

# Acute beetroot juice supplementation improves exercise tolerance and cycling efficiency in adults with obesity

Christian E. Behrens Jr<sup>1</sup> | Khandaker Ahmed<sup>2</sup> | Karina Ricart<sup>2</sup> | Braxton Linder<sup>3</sup> | José Fernández<sup>1</sup> | Brenda Bertrand<sup>1</sup> | Rakesh P. Patel<sup>2</sup> | Gordon Fisher<sup>3</sup> 

<sup>1</sup>Department of Nutrition Sciences, The University of Alabama at Birmingham, Birmingham, AL, USA

<sup>2</sup>Department of Pathology and Center for Free Radical Biology, The University of Alabama at Birmingham, Birmingham, AL, USA

<sup>3</sup>Department of Human Studies, The University of Alabama at Birmingham, Birmingham, AL, USA

## Correspondence

Gordon Fisher, Department of Human Studies, University of Alabama at Birmingham, Education Building, 202, 901 13th St. South, Birmingham, AL 35294, USA.  
Email: grdnfs@uab.edu

## Funding information

This work was supported by a School of Education Faculty Development Grant awarded to Gordon Fisher. Christian Behrens is supported by a NIH T-32 Grant (T32HL105349) from the National Heart Lung and Blood Institute. Additional support provided by Award Number P30DK056336 – National Institute of Diabetes Digestive and Kidney Diseases.

## Abstract

**Background:** Exercise training improves health outcomes in individuals with obesity (IO); however, it remains challenging for IO to adhere to exercise. Thus, it is critical to identify novel strategies that improve exercise tolerance (ET) and adherence in IO. Beetroot juice (BRJ), high in inorganic dietary nitrate, consistently improves exercise performance in athletes, individuals with cardiopulmonary diseases, and nonobese lean individuals. These improvements may be explained by reduced oxygen uptake ( $\text{VO}_2$ ) during exercise, enhanced blood flow, and greater mitochondrial efficiency. To date, we are aware of no studies that have compared the effects of BRJ, sodium nitrate ( $\text{NaNO}_3$ ), and nitrate-depleted BRJ (PLA) for improving ET and cardiometabolic health in IO.

**Purpose:** Determine if BRJ improves ET, exercise efficiency (EE), and cardiometabolic health in IO and identify possible mechanisms of action.

**Methods:** Vascular hemodynamic, submaximal- and maximal-exercise  $\text{VO}_2$ , and time to exhaustion (TTE) were assessed in 16 participants 2.5 hr following consumption of: 1) BRJ, 2)  $\text{NaNO}_3$ , 3) PLA, or 4) CON.

**Results:** A significant treatment effect was observed for submaximal exercise  $\text{VO}_2$  ( $p = .003$ ), and TTE ( $p < .001$ ). Post hoc analyses revealed lower  $\text{VO}_2$  during submaximal exercise in BRJ compared to PLA ( $p = .009$ )  $\text{NaNO}_3$  ( $p = .042$ ) and CON (0.009), equating to an average improvement of ~ 7% with BRJ. TTE was greater for BRJ compared to other treatment arms, PLA ( $p = .008$ ),  $\text{NaNO}_3$  ( $p = .038$ ), and CON ( $p < 0.001$ ), equating to ~ 15% improvement with BRJ. No significant changes were observed for other outcomes.

**Conclusions:** Consumption of BRJ improved EE during submaximal exercise by 7%, and TTE by 15% compared to other conditions. These results suggest that BRJ may improve EE and exercise tolerance in IO.

## KEYWORDS

beetroot, exercise, nitrate, obesity

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2020 The Authors. Physiological Reports published by Wiley Periodicals LLC on behalf of The Physiological Society and the American Physiological Society

## 1 | INTRODUCTION

Obesity is ranked among the top growing health concerns worldwide. In the United States alone, obesity affects over 93 million adults inflicting hundreds of billions of dollars in obesity-related medical costs each year (Tremmel, Gerdtham, Nilsson, & Saha, 2017; Wolf & Colditz, 1998). This is especially concerning due to comorbidities associated with obesity such as hypertension, dyslipidemia, and type 2 diabetes. The successful treatment of obesity, particularly long term, has proven challenging, warranting continued investigation into alternative strategies that may reduce the incidence and severity of obesity and secondary cardiometabolic comorbidities.

Regular physical exercise has been shown to reduce the incidence of obesity and assist with weight maintenance following weight loss (Amisola & Jacobson, 2003; Ladabaum, Mannalithara, Myer, & Singh, 2014; McInnis, 2000; Shaw, Gennat, O'Rourke, & Del Mar, 2006). However, due to many of the biological consequences of obesity, individuals with obesity are often burdened by a diminished capacity and tolerance to exercise, often resulting in abstention and increased sedentarism (Bournat & Brown, 2010; Martínez-González, Alfredo Martínez, Hu, Gibney, & Kearney, 1999; Tryon, Goldberg, & Morrison, 1992). One potential mechanism underlying reduced exercise tolerance is impaired nitric oxide (NO) production and bioactivity that manifests in decreased systemic oxygen delivery and/or skeletal muscle oxygen utilization (Lee-Young et al., 2010; Pohl & Lamontagne, 1991). Obesity is associated with lower levels of NO production and bioavailability (Sansbury & Hill, 2014a, 2014b; Siervo, Jackson, & Bluck, 2011; Williams, Wheatcroft, Shah, & Kearney, 2002). Therefore, it is possible that strategies which increase NO production in individuals with obesity may improve oxygen delivery and/or utilization, increase exercise tolerance, and overall exercise participation, leading to improvements in secondary cardiometabolic comorbidities associated with obesity.

Previous work suggests supplementation with inorganic dietary nitrate ( $\text{NO}_3^-$ ) found in many plant-based sources such as beetroot, mustard leaf, spinach, and arugula, improves NO bioavailability and oxygen delivery and/or utilization leading to increased exercise tolerance and improvements in cardiometabolic health outcomes, such as blood pressure, endothelial function, and measures of inflammation and oxidative stress (Clements, Lee, & Bloomer, 2014; Clifford, Howatson, West, & Stevenson, 2015; Larsen et al., 2011; Lundberg & Govoni, 2004). Beetroot juice (BRJ), is exceptionally high in  $\text{NO}_3^-$  and is commonly used to assess potential improvements in human health and exercise performance (Jones, Thompson, Wylie, & Vanhatalo, 2018). The mechanism by which BRJ leads to these improvements is that  $\text{NO}_3^-$  is

reduced to nitrite ( $\text{NO}_2^-$ ) by oral bacterial nitrate reductase. Nitrite is then further reduced to NO in hypoxic and acidic compartments (Domínguez et al., 2017). The majority of these studies have focused on ergogenic benefits of dietary nitrate to trained athletes. Few clinical studies have investigated the effect on populations with chronic diseases, such as chronic obstructive pulmonary disease and heart failure with some reporting improvements in exercise and vascular outcomes, and others reporting no effect of treatment (Berry et al., 2015; Coggan et al., 2018; Eggebeen et al., 2016; Gilchrist et al., 2013). These studies have also provided insights into potential mechanisms underlying the ergogenic effects of BRJ including modulation of  $\text{O}_2$  metabolism characterized by faster pulmonary  $\text{O}_2$  consumption, (Breese et al., 2013). vasodilation increasing blood flow and oxygen delivery to muscles (Erzurum et al., 2007), improved ATP utilization by exercising myofibrils, and increased mitochondrial biogenesis (Larsen et al., 2011; Stamler & Meissner, 2001; Tong, Heim, & Wu, 2011). However, BRJ also contains many potent antioxidant compounds, which may confer additional benefits for exercise performance and cardiometabolic health. Thus, it is possible that BRJ can increase antioxidant capacity and blunt pro-inflammatory pathways, providing a protective benefit against elevated oxidative stress observed in IO (Clifford et al., 2015; Georgiev et al., 2010; Kanner, Harel, & Granit, 2001; Vulić et al., 2014; Zielinska-Przyjemka, Olejnik, Dobrowolska-Zachwieja, & Grajek, 2009). It remains to be determined if the improvements in exercise and health outcomes following BRJ intake are due to increases in nitrate, antioxidant capacity, or a synergistic effect of both (Georgiev et al., 2010; Kanner et al., 2001; Reddy, Alexander-Lindo, & Nair, 2005).

To date, there have been very few clinical investigations into the efficacy of BRJ supplementation on obesity, although previous investigations have been similar in scope (Beals et al., 2017; Lara et al., 2015; Lima Bezerra et al., 2019; Rasica et al., 2018), to our knowledge there have been no clinical investigations specifically examining its effect on exercise tolerance in adults with obesity. Thus, the primary purpose of this study is to determine if BRJ supplementation improves exercise tolerance and markers of cardiometabolic health in individuals with obesity, and secondly to determine if the ergogenic and cardiometabolic health benefits of BRJ are due to increased nitric oxide bioavailability, increased antioxidant capacity, or a combination of these two. We utilize a three treatment crossover design involving a nitrate-rich BRJ supplement containing all naturally occurring  $\text{NO}_3^-$  and antioxidant properties, a nitrate-depleted BRJ supplement containing only the naturally occurring antioxidant properties but negligible  $\text{NO}_3^-$ , and finally a nitrate-matched sodium nitrate solution used as a surrogate vehicle for  $\text{NO}_3^-$  delivery, without any added antioxidant compounds.

## 2 | METHODS

### 2.1 | Participants

The protocol for this study was reviewed and approved by the institutional review board at the University of Alabama at Birmingham (IRB-300001210). Men and women ( $N = 16$ ) were recruited for this randomized crossover study. Inclusion criteria were age 19 to 40 years, sedentary (not currently engaging in structured weekly physical activity), and obese (body mass index  $>30$  kg/m<sup>2</sup>). Female participants were enrolled during the luteal phase of their menstrual cycle, lasting 12–14 days. Confirmation of menstrual cycle phase was self-reported by female participants. Participants were non-smokers and otherwise considered healthy with no history of chronic or degenerative disease. Participants were not currently taking any form of supplement or medication known to interfere with cardiometabolic or exercise measures associated with the study protocol. Participants were also asked to abstain from using antibacterial mouthwash (shown to interfere with NO<sub>3</sub><sup>-</sup> metabolism) and, antioxidant and pre/probiotic supplements for the duration of the experimental period.

### 2.2 | Experimental design

Participants visited the laboratory on five separate occasions over approximately an 18-day period. Each visit was separated by at least a 72-hr washout period to assure that effects of supplementation returned to baseline. On *visit 1* participants gave written informed consent to participate in the study. Total body composition was measured using dual-energy X-ray absorptiometry (DXA) and participants performed a graded incremental exercise test on an electronically braked cycle ergometer for the determination of VO<sub>2peak</sub> and calculation of the gas exchange threshold (GET). *Visits 2–5* were identical in nature with the exception of the supplement consumed and consisted of blood and saliva collection at three separate time points, measurements of vascular health using pulse wave contour analysis (systolic/diastolic blood pressure, pulse rate, large, and small artery elasticity), and a time to exhaustion test (TTE). Study schematic detailed in Figure 1. Experimental visits all began at the same time of day (7:00 a.m.). Participants were asked to arrive to each experimental visit in a fasted state. All participants

were instructed to maintain their regular diet throughout the course of their enrollment in the study. Diet recalls (24 hr) were obtained by a registered dietitian at each experimental visit. Data from 24-hr recalls were entered into diet analysis software (NutriTiming® LLC, Atlanta, GA, USA) and examined for total calories, macronutrients, and other micronutrients known to influence antioxidant status. Dietary data were used to test for differences between and within participants for diet composition.

### 2.3 | Supplementation

In a single-blind crossover design, participants were randomized to one of four conditions at each visit: (a) NO<sub>3</sub><sup>-</sup> rich beetroot juice (BRJ; ~6.4 mmol of NO<sub>3</sub><sup>-</sup> per 70 ml; Beet it; James White Drinks, Ipswich, United Kingdom), (b) NO<sub>3</sub><sup>-</sup> depleted beetroot juice (PLA; ~0.04 mmol of NO<sub>3</sub><sup>-</sup> per 70 ml; Beet it; James White Drinks, Ipswich, United Kingdom), (c) sodium nitrate solution matched for the amount of NO<sub>3</sub><sup>-</sup> delivered by the BRJ (NaNO<sub>3</sub>; 0.5 grams of pure sodium nitrate, Sigma-Aldrich *ReagentPlus*®, dissolved in 50ml of deionized water), or (d) no supplement control (CON). Although the BRJ and PLA conditions were indiscernible to participants, we acknowledge a limitation to this study design was our inability to further blind participants to the NaNO<sub>3</sub> and CON conditions. At each visit, following baseline vascular measures and blood and saliva collection, participants were instructed to consume the supplement and then remained in the Clinical Research Unit in a seated and relaxed position for 2.5 hr.

### 2.4 | Body composition measurement

Body mass was obtained in minimal clothing on a digital platform scale (Seca 514 mBCA, Seca North America, Chino, CA). Height was measured with shoes removed using a digital wall-mounted stadiometer (Seca 264 Stadiometer, Seca North America, Chino, CA). Total body fat, lean mass, and visceral fat were measured by DXA with the use of a Lunar iDXA densitometer (GE Medical Systems, Madison, WI, enCORE Version 15). Participants were required to wear light clothing, remove all metal objects, and lie supine with arms at their sides during the scan.

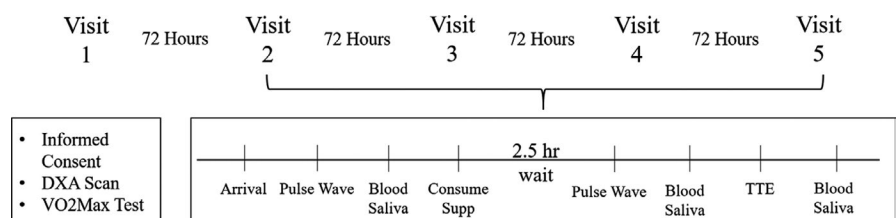


FIGURE 1 Study schematic

## 2.5 | Graded incremental exercise test

During *Visit 1* participants performed a graded incremental exercise test on an electronically braked cycle ergometer (Lode Excalibur Sport, Groningen, Netherlands) for the determination of  $\text{VO}_{2\text{peak}}$  and GET. Participants cycled for 3 min at 0 watts, after which the work rate was increased 30 watts per minute until the participant was unable to continue. Aerobic power was defined as the highest 30 s average volume of oxygen consumed before the participant's volitional exhaustion. Minute ventilation, oxygen uptake, and carbon dioxide production were continuously recorded by an open-circuit spirometry system (TrueOne 2400, ParvoMedics, Salt Lake City, UT). Additionally, heart rate was continuously recorded (Polar T31 Coded Heartrate Transmitter, Polar Electro Oy, Kempele, Finland).  $\text{VO}_{2\text{peak}}$  confirmation was determined by meeting two of the following three criteria: (a) a plateau in oxygen consumption as workload increases, (b) a respiratory exchange ratio greater than 1.15, (c) a max heart rate within 10–12 beats or the age predicted heart rate max. Breath-by-breath  $\text{VO}_2$  (L/min) and  $\text{VCO}_2$  (L/min) was obtained and the participants GET was calculated using the V-slope method (Schneider, Phillips, & Stoffolano, 1993).

## 2.6 | Time to exhaustion test

During *Visits 2–5* participants underwent a TTE test 2.5 hr postsupplementation. Minute ventilation, oxygen uptake, and carbon dioxide production were continuously recorded by an open-circuit spirometry system. Heart rate was also recorded throughout each test. Testing began with participants cycling for 3 min at a workload of 20 watts. After 3 min of warm-up, the workload was increased to 90% of the workload performed at the participants GET for 5 min. After 5 min the workload was increased to 90% of the workload performed at the participants  $\text{VO}_{2\text{peak}}$  where cycling continued until volitional exhaustion or cycling cadence decreased below 60 rpm. The duration of exercise during this intensity was used to determine the participants TTE. Gas exchange data collected during the last 3 min of the second stage was used for submaximal  $\text{VO}_2$  comparison between arms and all gas exchange collected during the third stage of the test used for maximal  $\text{VO}_2$  comparisons.

## 2.7 | Arterial elasticity test

Hemodynamic and arterial elasticity variables (Systolic/Diastolic blood pressure, mean arterial blood pressure, pulse rate, large/small artery elasticity total vascular impedance, systemic vascular resistance, cardiac output, estimated

cardiac index) were obtained from the radial artery using a noninvasive calibrated pulse wave tonometer (HDI-CR-2000, Hypertension Diagnostics Inc. Eagen, MN). This analysis is based on a modified Windkessel model that allows evaluation of the large conduit arteries and the small microcirculatory arteries (Cohn et al., 1995). Data from Pulse wave tonometry were obtained at two time points in triplicate during each visit (*Visits 2–5*). Upon arrival to the laboratory, participants were seated and stationary for at least 10 min prior to beginning the first pulse wave analysis. Following the first assessment, and baseline blood/saliva collection, participants received their randomized supplement and were asked to remain seated and stationary for 2.5 hr. Immediately following the 2.5 hr wait, a second pulse wave assessment was conducted. In short, in the seated position, a solid-state pressure transducer array (tonometer) was placed over the participant's radial artery at the wrist and secured with a Velcro strap around the radial bony prominence. A wrist stabilizer was used to support the participant's arm to assure positioning and minimize movement and an automated oscillatory blood pressure cuff was placed on the contralateral arm. The waveform was calibrated by the oscillometric method. Once a stable measurement was achieved, a 30 s analog tracing of the radial waveform was digitized at 200 samples per second. BP measurements were taken before, during, and after the waveform assessment. The first maximum waveform recorded represents the work of the arteries after cardiac ejection and is representative of the large arteries, whereas the following rebound wave is reflective of small artery compliance. Total vascular impedance was determined from the modified Windkessel model evaluated at the frequency of the measured heart rate (Hales, 1733).

## 2.8 | Blood and saliva collection

Blood and saliva samples were obtained at three time points during each experimental visit (*Visits 2–5*). Samples were obtained at baseline (hour 0) with supplementation immediately following, 2.5 hr postsupplementation (hour 2.5), and immediately following TTE test (hour 3). Using 2.7ml sodium citrate, and 4.0ml EDTA vacutainers (Vacutainer, Becton Dickinson, Franklin Lakes, NJ), whole blood was obtained at each time point from an IV-catheter placed in the antecubital vein. Within 2 min of collection samples obtained in sodium citrate tubes were centrifuged at 3,000 g for 2 min for subsequent nitrite and nitrate measurements. Samples obtained in EDTA tubes were centrifuged at 5,000 g for 10 min for subsequent antioxidant capacity measurements. Unstimulated saliva samples (~5ml) were collected in large 50ml conical tubes (Falcon) and subsequently separated into 1.5 ml Eppendorf tubes at each time point. Saliva samples were centrifuged at 3,000 g for ~5 min. Following centrifugation, plasma and saliva supernatant were separated into 500  $\mu\text{l}$



aliquots, snap frozen in liquid nitrogen, and stored at  $-80^{\circ}\text{C}$  for later determination of nitrite and nitrate concentration.

## 2.9 | Total antioxidant capacity

Total plasma antioxidant potential was determined by the Ferric Reducing Ability of Plasma (FRAP) assay according to the methodology of Benzie and Strain (Benzie & Strain, 1996). Working FRAP solution was placed in a water bath and warmed to  $37^{\circ}\text{C}$ . Then,  $10\ \mu\text{L}$  of blank, samples, and ascorbate STDs were transferred by micropipette into designated well of 96-well plate. Three hundred microliter of FRAP reagent was then added to all wells containing blank, samples, and standards. Ninety-six-well plate was then incubated for 4 min at  $37^{\circ}\text{C}$  before being read at 593 nm in a spectrophotometer (Molecular Devices – SpectraMax M3. Softmax Pro Version 6.3). Samples and standards were analyzed in triplicate, and FRAP values were expressed as vitamin C equivalents as determined by linear regression from a vitamin C curve (0–1000  $\mu\text{mol}$ ).

## 2.10 | Measurement of nitrite and nitrate

After thawing, methanol was added to plasma and saliva (2:1 ratio), vortex mixed and supernatant collected. Nitrite and nitrate was then measured on methanolic extracts by HPLC-coupled to the Griess reaction using the ENO-20 (EiCom, Japan). Nitrate and nitrite levels were calculated by comparison to standard curves generated daily and adjusted for extraction efficiency (Joshipura, Munoz-Torres, Morou-Bermudez, & Patel, 2017).

## 2.11 | Statistical analysis

Sample size calculations for this study were based on mean changes and standard deviations for systolic blood pressure and volume of oxygen consumed ( $\text{VO}_2$ ) during submaximal exercise established from previous work in this area (Jajja et al., 2014; Jakeman & Maxwell, 1993). Given the mean changes and  $SD$  in SBP and submaximal  $\text{VO}_2$  following BRJ supplementation ( $SD = 5\ \text{mmHg}$  and  $0.10\ \text{LO}_2/\text{min}$ ), a sample size of 11 for SBP and 6 for  $\text{VO}_2$  would be sufficient to yield a power of 80% to detect 20% differences with 95% confidence intervals. This study initially recruited 20 participants, with 16 participants completing all study requirements.

Participant characteristics from data collected during Visit 1 were analyzed using descriptive statistics, expressed as mean  $\pm SD$ . Each continuous variable was checked for normality using Kolmogorov-Smirnov tests. Normality was confirmed visually with QQ-plot observations. For analyses

**TABLE 1** Participant characteristics ( $n = 16$ )

	Men ( $n = 11$ )	Women ( $n = 5$ )
Age	$26 \pm 6.1$	$30 \pm 6.5$
Height (cm)	$179.3 \pm 5.7$	$163.3 \pm 6.5$
DXA Total (kg)	$114.1 \pm 19.5$	$89.0 \pm 11.7$
DXA Lean (kg)	$66.2 \pm 7.6$	$44.0 \pm 3.1$
DXA Fat %	$39.5 \pm 5.6$	$48.6 \pm 5.8$
$\text{VO}_2\text{Peak}$ ( $\text{mL}/\text{kg}/\text{min}$ )	$27.1 \pm 3.7$	$25.5 \pm 6.6$
SBP (mmHg)	$130 \pm 14$	$107 \pm 11$
DBP (mmHg)	$73 \pm 8$	$60 \pm 10$

Note: Values are means  $\pm SD$ .

assuming normally distributed data, log transformations were made for data not conforming to a normal distribution. Linear mixed models were used to determine main effects of treatment (BRJ, PLA, CON, and  $\text{NaNO}_3$ ) and time interaction (Visits 2–5) for all primary outcome variables (plasma and saliva  $\text{NO}_3^-/\text{NO}_2^-$ , FRAP, pulse wave variables, submaximal/maximal  $\text{VO}_2$ , and TTE). Time and treatment were set as fixed effects for each of these models with treatment order set as a random effect. Studentized residuals falling outside of three standard deviations were considered outliers and removed from analysis; this resulted in two data points being excluded for saliva NO metabolites. Significant findings were determined by an alpha level of 0.05 for pairwise comparisons. Tukey's post hoc correction was used where significant effects were observed, however, given the nature of our small sample size and the possibility for overcorrection, we chose to report significance based on unadjusted pairwise comparisons. Relationships between NO metabolites and exercise outcomes were further explored using Pearson's correlation coefficients. Statistical tests were conducted using SAS Version 9.4 (Cary, NC).

## 3 | RESULTS

Twenty participants were enrolled in this study. One participant was unable to perform exercise testing and 3 others were lost during follow-up for reasons unknown, leaving 16 participants (11 male/5 female) who completed the study. Baseline characteristics of completed participants are presented in Table 1. There were no adverse health events reported or observed from supplementation or exercise testing.

No significant differences were observed in diet composition within participants or across treatment arms (Table 2). Baseline saliva and plasma  $[\text{NO}_2^-]$  and  $[\text{NO}_3^-]$  were similar across participants and treatment arms. Absolute mean values and mean change in saliva and plasma  $[\text{NO}_2^-/\text{NO}_3^-]$  for each study arm are reported in Figure 2. Postsupplementation

	PLA	BRJ	NaNO <sub>3</sub>	CON
Total Calories	2086 ± 892	2,527 ± 1,408	1883 ± 319	2,590 ± 846
Carbohydrate (g)	276 ± 170	300 ± 149	193 ± 61	263 ± 103
Protein (g)	90 ± 26	96 ± 64	94 ± 37	118 ± 38
Fat (g)	112 ± 63	116 ± 86	84 ± 20	122 ± 51
Vitamin A (IU)	391 ± 137	355 ± 112	369 ± 119	384 ± 127
Vitamin C (mg)	25 ± 11	28 ± 16	30 ± 19	31 ± 24
Vitamin E (mg)	3.1 ± 2.3	4.7 ± 4.1	3.3 ± 1.6	4.6 ± 2.8
Copper (mcg)	0.53 ± 0.40	0.93 ± 0.42	0.60 ± 0.39	0.80 ± 0.46
Manganese (mg)	1.9 ± 1.5	1.9 ± 1.2	1.8 ± 1.4	1.5 ± 0.90
Selenium (mcg)	66 ± 32	84 ± 77	68 ± 38	71 ± 33
Zinc (mg)	4.4 ± 2.9	4.5 ± 2.5	3.9 ± 1.9	4.7 ± 2.3

Note: Values are means ± SD.

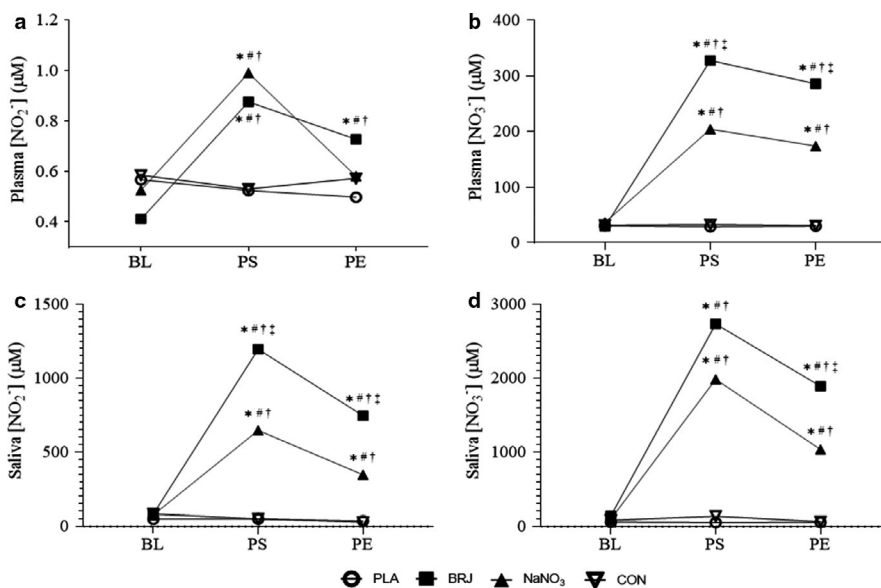


TABLE 2 Participant diet composition

FIGURE 2 Changes in Plasma/Saliva [NO<sub>3</sub><sup>-</sup>/NO<sub>2</sub><sup>-</sup>]. (a and B) Plasma measures. (c and d) Saliva measures. Baseline (BL), 2 hr postsupplementation (PS), and postexercise (PE). \*Significantly different from BL, #Significantly different from PLA, †Significantly different from CON, ‡Significantly different from NaNO<sub>3</sub>. ( $p < .05$ )

systolic and diastolic blood pressures were not different from baseline for any of the treatment arms (Table 3). Furthermore, there were no differences detected for other measures of vascular health (systolic/diastolic blood pressure, pulse rate, large and small artery elasticity) (Table 3). A significant main effect of treatment was observed for submaximal  $\dot{V}O_2$  ( $p = .003$ ) such that supplementation with BRJ resulted in a lower  $\dot{V}O_2$  compared to PLA ( $p = .009$ ), NaNO<sub>3</sub> ( $p = .042$ ), and CON ( $p = .009$ ) (Figure 3), which reflects a 7% mean improvement in submaximal exercise efficiency. A significant effect for treatment was observed for TTE ( $p \leq .001$ ), with BRJ eliciting a significantly greater improvement in TTE compared to PLA ( $p = .008$ ), NaNO<sub>3</sub> ( $p = .038$ ), and CON ( $p \leq .001$ ). A small but significant inverse association was observed for submaximal  $\dot{V}O_2$  and postsupplementation plasma  $\text{NO}_2^-$  ( $p = .04$ ) depicted in Table 4. No significant changes were observed for total antioxidant capacity (Figure 4), or other exercise or vascular related measures.

## 4 | DISCUSSION

The main purpose of this study was to determine the effects of acute supplementation with inorganic dietary  $\text{NO}_3^-$  in the form of BRJ, on exercise tolerance and cardiometabolic health among healthy men and women with obesity. Although recent studies have been similar in scope (Beals et al., 2017; Lara et al., 2015; Lima Bezerra et al., 2019; Rasica et al., 2018), to our knowledge, this is the first to report such findings in this population. The secondary purpose of this study was to explore the potential mechanisms by which these effects might occur by separating potentially salubrious components of BRJ (inorganic dietary nitrate and antioxidants) by treatment arm. In this study of 16 individuals with obesity we demonstrated that acute supplementation with inorganic dietary  $\text{NO}_3^-$  rich BRJ increased plasma and saliva  $\text{NO}_3^-$  and  $\text{NO}_2^-$ , decreased  $\dot{V}O_2$  during submaximal exercise, and extended time to exhaustion. These results are reflective of increased

**TABLE 3** Blood pressure, Pulse rate, and Elasticity indices in response to supplementation ( $n = 16$ )

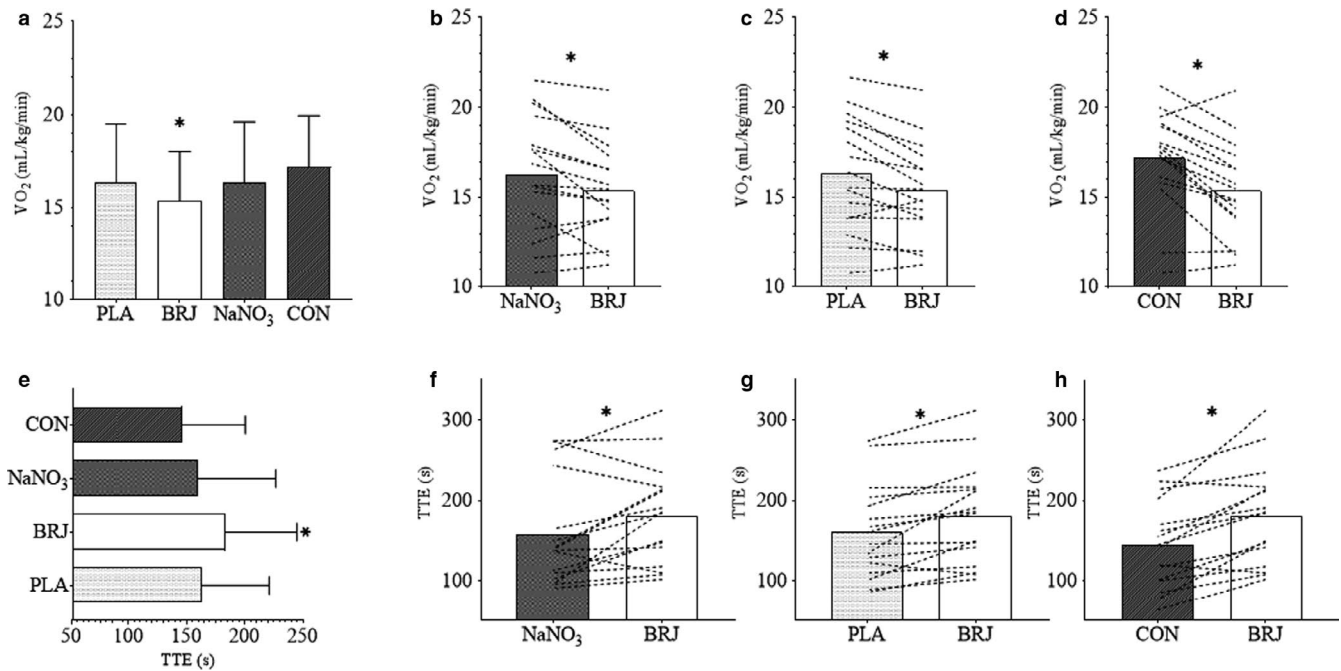
	PLA			BRJ			NaNO <sub>3</sub>			CON		
	Pre	Post	Δ	Pre	Post	Δ	Pre	Post	Δ	Pre	Post	Δ
SBP (mmHg)	121 ± 16	123 ± 14	2 ± 5	123 ± 15	121 ± 17	-2 ± 7	122 ± 17	122 ± 14	0 ± 6.5	124 ± 20	123 ± 17	-1 ± 12
DBP (mmHg)	68 ± 11	68 ± 9	0 ± 6	70 ± 11	69 ± 12	-1 ± 6	69 ± 10	69 ± 10	0 ± 4	70 ± 12	69 ± 11	-1.0 ± 5
PR (bpm)	69 ± 11	68 ± 11	-1 ± 6	71 ± 12	67 ± 11	-4 ± 4	71 ± 10	67 ± 9	-4 ± 5	70 ± 10	68 ± 10	-2 ± 5
LAE	16.0 ± 4.3	16.7 ± 5.2	0.7 ± 5.5	16.4 ± 3.2	16.2 ± 3.8	-0.2 ± 4.2	15.9 ± 4.1	16.8 ± 5.8	0.9 ± 6.3	16.0 ± 3.5	18.6 ± 6.6	2.6 ± 5.3
SAE	11.3 ± 4.5	10.7 ± 3.7	-0.6 ± 3.7	13.8 ± 13.8	10.6 ± 3.8	-3.2 ± 14.6	9.9 ± 3.3	10.9 ± 3.4	1.0 ± 3.3	11.9 ± 4.8	10.6 ± 3.1	-1.3 ± 5.1

Note: Values are means ± SD.

Abbreviations: LAE, large artery elasticity; SAE, small artery elasticity.

submaximal exercise efficiency, and improved exercise tolerance. Despite previous investigations demonstrating improvements in blood pressure (Berry et al., 2015; Eggebeen et al., 2016; Jajja et al., 2014; Lima Bezerra et al., 2019; Webb et al., 2008), we did not find such improvements in our study.

We observed comparable results with previous studies in that supplementing with NaNO<sub>3</sub> or NO<sub>3</sub><sup>-</sup> rich BRJ, increased both plasma and saliva NO<sub>3</sub><sup>-</sup>/NO<sub>2</sub><sup>-</sup> (Bailey, Fulford, et al., 1985; Bailey, Winyard, et al., 1985; Kelly, Vanhatalo, Wilkerson, Wylie, & Jones, 2013; Lansley, Winyard, Fulford, et al., 2011; Larsen, Weitzberg, Lundberg, & Ekblom, 2007). Our study, similar to others (Flueck, Bogdanova, Mettler, & Perret, 2016), further aimed to assess the difference in these responses between BRJ and a NaNO<sub>3</sub> solution matched for nitrate content. Both arms resulted in a significant rise in plasma and saliva NO<sub>3</sub><sup>-</sup>/NO<sub>2</sub><sup>-</sup> from baseline. With the exception of plasma NO<sub>2</sub><sup>-</sup>, the BRJ treatment elicited a significantly greater response compared to NaNO<sub>3</sub>. These results suggest that other components in concentrated BRJ enhance nitrate metabolism (absorption/transport) into the blood and saliva and support data presented by Jonvik et al who demonstrated that acute supplementation with nitrate-matched beetroot, rocket salad, and spinach beverages increased plasma NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> to a greater extent than NaNO<sub>3</sub> (Jonvik et al., 2016). BRJ contains the amino acid betaine, and polyphenols quercetin and resveratrol. Although there were no differences detected in antioxidant capacity from baseline to postsupplementation across study arms, it is possible there may be synergistic interactions between dietary NO<sub>3</sub><sup>-</sup> and these compounds that lead to improved ergogenic effects. Notably, the antioxidant compounds in BRJ have been linked to improvements in mitochondrial efficiency, increased aerobic capacity, improvements in endurance, strength, and power (Alway et al., 2017; Davis, Murphy, Carmichael, & Davis, 2009; Hoffman, Ratamess, Kang, Rashti, & Faigenbaum, 2009). This idea is supported by results from the present study showing greater improvement in exercise-related outcomes following BRJ consumption compared to NaNO<sub>3</sub> despite the putative endpoint, plasma NO<sub>2</sub><sup>-</sup> being similar. Indeed, this suggests the involvement of other factors which may contribute to these NO mediated outcomes. Previous studies assessing the effectiveness of exogenous antioxidant supplementation alone are widely inconclusive, with some suggesting improvement in performance and/or physiological markers associated with exercise (Clarkson & Thompson, 2000; Jakeman & Maxwell, 1993), and others reporting no effect (Gey, Cooper, & Bottenberg, 1970). Differences in training status, affecting endogenous antioxidant production, diet, along with time, type, and intensity of exercise has all been cited as possible reasons for the inconsistencies observed with previous antioxidant-alone investigations. Antioxidant compounds in BRJ (vitamin C and



**FIGURE 3** Exercise-related Outcomes in Response to Treatment. (a) Mean  $\text{VO}_2$  response during submaximal exercise. (b-d) Mean submaximal  $\text{VO}_2$  for BRJ compared to  $\text{NaNO}_3$ , PLA, and CON, respectively. (e) Mean time to exhaustion (TTE). (f-h) TTE for BRJ compared to  $\text{NaNO}_3$ , PLA, and CON, respectively. \*Significant difference between conditions. ( $p < .05$ )

**TABLE 4** Pearson Correlation Matrix for NO Metabolites and Exercise Outcomes

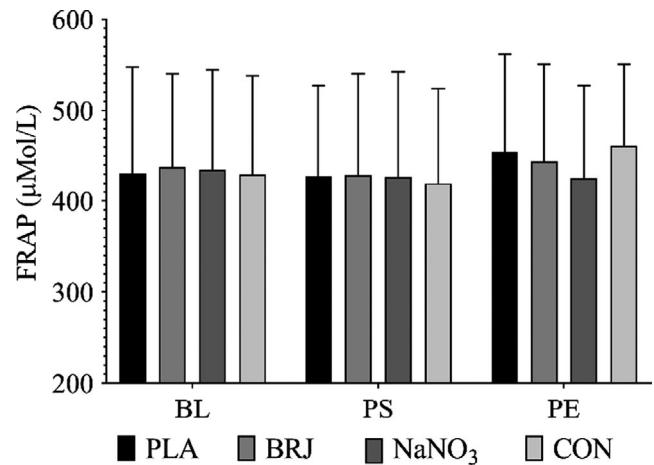
	$\text{PNO}_2^-$	$\text{PNO}_3^-$	$\text{SNO}_2^-$	$\text{SNO}_3^-$
Submax $\text{VO}_2$	<b>-0.36</b>	-0.17	<0.01	-0.07
Maximal $\text{VO}_2$	-0.11	0.11	0.19	0.18
TTE	0.051	0.09	0.17	0.14

Note: Bold values indicate  $p < .05$ .

P, plasma measures; S, saliva measures; TTE, time to exhaustion.

polyphenols) have been shown to facilitate  $\text{NO}_2^-$  reduction to NO in the gut (Peri et al., 2005; Rocha, Gago, Barbosa, & Laranjinha, 2009). A study by Ogawa *et al* found that the generation of NO from reduced  $\text{NO}_2^-$  was enhanced by high-phenolic beverages (Ogawa & Mochizuki, 2013). Therefore, the differential response may rise from more efficient conversion of  $\text{NO}_2^-$  to NO, catalyzed by phenolic compounds, vitamin C, and other antioxidants within BRJ. An interesting area of future research would be elucidating the independent and/or synergistic contributions these compounds may elicit, and further determine the dose/duration of supplementation necessary to detect optimal effects.

The majority of previous investigations have focused on the ergogenic potential of BRJ in athletic and healthy individuals, with many, but not all demonstrating improvement in performance time, submaximal  $\text{O}_2$  consumption, mitochondrial efficiency, and blood pressure (Cermak, Gibala, & van Loon, 2012; Domínguez et al., 2017; Lansley, Winyard,



**FIGURE 4** FRAP at baseline (BL), 2 hr postsupplementation (PS), and postexercise (PE). Values are expressed as  $\mu\text{M}$  ascorbate equivalents. No significant differences within or between treatments. ( $p < .05$ )

Bailey, et al., 2011; Larsen et al., 2007). Notably, effects of dietary nitrate are inconsistently observed in some disease settings such as type 2 diabetes (Beals et al., 2017; Gilchrist et al., 2013) underscoring the need to assess whether this pathway is functional in disease. Many individuals with obesity present with a reduced ability to participate in physical exercise characterized by reduced cardiorespiratory fitness and exercise intolerance (Salvadori et al., 1999; Tryon et al., 1992). Similar to results with healthy volunteer



(Bailey, Fulford, et al., 1985; Bailey, Winyard, et al., 1985; Lansley, Winyard, Fulford, et al., 2011; Larsen et al., 2007), we demonstrated that acute supplementation with BRJ improved  $\text{VO}_2$  during submaximal cycling by  $\sim 7\%$  over all other experimental arms in adults with obesity. It has been suggested that these improvements could in part be due to increased circulating  $\text{NO}_2^-$  which through a series of different mechanisms can undergo a one-electron reduction to NO (Lundberg & Weitzberg, 2009) This  $\text{NO}_2^-$  to NO reduction is partially regulated by declines in  $\text{O}_2$  tension and pH which mirrors increases in exercise intensity (Casey, Treichler, Ganger, Schneider, & Ueda, 2015; Gaebelein & Ladd, 1986) A study by Bailey *et al* attributed the reduced  $\text{O}_2$  cost of submaximal, moderate-intensity exercise to a reduction in ATP cost of force production (Bailey, Fulford, et al., 1985) This was further capitulated by Larsen *et al* showing lower  $\text{O}_2$  demand during submaximal work in healthy trained men (Larsen et al., 2007). Other potential mechanisms include improvements in mitochondrial and  $\text{Ca}^{2+}$  handling proteins that may improve metabolic and contractile function (Larsen et al., 2011). Obesity is associated with a higher  $\text{O}_2$  cost for submaximal exercise (Salvadori et al., 1999) and reduced  $\text{VO}_{2\text{peak}}$  (Green, O'Connor, Kiely, O'Shea, & Egana, 2018) which may require increased work and metabolic stress when participating in activities of daily living. The results from this study and others, suggest that acute  $\text{NO}_3^-$  supplementation decreases the cost of submaximal work. Future studies should more closely examine the effects of both acute and chronic supplementation on functional capacity during lower intensity activities of daily living in individuals with obesity, perhaps improving functional outcomes and quality of life.

We found that BRJ improved exercise tolerance during high-intensity cycling by  $\sim 15\%$  over all other treatment arms with as high as a 22% improvement when compared to CON. In addition to the aforementioned reasons, improvements in high-intensity exercise following BRJ supplementation is consistently attributed to alterations in  $\text{VO}_2$  kinetics, affecting finite substrate utilization and availability during exercise (i.e., rate of PCr breakdown and ATP generation from anaerobic glycolysis—metabolites of which are implicated in skeletal muscle fatigue) (Bailey, Varnham, et al., 1985; Breese et al., 2013; Murgatroyd, Ferguson, Ward, Whipp, & Rossiter, 1985). Although we did not measure  $\text{VO}_2$  kinetics in this study, Bailey et al and others have shown that BRJ supplementation speeds phase II  $\text{VO}_2$  kinetics during high-intensity exercise, possibly linked to enhanced muscle vasodilation, blood flow, and  $\text{O}_2$  supply (Bailey, Varnham, et al., 1985). This is perhaps most important in type II muscle fibers where  $\text{O}_2$  delivery often lags behind  $\text{O}_2$  demand ultimately leading to oxidative inefficiency and fatigue during severe-intensity exercise (Grassi, Rossiter, & Zoladz, 2015; McDonough, Behnke, Padilla, Musch, & Poole, 2005). Indeed, a similar study by Rasica *et al* illustrated a reduced rate of  $\text{VO}_2$

increase during severe-intensity exercise with BRJ supplementation, in adolescents with obesity, suggesting improved efficiency (Rasica et al., 2018). The observed improvement in exercise tolerance with BRJ during high-intensity exercise is noteworthy considering that high-intensity interval training has been shown to be a highly effective intervention for improvement in weight and health status in individuals with obesity (Fisher et al., 2015; Gillen et al., 2016).

Strengths of this study include its design, utilizing a partially blinded, four condition, crossover design including a BRJ supplement, a placebo supplement with negligible  $\text{NO}_3^-$  content, a  $\text{NaNO}_3$  solution matched for BRJ  $\text{NO}_3^-$  content, and a no treatment control. Many previous investigations have speculated other bioactive compounds within BRJ may act synergistically with  $\text{NO}_3^-$ . By including a  $\text{NO}_3^-$  matched  $\text{NaNO}_3$  treatment in our study, we were able to examine potentially mechanistic differences in response between BRJ, PLA, and  $\text{NaNO}_3$  with our results indeed suggesting the presence of a synergistic effect from BRJ. Weaknesses in our study include the inability to blind participants to treatment ( $\text{NaNO}_3$  treatment and CON), a small sample size, and racial homogeneity, making it difficult to generalize our findings. However, only 2 of our 16 participants failed to significantly improve their TTE when supplemented with BRJ compared to CON.

In conclusion, in individuals with obesity, acute supplementation with inorganic dietary nitrate-rich BRJ, significantly increased  $[\text{NO}_3^-]$  and  $[\text{NO}_2^-]$  measured in both saliva and plasma. Additionally,  $\text{VO}_2$  during submaximal exercise was decreased by  $\sim 7\%$ , and perhaps most notable was a  $\sim 22\%$  improvement in time to exhaustion observed in the BRJ condition over control. These results taken together suggest that acute supplementation with BRJ is an effective strategy for improving exercise tolerance and efficiency in individuals with obesity. With reduced participation in physical exercise being hallmark of those with obesity, perhaps in part due to insufficient or impaired NO bioavailability, regular supplementation with BRJ may improve NO status and in turn improve exercise-related outcomes in individuals with obesity. Regular physical exercise has been implicated in the long-term success of weight loss maintenance. Thus, identifying novel strategies to improve exercise participation among individuals with obesity is of critical importance. Outcomes from this study suggest that BRJ, may be an easily obtained, cost-effective nutraceutical to assist easing the burden of exercise and perhaps improve participation in and adherence to exercise.

## ACKNOWLEDGMENTS

This work was supported by a School of Education Faculty Development Grant awarded to Gordon Fisher. Christian Behrens is supported by a NIH T-32 Grant (T32HL105349) from the National Heart Lung and Blood Institute. Additional

support provided by Award Number P30DK056336 – National Institute of Diabetes Digestive and Kidney Diseases. The authors thank the staff of the Clinical Research Unit at The University of Alabama at Birmingham, along with Wesley Calhoun, Mary Kopp, Sydney Bell, Rima Patel, and Emily Stuckey for their assistance in data acquisition.

## CONFLICT OF INTEREST

The authors have no conflict of interest.

## AUTHOR CONTRIBUTION

GF, CB, RP, KR, BL, and KA performed the experiments, GF, CB, KA, KR, and JF analyzed data and interpreted results of experiments. CB drafted manuscript. GF, RP, and JF edited and revised manuscript. GF, CB, and RP conceptualized and designed the experiments. All authors approved final version of manuscript.

## ETHICAL STATEMENT

The protocol for this study was reviewed and approved by the institutional review board at the University of Alabama at Birmingham. Informed consent was obtained from all study participants.

## ORCID

Gordon Fisher  <https://orcid.org/0000-0002-9191-3774>

## REFERENCES

- Alway, S. E., McCrory, J. L., Kearcher, K., Vickers, A., Frear, B., Gilleland, D. L., ... Mohamed, J. S. (2017). Resveratrol enhances exercise-induced cellular and functional adaptations of skeletal muscle in older men and women. *The Journals of Gerontology: Series A*, *72*, 1595–1606. <https://doi.org/10.1093/gerona/glx089>
- Amisola, R. V., & Jacobson, M. S. (2003). Physical activity, exercise, and sedentary activity: Relationship to the causes and treatment of obesity. *Adolescent Medicine (Philadelphia, PA)*, *14*, 23–35.
- Bailey, S. J., Fulford, J., Vanhatalo, A., Winyard, P. G., Blackwell, J. R., DiMenna, F. J., ... Jones, A. M. (1985). Dietary nitrate supplementation enhances muscle contractile efficiency during knee-extensor exercise in humans. *Journal of Applied Physiology (Bethesda, MD)*, *109*(1), 135–148. <https://doi.org/10.1152/jappphysiol.00046.2010>
- Bailey, S. J., Varnham, R. L., DiMenna, F. J., Breese, B. C., Wylie, L. J., & Jones, A. M. (1985). Inorganic nitrate supplementation improves muscle oxygenation, O<sub>2</sub> uptake kinetics, and exercise tolerance at high but not low pedal rates. *Journal of Applied Physiology (Bethesda, MD)*, *118*:1396–1405, 2015.
- Bailey, S. J., Winyard, P., Vanhatalo, A., Blackwell, J. R., Dimenna, F. J., Wilkerson, D. P., ... Jones, A. M. (1985). Dietary nitrate supplementation reduces the O<sub>2</sub> cost of low-intensity exercise and enhances tolerance to high-intensity exercise in humans. *Journal of Applied Physiology (Bethesda, MD)*, *107*:1144–1155, 2009.
- Beals, J. W., Binns, S. E., Davis, J. L., Giordano, G. R., Klochak, A. L., Paris, H. L., ... Bell, C. (2017). Concurrent Beet Juice and Carbohydrate Ingestion: Influence on Glucose Tolerance in Obese and Nonobese Adults. *Journal of Nutrition and Metabolism*, *2017*, 6436783.
- Benzie, I. F., & Strain, J. J. (1996). The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": The FRAP assay. *Analytical Biochemistry*, *239*, 70–76.
- Berry, M. J., Justus, N. W., Hauser, J. I., Case, A. H., Helms, C. C., Basu, S., ... Miller, G. D. (2015). Dietary nitrate supplementation improves exercise performance and decreases blood pressure in COPD patients. *Nitric Oxide: Biology and Chemistry*, *48*, 22–30. <https://doi.org/10.1016/j.niox.2014.10.007>
- Bournat, J. C., & Brown, C. W. (2010). Mitochondrial dysfunction in obesity. *Current Opinion in Endocrinology, Diabetes, and Obesity*, *17*, 446–452. <https://doi.org/10.1097/MED.0b013e32833c3026>
- Breese, B. C., McNarry, M. A., Marwood, S., Blackwell, J. R., Bailey, S. J., & Jones, A. M. (2013). Beetroot juice supplementation speeds O<sub>2</sub> uptake kinetics and improves exercise tolerance during severe-intensity exercise initiated from an elevated metabolic rate. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology*, *305*, R1441–R1450.
- Casey, D. P., Treichler, D. P., Ganger, C. T., Schneider, A. C., & Ueda, K. (2015). Acute dietary nitrate supplementation enhances compensatory vasodilation during hypoxic exercise in older adults. *Journal of Applied Physiology*, *118*, 178–186. <https://doi.org/10.1152/jappphysiol.00662.2014>
- Cermak, N. M., Gibala, M. J., & van Loon, L. J. (2012). Nitrate supplementation's improvement of 10-km time-trial performance in trained cyclists. *International Journal of Sport Nutrition and Exercise Metabolism*, *22*, 64–71. <https://doi.org/10.1123/ijnsnem.22.1.64>
- Clarkson, P. M., & Thompson, H. S. (2000). Antioxidants: What role do they play in physical activity and health? *The American Journal of Clinical Nutrition*, *72*, 637s–646s. <https://doi.org/10.1093/ajcn/72.2.637S>
- Clements, W. T., Lee, S. R., & Bloomer, R. J. (2014). Nitrate ingestion: A review of the health and physical performance effects. *Nutrients*, *6*, 5224–5264.
- Clifford, T., Howatson, G., West, D. J., & Stevenson, E. J. (2015). The potential benefits of red beetroot supplementation in health and disease. *Nutrients*, *7*, 2801–2822. <https://doi.org/10.3390/nu7042801>
- Coggan, A. R., Broadstreet, S. R., Mahmood, K., Mikhalkova, D., Madigan, M., Bole, I., ... Peterson, L. R. (2018). Dietary nitrate increases VO<sub>2peak</sub> and performance but does not alter ventilation or efficiency in patients with heart failure with reduced ejection fraction. *Journal of Cardiac Failure*, *24*, 65–73. <https://doi.org/10.1016/j.cardfail.2017.09.004>
- Cohn, J. N., Finkelstein, S., McVeigh, G., Morgan, D., LeMay, L., Robinson, J., & Mock, J. (1995). Noninvasive pulse wave analysis for the early detection of vascular disease. *Hypertension*, *26*, 503–508. <https://doi.org/10.1161/01.HYP.26.3.503>
- Davis, J. M., Murphy, E. A., Carmichael, M. D., & Davis, B. (2009). Quercetin increases brain and muscle mitochondrial biogenesis and exercise tolerance. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology*, *296*, R1071–R1077. <https://doi.org/10.1152/ajpregu.90925.2008>
- de Lima Bezerra, Á. D., Costa, E. C., Pacheco, D. A., Souza, D. C., Farias-Junior, L. F., Ritti-Dia, R. M., ... Fayh, A. P. T. (2019). Effect of acute dietary nitrate supplementation on the post-exercise ambulatory blood pressure in obese males: A randomized, controlled, crossover trial. *Journal of Sports Science and Medicine*, *18*, 118–127.
- Domínguez, R., Cuenca, E., Maté-Muñoz, J., García-Fernández, P., Serra-Paya, N., Estevan, M., ... Garnacho-Castaño, M. (2017). Effects of beetroot juice supplementation on cardiorespiratory

- endurance in athletes. A systematic review. *Nutrients*, *9*(1), 43. <https://doi.org/10.3390/nu9010043>
- Ergebeben, J., Kim-Shapiro, D. B., Haykowsky, M., Morgan, T. M., Basu, S., Brubaker, P., ... Kitzman, D. W. (2016). One week of daily dosing with beetroot juice improves submaximal endurance and blood pressure in older patients with heart failure and preserved ejection fraction. *JACC Heart Failure*, *4*, 428–437.
- Erzurum, S. C., Ghosh, S., Janocha, A. J., Xu, W., Bauer, S., Bryan, N. S., ... Beall, C. M. (2007). Higher blood flow and circulating NO products offset high-altitude hypoxia among Tibetans. *Proceedings of the National Academy of Sciences of the United States of America*, *104*, 17593–17598. <https://doi.org/10.1073/pnas.0707462104>
- Fisher, G., Brown, A. W., Bohan Brown, M. M., Alcorn, A., Noles, C., Winwood, L., ... Allison, D. B. (2015). High intensity interval- vs moderate intensity- training for improving cardiometabolic health in overweight or obese males: A randomized controlled trial. *PLoS One*, *10*, e0138853.
- Flueck, J. L., Bogdanova, A., Mettler, S., & Perret, C. (2016). Is beetroot juice more effective than sodium nitrate? The effects of equimolar nitrate dosages of nitrate-rich beetroot juice and sodium nitrate on oxygen consumption during exercise. *Applied Physiology, Nutrition and Metabolism*, *41*, 421–429. <https://doi.org/10.1139/apnm-2015-0458>
- Gaebelein, C. J., & Ladd, C. M. (1986). Blood flow, PO<sub>2</sub>, PCO<sub>2</sub> and pH during progressive working contractions in a whole muscle group. *European Journal of Applied Physiology and Occupational Physiology*, *54*, 638–642.
- Georgiev, V. G., Weber, J., Kneschke, E. M., Denev, P. N., Bley, T., & Pavlov, A. I. (2010). Antioxidant activity and phenolic content of betalain extracts from intact plants and hairy root cultures of the red beetroot *Beta vulgaris* cv. Detroit dark red. *Plant Foods for Human Nutrition (Dordrecht Netherlands)*, *65*, 105–111. <https://doi.org/10.1007/s11130-010-0156-6>
- Gey, G. O., Cooper, K. H., & Bottenberg, R. A. (1970). Effect of ascorbic acid on endurance performance and athletic injury. *JAMA*, *211*, 105. <https://doi.org/10.1001/jama.211.1.105>
- Gilchrist, M., Winyard, P. G., Aizawa, K., Anning, C., Shore, A., & Benjamin, N. (2013). Effect of dietary nitrate on blood pressure, endothelial function, and insulin sensitivity in type 2 diabetes. *Free Radical Biology & Medicine*, *60*, 89–97. <https://doi.org/10.1016/j.freeradbiomed.2013.01.024>
- Gillen, J. B., Martin, B. J., MacInnis, M. J., Skelly, L. E., Tarnopolsky, M. A., & Gibala, M. J. (2016). Twelve weeks of sprint interval training improves indices of cardiometabolic health similar to traditional endurance training despite a five-fold lower exercise volume and time commitment. *PLoS One*, *11*, e0154075. <https://doi.org/10.1371/journal.pone.0154075>
- Grassi, B., Rossiter, H. B., & Zoladz, J. A. (2015). Skeletal muscle fatigue and decreased efficiency: Two sides of the same coin? *Exercise and Sport Sciences Reviews*, *43*, 75–83. <https://doi.org/10.1249/JES.0000000000000043>
- Green, S., O'Connor, E., Kiely, C., O'Shea, D., & Egana, M. (2018). Effect of obesity on oxygen uptake and cardiovascular dynamics during whole-body and leg exercise in adult males and females. *Physiological Reports*, *6*, e13705. <https://doi.org/10.14814/phy2.13705>
- Hales, S. (1733). *Statistical Essays Vol II. Haemostatics. Innings and Manby, London.*
- Hoffman, J. R., Ratamess, N. A., Kang, J., Rashti, S. L., & Faigenbaum, A. D. (2009). Effect of betaine supplementation on power performance and fatigue. *Journal of the International Society of Sports Nutrition*, *6*, 7. <https://doi.org/10.1186/1550-2783-6-7>
- Jajja, A., Sutyarjoko, A., Lara, J., Rennie, K., Brandt, K., Qadir, O., & Siervo, M. (2014). Beetroot supplementation lowers daily systolic blood pressure in older, overweight subjects. *Nutrition Research (New York, NY)*, *34*, 868–875. <https://doi.org/10.1016/j.nutres.2014.09.007>
- Jakeman, P., & Maxwell, S. (1993). Effect of antioxidant vitamin supplementation on muscle function after eccentric exercise. *European Journal of Applied Physiology and Occupational Physiology*, *67*, 426–430. <https://doi.org/10.1007/BF00376459>
- Jones, A. M., Thompson, C., Wylie, L. J., & Vanhatalo, A. (2018). Dietary nitrate and physical performance. *Annual Review of Nutrition*, *38*, 303–328. <https://doi.org/10.1146/annurev-nutr-082117-051622>
- Jonvik, K. L., Nyakayiru, J., Pinckaers, P. J., Senden, J. M., van Loon, L. J., & Verdijk, L. B. (2016). Nitrate-rich vegetables increase plasma nitrate and nitrite concentrations and lower blood pressure in healthy adults. *The Journal of Nutrition*, *146*, 986–993. <https://doi.org/10.3945/jn.116.229807>
- Joshiyura, K. J., Munoz-Torres, F. J., Morou-Bermudez, E., & Patel, R. P. (2017). Over-the-counter mouthwash use and risk of pre-diabetes/diabetes. *Nitric Oxide: Biology and Chemistry*, *71*, 14–20. <https://doi.org/10.1016/j.niox.2017.09.004>
- Kanner, J., Harel, S., & Granit, R. (2001). Betalains—a new class of dietary cationized antioxidants. *Journal of Agricultural and Food Chemistry*, *49*, 5178–5185. <https://doi.org/10.1021/jf010456f>
- Kelly, J., Vanhatalo, A., Wilkerson, D. P., Wylie, L. J., & Jones, A. M. (2013). Effects of nitrate on the power-duration relationship for severe-intensity exercise. *Medicine and Science in Sports and Exercise*, *45*, 1798–1806. <https://doi.org/10.1249/MSS.0b013e31828e885c>
- Ladabaum, U., Mannalithara, A., Myer, P. A., & Singh, G. (2014). Obesity, abdominal obesity, physical activity, and caloric intake in US adults: 1988 to 2010. *The American Journal of Medicine*, *127*, 717–727. <https://doi.org/10.1016/j.amj.2014.07.012>
- Lansley, K. E., Winyard, P. G., Bailey, S. J., Vanhatalo, A., Wilkerson, D. P., Blackwell, J. R., ... Jones, A. M. (2011). Acute dietary nitrate supplementation improves cycling time trial performance. *Medicine and Science in Sports and Exercise*, *43*, 1125–1131. <https://doi.org/10.1249/MSS.0b013e31821597b4>
- Lansley, K. E., Winyard, P. G., Fulford, J., Vanhatalo, A., Bailey, S. J., Blackwell, J. R., ... Jones, A. M. (2011). Dietary nitrate supplementation reduces the O<sub>2</sub> cost of walking and running: A placebo-controlled study. *Journal of Applied Physiology*, *110*, 591–600.
- Lara, J., Ogbonmwan, I., Oggioni, C., Zheng, D., Qadir, O., Ashor, A., ... Siervo, M. (2015). Effects of handgrip exercise or inorganic nitrate supplementation on 24-h ambulatory blood pressure and peripheral arterial function in overweight and obese middle age and older adults: A pilot RCT. *Maturitas*, *82*, 228–235.
- Larsen, F. J., Schiffer, T. A., Borniquel, S., Sahlin, K., Ekblom, B., Lundberg, J. O., & Weitzberg, E. (2011). Dietary inorganic nitrate improves mitochondrial efficiency in humans. *Cell Metabolism*, *13*, 149–159. <https://doi.org/10.1016/j.cmet.2011.01.004>
- Larsen, F. J., Weitzberg, E., Lundberg, J. O., & Ekblom, B. (2007). Effects of dietary nitrate on oxygen cost during exercise. *Acta Physiologica*, *191*, 59–66. <https://doi.org/10.1111/j.1748-1716.2007.01713.x>
- Lee-Young, R. S., Ayala, J. E., Hunley, C. F., James, F. D., Bracy, D. P., Kang, L., & Wasserman, D. H. (2010). Endothelial nitric oxide synthase is central to skeletal muscle metabolic regulation and



- enzymatic signaling during exercise in vivo. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology*, 298, R1399–R1408. <https://doi.org/10.1152/ajpregu.00004.2010>
- Lundberg, J. O., & Govoni, M. (2004). Inorganic nitrate is a possible source for systemic generation of nitric oxide. *Free Radical Biology and Medicine*, 37, 395–400. <https://doi.org/10.1016/j.freeradbiomed.2004.04.027>
- Lundberg, J. O., & Weitzberg, E. (2009). NO generation from inorganic nitrate and nitrite: Role in physiology, nutrition and therapeutics. *Archives of Pharmacological Research*, 32, 1119–1126.
- Martínez-González, M. Á., Alfredo Martínez, J., Hu, F. B., Gibney, M. J., & Kearney, J. (1999). Physical inactivity, sedentary lifestyle and obesity in the European Union. *International Journal of Obesity*, 23, 1192–1201. <https://doi.org/10.1038/sj.ijo.0801049>
- McDonough, P., Behnke, B. J., Padilla, D. J., Musch, T. I., & Poole, D. C. (2005). Control of microvascular oxygen pressures in rat muscles comprised of different fibre types. *The Journal of Physiology*, 563, 903–913. <https://doi.org/10.1113/jphysiol.2004.079533>
- McInnis, K. J. (2000). Exercise and obesity. *Coronary Artery Disease*, 11, 111–116. <https://doi.org/10.1097/00019501-200003000-00004>
- Murgatroyd, S. R., Ferguson, C., Ward, S. A., Whipp, B. J., & Rossiter, H. B. (1985). Pulmonary O<sub>2</sub> uptake kinetics as a determinant of high-intensity exercise tolerance in humans. *Journal of Applied Physiology (Bethesda, MD)*, 110, 1598–1606, 2011.
- Ogawa, T., & Mochizuki, S. (2013). P20: Evaluation of generation of nitric oxide by reducing nitrite with antioxidants in drinks. *Nitric Oxide: Biology and Chemistry*, 31, S21.
- Peri, L., Pietraforte, D., Scorza, G., Napolitano, A., Fogliano, V., & Minetti, M. (2005). Apples increase nitric oxide production by human saliva at the acidic pH of the stomach: A new biological function for polyphenols with a catechol group? *Free Radical Biology & Medicine*, 39, 668–681. <https://doi.org/10.1016/j.freeradbiomed.2005.04.021>
- Pohl, U., & Lamontagne, D. (1991). Impaired tissue perfusion after inhibition of endothelium-derived nitric oxide. *Basic Research in Cardiology*, 86(Suppl 2), 97–105.
- Rasica, L., Porcelli, S., Marzorati, M., Salvadego, D., Vezzoli, A., Agosti, F., ... Grassi, B. (2018). Ergogenic effects of beetroot juice supplementation during severe-intensity exercise in obese adolescents. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology*, 315, R453–r460. <https://doi.org/10.1152/ajpregu.00017.2018>
- Reddy, M. K., Alexander-Lindo, R. L., & Nair, M. G. (2005). Relative inhibition of lipid peroxidation, cyclooxygenase enzymes, and human tumor cell proliferation by natural food colors. *Journal of Agricultural and Food Chemistry*, 53, 9268–9273. <https://doi.org/10.1021/jf051399j>
- Rocha, B. S., Gago, B., Barbosa, R. M., & Laranjinha, J. (2009). Dietary polyphenols generate nitric oxide from nitrite in the stomach and induce smooth muscle relaxation. *Toxicology*, 265, 41–48. <https://doi.org/10.1016/j.tox.2009.09.008>
- Salvadori, A., Fanari, P., Fontana, M., Buontempi, L., Saezza, A., Baudo, S., ... Longhini, E. (1999). Oxygen uptake and cardiac performance in obese and normal subjects during exercise. *Respiration*, 66, 25–33. <https://doi.org/10.1159/000029333>
- Sansbury, B. E., & Hill, B. G. (2014). Chapter thirteen - antiobesogenic role of endothelial nitric oxide synthase. In: *Vitamins & Hormones* (p. 323–346), edited by Litwack G Academic Press.
- Sansbury, B. E., & Hill, B. G. (2014). Regulation of obesity and insulin resistance by nitric oxide. *Free Radical Biology and Medicine*, 73, 383–399. <https://doi.org/10.1016/j.freeradbiomed.2014.05.016>
- Schneider, D. A., Phillips, S. E., & Stoffolano, S. (1993). The simplified V-slope method of detecting the gas exchange threshold. *Medicine and Science in Sports and Exercise*, 25, 1180–1184. <https://doi.org/10.1249/00005768-199310000-00015>
- Shaw, K. A., Gennat, H. C., O'Rourke, P., & Del Mar, C. (2006). Exercise for overweight or obesity. *Cochrane Database of Systematic Reviews*. <https://doi.org/10.1002/14651858.CD003817.pub3>
- Siervo, M., Jackson, S. J., & Bluck, L. J. (2011). In-vivo nitric oxide synthesis is reduced in obese patients with metabolic syndrome: Application of a novel stable isotopic method. *Journal of Hypertension*, 29, 1515–1527.
- Stamler, J. S., & Meissner, G. (2001). Physiology of nitric oxide in skeletal muscle. *Physiological Reviews*, 81, 209–237. <https://doi.org/10.1152/physrev.2001.81.1.209>
- Tong, L., Heim, R. A., & Wu, S. (2011). Nitric oxide: A regulator of eukaryotic initiation factor 2 kinases. *Free Radical Biology & Medicine*, 50, 1717–1725.
- Tremmel, M., Gerdtham, U. G., Nilsson, P. M., & Saha, S. (2017). Economic burden of obesity: A systematic literature review. *International Journal of Environmental Research and Public Health*, 14. <https://doi.org/10.3390/ijerph14040435>
- Tryon, W. W., Goldberg, J. L., & Morrison, D. F. (1992). Activity decreases as percentage overweight increases. *International Journal of Obesity and Related Metabolic Disorders Journal of the International Association for the Study of Obesity*, 16, 591–595.
- Vulić, J. J., Čebović, T. N., Čanadanović-Brunet, J. M., Četković, G. S., Čanadanović, V. M., Djilas, S. M., & Tumbas Šaponjac, V. T. (2014). In vivo and in vitro antioxidant effects of beetroot pomace extracts. *Journal of Functional Foods*, 6, 168–175. <https://doi.org/10.1016/j.jff.2013.10.003>
- Webb, A. J., Patel, N., Loukogeorgakis, S., Okorie, M., Aboud, Z., Misra, S., ... Ahluwalia, A. (2008). Acute blood pressure lowering, vasoprotective, and antiplatelet properties of dietary nitrate via bioconversion to nitrite. *Hypertension*, 51, 784–790. <https://doi.org/10.1161/HYPERTENSIONAHA.107.103523>
- Williams, I. L., Wheatcroft, S. B., Shah, A. M., & Kearney, M. T. (2002). Obesity, atherosclerosis and the vascular endothelium: Mechanisms of reduced nitric oxide bioavailability in obese humans. *International Journal of Obesity*, 26, 754–764.
- Wolf, A. M., & Colditz, G. A. (1998). current estimates of the economic cost of obesity in the United States. *Obesity Research*, 6, 97–106. <https://doi.org/10.1002/j.1550-8528.1998.tb00322.x>
- Zielinska-Przyjemska, M., Olejnik, A., Dobrowolska-Zachwieja, A., & Grajek, W. (2009). In vitro effects of beetroot juice and chips on oxidative metabolism and apoptosis in neutrophils from obese individuals. *Phytotherapy Research: PTR*, 23, 49–55.

**How to cite this article:** Behrens CE Jr , Ahmed K, Ricart K, et al. Acute beetroot juice supplementation improves exercise tolerance and cycling efficiency in adults with obesity. *Physiol Rep*. 2020;8:e14574. <https://doi.org/10.14814/phy2.14574>