BRIEF COMMUNICATION



Coronavirus Persistence on a Plastic Carrier Under Refrigeration Conditions and Its Reduction Using Wet Wiping Technique, with Respect to Food Safety

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Abstract

The global SARS-CoV-2 pandemic dictates that anti-contagion strategies should become matters of essential routine in everyday life. Fomite transference is one of the routes of transmission that has been considered for this virus. However, the risks associated with contaminated surfaces of food packaging kept in refrigerators have not yet been adequately assessed. In this study, a surrogate virus, *Alphacoronavirus 1*, was used to investigate the persistence of coronavirus dried on a plastic carrier at 4 °C. Techniques of wet wiping, with or without disinfectant saturation, were employed to evaluate their effectiveness in the elimination of the virus. If not wiped, the loss of infectivity of the virus on plastic surfaces was, on average, 0.93 \log_{10} (i.e. 83%) per day of storage at 4 °C. Wiping with water-saturated material reduced the initial virus titre on the plastic carrier rier by 2.4 \log_{10} (99.6%); the same results were achieved through wiping with bactericidal wipes containing ethanol. Wipes saturated with a combination of disinfectant agents (didecyl-dimethyl-ammonium chloride, hydrogen peroxide) decreased the virus titre still more efficiently, by 3.8 \log_{10} (99.98%) and also significantly prevented further transfer of the virus to a secondary surface through wiping. Thus SARS-CoV-2 transmission potential via contaminated plastic packaging and food may be efficiently eliminated by wet-wiping, especially when wipes saturated with specific disinfectants are used.

Keywords Covid-19 · Alphacoronavirus 1 · Surface contamination · Food safety · Virus survival · Refrigerated food

Introduction

COVID-19 is an infectious respiratory disease caused by the recently discovered SARS-CoV-2 coronavirus. It has now reached pandemic proportions worldwide. SARS-CoV-2 spreads primarily through respiratory secretions, droplets of which may lead to infection through inhalation or via contaminated hands touching the mucosal membranes of the nose, mouth or eyes (Miller and Englund 2020). Fomite transmission probably plays a significant role in indirect infection and transmission of SARS-CoV-2 (Fiorillo et al. 2020).

There is only limited evidence that virus particles on food products or food packages lead to the disease (Seymour et al.

Hana Malenovská malenovska@vri.cz 2020). However, refrigeration and the plastic materials that make up most packaging provide ideal conditions for the preservation of coronavirus infectivity, since the virus can survive for days in such an environment (Geller et al. 2012; Desai and Aronoff 2020; Eslami and Jalili 2020; Ren et al. 2020). Frequent handling of goods by manufacturers, suppliers, store staff and customers further increases the risk of contamination. Recently, a local outbreak of the SARS-CoV-2 from a food store was reported by a press release from the Ministry of Health of the Czech Republic (2020). The outbreak of the virus originated with an infected store employee. Even though all staff and customers were wearing masks when working and shopping, eleven customers (including one who even wore gloves in the shop and bought only a single tub of yogurt) became infected. This suggests that COVID-19 infection due to contaminated food packages may be rare, but it is possible. The Centre of Disease Control and Prevention (CDC) considers COVID-19 infection from handling contaminated food packages low-risk, but nonetheless recommends cleaning and disinfection (Seymour et al. 2020).

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The aim of this study was to investigate coronavirus stability under refrigeration conditions on a plastic surface simulating food packaging and to evaluate the efficiency of a simple method of virus elimination: wet wiping that can be applied in any household. Alphacoronavirus 1 (Fig. 1) was used as a surrogate for SARS-CoV-2. It has previously been employed as a surrogate for SARS-CoV (Casanova et al. 2009; Goyal et al. 2014). In general, members of the same virus family are inactivated similarly under the same physical and chemical stresses (Malenovska 2014), although some differences even between closely related viruses may exist. Nevertheless, Alphacoronavirus 1 and SARS-CoV have been shown to have similar persistence on surfaces (Fiorillo et al. 2020; Kampf et al. 2020) and surface stability of SARS-CoV is markedly similar to SARS-CoV-2 (van Doremalen et al. 2020). Apart from its intrinsically lower risk factor with regard to human health,



Fig. 1 TEM photograph of viral particles of *Alphacoronavirus 1*, strain M -42 (CAPM V-126) used in this study as a surrogate for SARS-CoV-2. Virus suspension was applied to a formvar-carbon-coated 400-mrsh copper grid, stained with 1% ammonium molyb-date for 1 min and viewed in a Philips EM 208 transmission electron microscope

the suitability of *Alphacoronavirus 1* for infectivity experiments consists in its replication to high titres. In contrast, SARS-CoV-2 cultivated in VERO cells achieves only low titres, placing limitations on such experiments (Keil et al. 2020; Sarkale et al. 2020).

Materials and Methods

Viruses and Cells

The *Alphacoronavirus 1* (strain M 42, CAPM V-126) was obtained from the Collection of Animal Pathogenic Microorganisms (CAPM), incorporated into the National Programme of Protection of Genetic Resources of Economically Significant Microorganisms and Animals of the Czech Republic. The virus was propagated in a PD 5 cell-line obtained from the Centro Substrati Cellulari Laboratory (Brescia, Italy). The cell culture with the virus was cultivated at 37 °C in Dulbecco's modified Eagle's medium (Biosera, Nuaille, France) supplemented with 3% heat-inactivated foetal calf serum (HyClone, South Logan, Utah, USA) until complete cytopathology. The virus was then aliquot-divided and frozen at - 80 °C.

Wipes

All wipes used in the experiments were purchased as hygiene items routinely available in the Czech Republic and were chosen to represent the types of wipes in widespread household use. Three different products were used for the experiments (Table 1): (i) wet wipes containing 99% water; (ii) wet hygiene wipes containing ethanol (designed for hand and surface hygiene), and (iii) wet wipes for surface disinfection containing didecyl-dimethyl-ammonium chloride and hydrogen peroxide.

Table 1 Wipes used for evaluation of efficacy of removal of SARS-CoV-2 surrogate Alphacoronavirus 1 from a plastic carrier

Product	Wet wipes Babydream	Hygiene wipes Balea	Disinfectant wipes SAVO
Substances	99% water, caprylyl/capryl glucoside, cit- ric acid, sodium citrate, sodium benzoate	Water, ethanol (0.6 g/wipe)*, glycerine, <i>Aloe barbadensis</i> leaf extract, chlorhex- idine digluconate	0.75% didecyl-dimethyl-ammonium chloride, 0.5% hydrogen peroxide, less than 5% non-ionic surface active agent, cationic surface active agent, bleach- ing agent based on oxygen, perfume, limonene, iodopropynyl butylcarbamate
Producer	Produced for Dirk Rossmann GmbH, Burgwedel, Germany	DM-Drogerie Markt, Karlsruhe, Germany	Unilever ČR, Czech Republic

*16.5% ethanol solution (concentration was calculated based on a comparison of the average weight of wipes before and after warm air-drying)

Virus Spiking on a Plastic Carrier and Wiping

A sample of 30 µl of the virus (7.5 $\log_{10} \text{TCID}_{50}$) was applied to the middle of the bottom of a Ø 60-mm plastic Petri dish. The drop was lightly dispersed with a pipette tip to form a stain of approximately 0.5×2 cm, and allowed to dry for one hour at room temperature in a biosafety cabinet. Wiping was performed by the same person as follows: a gloved forefinger was covered with the wipe and one single direct wipe, using the fingertip, was performed with a moderate pressure over the whole area of the dried virus spot for 2 s. The Petri dishes were then kept in a refrigerator at 4 °C (±0.5 °C) before virus elution and titration, which was performed: (i) immediately after virus drying and 5 min after wiping (if applicable), (ii) every 24 h over 5 consecutive days of storage at 4 °C. All experiments were performed in triplicate.

Virus Elution from the Carrier and Titration

The virus was eluted from the carrier in 3 ml of Dulbecco's modified Eagle's medium (Biosera, Nuaille, France) at 4 °C, supplemented with 10% heat-inactivated foetal calf serum (HyClone, South Logan, Utah, USA). The medium was transferred to a Petri dish and the virus left to dissolve for 5 min, after which it was re-suspended using a 5-ml plastic syringe with a needle. The virus suspension was filter-sterilised with a 0.43- μ m PES syringe filter.

The virus was tenfold serially diluted. Each dilution, at a volume of 100 µl, was inoculated into five wells of a 48-well microtitration plate with PD 5 cell-culture monolayer. Eight non-inoculated wells were used as negative controls. Culture medium was added after one hour of incubation. The plates were incubated for 5 days at 37 °C in a 5% CO₂ atmosphere and then the cultures were microscopically examined for virus cytopathic effect. The dilution of suspension that gave rise to cytopathology in half of the cultures (the median tissue culture infective dose, \log_{10} TCID₅₀/mL) was then calculated (Kaerber 1931).

Transfer of the Virus

To study a possible virus spread to a secondary surface through wiping, virus spiking on a Petri dish and wiping of a virus spot were performed following the same method as specified above. Immediately afterwards, another single direct wipe using the same finger covered with the contaminated wipe was performed across a clean plastic \emptyset 60-mm Petri dish. Then, the virus was eluted and titrated following the method described above. Experiments with each type of wipe were performed in triplicate.

Virus Inactivation on Wipes

A drop of 30 µl of the virus (7.5 $\log_{10} \text{TCID}_{50}$) was distributed over a freshly prepared wipe cut (2×2 cm) placed on a Ø 60-mm plastic Petri dish. Wipe cuts with the virus were then kept for 5 min at room temperature in a biosafety cabinet. Subsequently, the virus was eluted from the wipe and titrated. Elution and titration were performed following the same method as the elution from the carrier. To reveal any cytotoxicity of chemicals eluted from wipes that would interfere with cytopathic effect on cell culture during virus titration, virus-free wipe cuts were used as a control in elution and titration. Experiments with each type of wipe were performed in triplicate.

Statistical Analysis

The virus titres acquired in triplicates are presented as mean \pm SD. Differences in the impacts of the different wipes on virus reduction were tested with one-way ANOVA, followed by Tukey's post-hoc test for pairwise comparisons. Results were considered significant at *P* < 0.05. All calculations were performed in Statistica v. 13 (TIBCO Software Inc.).

Results

The results for coronavirus survival on a plastic carrier are summarised in Table 2. During storage at 4 °C, the average rate of natural inactivation of the virus dried on a plastic surface was 0.93 ± 0.41 SD log₁₀ TCID₅₀ per day. The virus remained detectable by titration in some of the replications on the 5th day of storage. When the virus was treated by wiping, titres on the carrier decreased dramatically in comparison with the initial titre. The wipes saturated with a combination of disinfectants (Table 1) decreased the virus titre significantly more efficiently than the other wipes (wet and hygiene) at P < 0.05, by 3.8 $\log_{10} \text{TCID}_{50}$. For both wet wipes and hygiene wipes, infectivity decreased by 2.4 log₁₀ TCID₅₀ after wiping. The subsequent rate of inactivation of the virus on a carrier that had been previously wiped and kept at 4 °C was similar to the unwiped control decreasing by less than $1 \log_{10} \text{TCID}_{50}$ per day of storage due to environmental factors.

The average titres of *Alphacoronavirus 1* after 5 min inactivation on the wipes are listed in Table 3. When the virus was spiked on wet wipes containing no disinfectant, the virus recovery achieved on average 7.0 $\log_{10} \text{TCID}_{50}$ out of 7.5 $\log_{10} \text{TCID}_{50}$ of the original virus suspension distributed over the wipe. Wet hygiene wipes saturated with ethanol performed in inactivation of the virus bound to the wipe significantly better than the wet wipes: the virus Table 2 Residual titres of SARS-CoV-2 surrogate Alphacoronavirus 1 dried on a plastic carrier and kept in a refrigerator at 4 °C, values in the table correspond to mean \pm SD of lg TCID₅₀

	Dried virus/no treatment	Dried virus/wet wipes**	Dried virus/hygiene wipes**	Dried virus/ disinfectant wipes**
Virus recovery before storage*	7.2 ± 0.4^{a}	$4.8\pm0.7^{\mathrm{b}}$	4.8 ± 0.5^{b}	$3.4 \pm 0.5^{\circ}$
24 h at 4 °C	6.0 ± 0.4	4.1 ± 0.9	4.2 ± 0.8	≤2.7
48 h at 4 °C	5.6 ± 0.2	3.0 ± 0.3	3.4 ± 0.6	≤2.5
72 h at 4 °C	4.8 ± 0.2	≤2.6	≤3.0	≤2.5
96 h at 4 °C	3.5 ± 0.4	≤2.5	≤2.5	ND
120 h at 4 °C	≤2.8	≤2.5	≤2.5	ND

ND not performed

*Virus elution and titration was arranged immediately after visible drying of the virus drop on the carrier and 5 min after wiping (if applicable)

**Active substances are listed in Table 1

^{abc}Values with different letters indicate significant differences between treatments at P < 0.05 (one-way ANOVA and post-hoc Tukey's test)

Table 3 Inactivation of SARS-CoV-2 surrogate *Alphacoronavirus 1* distributed over wipes at RT and virus spread to a secondary surface through wiping, values in the table correspond to mean \pm SD of *lg* TCID₅₀

	Wet wipes*	Hygiene wipes*	Disin- fectant wipes*
Residual titres after 5 min inactivation on wipes	7.0 ± 0.3^{a}	5.7 ± 0.2^{b}	≤3.5 ^c
Spread to a secondary surface	3.0 ± 0.4^{a}	2.8 ± 0.2^{a}	≤2.5 ^a

*Active substances are listed in Table 1

^{abc}Different letters indicate significant differences between results of different wipes at P < 0.05 (one-way ANOVA, Tukey's post-hoc test)

recovery was in average $5.7 \log_{10} \text{TCID}_{50}$. The virus titres after 5 min inactivation on disinfectant wipes were under the detection limit of $3.5 \log_{10} \text{TCID}_{50}$. This limit was caused by the cytotoxicity of the chemicals eluted from the wipes that interfered with the virus titration.

The results of the experimental virus transfer to a secondary surface through wiping are listed in Table 3. The quantities of the virus detected on the surface after wiping with contaminated wet and hygiene wipes did not differ significantly, being on average 3.0 and 2.8 $\log_{10} \text{TCID}_{50}$, respectively. Transfer of the virus through wiping with disinfectant wipes was not confirmed, as the results were under the detection limit of 2.5 $\log_{10} \text{TCID}_{50}$.

Discussion

The persistence of the coronavirus on a plastic carrier simulating contaminated food packages stored in a refrigerator at 4 °C was investigated herein using a SARS-CoV-2 surrogate *Alphacoronavirus 1*. In agreement with the findings of previous studies (Geller et al. 2012; Ren et al. 2020), these results confirmed high coronavirus persistence, especially at a low temperature and on a plastic surface; this persistence is higher even in comparison with other enveloped respiratory viruses, e. g. the influenza virus (Otter et al. 2016; Zhang and Li 2018). However, according to Mullis et al. (2012), the infectivity of *Betacoronavirus 1* on the surface of romaine lettuce under household refrigeration conditions extended to 14 days. Fomite transmission of viral respiratory diseases, including coronaviruses and less persistent viruses, has been previously proven or suspected (Boone and Gerba 2007; Xiao et al. 2017). Those findings support the possibility of COVID-19 spread through food and food packaging.

The second aim of this study was to investigate the efficacy of wet-wiping in removing the virus contamination from a plastic surface, and specifically to determine any differences between wiping with or without disinfectants. An efficient reduction, of 2.4 \log_{10} (99.6%), of the virus was achieved with a single direct wipe over the contaminated area even with a wipe containing no disinfectant. Similarly, wet wiping has been proven as an efficient tool for removing coronavirus from the hands (Ma et al. 2020). Hygienic wipes containing 16.5% ethanol (Table 1) gave the same results as wet wipes. Wiping is a short duration activity and the virucidal effect of disinfectant is required within seconds (Verhaelen et al. 2014). Ethanol is widely assumed to be very effective against coronaviruses, especially at concentrations of around 70% with a contact time of 30 s or longer (Geller et al. 2012). Ethanol in the formulation tested here had no effect on the reduction of the virus during wiping from the surface. In the concentration tested, it had also no effect on the transfer of the virus to another surface if the latter was wiped immediately after the contamination of the wiping cloth, which supports previous findings (Kampf 2018). On the other hand, a longer exposure (5 min), when the virus was bound to the hygienic wipe, resulted in a significant reduction of the virus in comparison with wet wipes. Therefore, certain reduction of the risk of further transfer of the virus to another surface during wiping was proven even for wipes containing 16.5% ethanol. Surface disinfectant wipes containing a combination of disinfectant agents, including didecyl-dimethyl-ammonium chloride and hydrogen peroxide (Table 1), reduced Alphacoronavirus 1 significantly more effectively than the other wipes tested. Both the disinfectant agents mentioned have been demonstrated as active against coronavirus in concentrations used in experiments in this study (Pratelli 2007; Kampf et al. 2020; Lin et al. 2020). Disinfectant wipes were also the only product tested that reduced the virus under the detection limit after 5 min binding to the wipes. Furthermore, the virus was undetectable also after a transfer to a secondary surface through wiping with a contaminated disinfectant wipe. The significance of the use of disinfectants to prevent a transfer of the virus to a larger surface is in accordance with previous findings (Barker et al. 2004).

The results of this study reveal the possible benefit of cleaning and disinfection of food and food packages before their use or storage in a refrigerator, a procedure that has also been recommended by the CDS (Seymour et al. 2020). Wet wiping using readily-available drugstore products for the household appears to be an efficient cleaning method for reduction of any SARS-CoV-2 transmission risk that may arise out of contaminated plastic packaging and food. Satisfactory reduction of the virus can be achieved even with wipes lacking any disinfectant, thus leaving no toxic residues. However, to prevent any transfer of the virus through contaminated wipes to other surfaces, the saturation of the wipes with a combination of disinfectants in sufficient concentrations is crucial. Furthermore, the wipes should be changed frequently and discarded to keep the effect of wiping optimal and minimise cross-contamination and transfer of the virus.

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Compliance with ethical standards

Conflict of interest The author declares that she has no conflict of interest.

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