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Dimeric small molecule agonists of EphA2 receptor inhibit glioblastoma cell growth

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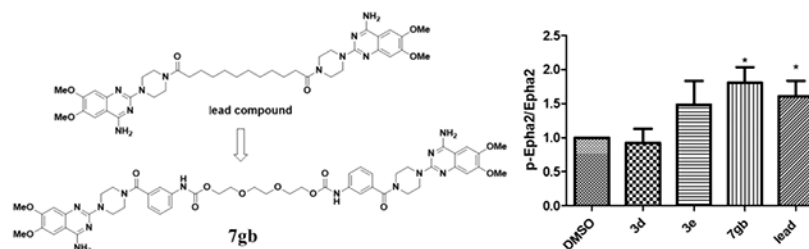
Abstract

EphA2 receptor kinase could become a novel target for anti-glioblastoma treatment. Doxazosin previously identified acts like the endogenous ligand of EphA2 and induces cell apoptosis. Through lead structure modification a derivative of Doxazosin possessing unique dimeric structure showed an improvement in the activity. In the current study, we expanded the dimeric scaffold by lead optimization to explore the chemical space of the conjoining moieties and a slight variation to the core structure. 27 new derivatives were synthesized and examined with EphA2 overexpressed and wild type glioblastoma cell lines for cell proliferation and EphA2 activation. Three new compounds **3d**, **3e**, and **7bg** showed potent and selective activities against the growth of EphA2 overexpressed glioblastoma cells. Dimer **3d** modification replaces the long alkyl chain with a short polyethylene glycol chain. Dimer **7bg** has a relatively longer polyethylene glycol chain in comparison to compound **3d** and the length is more similar to the lead compound. Whereas dimer **3e** has a rigid aromatic linker exploring the chemical space. The diversity of the linkers in the active suggest additional hydrogen binding sites has a positive correlation to the activity. All three dimers showed selective activity in EphA2 overexpressed cells, indicating the activity is correlated to the EphA2 targeting effect.

Graphical Abstract

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Keywords

EphA2; glioblastoma; Doxazosin; agonist; dimer

1. Introduction

Glioblastoma multiforme (GBM) is the most common and malignant primary neurological brain tumor in adults.¹⁻³ Treatment options for GBM are still very limited, and the standard available chemotherapy agent for GBM is Temozolomide (TMZ), a DNA-alkylating reagent.⁴ This current treatment often fails due to resistance.⁵ Exploring other pathways to inhibit malignancy growth is critical in extending patient life expectancy. Erythropoietin-producing hepatocellular receptor 2 (EphA2) has shown to play an important role in cancer migration.^{2,3,6} It has been well documented that EphA2 is overexpressed in GBM, and the overexpression is correlated with malignant progression and poor prognosis.^{7,8} Studies indicate that EphA2 activation by its ligand ephrin-A1 has the ability to suppress tumor progression. In addition, the activation causes induction of apoptosis, inhibition of cell proliferation, and suppression of cell migration.⁹⁻¹² *In vivo* studies demonstrated restoring basal levels of ephrin-A1 by systemic administration decreases tumorigenicity and invasiveness of carcinoma xenografts.^{13,14} Unfortunately, EphA2 overexpression is often accompanied by loss of expression or mis-localization of ephrin-A1 in tumor tissue.^{3,6,15} The reduced ligand coupled with increased EphA2 expression results in frequent Akt activation, which promotes ligand-independent pro-invasive Akt-EphA2 crosstalk.^{3,16,17} This interaction may be in part responsible for EphA2 overexpression during tumor progression and the positive correlation of EphA2 expression and unfavorable prognosis.^{1,17} From a therapeutic point of view, restoring the ligand for EphA2 could recover the tumor suppressing activity of EphA2, and provide a new approach for the treatment of EphA2 overexpressed tumor. Unfortunately, the endogenous ligand of EphA2, ephrin-A1, could not effectively pass blood brain barrier (BBB), which makes this endogenous ligand less useful for the treatment of glioblastoma with overexpression of EphA2. This fact led us to hypothesize that a small molecule agonists for EphA2 can be exploited as novel cancer therapeutic.^{18,19}

In previous studies, we identified a small molecule EphA2 agonist Doxazosin using a combination of structure-based virtual screening and cell-based assays.¹⁹ Further lead optimization of Doxazosin to activate EphA2 resulted in one particular compound with a unique dimeric structure and superior activity compared to Doxazosin (Figure 1).¹⁸

Furthermore, this dimer also has the capability to cross BBB.²⁰ The novel symmetric structure provides us a new scaffold for further optimization.

In this current study, we performed optimization of the dimeric lead compound. 27 new derivatives were synthesized and examined with glioblastoma cell lines. Three new dimers **3d**, **3e**, and **7bg** showed similar activities compared to the lead compound (scheme 1-3). Dimer **3d** exchanges the long alkyl chain with a short polyethylene glycol linker, this modification reduces the molecular weight and introduces hydrogen bonding acceptors into the structure allowing different interactions to happen inside the ligand binding domain. Dimer **7bg** has a relatively longer polyethylene glycol chain compared to compound **3d**, and has an additional benzamide moiety exploring the effect of the use of the polyethylene glycol chain. Dimer **3e** has a rigid aromatic linker, suggesting both the flexibility and bulkiness of the compound are not critical to retain activity. All three dimers showed better activity in EphA2 overexpressed cells versus wild type glioma cells, indicating the targeting effect is through EphA2 pathways. However, only **7bg** showed significant EphA2 activation, suggesting that the introduction of a benzamide moiety increases the ability to bind to the EphA2 binding domain in comparison to compound **3d** which also contains a polyethylene glycol linker. Compounds **3d** and **3e** failed to cause EphA2 activation as effectively as the lead compound indicating that the modification may lead to other targeting effects to the pathway instead of EphA2 activation.

2. Results and discussion

2.1. Development of an EphA2 overexpressed glioblastoma cell line

Targeting EphA2 to induce cell apoptosis provides a benefit due to lower systematic toxicity in comparison to current treatment by TMZ, a chemotherapy agent for GBM. The ideal drug candidates should selectively decrease the growth of EphA2 overexpressed cells. To examine the selectivity of the compounds, we constructed a U251 GBM cell line with overexpression of EphA2.

The western blotting results indicate that the basal EphA2 expression in U251 cells is negligible and similar to the empty vector transfected cells, whereas EphA2 transfected cells have a 5-fold increase (Figure 2). Previously the lead compound resulted in inhibition of growth based on these three cell lines wild type with an IC₅₀ of 5.2 μM, the empty vector with an IC₅₀ of 4.8 μM, and the EphA2 overexpressed with an IC₅₀ of 2.1 μM. The results are consistent to our previous study that the dimeric lead compound is a selective EphA2 ligand,¹⁸ and inhibits cell growth via activation of EphA2.

2.2. Lead optimization and biological evaluation

Our previous efforts resulted in a new EphA2 agonist with a dimeric structure (Figure 1).¹⁸ It significantly activated EphA2 at 2 μM *in vitro*, and induced EphA2 kinase internalization at 2 μM as well. In this study, a total of 27 new derivatives were synthesized using combinatorial chemistry strategy, mainly focused on the exploration of the dimeric structures.

First, we focused on the modification of the linker of this lead compound (Scheme 1). The alkyl linker cannot serve as either hydrogen bond acceptor or donor. To explore the ligand binding domain, these linkers were exchanged with various structures possessing hydrogen bonding capability. Therefore, we introduced polyethylene glycol linkers to overcome the possibility that introduction of a hydrogen bond acceptor or donor could increase the binding activity (compound **3c** and **3d**). To explore if length could be a contributing factor, we also generated analogs with short alkyl linkers (compounds **3a**, **3b**). We also wanted to explore if flexibility and bulkiness mattered in the ligand binding domain, therefore rigid aromatic linkers were used too (compounds **3e**, **3f**). The first set of new linkers for the dimers are listed below (Table 1).

We also modified the original core of the structure by adding benzamide moieties and varying the aromatic nitro group (**5a-5f**). These nitro groups were reduced to amino group (**7a-7f**) (Scheme 2). To explore if this modification retained activity these compounds were tested with the U251 cell lines. Of these new analogs we varied the combination of nitro group with different substitutions to examine their ability to target EphA2 (Table 2).

From the new analogs we then attempted to link the two together with the same set of linkers used beforehand (Scheme 3). Unfortunately, with the additional benzamide moiety, the formed dimers have poor solubility in various solvents making the purification not practicable. Overall, 9 new dimers were generated with this process (Table 3).

The 27 new analogs including 15 dimers and 12 monomers show diverse substitution on the linker, and the biological results of the compounds can provide us a detailed SAR to elucidate the structural requirement for the targeting of EphA2.

To evaluate the compounds, we utilized the above developed EphA2 overexpressed U251 cells. To verify the selectivity of the compounds, we also screened the compounds with wild type U251 cells with low EphA2 expression. The selective index is calculated based on the IC₅₀s of the two cell lines (Table 4).

All the compounds were examined with multiple doses, and the cells were treated with the compounds for 48 hr, and cell viability was determined after. We used the lead compound and 0.1% DMSO as the positive and negative control, respectively.

Among the first set of dimers (scheme 1 and table 1) the activity varied. Compounds **3a** and **3b** with reduced length of the alkyl chain resulted in decreased activity, which is consistent with our previous results and should be eliminated as an optimization direction in the future. Compounds **3d-3f** all showed similar or improved activity compared to the lead, which possible is due to the introduction of hydrogen binding site or by reducing the flexibility. **3e** and **3f** both have a rigid and aromatic linker, and both inhibited the growth of EphA2 overexpressed cells with IC₅₀s in the low micro mole range, suggesting that keeping the two monomers far away from each other benefit the activity. Compounds **3c** and **3d** have polyethylene glycol linkers which may contribute to the binding of the ligand to EphA2 via hydrogen bonding. Compound **3c** lost the potency but retained the selectivity for EphA2 compared to the wild type cell lines. However, **3d** has a different polyethylene glycol linker,

and the potency dramatically increased and the selectivity remained. Of the first set of dimmers compound **3d** showed two-fold more selectivity to the EphA2 overexpressed cells than the wild type and improved potency compared to the lead.

In the next scheme we varied the core structure by adding nitro containing benzamide moieties and then reducing the nitro group to an amino group (Scheme 2 and Table 2). For monomers **5a--6f**, most of them have the IC₅₀s above 10 μM, regardless if it is the nitro group or the amino group on the benzamide moiety. Only compounds **6d**, **6e** and **6f** showed relatively better activity, with an IC₅₀ of 3.6, 3.0 and 2.7 μM, respectively. The position of the amino group is critical to the activity and the para position has the greatest activity of the monomers. Compound **6f** had the best selectivity index among all the compounds with a value of 17.78, suggesting the reduction of the nitro group to the amine adds the possibility of improved ligand binding to EphA2 receptor. Overall, the trend in the reduction from a nitro group to an amino group benefitted the activity, suggesting that hydrogen bond donors are important for the activity on the benzamide moiety.

After synthesis of the dimers based on the amino containing benzamide, the final set of dimmers were produced (Scheme 3 and Table 3). For the 9 dimers among this group, the activities of the majority were not appealing with the exception of compound **7bg**. It showed an IC₅₀ of 1.9 μM for EphA2 overexpressed cells and 7.9 μM for wild type of cells, which results in a selective index of 4.2. Although the potency was reduced by the addition of the benzamide moiety, it is clear that introduction of hydrogen binding in the linker has a positive effect on the potency of the compounds. In most cases when the polyethylene glycol linkers are used the dimeric compounds had improved activity compared to the similar monomers. But in comparison to the original core structure, the dimers formed by monomers with benzamide moiety did not show better activity compared to dimers **3a-3f**.

Considering both of the potency and selectivity, we narrow down to compounds **3d**, **3e**, **7bg** for further mechanism investigation.

2.3. EphA2 activation

EphA2 overexpression has been identified in various different malignant cells.^{2,3,24,6,7,12,13,17,21-23} Decreased levels of EphA2-ligand combined with the overexpression of EphA2 in cancer cells lead to the aggressive growth of the tumor, which can be overcome *in vitro* by using endogenous EphA2 ligand or a small molecule ligand.^{12,18,19,22,25} Our lead compound could induce EphA2 phosphorylation just as the endogenous ligand ephrin-A1, and it is necessary to determine if the new analogs could cause EphA2 activation as well.¹⁸

Dimers **3e** and **7bg** showed better activity to induce EphA2 phosphorylation at 2 μM, with only **7bg** showing statistical significance compared to control (Figure 3). To our surprise, dimer **3d** did not show clear EphA2 activation. Even **3d** showed great activity and selectivity to EphA2 overexpressed cells. It is possible that **3d** targets EphA2 with some unknown different mechanism, which needs further investigation.

2.4. Log P value

Since the compounds are designed to be used for the treatment of GBM, whether they can cross BBB is very critical for the potential application. Hansch and Leo found that blood-brain barrier penetration is optimal when the LogP values of the compounds are in the range of 1.5-2.7, with the mean value of 2.1.^{26,27} Therefore, we measured LogP value of compounds **3d**, **3e**, and **7bg** to predict the potential capability of these compounds to cross BBB, and the lead compound was measured as a positive control. The results are listed in Table 5. The data reported here showed that dimers **3e** and **7bg** showed similar LogP values compared to the lead compounds. Although the value is out of the range of the preferred LogP values indicated by Hansch, the lead compound has the ability to cross the BBB shown in our previous study.²⁰ We speculate that for these types of dimeric structures, may cross the BBB with an unknown mechanism instead of passive lipid penetration.

3. Conclusion

Based on the previous structure activity relationship, the dimeric structure of Doxazosin analog showed promising ligand binding activity to EphA2. To further study this unique structure a variety of structural moieties examined if they could further improve the activity, we synthesized 27 new derivatives and 15 which retain the dimeric structure. The dimers consist of several different monomers and various linkers including alkyl, polyethylene glycol and aromatic or rigid structures. Two compounds **7bg** and **3d** showed great potency and selectivity toward EphA2 overexpressed GBM cells. They also stimulated EphA2 phosphorylation, which demonstrated their targeting effect to EphA2 receptor. The LogP values of the two identified dimers are not in the ideal range for their capability to cross the BBB. However, the lead compound has a similar LogP value and does cross the BBB in our previous study,²⁰ suggesting that these types of compounds may possess a novel mechanisms to cross the BBB other than passive lipid penetration, which will need further investigation.

4. Experimental section

4.1. Chemistry

Chemicals were commercially available and used as received without further purification unless otherwise noted. Moisture sensitive reactions were carried out under a dry argon atmosphere in flame-dried glassware. Solvents were distilled before use under argon. Thin-layer chromatography was performed on precoated silica gel F254 plates (Whatman). Silica gel column chromatography was performed using silica gel 60Å (Merck, 230-400 Mesh), and ethanol/dichloromethane was used as the elution solvent. Mass spectra were obtained on the electrospray mass spectrometer at Cleveland State University MS facility Center. The molecular weight of the compounds was examined with LC-MS. All the NMR spectra were recorded on a Bruker 400 MHz in DMSO-*d*₆. Chemical shifts (δ) for ¹H NMR spectra are reported in parts per million to residual solvent protons.

Compounds 3a-3f were prepared as the following procedure.—The dimethoxyquinazolin and piperazine were dissolved in n-BuOH with 1 to 5 mole ratio, and

the mixture was heated to 100°C for 24 hours, then it was cooled down to room temperature. The precipitated white solid was collected via filtration and washed with n-BuOH. After it was dried, the solid was mixed with various different linkers and K₂CO₃ in DMF. It was stirred until the reaction was completed. The reaction was quenched with Na₂CO₃ aqueous solution and stirred overnight, and the precipitated product was collected via filtration and washed with water and dried to give the corresponding compounds.

1,4-bis(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)butane-1,4-dione

(3a): Light pink solid, 80% yield for the last step; ¹H-NMR (400MHz, DMSO-d₆) δ 7.44 (2H, s), 7.14 (4H, b), 6.76 (2H, s), 3.84 (6H, s), 3.81(10H, s) 3.70 (4H, s), 3.55-3.52 (4H, s), 2.63 (4H, s). ¹³C-NMR (400MHz, DMSO-d₆) δ 170.67, 161.62, 159.10, 158.82, 154.73, 149.35, 149.26, 145.50, 145.39, 105.73, 104.20, 103.44, 56.33, 55.89, 45.33, 44.25, 43.93, 41.73, 28.15. DUIS-MS calculated for C₃₂H₄₀N₁₀O₆ [M + H]⁺: 660.31, found : 661.10

1,5-bis(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)pentane-1,5-dione

(3b): Light pink solid, 80% yield for the last step; ¹H-NMR (400MHz, DMSO-d₆) δ 7.43 (2H, s), 7.14 (4H, b), 6.75 (2H, s), 3.84 (6H, s), 3.80 (6H, s), 3.75 (4H, s), 3.70 (4H, s), 3.52 (8H, s), 2.43-2.40 (3H, t, J=7.1 Hz), 1.78 (2H, t, J= 14.0 Hz). ¹³C-NMR (400MHz, DMSO-d₆) δ 171.09, 161.62, 159.10, 158.79, 154.73, 149.35, 149.26, 145.50, 145.39, 105.72, 104.19, 103.44, 56.33, 55.89, 45.34, 44.33, 44.18, 43.94, 41.54, 32.41, 21.09. DUIS-MS calculated for C₃₃H₄₂N₁₀O₆ [M + H]⁺: 674.33, found : 675.15

2,2'-oxybis(1-(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)ethanone)

(3c): Light pink solid, 72% yield for the last step; ¹H-NMR (400MHz, DMSO-d₆) δ 7.43 (2H, s), 7.15 (4H, b), 6.76 (4H, s), 4.31 (4H, s), 3.84 (6H, s), 3.79 (6H, s), 3.75(8H,s) 3.52 (8H, s) ¹³C-NMR (400MHz, DMSO-d₆)) δ 167.62, 161.63, 158.76, 154.73, 149.25, 145.52, 105.74, 104.18, 103.45, 69.64, 56.33, 55.89, 44.72, 44.28, 43.87, 41.66. DUIS-MS calculated for C₃₂H₄₀N₁₀O₇ [M + H]⁺: 676.31, found : 677.20

ethane-1,2-diyl bis(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazine-1-carboxylate)

(3d): Light pink solid, 43% yield for the last step; ¹H-NMR (400MHz, DMSO-d₆) δ 7.43 (2H, s), 7.14 (4H, b), 6.75 (4H, s), 4.27 (4H, s), 3.83 (6H, s), 3.79 (6H, s), 3.78(8H, s), 3.45 (8H, s) ¹³C-NMR (400MHz, DMSO-d₆) δ 162.77, 161.63, 158.76, 155.00, 149.22, 145.52, 105.72, 104.19, 103.46, 63.67, 56.32, 55.87, 43.98, 43.80. DUIS-MS calculated for C₃₂H₄₀N₁₀O₈ [M + H]⁺: 692.30, found : 693.15

propane-2,2-diylbis(4,1-phenylene)bis(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazine-1-carboxylate) **(3e):**

light pink solid, 31% yield for the last step; ¹H-NMR (400MHz, DMSO-d₆) δ 7.45 (2H, s), 7.24-7.18 (8H, m), 7.07-7.06 (4H, m), 6.78(2H, s) 3.84 (9H, s), 3.80 (9H, s), 3.64 (4H, m) 3.50 (4H, m), 1.66 (6H, s). ¹³C-NMR (400MHz, DMSO-d₆) δ 161.67, 158.68, 154.77, 153.60, 149.51, 149.09, 147.45, 145.58, 127.75, 121.74. 105.66, 104.23, 103.48, 56.34, 55.91, 43.78, 42.38, 31.02. DUIS-MS calculated for C₄₅H₅₀N₁₀O₈ [M + H]⁺: 858.38, found : 859.15

cyclohexane-1,1-diylbis(4,1-phenylene)bis(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazine-1-carboxylate) **(3f):**

light pink solid, 31% yield for the last step; ¹H-NMR

(400MHz, DMSO- d_6) δ 7.45 (2H, s), 7.31 (2H, s), 7.28 (2H, s), 7.18(4H, b) 7.06 (2H, s), 7.04 (2H, s), 6.77 (2H, s) 3.84 (6H, s), 3.80 (13H, s), 3.63 (5H, m) 3.50 (6H, m), 2.25 (4H, s), 1.47 (6H, s). ^{13}C -NMR (400MHz, DMSO- d_6) δ 161.66, 158.64, 154.78, 153.56, 149.26, 149.04, 145.59, 145.42, 128.04, 121.91, 105.64, 104.24, 103.47, 56.35, 55.92, 45.52, 43.78, 36.80, 26.17, 22.97. DUIS-MS calculated for $\text{C}_{48}\text{H}_{54}\text{N}_{10}\text{O}_8$ $[\text{M} + \text{H}]^+$: 898.41, found : 899.25

Compounds 5a-5f were prepared as the following procedure.—The dimethoxyquinazolin and piperazine product was mixed with various different substituted acetyl chloride and K_2CO_3 in THF / H_2O one to one mixture. It was stirred until the reaction was completed. The reaction was quenched with Na_2CO_3 aqueous solution and stirred overnight, and the precipitated product was collected via filtration and washed with water and dried to give the corresponding compounds.

(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)(4-nitrophenyl)methanone

(5a): Light yellow solid, 87% yield for the step; ^1H -NMR (400MHz, DMSO- d_6) 8.30, 8.28, (1H, d, $J = 7.9$ Hz), 8.25-8.23 (1H, d, $J = 7.9$), 8.13-8.11 (1H, d, $J = 8.1$ Hz), 7.74-7.72 (1H, d, $J = 8.0$ Hz), 7.47-7.45 (1H, d, $J = 7.0$ Hz) 7.19 (2H, b), 6.76-6.75(1H, d, $J = 5.9$ Hz), 3.94 (2H, s), 3.83 (4H, s), 3.80 (3H, s), 3.73 (2H, s) 3.35 (1H, s), 3.12 (1H, s). ^{13}C -NMR (400MHz, DMSO- d_6) δ 167.73, 167.39, 161.74, 161.65, 158.66, 158.35, 154.78, 154.73, 149.38, 149.05, 148.95, 148.29, 145.78, 145.58, 142.80, 130.77, 128.86, 124.25, 123.64, 105.70, 105.64, 104.19, 103.61, 103.48, 56.33, 55.90, 47.43, 44.22, 43.77, 43.08, 42.16, 41.36. DUIS-MS calculated for $\text{C}_{21}\text{H}_{22}\text{N}_6\text{O}_5$ $[\text{M} + \text{H}]^+$: 438.17, found : 439.15

(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)(3-nitrophenyl)methanone

(5b): Light yellow solid, 80% yield for the step; ^1H -NMR (400MHz, DMSO- d_6) δ 8.33-8.31 (1H, d, $J = 8.2$ Hz), 8.27 (1H, s), 7.92-7.90 (1H, d, $J = 7.6$ Hz), 7.79-7.75(1H, t, $J = 15.9$ Hz) 7.15(2H, b), 6.75 (1H, s), 3.84 (4H, s), 3.80 (4H, s), 3.74 (3H, m), 3.40 (1H, m). ^{13}C -NMR (400MHz, DMSO- d_6) δ 167.39, 161.65, 158.78, 154.75, 149.22, 148.21, 145.58, 138.01, 133.92, 130.71, 124.72, 122.46, 105.73, 104.20, 103.50, 56.33, 55.89, 44.19, 42.23. DUIS-MS calculated for $\text{C}_{21}\text{H}_{22}\text{N}_6\text{O}_5$ $[\text{M} + \text{H}]^+$: 438.17, found : 439.10

(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)(2-nitrophenyl)methanone

(5c): Light yellow solid, 85% yield for the step; ^1H -NMR (400MHz, DMSO- d_6) δ 8.23-8.212, (1H, d, $J = 8.0$ Hz), 7.87 (1H, t, $J = 6.9$ Hz), 7.72 (1H, t, $J = 7.3$ Hz), 7.60-7.58 (1H, d, $J = 7$ Hz) 7.44 (1H, s), 7.17 (2H, b) 6.74 (1H, s), 3.83 (5H, s), 3.79 (4H, s), 3.72 (3H, m), 3.26 (2H, m)-NMR (400MHz, DMSO- d_6) δ 166.02, 161.65, 158.77, 154.72, 149.16, 145.91, 145.59, 133.36, 132.89, 130.77, 128.62, 125.23, 105.71, 104.15, 103.51, 56.31, 55.89, 46.95, 43.95, 43.65, 41.88. DUIS-MS calculated for $\text{C}_{21}\text{H}_{22}\text{N}_6\text{O}_5$ $[\text{M} + \text{H}]^+$: 438.17 found : 439.10

(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)(4-chloro-3-

nitrophenyl)methanone (5d): Light yellow solid, 72% yield for the step; ^1H -NMR (400MHz, DMSO- d_6) δ 8.18, (1H, s), 7.88-7.87 (1H, s), 7.80 (1H, s), 7.44 (1H, s) 7.17 (2H, b), 6.74 (1H, s), 3.83 (6H, s), 3.79 (5H, s), 3.73-3.60 (4H, m), 3.41 (2H, m). ^{13}C -NMR (400MHz, DMSO- d_6) δ 166.45, 161.63, 158.71, 154.72, 149.12, 148.03, 145.56, 136.69,

132.76, 132.39, 126.40, 124.90, 105.66, 104.12, 103.46, 56.30, 55.89, 47.55, 44.18, 43.71, 42.32. DUIS-MS calculated for $C_{21}H_{21}ClN_6O_5$ [M + H]⁺: 472.13, found : 472.95

(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)(2-chloro-4-nitrophenyl)methanone (5e): Light yellow solid, 79% yield for the step; ¹H-NMR (400MHz, DMSO-d₆) δ 8.42 (1H, s), 8.30-8.28 (1H, d, J= 7.5 Hz), 7.76-7.74 (1H, d, J= 7.8) 7.43 (1H, s), 7.17 (2H, b), 6.76-6.74 (1H, d, J= 7.0), 3.83 (4H, s), 3.79 (4H, s), 3.73 (3H, s), 3.22 (1H, s). ¹³C-NMR (400MHz, DMSO-d₆) δ 164.61, 161.64, 161.58, 159.09, 158.68, 154.72, 154.67, 149.33, 149.13, 148.60, 145.60, 145.37, 145.31, 130.86, 129.71, 125.15, 123.39, 105.70, 104.20, 104.12, 103.50, 103.39, 56.31, 55.89, 46.62, 44.21, 44.17, 43.76, 41.81. DUIS-MS calculated for $C_{21}H_{21}ClN_6O_5$ [M + H]⁺: 472.13, found : 473.00

(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)(4-methyl-3-nitrophenyl)methanone (5f): Light yellow solid, 83% yield for the step; ¹H-NMR (400MHz, DMSO-d₆) δ 8.05, (1H, s), 7.70 (1H, s), 7.60 (1H, s), 7.44 (1H, s) 7.16 (2H, b), 6.74 (1H, s), 3.83 (4H, s), 3.80 (4H, s), 3.74 (4H, m), 3.42 (2H, m), 2.56(3H, s). ¹³C-NMR (400MHz, DMSO-d₆) δ 167.37, 161.63, 158.76, 154.72, 149.28, 149.19, 145.55, 135.45, 134.52, 133.47, 132.12, 123.67, 105.70, 104.10, 103.48, 56.30, 55.88, 47.65, 44.19, 43.82, 42.23, 19.82. DUIS-MS calculated for $C_{22}H_{24}N_6O_5$ [M + H]⁺: 452.18, found : 453.15

Compounds 6a-6f was prepared as the following procedure.—Last step products with the nitro group were reduced by using NH₄Cl 5 molar equivalence and Zn 15 molar equivalence to the nitro compounds. This was monitored by TLC in DCM/MeOH until reaction was complete it was then stopped by using an Acid base extraction. Acid was added until all zinc was dissolved then filtered to remove any undissolved zinc or other contaminates. Follow by readjusting the pH to a basic 12 by adding sodium carbonate aqueous solution. This mixture was then extracted by DCM and ethyl acetate then was dried with MgSO₄ and washed with diethyl ether and hexane to give the corresponding compounds.

(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)(4-aminophenyl)methanone (6a): Light yellow solid, 78% yield for the step; ¹H-NMR (400MHz, DMSO-d₆) δ 7.43 (1H, s), 7.19-7.17 (3H, m), 6.74 (1H, s), 6.59 (1H, s) 6.56 (1H, d), 5.53 (2H, s), 3.83 (3H, s), 3.79 (3H, s), 3.76 (4H, s) 3.56 (4H, s). ¹³C-NMR (400MHz, DMSO-d₆) 170.55, 161.61, 158.82, 154.70, 150.99, 149.24, 145.48, 129.80, 122.44, 113.19, 105.70, 104.16, 103.44, 56.31, 55.88, 44.22. DUIS-MS calculated for $C_{21}H_{24}N_6O_3$ [M + H]⁺: 408.19, found : 409.15

(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)(3-aminophenyl)methanone (6b): Light yellow solid, 89% yield for the last step; ¹H-NMR (400MHz, DMSO-d₆) δ 7.43 (1H, s), 7.15 (2H, b), 7.10-7.06 (1H, t, J= 7.7Hz), 6.74 (1H, s), 6.64-6.2 (1H, d, J= 8.0Hz), 6.59 (1H, s), 6.53-6.51 (1H, d, J= 7.4Hz), 3.83 (3H, s), 3.79 (3H, s), 3.75 (4H, m), 3.62-3.59 (2H, t, J= 16.5Hz), 3.42 (2H, b). ¹³C-NMR (400MHz, DMSO-d₆) δ 170.30, 161.61, 158.80, 154.72, 149.20, 145.54, 137.13, 129.32, 115.25, 114.43, 112.63 105.71, 104.18, 103.47, 56.33, 55.89, 44.17. DUIS-MS calculated for $C_{21}H_{21}ClN_6O_5$ [M + H]⁺: 408.19, found : 409.05

(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)(2-aminophenyl)methanone (6c): Light yellow solid, 78% yield for the step; ¹H-NMR (400MHz, DMSO-d₆) δ 7.43 (1H, s), 7.15-7.11 (3H, m), 7.04 (1H, s), 6.74 (2H, s), 6.58 (1H, b), 5.19 (2H, s), 3.83 (4H, s), 3.79 (7H, s), 3.53 (4H, m). ¹³C-NMR (400MHz, DMSO-d₆) δ 169.28, 161.61, 158.81, 154.70, 149.22, 146.32, 145.50, 130.50, 128.29, 119.80, 116.01, 105.70, 104.14, 103.45, 56.31, 55.88, 44.20, 30.89. DUIS-MS calculated for C₂₁H₂₄N₆O₃ [M + H]⁺: 408.19, found : 409.15

(3-amino-4-chlorophenyl)(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)methanone (6d): Light yellow solid, 77% yield for the step; ¹H-NMR (400MHz, DMSO-d₆) δ 7.44 (1H, s), 7.25 (1H, s), 7.18, (2H, s), 6.84 (1H, s), 6.74 (1H, s), 6.57 (1H, s) 5.57 (2H, b), 3.83 (3H, s), 3.79 (5H, m), 3.64 (5H, m). ¹³C-NMR (400MHz, DMSO-d₆) δ 169.27, 161.62, 158.78, 154.69, 149.17, 145.52, 145.22, 135.92, 129.46, 118.40, 115.58, 114.29, 105.68, 104.12, 103.48, 56.30, 55.89, 47.59, 44.17, 42.17. DUIS-MS calculated for C₁₇H₂₃N₅O₃ [M + H]⁺: 442.15, found : 443.10

(4-amino-2-chlorophenyl)(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)methanone (6e): Light yellow solid, 72% yield for the step; ¹H-NMR (400MHz, DMSO-d₆) δ 7.43 (1H, s), 7.14, (2H, b), 7.01-6.99, (1H, d, *J* = 7.5 Hz), 6.73 (1H, s) 6.63 (1H, s), 6.55- 6.53 (1H, d, *J* = 7.5 Hz), 5.63 (2H, s) 3.83 (4H, s) 3.79 (5H, s), 3.65-3.60 (8H, m), 3.25 (2H, s), 3.17(2H, s) ¹³C-NMR (400MHz, DMSO-d₆) δ 167.25, 161.61, 158.77, 154.70, 151.04, 149.17, 145.54, 130.48, 129.40, 122.63, 113.56, 112.90, 105.70, 104.13, 103.47, 56.32, 55.90, 49.07, 46.91, 44.34, 43.94, 41.83. DUIS-MS calculated for C₁₉H₂₁N₅O₄ [M + H]⁺: 442.15, found : 443.05

(3-amino-4-methylphenyl)(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazin-1-yl)methanone (6f): Light yellow solid, 77% yield for the step; ¹H-NMR (400MHz, DMSO-d₆) δ 7.43 (1H, s), 7.14 (2H, b), 6.99-6.97 (1H, d, *J* = 7.7 Hz), 6.74 (1H, s), 6.67 (1H, s), 6.50-6.49 (1H, s, *J* = 3 Hz), 5.02 (2H, b) 3.83 (3H, s), 3.80 (4H, m), 3.723-3.40 (8H, m), 2.08(3H, s). ¹³C-NMR (400MHz, DMSO-d₆) δ 170.41, 161.65, 154.82, 147.06, 145.76, 134.53, 130.16, 123.04, 1114.97, 112.95, 104.51, 103.31, 56.45, 56.10, 55.98, 44.37, 17.80. DUIS-MS calculated for C₂₂H₂₆N₆O₃ [M + H]⁺: 422.21, found : 423.10

Compounds 7aa-7fc were prepared as the following procedure.—The corresponding reduced benzamides 6a-6f were mixed with different dichloride linkers. They were dissolved in anhydrous DMF in a 2 to 1 ratio under argon protection. The dichloride linkers were diluted in anhydrous DMF and added slowly. It was stirred overnight and monitored by TLC after 24 hours. The reaction was stopped by reducing down the solvent and purified with PTLC in DCM/EtOH.

N¹,N⁵-bis(4-(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazine-1-carbonyl)phenyl)glutaramide (7ab): Light yellow solid, 70% yield for the last step; ¹H-NMR (400MHz, DMSO-d₆) 10.15 (2H, s), 7.71-7.69 (4H, m), 7.44-7.40 (7H, d), 7.16 (6H, m), 6.74 (3H, d), 3.83 (10H, m), 3.79 (13H, m), 3.56 (9H, m) 2.44 (4H, s) 1.93 (2H, s). ¹³C-NMR (400MHz, DMSO-d₆) δ 171.61, 169.51, 161.62, 158.78, 154.70, 154.52, 149.20, 145.51, 140.99, 130.58, 128.94, 128.60, 120.88, 118.94, 105.69, 104.15, 103.45, 56.31,

55.88, 44.17, 36.06, 21.29. DUIS-MS calculated for $C_{47}H_{52}N_{12}O_8$ $[M + H]^+$: 912.40, found : 913.20

2,2'-oxybis(N-(4-(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazine-1-carbonyl)phenyl)acetamid e)(7ac): Light yellow solid, 70% yield for the last step; 1H -NMR (400MHz, DMSO- d_6) δ 10.27 (2H, s), 7.78 (2H, s), 7.77 (2H, s), 7.47 (2H, s), 7.45 (2H, s) 7.43 (2H, s), 7.15 (4H, m) , 6.74 (2H, s), 4.33 (4H, s) 3.83(10H, s), 3.79 (11H, m) 3.61-3.56 (8H, m). ^{13}C -NMR (400MHz, DMSO- d_6) δ 169.40, 168.88, 161.62, 158.77, 154.71, 149.18, 145.52, 139.99, 131.34, 128.64, 119.61, 105.68, 104.15, 103.45, 71.33, 56.31, 55.88, 44.13. DUIS-MS calculated for $C_{46}H_{50}N_{12}O_9$ $[M + H]^+$: 914.38, found : 915.15

N^1, N^4 -bis(3-(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazine-1-carbonyl)phenyl)succinamide (7ba): Light yellow solid, 70% yield for the last step; 1H -NMR (400MHz, DMSO- d_6) δ 10.32 (2H, s), 8.55 (4H, s), 7.83(2H, s), 7.70 (2H, s) 7.64-7.62 (2H, d, $J = 7.0$ Hz), 7.42-7.38(2H, t, $J = 14$ Hz), 7.12-7.10 (2H, d, $J = 7.8$ Hz), 3.92 (8H, m), 3.86 (8H, s), 3.84 (7H, s) 3.76 (4H, m) 3.53 (5H, m) 2.71 (4H, s). ^{13}C -NMR (400MHz, DMSO- d_6) δ 171.18, 169.60, 161.74, 155.59, 147.08, 139.91, 129.38, 121.93, 120.57, 118.05, 105.29, 102.39, 56.68, 56.43, 45.00, 31.68. DUIS-MS calculated for $C_{46}H_{50}N_{12}O_8$ $[M + H]^+$: 898.39, found : 899.20

N^1, N^5 -bis(3-(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazine-1-carbonyl)phenyl)glutaramide (7bb): Light yellow solid, 70% yield for the last step; 1H -NMR (400MHz, DMSO- d_6) 10.09 (2H, s), 7.45 (2H, s), 7.64-7.62 (2H, m), 7.43 (2H, s) 7.40-7.38 (4H, m), 7.14-7.10(6H, m), 6.74 (2H, d), 3.92 (8H, m), 3.83 (9H, m), 3.79 (18H, m) 2.41 (4H, s) 1.93 (2H, s). ^{13}C -NMR (400MHz, DMSO- d_6) δ 171.57, 169.44, 161.62, 158.79, 154.72, 149.18, 145.55, 139.77, 136.78, 129.33, 121.98, 120.50, 118.15, 105.71, 104.15, 103.48, 56.31, 55.89, 47.55, 44.06, 36.03, 21.28. DUIS-MS calculated for $C_{47}H_{52}N_{12}O_8$ $[M + H]^+$: 912.40, found : 913.20

2,2'-oxybis(N-(3-(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazine-1-carbonyl)phenyl)acetamid e) (7bc): Light yellow solid, 73% yield for the last step; 1H -NMR (400MHz, DMSO- d_6) δ 10.26 (2H, s), 7.82 (2H, s), 7.74-7.73 (2H, d, $J = 6.7$ Hz), 7.44 (4H, m) 7.15 (5H, b), 6.74 (2H, d), 4.31 (3H, s) 3.82 (8H, s), 3.79 (9H, m). ^{13}C -NMR (400MHz, DMSO- d_6) δ 169.30, 168.82, 162.78, 161.62, 158.80, 154.71, 149.19, 145.55, 138.83, 129.44, 122.71, 121.16, 118.79, 105.71, 104.17, 103.49, 71.31, 56.32, 55.88, 47.64, 44.18, 42.26, 36.24, 31.25. DUIS-MS calculated for $C_{46}H_{50}N_{12}O_9$ $[M + H]^+$: 914.38, found : 915.10

ethane-1,2-diylbis((3-(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazine-1-carbonyl)phenyl)carbamate) (7bd): light yellow solid, 30% yield for the last step; 1H -NMR (400MHz, DMSO- d_6) δ 9.94 (2H, s), 7.57-7.54 (4H, m), 7.43 (3H, s), 7.37 (2H, s) 7.14 (5H, b), 7.07-7.05 (2H, d), 6.74 (3H, s) 4.37 (4H, s) 3.83 (11H, s), 3.79 (11H, m). ^{13}C -NMR (400MHz, DMSO- d_6) δ 169.38, 161.62, 158.78, 154.71, 153.81, 149.18, 145.54, 139.58, 136.91, 129.44, 121.52, 119.75, 117.30, 105.70, 104.15, 103.47, 63.27, 62.49,

56.31, 55.88, 47.56, 44.12, 42.18, 25.95. DUIS-MS calculated for $C_{46}H_{50}N_{12}O_{10}$ $[M + H]^+$: 930.38, found : 931.20

(ethane-1,2-diylbis(oxy))bis(ethane-2,1-diyl)bis((3-(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazine-1-carbonyl)phenyl)carbamate) (7bg): Light yellow solid, 23% yield for the last step; 1H -NMR (400MHz, DMSO- d_6) δ 9.92 (2H, s), 7.57-7.53 (4H, m), 7.43 (2H, s), 7.37-7.36 (2H, s) 7.14 (4H, b), 7.06-7.04 (2H, d), 6.74 (2H, s) 4.22 (3H, s) 3.83 (9H, s), 3.79 (1 OH, m), 3.67 (4H, s), 3.59 (4H, s). ^{13}C -NMR (400MHz, DMSO- d_6) δ 169.41, 162.77, 161.62, 158.78, 154.70, 153.96, 149.19, 145.53, 139.70, 136.90, 129.41, 121.38, 119.61, 117.15, 105.71, 104.14, 103.47, 70.18, 69.15, 64.12, 60.69, 56.31, 55.88, 47.64, 44.17, 42.24, 36.24, 31.25. DUIS-MS calculated for $C_{50}H_{58}N_{12}O_{12}$ $[M + H]^+$: 1018.43, found : 1019.20

2,2'-oxybis(N-(4-(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazine-1-carbonyl)-3-chlorophenyl) acetamide) (7ec): Light yellow solid, 43% yield for the last step; 1H -NMR (400MHz, DMSO- d_6) δ 10.32 (2H, s), 7.97 (2H, s), 7.68-7.66 (2H, d, $J = 8.0$ Hz), 7.44 (2H, s), 7.41-7.39 (2H, s), 7.18 (5H, m), 6.74 (3H, s), 4.33 (4H, s) 3.83 (13H, s), 3.75 (11H, s), 3.72 (11H, m) 3.24 (6H, m). ^{13}C -NMR (400MHz, DMSO- d_6) δ 169.09, 166.09, 161.63, 158.64, 154.74, 148.97, 145.60, 145.48, 140.35, 131.20, 129.89, 129.02, 120.15, 118.86, 105.61, 105.42, 104.27, 104.16, 103.48, 103.35, 71.13, 56.32, 55.90, 46.78, 44.31, 44.16, 43.88, 41.79. DUIS-MS calculated for $C_{46}H_{48}C_{12}N_{12}O_9$ $[M + H]^+$: 982.30, found : 983.10

2,2'-oxybis(N-(5-(4-(4-amino-6,7-dimethoxyquinazolin-2-yl)piperazine-1-carbonyl)-2-methylphenyl)acetamide) (7fc): Light yellow solid, 23% yield for the last step; 1H -NMR (400MHz, DMSO- d_6) δ 9.62 (2H, s), 7.60 (2H, s), 7.43 (2H, s), 7.34-7.32 (2H, d, $J = 7.0$ Hz), 7.21-7.19 (2H, d, $J = 7.6$ Hz), 7.15 (4H, b), 6.74 (2H, s), 4.33 (4H, s) 3.83 (9H, s), 3.79 (11H, s), 2.29 (6H, m) 1.36 (3H, s). ^{13}C -NMR (400MHz, DMSO- d_6) δ 169.16, 168.46, 161.62, 158.77, 154.71, 149.20, 145.52, 135.90, 134.08, 133.91, 130.91, 124.73, 124.29, 105.71, 104.15, 103.46, 71.09, 56.31, 55.88, 44.16, 34.85, 30.90, 18.09. DUIS-MS calculated for $C_{48}H_{54}N_{12}O_9$ $[M + H]^+$: 942.41, found : 943.20

4.2. LogP measurement

Compounds **3d**, **3e**, **7gb** and lead compound were used in the determination of their log P values. This method follows the OECD test guidelines for the slow stir method, with slight modification due to the data collection method. The data was collected on a UV-Vis spectrophotometer Shimadzu UV-1280 and standard UV Quartz 10 mm cuvette was used. The chemical was purchased from VWR brand EMPLURA 1-octanol 99% purity. The Flask where equipped with Teflon coated magnetic stirrers and Corning PC420D stir plates where used. The 1-octanol and DDH₂O where placed into two separate containers containing both phases to become mutual saturated by allowing them to stir for 48 h. Then the two phases where separated by a separation funnel. Compounds **3d**, **3e**, **7gb** and lead now were dissolved in 1-octanol pre-saturated with water. These stock solutions are then centrifuged at 5,000 rpm for 5 min. The supernatant was taken to be the working solution. Take 5 mL working solution of each into new clear flask, and add identical volume water saturated with 1-octanol, then stir at room temperature for 48 h. After that, stop stirring; take two-phase

solution out, centrifuge at 5,000 rpm for 5 min to separate this two-phase system. Removal of the 1-octanol phase from the solution then transferred for UV spectrophotometry to achieve absorbance at the correct wavelength. According to the procedure, concentrations were determined by UV measurement so, equation written as:

$$\text{Log P} = \log \frac{A_x}{A_0 - A_x}$$

Where A_0 and A_x are the initial and final absorbance values of organic layer.

HPLC-PDA analysis of the purity of the compounds.—Reversed-phase HPLC analysis of compounds were conducted on Beckman HPLC system with AutoSampler (table 6). The chromatographic separation was performed on a C18 column (2.0 mm × 150 mm, 5 μm) from Phenomenex (Torrance, CA, USA), one mobile phase was employed for isocratic elution with a flow rate of 0.2 mL/min. The injection volume was 6 μL and the PDA detector was set up 346nm

4.3. Biological studies

4.3.1. Cell Culture and Antibodies.—U251 cells were obtained from ATCC (Rockville, MD). The cells were maintained in Dulbecco's Modified Eagle Medium (DMEM) supplemented with 10% fetal bovine serum (FBS), 10 mg/ml L-Glutamine, 100 U/mL penicillin-streptomycin, and 0.1 mg/ml streptomycin. EphA2 overexpression cells were grown in the presence of 1 μg/ml puromycin. Cell cultures were grown at 37 °C, in a humidified atmosphere of 5% CO₂ in a Thermo CO₂ incubator (Grand Island, NY). Antibodies used included rabbit polyclonal antibodies against pEphA/B (Santa Cruz, Santa Cruz, CA), as well as mouse monoclonal antibody to EphA2 (clone D7, Millipore, Billerica, MA).

4.3.2. Mammalian cell viability analysis—The MTT assay was used to examine the effect of the compounds on the growth of U251 and U251 over expression EphA2 cells in four replications. 3000 cells per well were seeded with RPMI1640 medium in 96-well flat-bottomed plates for 24 h and were then exposed to various concentrations of test compounds dissolved in DMSO (final concentration 0.1%) in medium for 48 h. Controls received DMSO at a same concentration as that in drug-treated cells. Cells were incubated in 200 μL of 0.5 mg/mL of MTT reagent diluted in fresh media at 37°C for 3 h. Supernatants were removed from the wells, and the reduced MTT dye was solubilized with 200 μL/well DMSO. Absorbance at 570 nm was determined on a SpectraMax Plus384 spectrophotometer (Molecular Devices). Data obtained with quadruplication were normalized and fitted to a dose–response curve using GraphPad Prism v.5 (GraphPad).

4.3.3. EphA2 activation.—U251 EphA2-overexpressing cells were plated in 6-well dishes at a density of 100,000 cells/well and grown for 24 hours prior to stimulation with appropriate compounds for 30 min. 0.1% DMSO was used as vehicle control. Following treatment, cells were washed and lysed in modified RIPA Buffer (20 mM Tris-HCl pH 7.4, 20 mM NaF, 150 mM NaCl, 10% glycerol, 0.1% SDS, 0.5% DCA, 2 mM EDTA, 1% Triton

X-100, 2 mM Na₃VO₄, and protease inhibitors, including 1 mM phenylmethylsulphonyl fluoride, and 2 µg/ml each of aprotinin and leupeptin) for 20 min, followed by immunoprecipitation. Samples were boiled 5 min and run on 4-12% Bis-Tris Plus gels (Thermo Fisher), followed by transfer to Immobilon-P PVDF membranes (Millipore, Billerica, MA). Membranes were blocked 1 hr at room temperature in 3% BSA in TBS containing 0.05% Tween-20 (TBS-T) followed by overnight incubation with pEphA/B (1:500) and EphA2 (Santa Cruz) (1:1000) primary antibodies. Membranes were washed in TBS-T and incubated with goat anti-rabbit-HRP (1:5000) secondary antibody 1 hr at room temperature, followed by washing and developing with Luminol Reagent (Santa Cruz). Band intensities were quantified using ImageJ (NIH) to generate the figure.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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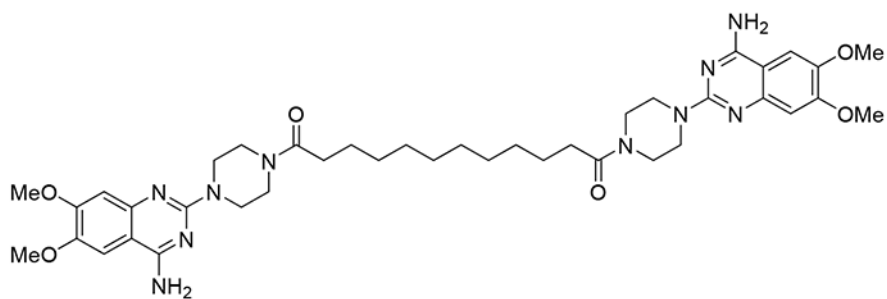


Figure 1.
EphA2 agonist with a dimeric structure

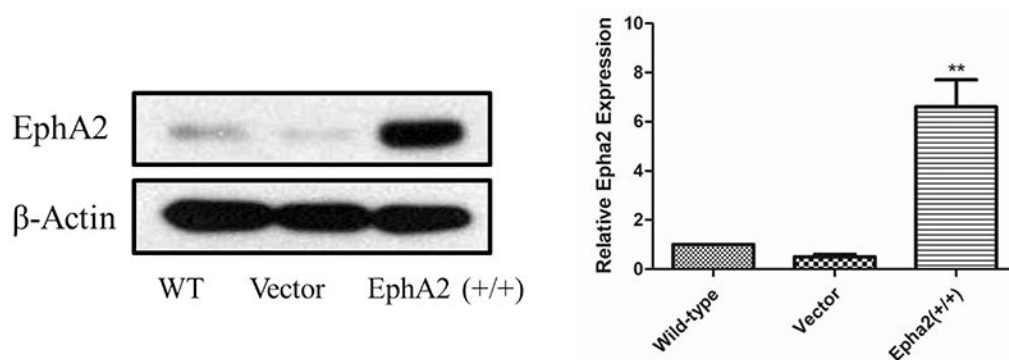


Figure 2.

Construction of U251/EphA2 cell line based on U251 wild type cells. The intensities of the bands were quantified using ImageJ (NIH) to generate the figure. The figures are representative of 3 experiments. ** $p < 0.001$ EphA2 vs. wild type by unpaired t -test.

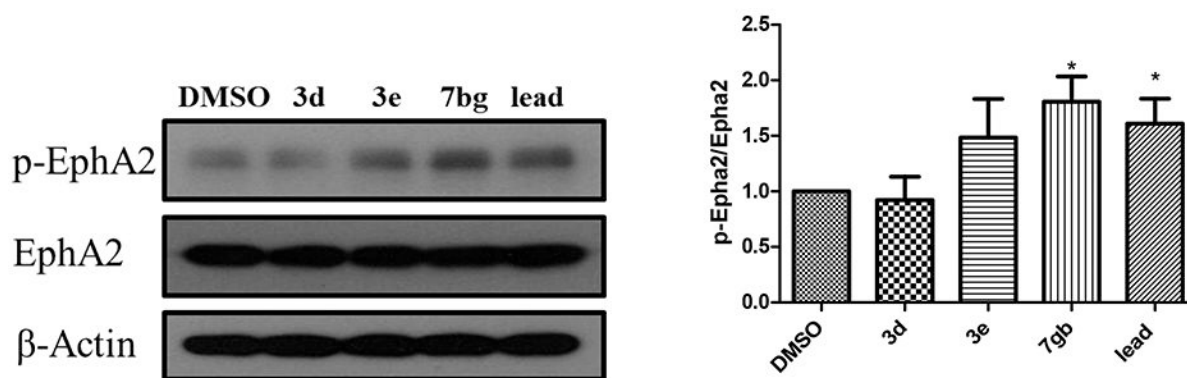
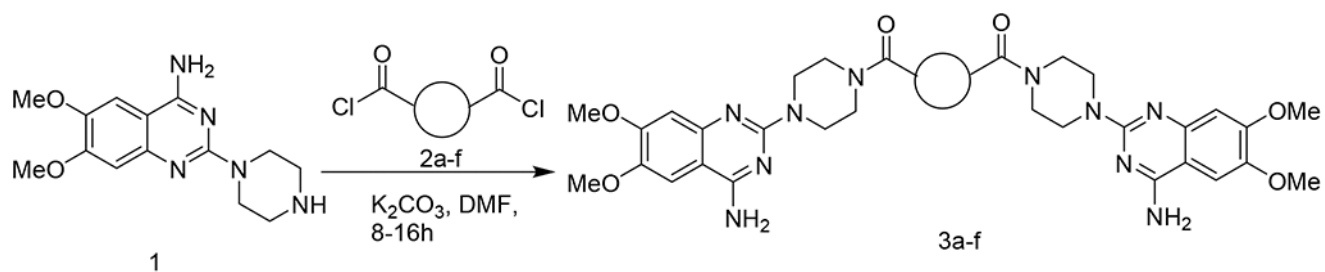
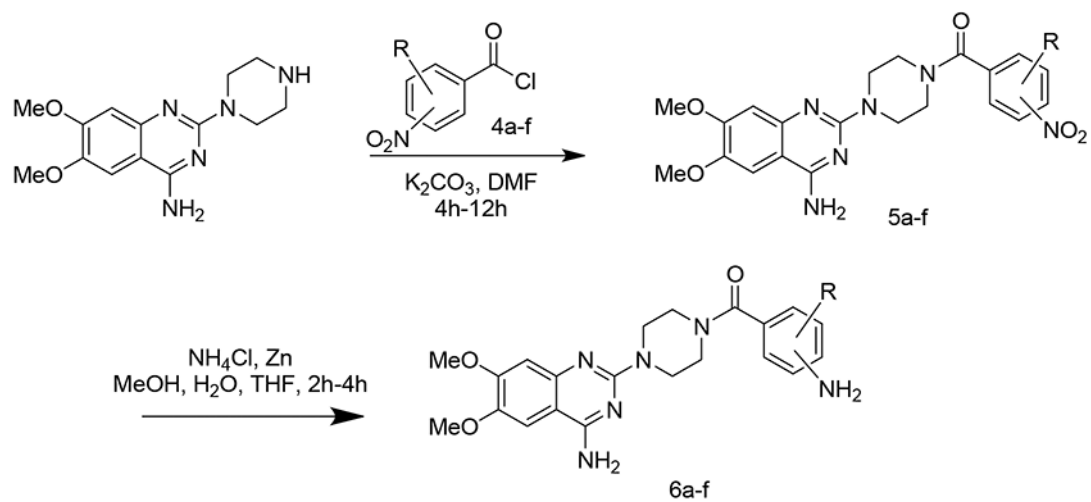


Figure 3.

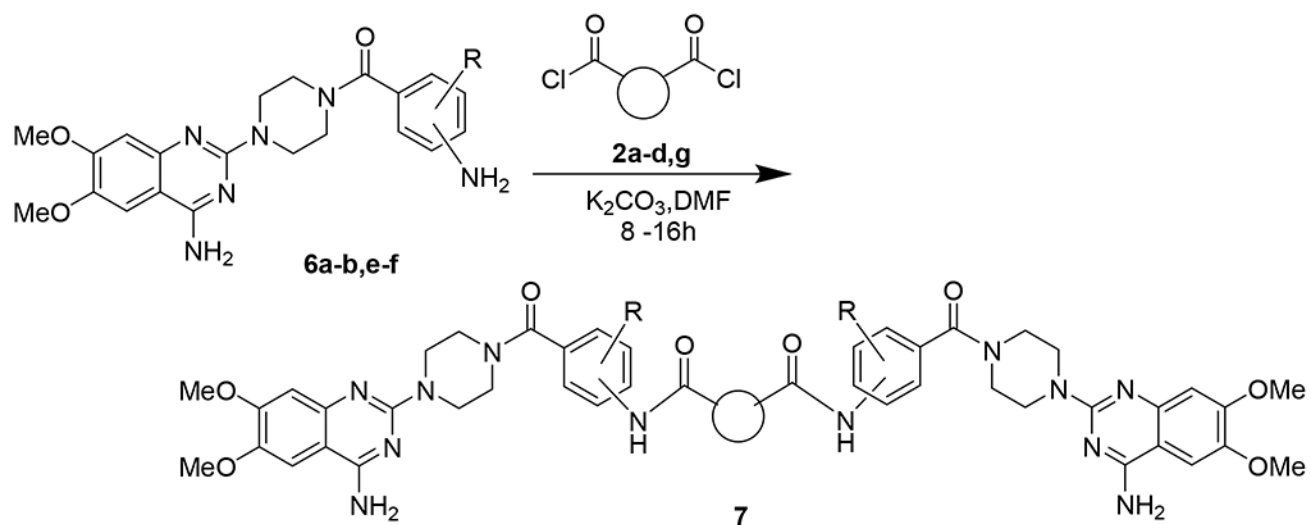
EphA2 activation of the new derivatives. U251 EphA2 cells were treated with the compounds with 2 μ M (in 0.1% DMSO) for 1hr and cell lysates were subject to immunoprecipitation and immunoblot for phosphorylated EphA/B kinases (pEphA/B) and total EphA2. Treatment with 2 μ M of lead was served as positive control. Treatment with 0.1% DMSO was served as vehicle control. The band intensities of the bands were quantified using ImageJ (NIH) to generate the figure. The figures are representative of 3 experiments. * $p < 0.05$ treatment vs. DMSO by unpaired t -test.

**Scheme 1.**

Synthesis of dimers based on 6,7-dimethoxy-2-(piperazin-1-yl)quinazolin-4-amine as the monomer

**Scheme 2.**

Addition of acyl chlorides to produce a small set of monomeric compounds

**Scheme 3.**

The monomeric compounds from scheme 2 introduced to the linkers from scheme 1.

Table 1.

compound list of the original core with the new linkers.

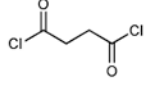
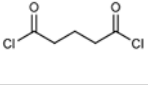
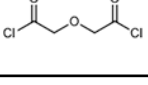
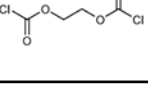
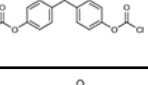
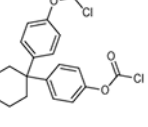
Entry	Addition of linkers: 2a-f	% yield of 3a-f	Molecular weight 3a-f
1	2a: 	80%	660.72
2	2b: 	80%	674.75
3	2c: 	57%	676.72
4	2d: 	43%	692.72
5	2e: 	31%	858.94
6	2f: 	31%	899.00

Table 2.

compound bearing the new aromatic core as monomers.

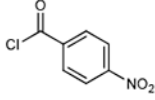
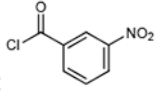
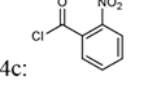
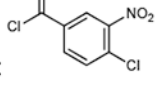
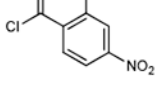
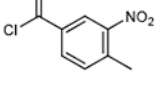
Entry	Addition of acyl chlorides: 4a-f	% yield of 5a-f	% yield of 6a-f	Molecular weight 5a-f	Molecular weight 6a-f
1	4a: 	87%	78%	438.44	408.45
2	4b: 	87%	89%	438.44	408.45
3	4c: 	85%	78%	438.44	408.45
4	4d: 	72%	77%	472.88	472.15
5	4e: 	79%	72%	274.88	442.90
6	4f: 	83%	77%	452.46	422.48

Table 3.

Dimeric compounds bearing the aromatic core with the new linkers.

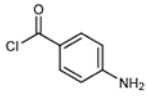
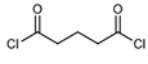
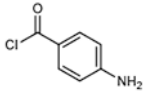
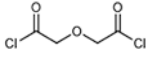
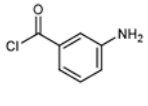
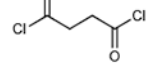
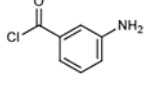
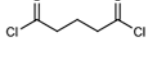
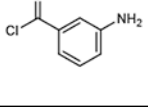
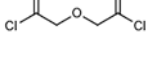
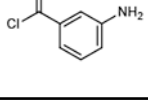
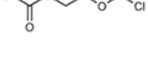
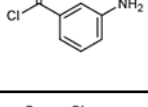
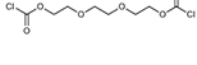
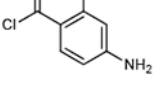
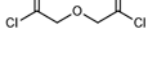
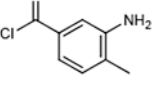
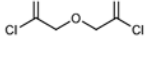
Entry	Acyl chloride from 6	Addition of linker 2a-d,g	% yield of 7a-d, g	Molecular weight
1	6a: 	2b: 	7ab: 70%	912.99
2	6a: 	2c: 	7ac: 70%	914.96
3	6b: 	2a: 	7ba: 73%	898.96
4	6b: 	2b: 	7bb: 30%	912.99
5	6b: 	2c: 	7bc: 23%	914.96
6	6b: 	2d: 	7bd: 70%	930.96
7	6b: 	2g: 	7bg: 70%	1019.07
8	6e: 	2c: 	7ec: 43%	983.85
9	6f: 	2c: 	7fc: 23%	943.02

Table 4.

Inhibition of the cell proliferation of U251/U251 EphA2 cells.

Comp	U251 EphA2 overexpressed* (μM)	U251 Wild type* (μM)	Selectivity index wild/overexpressed**
Lead	2.1 \pm 1.05	5.2 \pm 2.56	2.5 \pm 2.44
3a	6.75 \pm 2.40	12.62 \pm 6.62	1.9 \pm 1.18
3b	7.47 \pm 3.98	8.07 \pm 4.37	1.1 \pm 0.82
3c	51.86 \pm 26.07	160 \pm 80.62	3.1 \pm 2.20
3d	1.60 \pm 0.40	3.05 \pm 0.82	1.9 \pm 0.70
3e	2.45 \pm 1.11	9.22 \pm 5.56	3.8 \pm 2.84
3f	3.18 \pm 1.44	7.48 \pm 4.03	2.4 \pm 1.66
5a	12.70 \pm 8.72	12.23 \pm 8.35	1.0 \pm .93
5b	27.41 \pm 15.05	24.80 \pm 16.01	0.9 \pm .77
5c	7.41 \pm 2.98	8.34 \pm 4.42	1.1 \pm .75
5d	12.06 \pm 5.98	14.73 \pm 9.20	1.2 \pm .97
5e	26.43 \pm 11.80	53.01 \pm 29.78	2.0 \pm 1.44
5f	4.41 \pm 2.36	10.77 \pm 7.23	2.4 \pm 2.10
6a	11.14 \pm 7.26	8.98 \pm 4.61	0.8 \pm 0.67
6b	16.50 \pm 9.16	8.84 \pm 4.36	0.5 \pm 0.40
6c	14.60 \pm 8.67	35.29 \pm 21.98	2.4 \pm 2.10
6d	5.15 \pm 3.01	17.91 \pm 12.66	3.5 \pm 3.19
6e	3.00 \pm 1.26	16.49 \pm 10.56	5.5 \pm 4.21
6f	2.70 \pm 0.55	47.94 \pm 21.13	17.8 \pm 8.62
7ab	27.99 \pm 14.87	45.72 \pm 34.86	1.6 \pm 1.52
7ac	30.44 \pm 12.63	174.01 \pm 129.21	5.7 \pm 4.86
7ba	41.41 \pm 25.89	47.86 \pm 26.93	1.2 \pm 0.97
7bb	122.30 \pm 59.31	128.10 \pm 53.59	1.0 \pm 0.67
7bc	9.92 \pm 6.20	8.11 \pm 4.42	0.8 \pm 0.68
7bd	4.99 \pm 2.32	10.12 \pm 6.11	2.0 \pm 1.55
7bg	1.90 \pm 0.55	7.91 \pm 2.28	4.2 \pm 1.70
7ec	3.41 \pm 1.50	2.59 \pm 1.24	0.8 \pm 0.49
7fc	5.66 \pm 2.94	11.69 \pm 7.36	2.1 \pm 1.69

The area shaded in gray represent the monomer compounds.

* IC50 values are reported in μM . included Mean \pm SD.(n=4)

** Selectivity index is calculated by taking the wild type average/Overexpressed average included is the error of propagation

Table 5.

The measurement was repeated three times with water and 1-octanol as the two solvents, and the results are presented as Mean±SD.

Compounds	Log P (Mean±SD)
3d	1.28±0.03
3e	0.75±0.01
7bg	0.78±0.03
Lead comp	0.71±0.02

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Table 6.

Purity of the compounds

H₂O/MeOH (20%/80%)		
Compound	Retention time (R_t) in minutes	Purity
3a	1.90	99.99%
3b	1.90	99.99%
3c	1.98	97.59%
3d	1.88	99.99%
3e	1.86	99.61%
3f	1.92	90.71%
5a	1.75	99.97%
5b	1.76	99.95%
5c	1.76	99.91%
5d	1.76	99.90%
5e	1.75	99.75%
5f	1.76	99.95%
6a	1.74	99.95%
6b	1.75	99.95%
6c	1.75	99.94%
6d	1.75	99.77%
6e	1.78	99.54%
6f	1.74	97.79%
7ab	1.79	99.29%
7ac	1.82	98.93%
7ba	1.86	98.95%
7bb	1.86	98.88%
7bc	1.84	97.08%
7bd	1.85	97.23%
7bg	1.83	98.15%
7ec	1.84	98.01%
7fc	1.85	99.44%