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## Free-Floating 2D Nanosheets with a Superlattice Assembled from Fe<sub>3</sub>O<sub>4</sub> Nanoparticles for Peroxidase-Mimicking Activity

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### Abstract

The organization of nanoparticles (NPs) with controlled chemical composition and size distribution into well-defined sheets will find many practical applications, but the chemistry remains problematic. Therefore, we report a facile method to assemble NPs to free-floating two-dimensional (2D) nanosheets with a superlattice and thicknesses reaching 22.8 nm. The ligand oleic acid is critical in the formation of nanosheets. As assembled, these free-floating 2D nanosheets remain intact in both polar and nonpolar solvents, e.g., deionized water, ethanol, *N,N*-dimethylformamide, dimethyl sulfoxide, toluene, hexane, and chloroform, without any

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Supporting Information

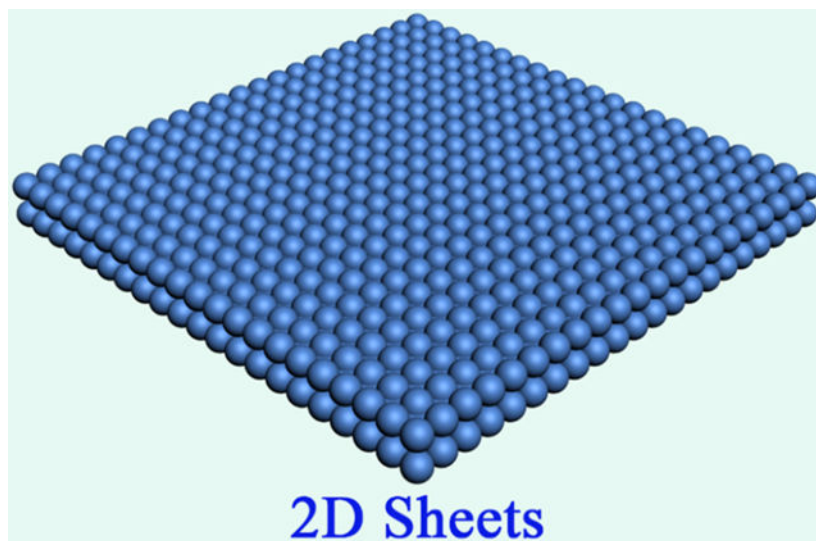
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Characterization of Fe<sub>3</sub>O<sub>4</sub> NPs and Fe<sub>3</sub>O<sub>4</sub> NP-micelles, stability of 2D nanosheets, FTIR and <sup>1</sup>H NMR spectra, TEM image of hybrid 2D nanosheets, and comparisons of the kinetic parameters of Fe<sub>3</sub>O<sub>4</sub> nanosheets and Fe<sub>3</sub>O<sub>4</sub> NPs (PDF)

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disassembly. Compared to  $\text{Fe}_3\text{O}_4$  NP building blocks, these 2D nanosheets show more favorable catalytic properties and enhanced catalytic reactivity, which can be exploited to mimic natural enzymes. Our work is expected to open up a new avenue for synthesizing free-floating 2D supersheets by NP assembly, leading to a new generation of materials with enriched functions and broader applications.

## Graphical Abstract



## Keywords

superlattice; nanosheets; nanoparticles; self-assembly; catalysis

The assembly of nanoparticles (NPs) into higher-order architectures, e.g., one-, two-, or three-dimensional (1D, 2D, or 3D) superstructures, can result in materials with uniquely or synergistically enhanced properties,<sup>1-4</sup> and such materials have been used in many fields, including catalysis,<sup>5</sup> biomedical diagnosis,<sup>6</sup> sensors,<sup>7</sup> magnetic resonance imaging,<sup>8</sup> plasmonics,<sup>9</sup> and drug delivery.<sup>10</sup> Accordingly, various methods have been developed to fabricate assembled structures of different types and with different properties.<sup>11,12</sup> For example, binary superlattices prepared by the coassembly of PbTe and  $\text{Ag}_2\text{Te}$  NPs show higher electrical conductivity compared to those of each individual component (i.e., PbTe or  $\text{Ag}_2\text{Te}$  NPs).<sup>13</sup> An interfacial self-assembly approach was explored to prepare 2D metal chalcogenide semiconductors with a honeycomb superlattice through the oriented attachment of nanocrystals,<sup>14</sup> and a facile bottom-up process was employed to fabricate binary nanocrystal superlattices by the slow evaporation of colloidal dispersions on tilted substrates.<sup>15</sup> Through a reel-to-reel-compatible large-area-coating technique, diverse single-component and binary superlattices were formed by depositing colloidal nanocrystals onto various substrates.<sup>16</sup> Among these works, 2D structures by aligning NPs on specific substrates, or templates, have been reported,<sup>17-19</sup> but it remains a formidable challenge to achieve free-floating 2D sheets from NP assembly, e.g., stable 2D nanosheets with a superlattice, by a template/substrate-free assembly process.

Colloidal assembly is a well-known strategy to achieve hierarchical structure in two or three dimensions by manipulating NPs in solution.<sup>20–22</sup> On the basis of this strategy, we developed a facile route to fabricate free-floating 2D nanosheets with a superlattice and long-range ordering up to several micrometers (Scheme 1). The thicknesses of these nanosheets are around 22.8 nm when piled up by approximately three layers of NPs. Oleic acid (OA), one of the aliphatic ligands used to stabilize Fe<sub>3</sub>O<sub>4</sub> NPs, is critical for the formation of well-defined nanosheets. It is noteworthy that these 2D nanosheets are stable and remain intact in both polar and nonpolar solvents. Compared to the original Fe<sub>3</sub>O<sub>4</sub> NP building blocks, these 2D nanosheets show more favorable catalytic properties and enhanced catalytic reactivity, which could be quite promising for application in mimicking natural enzymes.

OA-stabilized Fe<sub>3</sub>O<sub>4</sub> NPs (size ~7 nm; Figure S1) were prepared according to a previously reported method.<sup>23</sup> Then, an aqueous solution of cetyltrimethylammonium bromide (CTAB) was added to the above NP solution with vigorous stirring. A subsequent heating process evaporated the chloroform and transferred the NPs into the aqueous phase, forming an NP–micelle solution; then, the mixed solvent [diethylene glycol (DEG) and poly(vinylpyrrolidone) (PVP)] was injected into the as-prepared micelle solution. After the solution was held at 83 °C for 2 h, assembly of the NPs occurred. Finally, these products were isolated by centrifugation and redispersed in ethanol. Transmission electron microscopy (TEM) shows that as-prepared products are 2D nanosheets with lateral dimensions up to several micrometers (Figure 1a). These 2D nanosheets were washed, isolated by centrifugation, and redispersed in different solutions, which display the free-floating feature (Figure S2). The enlarged TEM image of one typical sheet in Figure 1b,c further reveals that it is composed of well-aligned NPs and shows a high degree of crystallinity (superlattice) with sharp electron diffraction spots (inset in Figure 1c). The thicknesses of 2D nanosheets were then measured by atomic force microscopy (AFM). The thickness of a typical sheet is  $22.8 \pm 0.5$  nm, corresponding to approximately three layers of NPs (Figure 1d). The nanosheet exhibits on-axis superlattice–fringe patterns related to a face-centered-cubic (fcc) superlattice structure (Figure 1b,c), which was further investigated by an ensemble measurement of these nanosheets using small-angle X-ray scattering (SAXS). The SAXS data exhibit two distinguishable peaks that could be indexed as the (110) and (221) Bragg diffractions of an fcc structure with a lattice constant (*a*) of  $8.7 \pm 0.2$  nm (Figure 1e), which is consistent with the value of  $8.8 \pm 0.2$  nm obtained from the TEM images (Figure 1b,c). As confirmed by our experiments, these assembled nanosheets display excellent stability in different solvents, both polar and nonpolar, e.g. deionized water, ethanol, *N,N*-dimethylformamide, dimethyl sulfoxide, toluene, hexane, and chloroform (Figure S2).

For a better understanding of the growth mechanism of the nanosheets, reaction parameters were systematically tuned, and TEM was used to investigate the formation of these products. First, PVP and DEG work as a mixed solvent in this system. For instance, without PVP, no assembled product was collected. Second, CTAB works as a “wrapping” agent to form NP–micelles because the amphiphilic structure of CTAB mediated the interaction (dispersion) between NPs in the mixed solvent. Without CTAB, only irregular aggregates were formed (Figure S3). Third, OA is a very critical factor. Without OA as the ligand, Fe<sub>3</sub>O<sub>4</sub> NPs easily

aggregated into bulk materials after injection into the mixed solvent (DEG and PVP; Figure 2b).<sup>24</sup> When the amount of OA was increased to 30  $\mu\text{L}$ , monodispersed 3D particles composed of NPs were formed (Figure 2c). A further increase of OA to 35  $\mu\text{L}$  resulted in the formation of semitransparent particles/vesicles (Figure 2d), which later merged together when 40  $\mu\text{L}$  of OA was added (Figure S4b). With more OA added (50  $\mu\text{L}$ ), surface wrinkles, a characteristic feature of 2D sheets, could be clearly observed (Figure 2e). Finally, well-defined 2D sheets were achieved when 60  $\mu\text{L}$  of OA was used (Figure 1). These results demonstrate that OA plays a critical role in avoiding the aggregation of large particles (Figure 2b), guiding the assembly of NPs to form monodispersed 3D particles (Figure 2c), and, finally, layering them into 2D sheets (Figure 2e,f).

On the basis of the above observations, we propose the following formation mechanism for free-floating 2D nanosheets with a superlattice (Figures 3 and 4). Prior to the assembly,  $\text{Fe}_3\text{O}_4$  NP–micelles adopt an interdigitated bilayer structure with OA ligands as the inner layer and CTAB as the outer layer (Figure 3). This results from hydrophobic van der Waals interactions between the aliphatic chains of OA and CTAB.<sup>25</sup> These NP–micelles are uniformly dispersed in a mixed solvent (DEG and PVP), owing to the amphiphilic nature of CTAB (Figure 4a). During the heating process (83 °C and 2 h), with subsequent evaporation of chloroform, the CTAB layer (outer layer, Figure 3b) detaches from the surface of  $\text{Fe}_3\text{O}_4$  NP–micelles because of its increased solubility in the mixed solvent upon heating, leaving OA-capped NPs “wrapping” inside the solvent (Figure 4b).  $^1\text{H}$  NMR and FTIR spectra confirmed the self-dissociation of the CTAB layer and the presence of OA (Figures S5 and S6). When the OA volume is less than 30  $\mu\text{L}$  (Figure 2f), a few OA molecules adsorb onto the surface of the  $\text{Fe}_3\text{O}_4$  NPs (Figure 3a),<sup>26</sup> and these NPs tend to assemble into large particles (Figure 2b) because of the strong van der Waals attraction among NPs. The addition of more OA (35–40  $\mu\text{L}$ ) promotes the dispersion of NPs inside the mixed solvent, and the distance between NPs gradually increases, resulting in the swelling of 3D particles, as shown in Figure 4c, and, in turn, forming semitransparent vesicles, as shown in Figure 2d and S4b. A further increase of OA (50–60  $\mu\text{L}$ ) results in breakage of the particle vesicles, and free-floating nanosheets (Figures 2e and 1), namely, fragments of the broken vesicles, are formed. It is worth noting that the redundant OA adsorbed on the surface of the NPs (Figure 3a)<sup>27</sup> acts as a “lubricating agent” to alleviate strong inter-NP attractions during the rearrangement process (Figure 4c).<sup>28,29</sup>

As a demonstration of one potential application for 2D nanosheets, we tested the peroxidase-mimicking activities of  $\text{Fe}_3\text{O}_4$  nanosheets, using 3,3',5,5'-tetramethylbenzidine (TMB) and hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) as substrates. After the addition of  $\text{Fe}_3\text{O}_4$  nanosheets to the TMB– $\text{H}_2\text{O}_2$  solution (pH 4.5), the solution changed from colorless to deep blue within 5 min at room temperature, as shown by the photograph and UV/vis absorption curves in Figure 5a,b. The optimum pH and temperature for catalysis of  $\text{Fe}_3\text{O}_4$  nanosheets are approximately 4.5 and 25 °C, respectively, which are very close to the values of the natural enzyme horseradish peroxidase (HRP; Figure 5c,d).<sup>30</sup> The data in Figure 5e-h confirm that the catalytic activity of the  $\text{Fe}_3\text{O}_4$  nanosheets is better than that of their  $\text{Fe}_3\text{O}_4$  NP building blocks. As further revealed by the kinetic parameters presented in Table S1,  $K_m$  (Michaelis constant) is an indicator of the enzymatic affinity to the substrate, with a low  $K_m$  representing strong affinity and vice versa.<sup>31</sup> With  $\text{H}_2\text{O}_2$  as the substrate, the apparent  $K_m$  value is about

20 times lower than that with HRP as the substrate,<sup>30</sup> and the initial velocity ( $V_{\max}$ ) is about 2 times greater, indicating a better affinity between  $H_2O_2$  and  $Fe_3O_4$  nanosheets compared to that of HRP. This can be ascribed to the collective property of the building blocks, leading to more active sites for  $H_2O_2$  interaction (Table S1),<sup>32</sup> which, in turn, results in lower  $K_m$  and higher  $V_{\max}$ . These results show that the 2D nanosheets could serve as a better artificial enzyme system than  $Fe_3O_4$  NPs or the natural enzyme HRP.

In conclusion, we have developed a template-free assembly route to fabricate free-floating 2D nanosheets with a superlattice from NPs. By fine-tuning aliphatic ligands, NPs can be effectively assembled into 2D nanosheets with lateral dimensions of several micrometers and thicknesses reaching 22.8 nm, and they display distinct superlattice structures. As a proof of concept, an artificial enzyme system composed of 2D nanosheets exhibits higher catalytic peroxidase activity compared to that of their building blocks and demonstrates functional mimicry of a natural enzyme. As revealed by these results, the flexible fabrication of free-floating 2D nanosheets from simple building blocks can be achieved. The success of this work will open a new avenue toward the design of stable and free-floating nanosheets, which will broaden the application of a new generation of functional materials.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## ACKNOWLEDGMENTS

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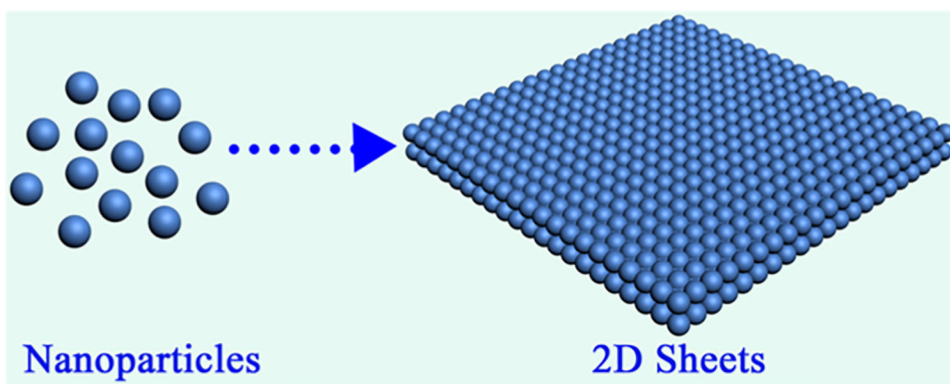
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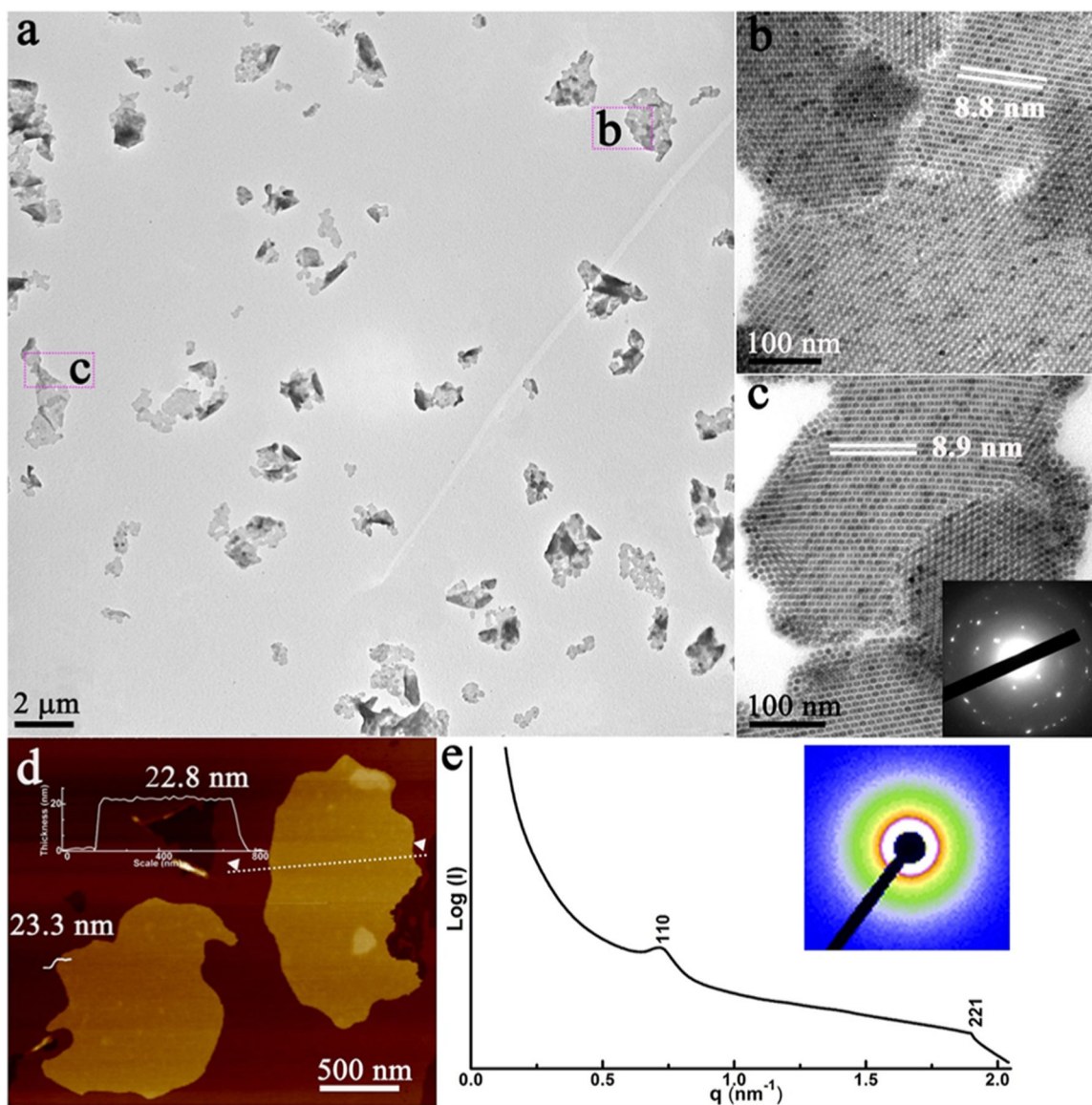
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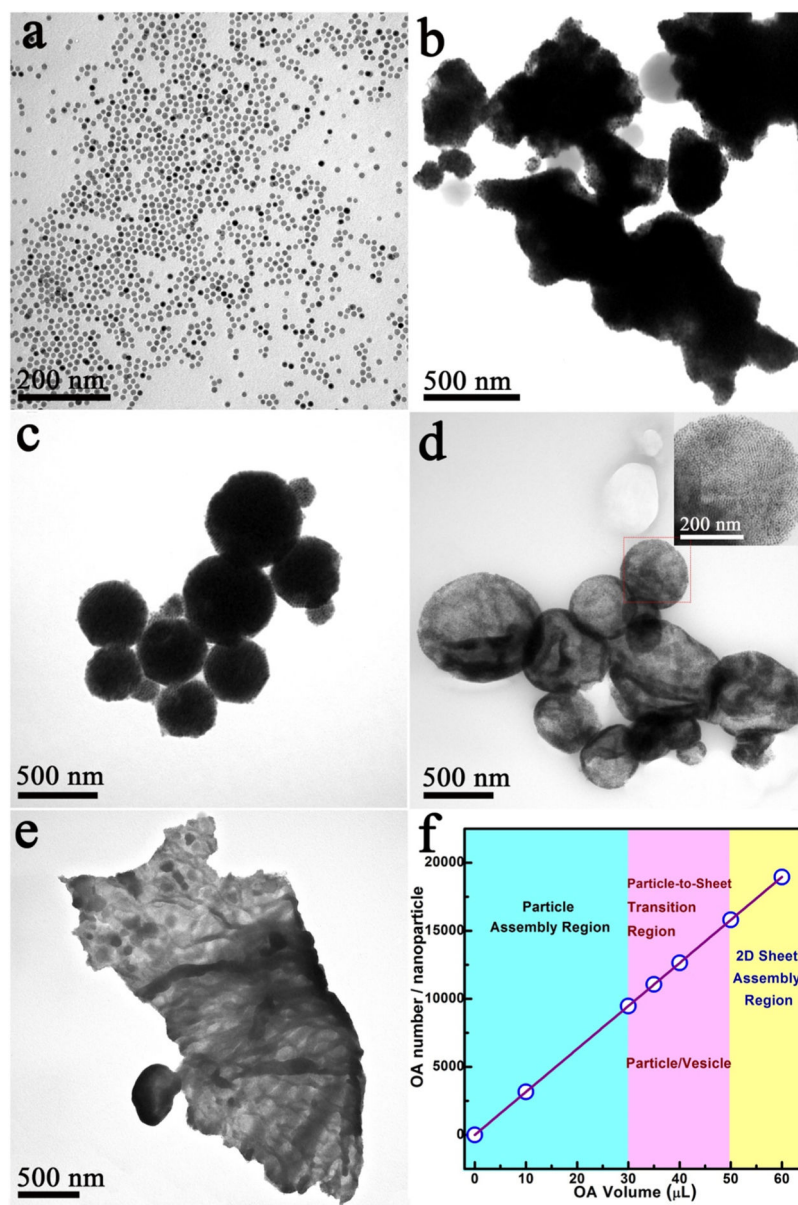
**Scheme 1.**

Formation of Free-Floating 2D Nanosheets through the Self-Assembly of NPs

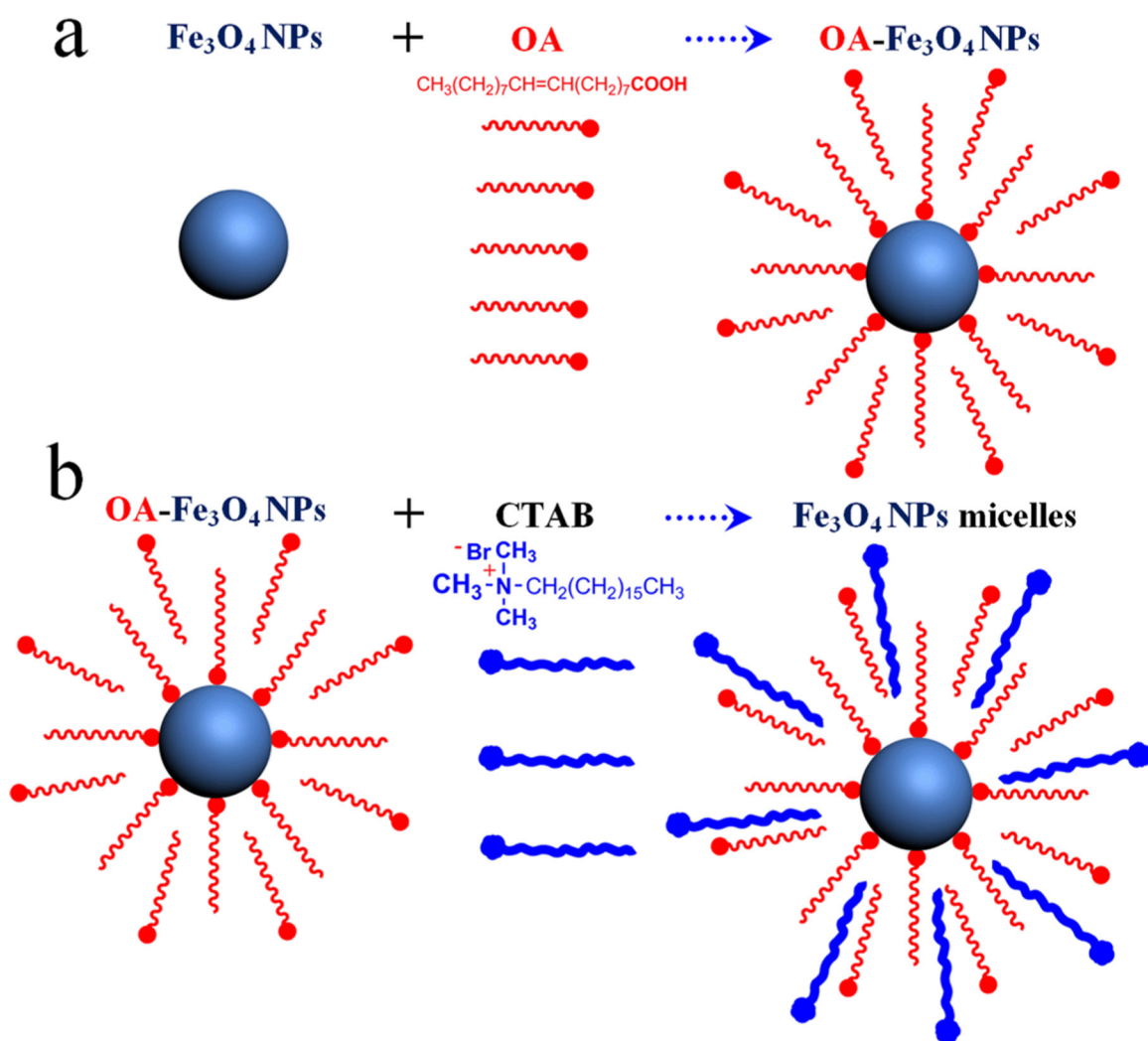




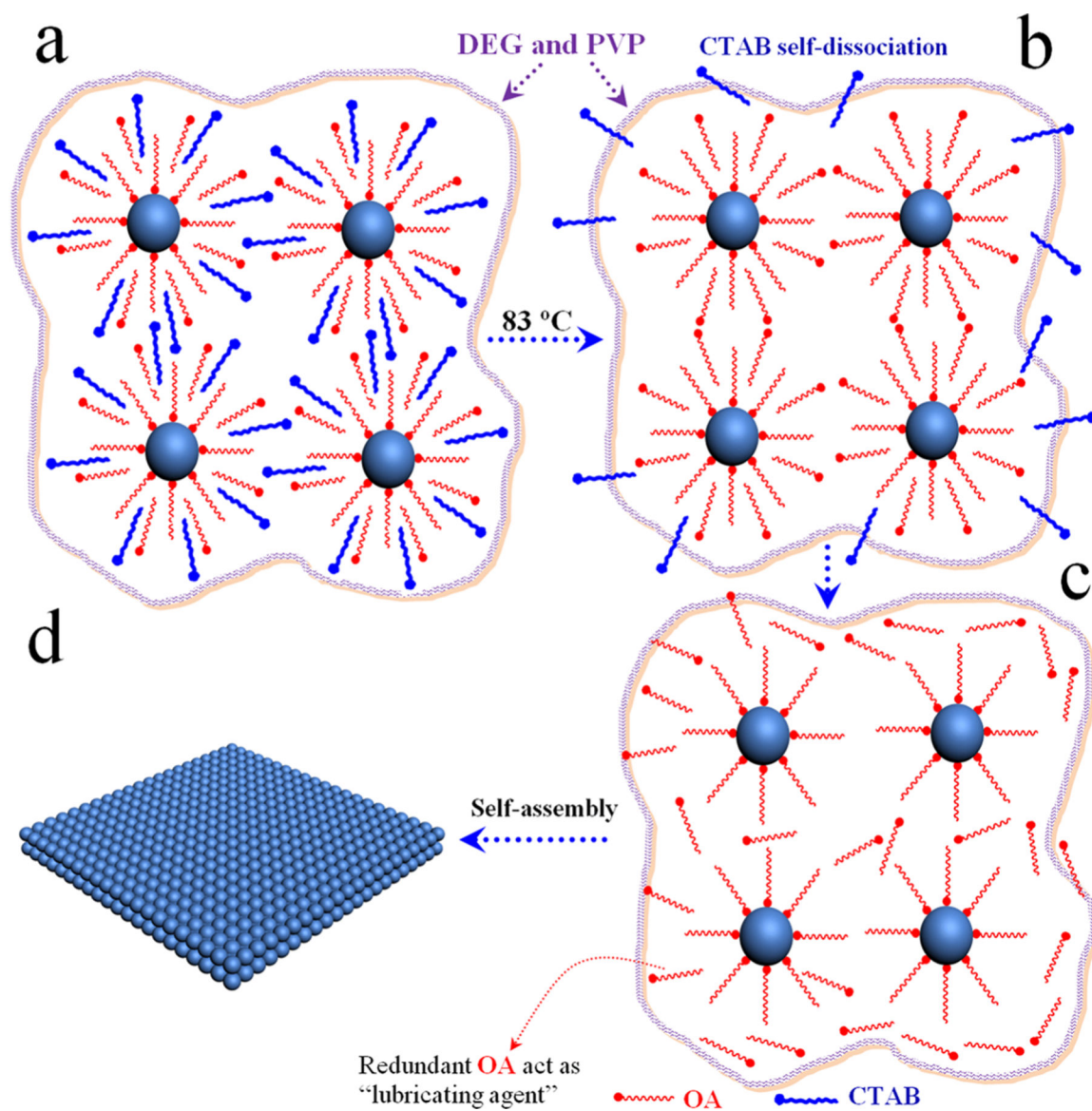
**Figure 1.** Characterization of free-floating 2D nanosheets: (a) low- and (b and c) high-magnification TEM images and electron diffraction pattern (inset); (d) AFM image; (e) SAXS pattern (inset) of the 2D nanosheets, where the color scale indicates the intensity.



**Figure 2.** TEM images of the assembly process for free-floating 2D nanosheet formation. Samples prepared from NPs assembled by adding different amounts of OA to guide the assembly process: (a) Fe<sub>3</sub>O<sub>4</sub> NPs; (b) 0 μL; (c) 30 μL; (d) 35 μL; (e) 50 μL; (f) relationship between the OA amount and assembled structures.

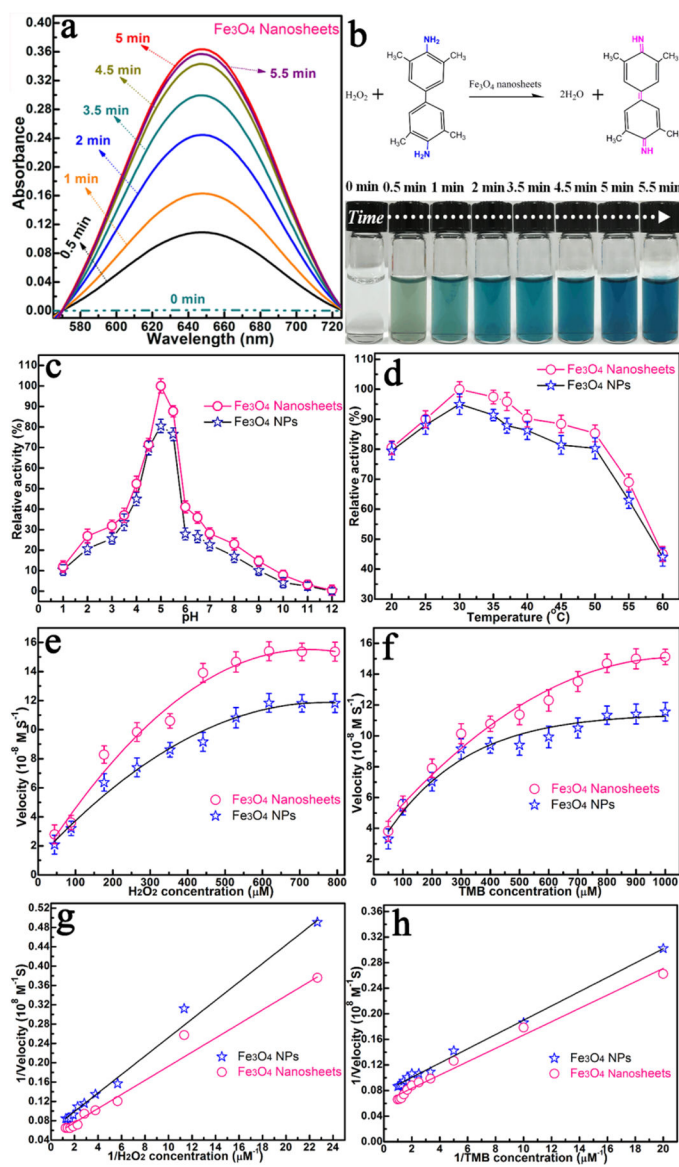


**Figure 3.** Schematic illustration of  $\text{Fe}_3\text{O}_4$  NP–micelles formation: (a) OA-stabilized  $\text{Fe}_3\text{O}_4$  NPs; (b)  $\text{Fe}_3\text{O}_4$  NP–micelles. These adopt an interdigitated bilayer structure with OA ligands as the inner layer and CTAB as the outer layer (when the OA volume is more than  $30 \mu\text{L}$ , many more OA molecules adsorb onto the surface of the  $\text{Fe}_3\text{O}_4$  NPs).



**Figure 4.** Scheme of the free-floating 2D nanosheet assembly: (a) Fe<sub>3</sub>O<sub>4</sub> NP-micelles in a mixed solvent; (b) self-dissociation of the CTAB layer; (c) OA acting as a "lubricating agent" to guide NP rearrangement; (d) formation of 2D nanosheets (the OA amount is 60  $\mu$ L).





**Figure 5.**

(a) UV/vis spectra. (b) Photographs of  $\text{Fe}_3\text{O}_4$  nanosheet catalysis using TMB as the substrate in the presence of  $\text{H}_2\text{O}_2$  (pH 4.5 and  $30^\circ\text{C}$ ). (c) pH and (d) temperature dependences of the peroxidase-like activity of the  $\text{Fe}_3\text{O}_4$  nanosheets and  $\text{Fe}_3\text{O}_4$  NPs. The velocity of the reaction was measured using  $\text{Fe}_3\text{O}_4$  NPs and  $\text{Fe}_3\text{O}_4$  nanosheets.  $\text{Fe}_3\text{O}_4$  nanosheets and  $\text{Fe}_3\text{O}_4$  NPs show optimal pH and temperature of 4.5 and  $30^\circ\text{C}$ , respectively. The maximum point in each curve was set as 100%. (e) Plot of the concentration of  $\text{H}_2\text{O}_2$  at  $530\ \mu\text{M}$  and TMB concentration varied. (f) Plot of the concentration of TMB at  $800\ \mu\text{M}$  and  $\text{H}_2\text{O}_2$  concentration varied. (g and h) Double-reciprocal plots with the concentration of one substrate ( $\text{H}_2\text{O}_2$  or TMB) fixed and the other varied. Experimental conditions:  $0.2\ \text{M}$  sodium acetate (NaAc) buffer, pH 4.5,  $30^\circ\text{C}$ .