



Population pharmacokinetic analysis of phase 1 bemarituzumab data to support phase 2 gastroesophageal adenocarcinoma FIGHT trial

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Abstract

Purpose To report population pharmacokinetic (PK) analysis of the phase 1 study (FPA144-001, NCT02318329) and to select a clinical dose and schedule that will achieve an empirical target trough concentration (C_{trough}) for an anti-fibroblast growth factor receptor 2b antibody, bemarituzumab.

Methods Nonlinear mixed-effect modeling was used to analyse PK data. In vitro binding affinity and receptor occupancy of bemarituzumab were determined. Simulation was conducted to estimate dose and schedule to achieve an empirical target C_{trough} in a phase 2 trial (FIGHT, NCT03694522) for patients receiving first-line treatment combined with modified 5-fluorouracil, oxaliplatin and leucovorin (mFOLFOX6) for gastric and gastroesophageal junction adenocarcinoma.

Results Bemarituzumab PK is best described by a two-compartment model with parallel linear and nonlinear (Michaelis–Menten) elimination from the central compartment. Albumin, gender, and body weight were identified as the covariates on the linear clearance and/or volume of distribution in the central compartment, and no dose adjustment was warranted. An empirical target of bemarituzumab C_{trough} of ≥ 60 $\mu\text{g/mL}$ was projected to achieve $> 95\%$ receptor occupancy based on in vitro data. Fifteen mg/kg every 2 weeks, with a single dose of 7.5 mg/kg on Cycle 1 Day 8, was projected to achieve the target C_{trough} on Day 15 in 98% of patients with 96% maintaining the target at steady state, which was confirmed in the FIGHT trial.

Conclusion A projected dose and schedule to achieve the target C_{trough} was validated in phase 1 of the FIGHT trial which supported selection of the phase 2 dose and schedule for bemarituzumab.

Keywords Bemarituzumab · Anti-fibroblast growth factor receptor 2b · Population PK analysis · Target-mediated clearance · Dose selection · Gastric and gastroesophageal junction adenocarcinoma

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Introduction

Gastric and gastroesophageal junction adenocarcinoma (GEA) represents the fourth most common cancer worldwide and is a highly lethal disease, with 5-year overall survival (OS) rates below 33% in the United States (US) [1–3]. Chemotherapy as standard first-line treatment has demonstrated an improvement in OS compared to best supportive care and additional progress has been made with the use of targeted therapies, such as trastuzumab and ramucirumab. However, even with these therapies, prognosis is poor. Therefore, the identification of new therapeutics with acceptable toxicities is important for this patient population [4, 5].

The fibroblast growth factor/fibroblast growth factor receptor (FGF/FGFR) pathway can stimulate the

transformation and proliferation of tumor cells and angiogenesis. FGF signaling is mediated by a family of transmembrane tyrosine kinase receptors encoded by four distinct genes producing FGF receptor subtypes termed FGFR1–4 [6]. Fibroblast growth factor receptor 2b (FGFR2b), one of 2 FGFR2 splicing variants, is expressed in tissues of epithelial origin (e.g., stomach, skin) with FGF7, FGF10, and FGF22 as three of its major ligands [7]. Alterations in signaling in the FGF/FGFR2 pathway (e.g., overexpression of FGFR2 protein or amplification of *FGFR2* gene) have been associated with GEA, breast, and other cancers and with a decreased prognosis [6, 8–11], suggesting that inhibition of FGFR2 may be a rational target for cancer therapy [12, 13].

Bemarituzumab (FPA144) is a first-in-class humanized Immunoglobulin G1 (IgG1) monoclonal antibody specific to the human FGFR2b receptor that blocks FGF binding to the receptor. It is also glycoengineered for increased affinity for human Fc gamma receptor IIIA (FcγRIIIa) expressed on natural killer cells, resulting in enhanced antibody-dependent cell-mediated cytotoxicity (ADCC) against FGFR2b-overexpressing tumors. These two mechanisms of action of bemarituzumab may together lead to improved OS in patients with GEA whose tumors overexpress FGFR2b.

Bemarituzumab has been evaluated in patients receiving treatment for late-line solid tumors, including those with GEA with or without overexpression of FGFR2b in the FPA144-001 trial (NCT02318329). Promising single agent activity with acceptable tolerability was demonstrated in patients whose GEA overexpressed FGFR2b [14]. GEA tends to be highly heterogeneous within the same tumor, and when present, FGFR2b may not be uniformly distributed throughout the tumor specimen [15, 16]. Combining bemarituzumab with chemotherapeutic agents that will target FGFR2 negative tumor cells is likely to improve the clinical benefit over bemarituzumab alone. Currently, bemarituzumab is being evaluated in combination with modified 5-fluorouracil, oxaliplatin, and leucovorin (mFOLFOX6) in the phase 2 placebo-controlled trial, A Study of Bemarituzumab Combined with Modified FOLFOX6 in Gastric/Gastroesophageal Junction Cancer (FIGHT, NCT03694522).

Dose selection is frequently a challenge for novel biological oncology drugs because the maximum tolerated dose approach may identify a dose that is significantly higher than the dose required to achieve maximum efficacy [17]. There are additional challenges for an orphan drug like bemarituzumab because of the lower prevalence of tumors expressing the target FGFR2b, which may limit the feasibility of an extensive clinical dose-ranging study. Therefore, an efficient approach was needed to advance bemarituzumab dose selection. To achieve this goal, multiple factors had to be considered, including population pharmacokinetic analysis (PopPK), which is the focus of this manuscript. On the basis of the mechanism of action for bemarituzumab (including

blocking of FGFR2b binding to its ligands), it is projected that keeping trough concentration (C_{trough}) above a threshold to saturate the target between-dose interval might provide maximum anti-tumor activity. Therefore, an empirical target C_{trough} estimated from in vitro data was included as an exposure target to support dose selection. This approach was used in multiple targeted antibody therapies including bevacizumab and trastuzumab as well as immunotherapies, such as ipilimumab [18–20]. In addition, a concentration associated with a high percentage of saturation of target-mediated clearance for bemarituzumab can be used as the target C_{trough} .

Here, we describe results from the PopPK analysis of bemarituzumab using serum concentration data from a first-in-human phase 1 study (FPA144-001) to support dose selection for the phase 2 trial. The key objectives of the analysis were to assess pharmacokinetic (PK) and covariates of bemarituzumab in humans using PopPK analysis, to describe the in vitro data used to derive an empirical target C_{trough} , to report the simulation results which identified the dose and regimen necessary to achieve an empirical target C_{trough} in the majority of patients, and to provide clinical data to support the projection from the simulation.

Materials and methods

Antibody reagents

Bemarituzumab was produced in a Chinese hamster ovary cell line that lacks the *FUT8* gene (α 1, 6-fucosyltransferase) at Five Prime Therapeutics, Inc. (South San Francisco, CA) for nonclinical studies, at AbbVie Bioresearch Center (Worcester, MA) for the FPA144-001 trial, and at Patheon (St. Louis, MO) for the FIGHT trial.

Bemarituzumab serum concentration in humans

Human serum concentration data were collected from 75 patients enrolled in the FPA144-001 clinical trial; 22 patients with a variety of solid tumors and 53 patients with GEA. Among the 75 patients, 27 were enrolled in the dose escalation portion of the trial and received a 30-min intravenous (IV) infusion of bemarituzumab once every 2 weeks (Q2W) in a range of doses [0.3 ($n=3$), 1 ($n=4$), 3 ($n=4$), 6 ($n=4$), 10 ($n=9$), and 15 ($n=3$) mg/kg; Part 1]. An additional 48 patients received 15 mg/kg Q2W in a 30-min IV infusion (Part 2). Each cycle had two doses with a Q2W dosing regimen. Three to five serum concentration samples were collected within 8 h on the day of dosing followed by collections on Days 2, 4, and 8 post-first dose of Cycle 1. Additional collections were done both before and at the end of infusion for the second dose of Cycle 1 and the first dose

of Cycles 2 through 5, every other cycle thereafter, and at the End of Treatment Follow-up Visit. Part 2 had the same serum concentration sample collection time points without collection of samples on Days 2 and 4 post-first dose.

FIGHT is a phase 2 trial preceded by dose-finding in phase 1. C_{trough} data collected during Cycle 1 through Cycle 5 from ten patients ($n=3$ for 6 mg/kg and $n=7$ for 15 mg/kg Q2W with a single dose of 7.5 mg/kg on Cycle 1 Day 8) in phase 1 of the FIGHT trial made up the observed serum concentration data displayed in Fig. 4.

Anti-drug antibody in humans

Anti-drug antibody (ADA) samples were collected from all patients in study FPA144-001 prior to dosing on Day 1 of Cycles 1 to 5, every other cycle after Cycle 5, and at the End of Treatment Follow-up Visit.

Determination of serum concentration of beemarituzumab in humans

The beemarituzumab serum concentration in humans was quantitatively measured at ICON Laboratory Services, Inc. (Whitesboro, NY) with a validated ELISA in which beemarituzumab was captured by plate-bound FGFR2b-Fc and detected by an anti-idiotypic antibody coupled to HRP. The LLOQ of the assay was 0.125 µg/mL. The accuracy expressed as percent relative error ranged from 0.250% to 4.80%. The intra- and inter-assay ranges were –0.240% to –4.80% and 5.74% to 7.45%, respectively.

Measurement of anti-drug antibody in patients

A validated bridging electrochemiluminescence assay that utilizes Meso Scale Discovery (MSD) technology was used to detect ADA in patient serum samples. The samples were acidified, then neutralized in a reaction mixture containing ruthenylated beemarituzumab and biotinylated beemarituzumab. After a 75-min incubation, the samples were transferred onto a streptavidin coated MSD assay plate. The plate was washed and a tripropylamine (TPA)-containing MSD read buffer added. In the presence of TPA, ruthenium produces a chemiluminescent signal when voltage is applied. Only samples that contained antibody bound to both biotinylated beemarituzumab and ruthenylated beemarituzumab produced the chemiluminescent signal, which was proportional to the amount of anti-beemarituzumab antibody present. The assay sensitivity was 32.7 ng/mL relative to the rabbit anti-FPA144 positive control. Drug tolerance was 2 µg/mL in the presence of 800 ng/mL of rabbit anti-beemarituzumab antibody.

Human population pharmacokinetic analysis

A PopPK model was developed to describe serum beemarituzumab concentration–time profiles. Various PK structural models were tested, including a one-compartment, two-compartment, two-compartment with time-varying clearance, and two-compartment with linear and nonlinear elimination components. The PK of beemarituzumab in the tested clinical dose range was best described by a two-compartment model with parallel linear and nonlinear (Michaelis–Menten) elimination pathways from the central compartment using the differential equations below [21, 22]:

$$\frac{dA_c}{dt} = - \left[\left(\frac{V_{\max}}{K_m + \frac{A_c}{V_c}} \right) / V_c + \frac{CL}{V_c} + \frac{Q}{V_c} \right] \times A_c + \frac{Q}{V_p} \times A_p \quad (1)$$

$$\frac{dA_p}{dt} = \frac{Q}{V_c} \times A_c - \frac{Q}{V_p} \times A_p \quad (2)$$

where A_c and A_p are the amount of drug in central and peripheral compartments, respectively. V_{\max} represents the maximum drug elimination by nonlinear clearance, and K_m , Michaelis–Menten constant, indicates the drug concentration at 50% V_{\max} . CL and Q represent linear clearance and distribution clearance, respectively, while V_c and V_p represent volume of distribution in central and peripheral compartments, respectively.

After an optimal structural model was identified (final base model), the effect of covariates (Table 1) including baseline demographic information, renal function, hepatic function, disease status, baseline tumor measurements, Eastern Cooperative Oncology Group (ECOG) status, and FGFR2b expression in patients with GEA were evaluated for their impacts on the CL and V_c . FGFR2b high was defined as $\geq 10\%$ of tumor cells with 3+ membranous staining using a centrally performed validated laboratory-developed prototype immunohistochemistry assay (LabCorp, Burlington, NC). Correlations between the PK parameters and the covariates were explored graphically, followed by linear regression (continuous covariates) and analysis of variance (ANOVA) testing (categorical covariates) using R software. These analyses were conducted on individual random effects (ETAs) for CL and V_c . Only covariates that showed a significant ($p < 0.01$) effect on the random effect, and that could be meaningfully explained from both a clinical and scientific perspective, were examined further using NONMEM. Covariates were selected based on their potential clinical relevance. The possible physiological basis for commonly evaluated covariates such as body size, age, gender, race, ADA, serum albumin, creatinine clearance, etc., were summarized by Thomas and Balthasar [23]. Selection of the final

Table 1 Covariate values of beemarituzumab population pharmacokinetic dataset

Variable	Study FPA144-001 (<i>n</i> = 75)
Age (year), median (range)	58 (25, 86)
Weight (kg), median (range)	61.4 (35.5, 148)
Gender, <i>n</i> (%)	
Female	42 (56%)
Male	33 (44%)
Race, <i>n</i> (%)	
White	29 (38.7%)
American Indian or Alaska Native	1 (1.33%)
Asian	44 (58.7%)
Black or African American	1 (1.33%)
Albumin (g/dL), median (range)	3.7 (1.9, 4.6)
Creatinine Clearance (mL/min), median (range)	75.3 (26.4, 200)
Total Bilirubin (mg/dL), median (range)	0.4 (0.1, 2.0)
ALT (U/L), median (range)	19 (6, 74)
AST (U/L), median (range)	24 (7, 106)
Baseline Tumor Size (mm), median (range)	50 (10, 244)
Tumor type, <i>n</i> (%)	
Gastric cancer	53 (70.7%)
Other solid tumors	22 (29.3%)
ECOG status, <i>n</i> (%)	
0	23 (30.7%)
1	52 (69.3%)
FGFR2b Expression in patients with gastric and gastroesophageal junction adenocarcinoma, <i>n</i> (%)	
FGFR2b high	26 (49%)
FGFR2b other	27 (51%)

covariate model (final PopPK model) was determined for its significance on the basis of likelihood ratio test at the $p < 0.01$ for forward inclusion and $p < 0.001$ for backward deletion.

The final PopPK model was evaluated with multiple internal model validations, including goodness-of-fit diagnostics, prediction-corrected visual predictive check (pcVPC) plots, numerical predictive check, bootstrap, and shrinkage assessments. The pcVPC was created to assess the predictive ability of the model. A total of 1000 replicates of the trials were simulated using the individual dosing history and covariates, the typical parameter estimates, and random sampled interindividual variability and residual errors. The 2.5th, 50th, and 97.5th percentiles of the observed data were overlaid on the 90% confidence interval (CI) of the 2.5th, 50th, and 97.5th simulated percentiles, and a visual inspection was performed. The sensitivity analysis was performed for the final PopPK model to examine the contribution of significant baseline covariates to the overall variability of the steady-state exposures including area under concentration–time curve at steady state (AUC_{ss}), maximum serum concentration at steady state ($C_{max,ss}$), and trough concentration at steady state ($C_{trough,ss}$) after Q2W dosing of 15 mg/kg beemarituzumab.

The PopPK analysis was performed using the nonlinear mixed effects modeling approach with the first-order conditional estimation with interaction method. Model parameter estimation and evaluation were implemented with NONMEM 7 (v. 7.3.0; ICON Development Solutions, Ellicott City, MD) with an Intel Fortran Compiler (v. 10.1.021; Intel, Santa Clara, CA), Perl-speaks-NONMEM (PsN, v. 3.2.12; Uppsala University, Uppsala, Sweden), and R 3.3.1.

Binding affinity using surface plasmon resonance

The binding affinity of beemarituzumab for human FGFR2b ECD-IgG1 Fc fusion protein (FGFR2b-Fc) was measured using surface plasmon resonance (SPR, Biacore T100, GE Healthcare Life Science, Marlborough, MA). Beemarituzumab was immobilized on a dextran chip using an amine coupling kit and 100 mM ethylenediamine in 100 mM Sodium Borate, pH 8.0, was used as the blocking reagent. Six different concentrations (0 nM to 500 nM) of FGFR2b-Fc proteins were diluted in HEPES buffered saline with 0.05% surfactant P20 running buffer and flowed over the immobilized antibody.

To determine binding affinity for FcγRIIIa (V158) by SPR, beemarituzumab was captured on the chip via protein A. Protein A was covalently attached to a dextran chip using

the same protocol as above. Five concentrations (0 nM to 1000 nM) of Fc γ R3a (V158) were diluted in running buffer and flowed over the captured antibody.

The association constant, dissociation constant, and affinity for beemarituzumab binding to human FGFR2b and human Fc γ R3a (V158) were calculated using the Biacore T100 Evaluation Software 1:1 binding model.

Cell lines

OCUM-2M is a *FGFR2* gene-amplified, FGFR2b protein overexpressing gastric cancer cell line obtained from Public University Corporation Osaka City University, Japan (Source: Dr Masakazu Yashiro). HSC-39 is a *FGFR2*-amplified gastric cell line and was kindly provided by Rebecca Fitzgerald (Hutchison/MRC Research Centre, Cambridge, UK). Gastric cell lines SNU-16 (with *FGFR2* amplification) and NCI-N87 (without *FGFR2* amplification or overexpression) were commercially acquired (ATCC, Manassas, VA). All four cell lines were mycoplasma negative tested by IDEXX Laboratories (Columbia, MO) using a real-time polymerase chain reaction.

Gastric cell lines-based binding

The cell surface expression of FGFR2b on OCUM-2 M, HSC-39, SNU-16, and NCI-N87 cells was calculated after incubation with 50 μ g/mL of either beemarituzumab or human IgG1 (human anti-hen egg lysozyme, Five Prime Therapeutics, Inc). Unbound antibody was washed with 1 \times phosphate-buffered saline with 2% fetal bovine serum (VWR Visalia, CA) and the cells were incubated with mouse anti-human IgG1-PE (Clone HP6001, lot L3117-M839, SouthernBiotech, Birmingham, AL). Cells were washed and data acquired on the BD LSRII Flow Cytometer (BD, San Jose, CA). Quantum Simply Cellular anti-mouse IgG Bang Beads (815B lot 12,380, Bangs Laboratories, Fishers, IN) were stained in parallel according to the manufacturer's instructions and acquired on the BD LSRII Flow Cytometer. The antibody binding capacity (ABC) was calculated using the Bang Laboratories QuickCal Template.

Separately, 2×10^5 OCUM-2 M, HSC-39, SNU-16, or NCI-N87 cells were incubated with varying concentrations of beemarituzumab or human IgG1, washed, stained with a mouse anti-human IgG1 PE secondary antibody (clone HP6001; SouthernBiotech, Birmingham, AL), washed, and acquired on the BD LSRII Flow Cytometer. A 4-parameter logistical regression curve fit for acquired mean fluorescence intensity versus beemarituzumab concentration data in semi-log plot was applied in GraphPad PRISM (La Jolla, CA) to estimate receptor occupancy.

Human pharmacokinetic simulation

Bemarituzumab exposures were simulated using the final PopPK model with different dose regimens. For each dose regimen, 1000 simulated model parameters constructed the distribution of model predictions for a typical population (61 kg male patients with albumin of 3.7 g/dL). The area under the concentration–time curve (AUC), C_{\max} , and C_{trough} after single and multiple treatment for each of the dose regimens were computed. The simulation was done for the dose cohorts tested in the safety lead-in phase 1 portion of the FIGHT trial: 6 mg/kg Q2W and 15 mg/kg Q2W with a single dose of 7.5 mg/kg on Cycle 1 Day 8.

Results

Bemarituzumab serum concentration in humans

Bemarituzumab serum concentration versus time data (group mean \pm SD) from Cycle 1 dose 1 in the FPA144-001 trial displayed a typical antibody serum concentration profile with a short distribution phase followed by a long elimination phase [14]. Bemarituzumab demonstrated dose dependent clearance in patients with solid tumors including GEA, with faster clearance at lower doses, suggesting target-mediated drug disposition. Clearance appeared non-linear from 0.3 mg/kg to 1 mg/kg and approximately linear from 1 mg/kg to 15 mg/kg.

The anti-drug antibody impact on beemarituzumab pharmacokinetics

A total of 75 patients from the FPA144-001 trial were tested for ADA. The patients were exposed to beemarituzumab for up to 966 days (median = 55 days). No patient sample was confirmed to be ADA positive after administration of beemarituzumab. A predose sample before beemarituzumab administration from one patient was ADA positive; however, all tested ADA samples post-bemarituzumab administration for this patient were negative. The predose ADA positive signal did not have a visible impact on the PK profile of this patient in comparison with the PK profiles from the other patients treated at the same dose level.

Population pharmacokinetic analysis

The development dataset for the final model included 814 beemarituzumab serum concentration data from 75 participants in the phase 1 study FPA144-001. A two-compartment model with parallel linear and nonlinear (Michaelis–Menten) elimination from the central compartment best described the beemarituzumab serum concentration data. No

time-varying CL was identified after beemarituzumab administration. Covariate analysis was conducted to understand the impact of the covariate values (Table 1) on the CL and V_c of beemarituzumab. The forward addition and backward deletion based on the final base model identified body weight, albumin, and gender as statistically significant covariates for beemarituzumab disposition parameters of CL and V_c , and this model is referred to as the final PopPK model:

$$CL_i = \exp\left(\theta_1 + \theta_7 \cdot \left(\frac{\text{Weight}}{61}\right) + \theta_9 \cdot \left(\frac{\text{ALB}}{3.7}\right) + \eta_{CL}\right) \quad (3)$$

$$V_{C_i} = \exp\left(\theta_2 + \theta_8 \cdot \left(\frac{\text{Weight}}{61}\right) + \theta_{10} \cdot \text{female} + \eta_{V_c}\right) \quad (4)$$

where CL_i is the individual linear clearance; V_{C_i} is the individual volume of distribution in the central compartment, η_{CL} and η_{V_c} are interindividual variability of CL and V_c , respectively.

The final PopPK model estimated a typical CL, saturation of target-mediated drug disposition, of 0.331 L/day, V_c of 3.70 L, Q of 0.788 L/day, V_p of 2.05 L, V_{\max} of 1.70 $\mu\text{g}/\text{day}$, and K_m of 4.58 $\mu\text{g}/\text{mL}$. The estimated linear clearance half-life was 12.8 days. The interindividual variability on CL, V_c , and V_p were 27.1%, 17.3%, and 60.0%, respectively.

Bootstrapping of 1000 datasets resulted in median parameter estimates and 95% CI similar to the estimates from the original dataset (Table 2), indicating that the final PopPK model provided good precision for

parameter estimation. Goodness-of-fit plots showed agreement between predicted and observed concentrations of beemarituzumab with no apparent bias in residual plots over time or across population-predicted concentrations (Supplementary Fig. 1). pcVPC plots (Fig. 1) demonstrated that the final PopPK model could reasonably describe the central tendency and variability of the beemarituzumab pharmacokinetic data.

Body weight was identified as a covariate for both CL and V_c , while albumin was a covariate for CL and gender was a covariate for V_c . Patients with higher body weight had statistically larger V_c and faster CL. Lower albumin had statistically faster CL and male patients had a higher V_c . Due to the limited number of patients ($n=75$), the covariate relationship should be confirmed in future studies. The sensitivity analysis suggested that albumin is the most important factor influencing the $C_{\text{trough ss}}$ of beemarituzumab. The population-predicted 5th and 95th percentile of total $C_{\text{trough ss}}$ for the actual patients' Q2W dosing of the 15 mg/kg were 65 and 196 $\mu\text{g}/\text{mL}$, respectively, corresponding to -47.5% and 58.3% variation around the $C_{\text{trough ss}}$ predicted for a typical patient (123.8 $\mu\text{g}/\text{mL}$). Of note, the extreme albumin (5th and 95th percentiles) corresponded to as high as -31.3% and 24.4% variation for $C_{\text{trough ss}}$ (Fig. 2). Body weight had a mild impact based on the sensitivity analysis (-9.2% and 8.7% variation on $C_{\text{trough ss}}$; Fig. 2). Gender was also an influential factor for the $C_{\text{trough ss}}$ with a -6.2% variation for female compared to male patients (Fig. 2). Albumin was the most influential

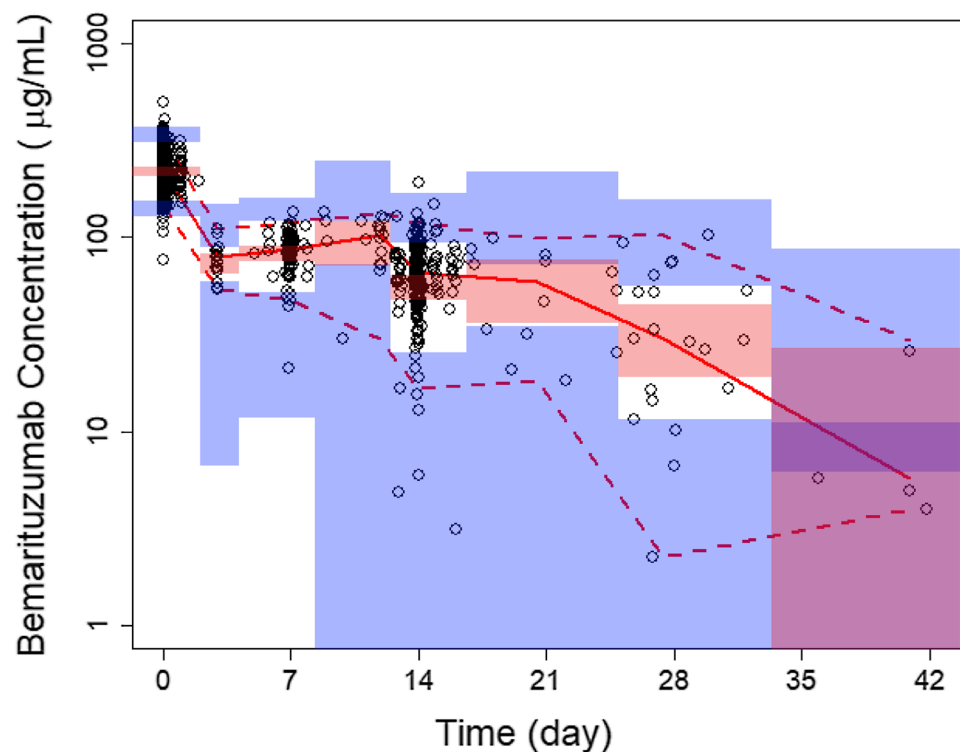
Table 2 Summary of population pharmacokinetic parameters

Parameter description	Base model estimates (%RSE)	Final model estimates (%RSE)	Bootstrap estimate median (2.5–97.5%tiles)
V_{\max} ($\mu\text{g}/\text{day}$)	2.75 (21.6%)	1.70 (14.3%)	1.82 (0.0901; 7.55)
K_M ($\mu\text{g}/\text{mL}$)	4.64 (42.5%)	4.58 (15.1%)	5.24 (0.533; 24.7)
Linear clearance, CL (L/day)	0.331 (4.73%)	0.331 (3.55%)	0.327 (0.275; 0.364)
Influence of body weight on CL	–	0.601 (19.5%)	0.641 (0.355; 0.991)
Influence of albumin on CL	–	-0.776 (19.9%)	-0.777 (-1.31 ; -0.262)
Volume of central compartment, V_c (L)	3.43 (2.76%)	3.70 (2.98%)	3.70 (3.51; 3.92)
Influence of body weight on V_c	–	0.303 (24.8%)	0.306 (0.109; 0.471)
Influence of sex on V_c	–	-0.191 (23.4%)	-0.194 (-0.284 ; -0.114)
Distribution clearance, Q (L/day)	0.772 (10.6%)	0.788 (8.99%)	0.791 (0.599; 1.11)
Volume of peripheral compartment, V_p (L)	2.07 (8.25%)	2.05 (7.73%)	2.08 (1.76; 2.42)
Interindividual variability of V_{\max}	113 (32.6%)	128 (23.3%)	122 (65.9; 437)
Interindividual variability of CL	36.0 (20.1%)	27.1 (19.7%)	25.5 (18.4; 31.7)
Interindividual variability of V_c	22.2 (18.2%)	17.3 (19.1%)	16.8 (13.7; 19.9)
Interindividual variability of V_p	59.3 (25.1%)	60.0 (24.1%)	56.0 (38.3; 74.9)
Covariance between CL and V_c	0.0319 (36.2%)	0.0141 (46.1%)	0.0137 (0.00111; 0.0254)
Residual variability (%CV)	14.4 (5.61%)	14.5 (5.89%)	14.4 (13.0; 15.7)

$$CL_i(\text{L}/\text{hr}) = \exp\left(-4.284 + 0.601 \times \log\left(\frac{\text{WT}}{61}\right) - 0.776 \times \log\left(\frac{\text{ALB}}{3.7}\right) + \eta_{CL,i}\right)$$

$$V_{C_i}(\text{L}) = \exp\left(1.308 + 0.303 \times \log\left(\frac{\text{WT}}{61}\right) - 0.191 \times \text{Female} + \eta_{V_{C,i}}\right)$$

Fig. 1 Prediction-corrected VPC of bezarituzumab serum concentration–time profile across dose groups. Black open circles are observed serum concentrations, solid red line represents the median observed value, and dashed red lines represent 2.5th and 97.5th percentile of the observed values, respectively. Pink shaded areas represent the spread of the median predicted values (5th to 95th percentile), and blue shaded areas represent the spread (5th and 95th percentile) of the 2.5th and 97.5th predicted percentile concentrations



factor for AUC_{ss} with -21.7% and 16.4% variation while body weight had the greatest impact on $C_{max,ss}$ with -20% to 25.7% variation (data not shown).

An empirical target trough concentration based on in vitro data

The affinity of bezarituzumab for FGFR2b and FcγRIIIa (V158) was measured by SPR. The binding affinity of bezarituzumab for FGFR2b was 0.58 nM and for FcγRIIIa (V158) was 9.2 nM. Therefore, 95% and 99% receptor occupancy by bezarituzumab for FGFR2b is estimated to be achieved at concentrations ≥ 1.59 $\mu\text{g/mL}$ and ≥ 8.26 $\mu\text{g/mL}$, respectively, and 95% and 99% receptor occupancy for FcγRIIIa (V158) is estimated to be achieved at concentrations of 25.1 $\mu\text{g/mL}$ and 131 $\mu\text{g/mL}$, respectively, using the following equation:

$$\% \text{Receptor occupancy} = (C / (KD + C)) \times 100. \quad (1)$$

where C is the concentration of the molecule and KD is the affinity of the molecule for its ligand/binding partner.

Among 3 FGFR2-amplified cell lines tested, FGFR2 expression level on the cell surface demonstrated the following order: OCUM-2 M, SNU-16, and HSC-39. The mean EC_{50} is similar among these three cell lines with values at 1.37 ± 0.0525 $\mu\text{g/mL}$, 1.81 ± 0.0289 $\mu\text{g/mL}$, and 2.53 ± 0.170 $\mu\text{g/mL}$ for OCUM-2 M, HSC-39, and

SNU-16, respectively (Fig. 3). The EC_{95} is 26.1 ± 1.00 $\mu\text{g/mL}$, 34.4 ± 0.548 $\mu\text{g/mL}$, and 48.1 ± 3.24 $\mu\text{g/mL}$, respectively.

Therefore, an empirical targeted C_{trough} of 60 $\mu\text{g/mL}$ was selected, which would achieve $>95\%$ receptor occupancy based on binding affinity and in vitro cell-based binding data from multiple FGFR2-amplified cell lines.

Simulation to support dose and regimen selection for bezarituzumab in combination with modified FOLFOX6 in the FIGHT trial

Using the final PopPK model, simulations were performed to determine whether alternative dosing schedules would allow patients to more rapidly reach target C_{trough} of ≥ 60 $\mu\text{g/mL}$ for a typical population (61 kg male patients with albumin of 3.7 g/dL). Accordingly, the dose cohorts tested in the safety lead-in phase 1 portion of the FIGHT trial were 6 mg/kg Q2W and 15 mg/kg Q2W with a single dose of 7.5 mg/kg on Cycle 1 Day 8. If the last dose cohort was not tolerable, then the Cycle 1 Day 8 dose was to be omitted and 15 mg/kg Q2W would be tested as a de-escalation dose cohort. The serum concentration versus time profiles for the two dose cohorts in the phase 1 portion of the FIGHT trial were projected (Fig. 4). The addition of a single dose of 7.5 mg/kg on Cycle 1 Day 8 for 15 mg/kg Q2W was predicated to allow a majority of patients (98%) to achieve target C_{trough} by

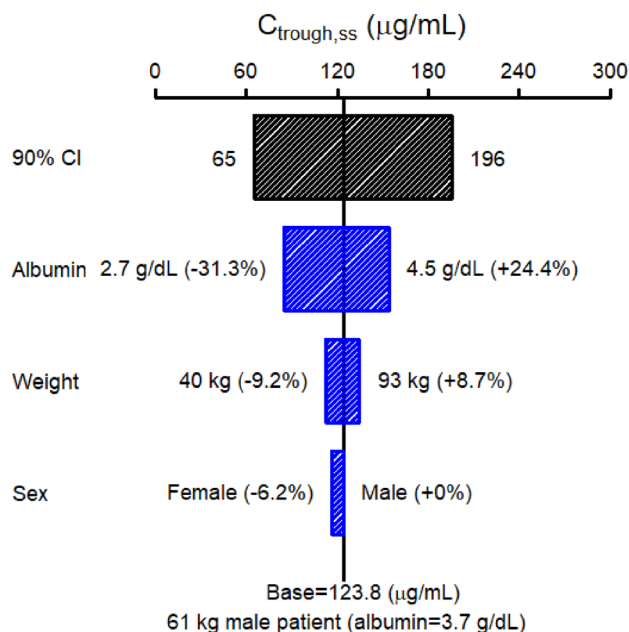


Fig. 2 Sensitivity plot comparing the effect of covariates on $C_{\text{trough,ss}}$ of bevacizumab. Base, as represented by the black vertical line and value, refers to the predicted typical $C_{\text{trough,ss}}$ of bevacizumab in a 61 kg male patient with an albumin of 3.7 g/dL kg after continuous Q2W dosing of 15 mg/kg bevacizumab for 6 months. The black horizontal bar with values at each end shows the 5th to 95th percentile $C_{\text{trough,ss}}$ range across the entire population. Each blue bar represents the influence of a single covariate on the $C_{\text{trough,ss}}$. The label at the left end of the bar represents the covariate being evaluated. The upper and lower values for each covariate capture 90% of the plausible range in the population. The length of each bar describes the potential impact of that particular covariate on bevacizumab $C_{\text{trough,ss}}$ with the percentage value in the parentheses at each end representing the percent change of $C_{\text{trough,ss}}$ from the base. The most influential covariate is at the top of the tornado plot

Day 15, compared to 96% achieving target C_{trough} by week 10 with good tolerance, thereby shortening the time to reach the target C_{trough} .

PK profile of bevacizumab is not affected by addition of mFOLFOX6

Bevacizumab in combination with mFOLFOX6 had an acceptable toxicity profile in previously treated patients with GEA in the phase 1 safety lead-in portion of the FIGHT trial at its highest dose tested-15 mg/kg Q2W with a single dose of 7.5 mg/kg on Cycle 1 Day 8 [24]. Bevacizumab concentration was not affected by the presence of mFOLFOX6. In addition, C_{trough} concentration from all seven patients who received 15 mg/kg with a single dose of 7.5 mg/kg on Cycle 1 Day 8 achieved target C_{trough} of 60 µg/mL (Fig. 4b).

Discussion

Bevacizumab, an FGFR2b inhibitor, demonstrated promising monotherapy activity in late-line GEA [14]. A phase 2 trial (FIGHT) was designed to evaluate bevacizumab combined with mFOLFOX6 chemotherapy in patients with previously untreated GEA whose tumors overexpress FGFR2b or have *FGFR2* amplification, however, efficient dose selection was required. A PK simulation in combination with an empirical target C_{trough} obtained from in vitro data was used to support the selection of the maximum test dose at 15 mg/kg Q2W with a single dose of 7.5 mg/kg on Cycle 1 Day 8 with mFOLFOX6 in the phase 1 safety lead-in portion of the FIGHT trial. This dose and schedule were selected for the phase 2 evaluation following the observed PK and safety data from phase 1 portion of FIGHT trial [24].

This is the first report of bevacizumab PopPK analysis and covariate assessment of the phase 1 data. Bevacizumab PK was best described by a two-compartment model with parallel linear and nonlinear (Michaelis–Menten) elimination from the central compartment. Target-mediated clearance observed with bevacizumab has been reported for multiple other antibodies, such as cetuximab, panitumumab, and anti-NRP1 [25–27].

Covariates impacting CL and V_c are variable for targeted antibodies. For example, body weight and gender were identified as the most significant covariates for CL and V_c of bevacizumab while baseline serum albumin and lean body weight were identified as significant covariates for pertuzumab CL [28, 29]. None of these covariates for bevacizumab or pertuzumab are significant enough to require a dose adjustment in the clinic for a specific patient population. The covariate analyses for bevacizumab similarly did not identify any covariates for exposure which would require a dose adjustment. Among 13 covariates evaluated (Table 1), only 3 had a significant impact on either CL or V_c . Body weight impacted both CL and V_c , albumin impacted CL, and gender impacted V_c . Sensitivity analyses indicated that the effects of these covariates on steady-state exposures including $C_{\text{trough,ss}}$, AUC_{ss} , and $C_{\text{max,ss}}$ were small compared to the overall between-subject variability in the population. Therefore, dose adjustment based on any of these parameters is not warranted. Due to the limited number of patients ($n = 75$) evaluated in the PopPK analysis, the covariate relationship will be further evaluated at the end of the phase 2 FIGHT trial. FGF2b expression was tested predose to select patients with GEA in the trial. FGFR2b status at baseline (high vs others) in patients with GEA was not a covariate for PK (Supplementary Fig. 2). In addition, there were no time-dependent changes in PK. Therefore, the PK profile

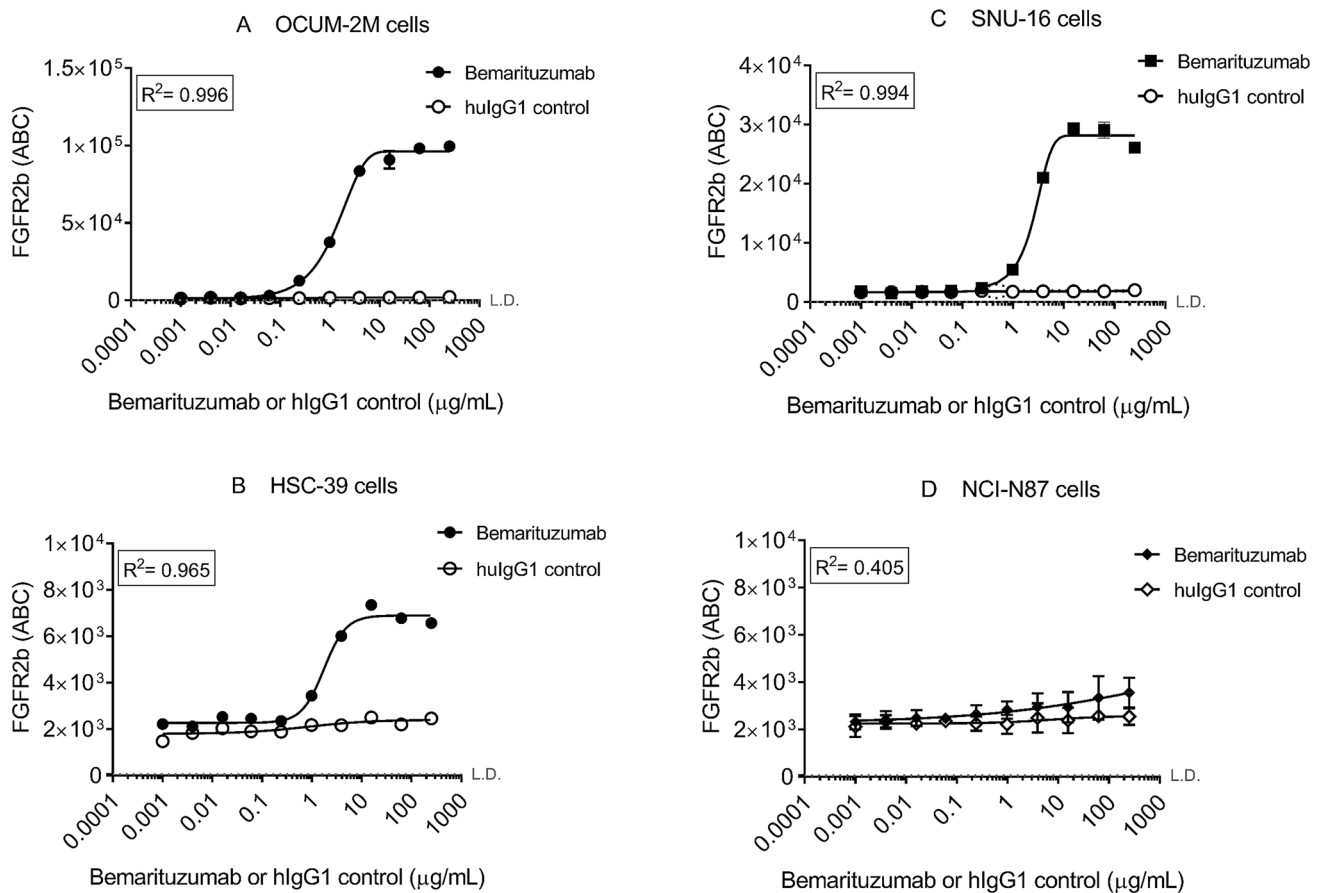


Fig. 3 In vitro cell-based binding for bemarituzumab using gastric cell lines. Mean fluorescence intensity (ABC) versus bemarituzumab concentration profile from receptor occupancy study in vitro are presented for FGFR2b-amplified cell lines OCUM-2 M, HSC-39, and

SNU-16 in Figs. 3A, 3B, and 3C, respectively. NCI-N87 is presented in 3D as a negative control. Symbols represent observed data ($n=3$ per time point). *L.D.* limit of detection, which is at 62 for ABC

should not be affected by changes in FGFR2b expression during treatment. Because no patients developed ADA after administration of bemarituzumab, there is no data to perform analysis of potential ADA impact on PK. When the impact of body weight is considered, gender does not play a significant role to impact PK parameters in general. However, the variability of CL and V_1 for bemarituzumab cannot be fully explained by either body weight or gender and, therefore, both are significant according to the method used for the PopPK analysis and statistical criteria described in the Materials and Methods section. This result suggests that gender had impact beyond body weight although the underlying reason is not understood. This observation was previously reported for other antibodies [28, 30, 31].

The empirical target C_{trough} of 60 $\mu\text{g/mL}$ for bemarituzumab was to achieve >95% receptor occupancy based on in vitro data, including receptor occupancy using multiple FGFR2-amplified gastric cancer cell lines. Although there is no robust clinical data to support the selection of the

empirical target C_{trough} for bemarituzumab, clinical data indicated that at 60 $\mu\text{g/mL}$, >90% of nonlinear clearance was projected to be saturated based on K_m obtained from the PopPK analysis. When K_m data from the clinic is available, it can be used directly to estimate target C_{trough} . In addition, all 6 patients with GEA and confirmed partial responses in the phase 1 monotherapy trial (5 with high FGFR2b overexpression), regardless of dose level, achieved the desired target $C_{\text{trough ss}}$ of $\geq 60 \mu\text{g/mL}$ although no dose-efficacy relationship was identified because of limited data [14 and Five Prime Therapeutics data on file]. It is understood that more clinical data is needed to support the empirical target C_{trough} and it is expected that the exposure–response relationship analysis post FIGHT trial should provide that data. For blocking antibodies, the C_{trough} has generally been used to select the target dose to maximize receptor occupancy throughout the treatment cycles [18–20]. Using trastuzumab as an example, two mechanisms of action are described: inhibition of the proliferation of human tumor cells that overexpress Her2 and ADCC. The minimum desired C_{trough}

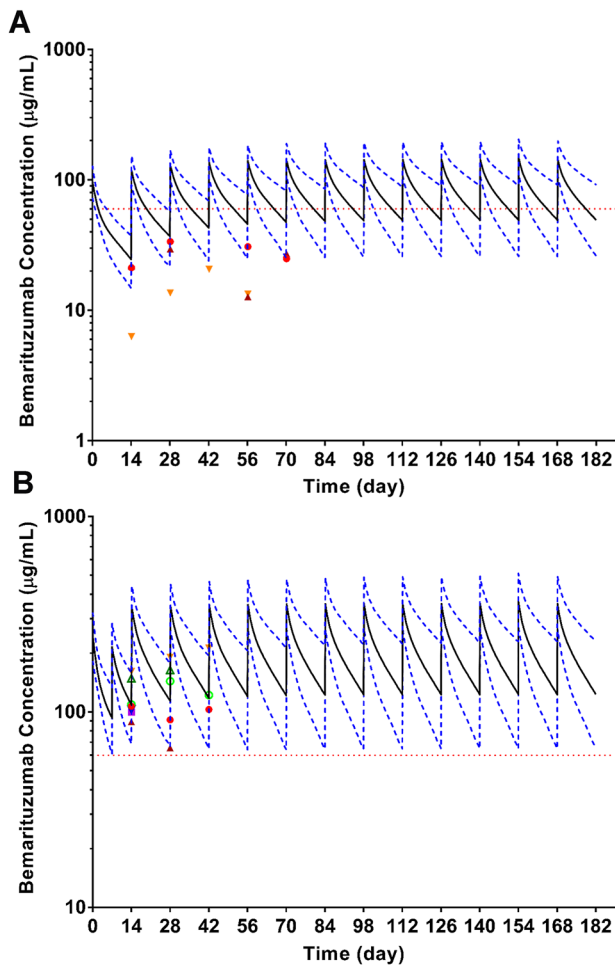


Fig. 4 Simulation results using PK parameters obtained from PopPK analysis versus observed C_{trough} from phase 1 of the FIGHT trial. Simulated serum concentration versus time profiles of bemarituzumab after continuous Q2W dosing of bemarituzumab at 6 mg/kg (a) or 15 mg/kg Q2W with a single dose of 7.5 mg/kg on Cycle 1 Day 8 (b) for 6 months based on the final PopPK model. For each dose regimen, 1000 simulated model parameters constructed the distribution of model predictions for a typical population (61 kg male patients with albumin of 3.7 g/dL). In each figure, solid black line represents the median predicted value and dashed blue lines represent the 5th and 95th percentile of the predicted concentrations and dashed red line is for C_{trough} of 60 $\mu\text{g/mL}$. Each symbol represents the individual bemarituzumab C_{trough} ($\mu\text{g/mL}$) in the presence of mFOL-FOX6 at 6 mg/kg (a, $n=3$) and 15 mg/kg Q2W with a single dose of 7.5 mg/kg on Cycle 1 Day 8 (b, $n=7$), respectively

for trastuzumab in the clinic was $> 10 \mu\text{g/mL}$ based on non-clinical data, which theoretically achieved $\geq 99\%$ receptor saturation in blood based on its affinity for Her2 (0.1 nM) and $< 50\%$ receptor saturation based on its affinity for Fc γ RIIIa (V158) (252 nM) [19, 32, 33], respectively. In the clinic, the observed mean $C_{\text{trough ss}}$ is 79 $\mu\text{g/mL}$, which is projected to achieve 99% receptor saturation of Her2, but not of Fc γ RIIIa [34]. Similar to trastuzumab, bemarituzumab

also has two mechanisms of action: blocking FGFR2b binding to its FGF ligands and enhanced ADCC. Therefore, C_{trough} was used as an exposure target to saturate FGFR2b receptors during the dose interval.

The two dose cohorts in the phase 1 part of the FIGHT trial were 6 mg/kg Q2W and 15 mg/kg Q2W with a single dose of 7.5 mg/kg on Cycle 1 Day 8. As advanced stage GEA is an aggressive disease with standard chemotherapy providing a median of only 6 months of disease control with a worse prognosis expected in patients whose tumors over-express FGFR2b, a method to shorten the time to achieve target C_{trough} levels was deemed clinically important. In the single agent phase 1 FPA144-001 trial, 15 mg/kg was the highest dose tested. Simulations using PopPK parameters were performed to estimate the doses required to achieve target C_{trough} . To maximize efficacy, simulation of the dosing regimen of 15 mg/kg q2w with a single dose of 7.5 mg/kg on Cycle 1 Day 8 showed that 98% of patients could be expected to achieve the target C_{trough} level of 60 $\mu\text{g/mL}$ on Day 15 and 96% of patients could be expected to maintain the target $C_{\text{trough ss}}$. To minimize a potential safety risk which could be associated with an increase in C_{max} , the addition of a single 7.5 mg/kg dose on Day 8 was instituted to maintain the projected C_{max} for this regimen within the level previously shown to be safe during the single agent study FPA144-001. It was anticipated that the addition of a single dose of 7.5 mg/kg on Cycle 1 Day 8 would shorten the time to achieve the target C_{trough} which was thought to be clinically important in the early control of an aggressive tumor such as GEA. Indeed, target C_{trough} was achieved by all patients in the phase 1 portion of the FIGHT trial on Day 15 using this dose and regimen. This PopPK analysis and clinical findings supported the selection of dose and schedule for initiation of the phase 2 randomized, double-blind portion of the global FIGHT trial which is currently ongoing worldwide [24].

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Compliance with ethical standards

Conflict of interest H. Xiang, A. Ahene, L. Dreiling, and H. Collins are employed with Five Prime Therapeutics, Inc. and M. Macal and AW Hsu are former Five Prime employees. They have ownership interest in the Five Prime stock. L. Liu and Y. Gao declare no potential conflicts of interest.

Ethical approval All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

Informed consent Informed consent was obtained from all individual participants included in the study.

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