



Correlation of liver-to-spleen ratio, lung CT scores, clinical, and laboratory findings of COVID-19 patients with two consecutive CT scans

Ezgi Guler¹ · Nalan Gulsen Unal² · Akin Cinkooglu¹ · Recep Savas¹ · Timur Kose³ · Husnu Pullukcu⁴ · Mustafa Harman¹ · Nevra Z. Elmas¹ · Nikhil H. Ramaiya⁵ · Ahmet Omer Ozutemiz²

Received: 5 July 2020 / Revised: 19 September 2020 / Accepted: 30 September 2020 / Published online: 13 October 2020
© Springer Science+Business Media, LLC, part of Springer Nature 2020

Abstract

Purpose Given the lack of information about abdominal imaging findings and correlation with clinical features of COVID-19, we aimed to evaluate the changes in hepatic attenuation during the course of disease. Our aim was to correlate the liver-to-spleen ratio (L/S), clinical, laboratory findings, and lung CT scores of patients with COVID-19 who had two consecutive chest CTs.

Methods A retrospective search was performed between March 1, 2020 and April 26, 2020 to identify patients who had positive RT-PCR tests and two unenhanced chest CTs. Scans that were obtained at hospital admission and follow-up were reviewed to assess L/S and lung CT scores. Patients were divided into two groups based on lung CT scores (non-progressive vs progressive). Patient demographics, laboratory findings, length of hospital stay, and survival were noted from electronic medical records.

Results Twenty patients in the progressive group and 7 patients in the non-progressive group were identified. The mean L/S of the progressive group (1.13 ± 0.3) was lower than that of the non-progressive group (1.21 ± 0.29) at hospital admission but there was no significant difference between the two groups ($p = 0.547$). L/S at follow-up was significantly different between the groups as the mean L/S values of the progressive and non-progressive groups were 1.02 ± 0.23 and 1.25 ± 0.29 , respectively ($p = 0.009$). L/S was negatively correlated with AST and ALT ($r = -0.46$, $p = 0.016$ and $r = -0.534$, $p = 0.004$, respectively). There were significant differences between the two groups in terms of WBC, neutrophil, lymphocyte, monocyte, and platelet counts that were obtained at hospital admission. Length of hospital stay was significantly longer in patients in the progressive group ($p = 0.035$).

Conclusions Decrease in L/S may be observed in patients with elevated lung CT scores at follow-up. WBC, neutrophil, lymphocyte, monocyte, and platelet counts at hospital admission may predict the progression of COVID-19.

Keywords Coronavirus · Multidetector computed tomography · Laboratory · Thorax · Liver

✉ Ezgi Guler
gulerezgi@yahoo.com

¹ Department of Radiology, Faculty of Medicine, Ege University, Bornova, Izmir 35100, Turkey

² Division of Gastroenterology, Department of Internal Medicine, Faculty of Medicine, Ege University, Izmir, Turkey

³ Department of Biostatistics, Faculty of Medicine, Ege University, Izmir, Turkey

⁴ Department of Infectious Diseases and Clinical Microbiology, Faculty of Medicine, Ege University, Izmir, Turkey

⁵ Department of Radiology, University Hospitals Cleveland Medical Center, Cleveland, USA

Introduction

Coronavirus disease 2019 (COVID-19), which is caused by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has been a threat to global public health [1]. Reverse transcription-polymerase chain reaction (RT-PCR) test is the gold standard for the molecular diagnosis of the disease [2]. Although it is a highly specific test, in clinical use it shows a variable sensitivity, which arises the need of other tools to support the diagnosis. Since chest computed tomography (CT) is widely available and can contribute to fast pneumonia diagnosis, it plays an important role in the diagnosis and management of COVID-19 patients [2–5].

COVID-19 demonstrates a wide clinical spectrum ranging from asymptomatic infection to severe pneumonia with acute respiratory distress syndrome, multiple organ failure, and death, with many patients being hospitalized [6–8]. Older age, comorbidities including hypertension, diabetes, cardiovascular disease, respiratory disease, and laboratory examinations such as white blood cells (WBC), aspartate aminotransferase (AST), creatinine, lactate dehydrogenase (LDH), and d-dimer have been reported to imply the progression of COVID-19 [9, 10]. Chest CT findings have been shown to be associated with severe or critical COVID-19 pneumonia as well as with the different stages of the disease [11, 12]. Although the involvement of the respiratory system is common, liver injury may occur especially in patients with severe COVID-19 [13].

Changes of hepatic density on unenhanced CT scans can occur in diffuse liver diseases, whether as a decrease in steatosis or an increase in hemochromatosis [14, 15]. Other entities that may alter hepatic density on CT include drug-induced toxicity, acute hepatitis, acute toxic hepatic injury, cirrhosis, and radiation-induced injury [15]. It has been reported that unenhanced CT scans demonstrate diffuse hypoattenuating areas in the liver of patients with viral acute liver failure [16]. Liver-to-spleen ratio (L/S) on unenhanced CT has been used to discriminate hepatic steatosis [17]. Thus, L/S may be helpful in the evaluation of liver injury as it reflects the changes of hepatic attenuation. Given the lack of information in the literature for the assessment of abdominal imaging findings in COVID-19 patients, which may be related to the limited evaluation of the upper abdomen on unenhanced chest CT scans, we aimed to investigate the changes in L/S during the course of disease. The purpose of this study is to correlate the L/S, clinical, laboratory findings, and lung CT scores of COVID-19 patients who had two consecutive chest CT scans.

Materials and methods

Patients

This retrospective study was approved by our institutional review board and the requirement for informed consent was waived. A database search between March 1, 2020 and April 26, 2020 was performed to identify patients with COVID-19 and who had chest CT scans. Inclusion criteria were as follows,

1. adult patients (age \geq 18 years),
2. patients who had positive RT-PCR tests for COVID-19,
3. patients who had two consecutive unenhanced chest CTs (obtained at hospital admission and follow-up).

Exclusion criteria were as follows,

1. patients who had positive RT-PCR tests but did not have chest CTs,
2. patients who had only one chest CT scan, and
3. patients who had only contrast-enhanced chest CTs.

Clinical and laboratory data

The electronic medical records were reviewed to note the laboratory examinations obtained within 1-day interval time of the corresponding dates of the first and second chest CT scans. Laboratory examinations including blood AST (normal limits, < 35 U/L), alanine aminotransferase (ALT) (normal limits, < 45 U/L), alkaline phosphatase (ALP) (35–104 U/L), gamma-glutamyltransferase (GGT) (normal limits, < 38 U/L), total bilirubin (0.1–1 mg/dL), creatinine (0.6–1.1 mg/dL), C-reactive protein (CRP) (0–5 mg/L), LDH (normal limits, < 225 IU/L), d-dimer (< 550 μ g/L FEU), WBC ($4.5\text{--}11 \times 10^3/\mu\text{L}$), neutrophil ($2.02\text{--}7.46 \times 10^3/\mu\text{L}$), lymphocyte ($1\text{--}3.38 \times 10^3/\mu\text{L}$), monocyte ($0\text{--}0.8 \times 10^3/\mu\text{L}$), hemoglobin (Hb) (11.7–16 g/dL), and platelet count ($150\text{--}450 \times 10^3/\mu\text{L}$) were recorded. Neutrophil/lymphocyte ratio was calculated. Comorbidities of the patients including diabetes, hypertension, chronic renal failure, and malignancy were noted. The treatment, length of hospital stay and survival of the patients were recorded. Empiric treatment of COVID-19 was given according to the guidelines of the National Ministry of Health. The agents including hydroxychloroquine, azithromycin, other antibiotics, oseltamivir, and favipiravir were used in the treatment according to the severity of pneumonia [18].

Imaging studies and analysis

All CT scans were performed using a 160-slice-CT scanner (Aquilion Prime, Canon Medical Systems, Tochigi, Japan). The axial images were acquired craniocaudally at shallow inspiration and included the body part extending from the thoracic inlet to upper abdomen (including the upper poles to midportion of the kidneys). High-resolution CT of the chest was obtained by using the following parameters; 120 kVp, 100–200 mA, 80×0.5 mm collimation and reconstructed at 0.5 mm slice thickness with a sharp reconstruction kernel. All of the patients underwent CT scans without contrast agent administration. CT images were reviewed in consensus by two thoracic radiologists (A.C and R.S). CT scores were assigned for the first scan that was obtained at hospital admission and follow-up CT scan of each patient to evaluate the extension of the lung involvement. A chest CT scoring system was used in which each lobe was given a score that was based on the following: score 0, no involvement; score 1, less than 5% involvement; score 2, 5–25%

involvement; score 3, 26–49% involvement; score 4, 50–75% involvement; and score 5, greater than 75% involvement of the lobe. According to this scoring system, the minimum and maximum values of lung CT scores were 0 and 25, respectively [19–21]. Patients were divided into two groups based on the lung CT scores as follows, non-progressive group: patients with decreased or stable CT scores on the follow-up CT scan and progressive group: patients with increased CT scores on the follow-up CT scan.

Two abdominal radiologists in consensus (E.G and N.Z.E) reviewed the upper abdominal parts that were included in the chest CT scans and calculated the L/S. In this study, L/S was obtained from two consecutive unenhanced chest CT scans of each COVID-19 patient to assess if a change occurred in hepatic density during the course of disease. Hepatic attenuation values were calculated by placing two regions of interest (ROI) greater than 100mm² in area, in the right liver lobe anteriorly and one ROI in the left liver lobe. Splenic attenuation was obtained by placing one ROI, greater than 100mm² in area (Fig. 1). L/S was calculated by taking the average Hounsfield unit (HU) measurement of both liver lobe ROIs and dividing it by the spleen HU value [22]. Attempts were made to place ROIs in the exact location as possible in both CT scans.

Statistical analysis

Statistical analyses were performed by using IBM SPSS Statistics for Windows, Version 25.0. (IBM Corp. Released 2017. Armonk, NY) and R software, version 3.5.2, package:

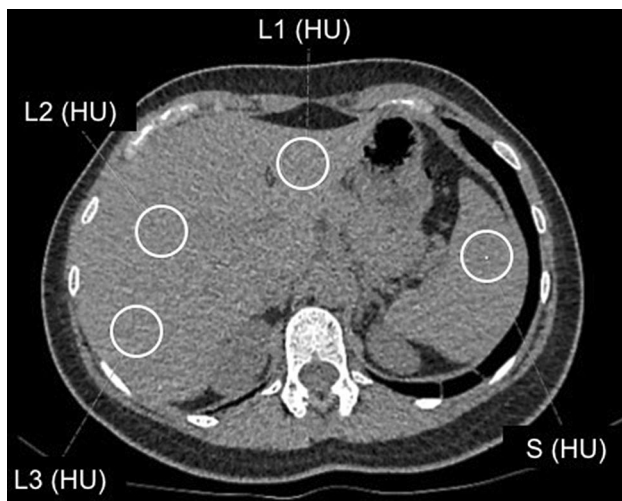


Fig. 1 29-year-old woman with COVID-19. Hepatic attenuation values were calculated by placing one ROI greater than 100mm² in area in the left liver lobe (L1), and two ROIs, in the right liver lobe (L2 and L3). Splenic attenuation was obtained by placing one ROI, greater than 100mm² in area (S). Liver-to-spleen ratio was calculated as $[(L1 + L2 + L3)/3]/S$

nparLD (R Foundation for Statistical Computing, Vienna, Austria; <https://r-project.org>).

Normally distributed continuous variables were displayed as mean \pm standard deviation, non-normally distributed continuous variables were displayed as median (minimum–maximum). Repeated measures ANOVA was used for normally distributed variables. When an interaction (interaction represents that the changes of parameters obtained at hospital admission and follow-up were different between the two groups) was found to be significant, paired *t*-test, two independent samples *t*-test, and analysis of covariance were performed. For the evaluation of non-normally distributed variables, the Brunner and Langer method (F1-LD-F1 model) was used. When an interaction was found to be significant, Mann–Whitney U test and Wilcoxon test were performed. Receiver Operating Characteristic (ROC) analysis was used for the assessment of the laboratory values obtained at the time of the first chest CT to predict the changes in the CT scores. Categorical variables were compared between the two groups of patients using the chi-square test and Fisher’s exact test. Spearman’s Rho analysis was used to evaluate the relationship between L/S and liver function tests.

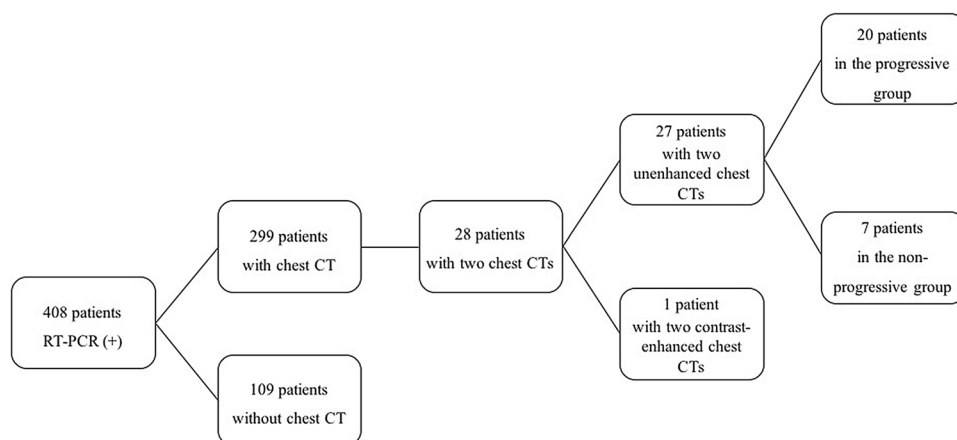
Results

Clinical and laboratory findings

Out of consecutive 408 patients who had positive RT-PCR tests, a total of 27 patients who met the inclusion criteria were identified. The flowchart of the study is shown in Fig. 2. There were 7 patients [4 men, 3 women; median age: 41 years (29–65 years)] in the non-progressive group and 20 patients [8 men, 12 women; median age: 57 years (23–81) years] in the progressive group. Table 1 demonstrates the clinical characteristics of the two groups of patients.

Three (42.8%) patients in the non-progressive group and 11 (55%) patients in the progressive group had comorbidities. Six (85.7%) patients in the non-progressive group and 18 (90%) patients in the progressive group received hydroxychloroquine therapy. Nine (45%) patients in the progressive group had favipiravir treatment while it was started in 1 (14.3%) patient in the non-progressive group. No significant differences of comorbidities and treatment were found between the two groups of patients ($p > 0.05$).

Patients in the progressive group had a significantly longer hospital stay than the patients in the non-progressive group ($p = 0.035$). Six (30%) patients who had increased lung CT scores were admitted to intensive care unit (ICU) whereas none of the patients with decreased or stable lung CT scores had ICU admission. One (5%) patient in progressive group died while none of the patients in the non-progressive group died.

Fig. 2 The flowchart of the study**Table 1** Patient demographics and clinical characteristics of the two groups of COVID-19 patients

Parameter	Non-progressive group (n = 7)	Progressive group (n = 20)	p
Age	41 (29–65)	57 (23–81)	0.653
Sex			0.662
Male	4 (57.1%)	8 (40%)	
Female	3 (42.8%)	12 (60%)	
Comorbidities	3 (42.8%)	11 (55%)	
Hypertension	1 (14.3%)	9 (45%)	0.204
Diabetes mellitus	3 (42.9%)	2 (10%)	0.091
Chronic renal failure	0 (0)	1 (5%)	1.000
Malignancy	1 (14.3%)	1 (5%)	0.459
Treatment			
Hydroxychloroquine	6 (85.7%)	18 (90%)	1.000
Azithromycin ± or other antibiotics	4 (57.1%)	11 (55%)	1.000
Oseltamivir	3 (42.9%)	7 (35%)	1.000
Favipiravir	1 (14.3%)	9 (45%)	0.204
LMWH	0 (0)	6 (30%)	0.155
Length of hospital stay (day)	5 (0–17)	12.5 (0–29)	0.035
ICU admission	0 (0)	6 (30%)	0.155
Survival			1.000
Dead	0 (0)	1 (5%)	
Alive	7 (100%)	19 (95%)	

Data are n (%) or median (minimum–maximum)

ICU intensive care unit, LMWH low-molecular-weight heparin

Laboratory parameters of the patients obtained at the time corresponding to the dates of the two chest CTs are detailed in Table 2. By using Brunner and Langer method, it was found that changes of parameters obtained at the time of hospital admission and follow-up were different between the two groups of patients for the following tests; AST, ALT, creatinine, and CRP. Δ values for each variable (differences between the values that were obtained at the time of the first

chest CT and the second chest CT) were compared for the two groups of patients. Among these parameters, Δ ALT values were found to be statistically significantly different between the two groups ($p = 0.03$). Additionally, patients with elevated lung CT scores had a significant change in Δ ALT values ($p = 0.02$). No significant difference was found for Δ AST values between the two groups.

In the progressive group, 14 (70%) patients and 18 (90%) of patients had elevated CRP levels at hospital admission and follow-up, respectively. The comparison of the two time intervals for CRP was significantly different in the progressive group ($p = 0.039$).

There were statistically significant differences between the two groups of patients in terms of WBC, neutrophil, lymphocyte, monocyte, and platelet counts that were obtained at hospital admission ($p < 0.05$). ROC analysis was used to determine cut-off values of the laboratory parameters obtained at the time of the first chest CT to predict the changes in the lung CT scores. Patients with a WBC count $< 6 \times 10^3/\mu\text{L}$, neutrophil count $< 3.3 \times 10^3/\mu\text{L}$, lymphocyte count $< 1.7 \times 10^3/\mu\text{L}$, monocyte count $< 0.55 \times 10^3/\mu\text{L}$, and platelet count $< 230 \times 10^3/\mu\text{L}$, which were obtained correspondingly at the time of the first chest CT, were predicted to have elevated lung CT scores in the consecutive scan (Table 3).

L/S and CT scores

The median time interval from the onset of symptoms of the disease to the first chest CT scan was 2 days (range 1–4 days). The median time interval between the first and second chest CT scans for the study population were 8 days (range 2–29 days). The median time interval between the first and second chest CT scans were 12 days (range 4–26) and 7.5 days (range 2–29) for the non-progressive group and progressive group, respectively. Table 4 shows lung CT scores, L/S, and liver densities on CT of the two groups of patients for

Table 2 Laboratory characteristics of the two groups of patients with COVID-19

Parameter	Non-progressive group (n=7)		Progressive group (n=20)		Between group comparison (p)	Hospital admission and follow-up comparison (p)		
	At hospital admission	Follow-up	At hospital admission	Follow-up				
AST*	19 (10–28)	17 (12–20)	18 (7–51)	22 (5–97)	0.825 ^a	0.071 ^b	0.149 ^c	0.141 ^d
Increased (n, %)	0 (0)	0 (0)	2 (10)	5 (25)				
ALT*	18 (5–58)	19 (6–50)	16 (6–50)	29 (9–127)	0.471 ^a	0.030^b	0.344 ^c	0.020^d
Increased (n, %)	2 (28.6)	1 (14.3)	1 (5)	5 (25)				
ALP	61 (39–109)	60 (39–248)	60 (36–98)	55.5 (30–124)	0.725		0.246	
Increased (n, %)	1 (14.3)	1 (14.3)	0 (0)	1 (5)				
GGT	20.5 (11–39)	21 (7–63)	17.5 (7–81)	24 (10–177)	0.950		0.148	
Increased (n, %)	1 (14.3)	2 (28.6)	2 (10)	6 (30)				
Total bilirubin	0.2 (0.1–0.4)	0.3 (0.1–0.4)	0.3 (0.01–1.3)	0.4 (0.2–1.2)	0.063		0.006	
Increased (n, %)	0 (0)	0 (0)	1 (5)	2 (10)				
Creatinine*	0.7 (0.4–0.9)	0.8 (0.4–1)	0.8 (0.1–3.7)	0.7 (0.5–2.7)	0.234 ^a	0.009^b	0.027^c	0.098 ^d
Increased (n, %)	0 (0)	0 (0)	2 (10)	1 (5)				
Urea	29 (14–33)	25 (13–34)	30 (12–235)	27 (16–221)	0.376		0.035	
Increased (n, %)	0 (0)	0 (0)	2 (10)	2 (10)				
CRP*	4.4 (0.3–135.6)	0.8 (0.3–211)	12.6 (0.5–310.9)	37.7 (1–176)	0.439 ^a	0.105 ^b	0.249 ^c	0.039^d
Increased (n, %)	3 (42.8)	1 (14.3)	14 (70)	18 (90)				
LDH	194 (132–384)	182 (166–376)	204 (138–300)	211 (117–364)	0.949		0.700	
Increased (n, %)	2 (28.6)	3 (42.8)	5 (25)	7 (35)				
D-dimer	275 (169–1355)	248 (168–1413)	647.5 (169–1919)	643.5 (169–2733)	0.114		0.856	
Increased (n, %)	2 (28.6)	3 (42.8)	11 (55)	8 (40)				
WBC	8.3 (4.3–13.9)	8.5 (5.1–13.2)	4.9 (0.6–8.9)	5.1 (0.5–12.1)	0.000^e		0.088	
Increased (n, %)	1 (14.3)	1 (14.3)	0 (0)	1 (5)				
Decreased (n, %)	1 (14.3)	0 (0)	7 (35)	6 (30)				
Neutrophil	4.7 (2–7.6)	5.3 (2.4–10.3)	3 (0.4–6.7)	3.3 (0.3–9.7)	0.037^e		0.208	
Increased (n, %)	1 (14.3)	1 (14.3)	0 (0)	1 (5)				
Decreased (n, %)	0 (0)	0 (0)	5 (25)	3 (15)				
Lymphocyte	2.4 (1.3–4.7)	2.3 (1.5–3.6)	1.4 (0.08–2.1)	(0.1–2)	0.000^e		0.777	
Increased (n, %)	2 (28.6)	0 (0)	0 (0)	0 (0)				
Decreased (n, %)	0 (0)	0 (0)	6 (30)	4 (20)				
Monocyte	0.9 (0.3–1.3)	0.8 (0.4–1)	0.5 (0.1–0.9)	0.5 (0.07–1)	0.009^e		0.731	
Increased (n, %)	4 (57.1)	2 (28.6)	2 (10)	2 (10)				
NLR	1.6 (0.8–5)	2 (0.9–6.1)	2.2 (1–4.8)	2.7 (0.9–14.9)	0.267		0.299	
Hb	11.3 (8.2–16.2)	11.5 (7.4–16.1)	13.3 (1.9–17)	12.7 (4.5–16.4)	0.475		0.050	
Decreased (n, %)	3 (42.8)	3 (42.8)	3 (15)	3 (15)				
Platelet	253 (203–399)	285 (191–409)	190 (68–247)	219 (86–336)	0.000^e		0.084	
Increased (n, %)	0 (0)	0 (0)	0 (0)	0 (0)				
Decreased (n, %)	0 (0)	0 (0)	2 (10)	2 (10)				

A p value of < 0.05 was considered to be statistically significant in bold

Data are median (minimum–maximum). *ALP* alkaline phosphatase (35–104 U/L), *ALT* alanine aminotransferase (normal limits, < 45 U/L), *AST* aspartate amino transferase (normal limits, < 35 U/L), creatinine (0.6–1.1 mg/dL), *CRP* C-reactive protein (0–5 mg/L), d-dimer (< 550 µg/L FEU), *GGT* gamma-glutamyltransferase (normal limits, < 38 U/L), *Hb* hemoglobin (11.7–16 g/dL), *LDH* lactate dehydrogenase (normal limits, < 225 IU/L), lymphocyte count ($1-3.38 \times 10^3/\mu\text{L}$), monocyte count ($0-0.8 \times 10^3/\mu\text{L}$), neutrophil count ($2.02-7.46 \times 10^3/\mu\text{L}$), *NLR* neutrophil / lymphocyte ratio, platelet count ($150-450 \times 10^3/\mu\text{L}$) total bilirubin (0.1–1 mg/dL), urea (10–50 mg/dL), *WBC* white blood cells ($4.5-11 \times 10^3/\mu\text{L}$)

*Interactions were found for these variables. Interactions represent that the changes of laboratory parameters obtained at hospital admission and follow-up were different between the two groups

^aComparison of the parameters between progressive and non-progressive groups at the time of hospital admission

^bComparison of Δ parameters between the progressive and non-progressive groups

^cComparison of differences of the parameters obtained at the time of follow-up and hospital admission in the non-progressive group

^dComparison of differences of the parameters obtained at the time of follow-up and hospital admission in the progressive group

^eStatistically significant difference was found between progressive and non-progressive groups in terms of laboratory parameters at hospital admission

Table 3 Predictors at hospital admission for the progression of lung CT score

Parameter	Cut-off value ^a	AUC	95% confidence interval		Sensitivity (%)	Specificity (%)	PPV (%)	NPV (%)	Std. Error	<i>p</i>
			Lower	Upper						
WBC count	6	0.850	0.651	1	85	85.7	94.4	66.7	0.102	0.007
Neutrophil count	3.3	0.757	0.531	0.983	75	71.4	88.2	50	0.115	0.046
Lymphocyte count	1.7	0.843	0.654	1	90	71.4	90.5	83.3	0.096	0.008
Monocyte count	0.55	0.757	0.510	1	70	57.1	82.4	40	0.126	0.046
Platelet count	230	0.929	0.813	1	95	85.7	95	85.7	0.059	0.001

^aCut-off value is given as ($\times 10^3/\mu\text{L}$). AUC area under curve, lymphocyte count ($1\text{--}3.38 \times 10^3/\mu\text{L}$), monocyte count ($0\text{--}0.8 \times 10^3/\mu\text{L}$), neutrophil count ($2.02\text{--}7.46 \times 10^3/\mu\text{L}$), NPV negative predictive value, platelet count ($150\text{--}450 \times 10^3/\mu\text{L}$), PPV positive predictive value, WBC white blood cells ($4.5\text{--}11 \times 10^3/\mu\text{L}$)

Table 4 Liver-to-spleen ratio of the two groups of COVID-19 patients

	Non-progressive group (<i>n</i> = 7)		Progressive group (<i>n</i> = 20)	
	At hospital admission	Follow-up	At hospital admission	Follow-up
CT score	1 (0–19)	1 (0–19)	6.5 (0–14)	12 (5–24)
L/S	1.21 ± 0.29^a	1.25 ± 0.29^a	1.13 ± 0.3^b	1.02 ± 0.23^b
Liver density (HU)	52.52 ± 12.4	55.19 ± 12.83	52.6 ± 12.8	49.87 ± 11.09
	56.67 (30.33–65.33)	58 (38–73.67)	55.5 (17.33–65)	52 (17.33–66)

Data are median (minimum–maximum) and mean \pm standard deviation. L/S liver-to-spleen ratio

^aIn the non-progressive group, there was no significant difference between the comparison of L/S values that were obtained at hospital admission and follow-up ($p = 0.465$) (paired *t*-test)

^bIn the progressive group, there was a significant difference between the comparison of L/S values that were obtained at hospital admission and follow-up ($p = 0.014$) (paired *t*-test)

There was no significant difference of L/S between the two groups at hospital admission ($p = 0.547$) (independent samples *t*-test)

Follow-up L/S was significantly different between the two groups ($p = 0.009$) (covariance analysis)

The change between the baseline and follow-up L/S was statistically significant for the progressive group ($p = 0.028$) (independent samples *t*-test)

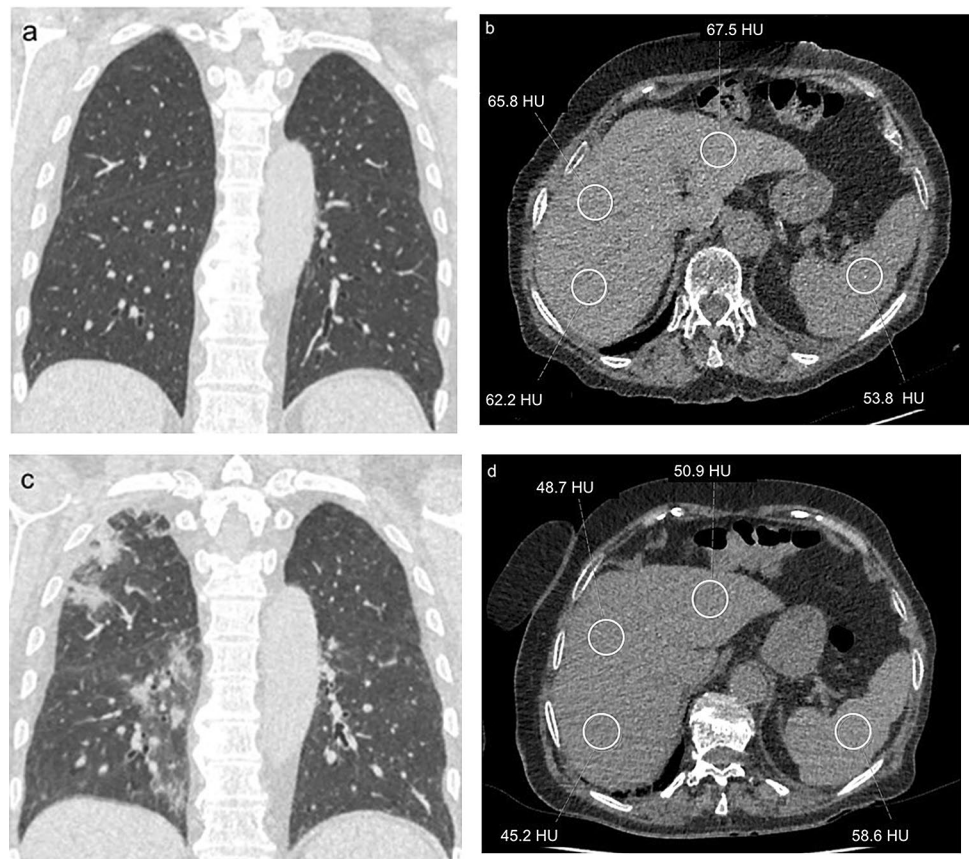
the two time points. The mean L/S of the progressive group was lower than that of the non-progressive group at hospital admission but there was no significant difference between the two groups ($p = 0.547$). However, L/S was found to be significantly different between the two groups of patients at follow-up ($p = 0.009$) (Fig. 3). For the intragroup analysis, there was a significant change in L/S in patients with increased lung CT score ($p = 0.014$). In the progressive group, as the median lung CT score was increased from 6.5 (range 0–14) to 12 (range 5–24) in two consecutive scans, the mean L/S was decreased from 1.13 ± 0.3 to 1.02 ± 0.23 . For the non-progressive group, L/S was elevated in the second CT scan, however the change in L/S was not statistically significant for the intragroup analysis ($p = 0.465$). The change between the baseline and follow-up L/S was found to be statistically significant for the progressive group ($p = 0.028$).

L/S and laboratory findings

For the whole group of patients, there was no significant correlation between L/S and ALT or AST that were obtained at hospital admission. However, negative correlations were found between L/S and both ALT and AST at follow-up ($r = -0.534$, $p = 0.004$ for L/S and ALT; $r = -0.46$, $p = 0.016$ for L/S and AST). As L/S decreased, both ALT and AST values increased.

An analysis was performed to evaluate the relationship between the therapies that were used for COVID-19 and liver function tests. When separate comparisons were performed between the patients who received hydroxychloroquine, azithromycin, other antibiotics, oseltamivir and who did not receive these agents, no significant differences of AST or ALT were found. However in patients who received favipiravir therapy, there was a significant change in both

Fig. 3 66-year old woman with COVID-19. **(a)** CT lung score was 0 at hospital admission. **(b)** Liver-to-spleen ratio at hospital admission was calculated as 1.2. **(c)** CT lung score was 12 at follow-up CT scan which was obtained 10 days later. **(d)** Liver-to-spleen ratio at follow-up was calculated as 0.82



AST and ALT values ($p < 0.05$). Patients who had favipiravir treatment had increased AST and ALT levels at follow-up, however there was no significant change in L/S between the patients who received favipiravir and who did not.

Discussion

In this study, we aimed to monitor the changes in clinical, laboratory features, and L/S of COVID-19 patients during the course of disease. Many patients who are hospitalized due to COVID-19 get multiple chest CT scans depending on the clinical progression. There are a few series in the literature that have demonstrated how evolution of thoracic imaging findings happen [2, 11, 20]. In spite of the commonly observed chest imaging findings associated with COVID-19, reports of abdominal imaging findings are limited. A study has demonstrated that bowel involvement and cholestasis are the most common abdominal imaging findings in COVID-19 patients [23]. It has been reported that 16–53% of patients with COVID-19 have abnormal liver function tests during disease progression [24–26]. However, how the changes in liver parenchyma will be seen on imaging is not clear. It is known that changes of hepatic attenuation may be observed in some diffuse liver diseases and L/S is commonly used to

assess the presence of liver fat [14, 15, 17]. In our study, we hypothesized the use of L/S to evaluate the changes in hepatic attenuation between the two consecutive scans and investigated if it may correlate with abnormal liver function tests. We found that L/S at follow-up was significantly different between the progressive and non-progressive groups. As the lung CT scores increased, L/S tended to decrease on follow-up CTs. Similarly, in our study, liver function tests did not differ between the two groups of patients at hospital admission however, the increase in ALT values at follow-up was significant in patients with progressed lung CT scores ($p = 0.02$). Additionally, in patients with decreased L/S, ALT levels were elevated at follow-up ($p = 0.004$).

Although it is difficult to determine the underlying mechanism, our study demonstrated the changes in hepatic attenuation during the course of disease in COVID-19 patients. It has been proposed that liver injury in COVID-19 patients might be the direct result of the infection of liver cells [13]. The derangement of liver function is generally mild and other explanations for the liver damage such as immune-mediated inflammation, cytokine storm, and pneumonia-associated hypoxia are suggested [13, 27]. Abnormal liver function tests may also be observed in drug-induced hepatotoxicity with the use of hydroxychloroquine, azithromycin, and favipiravir in COVID-19 patients [28–30]. In our

study, only in the group of patients who received favipiravir therapy had significantly elevated AST and ALT levels at follow-up, however, there was no change of L/S between the groups.

It has been suggested that hepatic steatosis might develop during the course of COVID-19 [31]. Histopathological examinations of the liver in COVID-19 patients have shown hepatic steatosis, mild lobular and portal inflammation [31, 32]. The decrease in L/S in the progressive group on follow-up that was identified in our study might be attributed to hepatic steatosis. In general, hepatic steatosis is most commonly associated with alcohol abuse and non-alcoholic fatty liver disease but it may be caused by various factors including viral hepatitis, ischemia, drug toxicity, and malnutrition [33]. Moreover, it has been demonstrated that SARS-CoV-2 infection may affect lipid metabolisms especially in patients with severe disease [34, 35]. Thus, the causes of decreased L/S in patients with progressive disease need to be further investigated.

Chest CT plays an important role in the diagnosis and evaluation of severity of COVID-19. In our study, patients with increased lung CT scores had a significantly longer hospital stay ($p=0.035$). Similarly, Li et al. reported that the CT scores of the patients with critical disease were higher than those of the ordinary COVID-19 patients and lung CT scores can be used to accurately differentiate severe patients [12]. In our study, in patients with progressed lung CT scores, CRP levels were found to be significantly elevated at follow-up. It has been suggested that increased values of CRP may be positively correlated with lung involvement and disease severity [36].

Laboratory parameters such as WBC, absolute values of lymphocytes, platelets, albumin, total bilirubin, creatinine, blood urea nitrogen, CRP, interleukin-6, AST, LDH, myoglobin, troponin, procalcitonin, and d-dimer have been reported to be risk factors for critical or mortal COVID-19 cases [10, 37]. In our study, WBC, neutrophil, lymphocyte, monocyte, and platelet levels at hospital admission were significantly different between the progressive and non-progressive groups of patients. Among these parameters, platelet count $< 230 \times 10^3/\mu\text{L}$ predicted the progression of lung CT score with a sensitivity of 95% and specificity of 85.7%.

Our study has several limitations including its retrospective design and small number of patients. Another limitation is that we did not evaluate the thoracic imaging characteristics of COVID-19. It has been reported that consolidation, linear opacities, crazy-paving pattern, bronchial wall thickening, and extrapulmonary lesions may be the imaging findings of severe COVID-19 patients. In our study, we evaluated the changes of L/S during the course of disease and used the lung CT scores to assess the extension and severity of the lung involvement in COVID-19 patients. Another limitation is the lack of contrast-enhanced liver

imaging. Since multiple unenhanced chest CT scans were performed for COVID-19 patients, we wanted to investigate the changes in upper abdomen and focused on L/S. As L/S can be affected by the pre-existing liver diseases, we only evaluated the changes on two consecutive scans and excluded COVID-19 patients who had single CT scans. However, prospective studies with larger number of patients, using different imaging modalities are required to confirm and radiologically detect the changes in liver parenchyma, which could be multifactorial, in patients with COVID-19.

In conclusion, decrease in L/S may be observed in COVID-19 patients with elevated lung CT scores at follow-up. Additionally, WBC, neutrophil, lymphocyte, monocyte, and platelet counts that are obtained at hospital admission may predict the progression of disease.

Compliance with ethical standards

Conflict of interest All the authors declared that they have no conflicts of interest.

References

1. World Health Organization Timeline – COVID-19. Available at: <https://www.who.int/news-room/detail/27-04-2020-who-timeline--covid-19> Accessed 6 May 2020.
2. Ai T, Yang Z, Hou H, et al (2020) Correlation of chest CT and RTPCR testing in coronavirus disease 2019 (COVID-19) in China: a report of 1014 cases. *Radiology* 200642. <https://doi.org/10.1148/radiol.2020200642>
3. Fang Y, Zhang H, Xie J et al (2020) Sensitivity of chest CT for COVID-19: comparison to RT-PCR. *Radiology* 200432. <https://doi.org/10.1148/radiol.2020200432>
4. Xie X, Zhong Z, Zhao W, et al (2020) Chest CT for typical 2019-nCoV pneumonia: relationship to negative RT-PCR testing. *Radiology* 200343. doi:<https://doi.org/10.1148/radiol.2020200343>
5. Li Y, Xia L (2020) Coronavirus disease 2019 (COVID-19): role of chest CT in diagnosis and management. *AJR Am J Roentgenol* 214:1280–1286. doi:<https://doi.org/10.2214/AJR.20.22954>
6. Huang C, Wang Y, Li X, et al (2020) Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 395:497–506. [https://doi.org/10.1016/S0140-6736\(20\)30183-5](https://doi.org/10.1016/S0140-6736(20)30183-5)
7. Wang D, Hu B, Hu C, et al (2020) Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China. *JAMA* 323:1061–1069. <https://doi.org/10.1001/jama.2020.1585>
8. Chen N, Zhou M, Dong X, et al (2020) Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet* 395:507–513. [https://doi.org/10.1016/S0140-6736\(20\)30211-7](https://doi.org/10.1016/S0140-6736(20)30211-7)
9. Zhou F, Yu T, Du R, et al (2020) Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet* 395:1054–1062. [https://doi.org/10.1016/S0140-6736\(20\)30566-3](https://doi.org/10.1016/S0140-6736(20)30566-3)
10. Zheng Z, Peng F, Xu B, et al (2020) Risk factors of critical & mortal COVID-19 cases: A systematic literature review and meta-analysis. *J Infect* S0163-4453(20)30234-6. doi:<https://doi.org/10.1016/j.jinf.2020.04.021>

11. Wan S, Li M, Ye Z, et al (2020) CT manifestations and clinical characteristics of 1115 patients with Coronavirus disease 2019 (COVID-19): a systematic review and meta-analysis. *Acad Radiol* 27:910-921. <https://doi.org/10.1016/j.acra.2020.04.033>
12. Li K, Wu J, Wu F, et al (2020) The clinical and chest CT features associated with severe and critical COVID-19 pneumonia. *Invest Radiol* 55:327-331. doi:<https://doi.org/10.1097/RLI.0000000000000672>
13. Zhang C, Shi L, Wang FS (2020) Liver injury in COVID-19: management and challenges. *Lancet Gastroenterol Hepatol* 5:428-430. [https://doi.org/10.1016/S2468-1253\(20\)30057-1](https://doi.org/10.1016/S2468-1253(20)30057-1)
14. Kodama Y, Ng CS, Wu TT et al (2007) Comparison of CT methods for determining the fat content of the liver. *AJR Am J Roentgenol* 188:1307-1312. <https://doi.org/10.2214/AJR.06.0992>
15. Ma X, Holalkere NS, Kambadakone R A, et al (2009) Imaging-based quantification of hepatic fat: methods and clinical applications. *Radiographics* 29:1253-1277. <https://doi.org/10.1148/rg.295085186>
16. Yasui S, Fujiwara K, Okitsu K, et al (2012) Importance of computed tomography imaging features for the diagnosis of autoimmune acute liver failure. *Hepatol Res* 42:42-50. <https://doi.org/10.1111/j.1872-034X.2011.00892.x>
17. Iwasaki M, Takada Y, Hayashi M, et al (2004) Noninvasive evaluation of graft steatosis in living donor liver transplantation. *Transplantation* 78:1501-1505. <https://doi.org/10.1097/01.tp.0000140499.23683.0d>
18. COVID-19 treatment algorithm for adults. Available at: <https://covid19bilgi.saglik.gov.tr/depo/algorithmalar/COVID19> Accessed 11 May 2020.
19. Chang YC, Yu CJ, Chang SC, et al (2005) Pulmonary sequelae in convalescent patients after severe acute respiratory syndrome: evaluation with thin-section CT. *Radiology* 236:1067-1075. <https://doi.org/10.1148/radiol.2363040958>
20. Pan F, Ye T, Sun P, et al (2020) Time course of lung changes on chest CT during recovery from 2019 novel coronavirus (COVID-19) pneumonia. *Radiology* 295:715-721. <https://doi.org/10.1148/radiol.2020200370>
21. Francone M, Iafrate F, Masci GM, et al (2020) Chest CT score in COVID-19 patients: correlation with disease severity and short-term prognosis [published online ahead of print, 2020 Jul 4]. *Eur Radiol* 1–10. <https://doi.org/10.1007/s00330-020-07033-y>
22. Zeb I, Li D, Nasir K, et al (2012) Computed tomography scans in the evaluation of fatty liver disease in a population based study: the multi-ethnic study of atherosclerosis. *Acad Radiol* 19:811-818. <https://doi.org/10.1016/j.acra.2012.02.022>
23. Bhayana R, Som A, Li MD, et al (2020) Abdominal imaging findings in COVID-19: preliminary observations. *Radiology* 201908. <https://doi.org/10.1148/radiol.2020201908>
24. Shi H, Han X, Jiang N, et al (2020) Radiological findings from 81 patients with COVID-19 pneumonia in Wuhan, China: a descriptive study. *Lancet Infect Dis* 20:425-434. [https://doi.org/10.1016/S1473-3099\(20\)30086-4](https://doi.org/10.1016/S1473-3099(20)30086-4)
25. Xu XW, Wu XX, Jiang XG, et al (2020) Clinical findings in a group of patients infected with the 2019 novel coronavirus (SARS-Cov-2) outside of Wuhan, China: retrospective case series. *BMJ* 368:m606. <https://doi.org/10.1136/bmj.m606>
26. Fan Z, Chen L, Li J, et al (2020) Clinical features of COVID-19-related liver functional abnormality. *Clin Gastroenterol Hepatol* 18:1561-1566. <https://doi.org/10.1016/j.cgh.2020.04.002>
27. Bangash MN, Patel J, Parekh D (2020) COVID-19 and the liver: little cause for concern. *Lancet Gastroenterol Hepatol* 5:529-530. [https://doi.org/10.1016/S2468-1253\(20\)30084-4](https://doi.org/10.1016/S2468-1253(20)30084-4)
28. Falcão MB, Pamplona de Góes Cavalcanti L, Filgueiras Filho NM, et al (2020) Case report: hepatotoxicity associated with the use of hydroxychloroquine in a patient with novel coronavirus disease (COVID-19). *Am J Trop Med Hyg* 102:1214-1216. <https://doi.org/10.4269/ajtmh.20-0276>
29. Boeckmans J, Rodrigues RM, Demuyser T, et al (2020) COVID-19 and drug-induced liver injury: a problem of plenty or a petty point? *Arch Toxicol* 94:1367-1369. doi:<https://doi.org/10.1007/s00204-020-02734-1>
30. Cai Q, Yang M, Liu D, et al (2020) Experimental treatment with favipiravir for COVID-19: an open-label control study. *Engineering (Beijing)* 10.1016/j.eng.2020.03.007. <https://doi.org/10.1016/j.eng.2020.03.007>
31. Lagana SM, Kudose S, Iuga AC, et al (2020) Hepatic pathology in patients dying of COVID-19: a series of 40 cases including clinical, histologic, and virologic data. *Mod Pathol* 1-9. doi:<https://doi.org/10.1038/s41379-020-00649-x>
32. Xu Z, Shi L, Wang Y, et al (2020) Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *Lancet Respir Med* 8:420-422. [https://doi.org/10.1016/S2213-2600\(20\)30076-X](https://doi.org/10.1016/S2213-2600(20)30076-X)
33. Milroy CM (2018) Fatty liver and the forensic pathologist. *Acad Forensic Pathol* 8:296-310. <https://doi.org/10.1177/1925362118782061>
34. Wei X, Zeng W, Su J, et al (2020) Hypolipidemia is associated with the severity of COVID-19. *J Clin Lipidol* 14:297-304. <https://doi.org/10.1016/j.jacl.2020.04.008>
35. Wang G, Zhang Q, Zhao X, et al (2020) Low high-density lipoprotein level is correlated with the severity of COVID-19 patients: an observational study. *Lipids Health Dis* 19:204. <https://doi.org/10.1186/s12944-020-01382-9>
36. Wang L (2020) C-reactive protein levels in the early stage of COVID-19. *Med Mal Infect* 50:332-334. <https://doi.org/10.1016/j.medmal.2020.03.007>
37. Ruan Q, Yang K, Wang W, et al (2020) Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan, China. *Intensive Care Med* 46:846-848. <https://doi.org/10.1007/s00134-020-05991-x>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.