistic paradigms of mammalian long non-coding RNAs. *Nucleic Acids Res.* **40**, 6391–6400, <https://doi.org/10.1093/nar/gks296>
- 5 Gupta, R.A., Shah, N., Wang, K.C., Kim, J., Horlings, H.M., Wong, D.J. et al. (2010) Long non-coding RNA HOTAIR reprograms chromatin state to promote cancer metastasis. *Nature* **464**, 1071–1076, <https://doi.org/10.1038/nature08975>
- 6 Nagano, T. and Fraser, P. (2011) No-nonsense functions for long noncoding RNAs. *Cell* **145**, 178–181, <https://doi.org/10.1016/j.cell.2011.03.014>
- 7 Singer, R.A., Arnes, L. and Sussel, L. (2015) Noncoding RNAs in beta cell biology. *Curr. Opin. Endocrinol. Diabetes Obes.* **22**, 77–85, <https://doi.org/10.1097/MED.000000000000141>
- 8 Mattick, J.S., Amaral, P.P., Dinger, M.E., Mercer, T.R. and Mehler, M.F. (2009) RNA regulation of epigenetic processes. *Bioessays* **31**, 51–59, <https://doi.org/10.1002/bies.080099>
- 9 Taucher, V., Mangge, H. and Haybaeck, J. (2016) Non-coding RNAs in pancreatic cancer: challenges and opportunities for clinical application. *Cell. Oncol. (Dordr.)* **39**, 295–318, <https://doi.org/10.1007/s13402-016-0275-7>
- 10 Cai, H., An, Y., Chen, X., Sun, D., Chen, T., Peng, Y. et al. (2016) Epigenetic inhibition of miR-663b by long non-coding RNA HOTAIR promotes pancreatic cancer cell proliferation via up-regulation of insulin-like growth factor 2. *Oncotarget* **7**, 86857–86870, <https://doi.org/10.18632/oncotarget.13490>
- 11 Lian, Y., Cai, Z., Gong, H., Xue, S., Wu, D. and Wang, K. (2016) HOTTIP: a critical oncogenic long non-coding RNA in human cancers. *Mol. Biosyst.* **12**, 3247–3253, <https://doi.org/10.1039/C6MB00475J>
- 12 Zhang, Z.Z., Shen, Z.Y., Shen, Y.Y., Zhao, E.H., Wang, M., Wang, C.J. et al. (2015) HOTAIR long noncoding RNA promotes gastric cancer metastasis through suppression of Poly (C)-Binding Protein (PCBP) 1. *Mol. Cancer Ther.* **14**, 1162–1170, <https://doi.org/10.1158/1535-7163.MCT-14-0695>
- 13 Liu, X.H., Sun, M., Nie, F.Q., Ge, Y.B., Zhang, E.B., Yin, D.D. et al. (2014) Lnc RNA HOTAIR functions as a competing endogenous RNA to regulate HER2 expression by sponging miR-331-3p in gastric cancer. *Mol. Cancer* **13**, 92, <https://doi.org/10.1186/1476-4598-13-92>
- 14 Yang, C., Tang, R., Ma, X., Wang, Y., Luo, D., Xu, Z. et al. (2015) Tag SNPs in long non-coding RNA H19 contribute to susceptibility to gastric cancer in the Chinese Han population. *Oncotarget* **6**, 15311–15320, <https://doi.org/10.18632/oncotarget.3840>
- 15 Wang, K.C., Yang, Y.W., Liu, B., Sanyal, A., Corces-Zimmerman, R., Chen, Y. et al. (2011) A long noncoding RNA maintains active chromatin to coordinate homeotic gene expression. *Nature* **472**, 120–124, <https://doi.org/10.1038/nature09819>
- 16 Fliss, I., St Laurent, M., Emond, E., Simard, R.E., Lemieux, R., Ettriki, A. et al. (1995) Anti-DNA.RNA antibodies: an efficient tool for non-isotopic detection of *Listeria* species through a liquid-phase hybridization assay. *Appl. Microbiol. Biotechnol.* **43**, 717–724
- 17 Ye, H., Liu, K. and Qian, K. (2016) Overexpression of long noncoding RNA HOTTIP promotes tumor invasion and predicts poor prognosis in gastric cancer. *Onco Targets Ther.* **9**, 2081–2088
- 18 Zhao, R., Zhang, Y., Zhang, X., Yang, Y., Zheng, X., Li, X. et al. (2018) Exosomal long noncoding RNA HOTTIP as potential novel diagnostic and prognostic biomarker test for gastric cancer. *Mol. Cancer* **17**, 68, <https://doi.org/10.1186/s12943-018-0817-x>
- 19 Long, Z.W., Yu, H.M., Wang, Y.N., Liu, D., Chen, Y.Z., Zhao, Y.X. et al. (2015) Association of IL-17 polymorphisms with gastric cancer risk in Asian populations. *World J. Gastroenterol.* **21**, 5707–5718, <https://doi.org/10.3748/wjg.v21.i18.5707>
- 20 Han, L., Lee, S.W., Yoon, J.H., Park, Y.G., Choi, Y.J., Nam, S.W. et al. (2013) Association of SOD1 and SOD2 single nucleotide polymorphisms with susceptibility to gastric cancer in a Korean population. *APMIS* **121**, 246–256, <https://doi.org/10.1111/j.1600-0463.2012.02963.x>
- 21 Sultana, Z., Bankura, B., Pattanayak, A.K., Sengupta, D., Sengupta, M., Saha, M.L. et al. (2018) Association of interleukin-1 beta and tumor necrosis factor-alpha genetic polymorphisms with gastric cancer in India. *Environ. Mol. Mutagen.* **59**, 653–667, <https://doi.org/10.1002/em.22208>
- 22 Hu, P., Qiao, O., Wang, J., Li, J., Jin, H., Li, Z. et al. (2017) rs1859168 A > C polymorphism regulates HOTTIP expression and reduces risk of pancreatic cancer in a Chinese population. *World J. Surg. Oncol.* **15**, 155

- 23 Tu, H., Sun, L., Dong, X., Gong, Y., Xu, Q., Jing, J. et al. (2015) Temporal changes in serum biomarkers and risk for progression of gastric precancerous lesions: a longitudinal study. *Int. J. Cancer* **136**, 425–434, <https://doi.org/10.1002/ijc.29005>
- 24 Wang, B.G., Xu, Q., Lv, Z., Fang, X.X., Ding, H.X., Wen, J. et al. (2018) Association of twelve polymorphisms in three onco-lncRNA genes with hepatocellular cancer risk and prognosis: a case-control study. *World J. Gastroenterol.* **24**, 2482–2490, <https://doi.org/10.3748/wjg.v24.i23.2482>
- 25 Xu, Q., Yuan, Y., Sun, L.P., Gong, Y.H., Xu, Y., Yu, X.W. et al. (2009) Risk of gastric cancer is associated with the MUC1 568 A/G polymorphism. *Int. J. Oncol.* **35**, 1313–1320
- 26 Xu, Q., Chen, M.Y., He, C.Y., Sun, L.P. and Yuan, Y. (2013) Promoter polymorphisms in trefoil factor 2 and trefoil factor 3 genes and susceptibility to gastric cancer and atrophic gastritis among Chinese population. *Gene* **529**, 104–112, <https://doi.org/10.1016/j.gene.2013.07.070>
- 27 Li, Z., Zhang, Z., He, Z., Tang, W., Li, T., Zeng, Z. et al. (2009) A partition-ligation-combination-subdivision EM algorithm for haplotype inference with multiallelic markers: update of the SHEsis (<http://analysis.bio-x.cn>). *Cell Res.* **19**, 519–523, <https://doi.org/10.1038/cr.2009.33>
- 28 Harrow, J., Frankish, A., Gonzalez, J.M., Tapanari, E., Diekhans, M., Kokocinski, F. et al. (2012) GENCODE: the reference human genome annotation for The ENCODE Project. *Genome Res.* **22**, 1760–1774, <https://doi.org/10.1101/gr.135350.111>
- 29 Xu, X.T., Tao, Z.Z., Song, Q.B., Yao, Y. and Ruan, P. (2014) SUZ12 RNA interference inhibits the invasion of gastric carcinoma cells. *Hepatogastroenterology* **61**, 697–702
- 30 Westra, H.J., Peters, M.J., Esko, T., Yaghootkar, H., Schurmann, C., Kettunen, J. et al. (2013) Systematic identification of trans eQTLs as putative drivers of known disease associations. *Nat. Genet.* **45**, 1238–1243, <https://doi.org/10.1038/ng.2756>