



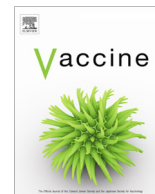
Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



Contents lists available at ScienceDirect

## Vaccine

journal homepage: [www.elsevier.com/locate/vaccine](http://www.elsevier.com/locate/vaccine)

## Commentary

COVID-19 vaccine and boosted immunity: Nothing *ad interim* to do?Luca Roncati<sup>a,\*</sup>, Maria Vadalà<sup>b,c</sup>, Veronica Corazzari<sup>c</sup>, Beniamino Palmieri<sup>b,c</sup><sup>a</sup>Institute of Pathology, University Hospital of Modena, Modena, Italy<sup>b</sup>Department of General Surgery and Surgical Specialties, University of Modena and Reggio Emilia, Modena, Italy<sup>c</sup>Second Opinion Medical Network, Modena, Italy

## ARTICLE INFO

## Article history:

Received 12 April 2020

Received in revised form 21 August 2020

Accepted 5 October 2020

Available online 9 October 2020

## Keywords:

Coronavirus Disease 2019 (COVID-19)

Severe Acute Respiratory Syndrome

Coronavirus 2 (SARS-CoV-2)

*Corynebacterium parvum* (*C. parvum*)*Propionibacterium acnes* (*P. acnes*)*Cutibacterium acnes* (*C. acnes*)

Bacillus Calmette-Guérin (BCG)

Hyaluronic acid

cluster of differentiation 44 (CD44)

T helper 1 (T<sub>H</sub>1)T helper 2 (T<sub>H</sub>2)

Vaccine

## ABSTRACT

Today, Coronavirus Disease 2019 (COVID-19) is a global public health emergency and vaccination measures to counter its diffusion are deemed necessary. Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2), the etiological agent of the disease, unleashes a T-helper 2 immune response in those patients requiring intensive care. Here, we illustrate the immunological mechanism to train the immune system towards a more effective and less symptomatic T-helper 1 immune response, to be exploited against SARS-CoV-2.

© 2020 Elsevier Ltd. All rights reserved.

In the 60–80 years of last century, an experimental wave of immunological studies attempted to fight cancer by exploiting live or killed bacteria, among which the most investigated were the *Bacillus Calmette-Guérin* (BCG), an attenuated strain of *Mycobacterium bovis*, and *Corynebacterium parvum* (*C. parvum*), later renamed *Propionibacterium acnes* and then *Cutibacterium acnes* [1,2]. Administered percutaneously or into the neoplastic mass, they proved able to induce the tumor lysis at some extent, and to delay or arrest the cancer growth through the innate immunity potentiation [3,4]. BCG gained approval since 1977 and is currently the standard of care for patients with non-muscle-invasive bladder cancer by means of mucosal instillation, besides to be registered as an anti-tuberculosis intradermal vaccination [5,6]. As of March 2020, BCG vaccine is furthermore in phase III or IV trials to prevent Coronavirus Disease 2019 (COVID-19) among healthcare workers in the Netherlands, Australia, USA, Germany, France, Denmark, Colombia, Mexico, Egypt, and South Africa [7–16]. Additional trials

in the Netherlands, Germany, Greece, and India are evaluating whether BCG vaccine provides protection against COVID-19 in the elderly and in middle age [17–20]; besides, randomized trials on volunteers over 18 years to test BCG vaccine in this context has been launched in Brazil and Canada [21,22]. *C. parvum* is an aerotolerant anaerobic rod-shaped Gram-positive bacterium largely commensal and part of the skin flora present on most healthy adults, but also associated to sarcoidosis and juvenile acne, hence its taxonomic renaming [23]. After an initial registration like immunoadjuvant and immunomodulator, *C. parvum* was added to the chemotherapy protocol for colon cancer by repeated injections in the form of formalin-killed freeze-dried vaccine preparation (Covarax<sup>®</sup>, Wellcome Research Laboratories, Beckenham, UK); however, it was discarded because no partial remission, overall survival or significant benefits were achieved in the treated cohorts of patients [24]. At that time, one of us (Prof. Palmieri) had the chance to perform a clinical pilot trial with *C. parvum* administration into subcutaneous and lymph nodal metastases from lungs, thyroid and breast malignancies, noting local shrinking and colliquative effect in 48–72 h, accompanied by mild symptoms and occasional febrile peaks. In a few cases, very rapid regressions of concomitant herpes infections involving the head, the thorax

\* Corresponding author at: Polyclinic Hospital, Largo del Pozzo, 71 - 41124 Modena (MO), Italy.

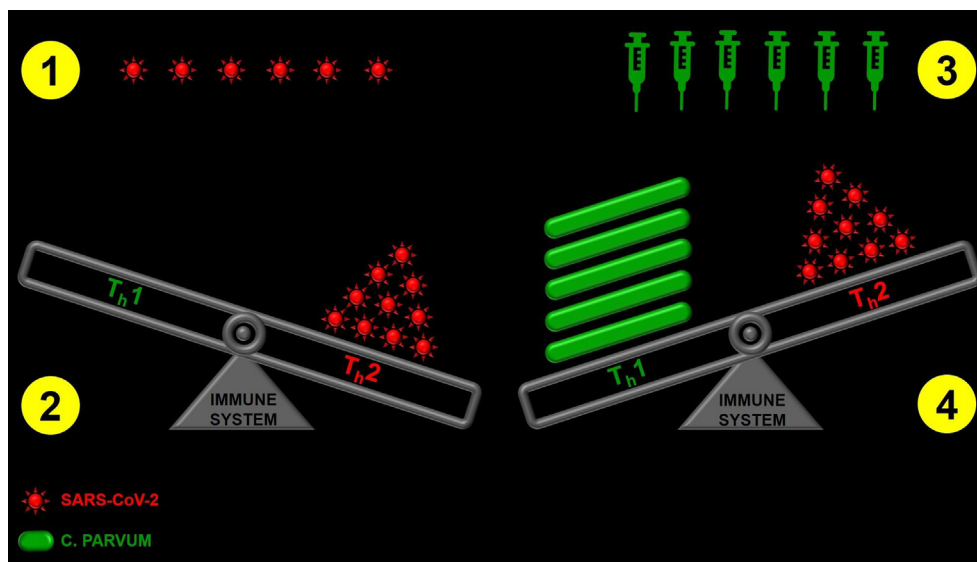
E-mail addresses: [roncati.luca@aou.mo.it](mailto:roncati.luca@aou.mo.it), [luca.roncati@unimore.it](mailto:luca.roncati@unimore.it), [emailmedica@gmail.com](mailto:emailmedica@gmail.com) (L. Roncati).

and the genitals were incidentally observed. By searching on the English biomedical literature, several studies from the past fully support the *C. parvum* antiviral power on man and animal against, for example, influenza virus, hepatitis B, rabies, encephalomyocarditis virus, herpes zoster and human papilloma virus [25–30]. In addition, a scientific evidence of *C. parvum* vaccine protection against a coronavirus (mouse hepatitis virus type 3), dating back to 1981 murine model by Schindler and colleagues, is also reported [31]. Always working on a murine model, Teixeira and collaborators proved in 2018 that *C. parvum* enhances the immunogenicity of the HIVBr18 vaccine, a vaccine against 18 epitopes of the human immunodeficiency virus, subtype B [32]. In the same year, Hsu and coworkers discovered 16 short RNA sequences from *C. parvum* similar to Ebola virus microRNAs, capable to protect the human host by influencing the thrombospondin 4 expression, a multifunctional protein which plays also an initiating role towards cell-mediated immunity in the skin [33]. Palmieri dropped out his clinical trial on bacterial immune stimulation against cancer in 1980, but he followed up with anecdotal compassionate treatments of severe herpes, and others heavy viral infections, such as influenza, mumps, varicella and measles, by subcutaneous injections of *C. parvum* cultured and killed in our microbiological lab, after signing an informed consent out of a total of 40 patients, 30 males and 10 females, aged between 5 and 97 years (mean age 49 years). During our evolving experience, we modified the subdermal injection formula ( $9 \times 10^9$  phenol-killed bacteria in 2 ml saline), adding high molecular weight hyaluronic acid as slow delivery system, having found aspecific virucide properties of this non-sulphated glycosaminoglycan, notoriously able to bind the cluster of differentiation 44 (CD44) receptors present on the surface of activated leukocytes [34,35]. The protocol varied from 1 single to 3–5 injections each other day or every 3 days accordingly with the clinical stage and response to the treatment. The *C. parvum* administration has been always safe and quickly effective showing clinical improvement after 24, 48, 72, 96 h, depending on the patient's performance status and the latency time between the infection outbreak and the beginning of the treatment (Fig. 1). As well known, naïve T-helper ( $T_H0$ ) cells can respond to novel infectious agents never encountered before, like the specific case of Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2), the newly identified betacoronavirus responsible for COVID-19, able to bind the angiotensin converting enzyme 2 receptors [36]. On the basis of the encountered pathogen,  $T_H0$  then polarize the immune response into T-helper 1 ( $T_H1$ ), the default response in immunocompetent subjects to intracellular or phagocytosable pathogens (e.g. viruses, bacteria, protozoa, fungi), mediated by macrophages and T-cytotoxic ( $T_C$ ) cells (cell-mediated immunity), or into T-helper 2 ( $T_H2$ ), classically directed against extracellular non-phagocytosable pathogens (e.g. helminths), whose main effectors are eosinophils, basophils, mastocytes and B cells (humoral immunity) [37]. In spite of this, severe SARS-CoV-2 infections are associated with marked  $T_C$  lymphopenia [38]. During our researches on COVID-19, we have disclosed that the immune system is forced to mount in critically ill patients a  $T_H2$  response, the only one still mountable in the attempt to counteract the viral load, rather than a  $T_H1$  response, which would keep the infection under control by means of macrophages and  $T_C$  cells [39,40]. Moreover, for the first time in worldwide literature, we have provide evidence that a life-threatening escalation from  $T_H2$  immune response to type 3 hypersensitivity (immune complex disease) in COVID-19 vasculitis takes place, and that the inflamed smooth muscle cells of blood vessels concur to the «cytokine storm» via interleukin (IL)-6 [41]. Therefore, we have proposed that an effective vaccination strategy should be able to prevent or limit the systemic imbalance of  $T_H2$  cytokines [42], inducing a protective  $T_H1$  response to be exploited against SARS-CoV-2 (Fig. 2). Among the  $T_H2$  cytokines, there are



**Fig. 1.** Illustrative clinical photographs taken by Prof. Palmieri before and after *C. parvum* vaccination: herpes zoster of the scalp pre-injection (A) and 48 h post-vaccine (B); herpes labialis before the injection (C) and after 24 h from the treatment (D); genital herpes at the time of subdermal injection (E) and 24 h post-vaccination (F).

IL-4, IL-5, IL-6, IL-9, IL-10, IL-13 and IL-25, while IL-2, IL-12, interferon- $\gamma$  (IFN- $\gamma$ ) and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) are the master  $T_H1$  cytokines [37]. Many researches have ascertained that *C. parvum* subcutaneous injection is able to induce a strong  $T_H1$  response favoring the production of IL-2, IL-12, IFN- $\gamma$  and TNF- $\alpha$ , in practice as BCG works, and that the characteristic allergic  $T_H2$  response can be counterbalanced by *C. parvum* vaccination [3,43–48]; besides, it has been found a natural killers (NK) and dendritic cells activator [49–51]. In 1985 Cioffi and colleagues reported that *C. parvum* protects splenectomized Sprague Dawley rats from respiratory challenge with *Streptococcus pneumoniae*, without alter the number or activity of lavageable alveolar macrophages, and they hypothesized that *C. parvum* protection is more likely due to an increased clearance of blood-borne bacteria by the expanded and enhanced reticuloendothelial system [52]. If we transfer this murine model to man, a  $T_H1$  cytokines release syndrome from activated pulmonary macrophages after *C. parvum* lysate subcutaneous injection, such as to aggravate a possible superimposed COVID-19, appears somewhat unlikely. Therefore, our long-standing experience lays the foundation to revalue *C. parvum* lysate as a further surrogate vaccine against COVID-19, in the attempt to prevent or mitigate the cumbersome pandemic morbidity and mortality. The accurate preparation of the lysate is a crucial time to avoid opportunistic bone implant-associated infections or acute septic polyarthritis, whose a single case has been described



**Fig. 2.** Illustrative scheme of our vaccination rationale: following SARS-CoV-2 entry (point 1), the immune system is forced to mount a  $T_{h2}$  response in those patients requiring intensive care, at the expense of a more effective and less symptomatic  $T_{h1}$  response, compromised by the viral load (point 2). Through *C. parvum* or BCG vaccine (point 3), it is theoretically possible to train and calibrate the immune system towards a  $T_{h1}$  response (point 4), able to prevent COVID-19 or to keep the disease under control in a paucisymptomatic or asymptomatic way thanks to activated reticuloendothelial system, NK,  $T_c$  and dendritic cells.

in 1983 after *C. parvum* instillation for malignant pleural effusion; an episode of prolonged fever from immune system hyperactivation has also been reported [53–55]. Previous efforts to develop subunit vaccines against the most lethal human coronaviruses, for instance the Severe Acute Respiratory Syndrome Coronavirus 1 (SARS-CoV-1) and the Middle East Respiratory Syndrome Coronavirus (MERS-CoV), have failed because not enough protective; a recent study on SARS-CoV-2 has confirmed that neutralizing antibodies decline during the weeks or 2–3 months following infection at least in asymptomatic or paucisymptomatic patients [56], a biological behavior comparable to that of other known human coronaviruses [57], and which complicates the road to develop a specific long-term protective COVID-19 vaccine. By boosting  $T_{h1}$  response and innate immunity rather than the humoral one, the disappearance of neutralizing antibodies and the adaptive mutational potential of SARS-CoV-2, limiting factors for the development of an effective subunit vaccine, could be so circumvent. In this regard, a further recent study on 10,022 SARS-CoV-2 genomes has identified 5,775 distinct genome variants, including 2,969 missense mutations, 1,965 synonymous mutations, 484 mutations in the non-coding regions, 142 non-coding deletions, 100 in-frame deletions, 66 non-coding insertions, 36 stop-gained variants, 11 frameshift deletions and 2 in-frame insertions, a series of genetic events which determine SARS-CoV-2 virulence, infectivity and transmissibility [58]. In conclusion, we have here illustrated our rationale for a strategic off-target vaccination against SARS-CoV-2, promptly available and safe for the patients.

#### Funding

None.

#### Authors contribution

LR conceived, designed and supervised the study, prepared figures and related legends, revised critically the first draft and wrote the final version of the manuscript; MV and VC analyzed and interpreted the data and performed the literature search; BP conceived and designed the study and wrote the first draft of the manuscript.

All the authors approved the final version of the manuscript and attested they meet the ICMJE criteria for authorship.

#### Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

#### References

- [1] Oettgen HF. Immunotherapy of cancer. *N Engl J Med* 1977;297:484–91.
- [2] Lipton A, Harvey HA, Lawrence B, Gottlieb R, Kukrika M, Dixon R, et al. *Corynebacterium parvum* versus BCG adjuvant immunotherapy in human malignant melanoma. *Cancer* 1983;51:57–60.
- [3] Tchaptchet S, Kirberg J, Freudenberg N, Schamel WW, Galanos C, Freudenberg MA. Innate, antigen-independent role for T cells in the activation of the immune system by *Propionibacterium acnes*. *Eur J Immunol* 2010;40:2506–16.
- [4] Angelidou A, Conti MG, Diray-Arce J, Benn CS, Shann F, Netea MG, et al. Licensed Bacille Calmette-Guérin (BCG) formulations differ markedly in bacterial viability, RNA content and innate immune activation. *Vaccine* 2020;38:2229–40.
- [5] World Health Organization. BCG vaccines: WHO position paper – February 2018. *Wkly Epidemiol Rec* 2018;93:73–96.
- [6] Brandau S, Suttman H. Thirty years of BCG immunotherapy for non-muscle invasive bladder cancer: a success story with room for improvement. *Biomed Pharmacother* 2007;61:299–305.
- [7] U.S. National Library of Medicine. Reducing health care workers absenteeism in Covid-19 pandemic through BCG vaccine (BCG-CORONA). *ClinicalTrials.gov* identifier: NCT04328441; 2020 [last accessed April 29, 2020].
- [8] U.S. National Library of Medicine. BCG vaccination to protect healthcare workers against COVID-19 (BRACE). *ClinicalTrials.gov* identifier: NCT04327206; 2020 [last accessed July 8, 2020].
- [9] U.S. National Library of Medicine. BCG vaccine for health care workers as defense against COVID-19 (BADAS). *ClinicalTrials.gov* identifier: NCT04348370; 2020 [last accessed May 27, 2020].
- [10] U.S. National Library of Medicine. Study to assess VPM1002 in reducing healthcare professionals' absenteeism in COVID-19 pandemic. *ClinicalTrials.gov* identifier: NCT04387409; 2020 [last accessed June 16, 2020].
- [11] U.S. National Library of Medicine. Efficacy of BCG vaccination in the prevention of COVID-19 via the strengthening of innate immunity in health care workers (COVID-BCG). *ClinicalTrials.gov* identifier: NCT04384549; 2020 [last accessed May 12, 2020].
- [12] U.S. National Library of Medicine. Using BCG vaccine to protect health care workers in the COVID-19 pandemic. *ClinicalTrials.gov* identifier: NCT04373291; 2020 [last accessed May 5, 2020].
- [13] U.S. National Library of Medicine. Performance evaluation of BCG vaccination in healthcare personnel to reduce the severity of SARS-COV-2 infection.

- ClinicalTrials.gov identifier: NCT04362124; 2020 [last accessed April 28, 2020].
- [14] U.S. National Library of Medicine. Prevention, efficacy, and safety of BCG vaccine in COVID-19 among healthcare workers. ClinicalTrials.gov identifier: NCT04461379; 2020 [last accessed July 14, 2020].
- [15] U.S. National Library of Medicine. Application of BCG vaccine for immunoprophylaxis among Egyptian healthcare workers during the pandemic of COVID-19. ClinicalTrials.gov identifier: NCT04350931; 2020 [last accessed April 20, 2020].
- [16] U.S. National Library of Medicine. BCG vaccination for healthcare workers in COVID-19 pandemic. ClinicalTrials.gov identifier: NCT04379336; 2020 [last accessed May 7, 2020].
- [17] U.S. National Library of Medicine. Reducing COVID-19 related hospital admission in elderly by BCG vaccination. ClinicalTrials.gov identifier: NCT04417335; 2020 [last accessed June 4, 2020].
- [18] U.S. National Library of Medicine. Study to assess VPM1002 in reducing hospital admissions and/or severe respiratory infectious diseases in elderly in COVID-19 pandemic. ClinicalTrials.gov identifier: NCT04435379; 2020 [last accessed August 6, 2020].
- [19] U.S. National Library of Medicine. Bacillus Calmette-Guérin vaccination to prevent COVID-19 (ACTIVATEII). ClinicalTrials.gov identifier: NCT04414267; 2020 [last accessed July 14, 2020].
- [20] U.S. National Library of Medicine. BCG vaccine in reducing morbidity and mortality in elderly individuals in COVID-19 hotspots. ClinicalTrials.gov identifier: NCT04475302; 2020 [last accessed July 23, 2020].
- [21] U.S. National Library of Medicine. COVID-19: BCG as therapeutic vaccine, transmission limitation, and immunoglobulin enhancement (BATTLE). ClinicalTrials.gov identifier: NCT04369794; 2020 [last accessed August 7, 2020].
- [22] U.S. National Library of Medicine. Efficacy and safety of VPM1002 in reducing SARS-CoV-2 (COVID-19) infection rate and severity (COBRA). ClinicalTrials.gov identifier: NCT04439045; 2020 [last accessed June 19, 2020].
- [23] Palmieri B, Vadalà M, Roncati L, Garelli A, Scandone F, Bondi M, et al. The long-standing history of *Corynebacterium parvum*, immunity, and viruses. *J Med Virol* 2020. <https://doi.org/10.1002/jmv.26100>.
- [24] Whisnant JK. *C. parvum* clinical protocols: prototypes and summary results in U.S. trials with Wellcome Coparvax. *Dev Biol Stand* 1977;38:559–66.
- [25] Mak NK, Schiltknecht E, Ada GL. Protection of mice against influenza virus infection: enhancement of nonspecific cellular responses by *Corynebacterium parvum*. *Cell Immunol* 1983;78:314–25.
- [26] Papaevangelou G, Sparros L, Vissoulis C, Kyriakidou A, Giokas G, Hadzimanolis J, et al. The effect of intradermal administration of *Corynebacterium parvum* on the immune response to hepatitis Bs antigen. *J Med Virol* 1977;1:15–9.
- [27] Megid J, Kaneno R, Nozaki CN, Brito CJ, Almeida MF. Increased interleukin-10 associated with low IL-6 concentration correlated with greater survival rates in mice infected by rabies virus vaccinated against it and immunomodulated with *P. acnes*. *Comp Immunol Microbiol Infect Dis* 2004;27:393–411.
- [28] Géniteau-Legendre M, Forestier F, Quérou AM, German A. Role of interferon, antibodies and macrophages in the protective effect of *Corynebacterium parvum* on encephalomyocarditis virus-induced disease in mice. *Antiviral Res* 1987;7:161–7.
- [29] Topciu V, Mihăilescu R. Immunomodulating and antiviral therapy in herpes zoster. *Rom J Virol* 1996;47:75–80.
- [30] Nasser N. Treatment of common warts with the immune stimulant *Propionibacterium parvum*. *An Bras Dermatol* 2012;87:585–9.
- [31] Schindler L, Streissle G, Kirchner H. Protection of mice against mouse hepatitis virus by *Corynebacterium parvum*. *Infect Immun* 1981;32:1128–31.
- [32] Teixeira D, Ishimura ME, Apostólico JS, Viel JM, Passarelli VC, Cunha-Neto E, et al. *Propionibacterium acnes* enhances the immunogenicity of HIVBr 18 human immunodeficiency virus-1 vaccine. *Front Immunol* 2018;9:177.
- [33] Hsu PC, Chiou BH, Huang CM. On revealing the gene targets of Ebola virus microRNAs involved in the human skin microbiome. *PeerJ* 2018;6:e4138.
- [34] Cermelli C, Cuoghi A, Scuri M, Bettua C, Neglia RG, Ardizzoni A, et al. In vitro evaluation of antiviral and virucidal activity of a high molecular weight hyaluronic acid. *Virol J* 2011;8:141.
- [35] Aruffo A, Stamenkovic I, Melnick M, Underhill CB, Seed B. CD44 is the principal cell surface receptor for hyaluronate. *Cell* 1990;61:1303–13.
- [36] Roncati L, Gallo G, Manenti A, Palmieri B. Renin-angiotensin system: the unexpected flaw inside the human immune system revealed by SARS-CoV-2. *Med Hypotheses* 2020;140:109686.
- [37] Spellberg B, Edwards Jr JE. Type 1/Type 2 immunity in infectious diseases. *Clin Infect Dis* 2001;32:76–102.
- [38] Zhao Q, Meng M, Kumar R, Wu Y, Huang J, Deng Y, et al. Lymphopenia is associated with severe coronavirus disease 2019 (COVID-19) infections: a systemic review and meta-analysis. *Int J Infect Dis*. 2020;96:131–5.
- [39] Roncati L, Nasillo V, Lusenti B, Riva G. Signals of Th2 immune response from COVID-19 patients requiring intensive care. *Ann Hematol* 2020;99:1419–20.
- [40] Roncati L, Lusenti B. The «moonlighting protein» able to explain the Th1 immune lockdown in severe COVID-19. *Med Hypotheses* 2020;143:110087.
- [41] Roncati L, Ligabue G, Fabbiani L, Malagoli C, Gallo G, Lusenti B, et al. Type 3 hypersensitivity in COVID-19 vasculitis. *Clin Immunol* 2020;217:108487.
- [42] Roncati L, Palmieri B. What about the original antigenic sin of the humans versus SARS-CoV-2? *Med Hypotheses* 2020;142:109824.
- [43] Kistowska M, Meier B, Proust T, Feldmeyer L, Cozzio A, Kuendig T, et al. *Propionibacterium acnes* promotes Th17 and Th17/Th1 responses in acne patients. *J Invest Dermatol* 2015;135:110–8.
- [44] Furusawa H, Suzuki Y, Miyazaki Y, Inase N, Eishi Y. Th1 and Th17 immune responses to viable *Propionibacterium acnes* in patients with sarcoidosis. *Respir Investig* 2012;50:104–9.
- [45] Tsuda K, Yamanaka K, Linan W, Miyahara Y, Akeda T, Nakanishi T, et al. Intratumoral injection of *Propionibacterium acnes* suppresses malignant melanoma by enhancing Th1 immune responses. *PLoS ONE* 2011;6:e29020.
- [46] Girvan RC, Knight DA, O'Loughlin CJ, Hayman CM, Hermans IF, Webster GA. MIS416, a non-toxic microparticle adjuvant derived from *Propionibacterium acnes* comprising immunostimulatory muramyl dipeptide and bacterial DNA promotes cross-priming and Th1 immunity. *Vaccine* 2011;29:545–57.
- [47] Kitagawa H, Yamanaka K, Kakeda M, Inada H, Imai Y, Gabazza EC, et al. *Propionibacterium acnes* vaccination induces regulatory T cells and Th1 immune responses and improves mouse atopic dermatitis. *Exp Dermatol* 2011;20:157–8.
- [48] O'Neill LAJ, Netea MG. BCG-induced trained immunity: can it offer protection against COVID-19? *Nat Rev Immunol* 2020;20:335–7.
- [49] Ananias RZ, Rodrigues EG, Braga EG, Squaiella CC, Mussalem JS, Longhini AL, et al. Modulatory effect of killed *Propionibacterium acnes* and its purified soluble polysaccharide on peritoneal exudate cells from C57Bl/6 mice: major NKT cell recruitment and increased cytotoxicity. *Scand J Immunol* 2007;65:538–48.
- [50] Balch CM, Smalley RV, Bartolucci AA, Burns D, Presant CA, Durant JR. A randomized prospective clinical trial of adjuvant *C. parvum* immunotherapy in 260 patients with clinically localized melanoma (Stage I): prognostic factors analysis and preliminary results of immunotherapy. *Cancer* 1982;49:1079–84.
- [51] Hirt HM, Schwentek M, Becker H, Kirchner H. Interferon production and lymphocyte stimulation in human leucocyte cultures stimulated by *Corynebacterium parvum*. *Clin Exp Immunol* 1978;32:471–6.
- [52] Cioffi WG, Hebert JC, Gamelli RL, Foster Jr RS. The quantity and function of pulmonary alveolar macrophages after splenectomy and *Corynebacterium parvum*. *J Trauma* 1985;25:405–9.
- [53] Dubus M, Varin J, Papa S, Rammal H, Chevrier J, Maisonneuve E, et al. Interaction of *Cutibacterium acnes* with human bone marrow derived mesenchymal stem cells: a step toward understanding bone implant-associated infection development. *Acta Biomater* 2020;104:124–34.
- [54] Lever AM, Forsythe J, Oxford P. Acute polyarthritides following the use of *Corynebacterium parvum* vaccine (Coparvax) for malignant pleural effusion. *Postgrad Med J* 1983;59:799–800.
- [55] Laroche CM, Britton M. Prolonged fever after pleural instillation of *Corynebacterium parvum* (Coparvax). *Thorax* 1987;42:823–4.
- [56] Long QX, Tang XJ, Shi QL, Li Q, Deng HJ, Yuan J, et al. Clinical and immunological assessment of asymptomatic SARS-CoV-2 infections. *Nat Med* 2020. <https://doi.org/10.1038/s41591-020-0965-6>.
- [57] Reed SE. The behaviour of recent isolates of human respiratory coronavirus in vitro and in volunteers: evidence of heterogeneity among 229E-related strains. *J Med Virol* 1984;13:179–92.
- [58] Koyama T, Platt D, Parida L. Variant analysis of SARS-CoV-2 genomes. *Bull World Health Organ* 2020;98:495–504.