Hindawi Disease Markers Volume 2020, Article ID 8831466, 10 pages https://doi.org/10.1155/2020/8831466

Research Article

Coexpression of Matrix Metalloproteinase-7 and Tissue Inhibitor of Metalloproteinase-1 as a Prognostic Biomarker in Gastric Cancer

Yangyu Zhang,¹ Lili Qin,¹ Xiaobo Ma ,² Yueqi Wang ,¹ Yanhua Wu ,¹ and Jing Jiang ,¹

Correspondence should be addressed to Yanhua Wu; wuyanhua@jlu.edu.cn and Jing Jiang; jiangjing19702000@jlu.edu.cn

Received 15 May 2020; Revised 24 July 2020; Accepted 1 August 2020; Published 14 September 2020

Academic Editor: Andrea Cabrera-Pastor

Copyright © 2020 Yangyu Zhang et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. Degradation of the extracellular matrix (ECM), an essential step in tumour invasion and metastasis, is mainly dependent on the activities of both matrix metalloproteinases (MMPs) and tissue inhibitors of metalloproteinases (TIMPs). This study aimed to explore whether expression of MMP-7 and TIMP-1 alone and in combination can be used as a prognostic marker for gastric cancer (GC). Method. A total of 285 patients who had undergone tumourectomy for GC were included. Gastric tumour tissues were stained immunohistochemically to evaluate expression of MMP-7 and TIMP-1. Results. Expression of MMP-7 was associated with tumour N stage and neural invasion. Multivariate Cox regression analysis suggested that expression of MMP-7 or TIMP-1 alone cannot serve as an indicator of patient prognosis; however, coexpression of MMP-7 and TIMP-1 was found to be an independent predictive factor of overall survival in patients with GC (HR = 1.74, 95% CI: 1.08-2.80). The results of stratified analysis also showed that the predictive value of MMP-7 and TIMP-1 coexpression was stronger in patients with N3 stage disease and not receiving chemotherapy. Conclusions. In conclusion, coexpression of MMP-7 and its inhibitor TIMP-1 in gastric tumour tissues is a potential prognostic marker for GC. Greater knowledge of protein expression will lead to new paradigms and possible improvements in therapeutics.

1. Introduction

Gastric cancer (GC) is the fifth most common malignant tumour and third leading cause of cancer death worldwide [1]. With the significant increase in discoveries and improvements in the diagnosis and treatment of GC, the mortality rate has declined notably during the past few decades. However, the prognosis of GC patients remains poor in China, with a 5-year survival rate of 35.9% [2]. Different recurrence and survival rates observed for GC patients with the same tumour stage indicate that clinical stage alone cannot sufficiently predict the biological behaviour of the tumour; novel biomarkers are needed to complement clinical parameters for individual therapy and improve survival outcomes [3].

Invasion and metastasis are estimated to account for more than 90% of patient mortality associated with solid tumours [4], and proteolytic degradation of the extracellular matrix (ECM) is an essential step in tumour invasion and metastasis [5, 6]. This degradation is mainly dependent on the activities of both matrix metalloproteinases (MMPs) and tissue inhibitors of metalloproteinases (TIMPs) [7]. MMPs are zinc-dependent proteinases that are capable of degrading almost all of the ECM in tissues and are overexpressed in many cancer tissues [8, 9]. TIMPs are secreted proteins that bind as a complex with MMPs to regulate their activity and strictly control ECM degradation [10]. However, in addition to their MMP inhibitory activity, TIMPs have various other functions, including inhibition of apoptosis by stimulating the Akt survival pathway [11], induction of angiogenesis [12] and direct stimulation of cell proliferation [13, 14], which may be directly involved in the progression and metastasis of cancer.

MMP-7 has been recognized as a critical member of the MMP family owing to its broad proteolytic activity against

¹Division of Clinical Research, First Hospital of Jilin University, Changchun, Jilin Province, China

²Department of Pathology, First Hospital of Jilin University, Changchun, Jilin Province, China

a variety of extracellular matrix substrates [15] and its ability to activate other MMPs (such as MMP-2 and MMP-9) [16]. TIMP-1 can block MMP-7 and coregulate ECM degradation. A number of studies have shown the poor prognostic effects of MMP-7 and TIMP-1 in gastric carcinoma [17-21], whereas others have reported conflicting results [22, 23]. Moreover, these studies focused on MMP-7 or TIMP-1 alone. Recently, Michal et al. [24] found a concomitant expression of MMP-7 and TIMP-1 in serum to be associated with worse survival in patients with colorectal cancer (CRC). However, the usefulness of MMP-7 and TIMP-1 coexpression in predicting survival outcome in GC patients has rarely been explored. Therefore, this study used immunohistochemistry to examine MMP-7 and TIMP-1 expression in gastric specimens from 285 GC patients and explore whether expression of MMP-7 and TIMP-1 alone and in combination can be used as a prognostic marker for GC.

2. Materials and Methods

- 2.1. Study Population. A total of 285 GC patients who had undergone tumourectomy were enrolled at the Gastric and Colorectal Department of the First Hospital of Jilin University (Changchun, China) from February 2011 to August 2018. Patients with positive surgical margins were excluded. None of the included patients had received preoperative chemotherapy or radiotherapy. The included clinical characteristics were age, gender, tumour size, postoperative chemotherapy, T stage, N stage, histological grade, vascular invasion, neural invasion, and follow-up data. Before enrolment, all participants signed informed consent forms, and this study was approved by the Ethics Committee of the First Hospital of Jilin University (Changchun, China, 2013-005).
- 2.2. Follow-Up Data Collection. Follow-up was scheduled in the third month, sixth month, and first year after the tumour-ectomy and then every year until death, the end of our study, or loss to follow-up. Patients who died of complications within 30 days after the operation or were lost to follow-up at the first appointment were excluded. Survival time was defined from the date of surgery to the date of death or final successful follow-up (whether the patients were alive or lost to follow-up).
- 2.3. Experimental Methods. In our study, immunohistochemistry staining was used to detect expression of MMP-7 and TIMP-1 proteins on tissue microarray slides containing formalin-fixed, paraffin-embedded tissue arrayed samples of GC patients. After routine baking and dewaxing, the slides were heated in a pressure cooker with EDTA for thirty minutes for antigen retrieval. The blocking of endogenous peroxidase was performed by using H₂O₂. The slides were incubated with primary antibodies against MMP-7 (ab205525, dilution: 1/150, Abcam, Cambridge, UK) and TIMP-1 (ab211926, dilution: 1/150, Abcam) for 1 hour and 30 minutes at room temperature, followed by further incubation with goat anti-rabbit IgG for 25 minutes at room temperature and washing 3 times with PBS for 5 minutes each time. 3,3′-Diaminobenzidine tetrahydrochloride (DAB) was

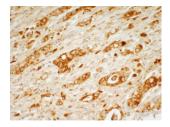


FIGURE 1: MMP-7 positive staining. The membrane or cytoplasm of gastric cancer cells was stained brown. Original magnification, ×400.

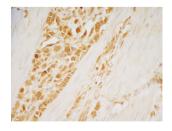


FIGURE 2: TIMP-1 positive staining. The membrane or cytoplasm of gastric cancer cells was stained brown. Original magnification, ×400.

applied to detect antigen-antibody complexes. Finally, the slides were counterstained with haematoxylin. Two pathologists who were blinded to the clinical data evaluated the immunohistochemical staining to determine expression of the two proteins. Positive expression was defined as the presence of brown granules in the cytoplasm and/or cell membrane of tumour cells.

2.4. Statistical Analysis. Discrete variables were compared by Pearson's χ^2 or Fisher's exact test, and the results are represented as frequencies (percentages). Differences in Kaplan-Meier survival curves were assessed by the log-rank test. Hazard ratios (HRs) and 95% confidence intervals (CIs) of possible prognostic factors were assessed using the Cox proportional hazards regression model. All of the analyses were two-sided, and P < 0.05 was considered to be statistically significant. SPSS (version 21.0, Chicago, IL, USA) was used to conduct the analyses.

3. Results

3.1. Expression of MMP-7 and TIMP-1 and Clinicopathological Characteristics of Gastric Carcinoma Patients. A total of 285 patients were enrolled in the study, and the median follow-up period was 66 months. During follow-up, 168 (58.9%) patients died of GC, 106 (37.2%) patients were still alive, and 11 (3.9%) patients were lost to follow-up. There were 219 (76.8%) males in our study, and the median age of the patients was 63 (range 26-91) years. Figures 1 and 2 illustrate the positive expression of MMP-7 and TIMP-1, respectively. Among the 285 gastric tumour tissues examined, the incidence of positive MMP-7 expression was 34.4% (98/285). A total of 78 (27.4%) GC patients had

Table 1: Clinicopathological characteristics of 285 patients and immunohistochemical staining for MMP-7 and TIMP-1.

Variables		N (%)	MMP-7		TIMP-1	
v arrables		IV (70)	+ (%)	P	+ (%)	P
				0.02		0.06
Sex	Male	219 (76.8)	83 (37.9)		54 (24.7)	
	Female	66 (23.2)	15 (22.7)		24 (36.4)	
				0.55		0.37
Age	<70	212 (74.4)	75 (35.4)		61 (28.8)	
	≥70	73 (25.6)	23 (31.5)		17 (23.3)	
				0.71		0.67
	T1	4 (1.4)	1 (25.0)		1 (25.0)	
T stage	T2	10 (3.5)	2 (20.0)		1 (10.0)	
	Т3	148 (51.9)	49 (33.1)		40 (27.0)	
	T4	123 (43.2)	46 (37.4)		36 (29.3)	
				0.03		0.67
	N0	47 (16.5)	8 (17.0)		11 (23.4)	
N stage	N1	48 (16.8)	16 (33.3)		13 (27.1)	
Ü	N2	77 (27)	33 (42.9)		25 (32.5)	
	N3	113 (39.6)	41 (36.3)		29 (25.7)	
Differentiation				0.74		0.48
	Middle	58 (20.4)	21 (36.2)		18 (31.0)	
	Low	227 (79.6)	77 (33.9)		60 (26.4)	
				0.59		0.73
Type	Adenocarcinoma and others	255 (89.5)	89 (34.9)		69 (27.4)	
-/[-	Signet-ring cell	30 (10.5)	9 (30.0)		9 (30.0)	
				0.30		0.61
Size	<5 cm	85 (29.8)	33 (38.8)		25 (29.4)	
	≥5 cm	200 (70.2)	65 (32.5)		53 (26.5)	
				0.76		0.63
Lymph vascular invasion	No	64 (22.5)	21 (32.8)	0.70	16 (25.0)	0.00
	Yes	221 (77.5)	77 (34.8)		62 (28.1)	
		221 (//10)	,, (6 116)	0.002	02 (2011)	0.11
Neural invasion	No	78 (27.4)	16 (20.5)	0.002	16 (20.5)	0.11
	Yes	207 (72.6)			62 (30.0)	
	1 es	207 (72.0)	82 (39.6)		02 (30.0)	
		4.50 (= 5.5)		0.13	2= (0.07
Chemotherapy	No	160 (56.1)	61 (38.1)		37 (23.1)	
	Yes	125 (43.9)	37 (29.6)		41 (32.8)	

^{+:} positive expression.

positive immunohistochemical staining for TIMP-1 in tumour tissues. The results of immunostaining for the presence of MMP-7 and TIMP-1 in tumour tissues in relation to clinicopathological tumour characteristics are presented in Table 1. Expression of MMP-7 was associated with sex, with MMP-7 more frequently expressed in males than in females (P = 0.02). Furthermore, expression of MMP-7 was associated with tumour N stage (P = 0.03) and neural invasion (P = 0.002). MMP-7 expression increased with lymph node metastasis, and patients with tumours positive for MMP-7 often had neural invasion. The expression of

TIMP-1 was not found to be associated with tumour size, histological type, depth of tumour invasion, lymph node metastasis, or lymph vascular/neural invasion.

3.2. Associations between Survival and Different Clinicopathological Characteristics and Expression of MMP-7 and TIMP-1. The associations we detected between survival and different clinicopathological characteristics and different expression patterns of MMP-7 and TIMP-1 in tumour tissues of patients are summarized in Table 2. The cumulative survival time for patients with and without MMP-7

Table 2: Association between the clinicopathological characteristics and prognosis of patients with gastric carcinoma.

Variables		5-year survival (%)	MST (months)	Log-rank F
Sex	Male	41.5	35.2	0.54
OCA	Female	37.3	26.2	
				<0.01
Age	<70	45.4	42.2	₹0.01
	≥70	26.9	24.3	
				<0.01
	T1	50.0	50.3 [†]	V0.01
T stage	T2	77.1	48.8 [†]	
	Т3	52.3	69.3	
	T4	23.9	22.3	
				<0.01
	N0	71.9	75.2 [†]	(0.01
N stage	N1	58.2	72.2	
	N2	38.6	27.3	
	N3	20.2	17.1	
				<0.01
Differentiation	Middle	58.6	72.2	V0.01
	Low	35.9	26.9	
				0.25
T	Adenocarcinoma and others	42.3	31.4	0.25
Туре	Signet-ring cell	25.6	28.2	
	Signet-Ting cen	23.0	26.2	
	_			0.08
Size	<5 cm	45.4	42.6	
	≥5 cm	38.6	26.9	
				< 0.01
Lymph vascular invasion	No	57.0	72.18	
	Yes	35.8	26.94	
				< 0.01
Neural invasion	No	64.3	83.15	
	Yes	31.8	24.48	
				< 0.01
Chemotherapy	No	32.4	22.28	
	Yes	51.1	83.15	
				0.39
MMP-7	_	41.9	35.58	
	+	37.8	24.81	
				0.71
TIMP-1	_	41.5	35.15	0.71
	+	38.1	27.27	
	·			0.10
TIMP-1&MMP-7	Both –	41.7	30.82	0.18
	-/+ or +/-	41.9	42.09	
	-/+ or +/- Both +	30.0	17.38	
	Dour I	20.0	17.50	0.07
TIMD 10-MMD 7	Nat bath.	41.7	27.10	0.07
TIMP-1&MMP-7	Not both +	41.7	37.19	
	Both +	30.0	17.38	

[†]Mean survival time was provided when MST could not be calculated; MST: median survival time, months; –: negative expression; +: positive expression; both –: patients whose tumour tissue expressed neither MMP-7 nor TIMP-1; –/+: patients whose tumour tissue expressed only TIMP-1; +/-: patients whose tumour tissue expressed only MMP-7; both +: patients whose tumour tissue expressed both MMP-7 and TIMP-1.

expression did not differ significantly (P = 0.39; Figure 3(a)). There was also no significant difference between patients with positive expression of TIMP-1 and negative expression of TIMP-1 (P = 0.71; Figure 3(b)). Three patterns were observed according to MMP-7 and TIMP-1 expression in gastric carcinoma tissues: A, negative expression of both MMP-7 and TIMP-1, n = 139 (48.8%); B, positive MMP-7 and negative TIMP-1 expression or negative MMP-7 and positive TIMP-1 expression, n = 116 (40.7%); and C, positive expression of both MMP-7 and TIMP-1, n = 30 (10.5%). The survival curves associated with the three patterns are depicted in Figure 3(c). As the curves of patterns A and B were very close and the 5-year survival rates were similar in the two groups, we combined them into one group; pattern C was treated as a separate group. The survival curves of the two groups are shown in Figure 3(d).

3.3. Association between Coexpression of MMP-7 and TIMP-1 and Overall Survival. A multivariate stepwise Cox regression model was performed to explore prognostic factors for GC. The results showed that age group, T stage, N stage, chemotherapy, and coexpression of MMP-7 and TIMP-1 in tumour tissues were associated with patient survival. Patients with positive expression of MMP-7 and TIMP-1 had a significantly increased risk of death compared with patients with other patterns of expression (HR = 1.74, 95% CI: 1.08-2.80) (Table 3).

3.4. Stratified Analysis of Expression of MMP-7 and TIMP-1 Associated with Gastric Cancer Prognosis. Associations between coexpression of MMP-7 and TIMP-1 in tumour tissues and the survival of GC patients were evaluated by a stratified analysis of age group, T stage, N stage, and chemotherapy. Compared to other patterns of expression, patients expressing both MMP-7 and TIMP-1 had a higher death risk in the subgroup of patients aged ≥ 70 years old (HR = 4.99, 95% CI: 1.81–13.77), patients with an advanced N stage (N stage III: HR = 2.54, 95% CI: 1.21–5.32) or patients with no postoperative chemotherapy (HR = 1.91, 95% CI: 1.02–3.58) (Figure 4).

4. Discussion

We analysed MMP-7 and TIMP-1 expression in GC by immunohistochemistry using 285 surgical specimens. Our results showed that although expression of MMP-7 or TIMP-1 alone may not serve as an indicator for patient prognosis, there was a significant association between concomitant positive expression of MMP-7 and TIMP-1 with poor survival of patients with gastric carcinoma. To our knowledge, this is the first study exploring the association of clinicopathological characteristics and the prognostic significance of positive expression of MMP-7 together with TIMP-1 in GC patients.

By modulating the degradation of ECM proteins and the regulation of other biomolecules in the body, MMPs are involved in many physiological and pathological conditions, such as growth, differentiation, inflammation, and angiogenesis [25, 26]. Moreover, these enzymes can affect essential

steps in the biological processes of tumours and play important roles in tumour invasion and tumour spread [27, 28]. As a unique member of this family, MMP-7 exhibits a wide spectrum of substrate specificity and potential for activating a cascade of MMPs, which indicates that it is a promising biomarker in tumorigenesis and cancer progression [29-32]. Our results showed that MMP-7 was more frequently expressed in male than in female patients with GC, which may be associated with males mounting a greater and often more damaging inflammatory response to infection compared to females and with the protective effects of female sex hormones [33, 34]. Ajisaka et al. [18] found positive expression of MMP-7 in gastric tumours to be associated with deep invasion of the gastric wall, nodal metastases, and infiltration of lymph vascular invasion. Koskensalo et al. [19] confirmed that MMP-7 overexpression is associated with lymph node and distant metastases in GC. The results of our study showed that expression of MMP-7 was related to nodal metastases and neural invasion but not to depth of invasion. The role of TIMP-1 in tumour invasion and metastasis appears to be complex and, in some cases, even divergent. For example, a study conducted by Zhang et al. [35] found significant negative correlations between TIMP-1 immunoreactivity and depth of invasion and lymph node metastasis, though a study conducted by Mroczko et al. [36] reported that expression of TIMP-1 in cancer tissue correlated positively with the depth of tumour invasion and nodal metastasis. We did not find associations between TIMP-1 expression and depth of invasion or lymph node metastasis, which was different from other studies. The inconsistent results may be due to the characteristics of GC patients in our study being different from those of the other studies. Indeed, more than 95% of our patients received radical gastrectomy and had T3- or T4-stage disease, which may have limited the statistical power for evaluating associations between the two proteinases and the tumour invasion process.

Although our results suggested that expression of MMP-7 or TIMP-1 alone cannot serve as an indicator for patient prognosis, there was a significant association between coexpression of MMP-7 and TIMP-1 with poor survival of patients with gastric carcinoma. On the one hand, expression of MMP-7 gradually increases with tumour progression, and eventually, highly expressed MMP-7 induces expression of TIMP-1 to regulate proteinase reactions [37]. The coexpression of MMP-7 and TIMP-1 in tumour tissue reflects a high level of activation of the entire proteinase system, which indicates an increase in tumour growth and metastasis [38]. On the other hand, TIMP-1 mediates constitutive activation of focal adhesion kinase (FAK), and phosphorylated FAK activates phosphatidylinositol 3-kinase (PI3K), which in turn activates Akt. The PI3K/Akt pathway is a canonical pathway involved in antiapoptosis and prosurvival, regulating a number of activities, such as proliferation, migration, and survival [39]. Moreover, the activated PI3K/Akt pathway can further promote expression of MMP-7 downstream [40]. Therefore, we speculate that coexpression of MMP-7 and TIMP-1 in tumour tissue may initiate a positive feedback pathway that directly accelerates tumour invasion and progression.

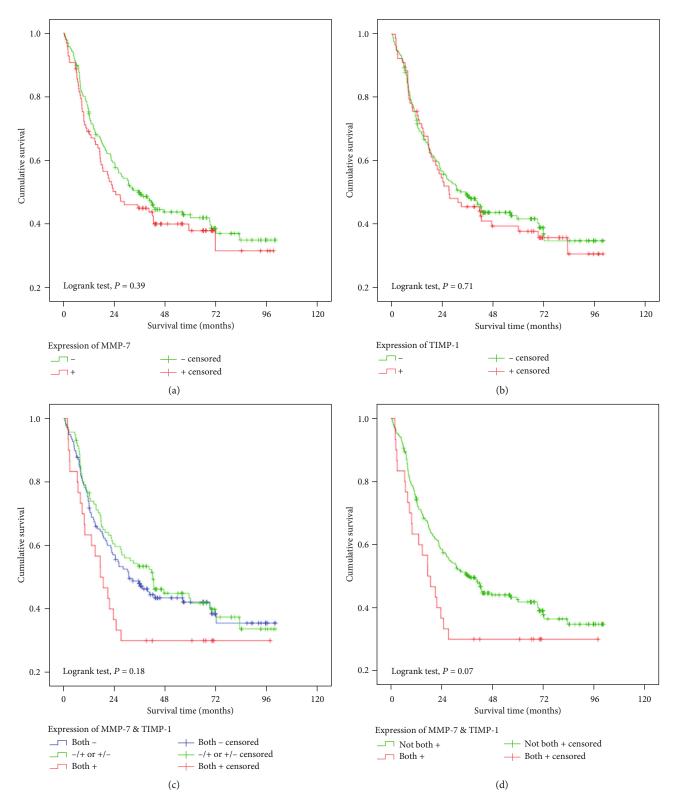


FIGURE 3: The association between expression of MMP-7 and TIMP-1 and postoperative survival in gastric carcinoma. (a) Comparison in terms of expression of MMP-7. (b) Comparison in terms of expression of TIMP-1. (c) Comparison between the three groups. (d) Comparison between the two groups. –: negative expression; +: positive expression; both –: patients whose tumour tissue expressed neither MMP-7 nor TIMP-1; –/+: patients whose tumour tissue expressed only TIMP-1; +/-: patients whose tumour tissue expressed only MMP-7; and both +: patients whose tumour tissue expressed both MMP-7 and TIMP-1.

Table 3: Stepwise Cox regression analysis of the survival of gastric cancer patients.

Variables	Adjusted HR (95% CI)	P
Age (≥70 vs <70)	1.56 (1.11-2.20)	0.01
T stage	1.39 (1.03-1.88)	0.03
N stage	1.63 (1.35-1.97)	< 0.01
Chemotherapy	0.56 (0.40-0.78)	0.01
TIMP-1&MMP-7 (both + vs not both +)	1.74(1.08-2.80)	0.02

Age group, sex, tumour size, T stage, N stage, differentiation, histological type, chemotherapy, lymph vascular invasion, neural invasion, and expression of MMP-7 and TIMP-1 were used as variables in the regression model. Both +: patients whose tumour tissue expressed both MMP-7 and TIMP-1.

Variables		HR (95% Cl)
Age	į	
<70	 - -	1.35 (0.77, 2.37)
≥70	-	4.99 (1.81, 13.77)
T stage	!	
I/II/III	ļ•	- 2.00 (0.98, 14.09)
IV	!	1.40 (0.70, 2.75)
N stage	!	
N0	-	1.49 (0.27, 8.24)
N1	-	0.89 (0.10, 7.93)
N2	:	1.81 (0.78, 4.20)
N3	-	2.54 (1.21, 5.32)
Chemotherapy		
No	 	1.91 (1.02, 3.58)
Yes		1.62 (0.76, 3.45)
	:	
	!	

FIGURE 4: Stratified analysis of the association between coexpression of MMP-7 and TIMP-1 and gastric cancer patient survival (both + vs not both +). Age group, sex, tumour size, T stage, N stage, differentiation, histological type, chemotherapy, lymph vascular invasion, and neural invasion were used as variables in the regression model. Both +: patients whose tumour tissues expressed both MMP-7 and TIMP-1.

Overall, concomitant positive expression of MMP-7 and TIMP-1 might comprehensively evaluate the state of tumour progression and predict the prognosis of patients with GC.

Our results indicate a positive correlation between concomitant expression of MMP-7 and TIMP-1 and the process of metastasis, which is consistent with the results of our stratified analysis showing that the predictive value was stronger in patients with N3-stage disease (HR: 2.54, 95% CI: 1.21-5.32). This also indicates that for patients with coexpression of MMP-7 and TIMP-1, more attention should be paid to lymph node detection to identify all possible metastatic lymph nodes. Furthermore, the results showed that in patients who did not receive chemotherapy, coexpression of MMP-7 and TIMP-1 could predict poor survival; however, in the subgroup of GC patients who received chemotherapy,

the difference disappeared. Previous studies have demonstrated that chemotherapy may have a direct effect on biomarker levels in some patients [41, 42]. TIMP-1 has broad tumour-promoting activity (proangiogenesis, antiapoptosis; promotion of cell growth, and proliferation) [43]. The majority of conventional chemotherapeutic drugs act by killing cancer cells by inducing apoptosis or function as epidermal growth factor receptor inhibitors or antiangiogenic drugs [44, 45]. The effect of apoptosis or antiangiogenesis induced by chemotherapy may affect the functions of TIMP-1 [46–48], which further influences MMP-7 downstream. Therefore, the predictive value of coexpression of MMP-7 and TIMP-1 in the subgroup of patients who received chemotherapy was overshadowed.

Elevation of blood MMP-7 and TIMP-1 levels is considered to result from reactive production of MMP-7 and TIMP-1 in cancerous tissues [49]. Increased blood MMP-7 and TIMP-1 levels occur when the proteins enter the circulation from tumour tissues [50]. Unfortunately, no study to date has investigated the correlation between serum/plasma levels of TIMP-1 and tumour tissue levels of TIMP-1 in GC patients [51]. And only a weak correlation between the serum level of MMP7 and the tumour tissue level of MMP7 has been reported in GC [52]. A study investigating the serum levels of TIMP-1 and MMP-7 in patients with metastatic CRC found that serum MMP-7 together with TIMP-1 can be used as effective tumour markers for detecting metastatic CRC [24]. In GC patients, most studies evaluating blood TIMP-1 levels [53, 54] have reported elevated TIMP-1 protein levels to be associated with a worse prognosis. However, the meta-analysis conducted by Soleyman-Jahi et al. [55] to explore the association between MMP7 and the prognosis of GC indicated that the pooled HR of two studies that used serum MMP7 was not statistically significant. The blood levels of MMP-7 together with TIMP-1 in predicting survival outcome in GC patients have not yet been reported. Noticeably, when using thawed serum samples, there is a substantial risk of falsely elevated levels of TIMP-1 because freezing and thawing of serum have been shown to cause high TIMP-1 levels due to activation and disintegration of platelets, which have a high TIMP-1 content [56]. In addition, serum or plasma levels of MMP-7 are elevated in Helicobacter pylori infection and periodontitis [57-59]. Therefore, it should be confirmed whether serum measurements of MMP-7 and TIMP-1 yield reliable results, and further studies are needed to evaluate whether the serum level and tissue expression of the same biomarker correlate in predicting the prognosis of GC.

The tissue microarray method allows analysis of only small regions of each tumour tissue; however, the samples from patients enrolled in a study can be stained as one batch without the risk of heterogeneity between staining experiments. The mechanism of the synergy between MMP-7 and TIMP-1 needs to be further confirmed. Moreover, the clinical application of immunohistochemical staining for MMP-7 and TIMP-1 in combination should be explored on a larger scale to estimate their value as potential predictive biomarkers for therapeutic decision-making and consequently improving survival outcomes.

5. Conclusions

In conclusion, coexpression of MMP-7 and its inhibitor TIMP-1 in gastric tumour tissues is a potential prognostic marker for GC. A better understanding of the expression of proteins will lead to new paradigms and possible improvements in therapeutics. Our results are promising but need to be validated in other patient cohorts.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare that they have no competing interests.

Authors' Contributions

Yangyu Zhang and Lili Qin contributed equally to this work.

Acknowledgments

We thank Ying Song for her work on follow-up of participants and the group of Gastric and Colorectal Surgery for their support. This work was supported by the National Natural Science Foundation of China (81874279, 81703286), China Postdoctoral Science Foundation (2019M650061), and Science and Technology Department of Jilin Province (20190201093JC).

References

- [1] F. Bray, J. Ferlay, I. Soerjomataram, R. L. Siegel, L. A. Torre, and A. Jemal, "Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries," *CA: a Cancer Journal for Clinicians*, vol. 68, no. 6, pp. 394–424, 2018.
- [2] L. Yang, R. Zheng, N. Wang et al., "Incidence and mortality of stomach cancer in China, 2014," *Chinese Journal of Cancer Research*, vol. 30, no. 3, pp. 291–298, 2018.
- [3] T. Sawada, M. Yashiro, K. Sentani et al., "New molecular staging with G-factor supplements TNM classification in gastric cancer: a multicenter collaborative research by the Japan Society for Gastroenterological Carcinogenesis G-Project committee," *Gastric Cancer*, vol. 18, no. 1, pp. 119–128, 2015.
- [4] W. G. Jiang, A. J. Sanders, M. Katoh et al., "Tissue invasion and metastasis: molecular, biological and clinical perspectives," *Seminars in Cancer Biology*, vol. 35, Suppl, pp. S244–S275, 2015.
- [5] H. C. Crawford and L. M. Matrisian, "Tumor and stromal expression of matrix metalloproteinases and their role in tumor progression," *Invasion & Metastasis*, vol. 14, no. 1-6, pp. 234–245, 1994.
- [6] W. G. Stetler-Stevenson, S. Aznavoorian, and L. A. Liotta, "Tumor cell interactions with the extracellular matrix during invasion and metastasis," *Annual Review of Cell Biology*, vol. 9, no. 1, pp. 541–573, 1993.

[7] H. W. Jackson, V. Defamie, P. Waterhouse, and R. Khokha, "TIMPs: versatile extracellular regulators in cancer," *Nature Reviews. Cancer*, vol. 17, no. 1, pp. 38–53, 2017.

- [8] J. R. MacDougall and L. M. Matrisian, "Contributions of tumor and stromal matrix metalloproteinases to tumor progression, invasion and metastasis," *Cancer Metastasis Reviews*, vol. 14, no. 4, pp. 351–362, 1995.
- [9] R. A. Wagenaar-Miller, L. Gorden, and L. M. Matrisian, "Matrix metalloproteinases in colorectal cancer: is it worth talking about?," *Cancer Metastasis Reviews*, vol. 23, no. 1/2, pp. 119–135, 2004.
- [10] S. Curran, S. R. Dundas, J. Buxton, M. F. Leeman, R. Ramsay, and G. I. Murray, "Matrix metalloproteinase/tissue inhibitors of matrix metalloproteinase phenotype identifies poor prognosis colorectal cancers," *Clinical Cancer Research*, vol. 10, no. 24, pp. 8229–8234, 2004.
- [11] X.-W. Liu, M. M. Bernardo, R. Fridman, and H.-R. C. Kim, "Tissue inhibitor of metalloproteinase-1 protects human breast epithelial cells against intrinsic apoptotic cell death via the focal adhesion kinase/phosphatidylinositol 3-kinase and MAPK signaling pathway," *The Journal of Biological Chemis*try, vol. 278, no. 41, pp. 40364–40372, 2003.
- [12] K. Kessenbrock, V. Plaks, and Z. Werb, "Matrix metalloproteinases: regulators of the tumor microenvironment," *Cell*, vol. 141, no. 1, pp. 52–67, 2010.
- [13] L. Guedez, W. G. Stetler-Stevenson, L. Wolff et al., "In vitro suppression of programmed cell death of B cells by tissue inhibitor of metalloproteinases-1," *The Journal of Clinical Investigation*, vol. 102, no. 11, pp. 2002–2010, 1998.
- [14] R. L. H. Bigelow, B. J. Williams, J. L. Carroll, L. K. Daves, and J. A. Cardelli, "TIMP-1 overexpression promotes tumorigenesis of MDA-MB-231 breast cancer cells and alters expression of a subset of cancer promoting genes in vivo distinct from those observed in vitro," *Breast Cancer Research and Treat*ment, vol. 117, no. 1, pp. 31–44, 2009.
- [15] C. Kapoor, S. Vaidya, V. Wadhwan, G. K. Hitesh, and A. Pathak, "Seesaw of matrix metalloproteinases (MMPs)," *Journal of Cancer Research and Therapeutics*, vol. 12, no. 1, pp. 28–35, 2016.
- [16] S. Chakraborti, M. Mandal, S. Das, A. Mandal, and T. Chakraborti, "Regulation of matrix metalloproteinases: an overview," *Molecular and Cellular Biochemistry*, vol. 253, no. 1/2, pp. 269–285, 2003.
- [17] X. P. Liu, S. Kawauchi, A. Oga et al., "Prognostic significance of matrix metalloproteinase-7 (MMP-7) expression at the invasive front in gastric carcinoma," *Japanese Journal of Cancer Research*, vol. 93, no. 3, pp. 291–295, 2002.
- [18] H. Ajisaka, Y. Yonemura, and K. Miwa, "Correlation of lymph node metastases and expression of matrix metalloproteinase-7 in patients with gastric cancer," *Hepato-Gastroenterology*, vol. 51, no. 57, pp. 900–905, 2004.
- [19] S. Koskensalo, J. Mrena, J.-P. Wiksten et al., "MMP-7 overexpression is an independent prognostic marker in gastric cancer," *Tumour Biology*, vol. 31, no. 3, pp. 149–155, 2010.
- [20] Y. E. Joo, K. S. Seo, H. S. Kim, J. S. Rew, C. S. Park, and S. J. Kim, "Expression of tissue inhibitors of metalloproteinases (TIMPs) in gastric cancer," *Digestive Diseases and Sciences*, vol. 45, no. 1, pp. 114–121, 2000.
- [21] T. Yoshikawa, A. Tsuburaya, O. Kobayashi et al., "Intratumoral concentrations of tissue inhibitor of matrix metalloproteinase 1 in patients with gastric carcinoma a new biomartker

for invasion and its impact on survival," *Cancer*, vol. 91, no. 9, pp. 1739–1744, 2001.

- [22] M. Blanco-Calvo, N. Tarrio, M. Reboredo, M. Haz-Conde, J. Garcia, M. Quindos et al., "Circulating levels of GDF15, MMP7 and miR-200c as a poor prognostic signature in gastric cancer," *Future Oncology*, vol. 10, no. 7, pp. 1187–1202, 2014.
- [23] M. F. Fanelli, L. T. Domingos Chinen, M. D. Begnami et al., "The influence of transforming growth factor-α, cyclooxygenase-2, matrix metalloproteinase (MMP)-7, MMP-9 and CXCR4 proteins involved in epithelial-mesenchymal transition on overall survival of patients with gastric cancer," *Histopathology*, vol. 61, no. 2, pp. 153–161, 2012.
- [24] M. Vočka, D. Langer, V. Fryba et al., "Serum levels of TIMP-1 and MMP-7 as potential biomarkers in patients with metastatic colorectal cancer," *The International journal of biological markers*, vol. 34, no. 3, pp. 292–301, 2019.
- [25] E. Hadler-Olsen, J. O. Winberg, and L. Uhlin-Hansen, "Matrix metalloproteinases in cancer: their value as diagnostic and prognostic markers and therapeutic targets," *Tumour Biology*, vol. 34, no. 4, pp. 2041–2051, 2013.
- [26] Z. W. Long, J. L. Wang, and Y. N. Wang, "Matrix metalloproteinase-7 mRNA and protein expression in gastric carcinoma: a meta-analysis," *Tumour Biology*, vol. 35, no. 11, pp. 11415–11426, 2014.
- [27] A. R. Nelson, B. Fingleton, M. L. Rothenberg, and L. M. Matrisian, "Matrix metalloproteinases: biologic activity and clinical implications," *Journal of Clinical Oncology*, vol. 18, no. 5, pp. 1135–1149, 2000.
- [28] B. Davies, J. Waxman, H. Wasan et al., "Levels of matrix metalloproteases in bladder cancer correlate with tumor grade and invasion," *Cancer Research*, vol. 53, no. 22, pp. 5365–5369, 1993.
- [29] Y. Adachi, H. Yamamoto, F. Itoh et al., "Clinicopathologic and prognostic significance of matrilysin expression at the invasive front in human colorectal cancers," *International Journal of Cancer*, vol. 95, no. 5, pp. 290–294, 2001.
- [30] M. Ii, H. Yamamoto, Y. Adachi, Y. Maruyama, and Y. Shinomura, "Role of matrix metalloproteinase-7 (matrilysin) in human cancer invasion, apoptosis, growth, and angiogenesis," *Experimental Biology and Medicine (Maywood, N.J.)*, vol. 231, no. 1, pp. 20–27, 2016.
- [31] W. S. Wang, P. M. Chen, H. S. Wang, W. Y. Liang, and Y. Su, "Matrix metalloproteinase-7 increases resistance to Fasmediated apoptosis and is a poor prognostic factor of patients with colorectal carcinoma," *Carcinogenesis*, vol. 27, no. 5, pp. 1113–1120, 2006.
- [32] H. Yamamoto, F. Itoh, S. Iku et al., "Expression of matrix metalloproteinases and tissue inhibitors of metalloproteinases in human pancreatic adenocarcinomas: clinicopathologic and prognostic significance of matrilysin expression," *Journal of Clinical Oncology*, vol. 19, no. 4, pp. 1118–1127, 2001
- [33] F. Guerra-Silveira and F. Abad-Franch, "Sex bias in infectious disease epidemiology: patterns and processes," *PLoS One*, vol. 8, no. 4, article e62390, 2013.
- [34] I. Marriott and Y. M. Huet-Hudson, "Sexual dimorphism in innate immune responses to infectious organisms," *Immunologic Research*, vol. 34, no. 3, pp. 177–192, 2006.
- [35] S. Zhang, L. Li, J. Y. Lin, and H. Lin, "Imbalance between expression of matrix metalloproteinase-9 and tissue inhibitor of metalloproteinase-1 in invasiveness and metastasis of

- human gastric carcinoma," World journal of gastroenterology, vol. 9, no. 5, pp. 899–904, 2003.
- [36] B. Mroczko, M. Lukaszewicz-Zajac, M. Groblewska et al., "Expression of tissue inhibitors of metalloproteinase 1 (TIMP-1) in gastric cancer tissue," Folia Histochemica et Cytobiologica, vol. 47, no. 3, pp. 511–516, 2010.
- [37] H. Sato, Y. Kida, M. Mai et al., "Expression of genes encoding type IV collagen-degrading metalloproteinases and tissue inhibitors of metalloproteinases in various human tumor cells," *Oncogene*, vol. 7, no. 1, pp. 77–83, 1992.
- [38] M. N. Holten-Andersen, R. W. Stephens, H. J. Nielsen et al., "High preoperative plasma tissue inhibitor of metalloproteinase-1 levels are associated with short survival of patients with colorectal cancer," *Clinical Cancer Research*, vol. 6, no. 11, pp. 4292–4299, 2000.
- [39] I. Vivanco and C. L. Sawyers, "The phosphatidylinositol 3-kinase AKT pathway in human cancer," *Nature Reviews. Cancer*, vol. 2, no. 7, pp. 489–501, 2002.
- [40] T. Bufu, X. Di, Z. Yilin, L. Gege, C. Xi, and W. Ling, "Celastrol inhibits colorectal cancer cell proliferation and migration through suppression of MMP3 and MMP7 by the PI3K/AKT signaling pathway," *Anti-Cancer Drugs*, vol. 29, no. 6, pp. 530–538, 2018.
- [41] B. Aldulaymi, P. Byström, A. Berglund et al., "High plasma TIMP-1 and serum CEA levels during combination chemotherapy for metastatic colorectal cancer are significantly associated with poor outcome," *Oncology*, vol. 79, no. 1-2, pp. 144–149, 2010.
- [42] H. Birgisson, H. J. Nielsen, I. J. Christensen, B. Glimelius, and N. Brünner, "Preoperative plasma TIMP-1 is an independent prognostic indicator in patients with primary colorectal cancer: a prospective validation study," *European journal of cancer*, vol. 46, no. 18, pp. 3323–3331, 2010.
- [43] F. Mannello and G. Gazzanelli, "Tissue inhibitors of metalloproteinases and programmed cell death: conundrums, controversies and potential implications," *Apoptosis: an international journal on programmed cell death*, vol. 6, no. 6, pp. 479–482, 2001.
- [44] P. Pfeiffer, C. Qvortrup, and J. G. Eriksen, "Current role of antibody therapy in patients with metastatic colorectal cancer," *Oncogene*, vol. 26, no. 25, pp. 3661–3678, 2007.
- [45] C. Frederiksen, C. Qvortrup, I. J. Christensen et al., "Plasma TIMP-1 levels and treatment outcome in patients treated with XELOX for metastatic colorectal cancer," *Annals of oncology : official journal of the European Society for Medical Oncology*, vol. 22, no. 2, pp. 369–375, 2011.
- [46] Z. Y. Fu, J. H Lv, C. Y. Ma, D. P. Yang, and T. Wang, "Tissue inhibitor of metalloproteinase-1 decreased chemosensitivity of MDA-435 breast cancer cells to chemotherapeutic drugs through the PI3K/AKT/NF-κB pathway," *Biomedicine & Pharmacotherapy*, vol. 65, no. 3, pp. 163–167, 2011.
- [47] M. Klintman, S. Ø. Würtz, I. J. Christensen et al., "Association between tumor tissue TIMP-1 levels and objective response to first-line chemotherapy in metastatic breast cancer," *Breast Cancer Research and Treatment*, vol. 121, no. 2, pp. 365–371, 2010.
- [48] M. L. Davidsen, S. Ø. Würtz, M. U. Rømer et al., "TIMP-1 gene deficiency increases tumour cell sensitivity to chemotherapyinduced apoptosis," *British Journal of Cancer*, vol. 95, no. 8, pp. 1114–1120, 2006.
- [49] S. Naruo, H. Kanayama, H. Takigawa, S. Kagawa, K. Yamashita, and T. Hayakawa, "Serum levels of a tissue

inhibitor of metalloproteinases-1 (TIMP-1) in bladder cancer patients," *International journal of urology: official journal of the Japanese Urological Association*, vol. 1, no. 3, pp. 228–231, 1994.

- [50] M. Holten-Andersen, I. J. Christensen, M. Nilbert et al., "Association between preoperative plasma levels of tissue inhibitor of metalloproteinases 1 and rectal cancer patient survival," *European Journal of Cancer*, vol. 40, no. 1, pp. 64–72, 2004.
- [51] M. Grunnet, M. Mau-Sorensen, and N. Brunner, "Tissue inhibitor of metalloproteinase 1 (TIMP-1) as a biomarker in gastric cancer: a review," *Scandinavian Journal of Gastroenter*ology, vol. 48, no. 8, pp. 899–905, 2013.
- [52] E. S. Gerstein, L. Sini, A. B. Ryabov et al., "Comparative enzyme immunoassay of matrix metalloproteinases-2, -7, -9 and their tissue inhibitor-2 in tumors and plasma of patients with gastric cancer," *Bulletin of Experimental Biology and Medicine*, vol. 148, no. 6, pp. 899–902, 2009.
- [53] O. Kemik, A. S. Kemik, A. Sümer et al., "Levels of matrix metalloproteinase-1 and tissue inhibitors of metalloproteinase-1 in gastric cancer," *World journal of gastroenterology*, vol. 17, no. 16, pp. 2109–2112, 2011.
- [54] B. Mroczko, M. Groblewska, M. Łukaszewicz-Zajac, R. Bandurski, B. Kedra, and M. Szmitkowski, "Pre-treatment serum and plasma levels of matrix metalloproteinase 9 (MMP-9) and tissue inhibitor of matrix metalloproteinases 1 (TIMP-1) in gastric cancer patients," *Clinical chemistry and laboratory medicine.*, vol. 47, no. 9, pp. 1133–1139, 2009.
- [55] S. Soleyman-Jahi, S. Nedjat, A. Abdirad, N. Hoorshad, R. Heidari, and K. Zendehdel, "Prognostic significance of matrix metalloproteinase-7 in gastric cancer survival: a metaanalysis," *PLoS One*, vol. 10, no. 4, article e0122316, 2014.
- [56] M. N. Holten-Andersen, G. Murphy, H. J. Nielsen et al., "Quantitation of TIMP-1 in plasma of healthy blood donors and patients with advanced cancer," *British journal of cancer.*, vol. 80, no. 3-4, pp. 495–503, 1999.
- [57] L. E. Wroblewski, P.-J. M. Noble, A. Pagliocca et al., "Stimulation of MMP-7 (matrilysin) by *Helicobacter pylori* in human gastric epithelial cells: role in epithelial cell migration," *Journal of Cell Science*, vol. 116, no. 14, pp. 3017–3026, 2003.
- [58] T. Sorsa, L. Tjäderhane, and T. Salo, "Matrix metalloproteinases (MMPs) in oral diseases," *Oral diseases.*, vol. 10, no. 6, pp. 311–318, 2004.
- [59] H. C. Crawford, U. S. Krishna, D. A. Israel, L. M. Matrisian, M. K. Washington, and R. M. Peek Jr., "Helicobacter pylori strain-selective induction of matrix metalloproteinase-7 in vitro and within gastric mucosa," Gastroenterology, vol. 125, no. 4, pp. 1125–1136, 2003.