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## **International Consensus Guidelines for the Optimal Use of the Polymyxins: Endorsed by the American College of Clinical Pharmacy (ACCP), European Society of Clinical Microbiology and Infectious Diseases (ESCMID), Infectious Diseases Society of America (IDSA), International Society for Anti-infective Pharmacology (ISAP), Society of Critical Care Medicine (SCCM), and Society of Infectious Diseases Pharmacists (SIDP)†**

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## Abstract

The polymyxin antibiotics colistin (polymyxin E) and polymyxin B became available in the 1950s and thus did not undergo contemporary drug development procedures. Their clinical use has recently resurged, assuming an important role as salvage therapy for otherwise untreatable gram-negative infections. Since their reintroduction into the clinic, significant confusion remains due to the existence of several different conventions used to describe doses of the polymyxins, differences in their formulations, outdated product information, and uncertainties about susceptibility testing that has led to lack of clarity on how to optimally utilize and dose colistin and polymyxin B. We report consensus therapeutic guidelines for agent selection and dosing of the polymyxin antibiotics for optimal use in adult patients, as endorsed by the American College of Clinical Pharmacy (ACCP), Infectious Diseases Society of America (IDSA), International Society of Anti-Infective Pharmacology (ISAP), Society for Critical Care Medicine (SCCM), and Society of Infectious Diseases Pharmacists (SIDP). The European Society for Clinical Microbiology and Infectious Diseases (ESCMID) endorses this document as a consensus statement. The overall conclusions in the document are endorsed by the European Committee on Antimicrobial Susceptibility Testing (EUCAST). We established a diverse international expert panel to make therapeutic recommendations regarding the pharmacokinetic and pharmacodynamic properties of the drugs and pharmacokinetic targets, polymyxin agent selection, dosing, dosage adjustment and monitoring of colistin and polymyxin B, use of polymyxin-based combination therapy, intrathecal therapy, inhalation therapy, toxicity, and prevention of renal failure. The treatment guidelines provide the first ever consensus recommendations for colistin and polymyxin B therapy that are intended to guide optimal clinical use.

## Keywords

polymyxin B; colistin; dosing guidelines

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This practice guideline provides consensus recommendations pertaining to the clinical use of the polymyxin antibiotics, colistin (polymyxin E) and polymyxin B, for the treatment of bacterial infections in adults. The polymyxin antibiotics became available clinically in the 1950s and thus did not undergo contemporary drug development procedures.<sup>1</sup> Polymyxins have a unique mechanism of action involving disruption of the outer membrane integrity of

gram-negative bacteria that in addition to providing rapid bactericidal activity may enhance the activity of other antibiotic classes.<sup>1</sup> Their clinical use has recently resurged, and the polymyxins have assumed an important role as salvage therapy for otherwise untreatable gram-negative infections, most notably multidrug-resistant (MDR) and extensively drug-resistant (XDR) strains of *Pseudomonas aeruginosa*, *Acinetobacter baumannii*, and Enterobacteriaceae.<sup>2</sup>

Since their reintroduction into the clinic in the 1980s through today, significant confusion remains regarding polymyxin use due to differences in the formulations. Colistin is administered as an inactive prodrug, colistimethate (also known as colistin methanesulfonate [CMS]), whereas polymyxin B is administered in its active form. Also, different conventions are used to describe dosing of the polymyxins, particularly colistin; product information is outdated; and uncertainties remain regarding susceptibility testing.<sup>3, 4</sup> Thus a lack of clarity remains about how optimally to utilize and dose colistin and polymyxin B.<sup>5, 6</sup> Unfortunately, polymyxins are highly nephrotoxic agents, and acute kidney injury (AKI) occurs frequently with conventional doses.<sup>7, 8</sup> Given the narrow therapeutic windows (low therapeutic indices) of polymyxins, this guideline provides clinicians with a practical framework for use in treating infections caused by MDR and XDR gram-negative pathogens.

## Methods

### Consensus Panel Composition

The Consensus Panel was composed of international experts. They represent membership in the endorsing organizations (American College of Clinical Pharmacy [ACCP], the European Society for Clinical Microbiology and Infectious Diseases [ESCMID], the Infectious Diseases Society of America [IDSA], International Society of Anti-Infective Pharmacology [ISAP], Society of Critical Care Medicine [SCCM], and the Society of Infectious Diseases Pharmacists [SIDP]).

### Consensus Development Based on Evidence

Consensus Panel members were assigned key topics that contribute to current knowledge and optimal utilization of the polymyxins. A draft document addressing these areas that included specific recommendations was reviewed and approved by all panel members. The panel conducted face-to-face meetings and teleconferences to complete the guideline work. The purpose of the meetings and teleconferences was to determine and discuss the clinical questions to be addressed, assign topics for review and writing of the initial draft, and develop recommendations. The entire panel reviewed all sections. After review by members of the ACCP, ESCMID, IDSA, SCCM, ISAP, and SIDP, the panel reviewed the submitted comments and recommendations. After careful discussion and consideration of these suggestions, the document was revised and circulated among the panel and supporting societies for final approval.

### Literature Review and Analysis

The recommendations in this guideline were developed following a review of studies published before December 31, 2018, in English. Studies were identified through Library of

Congress, LISTA (Library, Information Science & Technology Abstracts [EBSCO]), and PubMed database searches with no date restrictions using Medical Subject Headings. Examples of keywords used to conduct literature searches were *polymyxin*, *colistin*, *polymyxin B*, *nephrotoxicity*, *pharmacokinetics*, *pharmacodynamics*, *area under the curve*, *toxicodynamics*, *resistance*, *carbapenem*, *A. baumannii*, *P. aeruginosa*, and *Klebsiella pneumoniae*.

## Process Overview

To evaluate evidence, the panel followed a process consistent with other contemporary guidelines. The process for evaluation was based on the Grading of Recommendations Assessment, Development, and Evaluation (GRADE) system, a newly created system for grading the quality of evidence and strength of recommendations for health care.<sup>9</sup> Some topics were determined to be ungradable such as those that involved nonclinical evidence (such as recommendations for in vitro minimum inhibitory concentration [MIC] breakpoints) and thus were not evaluated according to the GRADE criteria. Some recommendations were labeled as best practice recommendations, particularly in scenarios where the recommendations lack sufficient randomized controlled trial (RCT) evidence. Panel members were divided into groups consisting of a primary lead author and coauthors for each section. Each author was asked to review the literature, evaluate the evidence, develop and determine the strength of recommendations, and provide an evidence summary supporting each recommendation. The panel reviewed all recommendations, the assigned strength of the recommendations, and quality of evidence. Discrepancies were discussed and resolved. We acknowledge this as a potential limitation. Similar to other guidelines, some of the evidence used to establish the recommendations was published by the authors writing each section.

## Clinical Questions and Recommendations

### Susceptibility and Pharmacokinetics/Pharmacodynamics

#### I. How Should Susceptibility Be Tested, and What Are the Minimum Inhibitory Concentration Breakpoints for the Polymyxins to Guide Therapy? Recommendation.

**RI:** The joint European Committee on Antimicrobial Susceptibility Testing (EUCAST) and Clinical and Laboratory Standards Institute (CLSI) polymyxin breakpoint working group recommended that standard broth microdilution ISO-74 20776<sup>10</sup> be used as the reference method for the MIC testing of colistin and be performed with cation-adjusted Mueller Hinton broth, with sulfate salts of colistin in plain polystyrene trays without additives such as polysorbate-80.<sup>11, 12</sup> Sulphate salts of polymyxins must be used (the methanesulfonate derivative of colistin must not be used - it is an inactive pro-drug that breaks down slowly in solution).<sup>11, 12</sup> Agar dilution, disk diffusion, and gradient diffusion are not currently recommended by CLSI-EUCAST as these methods yield unacceptably high error rates compared to broth microdilution.<sup>11-13</sup> We recommend that the CLSI/EUCAST Joint Working Group clinical breakpoints be used for colistin (Table 1).

**Evidence Summary.:** CLSI<sup>14</sup> and EUCAST<sup>15</sup> established a Joint Working Group regarding susceptibility testing and breakpoints for colistin.<sup>11, 12</sup> Polymyxin B was not addressed by this group. The CLSI/EUCAST Joint Working Group recommended clinical breakpoints that are harmonized for *Acinetobacter* sp and *P. aeruginosa*. These recommendations were approved by the CLSI Antimicrobial Susceptibility Testing Subcommittee in 2016.<sup>11,12</sup> Breakpoints for Enterobacteriaceae were also considered. However, there were insufficient data, and a clinical breakpoint was not established. Rather, an epidemiological cutoff value (ECV) was defined, based on the MIC distribution data for *Klebsiella aerogenes*, *Enterobacter cloacae*, *Escherichia coli*, *K. pneumoniae*, and *Raoultella ornithinolytica*. It is important to note that CLSI recommended that the ECV should be applied only to these species because wild-type MIC distributions may be different for other genera and species of Enterobacteriaceae. Thus the clinical breakpoints for colistin provided by the CLSI for *P. aeruginosa* and *Acinetobacter* sp were a susceptible breakpoint of 2 mg/L or lower and a resistant breakpoint of 4 mg/L or higher.<sup>12</sup> EUCAST breakpoints for colistin for *P. aeruginosa*, *Acinetobacter* sp and Enterobacteriaceae are a susceptible breakpoint of 2 mg/L or lower and a resistant breakpoint of greater than 2 mg/L (Table 1).<sup>15</sup>

**Future Research Needs.:** Research should be directed toward defining reliable testing methods for colistin that are more convenient than microdilution techniques. Rapid diagnostics of polymyxin resistance<sup>16-18</sup> and defining Enterobacteriaceae MIC breakpoints are necessary. Because polymyxin B breakpoints have not been established, future research is necessary to evaluate and define clinical breakpoints independently for all species.

## II. Is There a Recommended PK/PD Therapeutic Target for Maximization of Efficacy for Colistin and Polymyxin B?

### Recommendations.

**R2:** We recommend that for colistin, an area under the plasma concentration-time curve across 24 hours at steady state ( $AUC_{ss,24\text{ hr}}$ ) of  $\sim 50$  mg · hour/L is required that equates to a target average steady-state plasma concentration ( $C_{ss,avg}$ ) of  $\sim 2$  mg/L for total drug. Although this target might be suboptimal for lower respiratory tract infections, it is noted that this should be considered as a maximum tolerable exposure. Concentrations higher than this were shown to increase both the incidence and severity of AKI.

**R3:** We recommend similar targets for polymyxin B as those listed for colistin. However, we note that data are lacking for  $AUC_{ss,24\text{ hr}}$  targets for polymyxin B. Emerging evidence suggest a different toxicodynamic (TD) profile for polymyxin B than colistin. Some evidence indicates that an  $AUC_{ss,24\text{ hr}}$  target of 50–100 mg · hour/L, corresponding to a  $C_{ss,avg}$  of 2–4 mg/L, may be acceptable from a toxicity standpoint.

**R4:** We recommend that the exposures just described for polymyxin B and colistin should be considered the maximal tolerable exposures. Although these recommended exposures should achieve bacterial killing at the current MIC breakpoints based on the mouse thigh infection model, both colistin and polymyxin B when administered systemically (i.e., not directly into the lungs) were shown in the mouse lung infection model to be substantially less effective.

**Evidence Summary:** The in vitro PD activity of colistin and polymyxin B is virtually indistinguishable.<sup>19–21</sup> Both polymyxins demonstrate rapid bactericidal killing against susceptible strains of *P. aeruginosa*,<sup>22, 23</sup> *A. baumannii*,<sup>19, 20</sup> and *K. pneumoniae*.<sup>24, 25</sup> Concentrations above the MIC result in extremely rapid initial killing, with large decreases in colony-forming units per milliliter (cfu/mL) occurring as early as 5 minutes following exposure.<sup>22</sup> A modest postantibiotic effect is evident for high concentrations of colistin and polymyxin B.<sup>22</sup> The PK/PD linked parameter of the polymyxins was investigated in in vitro PK models and animal models. Clearly, for colistin, in vitro<sup>26, 27</sup> and animal studies<sup>28, 29</sup> point to the free-drug area under the concentration-time curve to MIC ratio (*f*AUC:MIC) as the PK/PD index that is best correlated with efficacy. Fewer preclinical data are available for polymyxin B.<sup>20, 21, 29, 30</sup> However, they also suggest that *f*AUC:MIC is the PK/PD index that correlates best with antibacterial activity. Because colistin and polymyxin B have very similar molecular structures and in vitro activity,<sup>1, 31</sup> it is reasonable to conclude that polymyxin B PK/PD indices and targets approach those of colistin.

Studies have elucidated the *f*AUC:MIC target for colistin in both in vitro systems and in animals. The most recent studies<sup>29</sup> of systemically administered colistin against *A. baumannii* and *P. aeruginosa* in murine thigh and lung infection models were used to determine *f*AUC:MIC targets for various magnitudes of bacterial kill and, as discussed earlier, to establish MIC breakpoints. For colistin, the *f*AUC:MIC values to obtain a 2 log<sub>10</sub> reduction in bacterial count in the experimental thigh infection model ranged from 7.4–13.7 for *P. aeruginosa* and from 7.4–17.6 for *A. baumannii*. The *f*AUC:MIC values to obtain a 1 log<sub>10</sub> reduction in bacterial count in experimental thigh infection ranged from 6.6–10.9 for *P. aeruginosa* and from 3.5–13.9 for *A. baumannii*. Target *f*AUC:MIC values for 1 and 2 log<sub>10</sub> kill in the lung infection model were substantially higher. Indeed, for *A. baumannii*, it was not even possible to achieve bacteriostasis for two of the three tested strains with the highest tolerable systemic dosage regimen of colistin.<sup>29</sup>

Based on these data, a target plasma colistin  $C_{ss,avg}$  of 2 mg/L was recommended for systemic administration of CMS.<sup>6, 32, 33</sup> This target is based on the following considerations. First, it accounts for the difference in the extent of protein binding between the plasma of mice and critically ill patients.<sup>6, 32, 33</sup> The protein binding in human plasma is ~50%. Second, based on the thigh infection model, this exposure would be expected to achieve bactericidal activity against an isolate with an MIC of 2 mg/L (the EUCAST and CLSI breakpoint). It is important to note that, unless the MIC of the infecting strain is well below the breakpoint, this target is very likely to be suboptimal for the systemic treatment of a lung infection.<sup>29, 30</sup> Third, it is considered unwise to target a higher plasma colistin  $C_{ss,avg}$  because pharmacokinetic/toxicodynamic (PK/TD) analyses in patients demonstrated that concentrations greater than 2 mg/L are associated with an increase in both the incidence and severity of AKI.<sup>34–36</sup> Therefore, the proposed target concentrations of colistin should be considered the maximal tolerable target. Finally, even though a plasma colistin  $C_{ss,avg}$  less than 2 mg/L may be adequate for an isolate with a low MIC, the susceptibility of the organism is often not known at the initiation of therapy, and therefore, a target of 2 mg/L is appropriate when starting CMS. Furthermore, given inaccuracies with antibiotic

susceptibility testing with the polymyxins, relying on the reported MIC may lead to suboptimal exposures.<sup>37</sup>

One group<sup>30</sup> recently reported the results of PK/PD studies for systemically administered polymyxin B against *K. pneumoniae* in murine thigh and lung infection models. The target values for 1 log<sub>10</sub> reduction in bacterial count in the thigh model (*f*AUC:MIC 3.72–28.0) were similar to those for colistin for the same magnitude of bacterial kill. Unlike colistin, 2 log<sub>10</sub> kill in the thigh model was not achieved even at the highest tolerated dose of polymyxin B. Similar to findings with colistin, polymyxin B was substantially less effective against lung infections and was not able to achieve stasis against any strain, even at the highest tolerated systemic dose.

For polymyxin B, clinical PK/TD data are scarce, and as described in detail later, it appears to differ from CMS with regard to the risk of AKI with currently used doses. In the absence of direct quantitative data to establish an exposure–toxicity relationship, clinicians should consider data derived from a meta-analysis of 16 studies involving a total of 971 subjects who received intravenous (IV) polymyxin B.<sup>38</sup> Pharmacokinetic exposures in patients in these studies were simulated based on patient characteristics and dosing information given in each study and published PK parameters for polymyxin B. The 25th, 50th and 75th percentiles of estimated polymyxin B AUC<sub>SS</sub> were 46.7, 58.6, and 78.1 mg · hour/L, respectively. Importantly, across all studies, 26.4% of patients displayed a 50% or higher decrease in creatinine clearance (Cl<sub>cr</sub>). Based on these findings, some experts suggest a target AUC<sub>SS,24 hr</sub> as high as 100 mg · hour/L for polymyxin B.<sup>39</sup> However, based on the recent lung infection model data for systemically administered polymyxin B against *K. pneumoniae*,<sup>30</sup> these higher exposures may still be insufficient to achieve killing in respiratory tract infections. Thus the benefit (and true toxicity risk) of these higher exposures remains unclear. Therefore, the panel recommends the same target exposures as for colistin (AUC<sub>SS</sub> of ~50 mg · hour/L).

It is important to note that the recommended PK/PD exposure targets have been derived from studies involving polymyxin monotherapy. Thus the PK/PD targets should apply to polymyxin monotherapy. Recent Hollow Fiber Infection Model studies conducted in vitro using a high bacterial density of organism and in the absence of an immune system demonstrated a paradoxical effect for the polymyxins whereby higher doses of polymyxin B and colistin further amplified high-level polymyxin resistance.<sup>20, 23, 40, 41</sup> An inoculum effect was demonstrated for polymyxin monotherapy with bacterial killing activity significantly attenuated at inocula consistent with ventilator-associated pneumonia (VAP) or health care–associated pneumonia (HAP).<sup>20, 23, 40, 41</sup>

**Future Research Needs.:** Future research should be directed toward defining optimal exposure targets in critically ill patients to establish the relationship between polymyxin exposure in relation to clinical success and failure in this patient population. The high proportion of patients who fail polymyxin therapy, and other patient-related factors, make the establishment of PK/PD relationships in critically ill patients extremely complex. PK/PD targets of polymyxins should also be considered in the context of combination therapy. The

concentrations of polymyxins necessary to potentiate other agents would help determine whether safer exposures can be given in combination regimens.

### Polymyxin Pharmacokinetics

#### III. Should I Preferentially Use One Polymyxin Over the Other?

##### Recommendations.

**R5:** We recommend that clinicians have access to parenteral products of both CMS and polymyxin B, so they can choose between the two in particular circumstances.

**R6:** We recommend polymyxin B as the preferred agent for routine systemic use in invasive infections. The rationale for this recommendation is that polymyxin B has superior PK characteristics in humans as well as a decreased potential to cause nephrotoxicity.

**R7:** We recommend colistin as the preferred polymyxin for the treatment of lower urinary tract infections given renal clearance of the prodrug CMS that then converts to the active moiety colistin in the urinary tract.

**Evidence Summary.:** There are several clinical pharmacologic differences between CMS/colistin and polymyxin B administered IV.<sup>42, 43</sup> We point the reader to an excellent review that highlights the key differences between polymyxin B and colistin.<sup>42, 43</sup> Polymyxin B appears to have superior clinical PK characteristics for infections where it is important to achieve rapidly and reliably and then maintain a desired concentration in plasma. In critically ill patients receiving IV CMS, plasma concentrations of formed colistin rise slowly. Even with a loading dose of CMS at the initiation of therapy, it may take several hours to achieve plasma colistin concentrations that may be effective. Polymyxin B is not administered as a prodrug, and therefore it is possible to use an IV dose to achieve plasma concentrations more rapidly that may be effective. In addition, dose selection is more difficult for CMS because the PK of CMS and formed colistin are subject to substantially greater interpatient variability than occurs with polymyxin B.<sup>42, 43</sup> Moreover, in patients with good renal function ( $Cl_{cr}$  greater than 80 mL/minute), it is not possible to attain reliably a plasma colistin  $C_{ss,avg}$  of 2 mg/L, a concentration regarded as a reasonable initial target when MIC is unknown (see Section II),<sup>6, 32, 33</sup> even with daily doses of CMS at the upper end of approved doses (see Section VI).<sup>6, 32</sup> The pharmacokinetics of polymyxin B are not similarly affected by renal function, and therefore it is possible to attain a plasma polymyxin B  $C_{ss,avg}$  of 2 mg/L reliably with approved daily doses, even in patients with  $Cl_{cr}$  greater than 80 mL/minute (see Section XI).<sup>44-47</sup>

The risk of AKI appears to be less with polymyxin B,<sup>48-54</sup> although some of the comparative studies are confounded by issues with different experimental designs.<sup>8, 42, 43</sup> Therapeutic drug monitoring (TDM) is inherently more difficult for colistin because of the need to ensure that samples are collected in such a way as to minimize ongoing in vitro conversion of CMS to colistin. However, CMS may be the preferred agent for the IV treatment of urinary tract infections. Urinary concentrations of colistin after administration of CMS (mainly cleared by renal excretion) can be high because of conversion of CMS to



colistin in the urinary tract.<sup>4, 43, 55, 56</sup> In contrast, polymyxin B is predominantly cleared by nonrenal mechanisms with a median urinary recovery of 4.0% in patients.<sup>44</sup>

**Future Research Needs.:** Although prospective RCTs comparing parenteral polymyxin B and colistin in patients with various types of infections are unlikely to be conducted, any comparative observational data would further elucidate the efficacy and toxicity differences between both polymyxins. In particular, well-controlled safety and efficacy studies comparing dose-optimized colistin versus polymyxin B are of great interest.

## Colistin Intravenous Dosing

### IV. For CMS, What Is the Relationship Between Different Dosing Units in the Literature?

#### Recommendation.

**R8:** We recommend that hospital guidelines and prescription orders specify doses of CMS in either number of international units (IU) or milligrams of colistin base activity (CBA), corresponding to the labeling convention used in the specific country. Because of the international scope of these guidelines, doses in the following sections are expressed in the approximate equivalents of both of these conventions. The conversion factor is 1 million IU is equivalent to ~33 mg CBA.

**Evidence Summary.:** Colistin is administered parenterally in the form of the inactive prodrug, CMS. Unfortunately, two different conventions are used in different parts of the world to label vials of parenteral CMS and to express doses for patients. Both conventions are based on microbiological assessment. The parenteral products of CMS available in Europe and some other parts of the world are labeled in terms of IU. In contrast, parenteral CMS vials available in North and South America and many other parts of the world are labeled in terms of CBA, another way of expressing microbiological activity.

As noted earlier, 1 million IU corresponds to ~33 mg CBA, and 1 million IU also corresponds to ~80 mg of the chemical CMS.<sup>57</sup> Thus it is critical that doses must not be prescribed in terms of milligrams of the chemical CMS.<sup>4</sup> When reading the scientific literature, clinicians must clearly understand whether doses reported in milligrams refer to CBA or the chemical CMS. Consistent global reporting of colistin doses is critically important to promote safe and effective use.<sup>58</sup>

**Future Research Needs.:** International harmonization is urgently needed to have a consistent approach to specify all doses in either numbers of IUs or milligrams of CBA.

### V. Do I Need to Administer an Intravenous Loading Dose When I Initiate Therapy with CMS?

#### Recommendation.

**R9:** We recommend initiating IV therapy with a CMS loading dose of 300 mg CBA (~9 million IU) infused over 0.5–1 hours and to administer the first maintenance dose 12–24 hours later.

**Evidence Summary.:** After initiation of CMS therapy in critically ill patients, plasma concentrations of formed colistin were reported to increase slowly over many hours or even days,<sup>33, 59–61</sup> although more rapid increases were also reported.<sup>62</sup> Such variation in the rate of concentration attainment of colistin is probably related to brand-to-brand or batch-to-batch differences in the complex chemical composition (degree of methanesulfonation) of the CMS administered to patients.<sup>63</sup> The case for a loading dose would be more compelling for a brand or batch that undergoes slow conversion. Unfortunately, there is no way of knowing (a priori) the rate of in vivo conversion for a particular batch. The impact of a loading dose on the risk of developing AKI is unclear.<sup>52, 54, 64</sup> Considering the need for timely antibiotic administration, the therapeutic benefits of a loading dose may justify the potential risk of AKI associated with loading dose.<sup>65–67</sup> The timing of the commencement of the maintenance dose should be based on the interval of the maintenance dose (e.g., if the patient is placed on colistin every 12 hours, the maintenance dose should start 12 hours later).

**Future Research Needs.:** More research is needed to define the brand-to-brand and batch-to-batch differences as they relate to degree of methanesulfonation and conversion to colistin. Additional data regarding the safety and efficacy of loading doses are needed.

## VI. What Should My Initial Daily Maintenance Dose of CMS Be in Patients with Normal Renal Function?

### Recommendation.

**RI0:** We recommend that for a patient with normal renal function, administer a daily dose of 300–360 mg CBA (~9–10.9 million IU), divided into two and infused over 0.5–1 hour at 12-hour intervals. Monitor renal function and adjust the daily dose accordingly using the recommendations in Table 2.

**Evidence Summary.:** Determining initial daily maintenance dose requires consideration of the desired target average steady-state plasma concentration ( $C_{ss,avg}$ ) of colistin. Based on translation of preclinical PK/PD data for *P. aeruginosa* and *A. baumannii* in murine thigh infection models and the ECV for *K. pneumoniae*,<sup>11, 12, 14, 29, 68</sup> clinical PK/TD data defining the relationship between plasma colistin exposure and risk of AKI in patients;<sup>34–36</sup> and the fact that the MIC of an isolate is often not known at initiation of therapy, a target plasma colistin  $C_{ss,avg}$  of 2 mg/L was suggested.<sup>6, 32</sup> This target may be appropriate for treatment of relatively accessible infections with organisms having colistin MICs of 2 mg/L or lower. However, it is important to recognize that murine lung infections with *P. aeruginosa* and *A. baumannii* were substantially more resilient to systemic treatment than were murine thigh infections.<sup>29</sup> Thus based on the preclinical data, a plasma colistin  $C_{ss,avg}$  of 2 mg/L achieved via IV administration may not be adequate for the treatment of lung infections in critically ill patients, especially those caused by organisms that have elevated MIC organisms.<sup>6, 29</sup>

The daily doses of CMS to achieve a target plasma colistin  $C_{ss,avg}$  of 2 mg/L (Table 2) were proposed based on analysis of PK data from more than 200 critically ill patients with a wide range of renal function.<sup>6</sup> For patients with a  $Cl_{cr}$  greater than 90 mL/minute, a suggested

maximum dose of 360 mg CBA (~10.9 million IU) per day was proposed because of limited clinical experience regarding the rate and impact of AKI with daily doses above this level. Even with the daily doses proposed for patients with  $Cl_{cr}$  greater than 90 mL/minute (Table 2), only 30–40% of patients are expected to achieve a plasma colistin  $C_{ss,avg}$  of 2 mg/L or more,<sup>6, 61</sup> although almost 80% of such patients may achieve a  $C_{ss,avg}$  of 1 mg/L or greater.<sup>6</sup>

Although weight-based dosing algorithms were proposed as alternatives in the U.S. package insert, such as those in a current RCT of colistin, <https://clinicaltrials.gov/ct2/show/NCT01597973?term=NCT01597973&rank=1>.<sup>69</sup> PK data do not support the need for weight-based dosing.

**Future Research Needs.:** The dose suggestions in Table 2 require validation by independent studies. In particular, these recommended doses need to be compared with lower historical dosing regimens to ensure the appropriate balance between safety and efficacy is achieved. Research is needed to define optimal dosing strategies in patients with  $Cl_{cr}$  greater than 80 mL/minute.

## VII. Do I Need to Adjust the Daily Maintenance Dose of CMS If the Patient Has Renal Impairment?

### Recommendation.

**R11:** We recommend that CMS dose adjustments be made in patients with renal insufficiency as provided in Table 2.

**Evidence Summary.:** The apparent clearance of colistin and hence the plasma colistin  $C_{ss,avg}$  achieved from a given daily dose of CMS is influenced by kidney function.<sup>6, 33, 62</sup> Therefore, the daily dose of CMS to target a plasma colistin  $C_{ss,avg}$  of 2 mg/L should be adjusted for renal impairment. Daily doses for patients with various degrees of renal function are provided in Table 2. The daily dose is divided into two doses, administered 12 hours apart, and each dose is infused over 0.5–1 hour. If the daily dose is not reduced in patients with decreased renal function, there is an increased probability that the plasma colistin  $C_{ss,avg}$  will be higher than 2 mg/L. This would be expected to increase antibacterial activity but is also expected to increase the likelihood of AKI.

**Future Research Needs.:** Although it is critical to adjust colistin doses in patients with renal impairment, definitive knowledge of the subsequent concentrations obtained requires TDM. Research is required to investigate the optimal approach to implementing TDM including identification of the patient groups most likely to benefit.

## VIII. Does Renal Replacement Therapy Have Implications for Selection of Intravenous CMS Dosage Regimens?

### Recommendations.

**R12:** We recommend that to target a plasma colistin  $C_{ss,avg}$  of 2 mg/L in a patient on intermittent hemodialysis (IHD), the following dosing schedule be utilized: On a nondialysis day, administer a CMS dose of 130 mg CBA/day (~3.95 million IU/day). On a dialysis day, administer a supplemental dose of CMS 40 mg CBA (~1.2 million IU) or 50 mg CBA (~1.6

million IU) for a 3- or 4-hour IHD session, respectively. If possible, the supplement to the baseline (nondialysis) daily dose should be administered with the next regular dose, after the dialysis session has ended. Conduct IHD sessions as late as possible within a CMS dosage interval to minimize the amount of CMS and formed colistin lost to the extracorporeal system.

**R13:** We recommend that to target a plasma colistin  $C_{ss,avg}$  of 2 mg/L in patients prescribed *sustained low-efficiency dialysis* (SLED) that 10% of the CMS dose be added to the baseline daily dose per 1 hour of SLED.

**R14:** We recommend that for patients prescribed continuous renal replacement therapy (CRRT), for a plasma colistin  $C_{ss,avg}$  of 2 mg/L, to administer CBA 440 mg/day (~13.3 million IU/day). This equates to 220 mg CBA every 12 hours (~6.65 million IU every 12 hours).

**Evidence Summary.:** Colistin methanesulfonate and formed colistin are efficiently cleared by intermittent and continuous renal support modalities; less information is available for SLED than for shorter forms of IHD and CRRT.<sup>6, 33, 70–77</sup> Supplementary doses of CMS are needed for patients receiving IHD or SLED. IHD, SLED, and CRRT, each removes ~10% of colistin an hour necessitating replacement of 10% of the daily dose per hour on these modalities. Because the duration of CRRT (24 hrs) is greater than the duration of SLED (often 8–10 hrs), which is greater than the duration of IHD (3–4 hrs), the supplementary doses needed differ significantly as a function of dialysis type. Apparent clearance of colistin and hence the dose requirements of CMS are greater in patients on CRRT than for patients with normal renal function.<sup>6, 33, 76</sup> Detailed dose suggestions for patients receiving renal support have been proposed.<sup>6, 75, 76</sup>

To target a plasma colistin  $C_{ss,avg}$  of 2 mg/L in patients prescribed SLED, it is recommended that 10% be added to the baseline daily dose per 1 hour of SLED. We provide the following practical example as an illustration.<sup>6</sup>

For a patient receiving a 10-hour nocturnal SLED session each day and receiving CMS every 12 hours:

- For a patient with  $Cl_{cr}$  of ~0 mL/minute, the CMS dose would be the sum of the baseline CMS dose (CBA dose of 130 mg/day [~3.95 million IU/day], Table 2) plus a supplementary dose comprising 10% of the baseline dose per hour × 10 hours.
- That is, for this patient, the CBA dose would be 260 mg/day (~7.9 million IU/day). In such a case, it may be most convenient and safe to administer 130 mg CBA every 12 hours (~3.95 million IU every 12 hours).

**Future Research Needs.:** Research is needed on colistin dosing in SLED patients, particularly with regard to the impact of different dialysis membranes on colistin removal. The preceding recommendations for SLED were based on small sample sizes with the use of

medium- to high-flux filters. Removal would be expected to be decreased with lower flux filters.

### Polymyxin B Intravenous Dosing

#### IX. Do I Need to Administer an Intravenous Loading Dose When I Initiate Therapy with Polymyxin B?

##### Recommendation.

**R15:** We recommend a loading dose of 2.0–2.5 mg/kg for polymyxin B, based on total body weight (TBW) (equivalent to 20,000–25,000 IU/kg) over 1 hour.

**Evidence Summary.:** A population PK study in critically ill patients showed that with a regimen of 1.25 mg/kg (equivalent to 12,500 IU/kg) every 12 hours, plasma polymyxin B concentrations achieved after the first dose were ~56–70% of the concentrations observed at steady state.<sup>44</sup> Using Monte Carlo simulations, it was estimated that with a loading dose of 2.0 mg/kg (equivalent to 20,000 IU/kg), day 1 exposures would likely be 76–94% of exposures at steady state.<sup>44</sup> There is a paucity of data regarding the clinical safety and efficacy of a polymyxin B loading dose strategy. However, one analysis found no association between loading dose of either polymyxin B or colistin and nephrotoxicity (adjusted hazard ratio [aHR] 0.78, 95% confidence interval [CI] 0.42–1.46).<sup>54</sup> In this analysis, 36 patients received an average polymyxin B loading dose of  $1.9 \pm 0.5$  mg/kg.<sup>54</sup> Conversely, although not statistically significant, loading doses were more frequently administered in patients who presented with neurotoxicity compared with patients who did not present with this adverse event (2 of 6 [33.3%] and 7 of 68 [10.3%], respectively;  $p=0.15$ ).<sup>78</sup>

Although it is reasonable to administer loading doses to all patients, priority should be given to those who are critically ill such as those with sepsis or septic shock. Pharmacokinetic data do not support capping upper absolute dose (i.e., expressed in milligrams) in obese patients. However, experience with the administration of more than 200 mg per infusion is limited,<sup>78, 79</sup> and infusion-related adverse effects, which include sudden thoracic pain, paresthesias, dizziness, dyspnea, and hypoxemia, were reported at a crude incidence of 0.9% (95% CI 0.2–3.2) and may increase with such doses.<sup>78</sup>

**Future Research Needs.:** Additional research is needed to define the safety and efficacy of high initial dose polymyxin B regimens. Although administration of doses higher than 3 mg/kg (equivalent to 30,000 IU/kg) were reported in patients,<sup>44, 78</sup> more data are needed on the safety as well as the clinical and microbiological impact of these regimens.

#### X. What Is the Recommended Initial Daily Maintenance Dose for Polymyxin B in Patients with Normal Renal Function?

##### Recommendation.

**R16:** We recommend that for patients with severe infections, a polymyxin B dose of 1.25–1.5 mg/kg (equivalent to 12,500–15,000 IU/kg TBW) every 12 hours is infused over 1 hour.

**Evidence Summary.:** As discussed earlier, considering that  $fAUC:MIC$  targets for 1  $\log_{10}$  kill for polymyxin B against *K. pneumoniae*<sup>30</sup> showed generally good agreement with the corresponding values for colistin against *P. aeruginosa* and *A. baumannii* in the murine thigh infection model,<sup>29</sup> and given the similar plasma unbound fractions (i.e., ~0.50) of polymyxin B<sup>44</sup> and colistin<sup>6</sup> in humans, a  $C_{ss,avg}$  of 2 mg/L seems to be an appropriate target for polymyxin B dosing guidance. This target may be revised as more information becomes available from preclinical studies to inform PK/PD relationships against gram-negative pathogens and from clinical studies to inform the PK/TD relationship for nephrotoxicity.

With doses of 2.5 and 3.0 mg/kg/day (equivalent to 25,000 and 30,000 IU/kg/day, respectively), 90% of patients, as determined by Monte Carlo simulations, would be expected to achieve an AUC of polymyxin B at steady state of at least 44.3 and 53.1 mg · hour/L, respectively,<sup>44</sup> that corresponds to  $C_{ss,avg}$  of 1.8 and 2.2 mg/L, respectively. Thus against isolates with polymyxin B MICs of 2 mg/L, the PK/PD target for 1  $\log_{10}$  kill of *P. aeruginosa*, *A. baumannii*, and *K. pneumoniae* in murine thigh infection<sup>29, 30</sup> will have an estimated probability of target attainment of more than 90% with either dosing strategy. Given the concerns mentioned earlier with antibacterial activity of systemically administered polymyxins in lung infections,<sup>29, 30</sup> higher plasma concentration targets might be necessary to achieve adequate antimicrobial activity in different infection sites. However, due to the lack of clinical safety data, a maintenance dose higher than 3 mg/kg/day (equivalent to 30,000 IU/kg/day) cannot be recommended at this time. A target of 2 mg/L is recommended even for isolates with an MIC lower than 2 mg/L in patients with severe infections.<sup>80</sup> Unfortunately, in the routine clinical microbiology laboratory setting, the MIC cannot be determined with enough accuracy at this stage, and a target of 2 mg/L therefore seems to be prudent in all cases.

Pharmacokinetic data do not support capping upper absolute doses (i.e., expressed in milligrams) in patients with high TBW. However, experience with infusions of more than 200 mg remains limited,<sup>78, 79</sup> and infusion-related adverse effects may increase with such doses.

No specific recommendation is available in the package insert concerning the duration of infusion. However, in recent PK analyses reflecting real-world use of polymyxin B, doses were safely administered over 1–4 hours in most patients.<sup>44–46, 81</sup> Because there might be a potential benefit on renal toxicity of higher peak-to-trough differences,<sup>82</sup> infusions over 1 hour might be preferred over longer infusions if well tolerated by patients.

**Future Research Needs.:** Additional research is needed to define the safety and efficacy associated with optimal maintenance dosing of polymyxin B.

## **XI. Do I Need to Adjust the Daily Polymyxin B Maintenance Dose If the Patient Has Renal Impairment?**

### **Recommendation.**

**R17:** We recommend that daily maintenance doses of polymyxin B should not be adjusted if the patient has renal impairment.

**Evidence Summary.:** Polymyxin B is not significantly eliminated by the kidneys, and clinical PK studies demonstrate that polymyxin B clearance does not depend on  $Cl_{cr}$ .<sup>44–47</sup> Therefore, no PK rationale exists for adjusting doses according to renal function. Lowering doses in patients with decreased  $Cl_{cr}$  will lead to lower polymyxin B plasma concentrations. The package insert for polymyxin B recommends dose reducing “downward for individuals with kidney impairment”; however, it is unclear what data spurred this recommendation.<sup>83</sup> More recent PK data as well as enhanced understanding of renal handling of polymyxin B refute this recommendation. If unnecessary renal dose adjustments are made in patients, there is potential for drug underexposure and clinical failure. Clinical literature supports this claim because doses of 1.2 mg/kg/day or less (equivalent to 12,000 IU/kg/day or less), which were commonly prescribed to patients with renal insufficiency, were associated with increased mortality in patients receiving polymyxin B.<sup>80</sup>

**Future Research Needs.:** Package insert dose adjustment for renal impairment should be revised because it is not supported by modern PK data. Furthermore, larger PK studies in patients with renal insufficiency are needed to validate the recommendations provided here.

## **XII. Does Renal Replacement Therapy Have Implications for Selection of Intravenous Polymyxin B Dosage Regimens?**

### **Recommendation.**

**RI8:** We recommend that neither the loading dose nor maintenance dose be adjusted in patients receiving renal replacement therapy.

**Evidence Summary.:** There are only two reports of the PK of polymyxin B in patients receiving renal replacement, and both involved CRRT. The first report involved two patients receiving continuous venovenous hemodialysis (CVVHD),<sup>79</sup> and the second described one patient receiving continuous venovenous hemofiltration (CVVHF).<sup>84</sup> In the former two patients, the CVVHD was responsible for 5.6% and 12.2% of polymyxin B total body clearance.<sup>79</sup> In the latter patient, the polymyxin B extraction across the extracorporeal cartridge was only 5.0%.<sup>84</sup> This degree of elimination is similar to the extent of renal elimination in critically ill patients not receiving extracorporeal modalities (median 4%, range 1.0–17.4%).<sup>44</sup> Although data are limited to these three cases, they suggest that CVVHD and CVVHF are unlikely to remove more than ~12% of total body polymyxin B, similar to the percentage recovered in the urine in patients not requiring renal replacement therapy.<sup>44</sup> Thus on the basis of these PK data, dose modifications are not warranted in patients receiving these forms of CRRT.

Clinical data also suggest that dose reductions in patients on CVVHD can potentially lead to underexposure and increased risk of poor outcomes.<sup>85</sup> Higher total daily doses were associated with lower 30-day mortality in bivariate analysis ( $p=0.04$ ), and a total daily dose of 200 mg or more (equivalent to 2 million IU or more) was associated with a lower risk of 30-day mortality in multivariate analysis ( $p=0.02$ ).<sup>85</sup> Thus dose reductions for patients receiving renal replacement therapy are not only unwarranted based on the limited amount of PK data, but the clinical evidence suggests they might potentially be harmful to patients.

Currently no PK data are available on polymyxin B in patients receiving intermittent renal replacement therapy.

**Future Research Needs.:** PK data are lacking for polymyxin B in patients receiving IHD and SLED, and only minimal data are available for CRRT. Larger PK analyses are urgently needed to further refine dosing recommendations.

### **XIII. Is There a Role for Therapeutic Drug Monitoring of Colistin or Polymyxin B?**

#### **Recommendation.**

**R19:** We recommend that TDM and adaptive feedback control (AFC) be used wherever possible for both colistin and polymyxin B.

**Evidence Summary.:** The polymyxins display characteristics that suggest TDM and AFC would be beneficial. Drug dose cannot be safely optimized using clinical observation and dosing algorithms alone, especially in the early treatment period that is a critical determinant of prognosis. Moreover, if therapy is unsuccessful, potential dire consequences may ensue (clinical ones for the patient concerned in addition to emergence of polymyxin resistance). In addition, based on the more abundant data for colistin, there are established relationships between plasma exposure and both antibacterial effect<sup>29</sup> and risk of AKI.<sup>34–36</sup> The therapeutic window is extremely narrow because plasma exposures required for antibacterial effect overlap those associated with increased AKI risk.<sup>6</sup> And the substantial interpatient variability in PK cannot be accounted for by known patient factors (such variability is substantially greater for IV colistin than polymyxin B).<sup>6, 44, 61</sup>

The use of TDM as an aid to dosing CMS was reported for a small number of patients,<sup>86, 87</sup> but the benefit was not demonstrated in appropriately designed studies.<sup>4</sup> For colistin, it is essential to ensure that sample collection, handling, and analysis are conducted appropriately to minimize ex vivo conversion of CMS to colistin.<sup>33, 59</sup> For colistin, by collecting blood samples just before the next dose (when CMS concentrations are the lowest), the potential for measurement of artificially elevated plasma colistin concentrations is minimized but not eliminated. For polymyxin B sample collection, handling and analysis for TDM are substantially less complicated because this polymyxin is administered directly, not as an inactive prodrug. As stated earlier, using TDM, the target concentration is 2 mg/L for susceptible microorganisms, irrespective of the MIC provided by the routine clinical microbiology laboratory.

**Future Research Needs.:** Real-time PK/PD/TD profiles obtained from patients during polymyxin therapy are needed so that maximally precise, patient-specific PK information can be obtained. Such data would inform evolving dose optimization at the individual patient level.



#### XIV. What Strategies Can Be Used to Decrease the Incidence of Acute Kidney Injury in Patients Receiving Colistin or Polymyxin B Therapy?

##### Recommendation.

**R20:** We recommend, wherever possible, that concomitant nephrotoxic agents should be avoided in patients receiving colistin or polymyxin B (*Strong recommendation, moderate quality evidence*).

**Remark:** This recommendation was initially graded with low confidence because data were observational. However, the evidence quality was upgraded due to the consistent large magnitude of the effect of administration of concomitant nephrotoxins on the incidence of AKI with no important threats to the validity of the data.

**Evidence Summary.:** Undoubtedly, nephrotoxicity is the most clinically relevant and dose-limiting adverse reaction of the polymyxins. The incidence of nephrotoxicity varies widely in the literature from 0% to more than 60% largely due to heterogeneous patient populations, differing definitions of nephrotoxicity, wide ranges of polymyxin doses administered, and differences in both severity of illness and the presence/absence of various other risk factors for the patients being studied.<sup>34, 36, 50, 54, 88–91</sup> Contemporary studies, using commonly accepted polymyxin doses and AKI definitions, place the rate of associated nephrotoxicity in the 20–50% range for both polymyxins.<sup>34, 36, 50, 54, 88–91</sup>

Risk factors vary between studies, but a few common factors are identified throughout the literature. More advanced age was identified as a risk factor in multiple analyses, although the so-called cutoff age for increased risk is inconsistent. Weight, irrespective of dose given, was shown to be a risk factor for nephrotoxicity for both colistin<sup>88</sup> and polymyxin B.<sup>92</sup> Chronic comorbid conditions and the presence of hypoalbuminemia were reported as risk factors for nephrotoxicity.<sup>88, 92</sup> Although these factors can help clinicians identify those patients at highest risk of AKI while receiving polymyxin therapy, they are not modifiable. Clinicians should work to address modifiable risk factors for AKI, and the recommendations represent the panel's view regarding how best to accomplish this.

Receipt of concomitant nephrotoxic agents is a consistent risk factor for AKI in patients receiving polymyxin therapy. Although many nephrotoxins have been identified as potential risk factors, only a few would be considered modifiable. For example, receipt of calcineurin inhibitors, acute administration of loop diuretics, and vasopressors have all been associated with polymyxin-associated nephrotoxicity; however, these exposures often cannot be avoided. Conversely, the use of IV contrast media for diagnostic testing, administering nonsteroidal antiinflammatory drug or angiotensin-converting enzyme inhibitor therapy, and/or receipt of other nephrotoxic antibiotics, most notably vancomycin, should be assessed by clinicians and avoided when possible.<sup>93, 94</sup> Although combination therapy with colistin and vancomycin showed both in vitro synergy<sup>95</sup> and select clinical data suggest a potential clinical benefit of this combination,<sup>93, 94</sup> multiple analyses with both colistin<sup>94, 96</sup> and polymyxin B<sup>89</sup> showed concomitant vancomycin to be an independent predictor of polymyxin-associated AKI; thus this combination should be avoided. In addition, analyses demonstrated rifampin<sup>90</sup> coadministration to increase the risk of nephrotoxicity.

Furthermore, concomitant aminoglycosides were also identified as independent predictors of colistin-associated AKI.<sup>91</sup> Given the emergence and spread of XDR gram-negative bacteria, including carbapenem-resistant Enterobacteriaceae (CRE), we acknowledge that aminoglycosides frequently are often one of the few agents to which these organisms are susceptible, and combination therapy involving aminoglycosides and polymyxins might be an attractive alternative and in some cases might be unavoidable.

**Future Research Needs.:** Data demonstrating the impact of purposeful avoidance of the nephrotoxic agents described earlier on prevention of AKI are lacking. Such data would enhance the quality of the evidence supporting this recommendation. Future research is needed to evaluate the safety and efficacy of polymyxin + aminoglycoside therapy. Timely monitoring of renal function is a critical aspect of detecting AKI for the polymyxins. As such, further research on biomarkers that respond rapidly to renal insult would be highly beneficial for toxicodynamic optimization.

### **Recommendation.**

**R21:** We recommend that doses greater than those listed in this guideline for colistin or polymyxin B be avoided in the absence of TDM (*Best practice recommendation*).

**Remark:** This recommendation was not assessed using GRADE. There is an absence of data testing this strategy. Higher doses have theoretical advantages, but the comparative safety and efficacy of those are unavailable based on the currently available literature. This recommendation prioritizes safety, due to the absence of efficacy data with higher dosing strategies. Furthermore, although dose increase or decrease based on serum concentrations is rational from a PK, PD, and TD standpoint, data are lacking to assess the safety and efficacy of such a strategy.

**Evidence Summary.:** The most important risk factor for polymyxin-associated AKI is the magnitude of polymyxin exposure. Higher CMS doses are consistently identified as a risk factor, with CBA doses higher than 5 mg/kg/day (equivalent to ~152,000 IU/kg/day) consistently posing the highest risk. Similarly, associations were seen with absolute polymyxin B doses of 150, 200,<sup>97</sup> and 250<sup>80</sup> mg/day or higher. Not surprisingly, colistin serum steady-state concentrations were also associated with AKI. Average steady-state concentrations of 1.9–2.3 mg/L were associated with higher degrees of toxicity than lower concentrations,<sup>35</sup> whereas day 3 trough concentrations of 3.33 and 2.42 mg/L or higher were associated with AKI at days 7 and 14, respectively.<sup>34</sup> Importantly, in the latter study, of the 26 patients who had colistin trough values higher than 2.2 mg/L on day 3, 17 (65%) and 22 (85%) had toxicity at days 7 and 14, respectively.<sup>34</sup> These PK/TD studies serve as the basis of the maximal tolerable dose described in earlier recommendations in these guidelines, and we would recommend against giving higher exposures.

**Future Research Needs.:** Studies are needed that weigh the risk-to-benefit ratio of clinical cure of infection with the development of nephrotoxicity. Furthermore, investigation regarding dosing regimens (i.e., once/day, multiple times/day, or continuous infusions) or other novel dosing strategies and their impact on nephrotoxicity should also be undertaken.

## Recommendation.

**R22:** In countries where both agents are available, we recommend preferential use of polymyxin B to limit the rate of polymyxin-associated AKI (*Weak recommendation, low-quality evidence*).

**Remark:** This recommendation started with low-quality evidence given the observational data used to make the recommendation. The confidence for the recommendation could not be significantly upgraded or downgraded based on the evidence. The relative consistency of the findings of the published literature and rate of AKI between the two polymyxins were considerations for upgrading the strength of the quality of the evidence. However, these were counter-balanced by some of the data that did not show a safety advantage with polymyxin B. In addition, comparative studies are also confounded by the different doses of colistin and polymyxin B used in comparing AKI. A strong recommendation cannot be made until adequately powered prospective dose-optimized studies are performed.

**Evidence Summary.:** When polymyxins reemerged in the 1980s, one of the main drivers of preferential use of CMS over polymyxin B was the historical belief, driven by anecdotes rather than evidence, that colistin was the safer option with respect to nephrotoxicity. Modern-day data have debunked this belief, and interestingly there is a suggestion that polymyxin B might in fact be safer, with respect to the kidneys, than colistin. Data from kidney cell lines<sup>98</sup> as well as animals<sup>99</sup> suggest that polymyxin B and colistin, as would be expected from their similar chemical structures, have similar toxic effects on the kidney.

However, in the six currently available clinical studies assessing comparative nephrotoxicity rates between the polymyxins, five displayed at least some suggestion of increased and/or more severe nephrotoxicity with colistin.<sup>48, 49, 52, 54, 98, 100</sup> The one outlier to this trend was limited by small numbers (only 30 and 39 patients receiving polymyxin B and CMS were evaluable for AKI, respectively).<sup>48, 49, 50, 51, 52, 54</sup> A systemic review and meta-analysis<sup>53</sup> summarizes the published studies. Taken together, these data suggest that polymyxin B is associated with less AKI in patients.

Regardless of the mechanism, the current data, although limited in quality, suggest that polymyxin B is less likely to cause nephrotoxicity than CMS. Until further evidence becomes available, clinicians should consider polymyxin B as the preferred alternative to decrease the risk of polymyxin-associated AKI. An exception to this would be for the treatment of urinary tract infections, where CMS/colistin may be the preferred agent.

**Future Research Needs.:** The main areas for prioritization of future research include prospective comparative trials assessing AKI rates with dose-optimized polymyxins, investigation into the mechanisms of potential discordant toxicity rates between the agents, and finally whether dose-optimized polymyxins differ in their rates of nonnephrotoxic adverse reactions, most notably neurotoxicity. In addition, studies comparing neurotoxicities and skin hyperpigmentation (e.g., skin darkness of face, ears, neck, and upper chest and head during therapy<sup>101</sup>) for polymyxin B versus colistin require future studies.

### Recommendation.

**R23:** Until further data become available, we do not recommend the routine use of antioxidants for the primary purpose of reducing polymyxin-associated nephrotoxicity (*Weak recommendation, very low-quality evidence*).

**Remark:** The quality of the evidence was initially low given that both underpowered randomized controlled and observational data were used for the assessment. The data suffered from every potential reason for downgrading the data (risk of bias, inconsistency, indirectness, imprecision, and publication bias) and therefore were rated as very low-quality evidence. The recommendation was weak, given that animal data support a potential protective effect as well as the general lack of risk of patient harm with the administration of antioxidants.

**Evidence Summary.:** Interest in using antioxidants, most notably ascorbic acid, has been increasing as a nephroprotective mechanism in patients receiving polymyxin therapy. This stems from preclinical observations that in polymyxin-induced nephrotoxicity, oxidative stress from reactive oxygen species initiates renal cell apoptosis. Animal models supported this protective role of ascorbic acid by demonstrating that administration can decrease kidney tissue apoptosis and subsequent tubular damage.<sup>102</sup>

Clinical data exploring the impact of ascorbic acid on limiting nephrotoxicity are scarce and have displayed conflicting results. One group<sup>103</sup> assessed nephrotoxicity rates with a novel dosing regimen based on recent PK advances. Interestingly, although not the primary intent of the analysis, both bivariate (30% vs 67%;  $p < 0.05$ ) and multivariate analyses (adjusted odds ratio [aOR] 0.27, 95% CI 0.13–0.57) suggested that concomitant administration of ascorbic acid was protective against nephrotoxicity. Conversely, a small RCT in 28 patients failed to show any benefit of 4 g/day of ascorbic acid on the rates of colistin-associated nephrotoxicity.<sup>104</sup> Therefore, although a promising therapy, the current data are insufficient to warrant a recommendation in favor of routine administration of ascorbic acid or any other antioxidant for the prevention of polymyxin-associated AKI.

**Future Research Needs.:** Adequately powered and sufficiently controlled prospective studies are warranted to assess the impact of ascorbic acid or other antioxidants on the incidence and/or severity of polymyxin-associated nephrotoxicity.

### XV. If My Patient Develops AKI While on Colistin or Polymyxin B, Should I Decrease the Dose?

#### Recommendations.

**R24:** We recommend that if a patient develops AKI while on colistin, the daily dose should be decreased to the appropriate renally adjusted dose for a plasma colistin  $C_{ss,avg}$  of 2 mg/L (Table 2).

**R25:** We recommend that doses should not be decreased, outside of the renal dosing recommendations for colistin, particularly in patients who develop AKI when colistin or polymyxin B is being administered for a life-threatening infection, a deep-seated infection,

or when the infecting pathogen has an MIC higher than 1 mg/L (*Strong recommendation, low-quality evidence*). If the MIC of the infecting pathogen and/or the nature of the infection suggest that targeting a lower plasma concentration may be adequate, consideration should be given to decreasing the dose to target a different  $C_{ss,avg}$  of colistin (*Best practice recommendation*).

**R26:** We recommend that cessation of therapy may be considered in patients who develop AKI if infection diagnosis is uncertain or when an alternative less nephrotoxic drug is available (*Best practice recommendation*).

**Evidence Summary.:** Although clinical PK data support the need for dose adjustment in AKI for colistin, they do not for polymyxin B.<sup>6, 44, 45</sup> It is a reasonable hypothesis that patients who develop AKI have “supratherapeutic” polymyxin plasma concentrations, but evidence from colistin studies suggests considerable overlap between the “therapeutic” and “nephrotoxic” plasma concentrations of polymyxins among patients who develop AKI.<sup>34–36</sup> It is also important to note that AKI may be precipitated by sepsis arising from inadequate treatment of infection.<sup>105</sup>

The rationale for the recommendation not to lower doses of polymyxin B in the setting of a decline in renal function is that lowering doses in these patients will ultimately lower serum concentrations of polymyxin B, and although that might limit toxicity, there is a greater concern that it would compromise therapeutic efficacy as was demonstrated in published studies. For polymyxin B, data suggested that higher doses, even in the setting of AKI, improve outcomes. One retrospective study with 276 patients showed a lower risk of in-hospital mortality (aOR 0.43, 95% CI 0.23–0.79, p=0.007) in patients receiving high-dose polymyxin B (200 mg/day or more) despite the development of moderate or severe renal injury, defined as 100% or more increase in serum creatinine from baseline or need for hemodialysis.<sup>97</sup> In a larger multicenter prospective cohort with 410 patients, a polymyxin B dose of 150 mg/day or higher was associated with a nonsignificant protective effect on 30-day mortality (aHR 0.74, 95% CI 0.51–1.07, p=0.11) in patients who developed AKI according to the RIFLE criteria.<sup>92</sup>

In patients who have less severe infections, are clinically stable, and are receiving combination therapy, or those with infecting organisms with MICs of 1 mg/L or lower, it is reasonable to reduce the dose in the setting of AKI. For such patients receiving colistin, a lower steady-state plasma concentration may be targeted by making proportional adjustment to the daily doses in Table 2 or by using the reported dosing algorithm.<sup>6</sup> Because the process regarding how exactly to achieve this and evidence to support this strategy is lacking, we find it reasonable to modify the dose to target a steady-state concentration of 1.5 mg/L in certain clinical scenarios. A similar strategy can be used for polymyxin B.

For polymyxin B, in similar clinical scenarios as described earlier for colistin, it would be reasonable to decrease the dose to the lower end of the package insert range. The evidence to support this strategy for polymyxin B and colistin is currently lacking, but it is considered appropriate in these settings because the likelihood of achieving only subtherapeutic drug exposure is significantly diminished, and continued declines in renal function might

adversely impact clinical outcomes. Similarly, clinical judgment should be used to decide whether or not to continue polymyxin therapy in patients who develop AKI and have an unconfirmed microbiological infectious etiology. The potential benefit of maintaining treatment should be weighed against the risk of worsening AKI on a case-by-case basis.

**Future Research Needs.:** Although research is emerging regarding the association between exposure of colistin and polymyxin B and toxicity, the precise PK/TD profile has yet to be fully elucidated as it relates to the time frame and onset of nephrotoxicity. Therefore, future research needs to further elucidate these targets. Data pertaining to clear dose modifications in the setting of AKI and the impact it has on the progression and/or resolution of AKI and clinical efficacy are also urgently needed.

### Polymyxin Combinations

Polymyxin combination therapy is a heavily debated and controversial topic. For multiple reasons, combination therapy might be advantageous. First, it is now very clear that plasma concentrations of colistin are suboptimal in a substantial proportion of patients, even when daily doses of CMS are at the upper limit of the approved product label.<sup>6, 33, 59–61</sup> Similarly, plasma polymyxin B concentrations achieved among patients receiving the current upper limit daily dose are not likely to be reliably efficacious in many clinical scenarios including respiratory tract infections.<sup>44</sup> Second, it is not possible to simply increase the daily doses of CMS or polymyxin B beyond doses recommended in this document due to the potential for nephrotoxicity that is the major dose-limiting adverse effect.<sup>8, 52, 90</sup> Third, the emerging body of evidence in preclinical lung infection models suggests poor in vivo response to the polymyxins when administered parenterally.<sup>29, 30</sup> Finally, polymyxin resistance is increasing worldwide with several recent reports of clinical failure and resistance emergence during monotherapy.<sup>106–109</sup> With the recent report of mobile colistin resistance genes,<sup>16–18, 110, 111</sup> the presence of heteroresistance,<sup>19</sup> and the association between colistin resistance and increased risk of in-hospital mortality,<sup>106</sup> there is mounting support for strategies to optimize polymyxins therapeutically including combination therapy.<sup>41, 111–113</sup> There is a mechanism-based rationale for using polymyxins in combination with other antimicrobials that display synergy with a membrane permeabilizer (such as the polymyxins) allowing for increased concentrations of companion antibacterial agents that have intracellular targets.<sup>109, 114–116</sup>

Unfortunately, the clinical literature on combination therapy versus monotherapy is difficult to interpret due to limitations in many studies.<sup>117, 118</sup> The first type of limitation relates to the characteristics of the critically ill patient population that develop infections due to carbapenem-resistant gram-negative bacilli. These are generally complex patients, with preexisting comorbidities who experience extremely high rates of treatment failure and death irrespective of infection-related outcome. Because the primary outcome in many analyses is all-cause mortality, defining the effectiveness of combination versus monotherapy based on this outcome is extremely challenging. In addition, patients requiring polymyxin therapy frequently have significant delays in time to appropriate therapy that may limit the clinical impact of treatment strategies. Furthermore, finding data comparing monotherapy and combination therapy where concomitant antibiotic exposure is minimized is unrealistic because critically ill patients frequently are treated empirically for concomitant infections

with a plethora of various different antimicrobials. Some of these antibiotics, such as vancomycin, that lack individual activity against gram-negative bacteria have displayed synergy with the polymyxins in vitro due to cell wall and membrane perturbations.<sup>94</sup> This leads to a potential scenario where patients in a so-called monotherapy group might not truly have received a monotherapeutic regimen. Another characteristic that makes these analyses difficult to interpret is that different types of carbapenem-resistant organisms are often grouped together. The assumption is that all carbapenem-resistant organisms classified dichotomously according to MIC breakpoints are identical and will respond identically to therapy, regardless of mechanism of resistance and specific MIC value, but it is unlikely that this is the case.

Furthermore, although more recent analyses have begun to examine dose-optimized polymyxin therapy, most publications to date do not describe the dosing of polymyxins or other combination agents, use suboptimal polymyxin doses, and/or do not clearly report renal dosing adjustments or MIC values of the polymyxins and/or other antimicrobials used in combination regimens for the pathogens. This is further complicated by the fact that the vast majority of previous combination studies used colistin, rather than polymyxin B, the latter of which has a more favorable and predictable PK profile. Most analyses are retrospective observational studies that have inherent biases (such as confounding by indication), making it difficult to interpret the results clearly.<sup>117</sup>

Finally, it is very important to consider site of infection in studies. Whereas most of the clinical studies with CRE evaluated bloodstream infection (BSI), most of the studies for carbapenem-resistant *A. baumannii* (CRAB) and carbapenem-resistant *P. aeruginosa* (CRPA) evaluated pneumonia. Polymyxins administered parenterally were shown to be far less effective in murine lung infection models than in thigh infection models.<sup>29, 30</sup> Therefore, although the clinical data, presented below, attempt to provide evidence toward the selection of polymyxin monotherapy versus polymyxin combination therapy, the inclusion of a variety of sites of infection within a given trial makes interpretation challenging because different pharmacologic considerations exist in the treatment of different infection sites.

In this section, we describe the latest of published evidence from clinical studies on polymyxin monotherapy versus combination therapy for the three major target organisms: CRE, CRAB, and CRPA. We assess the evidence regarding combination therapy in two different types of scenarios. The first is when the polymyxin is combined with an agent to which the infecting pathogen is susceptible (R27, R29, and R31). The second is when the polymyxin is combined with an agent to which the pathogen lacks in vitro susceptibility (i.e., a “nonsusceptible” agent) (R28, R30, and R32). We acknowledge the rigorous debate by noting the controversies surrounding polymyxin combination versus monotherapy, often in the absence of RCTs.

Given the controversies regarding monotherapy versus combination therapy for polymyxins, it is important to note the panel did not achieve unanimity on this topic, due to a variety of factors including limitations of published studies, lack of clear clinical evidence, and weighing the potential benefit-to-risk ratio of combination versus monotherapy. Therefore, a

decision was made for authors to vote on the recommendations R27 to R32. Some authors abstained from the vote. Based on these voting results, these guidelines provide the panel's consensus recommendations. In some cases, we labeled recommendations as "best practice recommendations," particularly in scenarios where the recommendations are in contrast to the currently published data and/or lack sufficient RCT evidence and represent the views of most panel members as opposed to quality published studies.

The recommendations voted on and thus serving as guideline recommendations R27 to R32 are *not* meant to serve as guideline recommendations for the optimal treatment of carbapenem-resistant organisms and are not recommending preferential use of polymyxin-based therapy for these organisms. Rather, the recommendations address scenarios where a clinician has already decided to use polymyxin-based therapy and is trying to decide between monotherapy and combination therapy.

## **XVI. Should Monotherapy or Combination Therapy for Polymyxin B or Colistin Be Used to Treat Patients with CRE Infections?**

### **Recommendations.**

**R27:** We recommend that for invasive infections due to CRE, polymyxin B or colistin be used in combination with one or more additional agents to which the pathogen displays a susceptible MIC (*Strong recommendation, very low-quality evidence*; panel vote 14–1 in favor of combination therapy).

**Remark:** The quality of the evidence was initially low given the observational data of the trials supporting combination therapy. The data were downgraded to very low for two major reasons. First, the results favoring combination therapy are inconsistent. Although several studies showed a mortality benefit of combination therapy, others failed to demonstrate this benefit, and more recent evidence suggests that such a benefit might be limited to severely ill patients. Second, although these combination studies included colistin as potential therapy, not all of the combination regimens in these studies were colistin based, making the exact role of polymyxin combination therapy difficult to tease out from other combination regimens.

**R28:** If a second active agent to which the infecting CRE displays a susceptible MIC is unavailable, we recommend that polymyxin B or colistin be used in combination with a second and/or third nonsusceptible agent (e.g., a carbapenem). Preference should be given to a non-susceptible agent with the lowest MIC relative to the respective susceptibility breakpoint (*Best practice recommendation*; panel voted 11–4 in favor of combination therapy).

**Evidence Summary.:** Perhaps the best evidence supporting polymyxin combination therapy comes from a series of retrospective observational studies evaluating outcomes of patients receiving combination or monotherapy for bloodstream infections (BSIs) due to carbapenemase-producing Enterobacteriaceae (largely, although not exclusively, producing *Klebsiella pneumoniae* carbapenemase [KPC]).<sup>119–122</sup> Two important features of these analyses warrant comment. First, combination therapy in each of the studies described in detail here



is defined as agents to which the infecting pathogen is susceptible according to the MIC. Second, although most of the combination regimens included a polymyxin (i.e., colistin), the multivariate models analyzing “combination therapy” also include regimens that did not include a polymyxin, and therefore, in some scenarios, the direct applicability of the findings to the polymyxins remains unclear. It is also important to note that there is no adequately powered published RCT to examine whether therapy with polymyxins (polymyxin B or colistin) administered in combination with another active agent is superior to polymyxin B or colistin monotherapy against CRE infections.

Two initial studies suggested a benefit with combination therapy for CRE BSIs.<sup>119, 120</sup> Although limited by small numbers of patients, both analyses showed dramatic associations between combination therapy and survival (infection-related mortality of 0/20 [0%] vs 7/15 [47%],  $p=0.001$ , and 28-day all-cause mortality of 2/15 [13%] vs 11/19 [57%],  $p=0.01$ , for patients receiving combination therapy vs monotherapy, respectively), and the association of combination regimens with survival remained significant in a multivariate model (OR 0.07, 95% CI 0.009–0.71). A report<sup>123</sup> on 125 patients with BSI due to KPC-producing *K. pneumoniae* furthered these findings because combination therapy with colistin + meropenem + tigecycline was independently associated with survival (OR 0.11, 95% CI 0.02–0.69) when compared with monotherapy. These findings were further supported in an analysis by two larger cohort studies,<sup>121, 124</sup> one from Greece and the other from Italy, including patients with infections caused by carbapenemase-producing Enterobacteriaceae where receipt of monotherapy (compared with combination therapy) was associated with an increased risk of death in the multivariate model. Of note, these two cohort studies pointed to a potential advantage of colistin-meropenem combination therapy when the meropenem MIC was 8 mg/L or less.<sup>121</sup> Interestingly, recent results from the INCREMENT trial<sup>122</sup> that included 437 patients with BSI due to CRE suggest that the true benefit of combination therapy might be limited to patients with a greater severity of illness. In this analysis, combination therapy was associated with lower mortality compared with monotherapy in the high-mortality-score stratum (30 [48%] of 63 vs 64 [62%] of 103; aHR 0.56, 95% CI 0.34–0.91) but not in the low-mortality-score stratum (17 [24%] of 72 vs 21 [20%] of 105; aOR 1.21, 95% CI 0.56–2.56,  $p=0.62$ ). It is important to note that most patients included in the studies just cited had BSI.

Based on the available literature, we recommend that when polymyxins are used for the management of invasive CRE infections, combination therapy including one or more additional agents with in vitro activity against the pathogen should be administered. The rationale for this recommendation is based on the available observational evidence suggesting decreased mortality with combination therapy as well as concerns regarding emergence of polymyxin resistance when monotherapy is used. Of note, none of the previously mentioned studies assessed the impact of combination regimens on the development of polymyxin resistance and were based on older definitions of meropenem susceptibility that have now changed to a breakpoint of 2 mg/L according to EUCAST/CLSI.<sup>12, 15</sup>

There are few studies which have assessed the impact of polymyxin combination therapy with a second nonsusceptible agent on outcomes in patients with invasive CRE infections.

Perhaps the best evidence suggesting a potential advantage of this strategy comes from a recently published RCT comparing colistin monotherapy versus colistin + meropenem combination therapy for the management of carbapenem-resistant gram-negative bacilli.<sup>125</sup> In this study, only nine patients (2%) had isolates susceptible (MICs of 8 mg/L or lower) to meropenem. Both clinical failure and 28-day mortality occurred in a lower proportion of patients with CRE receiving the colistin + meropenem combination than colistin monotherapy (failure rates 18/39 [46%] vs 23/34 [68%];  $p=0.19$ , and 28-day mortality of 21% vs 35%;  $p=0.24$ ), although statistical significance was not demonstrated.<sup>125</sup> Based on the lack of evidence clearly addressing this issue in CRE and the previously mentioned concerns/limitations with monotherapy, we recommend that if no second agents to which the infecting pathogen displays a susceptible MIC are available for combination therapy, a second and/or third “nonsusceptible” agent should be administered in combination with the polymyxin. Given the lack of supporting evidence, this is a best practice recommendation.

**Future Research Needs.:** Currently a second ongoing RCT is comparing colistin monotherapy with colistin + meropenem combination therapy for the management of invasive infections due to carbapenem-resistant gram-negative organisms (<https://clinicaltrials.gov/ct2/show/NCT01597973?term=NCT01597973&rank=1>). Data from this study should further elucidate the role of combinations in the management of CRE. Furthermore, given the potential advantages of polymyxin B over colistin, clinical data assessing the impact of polymyxin B–based combination regimens are needed. Future studies should also address the impact of infection site on the effectiveness of combination therapy.

## **XVII. Should Monotherapy or Combination Therapy for Polymyxin B or Colistin Be Used to Treat Patients with CRAB?**

### **Recommendations.**

**R29:** We recommend that for invasive infections due to CRAB, polymyxin B or colistin should be used in combination with one or more additional agents to which the pathogen displays a susceptible MIC (*Best practice recommendation*; panel voted 10–5 in favor of combination).

**R30:** If a second active agent to which the infecting CRAB displays a susceptible MIC is unavailable, we recommend that polymyxin B or colistin should be used alone as monotherapy (*Weak recommendation, moderate quality evidence*; panel voted 8–7 in favor of monotherapy).

**Remark:** The quality of the evidence for this recommendation began as high based on the previously mentioned RCTs. However, the quality of the evidence was finally graded as moderate due to the open-label nature of the RCTs, the use of nonstudy anti–gram-negative therapies, and relatively low numbers of patients in the rifampin and fosfomycin studies. The strength of the recommendation is weak due to the dichotomy in our panel with regard to optimal management of these patients, potential bias in the studies, lack of AFC to optimize polymyxin concentrations, and dosing concerns in the rifampin trial.

**Evidence Summary.:** Perhaps more than any other organism mentioned in these guidelines, the retrospective CRAB literature surrounding combination therapy versus monotherapy is nearly uninterpretable due to confounding by indication, poorly described dosing, and a lack of clarity regarding the timing of initial administration of therapy (and subsequently time to appropriate therapy) that are incompletely described. In addition, the published literature is often unclear as to whether or not patients had infection versus colonization because the infection site is often described as “respiratory” without a clear explanation of how infection was defined. Also, as previously discussed, given the complexity of study patients and the lack of a true definition of infection, the primary end point of mortality (all-cause, in-hospital, or 30-day) is suboptimal because often many competing mortality risks exist. In these studies, it is not always clear whether death was clearly associated with infection.

Therefore, the studies reviewed in this section are limited to the three major randomized open-label trials that compared colistin with a second nonsusceptible agent including rifampin, fosfomycin, or meropenem.<sup>125–127</sup> Some isolates in the rifampin and fosfomycin studies were defined as having in vitro susceptibility to these agents. However, for the purposes of these guidelines, the panel considered these isolates to be nonsusceptible due to a lack of uniform susceptibility in the isolates included in these studies (not all isolates were defined as susceptible) and a lack of dose optimization strategies used for these agents.<sup>126, 127</sup> Taken together with the insufficient clinical data to support efficacy, concerns for resistance development and the routine avoidance by clinicians for fosfomycin and rifampin monotherapy provide a further rationale as to why both of these agents were considered nonsusceptible. There are currently no prospective randomized trials that study polymyxin combinations involving a second agent to which the infecting pathogen displays a susceptible MIC. Therefore, there are no clinical data assessing combination therapy with a polymyxin and a second in vitro active agent, and thus the best practice recommendation for using this strategy is an extrapolation from the CRE data. The three RCTs compared combination with a nonsusceptible agent versus monotherapy.

The first of the three open-label RCTs comparing combinations with monotherapy was a prospective study<sup>126</sup> that enrolled 210 patients to receive colistin or colistin + rifampin randomly for the treatment of life-threatening XDR *A. baumannii* infections. No colistin loading dose was administered, and the maximum daily maintenance dose was low by current standards. Patients were randomly allocated (1:1) to either colistin alone, 2 million IU every 8 hours IV, or colistin plus rifampin 600 mg every 12 hours IV. The colistin MIC was 0.5 mg/L or lower for all isolates at randomization. This analysis reported that the risk of death within 30 days was similar between combination therapy and monotherapy (OR = 0.88, 95% CI 0.46–1.69, p=0.71) despite a significantly improved microbiological cure rate in patients receiving colistin + rifampin (p=0.034).

Furthermore, no patients developed colistin-resistant isolates in either arm. This improvement in microbiological cure was consistent with another small randomized trial (n=43) that compared colistin and colistin + rifampin, where time to microbiological clearance was reduced in the colistin + rifampin arm (3.1 vs 4.5 days; p=0.029).<sup>128</sup> It is important to note that although rifampin displays potent in vitro synergy with polymyxins, many suboptimal pharmacologic characteristics are associated with the drug. In addition to

drug interaction concerns due to induction of drug metabolism, rifampin is also associated with adverse drug events including hepatotoxicity. A nonsignificantly higher rate of hepatotoxicity in the colistin + rifampin arm was identified in one trial<sup>126</sup> (20.8% in the colistin + rifampin arm vs 11.9% in the colistin arm;  $p=0.13$ ). In fact, 10 patients in the combination therapy arm had rifampin discontinued due to this adverse event. In such an open-label study, in patients receiving “monotherapy,” it is difficult to avoid use of agents that might provide a combinatorial benefit with polymyxins. As an example, ~70% of patients in the monotherapy and combination groups received other antibiotics including agents such as meropenem (prescribed more commonly in the monotherapy than combination therapy arm [15.9% vs 3.9%, respectively]).

In another open-label, prospective, randomized trial of 94 patients with CRAB infections, subjects were randomized to receive colistin alone or colistin + fosfomycin.<sup>127</sup> Some patients in both groups received other antibiotics; for example, 17.0% and 8.5% of patients in the monotherapy and combination groups, respectively, received a carbapenem. No significant differences were observed between monotherapy and combination therapy arms in infection-related (23.1% vs 16.3%;  $p=0.507$ ) or all-cause mortality (57.4% vs 46.8%;  $p=0.41$ ). Interestingly, microbiological cure in the first 72 hours (65.7% vs 78.8%;  $p=0.028$ ) and at the end of treatment (84.5% vs 100%;  $p=0.023$ ) occurred more frequently in the combination arm.

Recently, the largest RCT to date (AIDA study)<sup>125</sup> compared colistin monotherapy with colistin (9 million IU or 300 mg CBA/day) + high-dose extended-infusion meropenem combination therapy for the treatment of carbapenem-resistant gram-negative bacilli. Although this study included CRE and carbapenem-resistant *P. aeruginosa*, 312 of 406 enrolled patients (77%) had CRAB. No significant difference was observed in the rate of clinical failure or 28-day mortality between monotherapy and combination therapy in the entire cohort (156/198 [79%] vs 152/208 [73%];  $p=0.17$  for clinical failure, and 43% vs 45%;  $p=0.78$  for 28-day mortality) or the subset of patients with *A. baumannii* infections (125/151 [83%] vs 81%;  $p=0.64$  for clinical failure, and 46% vs 52%;  $p=0.40$  for mortality). A total of 94% of patients in this study had either bacteremia or pneumonia with nearly an even split between the two. Importantly, there was also no significant difference between groups in the identification of colistin resistance in clinical samples by day 28 (6% for monotherapy vs 5% for combination therapy;  $p=0.77$ ) or microbiological failure (31% for monotherapy vs 35% for combination therapy;  $p=0.49$ ).

In summary, the data comparing monotherapy to combination therapy do not support the addition of that second nonsusceptible agent. Therefore, the evidence-based recommendation is in support of monotherapy. There was significant debate and disagreement among the panel members surrounding this recommendation. Many members of the panel were concerned that even though the clinical evidence does not support combination therapy, the PK/PD limitations of the polymyxins and the development of resistance remain great concerns. The small numbers and large percentage of patients with pneumonia in the rifampin and fosfomycin studies as well as the limitations of the AIDA study (e.g., open label, the large number of patients treated for pneumonia, and low

Sequential Organ Failure Assessment scores) are why many panel members voted for combination therapy. However, the final vote was in favor of monotherapy.

**Future Research Needs.:** An ongoing double-blind RCT will help further shed light on the role of combinations in the management of gram-negative infections including those caused by CRAB. Clinical data assessing the impact of polymyxin B–based combination regimens are needed. Future studies should also address the impact of infection site on the relative effectiveness of combination therapy versus monotherapy.

## XVIII. Should Monotherapy or Combination Therapy for Polymyxin B or Colistin Be Used to Treat Patients with CRPA?

### Recommendations.

**R31:** We recommend that for invasive infections due to CRPA, polymyxin B or colistin should be used in combination with one or more additional agents to which the pathogen displays a susceptible MIC (*Best practice recommendation*; panel voted 14–1 in favor of combination therapy).

**R32:** If a second active agent to which the infecting CRPA displays a susceptible MIC is unavailable, we recommend polymyxin B and colistin be used in combination with a second and/or third nonsusceptible agent (e.g., a carbapenem). Preference should be given to a non-susceptible agent with the lowest MIC relative to the respective susceptibility breakpoint (*Best practice recommendation*; panel voted 11–4 in favor of combination therapy).

**Evidence Summary.:** Very little evidence has assessed the comparative outcomes of polymyxin monotherapy and combination therapy for MDR/XDR *P. aeruginosa* infections. The primary short-coming of the available literature is that all of the analyses are retrospective and observational, and when analyzed, *P. aeruginosa* is often lumped together with other carbapenem-resistant pathogens. Therefore, many of the studies are difficult to interpret with regard to the independent impact of polymyxin combination therapy on *P. aeruginosa* infection. This section only includes those analyses that specifically focused on outcomes in *P. aeruginosa* infections.

In a single-center retrospective study of 74 patients with HAP caused by MDR *P. aeruginosa* who were treated with polymyxin B, there was no statistically significant difference in clinical cure rates between patients receiving polymyxin B plus another agent (mainly imipenem) and patients receiving polymyxin B monotherapy (14/28 [50%] vs 21/46 [46%];  $p=0.71$ ).<sup>129</sup> In an additional retrospective single-center study of 258 patients with documented infections (mainly pneumonia) due to MDR gram-negative organisms, 68 (26.4%) of which were caused by MDR *P. aeruginosa*, rates of clinical cure in patients with *P. aeruginosa* infection who received colistin monotherapy, colistin + meropenem, colistin + piperacillin/tazobactam, colistin + ampicillin/sulbactam, and colistin + other agents were 75.0% (9/12), 85.7% (24/28), 60% (6/10), 100% (1/1), and 64.7% (11/17), respectively.<sup>130</sup> In a retrospective multicenter study, among 89 cancer patients with *P. aeruginosa* infection (mainly bacteremia), only 15 were treated with colistin (17%). Mortality occurred in 3 of 8 patients (37.5%) treated with colistin monotherapy and 4 of 7 (57.1%) receiving colistin plus

another agent, mostly a  $\beta$ -lactam ( $p=0.8$ ).<sup>131</sup> In a multicenter retrospective study,<sup>132</sup> the authors compared polymyxin B plus other agents with polymyxin B monotherapy for treating infections caused by *A. baumannii* and *P. aeruginosa* (mainly respiratory infections) in 101 critically ill patients. Most infections were caused by *A. baumannii* (83 [82.2%]), and only 18 (17.8%) were due to *P. aeruginosa*. Overall, 3 of 18 patients with *P. aeruginosa* infections received combination therapy and all survived; 14 of 15 patients treated with polymyxin B monotherapy died within 30 days ( $p=0.005$ ).<sup>132</sup>

The results of a single-center retrospective cohort of 34 patients with osteoarticular infections due to MDR *P. aeruginosa* were reported; 15 (44.1%) had prosthetic joint infections and 19 (55.9%) osteoarthritis. Patients were treated with IV antibiotics for 6 weeks.<sup>133</sup> Combination therapy (mainly colistin plus a  $\beta$ -lactam) was associated with higher cure rates than monotherapy with colistin or a  $\beta$ -lactam (11/15 [73.3%] vs 6/19 [31.6%], respectively;  $p=0.016$ ).<sup>133</sup> Finally, a single-center prospective study<sup>134</sup> was conducted on 91 patients with infections (most commonly pneumonia, followed by urinary tract infection) caused by colistin-susceptible *P. aeruginosa* who were treated with colistin. No association was detected between receipt of monotherapy or combination therapy and either clinical failure or mortality.<sup>134</sup>

The small numbers, discordant results, retrospective nature of most studies, and inconsistencies regarding other agents being included in combination regimens preclude any definitive conclusion with regard to polymyxin combination therapy versus monotherapy for *P. aeruginosa*. Until further evidence becomes available, the panel recommends that when polymyxins are used to treat invasive infections caused by *P. aeruginosa* that they be used in combination with one or more additional agents to which the pathogen displays a susceptible MIC. The rationale for this recommendation is based on extrapolation of the available evidence for CRE and the potential risk of clinical failure or emergence of resistance when monotherapy is used. If no active agents are available, additional nonsusceptible agents should be administered based on MIC value. Preference should be given to nonsusceptible agents to which the pathogen demonstrates the lowest MIC respective to the breakpoint.

**Future Research Needs.:** Additional data, even observational in nature, assessing outcomes of polymyxin monotherapy versus combination therapy for MDR/XDR *P. aeruginosa* are needed. Care should be taken by investigators to clearly describe polymyxin dosing, other antimicrobials administered, and the degree of susceptibility of the pathogen to the agents included in the treatment regimens for a given isolate. Future studies should also address the impact of infection site on the relative effectiveness of combination therapy.

## **XIX. Should Inhaled Polymyxins Be Administered to Patients with HAP/VAP, and If So, Which Agent Is Preferred?**

### **Recommendations.**

**R33:** We recommend that patients requiring IV polymyxin therapy for suspected or documented XDR gram-negative HAP or VAP should receive adjunctive polymyxin aerosol therapy (*Weak recommendation, low-quality evidence*).

**R34:** We recommend that for polymyxin aerosol therapy, either colistin or polymyxin B is appropriate (*Weak recommendation; very low-quality evidence*).

**Evidence Summary.:** An RCT was performed comparing empirical CMS aerosol with placebo aerosol.<sup>135</sup> Patients were randomized to receive either 4 mL nebulized sterile normal saline or CMS equivalent to 75 mg colistin base activity reconstituted in 4 mL nebulized sterile normal saline that was delivered immediately via a jet or ultrasonic nebulizer for 10 minutes or until the nebulized solution container was empty.<sup>135</sup> The regimen and duration of the systemic antibiotic (s) were chosen by the patient's responsible physician. No benefit in clinical cure or mortality with adjunctive aerosol CMS was demonstrated in this trial.<sup>135</sup> A second RCT has been performed in 149 critically ill adults who developed gram-negative VAP.<sup>136</sup> Patients were randomized to receive 4 million IU of aerosolized CMS by nebulisation for 30 minutes three times/day in addition to IV imipenem 1 g three times/day compared to IV CMS given as a loading dose of 9 million IU followed by 4.5 million IU two times/24 hours in addition to IV imipenem 1 g three times/day. The clinical cure rate was 67.1 % in aerosol group and 72% in IV group (p=0.59). When administered in monotherapy or in combination, the aerosol CMS was as effective as IV regimen. Patients in aerosol group had significantly lower incidence of acute renal failure.<sup>136</sup> In contrast, a 2015 meta-analysis found that clinical response was improved (OR 1.57, 95% CI 1.14–2.15) and mortality was lower (OR 0.89, 95% CI 0.51–1.01) with adjunctive aerosol CMS. All analyses were imprecise and demonstrated inconsistency except for microbiological eradication.<sup>137</sup> Since this meta-analysis, only one retrospective cohort study in pediatric patients has been published that found essentially the same results for clinical response.<sup>138</sup>

Most of the studies included in the meta-analysis focused on MDR pathogens, mainly *Pseudomonas*, *Acinetobacter*, and CRE.<sup>137</sup> Most had carbapenem-resistant or colistin-only susceptible isolates. In many cases, polymyxin aerosols were only added after culture results were known. As such, early effective empirical antibiotic therapy, critical for good outcomes in HAP/VAP, may have been inadequate even in those receiving polymyxin aerosols.

The assumption is that IV colistin may be considered in the patients with pneumonia due to XDR pathogens. Poor results with lower dose IV therapy and higher nephrotoxicity with high-dose therapy,<sup>139</sup> safety concerns when combination therapy includes other nephrotoxic agents, and poor response to polymyxins in preclinical lung infection murine models all warrant consideration of polymyxin aerosols as an adjunctive therapy to IV polymyxins. Use of aerosolized CMS, mainly monotherapy without any IV therapy, for all XDR *Pseudomonas* and *Acinetobacter* VAPs, had equivalent results to IV therapy of less resistant strains.<sup>140</sup> An increase in nephrotoxicity is difficult to detect in the meta-analysis<sup>137</sup> because all studies used IV colistin in addition to aerosol and used various doses of IV colistin, but overall nephrotoxicity rates were high in most studies. More recent studies including a recent meta-analysis of inhaled CMS and a retrospective review comparing 95 critically ill surgical patients who were diagnosed with *A. baumannii* VAP support the notion that nebulized CMS may have less nephrotoxicity and provide similar clinical results, compared to IV CMS.<sup>141</sup> Interestingly, in a systemic review the combination of inhaled CMS + IV CMS was significantly superior to IV CMS for patient survival and clinical cure for the

treatment of resistant *A. baumannii* pneumonia in critically ill patients.<sup>142, 143</sup> These recommendations place high value on pharmacologic considerations.

The overwhelming number of case-control studies, significant clinical experience, and the RCTs<sup>135, 136</sup> have utilized CMS. No direct comparison of CMS and polymyxin B has been performed and appear to have equivalent adverse events, mainly bronchospasm. It is important to note that it has been suggested that aerosolized CMS therapy may have a substantial targeting advantage over IV therapy. Very high concentrations of formed colistin in ELF have been reported after aerosolized CMS delivery in critically ill patients. ELF colistin concentrations from CMS aerosol delivery (9.53 to 1137 mg/L) have been shown to be much higher than those in plasma (0.15 to 0.73 mg/L) after intravenous administration of CMS.<sup>146</sup> Typically, 9% of the CMS dose reaches the alveolar level.<sup>146</sup> Colistin levels achieved in alveolar fluid at the end of an 8-hour interval may be below the MIC of MDR pathogens, raising the possibility of failure.<sup>147</sup> However, colistin was shown to bind to secretory mucin in sputum or epithelial mucin that lines airways that may reduce the antibacterial efficacy of inhaled or IV administered colistin.<sup>148</sup> Furthermore, a major concern is the actual aerosol delivery.<sup>149</sup> Experimental studies demonstrated significant variation in the amount of drug deposited at the alveolar level in mechanically ventilated patients.<sup>150</sup> A survey found that 30% of intensivists in Europe and France have used aerosolized antibiotics at least every other month.<sup>151</sup> However, most did not vary ventilator settings to optimize delivery of the antibiotic to the alveolar level. Therefore, optimizing ventilator settings and aerosol generator capabilities likely played a much greater role in clinical response in studies in which polymyxin was used.

It is important to acknowledge that ESCMID has recently published a position paper on the use of nebulized antibiotics in invasively mechanically ventilated adults.<sup>152</sup> Overall, ESCMID's position paper recommends avoiding the use of nebulized antibiotics in clinical practice due to a weak level of evidence regarding their efficacy and the high potential for underestimated risks of adverse events (particularly, respiratory complications). Specifically, ESCMID's position paper suggests avoiding the routine addition of nebulized antibiotics such as colistin, to conventional IV antibiotic therapy that already includes IV colistin for the treatment of VAP caused by resistant pathogens.

However, the authors of the polymyxin guidelines elected to place higher value on the potential benefit of adjunctive polymyxin aerosol therapy that outweighed potential risks. These include placing higher value on data from randomized controlled trials in patients with lower respiratory tract infections due to XDR gramnegative bacilli who have been treated with intravenous polymyxin therapy and have experienced high rates of mortality and clinical failure. Preclinical evidence suggests negligible antimicrobial effect of systemic polymyxins (unlike aminoglycosides) in lung infection models although additional clinical studies are urgently needed to better evaluate this. Therefore, although there are safety concerns and poor quality of the evidence with regards to polymyxin aerosol therapy, the guidelines committee believes that the potential benefits outweighs the risks, which is the basis for the recommendation.



**Future Research Needs.:** Prospective clinical trials evaluating adjunctive polymyxin aerosol therapy in addition to IV therapy are necessary. Pharmacokinetic and PK/PD studies in lung infection using aerosol therapy, adjunctive aerosol therapy in combination with IV polymyxin therapy, and adjunctive aerosol therapy in combination with IV polymyxin together with other IV active antibiotics therapy are necessary. Comparative studies between aerosol polymyxin B and colistin are also needed.

### Intrathecal and Intraventricular Administration of Polymyxins

#### XX. Should Intraventricular or Intrathecal Administration of Polymyxins Be Considered in Meningitis or Ventriculitis?

##### Recommendations.

**R35:** Intraventricular (IVT) or intrathecal (ITH) administration of polymyxins at a dosage of 125,000 IU CMS (~4.1 mg CBA) or 5 mg (50,000 IU) polymyxin B per day with concomitant IV polymyxin is recommended for ventriculitis or meningitis caused by MDR and XDR gram-negative pathogens.

**R36:** Due to limited experience with polymyxin B, CMS is the preferred polymyxin for intraventricular or intrathecal administration.

**Evidence Summary.:** Health care–associated ventriculitis and meningitis are an evolving occurrence due to the increasing rates of neurosurgery procedures. The most prevalent pathogens are staphylococci and MDR and XDR gram negatives (*A. baumannii*, *P. aeruginosa*, and *K. pneumoniae*) depending on local epidemiology data.<sup>153, 154</sup> Therapeutic treatment has become increasingly challenging due to the increasing emergence of MDR, and in some cases, colistin or polymyxin B is the only available antimicrobial agent active against meningitis pathogens.<sup>155</sup> Colistin exhibits limited penetration into the cerebrospinal fluid (CSF), with only 5% of serum colistin levels detected in the CSF after IV administration.<sup>156</sup> In the presence of meningitis, an increase in CSF colistin concentrations (34–67% of serum colistin levels) was reported after IV administration, although CSF colistin levels of only 0.5 mg/L were reported in the setting of meningitis, suggesting potentially subtherapeutic colistin CSF concentrations following IV colistin administration.<sup>157</sup> However, IVT administration of colistin in nine neurosurgery patients with XDR gram-negative infections achieved an estimated average steady-state concentrations of colistin in the CSF ranging from 3.0–12.2 mg/L; in the eight patients who were administered CMS IVT at a dosage of 60,000–125,000 IU (this relates to 1.8–4.1 mg CBA) per day, trough CSF levels were between 2.0 and 9.7 mg/L.<sup>158</sup> Thus the measured CSF concentrations in these patients were continuously above the colistin MIC breakpoint of 2 mg/L, and clearance of colistin in the CSF depended on the amount of CSF drained. It is clear that administration of CMS directly into the CSF achieves concentrations of colistin that could not be safely obtained with IV administration alone. Information is lacking on the CSF pharmacokinetics of polymyxin B.

Superiority of combined treatment with IV and IVT colistin treatment with greater potential for eradication of gram-negative bacilli from CSF was documented with no evidence of drug accumulation over time.<sup>159</sup> Intraventricular polymyxin dose is diluted with 3–4 mL of sterile

normal saline and given after removal of an equal volume of CSF. After polymyxin administration, the ventricular drainage is flushed with 2 mL saline solution to minimize the dose remaining in the drainage and given through an external ventricular drain that is clamped for 1 hour. Intrathecal polymyxin is administered through a lumbar drain.<sup>160</sup> The recommended daily dose by the European Medicines Agency and Infectious Diseases Society of America (IDSA) for IVT/ITH colistin is 125,000 IU (~4.1 mg CBA),<sup>57, 153</sup> whereas for polymyxin B, 50,000 IU for adults is recommended by the IDSA.<sup>153</sup>

A systematic review of the evidence regarding clinical efficacy and safety of IVT or ITH colistin or polymyxin B was conducted.<sup>159–176</sup> A total of 234 cases of gram-negative health care–associated ventriculitis or meningitis treated with IVT or ITH colistin or polymyxin B were reported. Intraventricular or ITH colistin was administered in 87% of cases and polymyxin B in the remaining 13%. In most cases (90%), IVT/ITH polymyxins were administered once/day. Monotherapy with IVT/ITH polymyxins was given in 24 cases, whereas in the remaining cases, a variety of parenteral antimicrobials (including polymyxins) was also administered. The median dose of CMS administered through the IVT or ITH route was 125,000 IU (~4.1 mg CBA) per day, whereas for polymyxin B, it was 50,000 IU (5 mg) per day with a mean duration of 18 days. Antimicrobial therapy was administered via a ventricular drain in cases of ventriculitis and clamped for 60 minutes. Successful outcomes were reported in 85% of cases: 144 of 167 cases (86%) caused by *A. baumannii*, 39 of 46 (85%) caused by *P. aeruginosa*, and 17 of 21 (81%) caused by *K. pneumoniae*. Toxicity was noted in 16 cases (7%), mostly presenting as chemical ventriculitis or meningitis in two and nine cases, respectively. Seizures were reported in three cases, numbness of extremities in two cases, and cauda equina syndrome in one.<sup>159–161</sup>

**Future Research Needs.:** Any additional data, even observational in nature, assessing polymyxin IVT and ITH administration are urgently needed to improve the recommendations in this section.

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## References

1. Li J, Nation RL, Turnidge JD, et al. Colistin: the re-emerging antibiotic for multidrug-resistant gram-negative bacterial infections. *Lancet Infect Dis* 2006;9:589–601.

2. Lim LM, Ly N, Anderson D, et al. Resurgence of colistin: a review of resistance, toxicity, pharmacodynamics, and dosing. *Pharmacotherapy* 2010;12:1279–91.
3. Nation RL, Li J, Turnidge JD. The urgent need for clear and accurate information on the polymyxins. *Clin Infect Dis* 2013;11:1656–7.
4. Nation RL, Li J, Cars O, et al. Framework for optimisation of the clinical use of colistin and polymyxin B: the Prato polymyxin consensus. *Lancet Infect Dis* 2015;2:225–34.
5. Onufrak NJ, Rao GG, Forrest A, et al. Critical need for clarity in polymyxin B dosing. *Antimicrob Agents Chemother* 2017;61(5). pii:e00208–17. 10.1128/AAC.00208-17. [PubMed: 28438796]
6. Nation RL, Garonzik SM, Thamlikitkul V, et al. Dosing guidance for intravenous colistin in critically ill patients. *Clin Infect Dis* 2017;5:565–71.
7. Pogue JM, Ortwine JK, Kaye KS. Optimal usage of colistin: are we any closer? *Clin Infect Dis* 2015;12:1778–80.
8. Zavascki AP, Nation RL. Nephrotoxicity of polymyxins: is there any difference between colistimethate and polymyxin B? *Antimicrob Agents Chemother* 2017;61(3). pii:e02319–16. 10.1128/AAC.02319-16. [PubMed: 27993859]
9. Guyatt GH, Oxman AD, Vist GE, et al. GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. *BMJ* 2008;7650:924–6.
10. ISO. Clinical laboratory testing and in vitro diagnostic test systems—susceptibility testing of infectious agents and evaluation of performance of antimicrobial susceptibility test devices—Part 1: Reference method for testing the in vitro activity of antimicrobial agents against rapidly growing aerobic bacteria involved in infectious diseases. ISO 20776–1:2006
11. EUCAST. Recommendations for MIC determination of colistin (polymyxin E) as recommended by the joint CLSI-EUCAST Polymyxin Breakpoints Working Group. EUCAST, 2016 Available from [http://www.eucast.org/ast\\_of\\_bacteria/guidance\\_documents/](http://www.eucast.org/ast_of_bacteria/guidance_documents/). Accessed January 24, 2019.
12. CLSI. Available from <https://clsi.org/media/1700/clsi-newswinter-2016.pdf>. Accessed January 24, 2019.
13. Hindler JA, Humphries RM. Colistin MIC variability by method for contemporary clinical isolates of multidrug-resistant gram-negative bacilli. *J Clin Microbiol.* 2013 6;51 (6):1678–84. 10.1128/JCM.03385-12.14. [PubMed: 23486719]
14. CLSI. M100-S27. Performance Standards for Antimicrobial Susceptibility Testing: 26th Informational Supplement. Wayne, PA: CLSI; 2017.
15. EUCAST. European Committee on Antimicrobial Susceptibility Testing breakpoint tables for interpretation of MICs and zone diameters. Version 7.1. 2017 Available from [http://www.eucast.org/clinical\\_breakpoints/](http://www.eucast.org/clinical_breakpoints/). Accessed January 24, 2019.
16. Liu YY, Wang Y, Walsh TR, et al. Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. *Lancet Infect Dis* 2016;2:161–8.
17. Mediavilla JR, Patrawalla A, Chen L, et al. Colistin- and carbapenem-resistant *Escherichia coli* harboring mcr-1 and blaNDM-5, causing a complicated urinary tract infection in a patient from the United States. *MBio* 2016;7(4). pii:e01191–16. 10.1128/mBio.01191-16. [PubMed: 27578755]
18. McGann P, Snestrud E, Maybank R, et al. *Escherichia coli* harboring mcr-1 and blaCTX-M on a novel IncF plasmid: first report of mcr-1 in the United States. *Antimicrob Agents Chemother* 2016;7:4420–1.
19. Li J, Rayner CR, Nation RL, et al. Heteroresistance to colistin in multidrug-resistant *Acinetobacter baumannii*. *Antimicrob Agents Chemother* 2006;9:2946–50.
20. Tsuji BT, Landersdorfer CB, Lenhard JR, et al. Paradoxical effect of polymyxin B: high drug exposure amplifies resistance in *Acinetobacter baumannii*. *Antimicrob Agents Chemother* 2016;7:3913–20.
21. Tam VH, Schilling AN, Vo G, et al. Pharmacodynamics of polymyxin B against *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother* 2005;9:3624–30.
22. Bergen PJ, Li J, Nation RL, Turnidge JD, Coulthard K, Milne RW. Comparison of once-, twice- and thrice-daily dosing of colistin on antibacterial effect and emergence of resistance: studies with *Pseudomonas aeruginosa* in an in vitro pharmacodynamic model. *J Antimicrob Chemother* 2008;3: 636–42.

23. Ly NS, Yang J, Bulitta JB, Tsuji BT. Impact of two-component regulatory systems PhoP-PhoQ and PmrA-PmrB on colistin pharmacodynamics in *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother* 2012;6:3453–6.
24. Bulman ZP, Satlin MJ, Chen L, et al. New polymyxin B dosing strategies to fortify old allies in the war against KPC-2-producing *Klebsiella pneumoniae*. *Antimicrob Agents Chemother* 2017;61(4). pii: e02023–16. 10.1128/AAC.02023-16. [PubMed: 28167549]
25. Deris ZZ, Yu HH, Davis K, et al. The combination of colistin and doripenem is synergistic against *Klebsiella pneumoniae* at multiple inocula and suppresses colistin resistance in an in vitro pharmacokinetic/pharmacodynamic model. *Antimicrob Agents Chemother* 2012;10:5103–12.
26. Bergen PJ, Bulitta JB, Forrest A, Tsuji BT, Li J, Nation RL. Pharmacokinetic/pharmacodynamic investigation of colistin against *Pseudomonas aeruginosa* using an in vitro model. *Antimicrob Agents Chemother* 2010;9:3783–9.
27. Khan DD, Friberg LE, Nielsen EI. A pharmacokinetic-pharmacodynamic (PKPD) model based on in vitro time-kill data predicts the in vivo PK/PD index of colistin. *J Antimicrob Chemother* 2016;7:1881–4.
28. Hengzhuang W, Wu H, Ciofu O, Song Z, Høiby N. In vivo pharmacokinetics/pharmacodynamics of colistin and imipenem in *Pseudomonas aeruginosa* biofilm infection. *Antimicrob Agents Chemother*. 2012;56(5):2683–90. 10.1128/AAC.06486-11. [PubMed: 22354300]
29. Cheah SE, Wang J, Nguyen VT, Turnidge JD, Li J, Nation RL. New pharmacokinetic/pharmacodynamic studies of systemically administered colistin against *Pseudomonas aeruginosa* and *Acinetobacter baumannii* in mouse thigh and lung infection models: smaller response in lung infection. *J Antimicrob Chemother* 2015;12:3291–7.
30. Landersdorfer CB, Wang J, Wirth V, et al. Pharmacokinetics/pharmacodynamics of systemically administered polymyxin B against *Klebsiella pneumoniae* in mouse thigh and lung infection models. *J Antimicrob Chemother* 2017;73 (2):462–468. 10.1093/jac/dkx409.
31. Sader HS, Rhomberg PR, Farrell DJ, Jones RN. Differences in potency and categorical agreement between colistin and polymyxin B when testing 15,377 clinical strains collected worldwide. *Diagn Microbiol Infect Dis* 2015;4:379–81.
32. Nation RL, Garonzik SM, Li J, et al. Updated US and European dose recommendations for intravenous colistin: how do they perform? *Clin Infect Dis* 2016;5:552–8.
33. Garonzik SM, Li J, Thamlikitkul V, et al. Population pharmacokinetics of colistin methanesulfonate and formed colistin in critically ill patients from a multicenter study provide dosing suggestions for various categories of patients. *Antimicrob Agents Chemother* 2011;7:3284–94.
34. Sorlí L, Luque S, Grau S, et al. Trough colistin plasma level is an independent risk factor for nephrotoxicity: a prospective observational cohort study. *BMC Infect Dis* 2013;13:380. [PubMed: 23957376]
35. Forrest A, Garonzik SM, Thamlikitkul V, et al. Pharmacokinetic/toxicodynamic analysis of colistin-associated acute kidney injury in critically ill patients. *Antimicrob Agents Chemother* 2017;61(11). pii:e01367–17. 10.1128/AAC.01367-17. [PubMed: 28893780]
36. Horcajada JP, Sorlí L, Luque S, et al. Validation of a colistin plasma concentration breakpoint as a predictor of nephrotoxicity in patients treated with colistin methanesulfonate. *Int J Antimicrob Agents* 2016;6:725–7.
37. Mouton JW, Muller AE, Canton R, Giske CG, Kahlmeter G, Turnidge J. MIC-based dose adjustment: facts and fables. *J Antimicrob Chemother* 2018;73(3):564–568. 10.1093/jac/dkx427. [PubMed: 29216348]
38. Forrest A, Silveira FP, Thamlikitkul V, et al. Toxicodynamics for colistin-associated changes in creatinine clearance. *Interscience Conference on Antimicrobial Agents and Chemotherapy* 2014, Washington, DC, 2014.
39. Lakota EA, Landersdorfer CB, Nation RL, et al. Personalizing polymyxin B dosing using an adaptive feedback control algorithm. *Antimicrob Agents Chemother* 2018;62(7):e00483–18. [PubMed: 29760144]

40. Bulitta JB, Yang JC, Yohonn L, et al. Attenuation of colistin bactericidal activity by high inoculum of *Pseudomonas aeruginosa* characterized by a new mechanism-based population pharmacodynamic model. *Antimicrob Agents Chemother* 2010;5:2051–62.
41. Ly NS, Bulman ZP, Bulitta JB, et al. Optimization of polymyxin B in combination with doripenem to combat mutator *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother* 2016;5:2870–80.
42. Kwa A, Kasiakou SK, Tam VH, Falagas ME. Polymyxin B: similarities to and differences from colistin (polymyxin E). *Expert Rev Anti Infect Ther* 2007;5:811–21. [PubMed: 17914915]
43. Nation RL, Velkov T, Li J. Colistin and polymyxin B: peas in a pod, or chalk and cheese? *Clin Infect Dis* 2014;59:88–94. [PubMed: 24700659]
44. Sandri AM, Landersdorfer CB, Jacob J, et al. Population pharmacokinetics of intravenous polymyxin B in critically ill patients: implications for selection of dosage regimens. *Clin Infect Dis* 2013;57(4):524–31. [PubMed: 23697744]
45. Thamlikitkul V, Dubrovskaya Y, Manchandani P, et al. Dosing and pharmacokinetics of polymyxin B in patients with renal insufficiency. *Antimicrob Agents Chemother* 2017;1(1). pii:e01337–16. 10.1128/AAC.01337-16.
46. Zavascki AP, Goldani LZ, Cao G, et al. Pharmacokinetics of intravenous polymyxin B in critically ill patients. *Clin Infect Dis* 2008;10:1298–304.
47. Kwa AL, Abdelraouf K, Low JG, Tam VH. Pharmacokinetics of polymyxin B in a patient with renal insufficiency: a case report. *Clin Infect Dis* 2011;10:1280–1.
48. Oliveira MS, Prado GV, Costa SF, Grinbaum RS, Levin AS. Polymyxin B and colistimethate are comparable as to efficacy and renal toxicity. *Diagn Microbiol Infect Dis* 2009;4:431–4.
49. Akajagbor DS, Wilson SL, Shere-Wolfe KD, Dakum P, Charurat ME, Gilliam BL. Higher incidence of acute kidney injury with intravenous colistimethate sodium compared with polymyxin B in critically ill patients at a tertiary care medical center. *Clin Infect Dis* 2013;9:1300–3.
50. Phe K, Lee Y, McDanel PM, et al. In vitro assessment and multicenter cohort study of comparative nephrotoxicity rates associated with colistimethate versus polymyxin B therapy. *Antimicrob Agents Chemother* 2014;5:2740–6.
51. Tuon FF, Rigatto MH, Lopes CK, Kamei LK, Rocha JL, Zavascki AP. Risk factors for acute kidney injury in patients treated with polymyxin B or colistin methanesulfonate sodium. *Int J Antimicrob Agents* 2014;4:349–52.
52. Rigatto MH, Oliveira MS, Perdigo-Neto LV, et al. Multicenter prospective cohort study of renal failure in patients treated with colistin versus polymyxin B. *Antimicrob Agents Chemother* 2016;4:2443–9.
53. Vardakas KZ, Falagas ME. Colistin versus polymyxin B for the treatment of patients with multidrug-resistant gram-negative infections: a systematic review and meta-analysis. *Int J Antimicrob Agents* 2017;2:233–8.
54. Crass RL, Rutter WC, Burgess DR, Martin CA, Burgess DS. Nephrotoxicity in patients with or without cystic fibrosis treated with polymyxin B compared to colistin. *Antimicrob Agents Chemother* 2017;61(4). pii:e02329–16. 10.1128/AAC.02329-16. [PubMed: 28167560]
55. Couet W, Gregoire N, Gobin P, et al. Pharmacokinetics of colistin and colistimethate sodium after a single 80-mg intravenous dose of CMS in young healthy volunteers. *Clin Pharmacol Ther* 2011;6:875–9.
56. Luque S, Escano C, Sorlí L, et al. Urinary concentrations of colistimethate and formed colistin after intravenous administration in patients with multidrug-resistant Gram-negative bacterial infections. *Antimicrob Agents Chemother* 2017;61 (8). pii:e02595–16. 10.1128/AAC.02595-16. [PubMed: 28559275]
57. European-Medicines-Agency. Assessment report on polymyxin-based products. Referral under Article 31 of Directive 2001/83/EC. Available from [https://www.ema.europa.eu/documents/referral/polymyxin-article-31-referral-assessment-report\\_en.pdf](https://www.ema.europa.eu/documents/referral/polymyxin-article-31-referral-assessment-report_en.pdf). Accessed January 24, 2019.
58. Nation RL, Li J, Cars O, et al. Consistent global approach on reporting of colistin doses to promote safe and effective use. *Clin Infect Dis* 2014;1:139–41.
59. Plachouras D, Karvanen M, Friberg LE, et al. Population pharmacokinetic analysis of colistin methanesulfonate and colistin after intravenous administration in critically ill patients with infections caused by gram-negative bacteria. *Antimicrob Agents Chemother* 2009;8:3430–6.

60. Mohamed AF, Karaiskos I, Plachouras D, et al. Application of a loading dose of colistin methanesulfonate in critically ill patients: population pharmacokinetics, protein binding, and prediction of bacterial kill. *Antimicrob Agents Chemother* 2012;8:4241–9.
61. Karaiskos I, Friberg LE, Pontikis K, et al. Colistin population pharmacokinetics after application of a loading dose of 9 MU colistin methanesulfonate in critically ill patients. *Antimicrob Agents Chemother* 2015;12:7240–8.
62. Gregoire N, Mimoz O, Megarbane B, et al. New colistin population pharmacokinetic data in critically ill patients suggesting an alternative loading dose rationale. *Antimicrob Agents Chemother* 2014;58(12):7324–30. 10.1128/AAC.03508-14. [PubMed: 25267662]
63. He H, Li JC, Nation RL, et al. Pharmacokinetics of four different brands of colistimethate and formed colistin in rats. *J Antimicrob Chemother* 2013;10:2311–7.
64. Shields RK, Anand R, Clarke LG, et al. Defining the incidence and risk factors of colistin-induced acute kidney injury by KDIGO criteria. *PLoS One* 2017;3:e0173286.
65. Kumar A, Roberts D, Wood KE, et al. Duration of hypotension before initiation of effective antimicrobial therapy is the critical determinant of survival in human septic shock. *Crit Care Med* 2006;6:1589–96.
66. Luna CM, Aruj P, Niederman MS, et al. Appropriateness and delay to initiate therapy in ventilator-associated pneumonia. *Eur Respir J* 2006;1:158–64.
67. Kumar A, Ellis P, Arabi Y, et al. Initiation of inappropriate antimicrobial therapy results in a fivefold reduction of survival in human septic shock. *Chest* 2009;5:1237–48.
68. EUCAST. European Committee on Antimicrobial Susceptibility Testing. MIC distributions and ECOFFs. Available from [http://www.eucast.org/mic\\_distributions\\_and\\_ecoffs/](http://www.eucast.org/mic_distributions_and_ecoffs/). Accessed January 24, 2019.
69. Pogue JM, Ortwine JK, Kaye KS. Clinical considerations for optimal use of the polymyxins: a focus on agent selection and dosing. *Clin Microbiol Infect* 2017;4:229–33.
70. Marchand S, Frat JP, Petitpas F, et al. Removal of colistin during intermittent haemodialysis in two critically ill patients. *J Antimicrob Chemother* 2010;8:1836–7.
71. Markou N, Fousteri M, Markantonis SL, et al. Colistin pharmacokinetics in intensive care unit patients on continuous venovenous haemodiafiltration: an observational study. *J Antimicrob Chemother* 2012;10:2459–62.
72. Karvanen M, Plachouras D, Friberg LE, et al. Colistin methanesulfonate and colistin pharmacokinetics in critically ill patients receiving continuous venovenous hemodiafiltration. *Antimicrob Agents Chemother* 2013;1:668–71.
73. Luque S, Sorli L, Li J, et al. Effective removal of colistin methanesulphonate and formed colistin during intermittent haemodialysis in a patient infected by polymyxin-only-susceptible *Pseudomonas aeruginosa*. *J Chemother* 2014;2: 122–4.
74. Mariano F, Leporati M, Carignano P, Stella M, Vincenti M, Biancone L. Efficient removal of colistin A and B in critically ill patients undergoing CVVHDF and sorbent technologies. *J Nephrol* 2015;5:623–31.
75. Jacobs M, Gregoire N, Megarbane B, et al. Population pharmacokinetics of colistin methanesulphonate (CMS) and colistin in critically ill patients with acute renal failure requiring intermittent haemodialysis. *Antimicrob Agents Chemother* 2016;60(3):1788–93. 10.1128/AAC.01868-15. [PubMed: 26729492]
76. Karaiskos I, Friberg LE, Galani L, et al. Challenge for higher colistin dosage in critically ill patients receiving continuous venovenous haemodiafiltration. *Int J Antimicrob Agents* 2016;3:337–41.
77. Strunk AK, Schmidt JJ, Baroke E, et al. Single- and multipledose pharmacokinetics and total removal of colistin in a patient with acute kidney injury undergoing extended daily dialysis. *J Antimicrob Chemother* 2014;7:2008–10.
78. John JF, Falci DR, Rigatto MH, Oliveira RD, Kremer TG, Zavascki AP. Severe infusion-related adverse events and renal failure in patients receiving high-dose intravenous polymyxin B. *Antimicrob Agents Chemother* 2018;62(1). pii:e01617–17. 10.1128/AAC.01617-17. [PubMed: 29038262]

79. Sandri AM, Landersdorfer CB, Jacob J, et al. Pharmacokinetics of polymyxin B in patients on continuous venovenous haemodialysis. *J Antimicrob Chemother* 2013;3:674–7.
80. Nelson BC, Eiras DP, Gomez-Simmonds A, et al. Clinical outcomes associated with polymyxin B dose in patients with bloodstream infections due to carbapenem-resistant Gram-negative rods. *Antimicrob Agents Chemother* 2015;11:7000–6.
81. Kwa AL, Lim TP, Low JG, et al. Pharmacokinetics of polymyxin B1 in patients with multidrug-resistant Gram-negative bacterial infections. *Diagn Microbiol Infect Dis* 2008;2:163–7.
82. Abdelraouf K, Braggs KH, Yin T, Truong LD, Hu M, Tam VH. Characterization of polymyxin B-induced nephrotoxicity: implications for dosing regimen design. *Antimicrob Agents Chemother* 2012;9:4625–9.
83. Polymyxin B [package insert]. Big Flats, NY: Xellia Pharmaceuticals; 2015.
84. Baird JS. Polymyxin B and haemofiltration in an adolescent with leukaemia. *J Antimicrob Chemother* 2014;5:1434.
85. Rigatto MH, Falci DR, Lopes NT, Zavascki AP. Clinical features and mortality of patients on renal replacement therapy receiving polymyxin B. *Int J Antimicrob Agents* 2016;2:146–50.
86. Spapen HD, Honore PM, Gregoire N, et al. Convulsions and apnoea in a patient infected with New Delhi metallo-beta-lactamase-1 *Escherichia coli* treated with colistin. *J Infect* 2011;6:468–70.
87. Bode-Boger SM, Schopp B, Troger U, Martens-Lobenhoffer J, Kalousis K, Mailander P. Intravenous colistin in a patient with serious burns and borderline syndrome: the benefits of therapeutic drug monitoring. *Int J Antimicrob Agents* 2013;4:357–60.
88. Gauthier TP, Wolowich WR, Reddy A, Cano E, Abbo L, Smith LB. Incidence and predictors of nephrotoxicity associated with intravenous colistin in overweight and obese patients. *Antimicrob Agents Chemother* 2012;5:2392–6.
89. Dubrovskaya Y, Prasad N, Lee Y, Esaian D, Figueroa DA, Tam VH. Risk factors for nephrotoxicity onset associated with polymyxin B therapy. *J Antimicrob Chemother* 2015;6:1903–7.
90. Pogue JM, Lee J, Marchaim D, et al. Incidence of and risk factors for colistin-associated nephrotoxicity in a large academic health system. *Clin Infect Dis* 2011;9:879–84.
91. Temocin F, Erdinc S, Tulek N, Demirelli M, Bulut C, Ertem G. Incidence and risk factors for colistin-associated nephrotoxicity. *Jpn J Infect Dis* 2015;4:318–20.
92. Rigatto MH, Behle TF, Falci DR, et al. Risk factors for acute kidney injury (AKI) in patients treated with polymyxin B and influence of AKI on mortality: a multicentre prospective cohort study. *J Antimicrob Chemother* 2015;5:1552–7.
93. Petrosillo N, Giannella M, Antonelli M, et al. Clinical experience of colistin-glycopeptide combination in critically ill patients infected with Gram-negative bacteria. *Antimicrob Agents Chemother* 2014;2:851–8.
94. Garnacho-Montero J, Amaya-Villar R, Gutierrez-Pizarra A, et al. Clinical efficacy and safety of the combination of colistin plus vancomycin for the treatment of severe infections caused by carbapenem-resistant *Acinetobacter baumannii*. *Chemotherapy* 2013;3:225–31.
95. Gordon NC, Png K, Wareham DW. Potent synergy and sustained bactericidal activity of a vancomycin-colistin combination versus multidrug-resistant strains of *Acinetobacter baumannii*. *Antimicrob Agents Chemother* 2010;12:5316–22.
96. Rattanaumpawan P, Ungprasert P, Thamlikitkul V. Risk factors for colistin-associated nephrotoxicity. *J Infect* 2011;2:187–90.
97. Elias LS, Konzen D, Krebs JM, Zavascki AP. The impact of polymyxin B dosage on in-hospital mortality of patients treated with this antibiotic. *J Antimicrob Chemother* 2010;10:2231–7.
98. Phe K, Shields RK, Tverdek FP, et al. Predicting the risk of nephrotoxicity in patients receiving colistimethate sodium: a multicentre, retrospective, cohort study. *J Antimicrob Chemother* 2016;71:3585–7. [PubMed: 27543655]
99. Roberts KD, Azad MA, Wang J, et al. Antimicrobial activity and toxicity of the major lipopeptide components of polymyxin B and colistin: last-line antibiotics against multidrug-resistant Gram-negative bacteria. *ACS Infect Dis* 2015;11:568–75.
100. Tuon FF, Aragao BZ, Santos TA, Gasparetto J, Cordova K, Abujamra M. Acute kidney injury in patients using amikacin in an era of carbapenem-resistant bacteria. *Infect Dis (Lond)* 2016;11–12:869–71.

101. Zavascki AP, Manfro RC, Maciel RA, Falci DR. Head and neck hyperpigmentation probably associated with Polymyxin B therapy. *Ann Pharmacother*. 2015;49(10):1171–2. 10.1177/1066028015595643. [PubMed: 26187742]
102. Yousef JM, Chen G, Hill PA, Nation RL, Li J. Ascorbic acid protects against the nephrotoxicity and apoptosis caused by colistin and affects its pharmacokinetics. *J Antimicrob Chemother* 2012;2:452–9.
103. Dalfino L, Puntillo F, Ondok MJ, et al. Colistin-associated acute kidney injury in severely ill patients: a step toward a better renal care? A prospective cohort study. *Clin Infect Dis* 2015;12:1771–7.
104. Sirijatuphat R, Limmahakhun S, Sirivatanauksorn V, Nation RL, Li J, Thamlikitkul V. Preliminary clinical study of the effect of ascorbic acid on colistin-associated nephrotoxicity. *Antimicrob Agents Chemother* 2015;6:3224–32.
105. Zarjou A, Agarwal A. Sepsis and acute kidney injury. *J Am Soc Nephrol* 2011;6:999–1006.
106. Rojas LJ, Salim M, Cober E, et al. Colistin resistance in carbapenem-resistant *Klebsiella pneumoniae*: laboratory detection and impact on mortality. *Clin Infect Dis* 2016;64(6):711–718. 10.1093/cid/ciw805.
107. Marchaim D, Chopra T, Pogue JM, et al. Outbreak of colistin-resistant, carbapenem-resistant *Klebsiella pneumoniae* in metropolitan Detroit, Michigan. *Antimicrob Agents Chemother* 2011;2:593–9.
108. Qureshi ZA, Hittle LE, O’Hara JA, et al. Colistin-resistant *Acinetobacter baumannii*: beyond carbapenem resistance. *Clin Infect Dis* 2015;9:1295–303.
109. Lenhard JR, Thamlikitkul V, Silveira FP, et al. Polymyxin-resistant, carbapenem-resistant *Acinetobacter baumannii* is eradicated by a triple combination of agents that lack individual activity. *J Antimicrob Chemother*. 2017;72(5):1415–20. 10.1093/jac/dkx002. [PubMed: 28333347]
110. Bulman ZP, Ly NS, Lenhard JR, Holden PN, Bulitta JB, Tsuji BT. Influence of rhlR and lasR on polymyxin pharmacodynamics in *Pseudomonas aeruginosa* and implications for quorum sensing inhibition with azithromycin. *Antimicrob Agents Chemother* 2017;61(4). pii: e00096–16. 10.1128/AAC.00096-16. [PubMed: 28096154]
111. Smith NM, Bulman ZP, Sieron AO, et al. Pharmacodynamics of dose-escalated ‘front-loading’ polymyxin B regimens against polymyxin-resistant mcr-1-harboring *Escherichia coli*. *J Antimicrob Chemother* 2017;8:2297–303.
112. Bulman ZP, Chen L, Walsh TJ, et al. Polymyxin combinations combat *Escherichia coli* harboring mcr-1 and bla<sub>NDM-5</sub>: preparation for a postantibiotic era. *MBio* 2017;8(4). pii: e00540–17. 10.1128/mBio.00540-17. [PubMed: 28743810]
113. Zhao M, Bulman ZP, Lenhard JR, et al. Pharmacodynamics of colistin and fosfomycin: a ‘treasure trove’ combination combats KPC-producing *Klebsiella pneumoniae*. *J Antimicrob Chemother* 2017;7:1985–90.
114. Landersdorfer CB, Ly NS, Xu H, Tsuji BT, Bulitta JB. Quantifying subpopulation synergy for antibiotic combinations via mechanism-based modeling and a sequential dosing design. *Antimicrob Agents Chemother* 2013;5:2343–51.
115. Lenhard JR, Smith NM, Bulman ZP, et al. High dose ampicillin/sulbactam combinations combat polymyxin-resistant *Acinetobacter baumannii* in a hollow-fiber infection model. *Antimicrob Agents Chemother* 2017;61(3). pii: e01268–16. 10.1128/AAC.01268-16.
116. Ly NS, Bulitta JB, Rao GG, et al. Colistin and doripenem combinations against *Pseudomonas aeruginosa*: profiling the time course of synergistic killing and prevention of resistance! *J Antimicrob Chemother* 2015;5:1434–42.
117. Paul M, Carmeli Y, Durante-Mangoni E, et al. Combination therapy for carbapenem-resistant Gram-negative bacteria. *J Antimicrob Chemother* 2014;9:2305–9. 10.1093/jac/dku168.
118. Zusman O, Avni T, Leibovici L, et al. Systematic review and meta-analysis of in vitro synergy of polymyxins and carbapenems. *Antimicrob Agents Chemother*. 2013;57 (10):5104–11. 10.1128/AAC.01230-13. [PubMed: 23917322]



119. Zarkotou O, Pournaras S, Tselioti P, et al. Predictors of mortality in patients with bloodstream infections caused by KPC-producing *Klebsiella pneumoniae* and impact of appropriate antimicrobial treatment. *Clin Microbiol Infect* 2011;12:1798–803.
120. Qureshi ZA, Paterson DL, Potoski BA, et al. Treatment outcome of bacteremia due to KPC-producing *Klebsiella pneumoniae*: superiority of combination antimicrobial regimens. *Antimicrob Agents Chemother* 2012;4:2108–13.
121. Daikos GL, Tsaousi S, Tzouveleki LS, et al. Carbapenemase-producing *Klebsiella pneumoniae* bloodstream infections: lowering mortality by antibiotic combination schemes and the role of carbapenems. *Antimicrob Agents Chemother* 2014;4:2322–8.
122. Gutierrez-Gutierrez B, Salamanca E, de Cueto M, et al. Effect of appropriate combination therapy on mortality of patients with bloodstream infections due to carbapenemase-producing Enterobacteriaceae (INCREMENT): a retrospective cohort study. *Lancet Infect Dis* 2017;7:726–34.
123. Tumbarello M, Viale P, Viscoli C, et al. Predictors of mortality in bloodstream infections caused by *Klebsiella pneumoniae* carbapenemase-producing *K. pneumoniae*: importance of combination therapy. *Clin Infect Dis* 2012;7:943–50.
124. Tumbarello M, Trecarichi EM, De Rosa FG, et al. Infections caused by KPC-producing *Klebsiella pneumoniae*: differences in therapy and mortality in a multicentre study. *J Antimicrob Chemother* 2015;70(7):2133–43. [PubMed: 25900159]
125. Paul M, Daikos GL, Durante-Mangoni E, et al. Colistin alone versus colistin plus meropenem for treatment of severe infections caused by carbapenem-resistant Gram-negative bacteria: an open-label, randomised controlled trial. *Lancet Infect Dis* 2018;18(4):391–400. [PubMed: 29456043]
126. Durante-Mangoni E, Signoriello G, Andini R, et al. Colistin and rifampicin compared with colistin alone for the treatment of serious infections due to extensively drug-resistant *Acinetobacter baumannii*: a multicenter, randomized clinical trial. *Clin Infect Dis* 2013;3:349–58.
127. Sirijatuphat R, Thamlikitkul V. Preliminary study of colistin versus colistin plus fosfomycin for treatment of carbapenem-resistant *Acinetobacter baumannii* infections. *Antimicrob Agents Chemother* 2014;9:5598–601.
128. Aydemir H, Akduman D, Piskin N, et al. Colistin vs. the combination of colistin and rifampicin for the treatment of carbapenem-resistant *Acinetobacter baumannii* ventilator-associated pneumonia. *Epidemiol Infect* 2013;6:1214–22.
129. Furtado GH, d’Azevedo PA, Santos AF, et al. Intravenous polymyxin B for the treatment of nosocomial pneumonia caused by multidrug-resistant *Pseudomonas aeruginosa*. *Int J Antimicrob Agents* 2007;30:315–9. [PubMed: 17631984]
130. Falagas ME, Rafailidis PI, Ioannidou E, et al. Colistin therapy for microbiologically documented multidrug-resistant gram-negative bacterial infections: a retrospective cohort study of 258 patients. *Int J Antimicrob Agents* 2010;35:194–9. [PubMed: 20006471]
131. Samonis G, Vardakas KZ, Kofteridis DP, et al. Characteristics, risk factors and outcomes of adult cancer patients with extensively drug-resistant *Pseudomonas aeruginosa* infections. *Infection* 2014;42:721–8. [PubMed: 24912861]
132. Rigatto MH, Vieira FJ, Antochévis LC, et al. Polymyxin B in combination with antimicrobials lacking in vitro activity versus polymyxin B in monotherapy in critically ill patients with *Acinetobacter baumannii* or *Pseudomonas aeruginosa* infections. *Antimicrob Agents Chemother* 2015;59:6575–80. [PubMed: 26259799]
133. Ribera A, Benavent E, Lora-Tamayo J, et al. Osteoarticular infection caused by MDR *Pseudomonas aeruginosa*: the benefits of combination therapy with colistin plus  $\beta$ -lactams. *J Antimicrob Chemother* 2015;70:3357–65. [PubMed: 26419763]
134. Sorlí L, Luque S, Segura C, et al. Impact of colistin plasma levels on the clinical outcome of patients with infections caused by extremely drug-resistant *Pseudomonas aeruginosa*. *BMC Infect Dis* 2017;17:11. [PubMed: 28056821]
135. Rattanaumpawan P, Lorsuthitham J, Ungprasert P, Angkasekwinai N, Thamlikitkul V. Randomized controlled trial of nebulized colistimethate sodium as adjunctive therapy of ventilator-associated pneumonia caused by Gram-negative bacteria. *J Antimicrob Chemother* 2010;12:2645–9.

136. Abdellatif S, Trifi A, Daly F, et al. Efficacy and toxicity of aerosolised colistin in ventilator-associated pneumonia: a prospective, randomised trial. *Ann Intensive Care*. 2016;6 (1):26-10.1186/s13613-016-0127-7. [PubMed: 27033711]
137. Valachis A, Samonis G, Kofteridis DP. The role of aerosolized colistin in the treatment of ventilator-associated pneumonia: a systematic review and metaanalysis. *Crit Care Med* 2015;3:527–33.
138. Polat M, Kara SS, Tapisiz A, Tezer H, Kalkan G, Dolgun A. Treatment of ventilator-associated pneumonia using intravenous colistin alone or in combination with inhaled colistin in critically ill children. *Paediatr Drugs* 2015;4:323–30.
139. Kalin G, Alp E, Coskun R, Demiraslan H, Gundogan K, Doganay M. Use of high-dose IV and aerosolized colistin for the treatment of multidrug-resistant *Acinetobacter baumannii* ventilator-associated pneumonia: do we really need this treatment? *J Infect Chemother* 2012;6:872–7.
140. Lu Q, Luo R, Bodin L, et al. Efficacy of high-dose nebulized colistin in ventilator-associated pneumonia caused by multidrug-resistant *Pseudomonas aeruginosa* and *Acinetobacter baumannii*. *Anesthesiology* 2012;6:1335–47.
141. Vardakas KZ, Voulgaris GL, Samonis G, Falagas ME. Inhaled colistin monotherapy for respiratory tract infections in adults without cystic fibrosis: a systematic review and meta-analysis. *Int J Antimicrob Agents*. 2018;51:1–9. 10.1016/j.ijantimicag.2017.05.016. [PubMed: 28669836]
142. Jung SY, Lee SH, Lee SY, et al. Antimicrobials for the treatment of drug-resistant *Acinetobacter baumannii* pneumonia in critically ill patients: a systemic review and Bayesian network meta-analysis. *Crit Care*. 2017;21:319 10.1186/s13054-017-1916-6. [PubMed: 29262831]
143. Jang JY, Kwon HY, Choi EH, et al. Efficacy and toxicity of high-dose nebulized colistin for critically ill surgical patients with ventilator-associated pneumonia caused by multidrug-resistant *Acinetobacter baumannii*. *J Crit Care*. 2017;40:251–6. 10.1016/j.jcrc.2017.04.004. [PubMed: 28458172]
144. Klick JM, du Moulin GC, Hedley-Whyte J, Teres D, Bushnell LS, Feingold DS. Prevention of gram-negative bacillary pneumonia using polymyxin aerosol as prophylaxis. II. Effect on the incidence of pneumonia in seriously ill patients. *J Clin Invest* 1975;3:514–9.
145. Feeley TW, Du Moulin GC, Hedley-Whyte J, Bushnell LS, Gilbert JP, Feingold DS. Aerosol polymyxin and pneumonia in seriously ill patients. *N Engl J Med* 1975;10:471–5.
146. Boisson M, Jacobs M, Gregoire N, et al. Comparison of intrapulmonary and systemic pharmacokinetics of colistin methanesulfonate (CMS) and colistin after aerosol delivery and intravenous administration of CMS in critically ill patients. *Antimicrob Agents Chemother* 2014;12:7331–9.
147. Athanassa ZE, Markantonis SL, Fousteri MZ, et al. Pharmacokinetics of inhaled colistimethate sodium (CMS) in mechanically ventilated critically ill patients. *Intensive Care Med* 2012;11:1779–86.
148. Huang JX, Blaskovich MA, Pelingon R, et al. Mucin binding reduces colistin antimicrobial activity. *Antimicrob Agents Chemother* 2015;59(10):5925–31. [PubMed: 26169405]
149. Wenzler E, Fraidenburg DR, Scardina T, Danziger LH. Inhaled antibiotics for Gram-negative respiratory infections. *Clin Microbiol Rev* 2016;3:581–632.
150. Rouby JJ, Bouhemad B, Monsel A, Brisson H, Arbelot C, Lu Q. Aerosolized antibiotics for ventilator-associated pneumonia: lessons from experimental studies. *Anesthesiology* 2012;6:1364–80.
151. Ehrmann S, Roche-Campo F, Sferrazza Papa GF, Isabey D, Brochard L, Apiou-Sbirlea G. Aerosol therapy during mechanical ventilation: an international survey. *Intensive Care Med* 2013;6:1048–56.
152. Rello J, Solé-Lleonart C, Rouby JJ, et al. Use of nebulized antimicrobials for the treatment of respiratory infections in invasively mechanically ventilated adults: a position paper from the European Society of Clinical Microbiology and Infectious Diseases. *Clin Microbiol Infect*. 2017;23(9):629–39. 10.1016/j.cmi.2017.04.011. [PubMed: 28412382]

153. Tunkel AR, Hasbun R, Bhimraj A, et al. 2017 Infectious Diseases Society of America's Clinical Practice Guidelines for Healthcare-Associated Ventriculitis and Meningitis. *Clin Infect Dis* 2017; 24(14):1093-1103. doi:10.1093/cid/ciw861.
154. Kim BN, Peleg AY, Lodise TP, et al. Management of meningitis due to antibiotic-resistant *Acinetobacter* species. *Lancet Infect Dis* 2009;4:245-55.
155. Karaiskos I, Giamarellou H. Multidrug-resistant and extensively drug-resistant gram-negative pathogens: current and emerging therapeutic approaches. *Expert Opin Pharmacother* 2014;10:1351-70.
156. Markantonis SL, Markou N, Fousteri M, et al. Penetration of colistin into cerebrospinal fluid. *Antimicrob Agents Chemother* 2009;11:4907-10.
157. Antachopoulos C, Karvanen M, Iosifidis E, et al. Serum and cerebrospinal fluid levels of colistin in pediatric patients. *Antimicrob Agents Chemother* 2010;9:3985-7.
158. Imberti R, Cusato M, Accetta G, et al. Pharmacokinetics of colistin in cerebrospinal fluid after intraventricular administration of colistin methanesulfonate. *Antimicrob Agents Chemother* 2012;8:4416-21.
159. Falagas ME, Bliziotis IA, Tam VH. Intraventricular or intrathecal use of polymyxins in patients with gram-negative meningitis: a systematic review of the available evidence. *Int J Antimicrob Agents* 2007;1:9-25.
160. Karaiskos I, Galani L, Baziaka F, Giamarellou H. Intraventricular and intrathecal colistin as the last therapeutic resort for the treatment of multidrug-resistant and extensively drug-resistant *Acinetobacter baumannii* ventriculitis and meningitis: a literature review. *Int J Antimicrob Agents* 2013;6:499-508.
161. Bargiacchi O, De Rosa FG. Intrathecal or intraventricular colistin: a review. *Infez Med* 2016;1:3-11.
162. Piparsania S, Rajput N, Bhatambare G. Intraventricular polymyxin B for the treatment of neonatal meningo-ventriculitis caused by multi-resistant *Acinetobacter baumannii*-case report and review of literature. *Turk J Pediatr* 2012;5:548-54.
163. Hoenigl M, Drescher M, Feierl G, et al. Successful management of nosocomial ventriculitis and meningitis caused by extensively drug-resistant *Acinetobacter baumannii* in Austria. *Can J Infect Dis Med Microbiol* 2013;3:e88-90.
164. Remes F, Tomas R, Jindrak V, Vanis V, Setlik M. Intraventricular and lumbar intrathecal administration of antibiotics in postneurosurgical patients with meningitis and/or ventriculitis in a serious clinical state. *J Neurosurg* 2013;6:1596-602.
165. Karagoz G, Kadanali A, Dede B, et al. Extensively drug-resistant *Pseudomonas aeruginosa* ventriculitis and meningitis treated with intrathecal colistin. *Int J Antimicrob Agents* 2014;1:93-4.
166. Bargiacchi O, Rossati A, Car P, et al. Intrathecal/intraventricular colistin in external ventricular device-related infections by multi-drug resistant gram negative bacteria: case reports and review. *Infection* 2014;5:801-9.
167. Santos AS, Iraneta A, Matos M, Brito MJ. Intraventricular colistin in gram-negative ventriculoperitoneal shunt infection in two pediatric patients. *Acta Neurochir (Wien)* 2015;12:2219-20.
168. Schiaroli E, Pasticci MB, Cassetta MI, et al. Management of meningitis caused by multi drug-resistant *Acinetobacter baumannii*: clinical, microbiological and pharmacokinetic results in a patient treated with colistin methanesulfonate. *Mediterr J Hematol Infect Dis* 2015;1:e2015055.
169. Shofty B, Neuberger A, Naffaa ME, et al. Intrathecal or intraventricular therapy for post-neurosurgical gram-negative meningitis: matched cohort study. *Clin Microbiol Infect* 2016;1:66-70.
170. Shrestha GS, Tamang S, Paneru HR, et al. Colistin and tige-cycline for management of external ventricular device-related ventriculitis due to multidrug-resistant *Acinetobacter baumannii*. *J Neurosci Rural Pract* 2016;3:450-2.
171. De Bonis P, Lofrese G, Scoppettuolo G, et al. Intraventricular versus intravenous colistin for the treatment of extensively drug resistant *Acinetobacter baumannii* meningitis. *Eur J Neurol* 2016;1:68-75.

172. Souhail D, Bouchra B, Belarj B, et al. Place of colistin-rifampicin association in the treatment of multidrug-resistant *Acinetobacter baumannii* meningitis: a case study. *Case Rep Infect Dis* 2016;2016:8794696. [PubMed: 27064923]
173. Fotakopoulos G, Makris D, Chatzi M, Tsimitrea E, Zakyntinos E, Fountas K. Outcomes in meningitis/ventriculitis treated with intravenous or intraventricular plus intravenous colistin. *Acta Neurochir (Wien)* 2016;3:603–10; discussion 610.
174. Inamasu J, Ishikawa K, Oheda M, Nakae S, Hirose Y, Yoshida S. Intrathecal administration of colistin for meningitis due to New Delhi metallo-beta-lactamase 1(NDM-1)-producing *Klebsiella pneumoniae*. *J Infect Chemother* 2016;3:184–6.
175. Ceylan B, Arslan F, Sipahi OR, et al. Variables determining mortality in patients with *Acinetobacter baumannii* meningitis/ventriculitis treated with intrathecal colistin. *Clin Neurol Neurosurg* 2017;153:43–9. [PubMed: 28013184]
176. Singh RK, Bhoi SK, Kalita J, Misra UK. Multidrug-resistant *Acinetobacter* meningitis treated by intrathecal colistin. *Ann Indian Acad Neurol* 2017;1:74–5.

**Table 1.**

## CLSI/EUCAST Breakpoints for Colistin

Organism	Colistin MIC, mg/L		
	Susceptible	Intermediate	Resistant
CLSI <sup>a</sup>			
<i>Acinetobacter</i> sp	2	-	4
<i>Pseudomonas aeruginosa</i>	2	-	4
EUCAST			
<i>Acinetobacter</i> sp	2		> 2
<i>P. aeruginosa</i>	2		> 2
Enterobacteriaceae	2		> 2

CLSI = Clinical and Laboratory Standards Institute; EUCAST = European Committee on Antimicrobial Susceptibility Testing; MIC = minimum inhibitory concentration.

<sup>a</sup>For isolates of *Enterobacter aerogenes*, *Enterobacter cloacae*, *Escherichia coli*, *Klebsiella pneumoniae*, and *Raoultella ornithinolytica*, CLSI define insufficient clinical and pharmacokinetic/pharmacodynamic (PK/PD) data to set a PK/PD-based breakpoint and cite epidemiological cutoff values (ECVs) of 2 mg/L based on MIC distribution data.<sup>11, 12</sup>

**Table 2.**Look-up Table of Daily Doses of CMS<sup>a</sup>

Daily dose of CMS for plasma colistin C <sub>ss,avg</sub> of 2 mg/L <sup>c</sup>		
Creatinine clearance, mL/minute <sup>b</sup>	mg CBA/day	Million IU/day
0	130	3.95
5 to < 10	145	4.40
10 to < 20	160	4.85
20 to < 30	175	5.30
30 to < 40	195	5.90
40 to < 50	220	6.65
50 to < 60	245	7.40
60 to < 70	275	8.35
70 to < 80	300	9.00
80 to < 90	340	10.3
90	360	10.9

CBA = colistin base activity; CMS = colistin methanesulfonate; C<sub>ss,avg</sub> = average steady-state plasma concentration;

<sup>a</sup>To achieve a desired target plasma colistin C<sub>ss,avg</sub> of 2 mg/L for patients with narrow windows of creatinine clearance. Reproduced from reference 6 with minor modifications.

<sup>b</sup>Adjusted body weight should be used to estimate creatinine clearance.

<sup>c</sup>Daily dose administered in two divided doses 12 hours apart.