

Fibroblast growth factor 23: a biomarker of fibrosis and prognosis in heart failure with preserved ejection fraction

Clotilde Roy MD^{1,2}, Sibille Lejeune MD^{1,2}, Alisson Slimani MD^{1,2}, Christophe de Meester PhD^{1,2}, Sylvie A. Ahn AS^{1,2}, Michel F. Rousseau MD, PhD^{1,2}, Amzulescu Mihaela MD, PhD^{1,2}, Audrey Ginion MS², Benjamin Ferracin³, Agnès Pasquet MD, PhD^{1,2}, David Vancraeynest MD, PhD^{1,2}, Christophe Beauloye MD, PhD^{1,2}, Jean-Louis Vanoverschelde MD, PhD^{1,2}, Sandrine Horman PhD², Damien Gruson PharmD, PhD³, Bernhard L. Gerber MD, PhD^{1,2} and Anne-Catherine Pouleur MD, PhD^{1,2*}

¹Division of Cardiology, Department of Cardiovascular Diseases, Cliniques Universitaires St. Luc, Avenue Hippocrate, 10, Brussels, 1200, Belgium; ²Pôle de Recherche Cardiovasculaire (CARD), Institut de Recherche Expérimentale et Clinique (IREC), Université Catholique de Louvain, Brussels, Belgium; ³Clinical Biology Department, Cliniques Universitaires St Luc, Université catholique de Louvain, Brussels, Belgium

Abstract

Aims Besides regulating calcium-phosphate metabolism, fibroblast growth factor 23 (FGF-23) has been associated with incident heart failure (HF) and left ventricular hypertrophy. However, data about FGF-23 in HF and preserved ejection fraction (HFpEF) remain limited. The aim of this study was to assess the association between FGF-23 levels, clinical and imaging characteristics, particularly diffuse myocardial fibrosis, and prognosis in HFpEF patients.

Methods and results We prospectively included 143 consecutive HFpEF patients (78 ± 8 years, 61% female patients) and 31 controls of similar age and gender (75 ± 6 years, 61% female patients). All subjects underwent a complete two-dimensional echocardiography and cardiac magnetic resonance with extracellular volume (ECV) assessment by T1 mapping. FGF-23 was measured at baseline. Among the patients, differences in clinical and imaging characteristics across tertiles of FGF-23 levels were analysed with a trend test across the ordered groups. Patients were followed over time for a primary endpoint of all-cause mortality and first HF hospitalization and a secondary endpoint of all-cause mortality. Median FGF-23 was significantly higher in HFpEF patients compared with controls of similar age and gender (247 [115; 548] RU/mL vs. 61 [51; 68] RU/mL, $P < 0.001$). Among HFpEF patients, higher FGF-23 levels were associated with female sex, higher incidence of atrial fibrillation, lower haemoglobin, worse renal function, and higher N terminal pro brain natriuretic peptide levels (P for trend < 0.05 for all). Regarding imaging characteristics, patients with higher FGF-23 levels had greater left atrial volumes, worse right ventricular systolic function, and more fibrosis estimated by ECV (P for trend < 0.05 for all). FGF-23 was moderately correlated with ECV ($r = 0.46$, $P < 0.001$). Over a mean follow-up of 30 ± 8 months, 43 patients (31%) died and 69 patients (49%) were hospitalized for HF. A total of 87 patients (62%) reached the primary composite endpoint of all-cause mortality and/or first HF hospitalization. In multivariate Cox regression analysis for the primary endpoint, FGF-23 (HR: 3.44 [2.01; 5.90], $P < 0.001$) and E wave velocities (HR: 1.01 [1.00; 1.02], $P = 0.034$) were independent predictors of the primary composite endpoint. In multivariate Cox regression analysis for the secondary endpoint, ferritin (HR: 1.02 [1.01; 1.03], $P < 0.001$), FGF-23 (HR: 2.85 [1.26; 6.44], $P = 0.012$), and ECV (HR: 1.26 [1.03; 1.23], $P = 0.008$) were independent predictors of all-cause mortality.

Conclusions Fibroblast growth factor 23 (FGF-23) levels were significantly higher in HFpEF patients compared with controls of similar age and gender. FGF-23 was correlated with fibrosis evaluated by ECV. High levels of FGF-23 were significantly associated with signs of disease severity such as worse renal function, larger left atrial volumes, and right ventricular dysfunction. Moreover, FGF-23 was a strong predictor of poor outcome (mortality and first HF hospitalization).

Keywords FGF-23; Biomarker; Troponin; Mortality; NT-proBNP; Heart failure; Fibrosis; Prognosis

Received: 16 November 2019; Revised: 5 March 2020; Accepted: 20 May 2020

*Correspondence to: Anne-Catherine Pouleur, MD, PhD, Cardiovascular Division, Department of Cardiovascular Diseases, Cliniques Universitaires St-Luc, Avenue Hippocrate, 10, Brussels 1200, Belgium. Tel: 32-2-7646561; Fax: 32-2-7648980. Email: anne-catherine.pouleur@uclouvain.be
Clotilde Roy and Sibille Lejeune contributed equally to this work.

Introduction

Heart failure (HF) with preserved ejection fraction (HFpEF) is a major cause of cardiovascular morbidity and mortality, especially among the elderly. Over the last years, a new paradigm has been proposed for HFpEF.¹ Systemic proinflammatory state induced by multiple co-morbidities has been identified as a potential cause of myocardial structural and functional alterations¹ and may play a key role in the genesis of vascular, skeletal muscle, and end-organ damages observed in HFpEF. Furthermore, several studies using autopsies, myocardial biopsies, or extracellular volume (ECV) measured by cardiac magnetic resonance (cMR) T1 mapping have highlighted the role of myocardial structural abnormalities and interstitial fibrosis in HFpEF.^{2–5}

Fibroblast growth factor 23 (FGF-23), a bone-derived hormone regulating renal phosphate homeostasis and vitamin D metabolism, has a direct action on the cardiovascular system and has been recently related to adverse cardiovascular events in chronic HF and implicated in cardiac remodelling.^{6–9} Several recent studies suggested that higher FGF-23 levels were associated with left ventricular hypertrophy¹⁰ and worse prognosis, particularly among patients with chronic kidney disease (CKD)^{11,12} but also in stable ischaemic cardiomyopathy¹³ and HF with reduced ejection fraction (HFrEF).^{6–8,14–16} FGF-23 has also been identified as a biomarker significantly related to atrial fibrillation.¹⁷

However, little is known about the association of FGF-23 with myocardial abnormalities prevalent in HFpEF, such as left ventricle (LV) diastolic dysfunction, LV hypertrophy, interstitial fibrosis, and severity of right ventricular (RV) dysfunction, nor about its role in the progression of myocardial dysfunction. In the present study, we sought to examine the relationship between FGF-23 and baseline biomarkers, demographic, and imaging characteristics, particularly fibrosis estimated by ECV using T1 mapping by cMR, and its prognostic value in a control population and in HFpEF patients.

Methods

Study population

Between December 2014 and June 2017, consecutive patients with HFpEF were prospectively evaluated for inclusion in the study.⁵

The following criteria had to be fulfilled for study inclusion: New York Heart Association functional class \geq II, typical signs of HF, N terminal pro brain natriuretic peptide (NT-

proBNP) $>$ 350 pg/mL and/or a hospitalization for HF in the previous 12 months, left ventricular ejection fraction \geq 50%, and relevant structural heart disease [LV hypertrophy/left atrial (LA) enlargement] and/or diastolic dysfunction assessed by echocardiography. The exclusion criteria were severe valvular disease, infiltrative or hypertrophic cardiomyopathy, acute coronary syndrome in the previous 30 days, chronic obstructive pulmonary disease GOLD 3 or 4, congenital heart disease, pericardial disease, atrial fibrillation with a ventricular response $>$ 140 bpm, and severe anaemia (haemoglobin $<$ 8 g/dL).

Patients were compared with 31 controls of similar age and gender without history of cardiovascular disease.¹⁸ All controls underwent a full clinical exam, electrocardiogram, echocardiography, and exercise stress test, which all had to be normal prior to inclusion.

All subjects underwent blood sampling, complete transthoracic echocardiography, and cMR.

The investigation conforms with the principles outlined in Declaration of Helsinki. The local ethics committee approved the study, and all patients gave written informed consent before study enrolment (Clinical trial NCT03197350).

Echocardiography

All subjects underwent a complete two-dimensional transthoracic echocardiography at inclusion (iE33 system Philips) to assess LV and RV structure, systolic and diastolic function, and measurements of left and right atrial volumes, as well as a valvular evaluation. Pulmonary pressures were estimated using tricuspid regurgitation velocity. Strain analysis was performed on acquired images with acceptable quality in TOMTEC Software (Munich, Germany). All measurements were averaged over three beats in atrial fibrillation.

Cardiac magnetic resonance

Cardiac magnetic resonance (cMR) was performed using a three-Tesla system (Ingenia, Philips Medical Systems, Best, The Netherlands). The different sequences have been previously described.⁵

Pre-contrast and post-contrast MOLLI images were processed using the open-source software MRmap v1.4 under IDL. Pre-myocardial and post-myocardial T1 times were measured in six regions of interest in the myocardium (anterior, anterolateral, inferolateral, inferior, inferoseptal, and anterosseptal). We calculated the average T1 time of the six different region of interests. Areas of ischaemic focal fibrosis

identified by late gadolinium enhancement were excluded from the analysis. ECV was then computed according to the formula.¹⁹

Biomarkers analysis

Blood samples were obtained by venipuncture at inclusion. After centrifugation at 3500 rpm for 10 min, aliquots of serum and plasma were stored at -80°C . High-sensitivity troponinT (hsTnT), NT-proBNP, and parathyroid hormone (PTH) were measured with electrochemiluminescent two sites immunoassay on Cobas 8000 platform (Roche Diagnostics, Mannheim, Germany). C-terminal FGF-23 (referred to as FGF-23 in the text) concentrations were determined with a second-generation C-terminal human enzyme-linked immunosorbent assay enzyme-linked immunosorbent assay (Immutopics, San Clemente, CA, USA). Soluble ST2 (sST2) was measured using the Presage[®] ST2 enzyme-linked immunosorbent assay (Critical Diagnostics, CA, USA).

Follow-up

Patients were prospectively followed by ambulatory visits and phone calls at 6-month intervals. Clinical and survival status were obtained by follow-up visits and by phone contact with the patients, their relatives, or their physician. The primary endpoint was a composite of all-cause mortality or hospitalization for HF, whichever came first. HF hospitalization was defined as patients treated in the emergency room or admitted to a hospital, diagnosed with decompensated HF, and requiring IV diuretics. The secondary endpoint was all-cause mortality.

Statistical analysis

Statistical analyses were performed using SPSS version 22 (SPSS Corp., Somers, New York). All tests were two sided, and a $P < 0.05$ was considered statistically significant. Continuous variables were expressed as mean \pm 1 SD if normally distributed or as median (25th and 75th percentiles) if not normally distributed. Categorical variables were expressed as counts and percentages. Biomarkers were log-transformed to establish normality. Comparison between groups was performed using unpaired *t*-tests or chi-square test, when appropriate. The population of HFpEF patients was divided into three equal groups according to tertiles of FGF-23 levels. Clinical, biological, and imaging parameters were compared among those groups using *P* for trend analysis.

Event-free and overall survival of HFpEF patients was estimated using Kaplan–Meier method and Cox regression analysis. All baseline and imaging variables were initially proposed

for inclusion in a univariate Cox proportional hazard model. Significant variables ($P < 0.10$) were entered into a stepwise forward multivariate Cox regression model. To avoid collinearity, the correlation coefficients between covariates were examined. In case of collinearity ($r > 0.50$), only the strongest of the two covariates was proposed for inclusion into the multivariate model. Kaplan Meier curves based on tertiles of FGF-23 were used to illustrate event-free and overall survival of HFpEF patients.

Results

Baseline characteristics

One hundred forty-three consecutive HFpEF patients (78 ± 8 years, 61% female patients) and 31 controls of similar age and gender (75 ± 6 years, 61% female patients) were included in the study. The baseline demographic and imaging characteristics of HFpEF patients and controls of similar age and gender are presented in *Table 1*. As expected, HFpEF patients had a higher incidence of cardiovascular risk factors and co-morbidities compared with controls. They had lower haemoglobin and lower estimated glomerular filtration rate (eGFR). Median NT-proBNP and FGF-23 were significantly higher in HFpEF (NT-proBNP: 1261 [589; 2663] pg/mL; and FGF-23: 247 [115; 548] RU/mL, respectively) than in controls (NT-proBNP: 117 [73; 158] pg/mL; and FGF-23: 61 [51; 68] RU/mL) (all $P < 0.001$) (*Figure 1*). HFpEF patients had higher LA volumes, higher E wave velocities, higher E/e' ratios, higher LV masses, higher pulmonary pressures, more signs of RV dysfunction, and higher ECV values, likely reflecting more diffuse myocardial fibrosis than controls. FGF-23 levels were significantly correlated to ECV in the whole population (*Figure 2*).

Comparison of clinical and imaging parameters across tertiles of fibroblast growth factor 23 in heart failure with preserved ejection fraction patients

Heart failure and preserved ejection fraction (HFpEF) patients in the highest FGF-23 tertile were more often female patients and had higher incidence of atrial fibrillation. They had lower haemoglobin, worse renal function, higher NT-proBNP and hsTnT, higher PTH, and higher sST2. The rise of FGF-23 was independent of the 25OH vitamin D status. Regarding imaging parameters, they had greater LA volumes with worse RV systolic function (lower RV FAC $P = 0.027$, lower TAPSE $P = 0.030$, and lower RVEF by cMR $P = 0.003$). Interestingly, ECV values significantly increased across tertiles of FGF-23 (*Table 2*).

Table 1 Baseline characteristics of HFpEF patients and controls of similar age and gender

	Controls (n = 31)	HFpEF (n = 143)	P-value
Baseline characteristics			
Age (years)	75 ± 6	78 ± 8	0.06
Female (n, %)	19 (61%)	87 (61%)	0.96
Body mass index (kg/m ²)	26 ± 4	29 ± 6	0.015
Mean blood pressure (mmHg)	113 ± 16	95 ± 14	<0.001
NYHA functional classes III and IV (n, %)	0 (0%)	62 (43%)	<0.001
Medical history			
Atrial fibrillation (n, %)	0 (0%)	88 (62%)	<0.001
Ischaemic cardiomyopathy (n, %)	0 (0%)	50 (35%)	<0.001
Previous heart failure episode (n, %)	0 (0%)	103 (72%)	<0.001
Previous valvular surgery (n, %)	0 (0%)	16 (11%)	0.051
Chronic obstructive pulmonary disease (n, %)	0 (0%)	14 (10%)	0.070
Sleep apnoea (n, %)	0 (0%)	18 (13%)	0.037
Cardiovascular risk factors			
Smoking (n, %)	7 (23%)	61 (43%)	0.038
Hypertension (n, %)	19 (61%)	134 (94%)	<0.001
Diabetes (n, %)	6 (19%)	55 (38%)	0.044
Family history of CV disease (n, %)	5 (16%)	30 (21%)	0.5
Hypercholesterolemia (n, %)	26 (84%)	95 (66%)	0,056
Medication			
Loop diuretic (n, %)	0 (0%)	101 (71%)	<0.001
Thiazide (n, %)	2 (6%)	30 (21%)	0.059
Mineralocorticoid receptor antagonist (n, %)	0 (0%)	29 (20%)	0.006
Beta-blocker (n, %)	3 (10%)	95 (66%)	<0.001
ACEI or ARB (n, %)	13 (42%)	99 (69%)	0.004
Antiaggregant (n, %)	8 (26%)	59 (42%)	0.10
Oral anticoagulant (n, %)	1 (3%)	79 (55%)	<0.001
Statins (n, %)	8 (26%)	64 (45%)	0.053
Biology			
Haemoglobin (g/dL)	13.9 ± 1.4	11.7 ± 2.0	<0.001
Total cholesterol (mg/dL)	203 ± 45	154 ± 45	<0.001
GFR (mL/min/1.73 m ²) by CK-EPI	63 ± 15	49 ± 19	<0.001
NT-proBNP (pg/mL)	117 (73; 158)	1,261 (589; 2,663)	<0.001
Hs TnT (pg/mL)	7 (6; 11)	26 (15; 38)	<0.001
Iron (µg/dL)	94 ± 33	77 ± 60	0.15
Ferritin (µg/L)	158 ± 167	231 ± 281	0.17
Calcium (mmol/L)	2.47 ± 0.14	2.39 ± 0.30	0.15
Phosphorus (mmol/L)	1.08 ± 0.17	1.19 ± 0.26	0.017
Intact PTH (pg/mL)	43 ± 23	68 ± 53	0.011
25OH-Vitamin D (ng/mL)	33 ± 15	25 ± 16	0.016
Soluble ST2 (ng/mL)	24 (21; 31)	42 (31; 60)	<0.001
FGF-23 (RU/mL)	61 (51; 68)	247 (115; 548)	<0.001
Echo study			
LA volume index (mL/m ²)	20 ± 6	46 ± 19	<0.001
LV EDV index (mL/m ²)	59 ± 10	67 ± 18	0.024
LV ejection fraction (%)	65 ± 5	63 ± 7	0.055
LV Endo GLS (%)	-21.0 ± 2.5	-16.5 ± 3.2	<0.001
LV mass index (g/m ²)	71 ± 16	96 ± 26	<0.001
E wave velocity (m/s)	55 ± 10	94 ± 31	<0.001
Septal e' (m/s)	7.3 ± 1.8	6.9 ± 2.2	0.003
E/e' septal ratio	9.5 ± 1.8	19.4 ± 8.7	<0.001
RV/RA gradient (mmHg)	18 ± 5	33 ± 11	<0.001
RV fractional area change (%)	47 ± 7	41 ± 9	0.001
TAPSE (mm)	24 ± 4	19 ± 5	<0.001
cMR study			
	n = 31	n = 121	
LA volume index (mL/m ²)	31 ± 9	67 ± 29	<0.001
LV EDV index (mL/m ²)	64 ± 11	74 ± 19	0.009
LV ejection fraction (%)	66 ± 6	63 ± 8	0.030
LV mass index (g/m ²)	57 ± 12	68 ± 15	0.001
RV EDV index (mL/m ²)	68 ± 10	82 ± 27	0.005
RV ejection fraction (%)	61 ± 7	57 ± 9	0.001
ECV (%)	27.8 ± 2.4	32.7 ± 4.9	0.001
Late gadolinium enhancement (%)	0 (0%)	1.5 ± 2.6	0.001

CK-EPI, Chronic Kidney Disease Epidemiology Collaboration; ECV, extracellular volume; EDV, end-diastolic volume; FGF-23, fibroblast growth factor 23; GFR, glomerular filtration rate; GLS, global longitudinal strain; HFpEF, heart failure with preserved ejection fraction;

hsTnT, high-sensitivity troponinT; LA, left atrium; LV, left ventricle; NT-proBNP, N terminal pro brain natriuretic peptide; PTH, parathormon; RV, right ventricle; ST2, soluble suppression tumourigenicity 2; TAPSE, tricuspid annular plane systolic excursion. Values are mean \pm SD or median [IQR: 0.25; 0.75].

Outcomes

Over a mean follow-up of 30 ± 8 months, 43 patients (31%) died and 69 patients (49%) were hospitalized for HF. A total of 87 patients (62%) reached the primary composite endpoint of all-cause mortality or HF hospitalization, whichever came first.

In univariate Cox regression analysis, New York Heart Association class, diabetes, haemoglobin, eGFR, HsTnT, phosphorus, intact PTH, sST2, FGF-23, treatment with loop diuretics and thiazides, E wave velocity, and E/e' ratio were predictors of the primary endpoint. Among cMR parameters, index RV volume and ECV were also significantly associated with the primary endpoint. In multivariate Cox regression analysis, only FGF-23 (HR: 3.44 [2.01; 5.90], $P < 0.001$) and E wave velocity (HR: 1.01 [1.00; 1.02], $P = 0.034$) were independent predictors of the primary endpoint (Table 3).

For the secondary endpoint, BMI, eGFR, hsTnT, ferritin, sST2, FGF-23, mineralocorticoid receptor antagonist medication, RV/RA gradient, and ECV were predictors of all-cause mortality in univariate Cox regression. In multivariate Cox regression analysis, only ferritin (HR: 1.02 [1.01; 1.03], $P < 0.001$), FGF-23 (HR: 2.85 [1.26; 6.44], $P = 0.012$), and ECV (HR: 1.26 [1.03; 1.23], $P = 0.008$) were independent predictors of all-cause mortality (Table 4).

Figure 3A and 3B shows the Kaplan Meier curves for the primary and secondary endpoint of HFpEF patients according to tertiles of FGF-23, respectively, illustrating that patients with the highest FGF-23 levels have the worse prognosis.

Discussion

In the present study, we sought to examine the relationship between FGF-23 and baseline biomarkers, demographic, and imaging characteristics, particularly diffuse myocardial fibrosis, in controls and HFpEF patients and the prognostic value of FGF-23 in HFpEF.

In this very well sub-phenotyped cohort of controls and HFpEF patients, FGF-23 was significantly higher in HFpEF patients compared with controls. FGF-23 was associated with some proinflammatory co-morbidities (i.e. renal dysfunction, diabetes, and atrial fibrillation), indices of cardiac dysfunction (i.e. LA dilatation and RV dysfunction), and biomarkers reflective of either ventricular stretch (NT-proBNP), myocardial necrosis (hsTnT), fibrosis (sST2), or bone and mineral metabolism (PTH). Moreover, FGF-23 was significantly correlated with myocardial fibrosis estimated by ECV. FGF-23 was

Figure 1 Box-plot of FGF-23 in heart failure with preserved ejection fraction (HFpEF) and controls of similar age and gender.

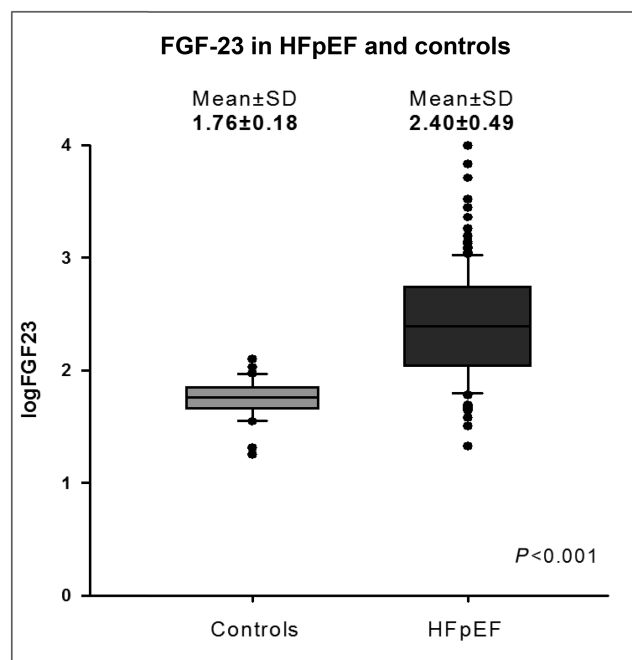
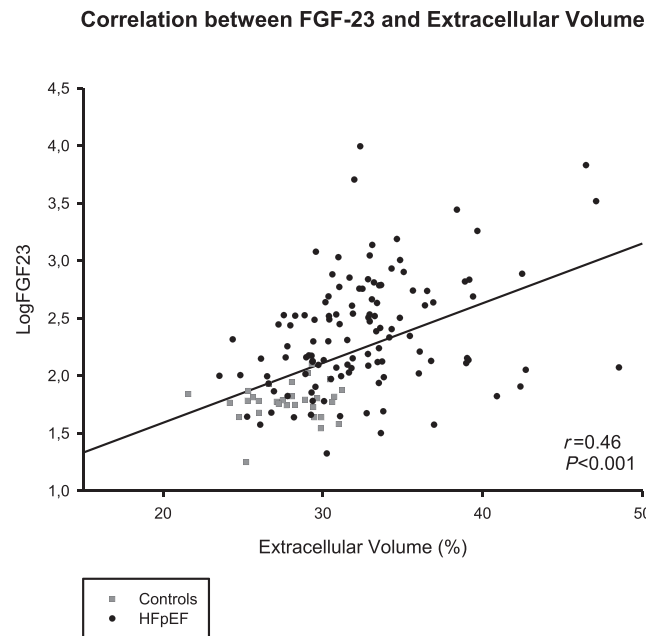


Figure 2 Correlation between fibroblast growth factor 23 (FGF-23) and extracellular volume (ECV) in heart failure with preserved ejection fraction (HFpEF) and controls of similar age and gender.



also a strong and independent predictor of the primary composite endpoint and the secondary endpoint of all-cause mortality, even after adjusting for confounding factors such as renal function and other clinically relevant variables. Interestingly, both the interplay with the other biomarkers and with the prognosis value of FGF-23 was independent of the vitamin D levels.

Mechanistic insight into the pathogenesis of disease

In HF, biomarkers are used primarily for diagnosis and risk stratification, natriuretic peptides playing a major role in this respect.²⁰ Nonetheless, biomarkers studies can also provide important insight into the pathophysiologic mechanisms leading to disease progression, which can then be targeted pharmacologically, such as the RAAS or the adrenergic system. However, this aspect of biomarker research is complicated by the fact that many biomarkers predictive of poor prognosis may reflect systemic organ failures rather than a specific mechanism underlying cardiac disease progression. Moreover, the plasma levels of most biomarkers are affected by multiple confounding factors such as age, sex, renal, and hepatic function. This complexity likely explains why progress has been slow in understanding the mechanisms responsible for the progression of HFpEF, as in this disease, LV systolic function remains by definition

preserved to the very end while systemic organ failures inexorably progress.

Heart failure and preserved ejection fraction (HFpEF) is a clinical syndrome that has already been associated with changes in the extracellular matrix and in which fibrosis seems to be a crucial component of cardiac remodelling.¹ Among all the regulators involved in the pathophysiology of cardiac fibrosis, FGF-23 seems to play an important role, probably still underestimated and understudied. Only few studies looked at the prognostic value of this biomarker in HFpEF, although experimental and in vivo data demonstrated the implication of FGF-23 in pathophysiological mechanisms involved in the development of the disease such as cardiac hypertrophy, fibrosis, angiogenesis, and cardiac remodelling.^{21–24} Indeed, in our study, we found a significant correlation between FGF-23 levels and diffuse myocardial fibrosis estimated by ECV. Whether the association between FGF-23 and ECV is causal cannot be concluded from our observational data, but previous studies demonstrated the implication of FGF-23 in pathophysiological mechanisms involved cardiac remodelling.^{22–24} It was demonstrated that FGF-23 promotes hypertrophic growth of cardiac myocytes via FGF receptor 4²¹ and that it contributes to myocardial fibrosis and diastolic dysfunction through the up-regulation of active β -catenin and TGF- β .²⁵ It was also shown that FGF-23 directly stimulates RAAS through the inhibition of ACE2²¹ and that the interplay of FGF-23 with PTH and RAAS might trigger adverse cardiovascular remodelling.^{26,27} Those data taken together suggest that FGF-23 is not only a marker

Table 2 Clinical and imaging parameters according to tertiles of FGF-23 in HFpEF

	FGF-23			P-for trend
	Tertile 1 n = 47 (<134 RU/mL)	Tertile 2 n = 47 (134–406 RU/mL)	Tertile 3 n = 48 (>406 RU/mL)	
Baseline characteristics				
Age (years)	78 ± 9	78 ± 8	79 ± 9	0.57
Female (n, %)	20 (43%)	33 (69%)	34 (71%)	0.005
Body mass index (kg/m ²)	28 ± 5	30 ± 7	27 ± 6	0.51
Mean blood pressure (mmHg)	97 ± 15	98 ± 11	90 ± 13	0.021
NYHA functional classes III and IV (n, %)	18 (38%)	19 (40%)	25 (52%)	0.18
Medical history				
Atrial fibrillation (n, %)	22 (47%)	30 (63%)	36 (75%)	0.005
Ischaemic cardiomyopathy (n, %)	18 (38%)	14 (29%)	18 (38%)	0.94
Previous heart failure episode (n, %)	29 (62%)	37 (77%)	37 (77%)	0.097
Previous valvular surgery (n, %)	5 (11%)	4 (8%)	7 (15%)	0.54
Chronic obstructive pulmonary disease (n, %)	5 (11%)	6 (13%)	3 (6%)	0.47
Sleep apnoea (n, %)	4 (9%)	8 (17%)	6 (13%)	0.57
Cardiovascular risk factors				
Smoking (n, %)	24 (51%)	17 (35%)	20 (42%)	0.36
Hypertension (n, %)	41 (87%)	47 (98%)	46 (96%)	0.087
Diabetes (n, %)	15 (32%)	17 (35%)	23 (48%)	0.11
Family history of CV disease (n, %)	12 (26%)	8 (17%)	10 (21%)	0.58
Hypercholesterolemia (n, %)	29 (62%)	32 (67%)	34 (71%)	0.35
Medication				
Loop diuretic (n, %)	27 (57%)	35 (73%)	40 (85%)	0.011
Thiazide (n, %)	13 (27%)	11 (23%)	6 (13%)	0.25
Mineralocorticoid receptor antagonist (n, %)	5 (10%)	10 (21%)	14 (30%)	0.06
Beta-blocker (n, %)	31 (65%)	33 (69%)	32 (68%)	0.90
ACEI or ARB (n, %)	33 (69%)	33 (69%)	32 (68%)	0.99
Antiaggregant (n, %)	23 (48%)	22 (46%)	14 (30%)	0.15
Oral anticoagulant (n, %)	20 (42%)	28 (58%)	32 (68%)	0.032
Statins (n, %)	24 (51%)	23 (49%)	17 (38%)	0.40
Biology				
Haemoglobin (g/dL)	12.6 ± 2.1	11.9 ± 1.7	10.8 ± 1.6	<0.001
GFR (mL/min/1.73 m ²) by CKD-EPI	60 ± 20	44 ± 17	44 ± 18	<0.001
NT-proBNP (pg/mL)	750 (360; 1289)	1201 (628; 2177)	2441 (1013; 4075)	<0.001
Hs TnT (pg/mL)	16 (12; 31)	28 (18; 37)	33 (18; 42)	<0.001
Iron (µg/dL)	84 ± 50	77 ± 49	71 ± 78	0.29
Ferritin (µg/L)	300 ± 360	230 ± 198	164 ± 254	0.021
Calcium (mmol/L)	2.35 ± 0.41	2.45 ± 0.23	2.37 ± 0.23	0.76
Phosphorus (mmol/L)	1.13 ± 0.15	1.23 ± 0.28	1.22 ± 0.31	0.078
Intact PTH (pg/mL)	50 ± 28	71 ± 39	84 ± 76	0.003
25OH-Vitamin D (ng/mL)	23 ± 15	27 ± 16	26 ± 18	0.39
Soluble ST2 (ng/mL)	35 (26; 56)	42 (32; 57)	47 (36; 69)	0.027
FGF-23 (RU/mL)	89 (60; 110)	245 (160; 311)	701 (549; 1131)	<0.001
Echo study				
LA volume index (mL/m ²)	44 ± 21	41 ± 12	52 ± 20	0.030
LV EDV index (mL/m ²)	69 ± 16	64 ± 16	68 ± 22	0.63
LV ejection fraction (%)	62 ± 7	63 ± 8	63 ± 8	0.58
LV Endo GLS (%)	-16.3 ± 3.5	-16.5 ± 3.1	-16.8 ± 2.9	0.45
LV mass index (g/m ²)	97 ± 26	92 ± 22	99 ± 31	0.79
E wave velocity (m/s)	89 ± 32	91 ± 29	101 ± 31	0.068
Septal e' (m/s)	5.1 ± 1.4	5.2 ± 1.4	5.2 ± 1.3	0.93
E/e' septal ratio	18.8 ± 9.2	18.5 ± 7.8	20.8 ± 9.2	0.27
RV/RA gradient (mmHg)	31 ± 9	32 ± 11	34 ± 12	0.17
RV/RA gradient >37 mmHg	14 (30%)	14 (29%)	18 (38%)	0.42
RV fractional area change (%)	44 ± 7	40 ± 10	40 ± 9	0.027
RV fractional area change <35% (n, %)	4 (5%)	12 (15%)	29 (35%)	<0.001
TAPSE (mm)	20 ± 5	19 ± 5	17 ± 6	0.030
TAPSE <17 mm (n, %)	6 (7%)	14 (17%)	42 (51%)	<0.001
cMR study				
LA volume index (mL/m ²)	62 ± 36	64 ± 21	76 ± 26	0.030
LV EDV index (mL/m ²)	76 ± 20	70 ± 18	74 ± 19	0.62
LV ejection fraction (%)	63 ± 8	62 ± 9	63 ± 7	0.94
LV mass index (g/m ²)	69 ± 15	65 ± 12	69 ± 18	0.82
RV EDV index (mL/m ²)	75 ± 20	78 ± 22	93 ± 34	0.003
RV ejection fraction (%)	59 ± 7	57 ± 8	54 ± 9	0.003
RV ejection fraction <45% (n, %)	5 (6%)	5 (6%)	11 (17%)	0.029
ECV (%)	31.7 ± 5.2	31.4 ± 3.3	35.5 ± 5.0	<0.001
Late gadolinium enhancement (%)	1.1 ± 2.6	1.7 ± 2.3	1.8 ± 2.8	0.26

CK-EPI, Chronic Kidney Disease Epidemiology Collaboration; ECV, extracellular volume; EDV, end-diastolic volume; FGF-23, fibroblast growth factor 23; GFR, glomerular filtration rate; GLS, global longitudinal strain; HFpEF, heart failure with preserved ejection fraction; hsTnT, high-sensitivity troponinT; LA, left atrium; LV, left ventricle; NT-proBNP, N terminal pro brain natriuretic peptide; PTH, parathormon; RV, right ventricle; ST2, soluble suppression tumourigenicity 2; TAPSE, tricuspid annular plane systolic excursion. Values are mean \pm SD or median [IQR 0.25; 0.75]. *P*-value obtained by *P* for trend analysis.

of risk but might contribute to the progression of the disease. The availability of FGF-23 antagonist or immunotherapy would allow testing that hypothesis.^{25,28} Sodium glucose

cotransporter-2 inhibitors, the most recently approved class of drug for type 2 diabetes, has recently shown benefit in HF patients, independently from its effect on diabetes.²⁹

Table 3 Cox regression analysis for the primary endpoint

Cox regression analysis Combined events	Univariate		Multivariate
	Hazard ratio 95% CI	<i>P</i> -value	
Age	1.01 [0.98; 1.04]	0.58	
Female	1.46 [0.94; 2.28]	0.088	
Body mass index	0.97 [0.94; 1.01]	0.10	
Mean blood pressure	0.99 [0.98; 1.01]	0.31	
NYHA functional classes III and IV	1.58 [1.03; 2.41]	0.037	
Cardiovascular risk factors			
Diabetes	1.65 [1.08; 2.52]	0.022	
Medical history			
Atrial fibrillation	1.25 [0.80; 1.95]	0.33	
Ischaemic cardiomyopathy	0.93 [0.60; 1.45]	0.76	
Medication			
Loop diuretic	2.16 [1.25; 3.72]	0.005	
Thiazide	0.55 [0.30; 0.99]	0.048	
Mineralocorticoid receptor antagonist	1.38 [0.85; 2.15]	0.20	
Beta-blocker	0.97 [0.62; 1.52]	0.90	
ACEI or ARB	0.90 [0.57; 1.40]	0.63	
Oral anticoagulant	1.46 [0.95; 2.27]	0.088	
Biology			
Haemoglobin (g/dL)	0.86 [0.76; 0.97]	0.013	
GFR (mL/min/1.73 m ²) by CKD-EPI	0.98 [0.97; 0.99]	<0.001	
NT-proBNP (pg/mL)	1.19 [0.97; 1.46]	0.10	
Hs TnT (pg/mL)	1.70 [1.25; 2.30]	0.001	
Iron (ug/dL)	1.00 [1.00; 1.00]	0.38	
Ferritin (ug/L)	1.00 [1.00; 1.00]	0.10	
Calcium (mmol/L)	0.89 [0.47; 1.68]	0.72	
Phosphorus (mmol/L)	2.31 [1.04; 5.14]	0.051	
Intact PTH (pf/mL)	1.01 [1.00; 1.01]	0.003	
25OH-Vitamin D (ng/mL)	1.00 [0.99; 1.01]	0.91	
Soluble ST2 (ng/mL)	3.46 [1.23; 9.74]	0.020	
FGF-23 (RU/mL)	2.21 [1.52; 3.22]	<0.001	3.15 [1.88; 5.31]
Echo study			
LA volume index (mL/m ²)	1.01 [1.00; 1.02]	0.17	
LV ejection fraction (%)	1.01 [0.98; 1.04]	0.69	
LV Endo GLS (%)	1.00 [0.93; 1.07]	0.95	
E wave velocity (m/s)	1.01 [1.00; 1.02]	0.002	1.01 [1.00; 1.02]
E/e' septal ratio	1.03 [1.01; 1.05]	0.018	
RV/RA gradient (mmHg)	1.02 [1.00; 1.04]	0.093	
RV fractional area change (%)	0.27 [0.02; 3.11]	0.29	
TAPSE	0.97 [0.93; 1.01]	0.13	
cMR study			
LA volume index (mL/m ²)	1.00 [1.00; 1.01]	0.31	
LV EDV index (mL/m ²)	1.00 [0.99; 1.02]	0.83	
LV ejection fraction (%)	1.01 [0.98; 1.04]	0.55	
LV mass index (g/m ²)	1.00 [0.99; 1.02]	0.64	
RV EDV index (mL/m ²)	1.01 [1.00; 1.02]	0.031	
RV ejection fraction (%)	0.98 [0.95; 1.01]	0.11	
Extracellular volume (%)	1.10 [1.05; 1.15]	<0.001	
Late gadolinium enhancement	1.09 [0.99; 1.20]	0.11	

ACEi, angiotensin-converting-enzyme inhibitor; ARB, Angiotensin II receptor blockers; ECV, extracellular volume; EDV, end-diastolic volume; FGF-23, fibroblast growth factor 23; GFR, glomerular filtration rate; GLS, global longitudinal strain; hsTnT, high-sensitivity troponinT; LA, left atrium; LV, left ventricle; NT-proBNP, N terminal pro brain natriuretic peptide; PTH, parathormon; RV, right ventricle; ST2, soluble suppression tumourigenicity 2; TAPSE, tricuspid annular plane systolic excursion.

Table 4 Cox regression analysis for the secondary endpoint

Cox regression analysis	Univariate		Multivariate	
	Hazard ratio 95% CI	P-value	Hazard ratio 95% CI	P-value
All-cause mortality				
Age	1.03 [0.99; 1.07]	0.13		
Female	1.18 [0.64; 2.20]	0.60		
Body mass index	0.91 [0.86; 0.96]	<0.001		
Mean blood pressure	0.98 [0.96; 1.00]	0.10		
NYHA functional classes III and IV	1.12 [0.61; 2.06]	0.72		
Cardiovascular risk factors				
Diabetes	1.53 [0.84; 2.78]	0.17		
Medical history				
Atrial fibrillation	0.85 [0.46; 1.56]	0.60		
Ischaemic cardiomyopathy	0.67 [0.34; 1.31]	0.23		
Medication				
Loop diuretic	1.71 [0.79; 3.69]	0.17		
Thiazide	0.79 [0.35; 1.79]	0.57		
Mineralocorticoid receptor antagonist	2.12 [1.12; 4.02]	0.021		
Beta-blocker	0.72 [0.39; 1.32]	0.28		
ACEI or ARB	0.62 [0.34; 1.15]	0.13		
Oral anticoagulant	1.19 [0.65; 2.20]	0.58		
Biology				
Haemoglobin	0.86 [0.73; 1.01]	0.068		
GFR	0.98 [0.97; 1.00]	0.045		
NT-proBNP	1.30 [0.98; 1.73]	0.078		
Hs TnT	1.62 [1.12; 2.34]	0.014		
Iron	1.00 [1.00; 1.01]	0.09		
Ferritin	1.00 [1.00; 1.00]	0.004	1.02 [1.01; 1.03]	<0.001
Calcium	0.85 [0.40; 1.78]	0.67		
Phosphorus	1.93 [0.64; 5.84]	0.26		
Intact PTH	1.00 [0.99; 1.01]	0.79		
25OH-Vitamin D	0.99 [0.97; 1.01]	0.25		
Soluble ST2	20.24 [4.88; 84.03]	<0.001		
FGF-23	2.12 [1.26; 3.67]	0.010	2.85 [1.26; 6.44]	0.012
Echo study				
LA volume index	1.01 [1.00; 1.02]	0.20		
LV ejection fraction	1.01 [0.97; 1.06]	0.53		
LV Endo GLS	1.04 [0.94; 1.15]	0.42		
E wave velocity	1.01 [1.00; 1.02]	0.11		
E/e' septal ratio	1.03 [1.00; 1.06]	0.084		
RV/RA gradient	1.03 [1.01; 1.06]	0.015		
RV fractional area change	0.16 [0.01; 4.88]	0.29		
TAPSE	0.97 [0.91; 1.02]	0.25		
cMR study				
LA volume index (mL/m ²)	1.00 [0.99; 1.01]	0.71		
LV EDV index (mL/m ²)	1.01 [0.99; 1.03]	0.26		
LV ejection fraction (%)	0.99 [0.94; 1.03]	0.60		
LV mass index (g/m ²)	1.02 [1.00; 1.04]	0.15		
RV EDV index (mL/m ²)	1.00 [0.99; 1.02]	0.66		
RV ejection fraction (%)	0.98 [0.93; 1.03]	0.39		
Extracellular volume (%)	1.12 [1.05; 1.20]	0.003	1.26 [1.03; 1.23]	0.008
Late gadolinium enhancement	1.13 [1.00; 1.28]	0.069		

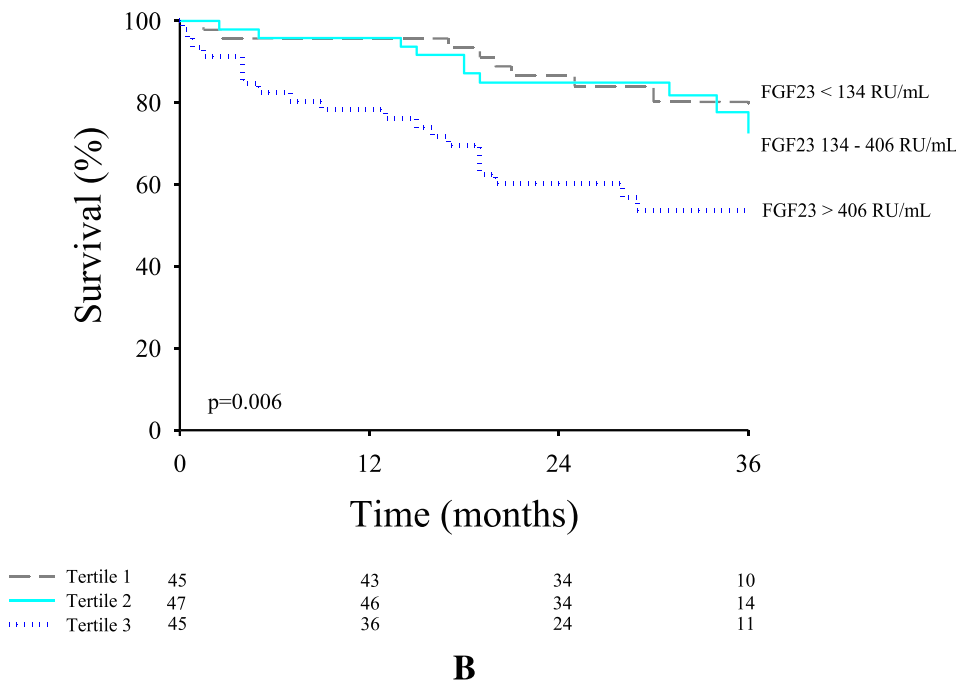
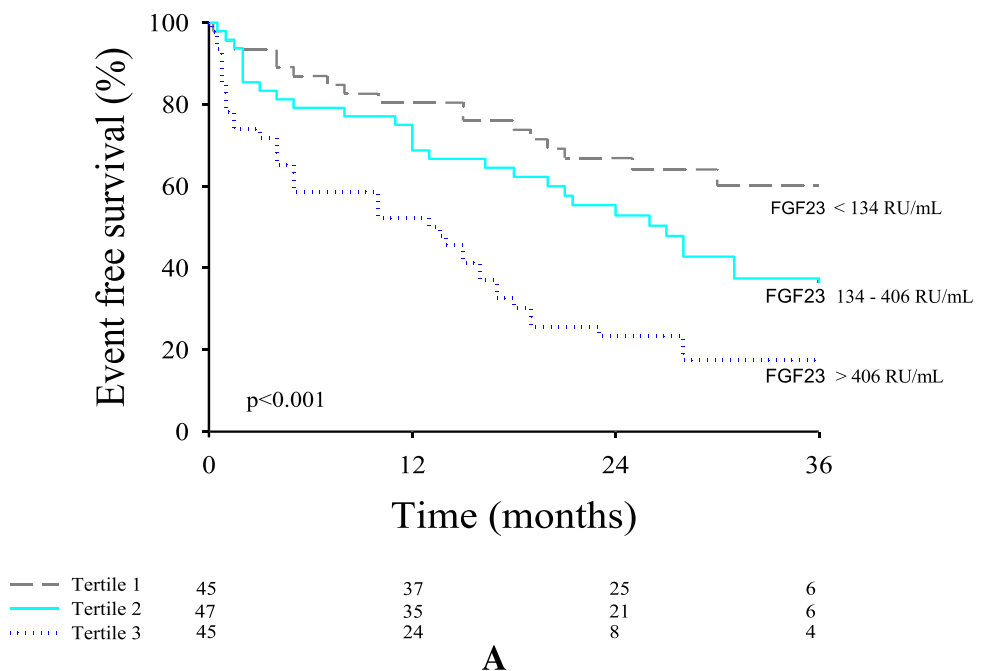
ACEi, angiotensin-converting-enzyme inhibitor; ARB, Angiotensin II receptor blockers; ECV, extracellular volume; EDV, end-diastolic volume; FGF-23, fibroblast growth factor 23; GFR, glomerular filtration rate; GLS, global longitudinal strain; hsTnT, high-sensitivity troponinT; LA, left atrium; LV, left ventricle; NT-proBNP, N terminal pro brain natriuretic peptide; PTH, parathormon; RV, right ventricle; ST2, soluble suppression tumourigenicity 2; TAPSE, tricuspid annular plane systolic excursion.

One hypothesis to explain this effect is that canagliflozin induces a prompt increase in serum phosphorus and triggers downstream changes in FGF-23.^{30,31} Studies (DELIVER and EMPEROR-Preserved) are currently going on to determine whether sodium glucose cotransporter-2 inhibitors will have a positive impact on the prognosis of HFpEF patients.³²

Association between fibroblast growth factor 23 levels and clinical characteristics

Although the mechanisms of sex differences in FGF-23 remain unclear, the presence of higher FGF-23 levels in female patients than in male patients has been described in previous studies, both in small children³³ and in adults with

Figure 3 Kaplan Meier curves for the primary endpoint according to tertiles of fibroblast growth factor 23 (FGF-23) in heart failure with preserved ejection fraction (HFpEF) patients (A) and Kaplan Meier curves for the secondary endpoint according to tertiles of FGF-23 in HFpEF patients (B).



cardiovascular risk factors.^{34,35} In the hypothesis that FGF-23 actively plays a part in the development of the disease, this might contribute to the overrepresentation of women among patients with HFpEF.

Atrial fibrillation, a highly prevalent co-morbidity in HFpEF, was also found to be associated with higher levels of FGF-

23.^{36–38} Two possible mechanisms were described to explain this interaction. First, FGF-23 could induce atrial fibrosis by increasing ROS production, subsequently activating STAT3 and SMAD3 signalling.³⁹ Second, FGF-23 increases pulmonary vein arrhythmogenesis through protein kinase C signalling and dysregulation of sodium and calcium homeostasis.⁴⁰

The association between elevated FGF-23 and impaired renal function is largely described. Levels of FGF-23 rise early in the course of CKD as part of the adaptive response to maintain neutral phosphate balance when renal excretory capacity declines. Chronic FGF-23 elevation in CKD is independently associated with development of HF and death.^{9,11} We also observed that haemoglobin decreased across increasing FGF-23 levels. This is not only mediated through the interaction with renal function (where anaemia is mediated by decreased erythropoietin production, low serum active vitamin D levels, and high renin-angiotensin-aldosterone activities) but also through a direct inhibition of erythropoiesis by FGF-23.⁴¹

Recent data from PARAMOUNT⁴² and RELAX³⁵ trials showed that sST2 levels were correlated with proinflammatory co-morbidities and with the severity of HFpEF, indicated by higher levels of NT-proBNP and signs of diastolic dysfunction. Authors from those trials concluded that patients with the more severe disease had a biomarker pattern associated with a more profibrotic milieu.⁴² This is corroborated by our study, where FGF-23 is associated both with the presence of myocardial fibrosis evaluated by ECV and with signs of disease severity (NT-proBNP, renal function, and RV dysfunction). On the other hand, we did not find a significant association between FGF-23 levels and E wave velocities or E/e' ratio. This is consistent with a recent study by Okamoto *et al.*²⁴ showing that alpha-klotho, a co-receptor of FGF-23 related to ageing suppression and organ protection, was significantly associated with E/e' ratio, while FGF-23 was not. However, in our study, higher FGF-23 levels were associated with larger LA volumes, which is also an important marker of elevated filling pressures and diastolic dysfunction.⁴³

Prognostic and risk stratification

Beyond assessment of natriuretic peptides, other biomarkers of mechanisms contributing to the pathophysiology of HF, such as inflammation (GDF-15 and sST2), LV hypertrophy (FGF-23), and myocardial necrosis (hsTnT), could add important prognostic information and identify patients at higher risk of events.^{44–47} Studies have already demonstrated the superiority of different biomarkers combinations, including NT-proBNP, sST2, and Galectin-3 for risk stratification of chronic HF patients.^{48,49} In 2015, we showed the additive value of FGF-23, Galectin 3, and sST2 compared with NT-proBNP for prediction of cardiovascular death in HFREF.⁵⁰ However, in the specific CORONA-HF population, a multimarker approach using a panel of 20 inflammatory and extracellular matrix biomarkers (including sST2 and galectin-3) was of limited clinical value for identifying the risk of adverse outcomes.⁵¹

Although FGF-23 has been associated with adverse cardiovascular outcomes in CKD and HFREF, data are lacking in

HFpEF. Koller *et al.*¹⁴ demonstrated that FGF-23 was independently associated with an increased risk of mortality in patients with HFREF ($n = 511$) but not in those with HFpEF ($n = 469$). Only one large trial showed a significant prognostic value of FGF-23 compared with NT-proBNP for a combined outcome of all-cause mortality and HF hospitalization in HFREF and in HFpEF.⁷ Our data corroborate the prognostic value of FGF-23 in HFpEF, demonstrating the importance of exploring physiopathological pathways for a better understanding of this syndrome. Surprisingly, NTproBNP was not associated with the primary and the secondary endpoints in our study (borderline for the prediction of mortality HR: 1.30 [0.98; 1.73] $P = 0.078$), probably mainly due to the high levels of NT-proBNP on average and the limited sample size.

Fibroblast growth factor 23 (FGF-23) might increase fluid retention by stimulating RAAS, probably explaining the link between FGF-23 and HF hospitalization (in univariate Cox regression analysis: HR 2.30 [1.51–3.49], $P < 0.001$) and might represent a potential therapeutic target. Even after adjustment for eGFR, FGF-23 remained predictive in our population. However, the strong predictive value of FGF-23 in our study might be related to the assay we used, and it might be hypothesized that assays targeting C-terminal FGF-23 fragments have stronger value for risk estimation than intact assays.⁵²

It could also be hypothesized that the rise of FGF-23 precedes the development of HF in patients at high risk, such as diabetic and hypertensive patients.^{53–55} The early sub-phenotyping of these patients including measurement of FGF-23 could represent an opportunity to precociously estimate their risk for developing HF and adjust their treatment accordingly. Further studies addressing the value of FGF-23 for risk stratification in this specific population are needed to confirm this hypothesis.

Strengths and limitations

To our knowledge, this is one of the first prospective study investigating the role of FGF-23 in a cohort of HFpEF patients and in a control group well characterized by echocardiography and cMR. Our cohort is comparable with PARAMOUNT⁴² and RELAX³⁵ biomarker analysis in terms of number of patients and demographic characteristics. However, this should be considered as exploratory and hypothesis generating and should be tested in larger mechanistic HFpEF studies.

Conclusions

Heart failure with preserved ejection fraction (HFpEF) patients showed significantly higher levels of circulating FGF-23 than controls. High FGF-23 levels were significantly

associated with LA volume (a marker of diastolic dysfunction), RV dysfunction, and with diffuse myocardial fibrosis evaluated by cMR. FGF-23 was also a strong predictor of poor outcome, even after adjusting for usual confounding factors. Sub-phenotyping of HFpEF patients with FGF-23 might therefore participate in a more personalized care.

Conflict of interest

The Cliniques St. Luc UCL has a master clinical research agreement with Philips Medical Instruments, and the MOLLI patch was supplied by Philips Medical under the terms of this agreement. The authors declare that they have no competing interests.

Funding

This work was funded by a grant of the Fondation Nationale de la Recherche Scientifique of the Belgian Government (FRMS CDR 23597851). Dr Pouleur is and Dr Beauloye was supported by a Post-doctorate Clinical Master Specialist of the Fondation Nationale de la Recherche Scientifique of the Belgian Government (FRSM: SPD 10844948). Dr Horman is a researcher associated of FNRS. Dr Roy is supported by Fondation Damman and Fondation Saint Luc for her fellowship. This work was funded by an unrestricted grant from AstraZeneca.

References

- Paulus WJ, Tschope C. A novel paradigm for heart failure with preserved ejection fraction: comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial inflammation. *J Am Coll Cardiol* 2013; **62**: 263–271.
- Duca F, Kammerlander AA, Zotter-Tufaro C, Aschauer S, Schwaiger ML, Marzluf BA, Bonderman D, Mascherbauer J. Interstitial fibrosis, functional status, and outcomes in heart failure with preserved ejection fraction: insights from a prospective cardiac magnetic resonance imaging study. *Circ Cardiovasc Imaging* 2016; **9**: e005277
- Kato S, Saito N, Kirigaya H, Gytoku D, Iinuma N, Kusakawa Y, Iguchi K, Nakachi T, Fukui K, Futaki M, Iwasawa T, Taguri M, Kimura K, Umemura S. Prognostic significance of quantitative assessment of focal myocardial fibrosis in patients with heart failure with preserved ejection fraction. *Int J Cardiol* 2015; **191**: 314–319.
- Mohammed SF, Hussain S, Mirzoyev SA, Edwards WD, Maleszewski JJ, Redfield MM. Coronary microvascular rarefaction and myocardial fibrosis in heart failure with preserved ejection fraction. *Circulation* 2015; **131**: 550–559.
- Roy C, Slimani A, de Meester C, Amzulescu M, Pasquet A, Vancraeynest D, Beauloye C, Vanoverschelde JL, Gerber BL, Pouleur AC. Associations and prognostic significance of diffuse myocardial fibrosis by cardiovascular magnetic resonance in heart failure with preserved ejection fraction. *J Cardiovasc Magn Reson* 2018; **20**: 55.
- Gruson D, Lepoutre T, Ketelslegers JM, Cumps J, Ahn SA, Rousseau MF. C-terminal FGF23 is a strong predictor of survival in systolic heart failure. *Peptides* 2012; **37**: 258–262.
- Ter Maaten JM, Voors AA, Damman K, van der Meer P, Anker SD, Cleland JG, Dickstein K, Filippatos G, van der Harst P, Hillege HL, Lang CC. Fibroblast growth factor 23 is related to profiles indicating volume overload, poor therapy optimization and prognosis in patients with new-onset and worsening heart failure. *Int J Cardiol* 2018; **253**: 84–90.
- Poelzl G, Trenkler C, Kliebhan J, Wuertinger P, Seger C, Kaser S, Mayer G, Pirklbauer M, Ulmer H, Griesmacher A. FGF23 is associated with disease severity and prognosis in chronic heart failure. *Eur J Clin Invest* 2014; **44**: 1150–1158.
- Niizuma S, Iwanaga Y, Yahata T, Miyazaki S. Renocardiovascular biomarkers: from the perspective of managing chronic kidney disease and cardiovascular disease. *Frontiers in cardiovascular medicine* 2017; **4**: 10.
- Faul C, Amaral AP, Oskoueï B, Hu MC, Sloan A, Isakova T, Gutiérrez OM, Aguillon-Prada R, Lincoln J, Hare JM, Mundel P, Morales A, Scialla J, Fischer M, Soliman EZ, Chen J, Go AS, Rosas SE, Nessel L, Townsend RR, Feldman HI, St. John Sutton M, Ojo A, Gadegbeku C, di Marco GS, Reuter S, Kentrup D, Tiemann K, Brand M, Hill JA, Moe OW, Kuro-o M, Kusek JW, Keane MG, Wolf M. FGF23 induces left ventricular hypertrophy. *J Clin Invest* 2011; **121**: 4393–4408.
- Isakova T, Xie H, Yang W, Xie D, Anderson AH, Scialla J, Wahl P, Gutiérrez OM, Steigerwalt S, He J, Schwartz S, Lo J, Ojo A, Sondheimer J, Hsu CY, Lash J, Leonard M, Kusek JW, Feldman HI, Wolf M, Chronic Renal Insufficiency Cohort (CRIC) Study Group. Fibroblast growth factor 23 and risks of mortality and end-stage renal disease in patients with chronic kidney disease. *JAMA* 2011; **305**: 2432–2439.
- Scialla JJ, Xie H, Rahman M, Anderson AH, Isakova T, Ojo A, Zhang X, Nessel L, Hamano T, Grunwald JE, Raj DS, Yang W, He J, Lash JP, Go AS, Kusek JW, Feldman H, Wolf M, the Chronic Renal Insufficiency Cohort (CRIC) Study Investigators. Fibroblast growth factor-23 and cardiovascular events in CKD. *Journal of the American Society of Nephrology: JASN* 2014; **25**: 349–360.
- Lutsey PL, Alonso A, Selvin E, Pankow JS, Michos ED, Agarwal SK, Loefer LR, Eckfeldt JH, Coresh J. Fibroblast growth factor-23 and incident coronary heart disease, heart failure, and cardiovascular mortality: the atherosclerosis risk in communities study. *J Am Heart Assoc* 2014; **3**: e000936.
- Koller L, Kleber ME, Brandenburg VM, Golasch G, Richter B, Sulzgruber P, Scharnagl H, Silbernagel G, Grammer TB, Delgado G, Tomaschitz A, Pilz S, Berger R, Mörtl D, Hülsmann M, Pacher R, März W, Niessner A. Fibroblast growth factor 23 is an independent and specific predictor of mortality in patients with heart failure and reduced ejection fraction. *Circ Heart Fail* 2015; **8**: 1059–1067.
- Plischke M, Neuhold S, Adlbrecht C, Bielez B, Shayganfar S, Bieglmayer C, Szekeres T, Hörl WH, Strunk G, Vavken P, Pacher R, Hülsmann M. Inorganic phosphate and FGF-23 predict outcome in stable systolic heart failure. *Eur J Clin Invest* 2012; **42**: 649–656.
- von Jeinsen B, Sopova K, Palapies L, Leistner DM, Fichtlscherer S, Seeger FH, Honold J, Dimmeler S, Aßmus B, Zeiher AM, Keller T. Bone marrow and

- plasma FGF-23 in heart failure patients: novel insights into the heart-bone axis. *ESC heart failure* 2019; **6**: 536–544.
17. Chua W, Purmah Y, Cardoso VR, Gkoutos GV, Tull SP, Neculau G, Thomas MR, Kotecha D, Lip GYH, Kirchhof P, Fabritz L. Data-driven discovery and validation of circulating blood-based biomarkers associated with prevalent atrial fibrillation. *Eur Heart J* 2019; **40**: 1268–1276.
 18. Roy C, Slimani A, de Meester C, Amzulescu M, Pasquet A, Vancraeynest D, Vanoverschelde JL, Pouleur AC, Gerber BL. Age and sex corrected normal reference values of T1, T2 T2* and ECV in healthy subjects at 3T CMR. *J Cardiovasc Magn Reson* 2017; **19**: 72.
 19. Kellman P, Wilson JR, Xue H, Ugander M, Arai AE. Extracellular volume fraction mapping in the myocardium, part 1: evaluation of an automated method. *J Cardiovasc Magn Reson* 2012; **14**: 63.
 20. Kristensen SL, Jhund PS, Kober L, McKelvie RS, Zile MR, Anand IS, Komajda M, Cleland JG, Carson PE, McMurray JJ. Relative importance of history of heart failure hospitalization and N-terminal pro-b-type natriuretic peptide level as predictors of outcomes in patients with heart failure and preserved ejection fraction. *JACC Heart failure* 2015; **3**: 478–486.
 21. Leifheit-Nestler M, Haffner D. Paracrine effects of FGF23 on the heart. *Front Endocrinol* 2018; **9**: 278.
 22. Hao H, Li X, Li Q, Lin H, Chen Z, Xie J, Xuan W, Liao W, Bin J, Huang X, Kitakaze M, Liao Y. FGF23 promotes myocardial fibrosis in mice through activation of beta-catenin. *Oncotarget* 2016; **7**: 64649–64664.
 23. Faul C. Cardiac actions of fibroblast growth factor 23. *Bone* 2016; **100**: 69–79.
 24. Okamoto Y, Fujita S, Morita H, Kizawa S, Ito T, Sakane K, Sohmiya K, Hoshiga M, Ishizaka N. Association between circulating FGF23, alpha-klotho, and left ventricular diastolic dysfunction among patients with preserved ejection fraction. *Heart Vessels* 2016; **31**: 66–73.
 25. Grabner A, Schramm K, Silswal N, Hendrix M, Yanucil C, Czaya B, Singh S, Wolf M, Hermann S, Stypmann J, di Marco GS, Brand M, Wacker MJ, Faul C. FGF23/FGFR4-mediated left ventricular hypertrophy is reversible. *Sci Rep* 2017; **7**: 1993.
 26. Hassan M, Qureshi W, Sroujeh LS, Albashaireh D, BouMalham S, Liroff M, Amjad W, Khalid F, Hadid H, Alirhayim Z. Interplay of parathyroid hormone and aldosterone antagonist in prevention of heart failure hospitalizations in chronic kidney disease. *Journal of the renin-angiotensin-aldosterone system: JRAAS* 2014; **15**: 278–285.
 27. Tomaschitz A, Pilz S, Rus-Machan J, Meinitzer A, Brandenburg VM, Scharnagl H, Kapl M, Grammer T, Ritz E, Horina JH, Kleber ME, Pieske B, Kraigher-Krainer E, Hartaigh B, Toplak H, van Balleghooijen AJ, Amrein K, Fahrleitner-Pammer A, März W. Interrelated aldosterone and parathyroid hormone mutually modify cardiovascular mortality risk. *Int J Cardiol* 2015; **184**: 710–716.
 28. von Lueder TG, Wang BH, Kompa AR, Huang L, Webb R, Jordaan P, Atar D, Krum H. Angiotensin receptor neprilysin inhibitor LCZ696 attenuates cardiac remodeling and dysfunction after myocardial infarction by reducing cardiac fibrosis and hypertrophy. *Circ Heart Fail* 2015; **8**: 71–78.
 29. McMurray JJV, Solomon SD, Inzucchi SE, Kober L, Kosiborod MN, Martinez FA, Ponikowski P, Sabatine MS, Anand IS, Böhlövék J, Böhm M. Dapagliflozin in patients with heart failure and reduced ejection fraction. *N Engl J Med* 2019; **381**: 1995–2008.
 30. Huang T, Lin X, Li Q, Luo W, Song L, Tan X, Wang W, Li X, Wu X. Selection of a novel FGF23-binding peptide antagonizing the inhibitory effect of FGF23 on phosphate uptake. *Appl Microbiol Biotechnol* 2015; **99**: 3169–3177.
 31. Blau JE, Bauman V, Conway EM, Piaggi P, Walter MF, Wright EC, Bernstein S, Courville AB, Collins MT, Rother KI, Taylor SI. Canagliflozin triggers the FGF23/1,25-dihydroxyvitamin D/PTH axis in healthy volunteers in a randomized crossover study. *JCI insight* 2018; **3**: e99123.
 32. Anker SD, Butler J, Filippatos GS, Jamal W, Salsali A, Schnee J, Kimura K, Zeller C, George J, Brueckmann M, Zannad F, Packer M, Packer M, Anker SD, Butler J, Filippatos GS, Zannad F, George J, Brueckmann M, Perrone S, Nicholls S, Janssens S, Bocchi E, Giannetti N, Verma S, Jian Z, Gomez Mesa JE, Spinar J, Böhm M, Merkely B, Chopra V, Senni M, Taddi S, Tsutsui H, Chuquiure E, la Rocca HBP, Ponikowski P, Vinereanu D, Sim D, Choi DJ, Juanatey JRG, Squire I, Butler J, Januzzi J, Pina I, Pocock SJ, Carson P, Doehner W, Miller A, Haas M, Pehrson S, Komajda M, Anand I, Teerlink J, Rabinstein A, Steiner T, Kamel H, Tsvigoulis G, Lewis J, Freston J, Kaplowitz N, Mann J, Petrie M, Bernstein R, Cheung A, Green J, Januzzi J, Kaul S, Ping CLS, Lip G, Marx N, McCullough P, Mehta C, Ponikowski P, Rosenstock J, Sattar N, Scirica B, Tsutsui H, Verma S, Wanner C, Welty FK, Parhofer KG, Clayton T, Pedersen TR, Lees KR, Konstam MA, Greenberg B, Palmer M. Evaluation of the effects of sodium-glucose co-transporter 2 inhibition with empagliflozin on morbidity and mortality in patients with chronic heart failure and a preserved ejection fraction: rationale for and design of the EMPEROR-Preserved Trial. *Eur J Heart Fail* 2019; **21**: 1279–1287.
 33. Holmlund-Suila E, Enlund-Cerullo M, Valkama S, Hauta-Alus H, Rosendahl J, Helve O, Hytinen T, Viljakainen H, Andersson S, Mäkitie O. Sex and iron modify fibroblast growth factor 23 concentration in 1-year-old children. *J Clin Endocrinol Metab* 2017; **102**: 4526–4533.
 34. di Giuseppe R, Kuhn T, Hirche F, Buijsse B, Dierkes J, Fritsche A, Kaaks R, Boeing H, Stangl GI, Weikert C. Potential predictors of plasma fibroblast growth factor 23 concentrations: cross-sectional analysis in the EPIC-Germany study. *PLoS ONE* 2015; **10**: e0133580.
 35. AbouEzzeddine OF, McKie PM, Dunlay SM, Stevens SR, Felker GM, Borlaug BA, Chen HH, Tracy RP, Braunwald E, Redfield MM. Suppression of tumorigenicity 2 in heart failure with preserved ejection fraction. *J Am Heart Assoc* 2017; **6**: e004382.
 36. Miyamura M, Fujita S, Morita H, Sakane K, Okamoto Y, Sohmiya K, Hoshiga M, Ishizaka N. Circulating fibroblast growth factor 23 has a U-shaped association with atrial fibrillation prevalence. *Circulation journal: official journal of the Japanese Circulation Society* 2015; **79**: 1742–1748.
 37. Mehta R, Cai X, Lee J, Scialla JJ, Bansal N, Sondheimer JH, Chen J, Hamm LL, Ricardo AC, Navaneethan SD, Deo R, Rahman M, Feldman HI, Go AS, Isakova T, Wolf M. Association of fibroblast growth factor 23 with atrial fibrillation in chronic kidney disease, from the chronic renal insufficiency cohort study. *JAMA Cardiol* 2016; **1**: 548–556.
 38. Mizia-Stec K, Wiczorek J, Polak M, Wybraniec MT, Wozniak-Skowarska I, Hoffmann A, Nowak S, Wikarek M, Wnuk-Wojnar A, Chudek J, Więcek A. Lower soluble klotho and higher fibroblast growth factor 23 serum levels are associated with episodes of atrial fibrillation. *Cytokine* 2018; **111**: 106–111.
 39. Dong Q, Li S, Wang W, Han L, Xia Z, Wu Y, Tang Y, Li J, Cheng X. FGF23 regulates atrial fibrosis in atrial fibrillation by mediating the STAT3 and SMAD3 pathways. *J Cell Physiol* 2019; **234**: 19502–19510.
 40. Huang SY, Chen YC, Kao YH, Hsieh MH, Lin YK, Chung CC, Lee TI, Tsai WC, Chen SA, Chen YJ. Fibroblast growth factor 23 dysregulates late sodium current and calcium homeostasis with enhanced arrhythmogenesis in pulmonary vein cardiomyocytes. *Oncotarget* 2016; **7**: 69231–69242.
 41. van Vuren AJ, Gaillard C, Eisenga MF, van Wijk R, van Beers EJ. The EPO-FGF23 signaling pathway in erythroid progenitor cells: opening a new area of research. *Front Physiol* 2019; **10**: 304.
 42. Zile MR, Jhund PS, Baicu CF, Claggett BL, Pieske B, Voors AA, Prescott MF, Shi V, Lefkowitz M, McMurray JJ, Solomon SD, Prospective Comparison of ARNI With ARB on Management of Heart Failure With Preserved Ejection Fraction (PARAMOUNT) Investigators. Plasma biomarkers reflecting profibrotic

- processes in heart failure with a preserved ejection fraction: data from the prospective comparison of ARNI with ARB on management of heart failure with preserved ejection fraction study. *Circ Heart Fail* 2016; **9**: e002551
43. Nagueh SF, Smiseth OA, Appleton CP, Byrd BF 3rd, Dokainish H, Edvardsen T, Flachskampf FA, Gillebert TC, Klein AL, Lancellotti P, Marino P. Recommendations for the evaluation of left ventricular diastolic function by echocardiography: an update from the American Society of Echocardiography and the European Association of Cardiovascular Imaging. *Eur Heart J Cardiovasc Imaging* 2016; **17**: 1321–1360.
 44. Bartunek J, Delrue L, Van Durme F, Muller O, Casselman F, De Wiest B, Croes R, Verstreken S, Goethals M, de Raedt H, Sarma J. Nonmyocardial production of ST2 protein in human hypertrophy and failure is related to diastolic load. *J Am Coll Cardiol* 2008; **52**: 2166–2174.
 45. Saunders JT, Nambi V, de Lemos JA, Chambless LE, Virani SS, Boerwinkle E, Hoogeveen RC, Liu X, Astor BC, Mosley TH, Folsom AR, Heiss G, Coresh J, Ballantyne CM. Cardiac troponin T measured by a highly sensitive assay predicts coronary heart disease, heart failure, and mortality in the atherosclerosis risk in communities study. *Circulation* 2011; **123**: 1367–1376.
 46. Tsutamoto T, Kawahara C, Yamaji M, Nishiyama K, Fujii M, Yamamoto T, Horie M. Relationship between renal function and serum cardiac troponin T in patients with chronic heart failure. *Eur J Heart Fail* 2009; **11**: 653–658.
 47. Smith K, de Filippi C, Isakova T, Gutierrez OM, Laliberte K, Seliger S, Kelley W, Duh S-H, Hise M, Christenson R, Wolf M, Januzzi J. Fibroblast growth factor 23, high-sensitivity cardiac troponin, and left ventricular hypertrophy in CKD. *Am J Kidney Dis* 2013; **61**: 67–73.
 48. Grande D, Leone M, Rizzo C, Terlizze P, Parisi G, Gioia MI, Leopizzi T, Segreto A, Guida P, Romito R, Ciccone M, Serio F, Iacoviello M. A multiparametric approach based on NT-proBNP, ST2, and Galectin3 for stratifying one year prognosis of chronic heart failure outpatients. *Journal of cardiovascular development and disease* 2017; **4**: 9.
 49. Lupon J, Sanders-van Wijk S, Januzzi JL, de Antonio M, Gaggin HK, Pfisterer M, Galán A, Shah R, Brunner-La Rocca HP, Bayes-Genis A. Prediction of survival and magnitude of reverse remodeling using the ST2-R2 score in heart failure: a multicenter study. *Int J Cardiol* 2016; **204**: 242–247.
 50. Gruson D, Ferracin B, Ahn SA, Rousseau MF. Comparison of fibroblast growth factor 23, soluble ST2 and Galectin-3 for prognostication of cardiovascular death in heart failure patients. *Int J Cardiol* 2015; **189**: 185–187.
 51. Nymo SH, Aukrust P, Kjekshus J, McMurray JJ, Cleland JG, Wikstrand J, Muntendam P, Wienhues-Thelen U, Latini R, Askevold ET, Gravning J, Dahl CP, Broch K, Yndestad A, Gullestad L, Ueland T, CORONA Study Group. Limited added value of circulating inflammatory biomarkers in chronic heart failure. *JACC Heart failure* 2017; **5**: 256–264.
 52. Fauconnier C, Roy T, Gillerot G, Roy C, Pouleur AC, Gruson D. FGF23: clinical usefulness and analytical evolution. *Clin Biochem* 2019; **66**: 1–12.
 53. Berezin AE, Berezin AA. Impaired function of fibroblast growth factor 23/klotho protein axis in prediabetes and diabetes mellitus: promising predictor of cardiovascular risk. *Diabetes & metabolic syndrome* 2019; **13**: 2549–2556.
 54. Frimodt-Moller M, von Scholten BJ, Reinhard H, Jacobsen PK, Hansen TW, Persson FI, Parving HH, Rossing P. Growth differentiation factor-15 and fibroblast growth factor-23 are associated with mortality in type 2 diabetes—an observational follow-up study. *PLoS ONE* 2018; **13**: e0196634.
 55. Gutierrez OM, Januzzi JL, Isakova T, Laliberte K, Smith K, Collierone G, Sarwar A, Hoffmann U, Coglianese E, Christenson R, Wang TJ. Fibroblast growth factor 23 and left ventricular hypertrophy in chronic kidney disease. *Circulation* 2009; **119**: 2545–2552.