



OPEN

Evaluation of biochemical and hematological parameters in adults with Down syndrome

David de Gonzalo-Calvo^{1,2,3}, Isabel Barroeta^{4,5}, Madalina Nicoleta Nan⁶, José Rives⁶, Diana Garzón^{4,5}, María Carmona-Iragui^{4,5,7}, Bessy Benejam^{4,5,7}, Laura Videla^{4,5,7}, Susana Fernández⁷, Miren Altuna^{4,5}, Sílvia Valldeneu^{4,5}, Rafael Blesa^{4,5}, Alberto Lleó^{4,5}, Francisco Blanco-Vaca^{6,8,9}, Juan Fortea^{4,5,7} & Mireia Tondo⁶✉

Down syndrome (DS) is the most common worldwide cause of intellectual disability of genetic origin and the most common chromosomal disorder affecting live-born infants. In addition to intellectual disability, individuals with DS have other comorbidities and complex medical conditions. The increase in the life expectancy of patients with DS requires expanding the knowledge about their clinical characteristics and related laboratory parameters. Several studies exploring laboratory tests in DS patients exist, but their focus is limited to specific areas of metabolism. Therefore, our main goal was to describe the biochemical and hematological findings in a DS cohort and to compare the values to those of a control population. A total of 248 DS individuals and 84 control subjects were enrolled. DS individuals had a higher frequency of several clinical conditions compared to control individuals and presented with significant differences with respect to the controls in both biochemical and hematological parameters. We found age- and sex-related differences in several of the parameters. A good understanding of the differences in our cohort might be of aid in the clinical follow-up of adults with DS, especially considering that the lifespan of DS individuals may reach 60 years of age in developed countries.

Abbreviations

AD	Alzheimer's disease
AF	Alkaline phosphatase
ALT	Alanine aminotransferase
AST	Aspartate aminotransferase
B12	Vitamin B12
CKD-EPI	Chronic kidney disease epidemiology collaboration
DS	Down syndrome
ESR	Erythrocyte sedimentation rate
FT4	Free thyroxine
eGFR	Estimated glomerular filtration rate
GGT	Gamma-glutamyl transferase
HbA1c	Glycated hemoglobin

¹Biomedical Research Institute Sant Pau (IIB Sant Pau), Barcelona, Spain. ²Institute of Biomedical Research of Barcelona (IIBB), Spanish National Research Council (CSIC), Barcelona, Spain. ³Translational Research in Respiratory Medicine, University Hospital Arnau de Vilanova and Santa Maria, IRBLleida, Lleida, Spain. ⁴Sant Pau Memory Unit, Department of Neurology, Hospital de La Santa Creu i Sant Pau, Biomedical Research Institute (IIB) Sant Pau, Universitat Autònoma de Barcelona, Barcelona, Spain. ⁵Center of Biomedical Investigation Network for Neurodegenerative Diseases (CIBERNED), Madrid, Spain. ⁶Department of Biochemistry, Hospital de La Santa Creu i Sant Pau, Biomedical Research Institute (IIB) Sant Pau, C/Sant Quintí 89, 08041 Barcelona, Spain. ⁷Barcelona Down Medical Center, Fundació Catalana de Síndrome de Down, Barcelona, Spain. ⁸Center of Biomedical Investigation Network for Diabetes and Metabolic Diseases (CIBERDEM), Madrid, Spain. ⁹Department of Biochemistry and Molecular Biology, Universitat Autònoma de Barcelona, Barcelona, Spain. ✉email: mtondo@santpau.cat

HDLc	High-density lipoprotein cholesterol
LDLc	Low-density lipoprotein cholesterol
MCH	Mean corpuscular hemoglobin
MCHC	Mean corpuscular hemoglobin concentration
MCV	Mean corpuscular volume
MDRD-4	Modification of diet in renal disease
MPV	Mean platelet volume
K+	Potassium
RDW	Red blood cell distribution width
Na+	Sodium
TG	Triglycerides
TSH	Thyroid stimulating hormone

Down syndrome (DS) is the most common worldwide cause of intellectual disability of genetic origin and the most common chromosomal disorder affecting live-born infants, with an estimated birth prevalence of 14 per 10,000 live births^{1–3}. Despite the shorter life expectancy when compared to healthy subjects and adults with other causes of intellectual disability⁴, there has been a progressive increase in the life expectancy of patients with DS in recent decades, currently reaching nearly 60 years⁵. This fact has increased the need to expand the knowledge about the clinical characteristics of DS individuals and the health problems differentiating them from both pediatric and adult populations⁶. DS is associated with a distinct phenotype involving many body systems. In addition to intellectual disability, individuals with DS present with a high number of comorbidities and complex medical conditions whose frequencies are modified throughout the lifespan of the individuals⁷. The increase in life expectancy has led to a higher prevalence of age-related pathologies, including premature Alzheimer's disease (AD)⁸.

Since optimal medical management is associated with improved quality of life and functioning among persons with DS^{9,10}, medical professionals, including pediatricians and other physicians, should closely supervise this population throughout their lifespan and evaluate their laboratory results. Previous investigations in DS cohorts have focused on select biochemical parameters, such as uric acid and thyroid function biomarkers, bone mineral density, nutritional zinc status, gonadal and endocrine function and glucose and lipid metabolism parameters^{11–16}. However, no previous work has described a comprehensive panel of biochemical and hematological parameters in a large cohort of DS patients.

Our hypothesis is that a thorough analysis of the biochemical and hematological parameters will provide a basis to establish whether commonly observed alterations in DS individuals are intrinsic of the disease or have clinical implications similarly as for the general population. Therefore, our goals were to describe the biochemical and hematological findings in our DS cohort and to compare the values to those of a control population.

Material and methods

Study participants. This was a single-center descriptive study of adults with DS recruited at Barcelona Down Medical Center (Fundació Catalana Síndrome de Down and Hospital de la Santa Creu i Sant Pau, Barcelona) in Catalonia, Spain, according to a population-based health plan to screen for neurological comorbidities^{17,18}. The Down Medical Center provides medical care specifically for individuals with DS and possesses over 2,500 medical records (more than 50% of the estimated Down syndrome population in Catalonia); therefore, it reflects the population with DS in our geographic area. The period of patient recruitment for this study was February 1, 2013, to June 30, 2018. In adults with DS (≥ 18 years), a biochemical and hematological analysis was performed as part of their annual health plan visit. A total of 254 patients were enrolled in the study. Six further patients were ultimately excluded for presenting with conditions unrelated to DS according to their medical records: 4 patients with hepatitis C, 1 patient with hepatitis B and 1 patient with breast cancer, resulting in a final total number of 248 DS individuals included (age range 18–63 years). A total of 84 healthy control participants in the same age range (23–65 years) were enrolled in the study. Volunteers were recruited from the SPIN (Sant Pau Initiative on Neurodegeneration) cohort (<https://santpauemoryunit.com/our-research/spin-cohort/>) or social media (@SantPauMemory). Further details on the clinical protocol of the SPIN cohort can be found elsewhere¹⁹.

Based on current guidelines^{17,20}, associated clinical conditions were obtained through a systematic review of the medical records, including the following: history of arterial hypertension, dyslipidemia, diabetes mellitus, congenital heart disease, gastrointestinal pathology, dermatological pathology, bone pathology, hypothyroidism, hearing problems, otolaryngology pathology, ophthalmological pathology, psychiatric pathology, epilepsy, and Alzheimer's disease. Treatment data, with a special focus on the treatment of hypothyroidism, were also collected.

Biochemical and hematological data. Analyzed biochemical and hematological parameters were selected according to a defined laboratory blood profile as recommended in the guidelines for management of patients with DS^{17,20}.

Blood collection and processing were performed in accordance with the Standard Operating Procedures for Serum and Plasma Collection from the Early Detection Research Network (EDRN) Consensus Statement and Standard Operating Procedure Integration Working Group²¹. Blood samples were collected by venipuncture after an overnight fast.

Whole blood samples were collected in VACUTAINER tubes and fractionated by centrifugation at 1,300 g for 15 min at room temperature to obtain serum. Serum was aliquoted into 1.5 mL tubes, and the following parameters were measured according to standard commercially available assays adapted to an Architect C4000 (Abbott Diagnostics, USA) using automated procedures: thyroid stimulating hormone (TSH), free thyroxine (FT4),

	Control		Down syndrome		p-value
	n	Median (P25–P75)/n (%)	n	Median (P25–P75)/n (%)	
Age (years)	84	55.0 (47.3–59.8)	248	43.0 (33.0–50.8)	<0.001
Male/female	84	21 (25.0)/63 (75.0)	248	132 (53.2)/116 (46.8)	<0.001
Arterial hypertension	84	16 (19.0)	248	3 (1.2)	<0.001
Dyslipidemia	84	24 (28.6)	248	37 (14.9)	0.009
Diabetes mellitus	84	6 (7.1)	248	6 (2.4)	0.082
Congenital heart disease	84	0 (0.0)	247	47 (19.0)	<0.001
Gastrointestinal pathology	84	1 (1.2)	247	40 (16.2)	<0.001
Dermatological pathology	84	2 (2.4)	247	83 (33.6)	<0.001
Bone pathology	84	3 (3.6)	247	36 (14.6)	0.006
Hypothyroidism	84	1 (1.2)	247	119 (48.2)	<0.001
Treatment for hypothyroidism	–		246	103 (41.9)	
Hearing problems	84	0 (0.0)	245	40 (16.3)	<0.001
Otolaryngology pathology	84	1 (1.2)	247	37 (15.0)	<0.001
Ophthalmological pathology	84	1 (1.2)	246	136 (55.3)	<0.001
Psychiatric pathology	84	18 (21.4)	246	45 (18.3)	0.524
Epilepsy	84	1 (1.2)	247	38 (15.4)	<0.001
Alzheimer's disease	84	0 (0.0)	247	50 (20.2)	<0.001

Table 1. Characteristics of the Study Population. Data are presented as frequencies (percentages) for categorical variables. Continuous variables are presented as median (interquartile range). Differences between groups were analyzed using Wilcoxon rank-sum test or Fisher's exact test.

sodium (Na⁺), potassium (K⁺), glucose, urea, creatinine, total bilirubin, triglycerides (TG), total cholesterol, aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (AF), gamma-glutamyl transferase (GGT), total proteins, vitamin B12, and folate. The estimated glomerular filtration rate (eGFR) was calculated according to the MDRD-4 (Modification of Diet in Renal Disease) and CKD-EPI (Chronic Kidney Disease Epidemiology Collaboration) formulas.

Whole blood samples in EDTA-K₃ were also obtained for determining blood cell count and indices. The tubes were immediately inverted 10 times to mix the anticoagulant additive with blood. The blood was processed within 2 h of extraction. Using the impedance channel of the automated hematology analyzer Sysmex XE-2100 (Roche Diagnostics, Kobe, Japan), the following parameters were determined: red blood cell count (RBC), white blood cell count (WBC), platelet count, hemoglobin, hematocrit, mean volume (MCV), mean corpuscular hemoglobin concentration (MCHC), mean corpuscular hemoglobin (MCH), red blood cell distribution width (RDW) and mean platelet volume (MPV). The erythrocyte sedimentation rate (ESR) was calculated with a VES cube 200 Sysmex Analyzer (Roche Diagnostics, Kobe, Japan).

Values were compared to normal reference ranges used in our laboratory established in a healthy population from our geographical area according to standardized guides²².

Statistical analysis. Descriptive statistics were used to summarize the characteristics of the study population. Data are presented as medians [25th percentile (P25)–75th percentile (P75)] for continuous variables and as frequencies (percentages) for categorical variables. Data normality was analyzed using the Kolmogorov–Smirnov test. Continuous variables were compared between groups using the Wilcoxon rank-sum test. ANCOVA models, adjusted for age and sex, were used to compare continuous variables across the study groups. Variables were log-transformed to achieve a normal distribution. For clarity, the original values are shown. Categorical variables were compared between groups using Fisher's exact test. Spearman's rho coefficient was used to assess the correlation between continuous variables. The statistical software package R (<https://www.r-project.org>) was used for statistical analyses. A P-value < 0.05 was considered statistically significant.

Ethical aspects. The study was approved by the Sant Pau Ethics Committee following the standards for medical research in humans recommended by the Declaration of Helsinki and in accordance with Spanish legislation for research in people with intellectual disabilities. All participants or their legally authorized representatives gave written informed consent before enrolment in accordance with the guidelines of the local ethics committee.

Results

Study cohort characteristics. We enrolled a total of 248 individuals with DS, 132 males (53.2%) and 116 females (46.8%), with a median age of 43.0 (33.0–50.8) years, and 84 control subjects, 21 males (25.0%) and 63 females (75.0%), with a median age of 55.0 (47.3–59.8) years. The clinical features of the DS and control populations are listed in Table 1. The frequency of the following clinical conditions was significantly higher in the DS group than in the control group: history of congenital heart disease, gastrointestinal pathology, dermatologi-

cal pathology, bone pathology, hypothyroidism, hearing problems, otolaryngology pathology, ophthalmological pathology, epilepsy, and AD. No differences were observed in the frequency of diabetes mellitus or psychiatric pathology for either group. DS individuals presented with a lower frequency of arterial hypertension and dyslipidemia compared to the control group. See Table 1 for further details on the cohort characteristics.

Biochemical and hematological parameters in patients with Down syndrome. We performed a detailed biochemical and hematological analysis of the DS cohort and compared the profiles obtained with our control population. The reference values of the studied parameters, the number and percentage of patients out of range, and the median (P25–P75) of the whole study population are shown in Table 2. Seventy-three percent of the studied hematological parameters and 53% of the studied biochemical parameters were significantly different between the DS individuals and the control population. The DS individuals presented with higher TSH, urea, creatinine, AST, hemoglobin, hematocrit, MCV, ESR, MCH and RDW values and lower TG, total cholesterol, folate, eGFR, MPV and WBC values. These differences remained significant, or close to significance, after adjusting for confounding factors such as age and sex. Statistical differences for RBC and MCHC were observed after adjustment. An additional analysis to evaluate the impact of hypothyroidism treatment on TSH was performed.

No differences were observed for TSH between both studied groups (treated DS individuals = 3.02 (1.25–4.27) vs. untreated DS individuals = 3.20 (1.84–3.98), P -value = 0.194). For categorical variables, the percentage of DS individuals out of range for some parameters was also statistically significant compared to the control population. Parameters with a higher percentage of values out of range in the DS group were TSH, urea, creatinine, total proteins, RBC, MCV, ESR, MCH, and WBC, whereas those with a lower percentage of values out of range were K^+ , TG, total cholesterol, and AST.

The differences in the biochemical and hematological parameters and the number and percentage of patients out of range between DS individuals and the control population according to sex are displayed in Supplemental Tables 1 and 2. For the female DS cohort, parameters with significantly higher values were TSH, urea, creatinine, AST, hemoglobin, hematocrit, MCV, ESR, MCH, and RDW, whereas those with significantly lower values were TG, total cholesterol, GGT, eGFR, RBC, MPV, and WBC. For categorical variables, parameters with significantly higher percentages of values out of range were TSH, creatinine, total proteins, MCV, ESR, MCH and WBC, whereas those with a significantly lower percentage of values out of range were total cholesterol and B12 (Supplemental Table 1). For the male DS cohort, parameters with significantly higher values were TSH, hemoglobin, hematocrit, MCV, ESR, MCH, and RDW, whereas those with significantly lower values were TG, total cholesterol, eGFR, MPV, and WBC. Regarding categorical variables, parameters with significantly higher percentages of values out of range were TSH, ESR, and MCH, whereas those with significantly lower percentages of values out of range were K^+ , TG, total cholesterol, and GGT (Supplemental Table 2).

The differences in the biochemical and hematological parameters between males and females as well as the frequency and percentage of patients out of range in the control and DS groups are displayed in Supplemental Table 3 and Table 3, respectively. For the control group, parameters with significantly higher values in the male subgroup were K^+ , creatinine, TG, ALT, hemoglobin, hematocrit, RBC, and MCHC, whereas those with significantly lower values were Al^{3+} , eGFR, and ESR. Among the categorical variables, K^+ had a significantly higher percentage of values out of range in the male subgroup, and ESR had a significantly lower percentage of values out of range (Supplemental Table 3). For the DS cohort, parameters with significantly higher values in the male subgroup were creatinine, total bilirubin, TG, ALT, GGT, hemoglobin, hematocrit, RBC, MCHC and WBC, whereas those with significantly lower values were folate, MCV, ESR, RDW, platelet count, and MPV. Regarding categorical variables, parameters with significantly higher percentages of values out of range in the male subgroup were total bilirubin, B12, RBC and MPV, whereas those with significantly lower percentages of values out of range were MCV, ESR and MCHC (Table 3).

The correlation between the biochemical and hematological data with age was also explored in both study groups. As shown in Table 4, for the control population, urea, creatinine, total cholesterol and AST showed a significant positive correlation with age, while eGFR showed a significant negative correlation. For the DS population, Na^+ , urea, creatinine, TG, total cholesterol, AST, Al^{3+} , MCV, ESR, MCH, and RDW showed a significant positive correlation with age, while eGFR, ALT, B12, hemoglobin, hematocrit, RBC, MCHC, and platelet count showed a significant negative correlation.

Discussion

The present study evaluated several biochemical and hematological parameters in a large sample of adults with DS. Several studies exploring laboratory tests in DS patients exist, but their focus is limited to specific areas of metabolism^{11–16}. DS is among the most complex genetic conditions compatible with life, characterized by accelerated aging and affecting gene expression beyond chromosome 21²³. The sheer number of affected genes and epigenetic changes suggests that numerous pathways of human metabolism are altered and subsequently might be reflected in laboratory test parameters. Here, we performed a comprehensive approach by analyzing parameters related to different physiological mechanisms. We found significant differences with respect to non-trisomic controls in both biochemical and hematological parameters, even after adjusting for potential confounding factors. Furthermore, we found age- and sex-related differences in several of the parameters. The fact that women with DS experience menopause earlier than healthy women²⁴ may explain some of these sex-related differences.

Clinically and as previously described^{4,8,9,25}, our DS cohort presented with a higher incidence of congenital heart disease, gastrointestinal pathology, dermatological pathology, bone pathology, hypothyroidism, otolaryngology pathology, ophthalmological pathology, epilepsy and AD than the control population. Arterial

Variable	Reference values	Control			Down syndrome			p-value (categorical)	p-value (continuous)	p-value (continuous, adjusted)
		n	n OOR (%)	Median (P25–P75)	n	n OOR (%)	Median P25–P75			
Biochemical parameters										
TSH (mUI/L)	(0.3–5.0)	84	1 (1.2)	1.2 (0.97–1.73)	247	46 (18.6)	2.8 (1.67–4.15)	<0.001	<0.001	<0.001
Na ⁺ (mmol/L)	(136–145)	84	1 (1.2)	140.0 (139.0–141.0)	248	4 (1.6)	140.0 (139.0–141.0)	1.000	0.696	0.889
K ⁺ (mmol/L)	(3.5–5.1)	83	4 (4.8)	4.3 (4.0–4.5)	245	1 (0.4)	4.3 (4.1–4.5)	0.016	0.236	0.472
Glucose (mmol/L)	(3.0–6.1)	84	3 (3.6)	5.0 (4.7–5.4)	248	17 (6.9)	5.0 (4.7–5.3)	0.426	0.559	0.649
Urea (mmol/L)	≤ 60 years (2.1–7.1) > 60 years (2.9–8.2)	84	11 (13.1)	5.5 (4.6–6.3)	248	62 (25.0)	6.1 (5.4–7.2)	0.023	<0.001	<0.001
Creatinine (μmol/L)	Females (<80) Males (<106)	84	2 (2.4)	64.0 (59.0–72.0)	248	24 (9.7)	74.0 (66.0–84.0)	0.033	<0.001	<0.001
eGFR (ml/min/1.73)	(<60)	84	0 (0.0)	97.4 (91.6–102.2)	248	9 (3.6)	90.0 (84.2–90.0)	0.119	<0.001	<0.001
Total bilirubin (μmol/L)	(<17)	84	8 (9.5)	9.0 (7.0–11.2)	234	30 (12.8)	10.0 (7.0–13.0)	0.557	0.141	0.778
TG (mmol/L)	(<1.65)	84	14 (16.7)	0.99 (0.71–1.52)	248	19 (7.7)	0.87 (0.71–1.05)	0.033	0.020	0.004
Total cholesterol (mmol/L)	(<6.2)	84	20 (23.8)	5.4 (4.8–6.2)	248	15 (6.0)	4.9 (4.3–5.4)	<0.001	<0.001	0.027
AST (U/L)	Females (<31) Males (<37)	84	10 (11.9)	19.0 (17.0–23.8)	245	12 (4.9)	21.0 (18.0–25.0)	0.040	0.008	0.089
ALT (U/L)	Females (<31) Males (<41)	84	10 (11.9)	19.0 (15.0–25.5)	248	25 (10.1)	20.0 (15.0–27.0)	0.682	0.648	0.783
AF (U/L)	Females (35–110) Males (40–130)	78	6 (7.7)	79.0 (63.5–95.5)	247	14 (5.7)	76.0 (64.0–87.0)	0.589	0.609	0.281
GGT (U/L)	Females (<43) Males (<54)	84	12 (14.3)	19.0 (14.0–31.5)	248	20 (8.1)	18.0 (13.0–26.0)	0.132	0.092	0.149
Total proteins (g/L)	(64–83)	84	2 (2.4)	69.6 (67.8–71.7)	233	24 (10.3)	68.5 (65.7–71.2)	0.021	0.016	0.021
B12 (pmol/L)	(150–650)	84	7 (8.3)	297.0 (235.0–401.3)	242	10 (4.1)	287.5 (221.8–350.3)	0.156	0.091	0.007
Folate (nmol/L)	(7–45)	84	6 (7.1)	14.7 (11.7–21.6)	243	16 (6.6)	12.3 (9.4–18.3)	0.805	0.009	0.488
Hematological parameters										
Hemoglobin (g/L)	Females (120–150) Males (130–170)	82	7 (8.5)	136.0 (128.0–142.0)	248	32 (12.9)	144.0 (136.0–154.0)	0.330	<0.001	0.005
Hematocrit (L/L)	Females (0.35–0.45) Males (0.4–0.5)	82	6 (7.3)	0.40 (0.38–0.42)	248	23 (9.3)	0.43 (0.41–0.46)	0.660	<0.001	<0.001
RBC (× 10 ¹² /L)	Females (3.9–5) Males (4.5–5.7)	82	9 (11.0)	4.5 (4.3–4.7)	248	58 (23.4)	4.6 (4.2–4.9)	0.017	0.915	0.002
MCV (fL)	(80–98)	82	1 (1.2)	88.4 (86.3–90.5)	248	60 (24.2)	95.4 (92.2–97.8)	<0.001	<0.001	<0.001
ESR (mm/h)	(1–10)	59	32 (54.2)	14.0 (5.0–25.0)	179	137 (76.5)	24.0 (11.0–39.0)	0.002	<0.001	<0.001
MCHC (g/L)	(320–360)	82	2 (2.4)	337.0 (330.0–345.0)	248	17 (6.9)	337.0 (330.3–343.0)	0.176	0.940	0.049
MCH (pg)	(27–32)	82	8 (9.8)	29.9 (28.8–30.8)	248	137 (55.2)	32.2 (31.0–33.0)	<0.001	<0.001	<0.001
RDW (%)	(12–15)	82	10 (12.2)	13.0 (12.3–13.7)	248	30 (12.1)	13.7 (13.2–14.4)	1.000	<0.001	<0.001
Platelet count (× 10 ⁹ /L)	(140–350)	81	4 (4.9)	251.0 (210.0–275.0)	248	9 (3.6)	235.0 (202.0–274.0)	0.531	0.109	0.054
MPV (fL)	(7.0–10.5)	82	7 (8.5)	8.4 (7.7–9.2)	248	18 (7.3)	7.8 (7.4–8.3)	0.810	<0.001	0.002
WBC (× 10 ⁹ /L)	(3.8–11.0)	82	3 (3.7)	6.2 (5.3–7.7)	248	30 (12.1)	5.2 (4.4–6.3)	0.032	<0.001	<0.001

Table 2. Biochemical and hematological parameters in the control group and the cohort of patients with Down Syndrome. Differences between groups were analyzed using Wilcoxon Rank-sum test, ANCOVA models adjusted for age and sex, or the Fisher's exact test. OOR out of range, NA not applicable.

hypertension and dyslipidemia were less prevalent, whereas no difference was observed regarding the diabetes mellitus incidence, as discussed below.

With respect to laboratory studies, the hematological profile was largely altered in DS individuals when compared to the control population. Of note, significant differences were found for almost all the hematological parameters when comparing males and females, suggesting the need to consider sex when evaluating the hematological profile in a DS individual. It is well known that trisomy 21 impacts hematopoietic cell biology through multiple and complex pathways. In adults, the metabolic and redox derangements observed in the RBCs from individuals with DS have been previously linked to alterations in cell survival and size, in particular macrocytosis²⁶. Different studies have also proposed that the additional copy of chromosome 21 has a profound impact on fetal hematopoiesis, which ultimately impacts the function and number of hematopoietic lineages^{27–31}. Additionally, between 4 and 10% of newborn infants with DS develop transient myeloproliferative disorder^{32–34}. Although the disease usually resolves without treatment in the first few months of life, it is estimated that 20–30% of individuals with transient myeloproliferative disorder will go on to develop subsequent leukemia^{35,36}. Finally, the fact that folate concentrations are significantly lower in DS individuals matches the observed hematological alterations. Taken together, these impaired hematological parameters suggest the existence of abnormalities

Variable	Female			Male			p-value (categorical)	p-value (continuous)
	n	n OOR (%)	Median (P25–P75)	n	n OOR (%)	Median (P25–P75)		
Biochemical parameters								
TSH (mUI/L)	115	24 (20.9)	2.79 (1.67–4.15)	132	22 (16.7)	2.85 (1.68–3.81)	0.417	0.877
Na ⁺ (mmol/L)	116	1 (0.9)	140.0 (139.0–141.0)	132	3 (2.3)	140.0 (139.0–141.0)	0.625	0.277
K ⁺ (mmol/L)	113	1 (0.9)	4.3 (4.2–4.5)	132	0 (0.0)	4.3 (4.1–4.6)	0.461	0.871
Glucose (mmol/L)	116	4 (3.4)	4.9 (4.6–5.2)	132	13 (9.8)	5.1 (4.8–5.4)	0.075	0.023
Urea (mmol/L)	116	29 (25.0)	6.2 (5.2–7.2)	132	33 (25.0)	6.0 (5.4–7.2)	1.000	0.861
Creatinine (μmol/L)	116	13 (11.2)	68.0 (62.0–75.0)	132	11 (8.3)	82.0 (73.0–94.0)	0.521	<0.001
eGFR (ml/min/1.73)	116	4 (3.4)	90.0 (83.1–90.0)	132	5 (3.8)	90.0 (87.0–90.0)	1.000	0.179
Total bilirubin (μmol/L)	111	9 (8.1)	9.0 (6.0–11.0)	123	21 (17.1)	10.0 (8.0–14.0)	0.050	<0.001
TG (mmol/L)	116	6 (5.2)	0.82 (0.70–0.95)	132	13 (9.8)	0.92 (0.72–1.21)	0.232	0.004
Total cholesterol (mmol/L)	116	9 (7.8)	4.9 (4.5–5.4)	132	6 (4.5)	4.9 (4.2–5.3)	0.302	0.315
AST (U/L)	113	8 (7.1)	22.0 (18.0–26.0)	132	4 (3.0)	21.0 (19.0–25.0)	0.234	0.754
ALT (U/L)	116	13 (11.2)	18.0 (14.0–25.0)	132	12 (9.1)	23.0 (16.0–29.0)	0.674	0.002
AF (U/L)	115	10 (8.7)	76.0 (65.0–88.0)	132	4 (3.0)	76.5 (64.0–87.0)	0.095	0.620
GGT (U/L)	116	9 (7.8)	16.0 (12.0–22.0)	132	11 (8.3)	19.0 (15.0–27.8)	1.000	0.001
Total proteins (g/L)	110	16 (14.5)	68.5 (65.3–71.4)	123	8 (6.5)	68.5 (66.0–70.9)	0.053	0.856
B12 (pmol/L)	112	1 (0.9)	292.0 (227.3–366.5)	130	9 (6.9)	286.5 (208.0–339.5)	0.022	0.278
Folate (nmol/L)	113	7 (6.2)	13.9 (9.9–20.6)	130	9 (6.9)	11.6 (8.8–16.8)	1.000	0.003
Hematological parameters								
Hemoglobin (g/L)	116	18 (15.5)	139.0 (132.0–145.0)	132	14 (10.6)	152.0 (143.0–160.0)	0.262	<0.001
Hematocrit (L/L)	116	10 (8.6)	0.42 (0.39–0.43)	132	13 (9.8)	0.45 (0.42–0.47)	0.828	<0.001
RBC (× 10 ¹² /L)	116	17 (14.7)	4.3 (4.1–4.6)	132	41 (31.1)	4.7 (4.4–5.0)	0.003	<0.001
MCV (fL)	116	37 (31.9)	96.2 (93.0–98.6)	132	23 (17.4)	94.6 (91.8–97.0)	0.011	0.004
ESR (mm/h)	81	75 (92.6)	32.0 (21.5–47.5)	98	62 (63.3)	14.5 (7.0–30.5)	<0.001	<0.001
MCHC (g/L)	116	12 (10.3)	334.0 (329.3–341.0)	132	5 (3.8)	338.5 (332.0–346.0)	0.047	0.003
MCH (pg)	116	71 (61.2)	32.4 (30.9–33.3)	132	66 (50.0)	32.1 (31.0–32.7)	0.096	0.189
RDW (%)	116	18 (15.5)	13.9 (13.4–14.6)	132	12 (9.1)	13.6 (13.1–14.3)	0.171	0.016
Platelet count (× 10 ⁹ /L)	116	3 (2.6)	240.0 (216.0–289.0)	132	6 (4.5)	232.0 (196.0–260.8)	0.508	0.024
MPV (fL)	116	3 (2.6)	7.9 (7.5–8.4)	132	15 (11.4)	7.8 (7.3–8.3)	0.012	0.040
WBC (× 10 ⁹ /L)	116	19 (16.4)	5.1 (4.2–6.0)	132	11 (8.3)	5.5 (4.5–6.7)	0.078	0.028

Table 3. Differences between sex in the Down syndrome group. Differences between groups were analyzed using Wilcoxon Rank-sum test or the Fisher's exact test. OOR out of range, NA not applicable.

in hematopoiesis and provide information on how an extra copy of chromosome 21 may lead to phenotypic consequences.

Concerning the biochemical profile, our results support the findings from previous independent studies. We showed that 18.6% of our DS individuals presented with values out of range for TSH level. Of those, 103 out of 119 were treated for hypothyroidism. Impaired TSH and FT4 levels have been largely described in DS populations³⁷. Moreover, subclinical hypothyroidism in children with DS is an abundantly common occurrence, with a prevalence of approximately 30%³⁸, and has been attributed to the dysregulation of the hypothalamic-pituitary-thyroid axis³⁷. Regarding urea metabolism, 25% of our DS individuals presented with a high urea concentration, which may be due to impaired renal function, among other causes. Indeed, and as previously reported³⁹, almost 10% of our DS individuals also presented with impaired creatinine values. Serum creatinine is the most reliable parameter for detecting kidney damage due to its high diagnostic specificity. From its concentration and based on formulas in which age, sex and weight are taken into account, it is possible to estimate the glomerular filtration rate (eGFR). Our DS cohort also presented with a lower eGFR, which is in agreement with a previous study exploring renal disease in DS individuals⁴⁰. Despite the significantly altered parameters related to renal function, our DS individuals presented with a very low frequency of arterial hypertension.

Concerning the lipid profile, we found significantly lower total cholesterol and TG concentrations in DS individuals compared to the control population. It would have been interesting to study the fractionated forms of cholesterol together with their apolipoprotein concentrations; however, because the current study was not designed to answer questions regarding lipid metabolism, low-density lipoprotein cholesterol (LDLc) and high-density lipoprotein cholesterol (HDLc) were not measured. Several works measuring circulating total cholesterol,

	Control			Down syndrome		
	n	Spearman's rho	p-value	n	Spearman's rho	p-value
Biochemical parameters						
TSH (mUI/L)	84	0.186	0.090	247	-0.018	0.777
Na ⁺ (mmol/L)	84	0.142	0.197	248	0.198	0.002
K ⁺ (mmol/L)	83	0.150	0.175	245	0.120	0.060
Glucose (mmol/L)	84	0.163	0.139	248	0.087	0.170
Urea (mmol/L)	84	0.283	0.009	248	0.253	<0.001
Creatinine (μmol/L)	84	0.319	0.003	248	0.176	0.005
eGFR (ml/min/1.73)	84	-0.727	<0.001	248	-0.498	<0.001
Total bilirubin (μmol/L)	84	-0.006	0.953	234	-0.017	0.801
TG (mmol/L)	84	0.023	0.838	248	0.150	0.018
Total cholesterol (mmol/L)	84	0.265	0.015	248	0.269	<0.001
AST (U/L)	84	0.267	0.014	245	0.142	0.026
ALT (U/L)	84	0.158	0.152	248	-0.153	0.016
AF (U/L)	78	-0.070	0.541	247	0.147	0.021
GGT (U/L)	84	0.206	0.060	248	0.027	0.671
Total Proteins (g/L)	84	-0.026	0.811	233	-0.066	0.315
B12 (pmol/L)	84	0.150	0.173	242	-0.202	0.002
Folate (nmol/L)	84	0.187	0.088	243	-0.102	0.113
Hematological parameters						
Hemoglobin (g/L)	82	0.195	0.079	248	-0.162	0.011
Hematocrit (L/L)	82	0.161	0.148	248	-0.129	0.042
RBC (× 10 ¹² /L)	82	0.148	0.185	248	-0.218	0.001
MCV (fL)	82	-0.022	0.843	248	0.291	<0.001
ESR (mm/h)	59	-0.095	0.474	179	0.305	<0.001
MCHC (g/L)	82	0.092	0.413	248	-0.160	0.011
MCH (pg)	82	0.047	0.675	248	0.153	0.016
RDW (%)	82	-0.003	0.979	248	0.172	0.007
Platelet count (× 10 ⁹ /L)	81	-0.053	0.640	248	-0.235	<0.001
MPV (fL)	82	-0.090	0.424	248	0.124	0.051
WBC (× 10 ⁹ /L)	82	-0.089	0.428	248	-0.031	0.629

Table 4. Correlations between biochemical and hematological parameters and age. NA not applicable.

LDLc, HDLc and TG concentrations in the DS population exist. However, they report contradictory results and prevent firm conclusions from being drawn. Some studies have reported an unfavorable^{41–45} or favorable lipid profile⁴⁶. However, most of the studies reported no change in serum TC, LDLc or HDLc in individuals with DS compared to a control group or to population norms^{41,45,47–51}. In our study, these lower total cholesterol and TG concentrations may have translated into a significantly lower prevalence of hyperlipidemia in DS individuals. It has been described that DS individuals may be protected against atherosclerosis^{47,52–54}, leading to a low incidence of cardiovascular events⁵³. However, a work carried out with 4,081 individuals with DS found that they were at high risk of cerebrovascular events, but a lower risk of coronary events in males⁵⁵. Therefore, risk of major cerebrovascular events in people with DS should not be ruled out. Concerning diabetes mellitus, a similar incidence of type 2 diabetes mellitus⁵⁰ and a higher incidence of type 1 diabetes mellitus has been described for individuals with DS⁵⁶. We found no difference in type 1 diabetes mellitus frequency among our DS and control populations as previously described in a different study¹⁶. In regard to arterial hypertension prevalence, our results are in line with numerous studies that have described a lower incidence of this condition in DS individuals^{50,51,57,58}. Despite these observations, cholesterol fractionated forms and glycated hemoglobin (HbA1c) concentrations were not measured, making it difficult to draw conclusions regarding dyslipidemia and diabetes mellitus in our cohort. Yet, an increased degree of hypolipidemia should not be ruled out. Overall, future studies elucidating the mechanisms behind the low cholesterol and TG concentrations and lower prevalence of arterial hypertension observed in our DS cohort should be performed.

It is important to emphasize that our main goal was to help determining if the observed biochemical and hematological alterations have direct clinical implications for DS individuals. While the altered biochemical and hematological profiles may be developmental features (i.e., a consequence of the specific genetic characteristics of individuals with DS) or the result of accelerated aging, it should be recalled that they may also be reflecting comorbidities or the use of medication. From a clinical standpoint, to elucidate if the observed differences are consequence of concomitant conditions or features of the syndrome itself could be of help in the management of DS individuals. Unfortunately, due to the design of our study, these questions remain unanswered. Future

studies focusing on specific areas of metabolism of DS individuals with different comorbidities could shed some light on this matter.

Our study has several strengths. We collected relevant clinical, biochemical and hematological data in a large DS cohort and performed a systematic analysis. The fact that our controls were chosen from a healthy background broadens the actual differences and strengthens the present results. Ultimately, according to the wide inclusion criteria and the broad range of represented ages, we believe that the results from our study may help clinicians when interpreting laboratory analyses in DS individuals. Some limitations should also be taken into account. The control and DS populations were not strictly age and sex matched and the control group had a reduced number of males when compared to females. Nonetheless, both populations were within the same age range and additional analysis including adjustment for age and sex were performed. Furthermore, despite our large cohort of DS individuals, the number was still not sufficient to perform statistical analysis stratification according to the observed clinical conditions. Moreover and as stated previously, some of the observed biochemical and/or hematological alterations may have been a consequence of the use of drugs for the treatment of other comorbidities. Finally, our defined clinical, biochemical and hematological profiles were somehow general and unable to cover all the possible comorbidities present in DS individuals.

In conclusion, adults with DS show a specific profile of biochemical and hematological parameters. A good understanding of the differences in our cohort with those in the general population might aid in the clinical follow-up of adults with DS, especially considering that the life span of DS individuals can now reach 60 years of age in developed countries.

Received: 26 March 2020; Accepted: 30 July 2020

Published online: 13 August 2020

References

- Parker, S. E. *et al.* Updated national birth prevalence estimates for selected birth defects in the United States, 2004–2006. *Birth Defects Res. A Clin. Mol. Teratol.* **88**, 1008–1016 (2010).
- Canfield, M. A. *et al.* National estimates and race/ethnic-specific variation of selected birth defects in the United States, 1999–2001. *Birth Defects Res. A Clin. Mol. Teratol.* **76**, 747–756 (2006).
- Besser, L. M., Shin, M., Kucik, J. E. & Correa, A. Prevalence of down syndrome among children and adolescents in metropolitan Atlanta. *Birth Defects Res. A Clin. Mol. Teratol.* **79**, 765–774 (2007).
- Yang, Q., Rasmussen, S. A. & Friedman, J. M. Mortality associated with Down's syndrome in the USA from 1983 to 1997: A population-based study. *Lancet* **359**, 1019–1025 (2002).
- Bittles, A. H. & Glasson, E. J. Clinical, social, and ethical implications of changing life expectancy in Down syndrome. *Dev. Med. Child Neurol.* **46**, 282–286 (2004).
- Morris, J. K. & Alberman, E. Trends in Down's syndrome live births and antenatal diagnoses in England and Wales from 1989 to 2008: Analysis of data from the National Down Syndrome Cytogenetic Register. *BMJ* **339**, b3794 (2009).
- Startin, C. M. *et al.* Health comorbidities and cognitive abilities across the lifespan in Down syndrome. *J. Neurodev. Disord.* **12**, 4 (2020).
- Hithersay, R. *et al.* Association of dementia with mortality among adults with Down syndrome older than 35 years. *JAMA Neurol.* **76**, 152–160 (2019).
- Bull, M. J. Health supervision for children with Down syndrome. *Pediatrics* **128**, 393–406 (2011).
- Roizen, N. J. & Patterson, D. Down's syndrome. *Lancet* **361**, 1281–1289 (2003).
- Hawli, Y., Nasrallah, M. & El-Hajj Fuleihan, G. Endocrine and musculoskeletal abnormalities in patients with Down syndrome. *Nat. Rev. Endocrinol.* **5**, 327–34 (2009).
- Sakadamis, A., Angelopoulou, N., Matziari, C., Papamelioti, V. & Souftas, V. Bone mass, gonadal function and biochemical assessment in young men with trisomy 21. *Eur. J. Obstet. Gynecol. Reprod. Biol.* **100**, 208–212 (2002).
- Niegawa, T. *et al.* Evaluation of uric acid levels, thyroid function, and anthropometric parameters in Japanese children with Down syndrome. *J. Clin. Biochem. Nutr.* **61**, 146–152 (2017).
- Costa, R. *et al.* Bone mineral density distribution curves in Spanish adults with Down syndrome. *J. Clin. Densitom.* **21**, 493–500 (2018).
- Lima, A. S., Cardoso, B. R. & Cozzolino, S. F. Nutritional status of zinc in children with Down syndrome. *Biol. Trace Elem. Res.* **133**, 20–28 (2010).
- Real de Asua, D., Parra, P., Costa, R., Moldenhauer, F. & Suarez, C. Evaluation of the impact of abdominal obesity on glucose and lipid metabolism disorders in adults with Down syndrome. *Res. Dev. Disabil.* **35**, 2942–9 (2014).
- Fortea, J. *et al.* Plasma and CSF biomarkers for the diagnosis of Alzheimer's disease in adults with Down syndrome: A cross-sectional study. *Lancet Neurol.* **17**, 860–869 (2018).
- Gimenez, S. *et al.* Prevalence of sleep disorders in adults with Down syndrome: A comparative study of self-reported, actigraphic, and polysomnographic findings. *J. Clin. Sleep Med.* **14**, 1725–1733 (2018).
- Alcolea, D. *et al.* The Sant Pau Initiative on Neurodegeneration (SPIN) cohort: A data set for biomarker discovery and validation in neurodegenerative disorders. *Alzheimers Dement. (N Y)* **5**, 597–609 (2019).
- Blesa, R., Trias, C., Fortea, J., Videla, S. Alzheimer's disease in adults with Down syndrome: a challenge. *T21 Res. Soc. Soc. Bull.* **2**, 4 (2015).
- Tuck, M. K. *et al.* Standard operating procedures for serum and plasma collection: Early detection research network consensus statement standard operating procedure integration working group. *J. Proteome Res.* **8**, 113–117 (2009).
- Medicine, C. a. L. S. I. F. o. C. C. a. L. *Defining, Establishing and Verifying Reference Intervals in the Clinical Laboratory: Approved Guideline*, 3rd edn. (2010).
- Patterson, D. Molecular genetic analysis of Down syndrome. *Hum. Genet.* **126**, 195–214 (2009).
- Romualdi, D. *et al.* Low AMH levels as a marker of reduced ovarian reserve in young women affected by Down's syndrome. *Menopause* **23**, 1247–1251 (2016).
- Cohen, W. I., Patterson, B. & Group, D. S. M. I. Health care guidelines for individuals with Down syndrome: 1999 revision (Down syndrome preventive medical checklist). **4**, 1–15 (1999).
- Culp-Hill, R. *et al.* Red blood cell metabolism in Down syndrome: Hints on metabolic derangements in aging. *Blood Adv.* **1**, 2776–2780 (2017).
- Chou, S. T. *et al.* Trisomy 21 enhances human fetal erythro-megakaryocytic development. *Blood* **112**, 4503–4506 (2008).

28. Tunstall-Pedoe, O. *et al.* Abnormalities in the myeloid progenitor compartment in Down syndrome fetal liver precede acquisition of GATA1 mutations. *Blood* **112**, 4507–4511 (2008).
29. Maclean, G. A. *et al.* Altered hematopoiesis in trisomy 21 as revealed through in vitro differentiation of isogenic human pluripotent cells. *Proc. Natl. Acad. Sci. U S A* **109**, 17567–17572 (2012).
30. Roy, A. *et al.* Perturbation of fetal liver hematopoietic stem and progenitor cell development by trisomy 21. *Proc. Natl. Acad. Sci. U S A* **109**, 17579–17584 (2012).
31. De Vita, S. *et al.* Trisomic dose of several chromosome 21 genes perturbs haematopoietic stem and progenitor cell differentiation in Down's syndrome. *Oncogene* **29**, 6102–6114 (2010).
32. Pine, S. R. *et al.* Incidence and clinical implications of GATA1 mutations in newborns with Down syndrome. *Blood* **110**, 2128–2131 (2007).
33. Zipursky, A., Brown, E., Christensen, H., Sutherland, R. & Doyle, J. Leukemia and/or myeloproliferative syndrome in neonates with Down syndrome. *Semin. Perinatol.* **21**, 97–101 (1997).
34. Bajwa, R. P., Skinner, R., Windebank, K. P. & Reid, M. M. Demographic study of leukaemia presenting within the first 3 months of life in the Northern Health Region of England. *J. Clin. Pathol.* **57**, 186–188 (2004).
35. Massey, G. V. *et al.* A prospective study of the natural history of transient leukemia (TL) in neonates with Down syndrome (DS): Children's Oncology Group (COG) study POG-9481. *Blood* **107**, 4606–4613 (2006).
36. Homans, A. C., Verissimo, A. M. & Vlacha, V. Transient abnormal myelopoiesis of infancy associated with trisomy 21. *Am. J. Pediatr. Hematol. Oncol.* **15**, 392–399 (1993).
37. Graber, E., Chacko, E., Regelman, M. O., Costin, G. & Rapaport, R. Down syndrome and thyroid function. *Endocrinol. Metab. Clin. N. Am.* **41**, 735–745 (2012).
38. O'Grady, M. J. & Cody, D. Subclinical hypothyroidism in childhood. *Arch. Dis. Child* **96**, 280–284 (2011).
39. Guzman, R., Campos, C., Lopez-Fernandez, E. & Casado, A. Biomarkers of age effect on renal function in Down syndrome. *Biomarkers* **16**, 679–685 (2011).
40. Malaga, S., Pardo, R., Malaga, I., Orejas, G. & Fernandez-Toral, J. Renal involvement in Down syndrome. *Pediatr. Nephrol.* **20**, 614–617 (2005).
41. Nishida, Y., Akaoka, I., Nishizawa, T., Maruki, M. & Maruki, K. Hyperlipidaemia in patients with Down's syndrome. *Atherosclerosis* **26**, 369–372 (1977).
42. Pueschel, S. M., Craig, W. Y. & Haddow, J. E. Lipids and lipoproteins in persons with Down's syndrome. *J. Intellect. Disabil. Res.* **36**(Pt 4), 365–369 (1992).
43. Adekan, T., Magge, S., Shults, J., Stallings, V. & Stettler, N. Lipid profiles of children with Down syndrome compared with their siblings. *Pediatrics* **129**, e1382–e1387 (2012).
44. Zamorano, A., Guzman, M., Aspillaga, M., Avendano, A. & Gatica, M. Concentrations of serum lipids in children with Down's syndrome. *Arch. Biol. Med. Exp. (Santiago)* **24**, 49–55 (1991).
45. Draheim, C. C., McCubbin, J. A. & Williams, D. P. Differences in cardiovascular disease risk between nondiabetic adults with mental retardation with and without Down syndrome. *Am. J. Ment. Retard* **107**, 201–211 (2002).
46. Licastro, F. *et al.* Does Down's syndrome support the homocysteine theory of atherogenesis? Experience in elderly subjects with trisomy 21. *Arch. Gerontol. Geriatr.* **43**, 381–387 (2006).
47. Draheim, C. C., Geijer, J. R. & Dengel, D. R. Comparison of intima-media thickness of the carotid artery and cardiovascular disease risk factors in adults with versus without the Down syndrome. *Am. J. Cardiol.* **106**, 1512–1516 (2010).
48. Braunschweig, C. L. *et al.* Nutritional status and risk factors for chronic disease in urban-dwelling adults with Down syndrome. *Am. J. Ment. Retard* **109**, 186–193 (2004).
49. Tansley, G., Holmes, D. T., Lutjohann, D., Head, E. & Wellington, C. L. Sterol lipid metabolism in down syndrome revisited: down syndrome is associated with a selective reduction in serum brassicasterol levels. *Curr. Gerontol. Geriatr. Res.* **2012**, 179318 (2012).
50. Real de Asua, D., Parra, P., Costa, R., Moldenhauer, F. & Suarez, C. A cross-sectional study of the phenotypes of obesity and insulin resistance in adults with Down syndrome. *Diabetes Metab. J.* **38**, 464–471 (2014).
51. Parra, P., Costa, R., de Asua, D. R., Moldenhauer, F. & Suarez, C. Atherosclerotic surrogate markers in adults with Down syndrome: A case-control study. *J. Clin. Hypertens. (Greenwich)* **19**, 205–211 (2017).
52. Rodrigues, A. N. *et al.* Stiffness of the large arteries in individuals with and without Down syndrome. *Vasc. Health Risk Manag.* **7**, 375–381 (2011).
53. Yla-Herttuala, S., Luoma, J., Nikkari, T. & Kivimaki, T. Down's syndrome and atherosclerosis. *Atherosclerosis* **76**, 269–272 (1989).
54. Murdoch, J. C., Rodger, J. C., Rao, S. S., Fletcher, C. D. & Dunnigan, M. G. Down's syndrome: an atheroma-free model?. *Br. Med. J.* **2**, 226–228 (1977).
55. Sobey, C. G. *et al.* Risk of major cardiovascular events in people with Down syndrome. *PLoS ONE* **10**, e0137093 (2015).
56. Bergholdt, R., Eising, S., Nerup, J. & Pociot, F. Increased prevalence of Down's syndrome in individuals with type 1 diabetes in Denmark: A nationwide population-based study. *Diabetologia* **49**, 1179–1182 (2006).
57. Richards, B. W. & Enver, F. Blood pressure in Down's syndrome. *J. Ment. Defic. Res.* **23**, 123–135 (1979).
58. van Schrojenstein Lantman-de Valk, H.M. *et al.* Prevalence and incidence of health problems in people with intellectual disability. *J. Intellect. Disabil. Res.* **41** (Pt 1), 42–51 (1997).

Acknowledgements

The authors would like to thank all the participants with Down syndrome, their families and their caretakers for their support of and dedication to this research. We also acknowledge the Fundació Catalana Síndrome de Down for global support; Laia Muñoz, Soraya Torres, and Raúl Núñez for laboratory and sample handling; Reyes Alcoverro, Marta Salinas, and Tania Martínez for administrative support; and Concepción Escolà for nursing handling. This study was supported by the Fondo de Investigaciones Sanitario (FIS), Instituto de Salud Carlos III (PI18/00164 to FB-V and MT, PI14/01126 and PI17/01019 to JF, PI13/01532 and PI16/01825 to RB, PI18/00335 to MCI and PI14/1561, PI17/01896 to AL), the CIBERNED program (Program 1, Alzheimer's Disease to Alberto Lleó and SIGNAL study, www.signalstudy.es), and CIBERDEM, partly jointly funded by the Fondo Europeo de Desarrollo Regional, Unión Europea, Una manera de hacer Europa. This work was also supported by the National Institutes of Health (NIA grants 1R01AG056850-01A1; R21AG056974 and R01AG061566 to JF), Departament de Salut de la Generalitat de Catalunya, Pla Estratègic de Recerca i Innovació en Salut (SLT002/16/00408 to AL), Fundació La Marató de TV3 (20141210 to JF and 044412 to RB). The Fundació Catalana Síndrome de Down and Fundació Víctor Grífols i Lucas partially supported this work. This work was also supported by the Generalitat de Catalunya (SLT006/17/00119 to JF) and a grant from the Fundació Bancaria La Caixa to RB. DdG-C was a recipient of a Juan de la Cierva-Incorporación grant from the Ministry of Science Innovation and Universities (IJCI-2016-29393).

Author contributions

Designing studies: D.d.G.-C., J.F. and M.T.; Conducting sample analysis: M.N.N. and J.R.; Analyzing data: D.d.G.-C., F.B., I.B., J.F. and M.T.; Patient handling: I.B., D.G., M.C.-I., B.B., L.V., S.F., M.A., S.V., R.B., A.L., and J.F.; Writing the manuscript: D.d.G.-C. and M.T. Editing the final version of the manuscript: D.d.G.-C., J.F., F.B. and M.T.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information is available for this paper at <https://doi.org/10.1038/s41598-020-70719-2>.

Correspondence and requests for materials should be addressed to M.T.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2020