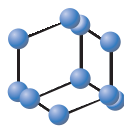


## REVIEW ARTICLE


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# Onychomycosis: An Updated Review



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**Abstract: Background:** Onychomycosis is a common fungal infection of the nail.

**Objective:** The study aimed to provide an update on the evaluation, diagnosis, and treatment of onychomycosis.

**Methods:** A PubMed search was completed in Clinical Queries using the key term "onychomycosis". The search was conducted in May 2019. The search strategy included meta-analyses, randomized controlled trials, clinical trials, observational studies, and reviews published within the past 20 years. The search was restricted to English literature. Patents were searched using the key term "onychomycosis" in www.freepatentsonline.com.

**Results:** Onychomycosis is a fungal infection of the nail unit. Approximately 90% of toenail and 75% of fingernail onychomycosis are caused by dermatophytes, notably *Trichophyton mentagrophytes* and *Trichophyton rubrum*. Clinical manifestations include discoloration of the nail, subungual hyperkeratosis, onycholysis, and onychia. The diagnosis can be confirmed by direct microscopic examination with a potassium hydroxide wet-mount preparation, histopathologic examination of the trimmed affected nail plate with a periodic-acid-Schiff stain, fungal culture, or polymerase chain reaction assays. Laboratory confirmation of onychomycosis before beginning a treatment regimen should be considered. Currently, oral terbinafine is the treatment of choice, followed by oral itraconazole. In general, topical monotherapy can be considered for mild to moderate onychomycosis and is a therapeutic option when oral antifungal agents are contraindicated or cannot be tolerated. Recent patents related to the management of onychomycosis are also discussed.

**Conclusion:** Oral antifungal therapies are effective, but significant adverse effects limit their use. Although topical antifungal therapies have minimal adverse events, they are less effective than oral antifungal therapies, due to poor nail penetration. Therefore, there is a need for exploring more effective and/or alternative treatment modalities for the treatment of onychomycosis which are safer and more effective.

**Keywords:** Dermatophytes, itraconazole, nail discoloration, onychia, onycholysis, subungual hyperkeratosis, terbinafine.

## 1. INTRODUCTION

Onychomycosis is an infection of the nail unit caused by fungi (dermatophytes, non-dermatophyte molds, and yeasts), presenting with discoloration of the nail, onycholysis, and

nail plate thickening [1, 2]. Any component of the nail unit, including the nail plate, nail matrix, and nail bed can be affected [3]. The term "onychomycosis" is derived from the Greek words "onyx" meaning nail and "mykes" meaning fungus [4]. Onychomycosis is the most common disorder affecting the nail unit and accounts for at least 50% of all nail diseases [2, 5, 6]. Laboratory confirmation of the clinical diagnosis of onychomycosis prior to initiating treatment is cost effective and is recommended [5]. In recent years, newer

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techniques enabling accurate and sensitive diagnosis of onychomycosis and novel treatments of this condition have emerged. The purpose of this communication is to provide readers with an update on current approaches to diagnosis and treatment of onychomycosis.

## 2. ETIOLOGY

Onychomycosis can be caused by dermatophytes (tinea unguium), non-dermatophyte molds and yeasts [1, 6, 7]. Approximately 90% of toenail and 75% of fingernail onychomycosis are caused by dermatophytes notably *Trichophyton mentagrophytes* and *Trichophyton rubrum* [8-12]. The remaining dermatophyte infections are caused by *Epidermophyton floccosum*, *Microsporum* species, *Trichophyton verrucosum*, *Trichophyton tonsurans*, *Trichophyton violaceum*, *Trichophyton soundanense*, *Trichophyton kraidenii*, *Trichophyton equinum*, and *Arthroderma* species [13-17]. Non-dermatophyte molds that can cause onychomycosis include *Aspergillus* species, *Scopulariopsis* species, *Fusarium* species, *Acremonium* species, *Syncephalastrum* species, *Scytalidium* species, *Paecilomyces* species, *Neoscytalidium* species, *Chaetomium* species, *Onychocola* species, and *Alternaria* species [11, 17, 18-31]. Non-dermatophyte molds account for approximately 10% of onychomycosis cases globally [6, 32]. Onychomycosis caused by yeasts is uncommon [33]. *Candida albicans* accounts for approximately 70% of onychomycosis caused by yeasts [1]. Other *Candida* species include *Candida tropicalis* and *Candida parapsilosis* [12, 17, 28, 33-35]. Patients with chronic mucocutaneous candidiasis and immunodeficiency are more likely infected with the yeast organism, especially in the fingernails [11, 28, 33, 36, 37].

## 3. EPIDEMIOLOGY

The overall worldwide prevalence of onychomycosis in the general population is approximately 5.5%, based on recently published epidemiological studies [6, 11, 14]. A 2013 systemic review of 11 population-based and 21 hospital-based studies showed that the mean prevalence of onychomycosis in North America and Europe was 4.3% (95% confidence interval: 1.9 to 6.8) in the population-based studies and 8.9% (95% confidence interval: 4.3 to 13.6) in the hospital-based studies [38]. There is evidence that the prevalence is rising, possibly because of longer life expectancy, use of occlusive modern footwear, increased prevalence of obesity, and increased urbanisation [17, 39, 40]. The condition is much more common in adults than in children and the prevalence increases with age [8, 14, 36, 41]. The prevalence in children in North America is approximately 0.4% [17], whereas the prevalence may be as high as 35% in the elderly (> 65 years of age) [42]. Toenail onychomycosis is more common in males whereas *Candida* fingernail onychomycosis is more common in females [43-45]. Other predisposing factors include fungal infection elsewhere on the body (in particular, tinea pedis), chronic paronychia, previous onychomycosis, wearing of occlusive and tight shoes, hyperhidrosis, participation in sports or fitness activities, nail trauma, poor nail grooming, use of commercial swimming pools, communal bathing, living with family members with fungal infection, poor health, genetic factors, immunodeficiency

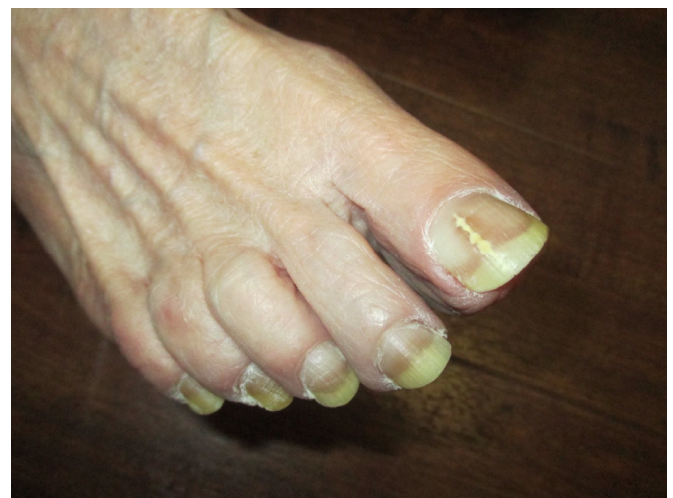
(in particular, acquired immune deficiency syndrome and transplant patients), diabetes mellitus, obesity, Down syndrome, psoriasis, smoking, peripheral vascular disease, venous insufficiency, hallux valgus, and asymmetric gait nail unit syndrome [44-56].

## 4. PATHOGENESIS

Onychomycosis is acquired through direct contact of the nail with dermatophytes, non-dermatophyte molds, or yeasts. Because the nail unit does not have effective cell-mediated immunity, it is susceptible to fungal infection [14]. Fungal production of enzymes that have proteolytic, keratinolytic, and lipolytic activities help to degrade the keratin in the nail plate and facilitate fungal invasion of the nail [36, 57]. Factors that compromise barriers to fungal infection may increase the risk for fungal infection [36]. The site and pattern of fungal invasion account for the production of different clinical subtypes of onychomycosis [57]. The formation of fungal biofilms allows the fungi to evade current antifungal therapies and contribute to antifungal resistance [58].

## 5. CLINICAL MANIFESTATIONS

Typically, onychomycosis presents as a white or yellow-brown discoloration of the nail [11, 46]. Violaceous, green, and black discoloration of the nail plate have also been observed [11, 14, 59]. Other clinical manifestations include subungual hyperkeratosis, detachment of the nail from the nail bed (onycholysis) and thickening of the nail plate (onychauxis) [14, 46, 60, 61]. Dermatophytoma presenting as linear, single or multiple white, yellow, orange or brown bands on the nail plate is specific for onychomycosis (Fig. 1) [14]. In general, toenails are affected seven to ten times more frequently than fingernails [6, 9]. The big toenails are most often affected [11]. Generally, several toenails are affected and tinea pedis is often present (Fig. 2) [14, 28]. Also, it is unusual to have more than one fingernail involved without concomitant toenail involvement unless the patient is immunocompromised or there is a history of trauma [14].



**Fig. (1).** Dermatophytoma presenting as a linear, yellow, band on the nail plate of the right big toe in a patient with distal lateral subungual onychomycosis.





**Fig. (2).** Onychomycosis in a patient with coexisting tinea pedis.

Based on the pattern of invasion, onychomycosis can be divided into the five clinical subtypes described below. It should be noted that patients may have a combination of these subtypes.

### 5.1. Distal Lateral Subungual Onychomycosis

Distal lateral subungual onychomycosis is the most common clinical subtype [35, 36, 62, 63]. In distal lateral subungual onychomycosis, the fungal invasion begins at the hyponychium and then progresses to involve the distal nail bed and subsequently the nail plate [57, 63, 64]. The fungus then migrates proximally through the nail plate, causing linear channels or "spikes" [35, 57]. This clinical subtype is usually caused by *Trichophyton rubrum* and less commonly by *Trichophyton mentagrophytes* [36, 57, 63]. Clinically, distal lateral subungual onychomycosis presents as yellowish, whitish, or brownish discoloration of a distal corner of a nail (Fig. 3) [36, 63]. Distal subungual hyperkeratosis, onycholysis, and/or onychauxis of the lateral and distal aspects of the nail plate are common [17, 28, 36].



**Fig. (3).** Distal lateral subungual onychomycosis: yellowish discoloration and onycholysis.

### 5.2. White Superficial Onychomycosis

In white superficial onychomycosis, the upper surface of the nail plate is affected by the fungus, notably *Trichophyton mentagrophytes* [36, 57, 63]. Typically, white superficial onychomycosis presents as white dots or patches on the surface of the nail plate (Fig. 4) [36, 57, 63]. The white dots and patches can be easily scraped off [8, 14, 65].



**Fig. (4).** White superficial onychomycosis.

### 5.3. Proximal Subungual Onychomycosis

Proximal subungual onychomycosis develops when the fungus invades the undersurface of the proximal nail fold in the vicinity of the cuticle and then extends distally (Fig. 5) [14, 36, 53]. This clinical subtype is usually caused by *Trichophyton rubrum* and *Fusarium* spp. [63]. Clinically, proximal subungual onychomycosis presents as an area of leukonychia in the proximal nail plate and moves distally with nail growth [57]. Proximal subungual onychomycosis usually occurs in patients with immunodeficiency, especially Acquired Immunodeficiency Syndrome (AIDS) [8, 14].



**Fig. (5).** Proximal subungual onychomycosis.

#### 5.4. Endonyx Onychomycosis

Endonyx onychomycosis is caused by fungal infection of the nail plate without infection of the nail bed [14, 28, 53, 63]. This clinical subtype is usually caused by *Trichophyton soundanense* and *Trichophyton violaceum* [14, 28, 63]. Clinically, endonyx onychomycosis is characterized by milky patches of the nail plate, indentations, and lamellar splitting [14, 28, 64]. The nail plate is firmly attached to the nail bed and subungual hyperkeratosis is absent [28, 36].

#### 5.5. Total Dystrophic Onychomycosis

Total dystrophic onychomycosis is characterized by the total destruction of the entire nail apparatus and is often the end-stage of onychomycosis that may follow any of the other subtypes [14, 53, 57, 63]. Clinically, total dystrophic onychomycosis presents with a severely dystrophic and crumbed nail plate which is yellowish, diffusely thickened, and friable (Fig. 6) [28, 57].



Fig. (6). Total dystrophic onychomycosis.

### 6. DIAGNOSIS AND DIAGNOSTIC STUDIES

A diagnosis of onychomycosis can be strongly suspected based on the typical clinical features such as nail discoloration, subungual hyperkeratosis/debris, onycholysis, and onychia [46, 66, 67]. In one study, the clinical diagnostic accuracy of onychomycosis among non-dermatologists and dermatologists was approximately 66% and 75%, respectively [68]. Dermoscopy of the nails is a useful, quick, non-invasive, and highly effective tool that may help to differentiate onychomycosis from other nail disorders [36, 69]. The most common dermoscopic pattern is a jagged proximal edge with spikes in the onycholytic area [8, 14, 70, 71]. Other dermoscopic findings include "ruined" appearance of the subungual hyperkeratosis, white to yellow longitudinal streaks/striae, leukonychia, chromonychia, parallel bands of different colors ("aurora borealis"), and dermatophytoma

[36, 69-75]. On the other hand, transverse onycholysis is suggestive of microtraumatic nail dystrophy [8].

The diagnosis can be confirmed by direct microscopic examination with a Potassium Hydroxide (KOH) wet-mount preparation, histopathologic examination of the trimmed affected nail plate with a Periodic-Acid-Schiff (PAS) stain, fungal culture, or Polymerase Chain Reaction (PCR) assays. The ideal test would identify the fungus and the species, determine its viability, be easy to perform with rapid result and low cost, and be highly specific and sensitive [14, 76].

Depending on the clinical presentation, nail clippings, nail plate scrapings, nail bed scrapings, and subungual scrapings may be necessary for sample collection [76, 77]. A sterile nail clipper should be used to clip the full thickness nail plate and a sterile curette or blade should be used to obtain subungual debris [57, 64, 78]. Nail plate dermoscopy can be used to identify the best location for localized abrasion to obtain adequate samples for mycological examination [79]. The clinical presentation also determines the site of sample collection [14]. For distal and lateral subungual onychomycosis, the sample should be obtained from the most proximal area of involvement (the most active area of infection where the highest concentration of hyphae is located) after clipping the distal onycholytic nail plate [17, 78]. In this regard, the hyphae at the distal end of the nail are less likely to be viable and to grow on culture media [57]. In white superficial onychomycosis, the specimen can be obtained by scraping the affected superficial aspect of the nail plate with a number 15 blade [3, 14, 36, 64, 78]. In proximal subungual onychomycosis, the upper nail plate of the proximal nail should be debrided or pared and the underlying nail debris collected with a curette [3, 14, 36, 64, 78]. Prior to sample collection, the nail plate and the surrounding soft tissue should be cleaned with 70% isopropyl or ethyl alcohol to prevent contamination [32, 64, 78]. Adequate samples should be collected, stored in sterile paper for transport, and delivered to the laboratory in a sterile container without delay [76, 78].

A potassium hydroxide preparation is a useful screening test to rule out the presence of fungi which provides almost immediate results at low cost [36, 68, 80]. The test is performed by adding a drop of 10 to 20% potassium hydroxide, to the nail specimen which is placed on a glass slide for examination by light microscopy [64, 78]. The potassium hydroxide dissolves the keratin, leaving behind easily visualized septate hyphae [64]. The specimen can be gently heated if no dimethyl sulfoxide is added to accelerate keratin dissolution [57, 64]. A positive test demonstrates fungal hyphae, spores, and yeasts cells [36, 64]. The test, however, does not provide information on the species of the fungus or fungus viability [6, 14, 36, 57]. A potassium hydroxide preparation is expertise dependent. The sensitivity has been reported as 48 to 60% [6, 57, 81]. The specificity ranges from 38 to 78% [36].

Histopathologic examination of the clipped affected nail plate with a PAS stain allows hyphae, pseudohyphae, spores, and yeasts to be visualized [32, 78]. The sensitivity ranges from 82 to 88% [57]. The sensitivity can be increased to 96% when PAS stain is combined with fungal culture [64]. PAS stain can produce results within 24 hours [6, 64]. However, the species of the offending fungus and its viability



cannot be determined in a PAS stain [32, 36, 78]. The cost associated with PAS stain is higher than potassium hydroxide preparation [36, 68, 78]. PAS stain is also more labor intensive than potassium hydroxide preparation [6].

Fungal cultures are specific (specificity 83 to 100%) [36] but are not very sensitive (sensitivity 60 to 65%) [6, 8]. Cultures are expensive and can take 2 to 4 weeks for results [6, 17]. Nevertheless, fungal cultures are useful for identifying the fungal species, provide information on fungal viability, and guide therapy [6, 14, 17]. For fungal cultures, the clinical sample is plated onto a properly selected culture medium such as Sabouraud dextrose agar [6]. Cycloheximide can be added to the Sabouraud dextrose agar to inhibit growth of non-dermatophyte molds [6, 76]. Gentamycin and chloramphenicol can be added to inhibit bacterial growth [6, 76]. To avoid false negative results, an adequate amount of scrapings is needed [14]. False negative results can also occur due to partial treatment or lingering antifungal drugs from previous treatment [6].

PCR testing allows for rapid and highly specific amplification of fungal DNA fragments [36, 82]. The PCR technique can accurately identify the causal dermatophyte [8, 40, 44, 45, 82]. Also, the results are available quickly (days instead of weeks) [8, 40, 44, 45, 82]. However, PCR assays are costly and not widely available which limit their use in general practice [78].

## 7. DIFFERENTIAL DIAGNOSIS

Differential diagnosis includes nail changes in psoriasis, lichen planus, alopecia areata, chronic dermatitis, onychogryphosis, chronic paronychia, pityriasis rubra pilaris, pachyonychia congenita, trachyonychia, onychogryphosis, median nail dystrophy, melanonychia striata, subungual melanoma, pemphigus vulgaris, pemphigoid, epidermolysis bullosa acquisita, bullous epidermolysis, subungual wart, subungual exostosis, subungual keratoacanthoma, rheumatoid arthritis, scleroderma, lupus erythematosus, scabies, tungiasis, twenty nail dystrophy, yellow nail syndrome, traumatic onychodystrophy, onychomatricoma, idiopathic onycholysis, porphyria, amyloidosis, myxoid cyst, fibroma, glomus tumor, Bowen disease, and squamous cell carcinoma [2, 3, 21, 24, 36, 44-46, 64, 83-87].

## 8. COMPLICATIONS

Onychomycosis may serve as a reservoir for cutaneous fungal infections such as tinea pedis, tinea corporis, and tinea cruris [11, 36]. The fungus may also disseminate to other nails [3]. There is an increased risk for bacterial infections such as cellulitis and paronychia, especially in immunocompromised individuals including diabetics [36, 88]. Severe onychomycosis may interfere with standing, walking, nail function, and daily activities [11, 53]. The condition, if left untreated, may cause discomfort, pain, paresthesia, nail deformities such as transverse over-curvature, difficulties in trimming thick nail plates, difficulties in fitting shoes, and low self-esteem [7, 9, 14, 37, 53, 78, 89]. In addition, onychomycosis can be unsightly and socially embarrassing (especially for females) and may have an adverse affect on quality of life [4, 6, 7, 90-92].

## 9. TREATMENT

Laboratory confirmation of onychomycosis before beginning a treatment regimen is cost-effective and should be considered to avoid misdiagnosis [14, 53, 66, 88, 93-95]. A misdiagnosis might result in unnecessary treatment and expose the patient to inherent risks of the side effects of the medications, potential negative drug-to-drug interactions associated with systemic antifungal medications, and therapeutic failure. It might also impose a financial burden to the patient [5]. However, empiric treatment of onychomycosis is still performed by many physicians [96]. Onychomycosis is notoriously difficult to treat because of the deep-seated nature of the fungus within the nail plate, the prolonged treatment required for resolution, poor patient compliance, and frequent recurrences [39]. Treatment options include oral antifungal therapy, topical antifungal therapy, laser therapy, photodynamic therapy, and surgical avulsion (e.g. very thick and chronic fungal nail).

### 9.1. Oral Antifungal Agents

Oral antifungal therapy is considered the gold standard for onychomycosis both in children and adults because of shorter courses of treatment and higher cure rates when compared with topical antifungal therapy [88, 95, 97, 98]. The incidence of adverse events associated with oral antifungal agents is lower in children [6]. Oral antifungal agents used for the treatment of onychomycosis include terbinafine (Lamisil), itraconazole (Sporanox, Sporaz, Orungal), and fluconazole (Diflucan, Celozole) [14, 53, 95]. Terbinafine, an antifungal agent of the allylamine group, is fungicidal. On the other hand, itraconazole and fluconazole are fungistatic and have more potential side effects and drug interactions than terbinafine [1, 88]. A 2017 Cochrane meta-analysis of 48 randomized controlled trials (n = 10, 200) assessing the effects of oral antifungal medications for the treatment of toenail onychomycosis found that terbinafine leads to better clinical and mycological cure rates than other treatments [99]. Currently, oral terbinafine (< 25kg, 125mg once daily; 25 to 35kg, 187.5mg once daily; > 35kg, 250mg once daily) is the drug of choice for the treatment of onychomycosis [39, 53, 62, 88, 100, 101]. Adverse events include headaches, taste disturbance, dermatitis, anorexia, vomiting, epigastric pain, diarrhea, drug-to-drug interactions and, rarely, depression, neutropenia, hepatic dysfunction, and Steven-Johnson syndrome [4, 17, 53, 81]. Generally, continuous terbinafine treatment has similar efficacy as pulse terbinafine treatment although some studies have shown superiority of continuous versus pulsed terbinafine treatment for toenail onychomycosis [102]. Oral itraconazole (children: < 20kg, 5mg/kg daily; 20 to 40kg, 100mg daily; > 40kg, 200mg daily for one week per month; adults: 200mg daily for one week per month for 3 to 6 months) should be considered for patients who cannot tolerate or fail to respond to oral terbinafine or whose onychomycosis is caused by non-dermatophyte molds or yeasts [17, 40, 61, 88]. Adverse events include headaches, gastrointestinal upset, upper respiratory infection, hypertriglyceridemia, hepatic dysfunction, and ventricular dysfunction [14, 44, 45, 53, 88]. Although oral fluconazole is approved for the treatment of onychomycosis in Europe, it is not approved by the United States Food and Drug Administration (FDA) for onychomycosis [14, 53]. Oral fluconazole (children: 3 to

6mg/kg once per week; adults: 150mg once per week) is used off-label for the treatment of onychomycosis in the United States, Canada, and Australia [14, 53]. The medication may be considered in patients who are unable to tolerate terbinafine or itraconazole [61]. Oral griseofulvin (Gris-peg, Grifulvin V) (not available in many countries, like Canada) is less effective, has more adverse events, and requires longer courses of treatment [1, 103]. For these reasons, oral griseofulvin is not the medication of choice in the treatment of onychomycosis [1, 61]. Likewise, oral ketoconazole (Nizoral) should not be used for the treatment of onychomycosis because of severe adverse events such as hepatotoxicity [61, 88].

Oral antifungal agents are recommended for all types of onychomycosis, especially when  $\geq 50\%$  of the nail is affected, multiple nails are infected, the nail matrix is involved, or dermatophytoma is present [1, 61, 63, 88, 104]. Oral antifungals, when used in combination with topical antifungals, increase the cure rate [8, 28, 63, 88]. Combination therapy can be used sequentially or in parallel. The treatment regimen should be tailored to the individual patient. Repeated treatment courses may be required, especially for chronic onychomycosis.

## 9.2. Topical Antifungal Agents

Topical antifungal therapy involves the use of nail lacquers and solutions in addition to topical antifungal agents [105, 106]. Generally, the nail is more permeable to topical antifungal agents formulated in aqueous vehicles [2]. The route of drug delivery is transungual, with the topical antifungal agent applied to the dorsal aspect of the nail [3, 60]. Although the efficacy of topical antifungal agents (e.g., efinaconazole) is not diminished with concurrent nail polish use, concurrent nail polish use may produce undesirable cosmetic changes to the quality of nail polish over time [3, 107, 108]. As such, concurrent nail polish use should be avoided. Commonly used topical antifungal agents include efinaconazole (Jublia, Clenafin) (10% nail solution), tavaborole (Kerydin) (5% nail solution), ciclopirox (Ciclodan, Penlac, Loprox) (8% nail lacquer or hydroxylacquer), amorolfine (Curanail, Loceryl, Locetar, Odenil) (5% nail lacquer), and terbinafine (Lamisil) (10% nail solution) [4, 6, 43, 53, 106, 109, 110]. Generally, topical antifungal agents are well-tolerated; adverse events are minimal and include periungual erythema and burning at the application site [6, 111, 112]. However, topical therapy requires longer treatment courses (often 48 weeks or longer) and may be less effective than oral treatment possibly because of insufficient nail plate penetration [4, 14, 88]. The impermeability of the nail can be attributed to the highly stable disulfide linkages and hydrogen bonds in the keratin network [105]. A 2019 meta-analysis of 26 randomized controlled trials ( $n = 8, 136$ ) assessing the efficacy of monotherapy for toenail onychomycosis found that the odds of mycological cure with continuous oral terbinafine or itraconazole are significantly greater than topical antifungal treatments [104]. In general, topical monotherapy may be considered for mild to moderate onychomycosis when less than 50% of the nail is affected without matrix involvement and only a few ( $< 3$ ) nails are infected [1, 2, 88]. In this regard, topical antifungal treatment is often sufficient for white superficial onychomycosis be-

cause of the superficial nature of the infection [2, 17, 61, 88]. Topical antifungal therapy is a therapeutic option when oral antifungal agents are contraindicated or cannot be tolerated [1, 2, 44, 45, 88]. Topical antifungals may also be used in combination with oral antifungal therapy as an adjunctive therapy to increase the cure rate due to synergistic antifungal action of the drugs [32, 88, 105, 113]. Because the rate of nail growth is faster and the nail plate is thinner in children, children are more likely to respond to topical antifungal therapy better than adults [17, 53, 88, 114, 115].

## 9.3. Lasers

Lasers have emerged as promising treatment options for the treatment of onychomycosis, although data are still lacking [116]. Most lasers use the principle of selective photothermolysis, whereby laser energy is preferentially absorbed by the fungal mycelia resulting in a rapid elevation of temperature within the fungal mycelia with resultant fungal cell death [39, 117]. As the treatment is targeted, the surrounding tissue is not affected, thus eliminating the potential of systemic adverse events [6, 7]. To be effective, lasers should have a wavelength between 750 and 1300 nm to penetrate the nail, a pulse duration that is shorter than the "thermal relaxation time" of the fungus, and a spatially uniform beam that does not cause "hot spots" [6, 107]. Several types of lasers have been used for the treatment of onychomycosis including long pulsed neodymium-doped yttrium aluminium garnet (Nd:YAG) laser, diode laser, and fractional carbon dioxide (CO<sub>2</sub>) laser [118-129]. Studies have shown that laser therapies are somewhat effective in achieving cosmetic endpoints in onychomycosis, but do not exceed or equal the efficacy of current topical and oral antifungal therapies in terms of achieving medical endpoints [39, 130]. Laser therapies are safe but expensive and may be considered for patients in whom systemic antifungal agents are contraindicated or as part of combination therapy to presumably increase the chances of successful fungal clearance [131].

## 9.4. Photodynamic Therapy

Photodynamic therapy involves photoactivation of a photosensitizer with light in specific wavelengths [132]. The photoactivation increases the energetic level of the photosensitizer [132]. The energy consequently generated reacts with the dissolved oxygen in the treated tissue, resulting in the formation of reactive oxygen species and free radicals which are cytotoxic [53, 132, 133]. The fungus absorbs the photosensitizer, making it more susceptible to destruction by apoptosis or necrosis than surrounding healthy tissue [3, 61]. Photosensitizers that have been used include 5-Aminolevulinic Acid (5-ALA), Methyl Aminolevulinic Acid (MAL), porphyrins, aluminium-phthalocyanine chloride, methylene blue, toluidine blue, and rose bengal [28, 39, 132, 134]. Data on the use of photodynamic therapy are scarce and mainly limited to case reports and non randomized trials. A 2016 systematic review of five *in vitro* and 12 *in vivo* studies found that photodynamic therapy may be of benefit in the treatment of onychomycosis [134]. Adverse events included mild pain, erythema, burning, edema, and blistering and were well tolerated [7, 134]. Two recent studies, each involving 20 patients with onychomycosis also showed the effectiveness of



photodynamic therapy in the treatment of onychomycosis [132, 133]. Well-designed, large-scale, randomized studies are necessary to confirm the efficacy of photodynamic therapy in order to make formal recommendations regarding its use in the treatment of onychomycosis.

### 9.5. Miscellaneous

Nail abrasion, trimming, avulsion, and debridement can be performed, if necessary, to enhance topical penetration of antifungal agents and reduce fungal load [2, 7, 60, 105]. White superficial onychomycosis can be treated by mechanical removal (e.g., scraping) of the involved area followed by topical antifungal therapy (vide supra) [88]. Surgical avulsion of the nail is painful and may cause disfigurement. Bristow *et al.* reported the use of a novel nail drill system that permits controlled micro-penetration of the nail without penetrating the nail bed beneath [135]. The authors reported the successful use of the device to deliver a topical antifungal agent directly and rapidly to the site of fungal infection with minimal adverse events, while maintaining the integrity of the nail. Chiu *et al.* used a dermaroller (Infinite Beauty, Birmingham, UK) to produce micropores on the nail surface [136]. PathFormer (Path Scientific, Carlisle, USA) is a FDA-approved device for this microporation technique [7].

Keratolytic agents such as urea, salicylic acid, lactic acid, and papain may enhance the delivery of topical antifungal agents into the nails [60]. Nam disclosed an invention for treating onychomycosis [137]. The invention comprises of urea, fumaric acid, 1, 3-buthylene glycol, a gel-forming polymer, a cross-linking agent, and 45 to 60 % by weight of water. The preparation has keratolytic and moisture-retention abilities and can be used in the treatment of onychomycosis. For patients with thick, dystrophic nails that are difficult to trim, the use of topical urea (40% ointment or cream) may be considered [88]. The topical application of urea to the treated area prior to therapy may soften the nail and increase the efficacy of the therapy [17, 132, 134]. A systematic review of six randomized, controlled trials (n = 407) showed that topical urea, as an adjunct to oral and topical antifungal treatment regimens, improves the efficacy of topical antifungal treatment [138]. K101 nail solution (Emtrix, Nalox, Naloc) is a combination of urea, propylene glycol, and lactic acid in a topical formulation. A retrospective study (n = 91) showed that combination therapy with topical K101 nail solution and oral terbinafine or itraconazole results in clearance of onychomycosis earlier than that of oral terbinafine or itraconazole alone [139]. Repka *et al.* showed that the topical application of phosphoric acid gel to the nail plate was effective to improve the permeability of the nail to topical ketoconazole; the treated nail showed sixfold higher ketoconazole permeation than the nonetched nail [140].

Preliminary studies have shown that iontophoresis could enhance the delivery of topical antifungal agents into the nail plate and other parts of the nail apparatus [7, 66, 88, 141, 142]. A tingling sensation may be experienced with the current application [7]. Additional studies are necessary to determine the efficacy and safety of iontophoresis in the treatment of onychomycosis.

## 10. PREVENTION

Because fungi thrive best in moist warm environments, patients should be advised to wear non-occlusive shoes, keep feet dry and cold, use absorbent socks, and clip nails short [66]. Tinea pedis, if present, should be treated [2, 143]. Also, family members with tinea pedis and onychomycosis should be appropriately treated [144]. To prevent recurrence, some authors suggest the use of topical antifungal therapy once weekly or twice monthly in high-risk patients for up to two years after completion of treatment [6, 40, 145, 146]. An ultraviolet C-based treatment device for footwear can be considered, as well as washing of running shoes (non-leather) in hot water.

## 11. PROGNOSIS

Generally, the prognosis is good with appropriate treatment. Yellow streaks along the lateral margin of the nail, the presence of dermatophytoma, and onychomycosis caused by non-dermatophyte molds (in particular, *Fusarium* species) are associated with a poor response to therapy. Other factors associated with a poor response include noncompliance, old age, advanced disease, nail matrix involvement, subungual hyperkeratosis greater than 2 mm, two feet-one hand syndrome, and immunodeficiency [14, 53, 64]. Poor response to therapy may also result from poor permeation of topical antifungal drugs across the nail plate and the deep-seated and recalcitrant nature of fungal infection [4]. Recurrences are not uncommon, with reported rates ranging from 10 to 53% [40, 67, 147]. Typically, recurrences occur within 3 years of completing therapy [14, 40]. Recurrence may be caused by relapse or reinfection [40].

## CONCLUSION

Onychomycosis is the most common nail disorder with a significant burden. The condition is most commonly caused by dermatophytes followed by non-dermatophyte molds and yeasts. Although the diagnosis can be strongly suspected based on clinical grounds, laboratory confirmation is necessary prior to treatment. Current treatment modalities of onychomycosis include oral antifungal therapies, topical antifungal therapies, and device-based therapies, alone or in combination. Oral antifungal therapies are recommended for onychomycosis especially for moderate to severe disease or when multiple nails are affected. On the other hand, topical antifungal therapies may be considered for mild to moderate disease. Topical antifungal therapy may also be used in combination with oral antifungal therapy to increase the cure rate of recalcitrant or severe disease. Laser therapies have shown promising results when used alone or in combination with oral and/or topical antifungal therapy. Very thick nails or nails stubborn to treatment can also be considered for surgical or chemical avulsion (e.g. high concentration urea).

Oral antifungal therapies are effective, but significant adverse effects and potential negative drug-to-drug interactions limit their use. Although topical antifungal therapies have minimal adverse events, they are less effective than oral antifungal therapies, due to poor nail penetration. As such, there is a need in exploring more effective and/or alternative treatment modalities for onychomycosis which are safer and

more effective. Combination therapies hold promise for improving patient outcomes.

## CURRENT & FUTURE DEVELOPMENTS

Novel broad spectrum oral antifungal agents that have shown promising results in the treatment of onychomycosis include posaconazole (Posanol, Noxafil), albaconazole (code name: UR-9825), ravuconazole (code names: BMS-207147, ER-30346), fosravuconazole, VT-1161, and P-3051 [53, 81, 88, 148-153]. Novel broad spectrum topical antifungal agents that have shown promising results in the treatment of onychomycosis include tazarotene (Tazorac), lanconazole (Astat), luliconazole (Luzu, Luzam, Lulicon), NCV-422, and ME1111 [53, 66]. Very recently, Mercer *et al.* disclosed a synthetic antifungal peptide in a water-based solution, NP213 (Novexatin), which is in the late-stage of development [154]. According to the authors, NP213 can penetrate the nail more effectively than other topical antifungal agents and shows promise as a drug of choice for topical treatment of onychomycosis. Thus far, these medications have not been approved by the FDA or Health Canada for the treatment of onychomycosis [81].

Viant disclosed antifungal preparations for topical treatment of onychomycosis [155]. The preparations comprise gamma-butyrolactone as the solvent vehicle, 1 to 6% by weight of salicylic acid, and a topical antifungal agent such as ciclopirox, amorolfine, and terbinafine. The therapeutic effects of these preparations are reinforced by the keratolytic action of the salicylic acid and the preparations may advantageously be made viscous, gelled or filmogenic, with or without a colorant. The author claimed that these antifungal preparations are very effective in the treatment of onychomycosis.

Zderic *et al.* disclosed an ultrasound-enhanced drug delivery method for the treatment of onychomycosis [156]. The method includes submerging a nail infected with onychomycosis in a solution containing at least one pharmaceutical agent and applying ultrasound to the infected nail for 1 to 10 minutes. Another method includes applying ultrasound to a subject's nail infected with onychomycosis first, and then applying a solution containing at least one pharmaceutical agent to the infected nail. The ultrasound device for applying ultrasound has a frequency of between 200kHz and 3,000kHz.

Mizutani *et al.* patented a film-forming preparation which contains efinaconazole or a salt thereof; 1 to 30% by weight of ethyl cellulose; 20% by weight or more of a volatile solvent (defined here as water or a liquid component having a boiling point lower than that of water); and 0.01 to 30% by weight of a surfactant [157]. The preparation does not leave a sticky surface but has sufficient cohesiveness after the preparation hardens and can be easily peeled off without washing. According to the authors, the preparation is more effective in the treatment of onychomycosis than the plain topical antifungal agent alone.

Lundahl disclosed a novel method of photodynamic therapy for the treatment of onychomycosis by applying a solution to the affected nail, wherein the solution comprises Aminolevulinic Acid (ALA) or its pharmaceutically accept-

able salts at a concentration of 5 to 20% [158]. After a waiting period of at least 10 days, the nail is exposed to a light source that emits red light in the 600-650nm wavelength range. The author claimed that the invention is safe and the long interval between the application of the photosensitizer and the photoactivating light increases the efficacy of the photodynamic therapy.

Nonthermal plasma is being investigated for the treatment of onychomycosis [53, 159]. Nonthermal plasma is created in the air by short pulses (about 10ns) of strong (about 20kV/mm peak) electric field which ionises air molecules, generating electrons, ions, hydroxy radicals, ozone, and nitric acid [53, 159]. Unlike thermal plasma which may cause extensive tissue heating and damage, nonthermal plasma with its small current and duration, does not cause substantial tissue damage [53, 159]. It has been shown that nonthermal plasma, generated by surface microdischarge technology, inhibited the growth of *Trichophyton rubrum in vitro* [160]. In a pilot study, Lipner *et al.* used nonthermal plasma to treat 19 patients with toenail onychomycosis [159]. The overall clinical cure was 53.8% and the mycological cure was 15.4% [159]. The authors concluded that nonthermal plasma may have a beneficial effect on toenail onychomycosis and that the procedure is safe. Well designed, large-scale clinical trials are needed to determine the true efficacy and safety of nonthermal plasma in treating onychomycosis.

Roe *et al.* disclosed a method that includes delivery of a redox gas solution to the affected nail for the treatment of onychomycosis, wherein the redox gas solution comprises a reactive species dissolved in a perfluorocarbon liquid [161]. The reactive species may include, alone or in combination, one or more of reactive oxygen, reactive nitrogen, reactive chlorine, or reactive bromine species [161]. The perfluorocarbon liquid may include perfluorodecalin. The treatment system includes a chamber assembly housing including a chamber formed therein; a plasma head assembly disposed within the chamber; and perfluorodecalin for forming a coating on a nail inserted within the chamber for treatment.

Topical non-prescription agents that have been used with varying success in the treatment of onychomycosis include tea tree (*Melaleuca alternifolia*) oil, cedar leaf oil (Vicks, VapoRub), snakeroot (*Ageratina pichinchensis*) extract, and a combination of cyanoacrylate, hydroquinone, and undecylenic acid (Renewed Nail) [64]. These agents have been evaluated in only a small number of studies involving a few patients. Larger, well-designed studies are necessary to evaluate the efficacy of these agents.

Fernandez and Leon disclosed a procedure for the preparation of a product based on the leaves of the plant *Sedum telephium* for the treatment of nails infected with onychomycosis [162]. The procedure consists of stripping away one side of the leaf of the *Sedum telephium* plant, after which the stripped surface is impregnated with sulfanilamide powder and then applied as a dressing on nails infected with onychomycosis.

Recently, Sonthalia *et al.* reported the successful use of Black peel (a combination of 0.5% salicylic acid, 50% black acetic acid, 6% tetrahydrojasmonic acid, 0.1% potassium



iodide, 10% bio sulphur) in two recalcitrant cases of onychomycosis [163]. Further studies have to be done to confirm or refute their findings.

Veiga and coworkers showed that propolis, an extract derived from an apiary of hives of honey bees (*Apis mellifera* L.) has shown promise in the treatment of onychomycosis [164]. This natural product is able to penetrate the nail without cytotoxicity and has good antifungal activity [165]. The authors used propolis with success in the treatment of three elderly patients with onychomycosis. This new finding warrants further investigations to further elucidate its clinical efficacy. Allergic contact dermatitis would be of some concern with this agent.

Mailland *et al.* disclosed an invention to treat onychomycosis by topically administering to the infected nail a composition which contains hydroxypropyl chitosan, water, and a lower alkanol as solvent [166]. The composition does not need to contain an antifungal agent or any nail penetration enhancer. After applying the composition, washing the nails should be avoided for at least six hours. The application should be repeated once daily for a minimum period of six months to one year. The authors claimed that the preparation is effective in the treatment of onychomycosis.

Most fungi produce biofilms [40]. Biofilms are sessile microbial communities that attach irreversibly to the epithelial surfaces such as the nail plate by means of an extracellular matrix [95, 167]. The extracellular matrix acts as a protective barrier to the organism such as a fungus within the biofilm [167]. As such, biofilms increase fungal resistance to antifungal agents by reducing penetration of these agents, along with protection from host defenses [81, 168]. Therefore, fungal biofilms may account for the persistence of onychomycosis as well as high rate of recurrence and relapse of onychomycosis [167]. Diagnostic tests to detect biofilms are needed. Device-based therapies that can destroy biofilms, enzymes such as with DNAase that can break down biofilms, dispersal agents and chitosan that can prevent biofilm adhesion, and antibodies that can inhibit matrix polysaccharides production, if available, may be used to improve treatment outcomes [6, 53, 167-171].

The inclusion of nanoparticles in topical antifungal agents enhances the drug profile and permeation, as small molecules can pass through nail pores easier than large molecules [7, 105]. The nanoparticles can be in the form of nanocapsules, nanoemulsion, nanovesicles, and polymeric nanoparticles [105, 172]. In a recent study, topical efinaconazole was combined with nitric oxide releasing nanoparticles [173]. The authors showed that the compound offered sustained nitric oxide release over time and enhanced efinaconazole penetration of the nail while exerting broad spectrum antifungal and immunomodulating properties [173]. Krainbring disclosed a novel pharmaceutically active composition which is able to permeate into the keratin sheets of the nails [174]. The composition is a microemulsion comprising ethoxy sulfate, ethoxylated glycerol fatty acid ester, and ethoxylated sorbitol or sorbitol anhydride fatty acid ester. The composition is particularly suitable for treating onychomycosis.

Definitions of treatment outcome measures in onychomycosis vary widely. As such, it is very difficult, if not im-

possible to compare data across studies. It is hoped that definitions of clinical cure, almost complete cure, clinical success, and clinical improvement need to be unified and consistently adhered to in all future studies.

#### LIST OF ABBREVIATIONS

AIDS	=	Acquired Immunodeficiency Syndrome
ALA	=	Aminolevulinic Acid
FDA	=	Food and Drug Administration
MAL	=	Methyl Aminolevulinate
PAS	=	Periodic-Acid-Schiff
PCR	=	Polymerase Chain Reaction

#### ETHICS APPROVAL AND CONSENT TO PARTICIPATE

Not applicable.

#### HUMAN AND ANIMAL RIGHTS

No Animals/Humans were used for studies that are base of this research.

#### CONSENT FOR PUBLICATION

Not applicable.

#### AVAILABILITY OF DATA AND MATERIALS

Not applicable.

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#### CONFLICT OF INTEREST

Professor Alexander K.C. Leung, Dr. Joseph M. Lam, Dr. Kin F. Leong, Professor Kam L. Hon, Dr. Benjamin Barankin, Dr. Amy A.M. Leung, and Dr. Alex H.C. Wong disclose no relevant financial relationship.

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