

Review

# Nutritional Composition of Edible Insects Consumed in Africa: A Systematic Review

Zabentungwa T. Hlongwane <sup>\*</sup>, Rob Slotow  and Thinandavha C. Munyai 

School of Life Sciences, University of KwaZulu-Natal, Private Bag X01, Scottsville 3209, South Africa; slotow@ukzn.ac.za (R.S.); munyaic2@ukzn.ac.za (T.C.M.)

\* Correspondence: nolwazihlongwane20@gmail.com

Received: 13 August 2020; Accepted: 3 September 2020; Published: 11 September 2020



**Abstract:** Edible insects are an important protein rich natural resource that can contribute to resilient food security. Edible insects not only play an important role in traditional diets, but are also an excellent source of protein in traditional dishes in Africa. We systematically searched Web-of-Science and Google Scholar from year 2000–2019 for studies on the consumption of insects and their nutritional composition in Africa, resulting in 98 eligible papers, listing 212 edible insect species from eight orders. These insects were rich in protein, fats, and fibre. The highest protein content was reported for Lepidoptera (range: 20–80%). Coleoptera had the highest carbohydrate content (7–54%), while Lepidoptera had the highest fat content (10–50%). Considering the excellent source of nutrition, and potential socio-economic benefits, from edible insects, they can contribute strongly to improved food security, and rural development in developing countries. In addition, edible insects can be used as a sustainable food source to combat food shortages in the future, for example, providing resilience during times of drought or other climate stressors.

**Keywords:** entomophagy; Africa; edible insects; nutrition; food security

## 1. Introduction

Consumption of insects has recently received more attention because of their promising potential for contributing to livelihoods and mitigating food security problems around the world [1–3]. Food security problems are caused by an enormous increase in the global human population, which is estimated to increase to approximately 9 billion people by 2050 [1], resulting in a 70% increase in food demand, and an increase in food prices [1,4,5]. The increase in food prices will prompt the search for cheap alternative sustainable protein sources [1]. Entomophagy, which refers to the consumption of insects by humans, is an environmentally friendly approach to increasing food for consumption, and contributing to food security across the world [2,5–7].

Edible insects might be a solution to food shortages, owing to their promising potential in contributing to livelihoods and mitigating food security problems around the world [1–3]. Insects are consumed as food in Thailand [8,9], China [10,11], Mexico [12–15], Latin America [16], Japan [17], and Africa [18]. According to van Huis [1], approximately 2 billion people worldwide regularly consume insects as part of their diets. The consumption of insects is not a new phenomenon, as it dates back to before the development of agriculture when humans relied on gathering plants and hunting wild animals [4,11,19].

Edible insects have played a very important traditional role in nutritious diets in various countries in Africa [18,20]. In addition, edible insects are an important natural resource that is used as a coping strategy, particularly in months of food shortage [21–23]. Unfavourable climatic conditions experienced in Africa affect small scale animal husbandry and reduce animal protein production, so diets are then supplemented with edible insect protein [22]. Edible insects provide significant socio-economic and ecological benefits for developing countries [24,25]. Approximately 500 species of edible insects are

consumed in Africa and form part of traditional diets [18]. Of these 500 species, 256 species were consumed in the Central African region, 164 in southern Africa, 100 species in eastern Africa, 91 in western Africa, and only eight species in northern Africa [18]. Insects are consumed among different African cultures because of their taste, cultural importance, and nutritive value, and as a supplementary food when staple food is limited [1,3,25–27].

Various studies in Africa have focused on studying the nutritional content of a single species, group, or genus [28–32]. Little is known about the diversity and nutritional content of various insects consumed in Africa. Therefore, the current study will review the existing literature on the diversity of insects, and their nutritional status in Africa, and, therefore, compile information on the nutrient composition of edible insects consumed in Africa. This will be done by asking the following questions: (1) What is the nutritional value of edible insects consumed in Africa, (2) what are the most consumed, and (3) the most studied insect species, in terms of nutrition, in Africa?

## 2. Materials and Methods

### 2.1. Search Strategy

To explore the diversity and nutritional status of edible insects in Africa, we followed the PRISMA guidelines for a systematic review. Peer-reviewed literature was obtained using the Thomson Reuters' Web of Science database (<https://apps.webofknowledge.com>) and google scholar (<https://scholar.google.co.za/>) looking for publications that researched entomophagy in Africa, edible insects, diversity, nutrient content of edible insects, and consumption of insects. To source information, the following key words and phrases were used, "entomophagy", "edible insects", "diversity of edible insects", "entomophagy in Africa", "edible insects in eastern Africa", "edible insects in north Africa", "edible insects in western Africa", "edible insects in Central Africa", "edible insects in southern Africa" and "nutrient content of edible insects". We also screened references included in selected articles in order to identify studies that might be relevant but did not appear in our search. We limited the search to literature published from 2000 to 2019. We started in the year 2000 because it was a starting point where most researchers began investigating the use of edible insects as a food source and as a solution to combat food insecurity problems [33,34].

### 2.2. Data Collection

Data from the selected articles were independently screened and extracted by a single author (Z.T.H). The search result was done by reading the title and abstract of the retrieved papers to determine if the article was relevant to the study. Once it was determined that the article was relevant, the full text of the selected articles was further analysed to extract relevant information. The information that was collected and extracted after full text reading from each article included year, study area and country, study insect species, reported nutrient composition of insects, consumption stage of an insect, main research findings, and conclusions. Collected articles were categorised by country and insect order.

### 2.3. Inclusion and Exclusion Criteria

#### 2.3.1. Inclusion Criteria

- Original research articles and review papers focusing on entomophagy, nutrient composition of single or multiple edible insect species.
- Articles published in English.
- Articles of work done in African countries.
- Articles that reported nutrient composition of edible insects.

#### 2.3.2. Exclusion Criteria

- Conference papers, editorial material, book chapters

- Articles on insect rearing and farming.

#### 2.4. Data Quality

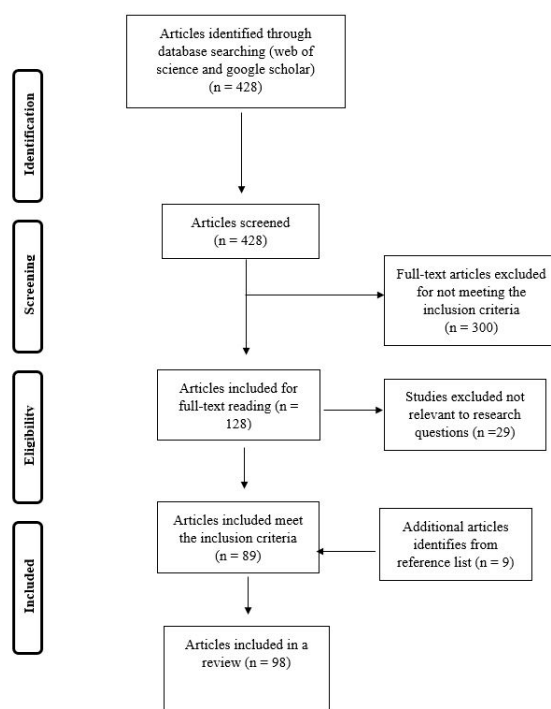
To evaluate the quality of studies included in this systematic review, we assessed quality based on the following criteria: (1) A clear food description (scientific name(s) of insects studied or genus), (2) a clear description on the part of the insects used for analysis, e.g., whole, head, abdomen, indication of geographic origin of the insects, and the country where it is used as food in Africa, (3) analytical method used, number of analytical samples, (4) clear indication of whether the nutritional composition was based on the dry weight. Studies were included if they meet all the above criteria.

#### 2.5. Data Analysis

The methods and data sources used in the included studies were highly heterogeneous and a statistical meta-analysis was not possible. Instead, a more narrative synthesis approach was used, and data from each study were tabulated. We synthesised the results according to study species and mean values of all insect species belonging to the same insect order were calculated and represented in bold, the nutritional composition of consumed species were presented in the table, most consumed species in different countries were presented graphically.

### 3. Results and Discussion

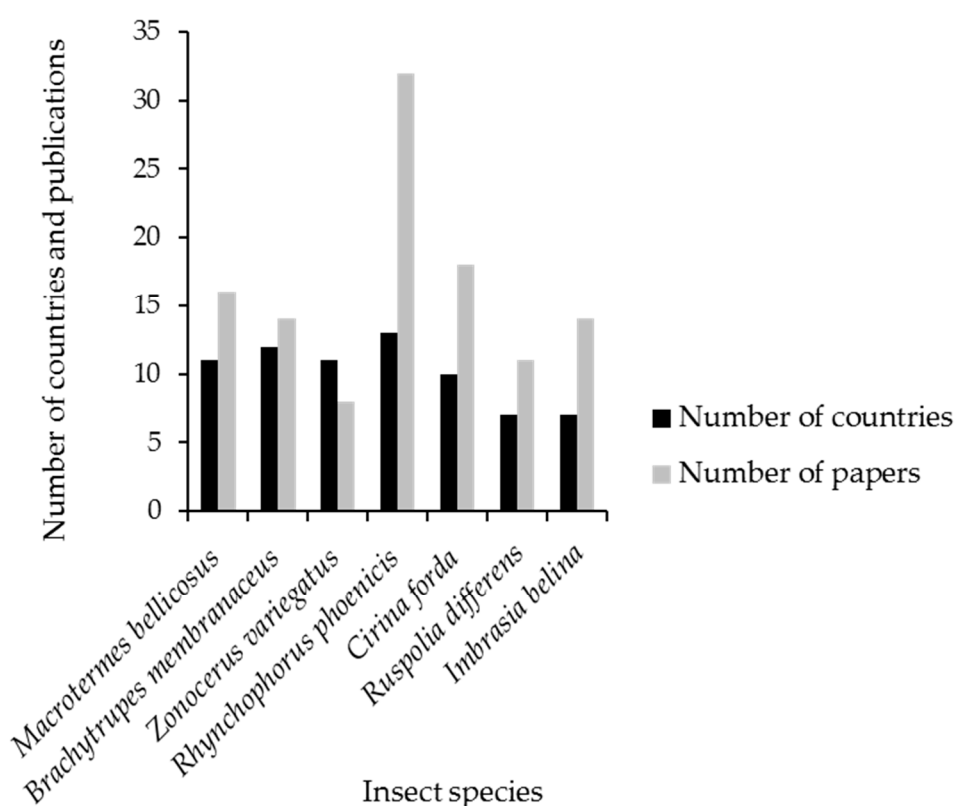
A total of 428 papers were identified for potential inclusion; after checking the title and abstract, 300 articles were excluded because they did not meet the inclusion criteria. From here, 128 articles were selected for full-text reading; from these, 29 articles were further excluded because they were not relevant or not conducted in Africa. After reading the full-text, 89 studies met all inclusion criteria, and a further nine articles were identified through screening references and confirming inclusion criteria were met. In total 98 articles were included in a systematic review (Figure 1).



**Figure 1.** Flow chart of the study selection process for systematic review of the nutritional composition of edible insects.

### 3.1. Consumption of Insect Patterns in Africa

For the research articles published since 2000, a total of 212 edible insect species from nine orders were recorded and are potentially consumed in different African countries (Appendix A). Of these, 41% were Lepidoptera, 23% Orthoptera, 15% Coleoptera, 12% Blattodea (including both cockroaches and termites as recently classified), 4% Hemiptera, and Hymenoptera, Diptera, Blattodea, and Mantodea each contributed <1%. *Rhynchophorus phoenicis* (African palm weevil) and *Cirina forda* (Pallid emperor moth) were the most studied species in Africa, with 32 publications from 12 countries, and 18 publications from 10 countries, respectively (Figure 2). Most research has been done in the western African countries, particularly in Nigeria, mainly on *Rhynchophorus phoenicis* and *Cirina forda*, which are the most consumed species in West Africa. However, southern African countries (Zimbabwe, South Africa, and Botswana) have the highest number of consumed species, but little research has been done on nutritional content and consumption patterns of edible insects.



**Figure 2.** The number of countries with journal peer-reviewed articles published on the most consumed and economically important insects in Africa.

### 3.2. Nutrient Composition of Edible Insects

A compilation of nutrient composition of 54 edible insects based on the dry matter is presented in Table 1. Percentage of fat, protein, moisture, and ash content were calculated based on dry weight of the insect when ready for preparation to eat, noting that, in some cases, the insects had been processed since collecting. The highest protein was reported in Lepidoptera (range: 12–79%) and Orthoptera (12–73%), while the lowest protein content ranging from (0–39%) was reported for Blattodea.

**Table 1.** Nutritional composition of edible insects, based on dry matter, from six orders consumed by people in Africa.

Scientific Name	Stage of Consumption	Protein (%)	Crude Fibre (%)	Moisture (%)	Ash (%)	Carb (%)	Vitamin A (mg/100 g)	Vitamin B2 (mg/100 g)	Vitamin C (mg/100 g)	Fe (mg/100 g)	Ca (mg/100 g)	Zn (mg/100 g)	P (mg/100 g)	Mg (mg/100 g)	Fats (mg/100 g)	Reference
<b>Blattodea (termites and cockroaches)</b>		<b>33.2 ± 14.5</b>	<b>4.7 ± 3.9</b>	<b>2.9 ± 0.1</b>	<b>5.2 ± 2.5</b>	<b>23.2 ± 0</b>	<b>2.7 ± 0.2</b>	<b>1.8 ± 0.2</b>	<b>3.2 ± 0.2</b>	<b>86 ± 96.8</b>	<b>54.1 ± 42.6</b>	<b>13.8 ± 3.5</b>	<b>125 ± 11</b>	<b>0.2 ± 0.1</b>	<b>22.2 ± 9.8</b>	
<i>Periplaneta Americana</i>	Adult	39.6	13.1		6.2											[35]
<i>Macrotermes nigeriensis</i>	Adult	35.9	5.5		5.8											[35]
<i>Macrotermes bellicosus</i>	Adult	20.4	2.7	2.8	11.3	23.2	2.9	2.0	3.4	27.0	21.0		136.0	0.2	36.1	[36]
<i>Macrotermes natalensis</i>	Adult	22.1	2.2	3.0	4.1		2.6	1.5	3.0	29.0	18.0		114.0	0.3	21.4	[36]
<i>Pseudacathoerms spinige</i>	Adult				6.8					332.0	84.7	11.9				[37]
<i>Macrotermes spp.</i>	Adult				2.4					93.9	83.7	8.1				[37]
<i>Macrotermes herus</i>	Adult				6.8					161.0	132.0	14.3				[37]
<i>Macrotermes bellicosus</i>	Adult	40.7			5.7					42.7		16.9			8.4	[38]
<i>Macrotermes bellicosus</i>	Adult	20.4	2.7	2.8	2.9		2.9	2.0	3.4	27.0	21.0		136.0	0.2		[36]
<i>Syntermes soldiers</i>	Adult	64.7			4.2					32.5		17.6			23.0	[38]
<i>Macrotermes natalensis</i>	Adult	22.1	2.2	3.0	1.9		2.6	1.5	3.0	29.0	18.0		114.0	0.3		[36]
<b>Coleoptera (beetles)</b>		<b>32.8 ± 11.5</b>	<b>6.2 ± 7.8</b>	<b>7.6 ± 15.7</b>	<b>4.7 ± 2.7</b>	<b>22.6 ± 13.2</b>	<b>11.2 ± 1.4</b>	<b>1.9 ± 0.9</b>	<b>5.4 ± 1.2</b>	<b>14.1 ± 8.9</b>	<b>43.6 ± 14.3</b>	<b>14.4 ± 12.1</b>	<b>109.6 ± 48.5</b>	<b>10.1 ± 4.2</b>	<b>29.1 ± 16.6</b>	
<i>Analeptes trifasciata</i>	Larvae	20.1	2.0	2.2	5.1		12.5	2.6	5.4	18.2	61.2		136.4	18.2		[36]
<i>Oryctes boas</i>	Larvae	26.0	1.5	1.9	1.5							2.3				[6,36]
<i>Oryctes monoceros</i>	Larvae	26.4		4.7	7.8	51.6										[39]
<i>Aphodius rufipes</i>	Larvae	22.4	28.1	3.3	2.7	13.1				30.9	42.2			11.7	30.5	[36]
<i>Rhynchophorus phoenicis</i>	Larvae	28.4	2.8	2.7	2.7		11.3	2.2	4.3	12.2	39.6	26.5	126.4	7.5	66.6	[6]
<i>Oryctes rhinoceros</i>	Larvae	50.5								4.5					38.1	[6]
<i>Oryctes owariensis</i>	Larvae	50.6		8.4	7.7	14.3									18.9	[40]

Table 1. Cont.

Scientific Name	Stage of Consumption	Protein (%)	Crude Fibre (%)	Moisture (%)	Ash (%)	Carb (%)	Vitamin A (mg/100 g)	Vitamin B2 (mg/100 g)	Vitamin C (mg/100 g)	Fe (mg/100 g)	Ca (mg/100 g)	Zn (mg/100 g)	P (mg/100 g)	Mg (mg/100 g)	Fats (mg/100 g)	Reference
<i>Eulopida mashona</i>	Larvae	46.3	14.8		10.9	16.2									11.8	[41]
<i>Heteroligus meles</i>	Larvae	38.1	3.0	1.0	5.8	20.1									32.0	[42]
<i>Rhynchophorus phoenicis</i>	Larvae	50.0	2.6	1.2	4.9	20.2									21.1	[42]
<i>Rhynchophorus phoenicis</i>	Larvae	28.4	2.8	2.7	2.7		11.3	2.2	4.3	12.2	39.6		126.4	7.5		[36]
<i>Analeptes trifasciata</i>	Larvae	29.6	2.0	2.2	4.2		12.5	2.6	5.4	18.2	61.3		136.4	6.1		[36]
<i>Oryctes boas</i>	Larvae	26.0	3.4	1.9	1.5		8.6	0.1	7.6	2.3	45.7		130.2	6.3		[36]
<i>Apomecyna parumpunctata</i>	Larvae	16.8	5.4	59.4	3.0						15.7		1.5	13.5	13.9	[43]
<b>Hemiptera (bugs)</b>		<b>39.3 ± 4.0</b>	<b>5.3 ± 0</b>	<b>4.9 ± 0</b>	<b>1.7 ± 0</b>	<b>6.3 ± 1.3</b>	<b>0.2 ± 0</b>	<b>0.9 ± 0</b>		<b>20.2 ± 0</b>	<b>91.0 ± 0</b>	<b>46.0 ± 0</b>	<b>57 ± 0</b>	<b>109 ± 0</b>		
<i>Encosternum delegorguei</i>		43.3	5.3	4.9	1.7	5.0	0.2	0.9		20.2	91.0		575.0	109.0	45.0	[6]
<i>Encosternum delegorguei</i>		35.2		4.9	1.7	7.6				20.2	91.0	46.0		109.0		[28]
<b>Hymenoptera (bees and ants)</b>		<b>33.9 ± 9.2</b>	<b>7.7 ± 4.6</b>	<b>3.9 ± 0.1</b>	<b>4.1 ± 3.2</b>		<b>12.4 ± 0</b>	<b>3.2 ± 0</b>	<b>10.3 ± 0</b>	<b>17.8 ± 6.6</b>	<b>21.6 ± 6.3</b>	<b>7.5 ± 2.5</b>	<b>115.6 ± 9.6</b>	<b>7.8 ± 2.6</b>	<b>42.9 ± 4.7</b>	
<i>Apis mellifera</i>	Adult	21.0	2.0	3.8	2.2		12.4	3.2	10.3	25.2	15.4		125.5	5.2		[6,36]
<i>Carebara vidua</i>	Adult	42.5	9.1		8,6					10.4	22.3	5.7	106.0	10.4	38.2	[6]
<i>Comptonotus spp.</i>	Adult	40.1	14.1		9.6											[35]
<i>Oecophylla longinoda</i>	Adult	37.8	12.3		7.3											[35]
<i>Crematogaster mimosa</i>	Adult				1.7					17.7	32.6	11.1				[37]
<i>Carebara vidua Smith</i>	Adult	40.8	6.9	3.9	1.6					10.7	22.2	5.7	106.0	10.4	47.5	[44]
<i>Apis mellifera</i>	Adult	21.0	2.0	3.8	2.2		12.4	3.2	10.3	25.2	15.4		125.0	5.2		[36]
<b>Lepidoptera (caterpillars)</b>		<b>46.3 ± 21.7</b>	<b>5.9 ± 5.4</b>	<b>29.3 ± 36.5</b>	<b>4.6 ± 2.2</b>	<b>18.0 ± 13.0</b>	<b>3.1 ± 0.2</b>	<b>1.7 ± 0.6</b>	<b>2.8 ± 1.0</b>	<b>15.4 ± 22.2</b>	<b>9.4 ± 2.3</b>	<b>10.6 ± 2.2</b>	<b>320.7 ± 367.9</b>	<b>18.9 ± 45.5</b>	<b>18.3 ± 14.8</b>	
<i>Anaphe venata</i>	Larvae	60.0	3.2	3.3			3.1	1.3	2.2	2.0	8.6		100.5	1.6		[6]
<i>Anaphe infracta</i>	Larvae	20.0	2.4	2.7			3.0	2.0	4.5	1.8	8.6		113.3	1.0		[6,36]
<i>Anaphe reticulata</i>	Larvae	23.0	3.1	3.2			3.4	2.0	2.2	2.2	10.5		102.4	2.6		[6,36]

Table 1. Cont.

Scientific Name	Stage of Consumption	Protein (%)	Crude Fibre (%)	Moisture (%)	Ash (%)	Carb (%)	Vitamin A (mg/100 g)	Vitamin B2 (mg/100 g)	Vitamin C (mg/100 g)	Fe (mg/100 g)	Ca (mg/100 g)	Zn (mg/100 g)	P (mg/100 g)	Mg (mg/100 g)	Fats (mg/100 g)	Reference
<i>Cirina forda</i>	Larvae	20.2	1.8	4.4			3.0	2.2	2.0	64.0	15.4	8.6	110.0	1.9		[6,36]
<i>Imbrasia epimethea</i>	Larvae	73.1		79.8						13.0		11.1	402.0		12.4	[36]
<i>Imbrasia obscura</i>	Larvae	62.3		83.0											12.2	[45]
<i>Gonimbrasia (Nudaurelia) alopia</i>	Larvae	62.3		85.7											1.9	[45]
<i>Gonimbrasia (Nudaurelia) dione</i>	Larvae															[45]
<i>Pseudantheraea discrepans</i>	Larvae	48.9		72.2											21.3	[45]
<i>Anaphe panda</i>	Larvae	53.2		83.4											55.0	[6,33]
<i>Cirina butyrospermi</i>	Larvae	62.7	5.0		5.1					13.0						[46]
<i>Imbrasia belina</i>	Larvae	55.3	16.0		8.3	8.2				31.0		14.0	543.0	160.0		[6,47]
<i>Gyanisa maia</i>	Larvae	51.1	16.2		7.7	14.1									16.4	[47]
<i>Loba leopardina</i>	Larvae	25.8	14.7		6.6	40.2									12.6	[47]
<i>Imbrasia macrothyris</i>	Larvae	75.4														[33]
<i>Nudaurelia macrothyris</i>	Larvae	75.4														[33]
<i>Gonimbrasia richelmanni</i>	Larvae	79.6														[33]
<i>Cirina spp.</i>	Larvae									64.0	7.0	8.6	1090.0	32.4		[48]
<i>Cirina butyrospermi</i>	Larvae	62.7			5.0								1160.0		14.3	[46]
<i>Hemijana variegata Rothschild,</i>	Larvae		8.3	5.9	5.2	9.5										[49]
<i>Anaphe infracta</i>	Larvae	20.0	2.4	2.7	1.6		3.0	2.0	4.5	1.8	8.6		111.3	1.0		[36]
<i>Anaphe reticulata</i>	Larvae	23.0	3.1	3.2	2.5		3.4	2.0	2.2	2.2	10.5		102.3	2.6		[36]
<i>Anaphe spp.</i>	Larvae	18.9	1.7	2.5	4.1		2.8	0.1	3.2	1.6	7.6		122.2	1.0		[36]
<i>Anaphe venata</i>	Larvae	25.7	2.3	3.3	3.2		3.1	1.3	2.2	2.0	8.6		100.5	1.6		[36]
<b>Orthoptera (grasshoppers, locust and crickets)</b>		<b>39.8 ± 21.1</b>	<b>6.4 ± 4.8</b>	<b>3.5 ± 1.7</b>	<b>5.5 ± 4.0</b>	<b>26.8 ± 14.5</b>	<b>3.0 ± 3.5</b>	<b>0.2 ± 0.4</b>	<b>2.9 ± 4.0</b>	<b>120.1 ± 298.8</b>	<b>17.3 ± 15.8</b>	<b>91.1 ± 99.8</b>	<b>119.7 ± 12.7</b>	<b>2.8 ± 3.8</b>	<b>20.8 ± 18.9</b>	

Table 1. Cont.

Scientific Name	Stage of Consumption	Protein (%)	Crude Fibre (%)	Moisture (%)	Ash (%)	Carb (%)	Vitamin A (mg/100 g)	Vitamin B2 (mg/100 g)	Vitamin C (mg/100 g)	Fe (mg/100 g)	Ca (mg/100 g)	Zn (mg/100 g)	P (mg/100 g)	Mg (mg/100 g)	Fats (mg/100 g)	Reference
<i>Brachytrypes membranaceus</i>	Adult	53.4	15.0	3.4	6.0	15.1	0.0	0.0	0.0	0.7	9.2		126.9	0.1	53.0	[6,47]
<i>Cytacanthacris naeruginosus unicolor</i>	Adult	12.1	2.1	2.6			1.0	0.1	1.0	0.4	4.4		100.2	0.1		[6,36]
<i>Zonocerus variegatus</i>	Adult	26.8	2.4	2.6			6.8	0.1	8.6	910.0	42.2		131.2	8.2		[6,36]
<i>Gryllotalpa africana</i>	Adult	22.0	7.5		12.6	47.2									10.8	[47]
<i>Henicus whellani</i>	Adult	53.6	10.6		14.0										4.3	[50]
<i>Cartarrtopsilus taeniolatus</i>	Adult	40.6	13.3		6.9											[35]
<i>Zulua cyanoptera</i>	Adult	33.7	13.3		6.6											[51]
<i>Ornithacris turbida</i>	Adult	42.7	2.0		4.5	18.2									2.0	[47]
<i>Ruspolia differens</i>	Adult	72.7	6.3		4.6			1.2	0.1	13.0	24.5	12.4	121.0	33,1	46.2	[6]
<i>Anacridium melanorhodon melanorhodon</i> (Walker)	Adult	66.2	8.4	7.5											12.4	[52]
<i>Zonocerus variegatus</i>	Adult	62.7	3.6		1.2		8.9	0.1	9.8		2.0	29.0				[6]
<i>Brachytrypes membranaceus</i> L.	Adult															[53]
<i>Zonocerus variegatus</i>	Adult	26.8	2.4	2.6	1.2					2.0	42.2		131.2	8.2		[36]
<i>Brachytrypes</i> spp.	Adult	65.4			4.9					33.6		232.0			16.9	[38]
<i>Brachytrypes</i> spp.	Adult	6.3	1.0	3.4	1.8		0.0	0.0	0.0	0.7	9.2		126.9	0.1		[36]
<i>Cytacanthacris aeruginosus unicolor</i>	Adult	12.1	1.5	2.6	2.1		1.0	0.1	1.0	0.4	4.4		100.2	0.1		[36]
* Recommended daily intakes (mg/day) for adults										45.0	7.5–58.8	1300.0	3.0–14.0	700.0	220–260	[37]

Note the mineral abbreviations are Fe: Iron; Zn: Zinc; Ca: Calcium; P: Phosphorus; Mg: Magnesium. \* Source [37]. Mean  $\pm$  standard deviation of insects belonging to the same insect order are highlighted in bold and species names are in italics.



The crude fibre was reported to be higher in Coleoptera (2–28%) and Lepidoptera (2–16%), while the crude fibre content was reported to be lowest in Hemiptera (0–5%). Lepidoptera had the highest moisture content (3–86%), while Blattodea had the lowest moisture content (2.8–3%) (Table 1).

The highest carbohydrate content was recorded in Coleoptera (13–52%) and Orthoptera (15–47%), while the lowest carbohydrate content was recorded in Blattodea (0–32%). Fat content was the highest in Lepidoptera (2–55%) and lowest in Orthoptera (2–16%) (Table 1).

Orthoptera had the highest iron content (0.3–910 mg/100 g) followed by Blattodea (27–332 mg/100 g), while Hemiptera had the lowest iron content (0–20 mg/100 g). Calcium content was higher in Blattodea (18–132 mg/100 g) and lowest in Lepidoptera (8–15 mg/100 g). The highest Phosphorus was recorded in Lepidoptera (100–730 mg/100 g) and the lowest in Orthoptera (106–125 mg/100 g). Magnesium content was the highest in order Lepidoptera (1–160 mg/100 g), while Blattodea had the lowest magnesium content (0.1–0.3 mg/100 g) (Table 1).

Edible insects are widely consumed in Africa, and play an important role in nutritious diets. However, the preference and consumption of insects vary with species and orders. Lepidoptera caterpillars were the most consumed order, and they are the most preferred species because of their nutritional value, they are rich in protein, fats, and essential micronutrients [6,54]. In addition, several caterpillar species play an important role in income generation in rural areas in southern Africa, Uganda, and Nigeria [18,22,55].

Studies from western and Central Africa indicated that *Rhynchophorus phoenic* (palm weevil), and *Cirina forda* (pallid emperor moth) were the commonly consumed species [18,24,56]. The palm weevil and pallid emperor moth are a delicacy in western and Central Africa, and, in addition, these species were of economic importance in Nigeria, Cameroon, Benin, and Ghana [57]. In southern Africa, the literature indicates that the most consumed or preferred species were *Imbrasia belina* (mopane worm), *Macrotermes natalensis*, *falciger*, and *bellicosus* (termites) [28,50,58]. While in eastern Africa, the most consumed species were *Ruspolia nitidula* and *differens* (grasshoppers), [22,59–61]. Mopane worms, and termites are an important part of food culture in different ethnic groups in southern Africa [18,59]. Moreover, the trade of mopane worms and termites plays an important role in rural food security and income generation, as it provides rural people with household income [28,50,57,58].

Edible insects are a good source of protein content, which ranges from 12–79% of dry matter, which is consistent with studies from China, Germany, and Asia [6,10]. The protein content reported in edible insects is higher than protein found in chicken (43%) or beef (54%) [28,62]. The high protein content found in edible insects could help to combat protein deficiency in Africa. Protein deficiency is a major contributor to human malnutrition [63], and, in Africa, protein deficiency is the most common form of malnutrition, which needs to be addressed to halt starvation [64]. Therefore, including edible insects in daily diets might help reduce malnutrition rates.

Moisture content ranged from 1–7.5%, which is relatively low, such that most edible insects have longer preservation periods, and the risk of microbial deterioration and spoilage is minimal [29,42,65]. Unlike beef or chicken, which are prone to decay (unless refrigerated), edible insects can be stored for longer periods, especially during the dry season when food shortage is higher [42]. However, three caterpillars (*Gonimbrasia* (Nudaurelia) *alopia*, *Anaphe panda*, and *Pseudontheraea discrepans*) had higher moisture (>60%), meaning they are prone to spoilage and their preservation period is shorter unless processed in some manner. Siulapwa et al. [29] reported similar results, where caterpillars *Imbrasia belina* and *Gynanisa maja* had higher moisture content than other species. To increase shelf life, caterpillars are usually degutted, washed in boiling salt water, or roasted before drying in the sun, then packed in large sacks and containers [23,66].

Edible insects contain fat content ranging from 1–67%. The fat content of edible insects are higher in the larval stage. For example, a palm weevil, which is a beetle larva that is consumed as a delicacy in western Africa, contained the highest fat content of 67%. These results are consistent with Bukkens [67], who reported that Lepidopteran caterpillars and palm weevil larvae contain higher fat than any other insect species. Edible insects can be used to provide essential fatty acids required by the

human body [10,68]. In addition, fat plays an important role in providing the human body with energy, which means that consuming insects such as *Rhynchophorus phoenicis*, *Imbrasia belina*, *Anaphe panda*, and *Brachytrupes membranaceus*, may help provide people with energy, thereby reducing malnutrition associated with energy deficiencies in developing countries [4,10,69].

Carbohydrates play a very important role in human nutrition as they are the primary source of energy. Carbohydrates found in edible insects varied from 5–51% [19,70]. Therefore, edible insects can be used as a source of carbohydrates, as they contain relatively high amounts of polysaccharides, which play an important role in enhancing the immune system of the human body [10]. In addition, carbohydrates are an essential nutritive element in the human body [29]. Species such as *Oryctes monoceros* and *Gryllotalpa africana*, reported in the current study, contained a high amount of carbohydrates; therefore, edible insects can be included in human diets to provide a good source of carbohydrates [29].

Excellent source of iron and zinc found in some edible insects indicate that edible insects could be used to combat malnutrition deficiencies such as zinc and iron deficiency anemia, which is prevalent in Africa [37]. Species such as *Zonocerus variegatus*, *Pseudacathotermes spinige*, and *Macrotermes herus* contained high iron content of 910, 332, and 161 mg/100 g respectively, which means that these species can be used as a good source of Iron. Zinc content was notably high in insects such as *Zonocerus variegatus* (29 mg/100 g) and *Rhynchophorus phoenicis* (26.5 mg/100 g) the Zinc content found in these insects exceed the daily recommended intake of 3.0–14 mg/100 g. Rumpold and Schluter [6] reported that Iron and Zinc content found in edible insects is generally higher than the Zinc and Iron content found in pork, beef, or chicken; therefore, edible insects might be a solution in fighting Iron and Zinc deficiency. Zinc and Iron deficiency are one of the health problems faced by many women of reproductive age and children in developing countries [37]. Therefore, consumption of edible insects might provide a solution to Iron deficiency health problems, such as anemia, reduced physical activity, and maternal mortality [37,71].

Edible insects reported in the current study contained a low amount of Vitamin A, B2, and C. The 100 g dry matter of edible insects reported in this study did not contain enough daily recommended Vitamin A (500–600 mg) or C (45 mg). As such, Chen et al. [10] reported that to meet the daily recommended amount of Vitamin C, insect tea derived from the excrement of insects is an option. This tea contains up to 15.04 mg of Vitamin C per 100 g, and the consumption of 300 mL of insect tea per day makes 45 mg of Vitamin C, which is the daily recommended amount of vitamin C for adults [10]. Contrary to findings reported in this study, Bukkens [67] reported that Vitamin B1, B2, and B3 content found in an edible house fly is richer than the Vitamin B1, B2, and B3 found in chicken, beef, or salmon. In addition, edible crickets contain twice more Vitamin B12 than the beef [69]. Igwe et al. [72] found that *Microtermes nigeriensis* contain a favourable high source of Niacin, Thiamine, Vitamin A, and C. Vitamins play an important role in human nutrition, as Vitamin C is important for human growth, development, and repair of various body tissues [73]. The excellent source of Vitamins found in some edible insects shows that insects have a great potential of being used as a healthy food supplement for malnourished people, or to prevent malnutrition [24].

There were several limitations to this review, which included studies reported in English only and excluded studies published in other languages used in Africa. There were significant gaps in data available on the nutritional composition of edible insects consumed in Africa. Most publications focused on a single macronutrient content, especially protein, carbohydrates, fats and fibre, and other nutrients, especially minerals, are not included in analyses. In addition, research focused on reporting the nutritional composition of economically important species such as *Imbrasia belina*, *Macrotermes natalensis*, *bellicosus* and *falciger*, *Rhynchophorus phoenicis*, and *Cirina forda*. Strengths of this review includes the robust approach to combine the nutritional composition of consumed insects in Africa, previous studies have focused on documenting the nutritional composition of single, or a group of, insects that are consumed in Africa.

This review reported combined nutritional data of consumed insects in Africa; this information can be useful to policy makers in the health and nutrition sector by including insects in food and

nutrition policies. Health officials need to motivate people to include insects in their daily diets, particularly the most vulnerable groups such as elderly people, women, and children, with the aim to improve the quality of life for people. In addition, farming and rearing of insects by the agricultural sector need to be adopted to ensure that insects are easily accessible and available all year even when they are out of season in nature. Insects can be included as an ingredient in other food products such as bread, maize powder, chocolate, and biscuits to overcome discomfort and fear associated with eating whole insects in some groups of people. Future studies are required to research sustainable ways of farming and rearing insects in Africa and the implication that might have on the environment.

#### 4. Conclusions

Meeting global food demand and halting poverty in Africa are among the greatest challenges, and these challenges are expected to continue if sustainable and innovative measures are not put into place. In 2017, approximately 256 million people were reported to be undernourished in Africa [74]. There is no doubt that Africa is far from achieving Sustainable Development Goal 2, which is to end hunger, achieve food security and improved nutrition, and promote sustainable agriculture by 2030. Edible insects are widely consumed in Africa, and they play an important socio-economic role for rural communities in Africa, by providing nutritious diets (this review), and income opportunities to traders and harvesters [22,75,76]. In addition, edible insects are a traditional delicacy, and are used as an emergency food source during times of food shortage [57]. They are rich in protein, carbohydrates, amino acids, and micronutrients such as Zinc and Iron. This implies that edible insects have a potential of contributing in sustainable diets, while assuring food security, and improving livelihoods of African people.

**Author Contributions:** Conceptualization of the study was done by Z.T.H., R.S. and T.C.M. methodology, Z.T.H.; data collection, Z.T.H.; writing—original draft preparation, Z.T.H.; writing—review and editing, R.S., T.C.M., and Z.T.H.; supervision, R.S. and T.C.M.; funding acquisition, R.S., and T.C.M. All authors have read and agreed to the published version of the manuscript.

**Funding:** This study was funded by the Sustainable and Healthy Food Systems (SHEFs) supported by the Wellcome Trust's Our Planet, our Health programme (grant number, 205200/Z/16/Z), and the National Research Foundation (NRF) (grant number, 114416).

**Acknowledgments:** This study forms part of the Sustainable and Healthy Food Systems (SHEFs) supported by the Wellcome Trust's Our Planet, our Health programme (grant number, 205200/Z/16/Z). We would like to thank Alan Dangour for providing constructive comments that helped improve this review. Lastly, we would like to thank the National Research Foundation (NRF) for financial support.

**Conflicts of Interest:** The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

## Appendix A

Table A1. Edible insects consumed in different African countries.

Order	Scientific Name/Morpho Species	Common Name	Country	Consumption Stage	References
Blattodea	<i>Periplaneta americana</i>	Common cockroach	Nigeria	Adult	[35]
Coleoptera	<i>Analeptes trifasciata</i>	Stem girdler	Nigeria	Larvae	[24,36,77]
Coleoptera	<i>Oryctes boas Fabr</i>	Rhinoceros beetle	Nigeria, Ivory Coast, Sierra Leone, Liberia, Democratic Republic of Congo, South Africa, Botswana, Namibia, Guinea Bissau	Larvae	[18,24,33,36,78,79]
Coleoptera	<i>Oryctes monoceros</i>	Rhinoceros beetle	Nigeria	Larvae	[24,33,36,39,56,79]
Coleoptera	<i>Aphodius rufipes</i>	Dung beetle	Nigeria	Larvae	[24,36,80]
Coleoptera	<i>Rhynchophorus phoenicis</i>	Palm weevil	Nigeria, Angola, Burkina Faso, Cameroon; Ghana, Cote D'ivoire, Democratic Republic of Congo, Liberia, Niger, Sao Tome, Togo, Benin, Guinea Bissau	Larvae, pupa and adult	[18,24,33,36,39,42,56,57,77,79,81–98]
Coleoptera	<i>Heteroligus meles</i>	Yam beetle	Nigeria	Larvae, pupa, adult	[24,36,42,77,79,91,99,100]
Coleoptera	<i>Eulepida mashona</i>	Beetle	Zimbabwe	Larvae/adult	[51,58]
Coleoptera	<i>Carbula marginella</i>	Beetle	Burkina Faso	Adult	[98]
Coleoptera	<i>oryctes sp.</i>	Beetle	Burkina Faso	Larvae	[98]
Coleoptera	<i>Oryctes rhinoceros larva</i>	Beetle	Nigeria; Cote D'ivoire	Larvae	[79,81,99,101,102]
Coleoptera	<i>Stenorcera orissa Buq</i>	Giant jewel beetle	Botswana, Zimbabwe	Winged adult	[58,78]
Coleoptera	<i>Eulepida anatine</i>	Beetle	Zimbabwe	Larvae	[58]
Coleoptera	<i>Eulepida nitidicollis</i>	Beetle	Zimbabwe	Larvae	[58]
Coleoptera	<i>Apomecyna parumpunctata</i>	African longhorned beetle	Nigeria	Larvae	[43]
Coleoptera	<i>Oryctes owariensis</i>	Beetle	Cote D'ivoire, Democratic Republic of Congo, South Africa, Angola, Malawi, Botswana, Mozambique, Zambia, Zimbabwe	Adult	[18,33,40]
Coleoptera	<i>Rhinoceros oryctes</i>	Beetle	Nigeria, Ivory Coast, Sierra Leone, Guinea, Ghana, Equatorial Guinea, Guinea Bissau	Larvae, pupa, adult	[91]
Coleoptera	<i>Sitophilus oryzae</i>	Rice weevil	Nigeria	Larvae, pupa, adult	[91]

Table A1. Cont.

Order	Scientific Name/Morpho Species	Common Name	Country	Consumption Stage	References
Coleoptera	<i>Callosobruchus maculatus</i>	Bean beetle	Nigeria	Larvae, pupa, adult	[91]
Coleoptera	<i>Dermestes maculatus</i>	Beetle	Nigeria	Larvae, pupa, adult	[91]
Coleoptera	<i>Cotinis nitida</i>	Beetle	Nigeria	Adult/larvae	[79]
Coleoptera	<i>Eulopida mashona</i>	Beetle	Zimbabwe	Adult/larvae	[47]
Coleoptera	<i>Sternocera funebris</i>	Beetle	Zimbabwe	Adult/larvae	[47]
Coleoptera	<i>Oryctes</i> spp Oliver	Beetle	Nigeria	Larvae	[77]
Coleoptera	<i>Augosoma centaurus</i>	Beetle	Cameroon	Adult, larvae	[57]
Coleoptera	<i>Phyllophaga nebulosa</i> (Harris)	Beetle larvae	Ghana	Larvae	[94,103]
Coleoptera	<i>Sitophilus zeamais</i>	Beetle	Ghana	Larvae, adult	[104]
Coleoptera	<i>Polycleis equestris</i>	Weevil	South Africa	Adult	[33]
Coleoptera	<i>Polycleis plumbeus</i>	Weevil	South Africa	Adult	[33]
Coleoptera	<i>Sipalus aloysii-sabaudiae</i>	Beetle	South Africa	Larvae	[33]
Coleoptera	<i>Teratobus flabellicornis</i>	Beetle	South Africa	Larvae	[33]
Coleoptera	<i>Sternocera orissa</i>	Beetle	South Africa	Larvae	[33]
Diptera	<i>Chaoborus edulis</i>		Malawi	Adult	[33]
Hemiptera	<i>Nezara viridula</i>	Southern green stink bug	Nigeria	Adult	[24,36,99]
Hemiptera	<i>Encosternum delegorgui</i> Spinola	Stink bug	South Africa, Zimbabwe, Swaziland, Malawi, Botswana, Namibia, Mozambique	Adult	[18,28,33,47,58,105]
Hemiptera	<i>Monomatapa insingnis</i> Distant	Cicada	Botswana	Adult	[78]
Hemiptera	<i>Aspongubus viduatus</i>	Melon bug	Sudan	Adult	[106]
Hemiptera	<i>Agonoscelis pubescens</i>	Sorghum bug	Sudan	Adult	[106]
Hemiptera	<i>Rhynchophorus</i> spp.	May bug	Nigeria, Cameroon	Larvae	[79,107]
Hemiptera	<i>Brevisana brevis</i>	African cicada	Zimbabwe	Adult	[47]
Hemiptera	<i>Ugada limbalis</i>	Cicada	Uganda		[108]
Hemiptera	<i>Pediculus capitata</i>		Angola, Malawi, South Africa, Zambia, Zimbabwe, Mozambique, Namibia, Botswana		[33]

Table A1. Cont.

Order	Scientific Name/Morpho Species	Common Name	Country	Consumption Stage	References
Hymenoptera	<i>Apis mellifera</i>	Honey bee	Nigeria, Botswana, Cote D'ivoire, Cameroon, Zambia, Zimbabwe, Botswana, Angola, Mozambique, Tanzania, Senegal, Ghana, Lesotho, Benin, South Africa	Egg, larva, pupa	[2,18,33,36,38,77,78,91,107,109,110]
Hymenoptera	<i>Carebara vidua</i>	African thief ant	Botswana, Zimbabwe; Kenya Burundi, South Africa, Malawi, Zambia, Sudan, Namibia, Mozambique	Winged adult	[18,33,44,47,58,78,81,108]
Hymenoptera	<i>Plebeina hildebrandti</i> Friese	Stingless bee	Botswana	Adult	[78]
Hymenoptera	<i>Hypotrigona gribodoi</i> Magretti	Stingless bee	Botswana	Adult	[78]
Hymenoptera	<i>Cossus cossus</i>	Capenter ant	Cote D'ivoire	Adult	[102]
Hymenoptera	<i>Componotus spp.</i>	Ant	Nigeria	Adult	[35]
Hymenoptera	<i>Oecophylla longinoda</i>	African weaver ant	Nigeria, Cameroon	Adult	[35,93,107]
Hymenoptera	<i>Carebara lignata</i>	Ant	Zambia, South Africa, Democratic Republic of Congo, Zimbabwe, Botswana, Mozambique, Namibia, Sudan	Adult	[18]
Blattodea	<i>Macrotermes nigeriensis</i>	Termite	Nigeria	Winged adult, queen	[24,33,35,36,72,111,112]
Blattodea	<i>Macrotermes bellicosus</i>	Termite	Nigeria, Kenya, Uganda, Democratic Republic of Congo, Cameroon, Cote D'ivoire, Sao Tome, Togo, Liberia, Burundi, Ghana, Zimbabwe,	Winged adult, queen	[18,24,33,36,59,77,79,82,94,109,111,113–115]
Blattodea	<i>Macrotermes natalensis</i>	Termite	Nigeria, South Africa, Zimbabwe, Cameroon, Democratic Republic of Congo, Burundi, Malawi	Winged adult, queen	[24,33,36,47,58,75,99]
Blattodea	<i>Macrotermes falciger</i>	Termite	Democratic Republic of Congo; South Africa, Zimbabwe, Burundi, Zambia, Burkina Faso, Benin	Winged adult	[18,33,58,75,108,115–117]
Blattodea	<i>Macrotermes michaelsoni</i>	Termite	South Africa	Winged adult	[75]
Blattodea	<i>Macrotermes subhyalinus</i>	Termite	Burkina Faso Zimbabwe, Cote D'ivoire, Rwanda, Uganda, Angola, Togo, Kenya	Adult	[18,33,58,98,102,108,118]



Table A1. Cont.

Order	Scientific Name/Morpho Species	Common Name	Country	Consumption Stage	References
Blattodea	<i>Hodotermes mossambicus</i> (Hagen)	Harvester termite	Botswana	Larvae	[78]
Blattodea	<i>Macrotermes sp.</i>	Termite	Nigeria, Uganda	Adult queen, soldiers	[35,38,91]
Blattodea	<i>Syntermes soldiers</i>	Termite	Uganda	Adult	[38]
Blattodea	<i>Pseudacanthotermes militaris</i>	Termite	Kenya, Uganda	Winged adult	[59,108,115]
Blattodea	<i>Pseudacanthotermes spiniger</i>	Termite	Kenya, Uganda, Burundi	Winged adult	[59,108,115]
Blattodea	<i>Odontotermes kibarensis</i>	Termite	Uganda	Winged adult	[108]
Blattodea	<i>Pseudacanthotermes sp.1</i>	Termite	Uganda	Winged adult	[108]
Blattodea	<i>Pseudacanthotermes sp.2</i>	Termite	Uganda	Winged adult	[108]
Blattodea	<i>Odontotermes spp.</i>	Termite	Uganda	Winged adult	[108]
Blattodea	<i>Pseudacanthotermes sp.5</i>	Termite	Uganda	Winged adult	[108]
Blattodea	<i>Pseudacanthotermes sp. 4</i>	Termite	Burundi	Adult	[108]
Blattodea	<i>Macrotermes spp.</i>	Termite	Rwanda, Cameroon	Winged adult	[93,107,108]
Blattodea	<i>Macrotermes swaziae</i>	Termite	Zimbabwe		[33]
Blattodea	<i>Microhodotermes viator</i>	Termite	South Africa		[33]
Blattodea	<i>Termes badius</i>	Termite	South Africa	Winged adult	[33]
Lepidoptera	<i>Anaphe venata</i>	African silkworm	Nigeria, Zambia, Cote D'ivoire, Sierra Leona, Guinea, Liberia, Guinea Bissau, Angola	Larvae	[18,24,33,36,77,96]
Lepidoptera	<i>Anaphe infracta</i>	African silkworm	Nigeria	Larvae	[24,33]
Lepidoptera	<i>Anaphe recticulata</i>	African silkworm	Nigeria	Larvae	[24,33,36]
Lepidoptera	<i>Bunaea alcinoe</i>	Emperor moth	Nigeria; Democratic Republic of Congo, Botswana, Zimbabwe, Cameroon, Zambia, South Africa, Democratic Republic of Congo, Tanzania	Larvae, pupa and adult	[18,24,36,58,77–79,87,93,99,117]
Lepidoptera	<i>Lepidoptara litoralia</i>	Caterpillar	Nigeria	Larvae	[24,36,119]
Lepidoptera	<i>Cirina forda</i>	Pallid emperor	Nigeria, Angola, Democratic Republic of Congo, Botswana, Zimbabwe; Togo, Zambia, Mozambique, Ghana, Namibia	Larvae	[18,24,30,31,33,36,48,56,58,78,96,112,117,120–123]

Table A1. Cont.

Order	Scientific Name/Morpho Species	Common Name	Country	Consumption Stage	References
Lepidoptera	<i>Imbrasia epimethea</i>	Caterpillar	Angola, Democratic Republic of Congo	Larvae	[18,33,45]
Lepidoptera	<i>Imbrasia obscura</i>	Caterpillar	Angola	Larvae	[96]
Lepidoptera	<i>Imbrasia truncata</i>	Caterpillar	Angola	Larvae	[96]
Lepidoptera	<i>Gonimbrasia (Nudaurelia) alopia</i>	Caterpillar	Angola	Larvae	[96]
Lepidoptera	<i>Gonimbrasia (Nudaurelia) dione</i>	Caterpillar	Angola	Larvae	[96]
Lepidoptera	<i>Pseudantheraea discrepans</i>	Caterpillar	Angola	Larvae	[96]
Lepidoptera	<i>Micragone cana</i>	Caterpillar	Angola, Democratic Republic of Congo	Larvae	[33,96]
Lepidoptera	<i>Anaphe panda</i>	Bagnest moth	Angola, Zimbabwe, Zambia, Cameroon, Democratic Republic of Congo, Nigeria, Tanzania	Larvae	[18,33,47,58,124]
Lepidoptera	<i>Notodontidae</i> sp. 1	Caterpillar	Angola	Larvae	[96]
Lepidoptera	<i>Notodontidae</i> sp. 2	Caterpillar	Angola	Larvae	[96]
Lepidoptera	<i>Notodontidae</i> sp. 3	Caterpillar	Angola	Larvae	[96]
Lepidoptera	<i>Notodontidae</i> sp. 4	Caterpillar	Angola	Larvae	[96]
Lepidoptera	<i>Gastropalakaeis rubroanalis</i>	Caterpillar	Angola	Larvae	[96]
Lepidoptera	<i>Sciatta inconcisa</i>	Caterpillar	Angola	Larvae	[96]
Lepidoptera	<i>Elaphrodes lactea</i> Gaede	Caterpillar	Democratic Republic of Congo	Larvae	[33,117]
Lepidoptera	<i>Lobobunaea saturnus</i> Fabricius	Caterpillar	Democratic Republic of Congo, Zimbabwe	Larvae	[33,58,117]
Lepidoptera	<i>Cinabra hyperbius</i> (Westwood)	Caterpillar	Democratic Republic of Congo	Larvae	[33,117]
Lepidoptera	<i>Gonimbrasia richelmanni</i> Weymer	Caterpillar	Democratic Republic of Congo	Larvae	[33,117]
Lepidoptera	<i>Antheua insignata</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33,117]
Lepidoptera	<i>Imbrasia rubra</i>	Caterpillar	Democratic Republic of Congo	Larvae	[117]
Lepidoptera	<i>Athletes semialba</i> (Sonthonnax)	Caterpillar	Democratic Republic of Congo, Zimbabwe, Zambia, South Africa, Namibia, Mozambique	Larvae	[33,58,117]



Table A1. Cont.

Order	Scientific Name/Morpho Species	Common Name	Country	Consumption Stage	References
Lepidoptera	<i>Cirina butyrospermi</i>	Caterpillar	Burkina Faso, Cote D'ivoire, Zambia, Zimbabwe, South Africa, Nigeria, Mali, Ghana	Larvae	[18,46,102,103,118,125]
Lepidoptera	<i>Hemijana variegata</i>	Caterpillar	South Africa	Larvae	[49]
Lepidoptera	<i>Imbrasia belina</i>	Mopane worm	Nigeria; Botswana; Zimbabwe, Namibia, South Africa, Malawi, Zambia, Angola, Mozambique	Larvae	[18,33,34,47,58,76,78,91,109,126–129]
Lepidoptera	<i>Isobertina paniculata</i>	Caterpillar	Zambia	Larvae	[127]
Lepidoptera	<i>Urota sinope</i>	Caterpillar	Zambia, Botswana	Larvae	[78,130]
Lepidoptera	<i>Gonimbrasia zambesina</i>	Caterpillar	Zambia; Zimbabwe, Democratic Republic of Congo	Larvae	[33,58,130]
Lepidoptera	<i>Lophostethus dumolinii</i>	Arrow sphinx	Botswana	Larvae	[78]
Lepidoptera	<i>Angas</i>				
Lepidoptera	<i>Daphnis nerii</i> L	Oleander hawk moth	Botswana	Larvae	[78]
Lepidoptera	<i>Heniocha</i> spp.	Marbled emperor moth	Botswana	Larvae	[78]
Lepidoptera	<i>Imbrasia tyrreha</i>	Willow emperor moth	Botswana	Larvae	[78]
Lepidoptera	<i>Sphingomorpha chlorea</i>	Sundown emperor moth	Botswana	Larvae	[78]
Lepidoptera	<i>Hippotion celerio</i> L.	Silver striped hawk	Botswana	Adult	[78]
Lepidoptera	<i>Agrius convolvuli</i> L.	Convolvulus hawk moth.	Botswana, South Africa, Angola, Zimbabwe, Zambia, Malawi, Mozambique, Namibia	Larvae	[78]
Lepidoptera	<i>Gonanisa maia</i>	Caterpillar	Zimbabwe, Botswana, Malawi, Democratic Republic of Congo, South Africa	Larvae	[33,47,58]
Lepidoptera	<i>Anthoera zambezina</i>	Caterpillar	Zimbabwe, Botswana, Malawi, Namibia, Zambia, South Africa, Mozambique, Angola	Larvae	[33,58]
Lepidoptera	<i>Athletes gigas</i>	Caterpillar	Zimbabwe, Botswana, Malawi, Namibia, Zambia, South Africa, Mozambique, Angola	Larvae	[33,58]
Lepidoptera	<i>Bombycomorpha pallida</i>	Moth	Zimbabwe, South Africa	Larvae	[33,58]

Table A1. Cont.

Order	Scientific Name/Morpho Species	Common Name	Country	Consumption Stage	References
Lepidoptera	<i>Bunaea caffra</i>	Moth	Zimbabwe, Zambia, South Africa, Namibia, Botswana, Mozambique, Angola	Larvae	[33,58]
Lepidoptera	<i>Bunaeopsis aurantica</i>	Moth	Zimbabwe, Democratic Republic of Congo	Larvae	[33,58]
Lepidoptera	<i>Gonometa postica</i>	Moth	Zimbabwe, South Africa	Larvae	[33,58]
Lepidoptera	<i>Heniocha dyops</i>	Moth	Zimbabwe, South Africa, Botswana, Zambia, Malawi, Namibia, Mozambique, Angola	Larvae	[33,58]
Lepidoptera	<i>Imbrasia epimethea</i>	Moth	Zimbabwe, Democratic Republic of Congo	Larvae	[33,58]
Lepidoptera	<i>Imbrasia ertli</i>	Caterpillar	Zimbabwe, South Africa, Cameroon, Democratic Republic of Congo, Angola, Zimbabwe, Botswana, Angola	Larvae	[18,33,58]
Lepidoptera	<i>Nudaurelia belina</i>	Moth	Zimbabwe, Malawi, Botswana, Mozambique, Namibia, Zambia, South Africa	Larvae	[33,58]
Lepidoptera	<i>Pseudobunaea irius</i>	Moth	Zimbabwe, South Africa, Zambia, Angola, Malawi, Namibia,	Larvae	[33,58]
Lepidoptera	<i>Loba leopardina</i>	Moth	Zimbabwe	Larvae	[58]
Lepidoptera	<i>Imbrasia oyemensis</i>	Caterpillar	Cote D'ivoire	Adult	[102]
Lepidoptera	<i>Imbrasia</i> spp.	Caterpillar	Cameroon	Larvae	[93]
Lepidoptera	<i>Eumeta</i> spp.	Caterpillar	Cameroon	Larvae	[107]
Lepidoptera	<i>Anaphe</i> spp.	Caterpillar	Cameroon	Larvae	[107]
Lepidoptera	<i>Dactyloceras</i> spp.	Caterpillar	Cameroon	Larvae	[107]
Lepidoptera	<i>Bunaea</i> spp.	Caterpillar	Cameroon	Larvae	[107]
Lepidoptera	<i>Dactyloceras lucina</i>	Caterpillar	Democratic Republic of Congo, Zambia, South Africa, Cameroon, Angola, Gabon, Sierra Leone, Equatorial Guinea, Sao Tome, Zambia, Democratic Republic of Congo, Sierra Leone, Rwanda, Burundi, Equatorial Guinea, Sao Tome,	Larvae	[18]
Lepidoptera	<i>Platysphinx stigmatica</i>	Caterpillar		Larvae	[18]

Table A1. Cont.

Order	Scientific Name/Morpho Species	Common Name	Country	Consumption Stage	References
Lepidoptera	<i>Epanaphe carteri</i>	Caterpillar	Democratic Republic of Congo, Angola, Gabon, Sierra Leone, Sao Tome, Equatorial Guinea	Larvae	[18]
Lepidoptera	<i>Gynanisa ata</i>	Caterpillar	Democratic Republic of Congo, Zambia, Malawi, Sudan	Larvae	[18]
Lepidoptera	<i>Eumeta cervina</i>	Caterpillar	Democratic Republic of Congo, Cameroon, Angola, Gabon, Sierra Leone, Sao Tome, Equatorial Guinea, Rwanda, Burundi, Liberia	Larvae	[18]
Lepidoptera	<i>Urota sinope</i>	Caterpillar	Democratic Republic of Congo, South Africa, Zimbabwe, Zimbabwe, Botswana, Gabon, Mozambique, Namibia	Larvae	[18,33]
Lepidoptera	<i>Anthoera caffraria</i>	Caterpillar	Angola, Malawi, South Africa, Zambia, Zimbabwe, Mozambique, Namibia, Botswana	Larvae	[33]
Lepidoptera	<i>Anthoera menippe</i>	Caterpillar	Angola, Malawi, South Africa, Zambia, Zimbabwe, Mozambique, Namibia, Botswana	Larvae	[33]
Lepidoptera	<i>Bunaea caffraria</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]
Lepidoptera	<i>Drapetides uniformis</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]
Lepidoptera	<i>Gonimbrasia hecate</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]
Lepidoptera	<i>Goodia kuntzei</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]
Lepidoptera	<i>Heniocha apollonia</i>	Caterpillar	Angola, Malawi, South Africa, Zambia, Zimbabwe, Mozambique, Namibia, Botswana	Larvae	[33]
Lepidoptera	<i>Heniocha marnois</i>	Caterpillar	Angola, Malawi, South Africa, Zambia, Zimbabwe, Mozambique, Namibia, Botswana	Larvae	[33]
Lepidoptera	<i>Herse convolvuli</i>	Caterpillar	South Africa	Larvae	[33]
Lepidoptera	<i>Imbrasia dione</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]
Lepidoptera	<i>Imbrasia macrothyris</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]
Lepidoptera	<i>Imbrasia rubra</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]
Lepidoptera	<i>Lobobunaea phaedusa</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]

Table A1. Cont.

Order	Scientific Name/Morpho Species	Common Name	Country	Consumption Stage	References
Lepidoptera	<i>Melanocera parva</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]
Lepidoptera	<i>Microgene cana</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]
Lepidoptera	<i>Nudaurelia macrothyrsus</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]
Lepidoptera	<i>Nyodes prasinodes</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]
Lepidoptera	<i>Rohaniella pygmaea</i>	Caterpillar	Angola, Malawi, South Africa, Zambia, Zimbabwe, Mozambique, Namibia, Botswana	Larvae	[33]
Lepidoptera	<i>Rheneae mediata</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]
Lepidoptera	<i>Tagoropsis flavinata</i>	Caterpillar	Democratic Republic of Congo	Larvae	[33]
Lepidoptera	<i>Usta terpisichore</i>	Caterpillar	Angola	Larvae	[33]
Lepidoptera	<i>Usta wallengreni</i>	Caterpillar	Angola, Malawi, South Africa, Zambia, Zimbabwe, Mozambique, Namibia, Botswana	Larvae	[33]
Mantodea	<i>Mantis religiosa</i>	African mantis	Nigeria, South Africa	Adult	[33,79]
Orthoptera	<i>Brachytrupes membranaceus</i>	Giant African cricket	Nigeria, Angola; Zimbabwe, Uganda; Cameroon, Democratic Republic of Congo, Burkina Faso, Tanzania, Angola, Togo, Benin; Malawi	Adult	[18,24,33,36,45,53,58,77,79,91,93,96,99,108,124]
Orthoptera	<i>Gymnogryllus lucens</i>	Cricket	Nigeria	Adult	[24,36,116]
Orthoptera	<i>Cytacanthacris naeruginosus</i>	Short horned grasshopper	Nigeria	Adult	[24,36]
Orthoptera	<i>Zonocerus variegatus</i>	Grasshopper	Nigeria, Cameroon, Uganda, Democratic Republic of Congo, Cote D'ivoire, Ghana, Guinea, Liberia, Sao Tome, Liberia, Guinea Bissau	Adult	[18,24,35,36,38,77,79,85,94,99,110,116,131]
Orthoptera	<i>Gryllotalpa africana</i>	Mole cricket	Nigeria; Zimbabwe; Malawi Kenya, Tanzania, Democratic Republic of Congo, Uganda, Zimbabwe, Rwanda, Cameroon, Uganda, Malawi, South Africa	Adult	[24,33,36,47,77,79,99,124]
Orthoptera	<i>Ruspolia differens</i>	Grasshopper	Democratic Republic of Congo, Uganda, Zimbabwe, Rwanda, Cameroon, Uganda, Malawi, South Africa	Adult	[33,58–60,108,112,115,117,132]
Orthoptera	<i>Melanoplus foedus</i>	Grasshopper	Nigeria	Adult	[112]
Orthoptera	<i>Gryllus assimilis</i>	Cricket	Nigeria; Ghana	Adult	[94,112]

Table A1. Cont.

Order	Scientific Name/Morpho Species	Common Name	Country	Consumption Stage	References
Orthoptera	<i>Henicus whellani</i>	Cricket	Zimbabwe	Adult	[50,51]
Orthoptera	<i>Kraussaria ongulifera</i>	Grasshopper	Burkina Faso	Adult	[98,118]
Orthoptera	<i>Gryllus campestris</i>	Field cricket	Burkina Faso; Cameroon, Malawi	Adult	[98,107,124]
Orthoptera	<i>Ruspolia nitidula</i>	Grasshopper	Uganda Botswana; Uganda, Zambia, South Africa, Democratic Republic of Congo, Zimbabwe, Botswana, Nigeria, Uganda, Tanzania, Malawi, Mozambique	Larvae and adult	[22,133]
Orthoptera	<i>Normadacris septemfasciata</i>	Red locust	Botswana, South Africa, Zimbabwe, Malawi, Libya	Adult	[18,78,108]
Orthoptera	<i>Locustana pardalina</i>	Brown locust	Botswana, Zambia, South Africa, Cameroon, Democratic Republic of Congo, Zimbabwe, Burkina, Faso, Malawi, Mali, Niger, Togo, Benin	Adult	[18,33,78]
Orthoptera	<i>Schistocerca gregaria</i>	Desert locust	Botswana	Adult	[78]
Orthoptera	<i>Cyrtacanthacris tatarica</i> L.	Brown-spotted locust	Botswana	Adult	[78]
Orthoptera	<i>Acrida acuminata</i>	Common stick grasshopper	Botswana	Adult	[78]
Orthoptera	<i>Zonocerus elegans</i>	Elegant grasshopper	Botswana, South Africa	Adult	[33,78]
Orthoptera	<i>Acrotylus</i> spp.	Burrowing grasshopper	Botswana	Adult	[78]
Orthoptera	<i>Homorocoryphus nitidulus</i>	Cricket	Cameroon	Larvae	[85,107]
Orthoptera	<i>Gynanisa maia</i>	Cricket	Zimbabwe, Malawi, South Africa	Larvae	[33,58]
Orthoptera	<i>Locusta migratoria</i>	migratory locust	Zimbabwe, Cote D'ivoire; Nigeria; Sudan, Zambia, Democratic Republic of Congo, Sudan, Ghana	Adult	[18,33,79,94,102,134,135]
Orthoptera	<i>Acheta domesticus</i>	Cricket	Cote D'ivoire; Nigeria, Ghana	Adult	[79,94,102]
Orthoptera	<i>Cartarrtopsilus taeniolatus</i>	Grasshopper	Nigeria	Adult	[35]
Orthoptera	<i>Zulua cyanoptera</i>	Grasshopper	Nigeria	Adult	[35]
Orthoptera	<i>Brachytrupes</i> spp.	Cricket	Uganda, Cameroon	Adult	[107,110]
Orthoptera	<i>Cyrtacanthacris aeruginosa unicolor</i>	Grasshopper	Uganda	Adult	[110]
Orthoptera	<i>Zonocerus</i> sp.	Grasshopper	Nigeria	Adult	[91]
Orthoptera	<i>Daraba (Sceloides) laisalis</i>	Locust	Nigeria	Larvae, pupa, adult	[91]
Orthoptera	<i>Ornithacris turbida</i>	Grasshopper	Zimbabwe	Adult	[47]

Table A1. Cont.

Order	Scientific Name/Morpho Species	Common Name	Country	Consumption Stage	References
Orthoptera	<i>Acanthoplus discoidalis</i>	Cricket	Zimbabwe	Adult	[47]
Orthoptera	<i>Acanthacris ruficornis</i>	Garden locust	Uganda, Zambia, South Africa, Cameroon, Democratic Republic of Congo, Zimbabwe, Burkina Faso, Malawi, Mali, Niger, Togo, Benin	Adult	[18,108]
Orthoptera	<i>Schistocerca</i> spp.	Grasshopper	Cameroon	Adult	[107]
Orthoptera	<i>Acanthacris</i> spp.	Grasshopper	Cameroon	Adult	[107]
Orthoptera	<i>Gastrimargus</i> spp.	Locust	Cameroon	Adult	[107]
Orthoptera	<i>Phymateus</i> spp.	Locust	Cameroon	Adult	[107]
Orthoptera	<i>Anacridium</i> spp.	Locust	Cameroon	Adult	[107]
Orthoptera	<i>Pyrgomorpha</i> spp.	Locust	Cameroon	Adult	[107]
Orthoptera	<i>Gastrimargus africanus</i>	Locust	Cameroon, Democratic Republic of Congo, Niger, Lesotho, Liberia	Adult	[18]
Orthoptera	<i>Phymateus viridipes brunneri</i> Bolivar	Locust	Zambia, South Africa, Democratic Republic of Congo, Zimbabwe, Botswana, Mozambique, Namibia	Adult	[18]
Orthoptera	<i>Gryllus bimaculatus</i>	Cricket	Togo, Nigeria, Guinea Bissau, Sierra Leone, Liberia, Benin, Democratic Republic of Congo, Kenya, Sudan, Zambia	Adult	[18]
Orthoptera	<i>Anacridium melanorhodon melanorhodon</i>	Cricket	Cameroon, Sudan, Niger	Adult	[18,52]
Orthoptera	<i>Paracinema tricolor</i>	Cricket	Cameroon, Malawi, Lesotho	Adult	[18]
Orthoptera	<i>Acheta</i> spp.	Cricket	Zambia, Zimbabwe, Kenya	Adult	[18]
Orthoptera	<i>Scapteriscus vicinus</i>	Field cricket	Ghana	Adult	[94,103]
Orthoptera	<i>Gryllotalpa gryllotalpa</i>	Mole cricket	Malawi	Adult	[124]
Orthoptera	<i>Homorocoryphus vicinus</i>	Cricket	Uganda	Adult	[33]
Orthoptera	<i>Nomadacris septumfasciata</i>	Cricket	South Africa	Adult	[33]
Orthoptera	<i>Schistocerca gregaria</i>	Cricket	Zimbabwe	Adult	[33]
Blattodea	<i>Pseudacathotermes spinige</i>	Termite	Kenya	Adult	[37]
Blattodea	<i>Macrotermes</i> spp.	Termite	Kenya	Adult	[37]
Blattodea	<i>Macrotermes subhylanus</i>	Termite	Kenya	Adult	[37]
Hymenoptera	<i>Crematogaster mimosae</i>	Ant	Kenya	Adult	[37]

## References

1. Van Huis, A. Potential of Insects as Food and Feed in Assuring Food Security. *Annu. Rev. Entomol.* **2013**, *58*, 563–583. [[CrossRef](#)]
2. Verbeke, W. Profiling consumers who are ready to adopt insects as a meat substitute in a Western society. *Food Qual. Prefer.* **2015**, *39*, 147–155. [[CrossRef](#)]
3. Nowak, V.; Persijn, D.; Rittenschober, D.; Charrondiere, U.R. Review of food composition data for edible insects. *Food Chem.* **2016**, *193*, 39–46. [[CrossRef](#)] [[PubMed](#)]
4. Van Huis, A. Edible insects are the future? *Proc. Nutr. Soc.* **2016**, *75*, 294–305. [[CrossRef](#)] [[PubMed](#)]
5. Myers, G.; Pettigrew, S. A qualitative exploration of the factors underlying seniors' receptiveness to entomophagy. *Food Res. Int.* **2017**, *103*, 163–169. [[CrossRef](#)] [[PubMed](#)]
6. Rumpold, B.A.; Schlüter, O.K. Nutritional composition and safety aspects of edible insects. *Mol. Nutr. Food Res.* **2013**, *57*, 802–823. [[CrossRef](#)] [[PubMed](#)]
7. Dobermann, D.; Swift, J.A.; Field, L.M. Opportunities and hurdles of edible insects for food and feed. *Nutr. Bull.* **2017**, *42*, 293–308. [[CrossRef](#)]
8. Hanboonsong, Y.; Jamjanya, T.; Durst, P.B. *Six-legged Livestock: Edible Insect Farming, Collect on and Market in Thailand*; Food and Agriculture Organization of the United Nations Regional Office for Asia and The Pacific: Bangkok, Thailand, 2013; pp. 1–57.
9. Halloran, A.; Roos, N.; Hanboonsong, Y. Cricket farming as a livelihood strategy in Thailand. *Geogr. J.* **2017**, *183*, 112–124. [[CrossRef](#)]
10. Chen, X.; Feng, Y.; Chen, Z. Common edible insects and their utilization in China: Invited review. *Entomol. Res.* **2009**, *39*, 299–303. [[CrossRef](#)]
11. Feng, Y.; Chen, X.M.; Zhao, M.; He, Z.; Sun, L.; Wang, C.Y.; Ding, W.F. Edible insects in China: Utilization and prospects. *Insect Sci.* **2018**, *25*, 184–198. [[CrossRef](#)]
12. Ramos-Elorduy, J. Insects: A sustainable source of food? *Ecol. Food Nutr.* **1997**, *36*, 247–276. [[CrossRef](#)]
13. Ramos-Elorduy, J.; Moreno, J.M.P. Edible insects of Chiapas, Mexico. *Ecol. Food Nutr.* **2002**, *41*, 271–299. [[CrossRef](#)]
14. Ramos-Elorduy, J.; Moreno, J.M.P.; Prado, E.E.; Perez, M.A.; Otero, J.L.; De Guevara, O.L. Nutritional value of edible insects from the state of Oaxaca, Mexico. *J. Food Compos. Anal.* **1997**, *10*, 142–157. [[CrossRef](#)]
15. Acuña, A.M.; Caso, L.; Aliphath, M.M.; Vergara, C.H. Edible insects as part of the traditional food system of the Popoloca Town of Los Reyes Metzontla, Mexico. *J. Ethnobiol.* **2011**, *31*, 150–169. [[CrossRef](#)]
16. Costa-Neto, E.M. Anthro-entomophagy in Latin America: An overview of the importance of edible insects to local communities. *J. Insects Food Feed* **2015**, *1*, 17–23. [[CrossRef](#)]
17. Mitsuhashi, J. Insects as traditional foods in Japan. *Ecol. Food Nutr.* **1997**, *36*, 187–199. [[CrossRef](#)]
18. Kelemu, S.; Niassy, S.; Torto, B.; Fiaboe, K.; Affognon, H.; Tonnang, H.; Maniania, N.K.; Ekesi, S. African edible insects for food and feed: Inventory, diversity, commonalities and contribution to food security. *J. Insects Food Feed* **2015**, *1*, 103–119. [[CrossRef](#)]
19. Kouřimská, L.; Adámková, A. Nutritional and sensory quality of edible insects. *NFS J.* **2016**, *4*, 22–26. [[CrossRef](#)]
20. Sogari, G.; Menozzi, D.; Mora, C. Exploring young foodies' knowledge and attitude regarding entomophagy: A qualitative study in Italy. *Int. J. Gastron. Food Sci.* **2017**, *7*, 16–19. [[CrossRef](#)]
21. De Foliar, G.R. An overview of the role of edible insects in preserving biodiversity. *Ecol. Food Nutr.* **1997**, *36*, 109–132. [[CrossRef](#)]
22. Agea, J.; Biryomumaisho, D.; Buyinza, M.; Nabanoga, G. Commercialization of *Ruspolia nitidula* (nseene grasshoppers) in Central Uganda. *Afr. J. Food Agric. Nutr. Dev.* **2008**, *8*, 319–332. [[CrossRef](#)]
23. Mutungi, C.; Irungu, F.G.; Nduko, J.; Mutua, F.; Affognon, H.; Nakimbugwe, D.; Ekesi, S.; Fiaboe, K.K.M. Postharvest processes of edible insects in Africa: A review of processing methods, and the implications for nutrition, safety and new products development. *Crit. Rev. Food Sci. Nutr.* **2019**, *59*, 276–298. [[CrossRef](#)] [[PubMed](#)]
24. Alamu, O.T.; Amao, O.A.; Nwokedi, I.C.; Oke, A.O.; Lawa, O. Diversity and nutritional status of edible insects in Nigeria: A review. *Int. J. Biodivers. Conser.* **2013**, *5*, 215–222. [[CrossRef](#)]
25. Nonaka, K. Feasting on insects. *Entomol. Res.* **2009**, *39*, 304–312. [[CrossRef](#)]



26. Shockley, M.; Dossey, A.T. Insects for Human Consumption. In *Mass Production of Beneficial Organisms: Invertebrates and Entomopathogens*; Academic Press: Cambridge, MA, USA, 2014; pp. 617–652. [[CrossRef](#)]
27. Nakimbugwe, D.; Ssepuuya, G.; Male, D.; Lutwama, V.; Mukisa, I.M.; Fiaboe, K.K.M. Status of the regulatory environment for utilization of insects as food and feed in Sub-Saharan Africa—a review. *Crit. Rev. Food Sci. Nutr.* **2020**, 1–10. [[CrossRef](#)]
28. Teffo, L.S.; Toms, R.B.; Eloff, J.N. Preliminary data on the nutritional composition of the edible stink-bug, *Encosternum delegorguei* Spinola, consumed in Limpopo province, South Africa. *S. Afr. J. Sci.* **2007**, *103*, 434–436.
29. Siulapwa, N.; Mwambungu, A.; Lungu, E.; Sichilima, W. Nutritional Value of Four Common Edible Insects in Zambia. *Int. J. Sci. Res.* **2012**, *3*, 2319–7064.
30. Adepoju, O.T.; Daboh, O.O. Nutrient composition of *Cirina forda* (westwood)-Enriched complementary foods. *Ann. Nutr. Metab.* **2013**, *63*, 139–144. [[CrossRef](#)]
31. Badanaro, F.; Amevoin, K.; Lamboni, C.; Amouzou, K. Edible *Cirina forda* (Westwood, 1849) (Lepidoptera: Saturniidae) caterpillar among Moba people of the savannah region in North Togo: From collector to consumer. *Asian J. Appl. Sci. Eng.* **2014**, *3*, 13. [[CrossRef](#)]
32. Adepoju, O.T.; Ajayi, K. Nutrient composition and adequacy of two locally formulated winged termite (*Macrotermes bellicosus*) enriched complementary Foods. *J. Food Res.* **2016**, *5*, 79. [[CrossRef](#)]
33. Illgner, P.; Nel, E. The geography of edible insects in Sub-Saharan Africa: A study of the mopane caterpillar. *Geogr. J.* **2000**, *166*, 336–351. [[CrossRef](#)]
34. Mpuchane, S.; Gashe, B.A.; Allotey, J.; Siame, B.; Teferra, G.; Dithlogo, M. Quality deterioration of phane, the edible caterpillar of an emperor moth *Imbrasia belina*. *Food Control* **2000**, *11*, 453–458. [[CrossRef](#)]
35. Mbah, C.; Elekima, G.O. Nutrient composition of some terrestrial insects in Ahmadu Bello University, Samaru Zaria Nigeria. *Sci. World J.* **2010**, *2*, 17–20. [[CrossRef](#)]
36. Banjo, A.D.; Lawal, O.A.; Songonuga, E.A. The nutritional value of fourteen species of edible insects in southwestern Nigeria. *Afr. J. Biotechnol.* **2006**, *5*, 298–301. [[CrossRef](#)]
37. Christensen, D.L.; Orech, F.O.; Mungai, M.N.; Larsen, T.; Friis, H.; Aagaard-Hansen, J. Entomophagy among the Luo of Kenya: A potential mineral source? *Int. J. Food Sci. Nutr.* **2006**, *57*, 198–203. [[CrossRef](#)] [[PubMed](#)]
38. Akullo, J.; Agea, J.G.; Obaa, B.B.; Okwee-Acai, J.; Nakimbugwe, D. Nutrient composition of commonly consumed edible insects in the Lango sub-region of northern Uganda. *Int. Food Res. J.* **2018**, *25*, 159–166.
39. Edijala, J.K.; Egbogbo, O.; Anigboro, A.A. Proximate composition and cholesterol concentrations of *Rhynchophorus phoenicis* and *Oryctes monoceros* larvae subjected to different heat treatments. *Afr. J. Biotechnol.* **2009**, *8*, 2346–2348. [[CrossRef](#)]
40. Assielou, B.; Due, E.; Koffi, M.; Dabonne, S.; Kouame, P. *Oryctes owariensis* Larvae as good alternative protein source: Nutritional and functional properties. *Annu. Res. Rev. Biol.* **2015**, *8*, 1–9. [[CrossRef](#)]
41. Manditsera, F.A.; Lakemond, C.M.M.; Fogliano, V.; Zvidzai, C.J.; Luning, P.A. Consumption patterns of edible insects in rural and urban areas of Zimbabwe: Taste, nutritional value and availability are key elements for keeping the insect eating habit. *Food Secur.* **2018**, *10*, 561–570. [[CrossRef](#)]
42. Jonathan, A.A. Proximate and anti-nutritional composition of two common edible insects: Yam beetle (*Heteroligus meles*) and palm weevil (*Rhynchophorus phoenicis*). *Elixir Food Sci.* **2012**, *49*, 9782–9786.
43. Thomas, C.N. Nutritional Potentials of Edible Larvae of Longhorned Beetle (*Apomecyna parumpunctata* Chev) (Coleoptera: Cerambycidae) in Niger Delta, Nigeria. *Int. J. Agric. Earth Sci.* **2018**, *4*, 46–51.
44. Ayieko, M.A.; Kinyuru, J.N.; Ndong'a, M.F.; Kenji, G.M. Nutritional value and consumption of black ants (*Carebara vidua* Smith) from the lake Victoria region in Kenya. *Adv. J. Food Sci. Technol.* **2012**, *4*, 39–45.
45. Lautenschläger, T.; Neinhuis, C.; Kikongo, E.; Henle, T.; Förster, A. Impact of different preparations on the nutritional value of the edible caterpillar *Imbrasia epimethea* from northern Angola. *Eur. Food Res. Technol.* **2017**, *243*, 1–10. [[CrossRef](#)]
46. Anvo, P.M.; Toguyéni, A.; Otchoumou, A.K.; Zoungrana-Kaboré, C.Y.; Koumelan, E.P. Nutritional qualities of edible caterpillars *Cirina butyrospermi* in southwestern of Burkina Faso. *Int. J. Innov. Appl. Stud.* **2016**, *18*, 639–645.
47. Musundire, R.; Zvidzai, C.J.; Chidewe, C.; Samende, B.K.; Chemura, A. Habitats and nutritional composition of selected edible insects in Zimbabwe. *J. Insects Food Feed* **2016**, *2*, 189–198. [[CrossRef](#)]
48. Akinnawo, O.; Ketiku, A.O. Chemical Composition and Fatty Acid Profile of Edible Larva of *Cirina forda* (Westwood). *Afr. J. Biomed. Res.* **2000**, *3*, 93–96.



49. Egan, B.A.; Toms, R.; Minter, L.R.; Addo-Bediako, A.; Masoko, P.; Mphosi, M.; Olivier, P.A.S. Nutritional Significance of the Edible Insect, *Hemijana variegata* Rothschild (Lepidoptera: Eupterotidae), of the Blouberg Region, Limpopo, South Africa. *Afr. Entomol.* **2014**, *22*, 15–23. [[CrossRef](#)]
50. Musundire, R.; Zvidzai, C.J.; Chidewe, C.; Samende, B.K.; Manditsera, F.A. Nutrient and anti-nutrient composition of *Henicus whellani* (Orthoptera: Stenopelmatidae), an edible ground cricket, in south-eastern Zimbabwe. *Int. J. Trop. Insect Sci.* **2014**, *34*, 223–231. [[CrossRef](#)]
51. Manditsera, F.A.; Luning, P.A.; Fogliano, V.; Lakemond, C.M.M. The contribution of wild harvested edible insects (*Eulepida mashona* and *Henicus whellani*) to nutrition security in Zimbabwe. *J. Food Compos. Anal.* **2019**, *75*, 17–25. [[CrossRef](#)]
52. El Hassan, N.M.; Hamed, S.Y.; Hassan, A.B.; Eltayeb, M.M.; Babiker, E.E. Nutritional evaluation and physiochemical properties of boiled and fried tree locust. *Pak. J. Nutr.* **2008**, *7*, 325–329. [[CrossRef](#)]
53. Adeyeye, E.I.; Awokunmi, E.E. Chemical composition of female and male giant African crickets, *Brachytrypes membranaceus* L. *Int. J. Pharma Bio Sci.* **2010**, *1*, 125–136.
54. Rumpold, B.A.; Schlüter, O. Insect-based protein sources and their potential for human consumption: Nutritional composition and processing. *Anim. Front.* **2015**, *5*, 20–24. [[CrossRef](#)]
55. Rahman, A.; Bordoloi, S.; Mazid, S. Entomophagy practiced among the Tiwa community of Morigaon district. *J. Entomol. Zool. Stud.* **2018**, *6*, 484–486.
56. Agbidye, F.; Nongo, N. Harvesting and processing techniques for the larvae of the pallid emperor moth. *J. Res. For. Wildl. Enviro.* **2009**, *1*, 123–132.
57. Muafor, F.J.; Levang, P.; Le Gall, P. A crispy delicacy: Augosoma beetle as alternative source of protein in East Cameroon. *Int. J. Biodivers.* **2014**, *2014*, 1–7. [[CrossRef](#)]
58. Dube, S.; Dlamini, N.R.; Mafunga, A.; Mukai, M.; Dhlamini, Z. A survey on entomophagy prevalence in Zimbabwe. *Afr. J. Food Agric. Nutr. Dev.* **2013**, *13*, 7242–7253. [[CrossRef](#)]
59. Kinyuru, J.N.; Konyole, S.O.; Roos, N.; Onyango, C.A.; Owino, V.O.; Owuor, B.O.; Estambale, B.B.; Friis, H.; Aagaard-Hansen, J.; Kenji, G.M. Nutrient composition of four species of winged termites consumed in western kenya. *J. Food Compos. Anal.* **2013**, *30*, 120–124. [[CrossRef](#)]
60. Rutaro, K.; Malinga, G.M.; Lehtovaara, V.J.; Opoke, R.; Valtonen, A.; Kwetegyeka, J.; Nyeko, P.; Roininen, H. The fatty acid composition of edible grasshopper *Ruspolia differens* (Serville) (Orthoptera: Tettigoniidae) feeding on diversifying diets of host plants. *Entomol. Res.* **2018**, *48*, 490–498. [[CrossRef](#)]
61. Malinga, G.M.; Valtonen, A.; Lehtovaara, V.J.; Rutaro, K.; Nyeko, R.O.P.; Roininen, H. Diet acceptance and preference of the edible grasshopper *Ruspolia differens* (Orthoptera; Tettigoniidae). *Appl. Entomol. Zool.* **2018**, *53*, 229–239. [[CrossRef](#)]
62. Blásquez, J.R.E.; Moreno, J.M.P.; Camacho, V.H.M. Could grasshoppers be a nutritive meal? *Food Nutr. Sci.* **2012**, *3*, 164–175. [[CrossRef](#)]
63. Henley, E.C.; Taylor, J.R.N.; Obukosia, S.D. The importance of dietary protein in human health: Combating protein deficiency in sub-Saharan Africa through transgenic biofortified sorghum. *Adv. Nutr Food Sci.* **2010**, *60*, 21–52.
64. De Onis, M.; Branca, F. Childhood stunting: A global perspective. *Matern. Child Nutr.* **2016**, *12*, 12–26. [[CrossRef](#)] [[PubMed](#)]
65. Bhattacharyya, B.; Choudhury, B.; Das, P.; Dutta, S.K.; Bhagawati, S.; Pathak, K. Nutritional composition of five soil-dwelling Scarab beetles (Coleoptera: Scarabaeidae) of Assam, India. *Coleopt. Bull.* **2018**, *72*, 339. [[CrossRef](#)]
66. Nyangena, D.N.; Mutungi, C.; Imathiu, S.; Kinyuru, J.; Affognon, H.; Ekesi, S.; Nakimbugwe, D.; Fiaboe, K.K.M. Effects of traditional processing techniques on the nutritional and microbiological quality of four edible insect species used for food and feed in East Africa. *Foods* **2020**, *9*, 574. [[CrossRef](#)]
67. Bukkens, S.G.F. The nutritional value of edible insects. *Ecol. Food Nutr.* **1997**, *36*, 287–319. [[CrossRef](#)]
68. Kewuyemi, Y.O.; Kesa, H.; Chinma, C.E.; Adebo, O.A. Fermented edible insects for promoting food security in Africa. *Insects* **2020**, *11*, 283. [[CrossRef](#)]
69. Akhtar, Y.; Isman, M.B. Insects as an alternative protein Source. *Food Sci. Nutr.* **2018**, *2*, 263–288. [[CrossRef](#)]
70. Adámková, A.; Mlček, J.; Kouřimská, L.; Borkovcová, M.; Bušina, T.; Adámek, M.; Bednářová, M.; Krajsa, J. Nutritional potential of selected insect species reared on the island of Sumatra. *Int. J. Environ. Res. Public Health* **2017**, *14*, 521. [[CrossRef](#)]

71. Mwangi, M.N.; Oonincx, D.G.A.B.; Stouten, T.; Veenenbos, M.; Melse-Boonstra, A.; Dicke, M.; Van Loon, J.J.A. Insects as sources of iron and zinc in human nutrition. *Nutr. Res. Rev.* **2018**, *31*, 248–255. [CrossRef]
72. Igwe, C.; Ujowundu, C.; Nwaogu, L. chemical analysis of an edible African termite, *Macrotermes nigeriensis*; a potential antidote to food security problem. *Biochem. Anal. Biochem.* **2012**, *1*, 1–4. [CrossRef]
73. Arigony, A.L.V.; De Oliveira, I.M.; Machado, M.; Bordin, D.L.; Bergter, L.; Pra, D.; Henriques, J.A.P. The influence of micronutrients in cell culture: A reflection on viability and genomic stability. *Biomed Res. Int.* **2013**, *2013*, 1–22. [CrossRef] [PubMed]
74. FAO. The State of Food Security and Nutrition in the World. 2019. Available online: <http://www.fao.org/3/ca5162en/ca5162en.pdf> (accessed on 21 May 2019).
75. Netshifhefhe, S.R.; Kunjeku, E.C.; Duncan, F.D. Human uses and indigenous knowledge of edible termites in Vhembe District, Limpopo Province, South Africa. *S. Afr. J. Sci.* **2018**, *114*, 1–10. [CrossRef]
76. Baiyegunhi, L.J.S.; Oppong, B.B.; Senyolo, M.G. Socio-economic factors influencing mopane worm (*Imbrasia belina*) harvesting in Limpopo Province, South Africa. *J. For. Res.* **2016**, *27*, 443–452. [CrossRef]
77. Adeoye, O.T.; Alebiosu, B.I.; Akinyemi, O.D.; Adeniran, O.A. Socio economic analysis of forest edible insects species consumed and its role in the livelihood of people in Lagos State. *J. Food Stud.* **2014**, *3*, 104. [CrossRef]
78. Obopile, M.; Seeletso, T.G. Eat or not eat: An analysis of the status of entomophagy in Botswana. *Food Secur.* **2013**, *5*, 817–824. [CrossRef]
79. Ebenebe, C.I.; Amobi, M.I.; Udegbala, C.; Ufele, A.N.; Nweze, B.O. Survey of edible insect consumption in south-eastern Nigeria. *J. Insects Food Feed* **2017**, *3*, 241–252. [CrossRef]
80. Paiko, Y.B.; Dauda, B.E.N.; Salau, R.B.; Jacob, J.O. Preliminary data on the nutritional potential of the larva of edible dung beetle consumed in Paikoro local government area of Niger state, Nigeria. *Cont. J. Food Sci. Technol.* **2012**, *6*, 3–42.
81. Onyeike, E.N.; Ayalogu, E.O.; Okaraonye, C.C. Nutritive value of the larvae of raphia palm beetle (*Oryctes rhinoceros*) and weevil (*Rhynchophorus phoenicis*). *J. Sci. Food Agric.* **2005**, *85*, 1822–1828. [CrossRef]
82. Ekpo, K.; Onigbinde, A.O. Nutritional potentials of the larva of *Rhynchophorus phoenicis*. (F). *Pak. J. Nutri.* **2005**, *4*, 287–289.
83. Omotoso, O.T.; Adedire, C.O. Nutrient composition, mineral content and the solubility of the proteins of palm weevil, *Rhynchophorus phoenicis* f. (Coleoptera: Curculionidae). *J. Zhejiang Univ. Sci. B* **2007**, *8*, 318–322. [CrossRef]
84. Ekpo, K.E.; Onigbinde, A.O.; Asia, I.O. Pharmaceutical potentials of the oils of some popular insects consumed in southern Nigeria. *Afr. J. Pharm. Pharmacol.* **2009**, *3*, 51–57.
85. Womeni, H.M.; Linder, M.; Tiencheu, B.; Mbiapo, F.T.; Villeneuve, P.; Fanni, J.; Parmentier, M. Oils of insects and larvae consumed in Africa: Potential sources of polyunsaturated fatty acids. *OCL Ol. Corps Gras Lipides* **2009**, *16*, 230–235. [CrossRef]
86. Idolo, I. Nutritional and quality attributes of wheat buns enriched with the larvae of *Rhynchophorus phoenicis* f. *Pak. J. Nutri.* **2010**, *9*, 1043–1046. [CrossRef]
87. Braide, W.; Nwaoguikpe, R. Assessment of microbiological quality and nutritional values of a processed edible weevil caterpillar (*Rhynchophorus phoenicis*) in Port Harcourt, southern Nigeria. *Int. J. Biol. Chem. Sci.* **2011**, *5*, 410–418. [CrossRef]
88. Ogbuagu, M.N.; Ohondu, I.; Nwigwe, C. Fatty Acid and amino acid profiles of the larva of raffia palm weevil: *Rhynchophorus phoenicis*. *Pac. J. Sci. Technol.* **2011**, *12*, 392–400.
89. Elemo, B.O.; Elemo, G.N.; Makinde, M.; Erukainure, O.L. Chemical evaluation of African palm weevil, *Rhynchophorus phoenicis*, larvae as a food source. *J. Insect Sci.* **2011**, *11*, 1–6. [CrossRef]
90. Womeni, H.M.; Tiencheu, B.; Linder, M.; Nabayo, E.M.C.; Tenyang, N.; Mbiapo, T.F.; Parmentier, M. Nutritional value and effect of cooking, drying and storage process on some functional properties of *Rhynchophorus phoenicis*. *Int. J. Life Sci. Pharma Res.* **2012**, *2*, 203–219.
91. Okore, O.; Awaaja, D.; Nwana, I. Edible insects of the Niger Delta area in Nigeria. *J. Nat. Sci. Res.* **2014**, *4*, 1–9.
92. Muafor, F.J.; Gnetegha, A.A.K.; Levang, P. *Final Project Report Promoting the Farming of Edible Larvae of Palm Weevils (Rhynchophorus Spp.) as an Alternative to Sustain Rural Livelihoods and Community Based Conservation in Cameroon Humid Forest Regions*; CIFOR LIFT IRD: Yaounde, Cameroon, 2016.
93. Meutchieye, F.; Tsafo, K.E.C.; Niassy, S. Inventory of edible insects and their harvesting methods in the Cameroon centre region. *J. Insects Food Feed* **2016**, *2*, 145–152. [CrossRef]

94. Anankware, J.P.; Osekre, E.A.; Obeng-Ofori, D.; Khamala, C. Identification and classification of common edible insects in Ghana. *Int. J. Entomol. Res.* **2016**, *1*, 2455–4758.
95. Laar, A.; Kotoh, A.; Parker, M.; Milani, P.; Tawiah, C.; Soor, S.; Anankware, J.P.; Kalra, N.; Manu, G.; Tandoh, A.; et al. An exploration of edible palm weevil larvae (Akoko) as a source of nutrition and livelihood: Perspectives from Ghanaian stakeholders. *Food Nutr. Bull.* **2017**, *38*, 455–467. [[CrossRef](#)] [[PubMed](#)]
96. Lautenschläger, T.; Neinhuis, C.; Monizi, M.; Mandombe, J.L.; Förster, A.; Henle, T.; Nuss, M. Edible insects of Northern Angola. *Afr. Invertebr.* **2017**, *58*, 55–82. [[CrossRef](#)]
97. Quaye, B.; Charity, C.; Donkoh, A. Nutritional potential and microbial status of African palm weevil nutritional potential and microbial status of African palm peevil (*Rhynchophorus phoenicis*) larvae raised on alternative feed resources. *ARJETS* **2018**, *48*, 45–52.
98. Séré, A.; Bougma, A.; Ouilly, J.T.; Traoré, M.; Sangaré, H.; Lykke, A.M.; Ouédraogo, A.; Gnankiné, O.; Bassolé, I.H.N. Traditional knowledge regarding edible insects in Burkina Faso. *J. Ethnobiol. Ethnomed.* **2018**, *14*, 1–12. [[CrossRef](#)]
99. Agbidye, F.; Ofuya, T.; Akindela, S. Some edible insect species consumed by the people of Benue State, Nigeria. *Pak. J. Nutr.* **2009**, *8*, 946–950. [[CrossRef](#)]
100. Tobih, F. Occurrence and seasonal variation of *Heteroligus meles* billb (Coleoptera: Dynastidae) in upper Niger Delta, Nigeria. *Agric. Biol. J. North Am.* **2011**, *2*, 826–831. [[CrossRef](#)]
101. Okaraonye, C.C.; Ikewuchi, J.C. Nutritional potential of *Oryctes rhinoceros* larva. *Pak. J. Nutr.* **2009**, *8*, 35–38. [[CrossRef](#)]
102. Ehounou, G.P.; Ouali-n'gorans, W.M.; Niassys, W.M. Assessment of entomophagy in Abidjan (Cote Divoire, West Africa). *Afr. J. Food Sci.* **2018**, *12*, 6–14. [[CrossRef](#)]
103. Anankware, J.P.; Osekre, E.A.; Fening, K.; Obeng-Ofori, D. Insects as food and feed. *Food Sci. Technol.* **2015**, *32*, 22–25. [[CrossRef](#)]
104. Aydoğan, Z.; Gürol, A.; İncekara, Ü.; Tahidu, O.D. Element content analysis of edible insect of Ghana (Curculionidae: *Sitophilus zeamais*) Using EDXRF Spectrometer. *Erzincan Üniversitesi Fen Bilim. Enstitüsü Derg.* **2016**, *9*, 86–94. [[CrossRef](#)]
105. Dzerefos, C.M.; Witkowski, E.T.F.; Toms, R. Life-history traits of the edible stinkbug, *Encosternum delegorguei* (Hem. Tessaratomidae), a traditional food in southern Africa. *J. Appl. Entomol.* **2009**, *133*, 749–759. [[CrossRef](#)]
106. Mariod, A.A.; Abdel-Wahab, S.I.; Ain, N.M. Proximate amino acid, fatty acid and mineral composition of two Sudanese edible pentatomid insects. *Int. J. Trop. Insect Sci.* **2011**, *31*, 145–153. [[CrossRef](#)]
107. Tamesse, J.L.; Kekeunou, S.; Tchatchouang, L.J.; Ndegue, O.L.M.; Aissatou, L.M.; Tombouck, D.; Youssa, B. Insects as food, traditional medicine and cultural rites in the west and south regions of Cameroon. *J. Insects Food Feed* **2016**, *2*, 153–160. [[CrossRef](#)]
108. Okia, C.A.; Odongo, W.; Nzabamwita, P.; Ndimubandi, J.; Nalika, N.; Nyeko, P. Local knowledge and practices on use and management of edible insects in Lake Victoria basin, East Africa. *J. Insects Food Feed* **2017**, *3*, 83–93. [[CrossRef](#)]
109. Ekpo, K.; Onigbinde, A.O. Characterization of lipids in winged reproductive of the termite *Macrotermes bellicosus*. *Pak. J. Nutr.* **2007**, *6*, 247–251.
110. Akullo, J.; Obaa, B.B.; Acai, J.O.; Nakimbugwe, D.; Agea, J.G. Knowledge, attitudes and practices on edible insects in Lango sub-region, northern Uganda. *J. Insects Food Feed* **2017**, *3*, 73–81. [[CrossRef](#)]
111. Ntukuyoh, A.I.; Udiong, D.S.; Ikpe, A.E.; Akpakpan, A.E. Evaluation of nutritional value of termites (*Macrotermes bellicosus*): Soldiers, workers, and queen in the Niger Delta Region of Nigeria. *Int. J. Food Nutr. Saf.* **2012**, *1*, 60–65.
112. Inje, O.F.; Olufunmilayo, A.H.; Audu, J.A.; Ndaman, S.A.; Chidi, E.E. Food science and human wellness protein quality of four indigenous edible insect species in Nigeria. *Food Sci. Hum. Wellness* **2018**, *7*, 175–183. [[CrossRef](#)]
113. Adepoju, O.T.; Omotayo, O.A. Nutrient composition and potential contribution of winged termites (*Macrotermes bellicosus* Smeathman) to micronutrient intake of consumers in Nigeria. *Curr. J. Appl. Sci. Technol.* **2014**, *4*, 1149–1158. [[CrossRef](#)]
114. Adepoju, O.T. Assessment of fatty acid profile, protein and macronutrient bioavailability of winged termites (*Macrotermes bellicosus*) using albino rats. *Malays. J. Nutr.* **2016**, *22*, 153–161.

115. Odongo, W.; Okia, C.A.; Nalika, N.; Nzabamwita, P.H.; Ndimubandi, J.; Nyeko, P. Marketing of edible insects in Lake Victoria basin: The case of Uganda and Burundi. *J. Insects Food Feed* **2018**, *4*, 285–293. [[CrossRef](#)]
116. Ajai, A.; Bankole, M.; Jacob, J.O.; Audu, U.A. Determination of some essential minerals in selected edible insects. *Afr. J. Pure Appl. Chem.* **2013**, *7*, 194–197. [[CrossRef](#)]
117. Bomolo, O.; Niassy, S.; Chocha, A.; Longanza, B.; Bugeme, D.M.; Ekesi, S.; Tanga, C.M. Ecological diversity of edible insects and their potential contribution to household food security in Haut-Katanga Province, Democratic Republic of Congo. *Afr. J. Ecol.* **2017**, *55*, 640–653. [[CrossRef](#)]
118. Rémy, D.A.; Hervé, B.B.; Sylvain, O.N. Study of some biological parameters of *Cirina butyrospermi* Vuillet (Lepidoptera, Attacidae), an edible insect and shea caterpillar (*Butyrospermum paradoxum* Gaertn. F.) in a context of climate change in Burkina Faso. *Adv. Entomol.* **2017**, *6*, 1–8. [[CrossRef](#)]
119. Solomon, M.; Prisca, N. Nutritive value of *Lepidoptara litoralia* (edible caterpillar) found in Jos Nigeria: Implications and food security and poverty alleviation. *AJFAND* **2012**, *12*, 6737–6747.
120. Ande, A.T.; Sciences, B. The lipid profile of the pallid emperor moth *Cirina forda* Westwood (Lepidoptera: Saturniidae) caterpillar. *Biokemistri* **2003**, *13*, 37–41.
121. Oyegoke, O.O.; Akintola, A.J.; Fasoranti, J.O. Dietary potentials of the edible larvae of *Cirina forda* (westwood) as a poultry feed. *Afr. J. Biotechnol.* **2006**, *5*, 1799–1802. [[CrossRef](#)]
122. Omotoso, O.T. Nutritional quality, functional properties and anti-nutrient compositions of the larva of *Cirina forda* (Westwood) (Lepidoptera: Saturniidae). *J. Zhejiang Univ. Sci. B* **2006**, *7*, 51–55. [[CrossRef](#)]
123. Osasona, A.I.; Olaofe, O. Nutritional and functional properties of *Cirina forda* larva from Ado-Ekiti, Nigeria. *Afr. J. Food Sci.* **2010**, *4*, 775–777.
124. Morris, B. Insects as food among hunter-gatherers. *Anthropol. Today* **2008**, *24*, 6–8. [[CrossRef](#)]
125. Bama, H.B.; Dabire, R.A.; Ouattara, D.; Niassy, S.; Ba, M.N.; Dakouo, D. Diapause disruption in *Cirina butyrospermi* Vuillet (Lepidoptera, Attacidae), the shea caterpillar, in Burkina Faso. *J. Insects Food Feed* **2018**, *4*, 239–245. [[CrossRef](#)]
126. Madibela, O.R.; Seitiso, T.K.; Thema, T.F.; Letso, M. Effect of traditional processing methods on chemical composition and in vitro true dry matter digestibility of the mopane worm (*Imbrasia belina*). *J. Arid Environ.* **2007**, *68*, 492–500. [[CrossRef](#)]
127. Ghally, A.E. The use of insects as human food in Zambia. *Online J. Biol. Sci.* **2009**, *9*, 93–104. [[CrossRef](#)]
128. Thomas, B. Sustainable harvesting and trading of mopane worms (*Imbrasia belina*) in Northern Namibia: An experience from the Uukwaluudhi area. *Int. J. Environ. Stud.* **2013**, *70*, 494–502. [[CrossRef](#)]
129. Pharithi, M.T.; Suping, S.M.; Yeboah, S.O. Variations of the fatty acid composition in the oil from the larval stages of the emperor moth caterpillar, *Imbrasia belina*. *Bull. Chem. Soc. Ethiop.* **2004**, *18*, 67–72. [[CrossRef](#)]
130. Mbata, K.J.; Chidumayo, E.N.; Lwatula, C.M. Traditional regulation of edible caterpillar exploitation in the Kopa area of Mpika district in northern Zambia. *J. Insect Conserv.* **2002**, *6*, 115–130. [[CrossRef](#)]
131. Adeyeye, E.I. Amino acid composition of variegated grasshopper, *Zonocerus variegatus*. *Trop. Sci.* **2005**, *45*, 141–143. [[CrossRef](#)]
132. Mmari, M.W.; Kinyuru, J.N.; Laswai, H.S.; Okoth, J.K. Traditions, beliefs and indigenous technologies in connection with the edible longhorn grasshopper *Ruspolia differens* (Serville 1838) in Tanzania. *J. Ethnobiol. Ethnomed.* **2017**, *13*, 1–11. [[CrossRef](#)]
133. Ssepunya, G.; Mukisa, I.M.; Nakimbugwe, D. Nutritional composition, quality, and shelf stability of processed *Ruspolia nitidula* (edible grasshoppers). *Food Sci. Nutr.* **2017**, *5*, 103–112. [[CrossRef](#)]
134. Mohamed, E.H.A. Determination of nutritive value of the edible migratory locust *Locusta migratoria*, Linnaeus, 1758 (Orthoptera: Acrididae). *Int. J. Adv. Pharm. Biol. Chem.* **2015**, *4*, 144–148.
135. Mohamed, E.H.A. Fatty acids contents of the edible migratory locust *Locusta migratoria*, Linnaeus, 1758 (Orthoptera: Acrididae). *Int. J. Adv. Pharm. Biol. Chem.* **2015**, *4*, 746–750.

