

HHS Public Access

Author manuscript *Urol Oncol.* Author manuscript; available in PMC 2022 June 01.

Published in final edited form as:

Urol Oncol. 2021 June ; 39(6): 322-326. doi:10.1016/j.urolonc.2020.03.016.

The Role of Heat Shock Protein-90 in the Pathogenesis of Birt-Hogg-Dubé and Tuberous Sclerosis Complex Syndromes

Mark R Woodford^{1,2,3}, Sarah J Backe^{1,2,3}, Rebecca A Sager^{1,2,3,4}, Dimitra Bourboulia^{1,2,3}, Gennady Bratslavsky^{1,2,3}, Mehdi Mollapour^{1,2,3}

¹Department of Urology, SUNY Upstate Medical University, Syracuse, NY, USA

²Department of Biochemistry and Molecular Biology, SUNY Upstate Medical University, Syracuse, NY, USA

³Upstate Cancer Center, SUNY Upstate Medical University, Syracuse, NY, USA

⁴College of Medicine, SUNY Upstate Medical University, Syracuse, NY, USA

Abstract

Birt-Hogg-Dubé (BHD) and tuberous sclerosis (TS) syndromes share many clinical features. These two diseases display distinct histologic subtypes of renal tumors: chromophobe renal cell carcinoma and renal angiomyolipoma, respectively. Early work suggested a role for mTOR dysregulation in the pathogenesis of these two diseases, however their detailed molecular link remains elusive. Interestingly, a growing number of case reports describe renal angiomyolipoma in BHD patients, suggesting a common molecular origin. The BHD-associated proteins FNIP1/2 and the TS protein Tsc1 were recently identified as regulators of the molecular chaperone Hsp90. Dysregulation of Hsp90 activity has previously been reported to support tumorigenesis, providing a potential explanation for the overlapping phenotypic manifestations in these two hereditary syndromes.

Keywords

Birt-Hogg-Dubé (BHD); FLCN; Tuberous Sclerosis Complex (TSC); Renal Angiomyolipoma; Tsc1 (Hamartin); Tsc2 (Tuberin); Heat Shock Protein-90; Chaperones

1. Introduction

Mutations in several distinct tumor suppressor genes are capable of causing hereditary kidney cancer. Numerous genes have been implicated in the dozen categorized histologic

[†]Correspondence: mollapom@upstate.edu.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Disclosure statement

The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed. No writing assistance was utilized in the production of this manuscript.

subtypes of hereditary kidney cancer, a topic that has recently been reviewed in detail [1–3]. Due to the well-established role of the nutrient-sensing kinase mammalian target of rapamycin (mTOR) signaling in normal kidney function [4], kidney cancer research has converged on dysregulation of this pathway. There are, however, unique phenotypic manifestations between diseases, which imply the existence of distinguishing molecular characteristics that may serve as novel therapeutic targets. Many of these hereditary renal cell carcinoma (RCC) syndromes lead to other clinical manifestations in addition to the predisposition to kidney tumor development. Two such syndromes, Birt-Hogg-Dubé syndrome and tuberous sclerosis (TS) syndrome, share many clinical and molecular commonalities.

2. Two Interrelated Subtypes of Kidney Cancer

Birt-Hogg-Dubé (BHD) syndrome is caused by germline mutations in the *FLCN* gene, located on chromosome 17p11.2 [5]. BHD manifests most commonly as cutaneous fibrofolliculomas, pulmonary cysts, spontaneous pneumothorax, and bilateral, multifocal renal cell carcinoma [6]. The most common histologic subtypes of BHD-associated RCC are chromophobe and renal oncocytoma, which occur in approximately one-third of patients [7].

FLCN is characterized as a tumor suppressor protein, though its precise cellular role remains unknown. Loss of *FLCN* has been previously shown to lead to mTOR activation, and opposing works show lysosomal-FLCN is a Rag-interacting protein with GAP activity for RagC/D or RagA/B [8–11]. However, the existence of conflicting data suggests a contextdependent role in mTOR regulation [12, 13]. Interestingly, facial fibrofolliculomas in patients with BHD failed to respond to topical rapamycin treatment in a recent trial [14], reinforcing an mTOR-independent role for FLCN-mediated pathogenesis. Additionally, a trial to evaluate the effect of mTOR inhibition on BHD-associated RCC began recruitment in 2015, however patient enrollment was scant, resulting in premature study termination (ClinicalTrials.gov Identifier: NCT02504892). There are currently no FDA-approved treatments specific to BHD-associated RCC.

Tuberous sclerosis (TS) syndrome is the result of germline mutations in the *TSC1* (chromosome 9q34) or, more commonly, *TSC2* (16p13.3) tumor suppressor genes [15]. Afflicted patients may suffer from TS-associated neurological disorders (TAND) including intellectual disability, autism, seizures, and subependymal giant cell astrocytomas [16] [16], as well as facial angiofibromas, pulmonary lymphangioleiomyomatosis [17], and renal angiomyolipoma (AML) in 50-80% of patients [18, 19].

TSC2 mutations are found in 70-90% of TS cases, identifying Tsc2 as the crucial signaling hub in this disease [20]. Tsc2 exerts its tumor suppressor function by acting as a GTPase activating protein for the mTOR activator Rheb [21]. Loss of Tsc2 promotes mTOR activation and a hyperproliferative phenotype [22]. The recent approval of the mTORC1 inhibitor everolimus (Afinitor®) for TS symptoms is based largely on the findings of the EXIST series of clinical trials and is currently approved as a first-line therapy for SEGA [23], AML [16, 17, 24, 25] and TS-associated seizures [26]. There are also six ongoing clinical trials assessing new therapeutic interventions in TS. Three unique topical mTOR

inhibitors in Phase 2 and 3 trials are under evaluation for the treatment of facial angiofibromas (ClinicalTrials.gov Identifier: NCT03363763, NCT03826628, NCT02860494). Three other studies are examining different strategies for the relief of epilepsy, including vigabatrin for epilepsy prevention in infants (Phase 2; NCT02849457), a Phase 2 trial of everolimus in treatment-resistant epilepsy (NCT02451696) and oral cannabidiol for seizure relief (Phase 3; NCT02544750). Despite this, a subset of patients do not clinically benefit from mTOR inhibition [26], suggesting additional opportunities for therapeutic intervention.

3. Co-incidence of BHD and TS

As described above, BHD and TS syndromes share many similarities in their clinical manifestations and both commonly affect the skin, lung, and kidney. This overlap suggests mutations of *FLCN* and *TSC1/2* contribute to a common pathway dysregulation capable of affecting these organs. In agreement with this, several groups have reported the occurrence of renal angiomyolipoma in patients with BHD [27–30]. The clinical observation of angiofibromas and Koenen's tumor in BHD [31, 32] and fibrofolliculomas in TS [33] further highlight the intimate interconnectivity of these two syndromes (Figure 1). Recent work has demonstrated a role for the molecular chaperone heat shock protein-90 (Hsp90) in the regulation of the pathways implicated in these syndromes. This connection provides a potential explanation for the significant overlap between these diseases.

4. FNIP1/2 and Tsc1 Regulate Hsp90

Heat shock protein-90 (Hsp90) is an ATP-dependent molecular chaperone involved in the stabilization, activation, and maturation of more than 200 intracellular proteins termed 'clients' [34]. The activity and function of Hsp90 is primarily regulated by a class of interacting proteins called co-chaperones. A co-chaperone is broadly defined as a protein that interacts with Hsp90 and influences client binding or activity, but is not itself dependent on Hsp90 [35]. Recent work has identified a critical role for Hsp90 in supporting the stability of the tumor suppressors FLCN and Tsc2. Specifically, the BHD- and TS-associated proteins Folliculin-Interacting Proteins1 and 2 (FNIP1/2) and Tsc1 function as co-chaperones of Hsp90. Broadly, this co-chaperone activity provides Hsp90 functional regulation and results in the stabilization of FLCN and Tsc2. These co-chaperones appear to affect Hsp90 similarly, both by decreasing the rate of ATP hydrolysis as well as providing a scaffold for Hsp90-client interactions [36–40].

FNIP1/2 were first identified in association with FLCN and were shown to be essential for FLCN stability [41, 42]. FNIP1/2 form stable dimers, and the resulting complex binds to the C-terminus of FLCN [41, 42]. Mutations in the *FLCN* gene commonly result in premature truncations of FLCN protein [43, 44], precluding the necessary binding of FNIP1/2. Recent work has unraveled that FNIP1/2 mediate FLCN stability through the Hsp90 chaperone system by acting as regulatory co-chaperones of Hsp90 and providing a scaffold for the chaperoning and stability of FLCN [36].

Tsc1 has long been known to stabilize Tsc2 [45] and was subsequently found to provide protection from HERC1-mediated ubiquitination and proteasomal degradation [46]. As Hsp90 is a crucial mediator of protein stability, these findings suggested a potential role for Hsp90 in the formation of the Tsc1/Tsc2 complex. Indeed, Tsc1 binds to Hsp90 and decreases its ATPase activity, promoting the folding and activity of client proteins. Tsc1 scaffolds Tsc2 to Hsp90, enhancing Tsc2 stability and preventing its ubiquitination and degradation [37]. These data also provide an explanation for the observed tumor suppressive role for Tsc1, as mutations in *TSC1* have been shown to cause TS [47]. In this model, Tsc1 loss prevents Tsc2 binding to Hsp90, leading to its degradation and loss of Tsc1/2 complex tumor suppressor activity.

5. Molecular Underpinnings of Clinical Overlap

An outstanding question in the regulation of Hsp90 concerns the dynamics of co-chaperone occupancy. Due to the sheer number of Hsp90 co-chaperones, there must be both spatial and temporal regulation of their binding to Hsp90. Indeed, many co-chaperones have been shown to either work in concert with or antagonize binding of other co-chaperones to Hsp90 [36, 37]. In light of the relative abundance of clinical reports on BHD and TS co-incidence, our group revisited the idea that Tsc1-Tsc2 and FNIP1/2-FLCN are dedicated, exclusive complexes. In fact, crosstalk between Tsc1 and FNIP1 affects Hsp90-mediated chaperoning of a patient-derived mutant FLCN, which in turn affects chaperoning of Tsc2 [30].

FLCN-L460QsX25 is a disease-associated destabilizing mutation of FLCN; a two base pair deletion leads to a frame-shift and premature stop codon resulting in C-terminal truncation [44, 48]. As such, this mutated protein is unable to bind FNIP1/2. Interestingly however, FLCN-L460QsX25 is capable of interacting with Tsc1, which partially compensates for FNIP1/2 binding and facilitates a low level of FLCN expression. Artificially increasing the expression of mutant FLCN compromises Tsc2 stability likely by occupying Tsc1 thereby, decreasing its availability for chaperoning of Tsc2 [30]. Decreased Tsc2 stability (and thus decreased tumor suppressor activity) potentially explains the finding of renal AML in this BHD patient (Figure 2). Notably, loss of both *FNIP1* and *TSC1* has been shown to synergistically induce mTOR activity [49], a finding that can be explained in part by their roles as Hsp90 co-chaperones [38].

6. Hsp90 is a Potential Therapeutic Target in RCC

Drugs targeting Hsp90 have had a prolonged courtship with therapeutic relevance [50, 51]. Despite their pre-clinical promise, Hsp90 inhibitors have been doomed by poor patient selection, a result of a lack of biomarkers to predict clinical applicability [52]. It has previously been shown that FNIP1/2 and Tsc1 can promote Hsp90 binding to its inhibitors [36, 37, 39]. In fact, recent work has demonstrated that Tsc1 expression promotes acetylation of Hsp90 and sensitizes bladder cancer cells to Hsp90 inhibitors [40], providing further evidence that biomarker discovery of is paramount to the success of Hsp90 inhibitors. As previously described, the requirement of the *FLCN-L460QsX25* mutant for Tsc1 co-chaperone activity likely leads to increased association of Tsc1 with Hsp90. It then follows that these BHD-associated tumors may be hypersensitive to Hsp90 inhibition,

though this remains untested. Interestingly, *FLCN* mutation has also been identified in colon and breast cancers [53]. Perhaps a similar phenomenon contributes to the pathogenesis of these *FLCN*-deficient diseases.

7. Concluding Remarks

Despite the early characterization of BHD and TS as diseases of mTOR dysregulation, the role each protein plays in this process is unclear. Mounting evidence suggests the mTOR suppressive effects of the FNIP/FLCN and Tsc1/2 systems are potentially mediated by the action of molecular chaperones. Further, it is possible that the action of FNIP1/2 and Tsc1 as Hsp90 co-chaperones is independent of mTOR altogether. In either case, the phenotypic convergence of BHD and TS suggest a shared underlying molecular pathology instrumental in the homeostasis of multiple organ systems. Uncovering this link is likely to present more broadly applicable therapeutic options.

Acknowledgments

This work was partly supported by the National Institute of General Medical Sciences of the NIH (grant R01GM124256 to M.M.). The content is solely the responsibility of the authors and does not necessarily represent the official views of the NIH. This work was also supported by funds from SUNY Upstate Medical University, the Upstate Foundation, and in part by the Urology Care Foundation Research Scholar Award Program and American Urological Association (M.M.). The authors are grateful to their colleagues Laura S. Schmidt, W. Marston Linehan for their stimulating discussions.

References

- Ball MW, Pinto PA. Familial and Hereditary Syndromes in Renal Cell Cancer. Renal Cancer2020. p. 35–48.
- [2]. Linehan WM, Schmidt LS, Crooks DR, Wei D, Srinivasan R, Lang M, et al. The Metabolic Basis of Kidney Cancer. Cancer Discov. 2019;9:1006–21. [PubMed: 31088840]
- [3]. Carlo MI, Hakimi AA, Stewart GD, Bratslavsky G, Brugarolas J, Chen YB, et al. Familial Kidney Cancer: Implications of New Syndromes and Molecular Insights. Eur Urol. 2019;76:754–64. [PubMed: 31326218]
- [4]. Fantus D, Rogers NM, Grahammer F, Huber TB, Thomson AW. Roles of mTOR complexes in the kidney: implications for renal disease and transplantation. Nat Rev Nephrol. 2016;12:587–609.
 [PubMed: 27477490]
- [5]. Schmidt LS, Warren MB, Nickerson ML, Weirich G, Matrosova V, Toro JR, et al. Birt-Hogg-Dubé syndrome, a genodermatosis associated with spontaneous pneumothorax and kidney neoplasia, maps to chromosome 17p11.2. Am J Hum Genet. 2001;69:876–82. [PubMed: 11533913]
- [6]. Birt AR, Hogg GR, Dubé WJ. Hereditary multiple fibrofolliculomas with trichodiscomas and acrochordons. Arch Dermatol. 1977;113:1674–7. [PubMed: 596896]
- [7]. Schmidt LS. Birt-Hogg-Dubé syndrome, a genodermatosis that increases risk for renal carcinoma. Curr Mol Med. 2004;4:877–85. [PubMed: 15579035]
- [8]. Lawrence RE, Fromm SA, Fu Y, Yokom AL, Kim DJ, Thelen AM, et al. Structural mechanism of a Rag GTPase activation checkpoint by the lysosomal folliculin complex. Science. 2019;366:971–7. [PubMed: 31672913]
- [9]. Petit CS, Roczniak-Ferguson A, Ferguson SM. Recruitment of folliculin to lysosomes supports the amino acid-dependent activation of Rag GTPases. J Cell Biol. 2013;202:1107–22. [PubMed: 24081491]
- [10]. Tsun ZY, Bar-Peled L, Chantranupong L, Zoncu R, Wang T, Kim C, et al. The folliculin tumor suppressor is a GAP for the RagC/D GTPases that signal amino acid levels to mTORC1. Mol Cell. 2013;52:495–505. [PubMed: 24095279]

- [11]. Shen K, Rogala KB, Chou HT, Huang RK, Yu Z, Sabatini DM. Cryo-EM Structure of the Human FLCN-FNIP2-Rag-Ragulator Complex. Cell. 2019;179:1319–29 e8. [PubMed: 31704029]
- [12]. Hudon V, Sabourin S, Dydensborg AB, Kottis V, Ghazi A, Paquet M, et al. Renal tumour suppressor function of the Birt-Hogg-Dubé syndrome gene product folliculin. J Med Genet. 2010;47:182–9. [PubMed: 19843504]
- [13]. Khabibullin D, Medvetz DA, Pinilla M, Hariharan V, Li C, Hergrueter A, et al. Folliculin regulates cell-cell adhesion, AMPK, and mTORC1 in a cell-type-specific manner in lung-derived cells. Physiol Rep. 2014;2.
- [14]. Gijezen LM, Vernooij M, Martens H, Oduber CE, Henquet CJ, Starink TM, et al. Topical rapamycin as a treatment for fibrofolliculomas in Birt-Hogg-Dubé syndrome: a double-blind placebo-controlled randomized split-face trial. PLoS One. 2014;9:e99071. [PubMed: 24910976]
- [15]. Henske EP, Jozwiak S, Kingswood JC, Sampson JR, Thiele EA. Tuberous sclerosis complex. Nat Rev Dis Primers. 2016;2:16035. [PubMed: 27226234]
- [16]. Kwiatkowski DJ, Palmer MR, Jozwiak S, Bissler J, Franz D, Segal S, et al. Response to everolimus is seen in TSC-associated SEGAs and angiomyolipomas independent of mutation type and site in TSC1 and TSC2. Eur J Hum Genet. 2015;23:1665–72. [PubMed: 25782670]
- [17]. Kingswood JC, Jozwiak S, Belousova ED, Frost MD, Kuperman RA, Bebin EM, et al. The effect of everolimus on renal angiomyolipoma in patients with tuberous sclerosis complex being treated for subependymal giant cell astrocytoma: subgroup results from the randomized, placebocontrolled, Phase 3 trial EXIST-1. Nephrol Dial Transplant. 2014;29:1203–10. [PubMed: 24729041]
- [18]. Kingswood JC, Belousova E, Benedik MP, Carter T, Cottin V, Curatolo P, et al. Renal angiomyolipoma in patients with tuberous sclerosis complex: findings from the TuberOus SClerosis registry to increase disease Awareness. Nephrol Dial Transplant. 2019;34:502–8. [PubMed: 29697822]
- [19]. Rakowski SK, Winterkorn EB, Paul E, Steele DJ, Halpern EF, Thiele EA. Renal manifestations of tuberous sclerosis complex: Incidence, prognosis, and predictive factors. Kidney Int. 2006;70:1777–82. [PubMed: 17003820]
- [20]. Rosset C, Netto CBO, Ashton-Prolla P. TSC1 and TSC2 gene mutations and their implications for treatment in Tuberous Sclerosis Complex: a review. Genet Mol Biol. 2017;40:69–79. [PubMed: 28222202]
- [21]. Inoki K, Li Y, Xu T, Guan KL. Rheb GTPase is a direct target of TSC2 GAP activity and regulates mTOR signaling. Genes Dev. 2003;17:1829–34. [PubMed: 12869586]
- [22]. El-Hashemite N, Zhang H, Henske EP, Kwiatkowski DJ. Mutation in TSC2 and activation of mammalian target of rapamycin signalling pathway in renal angiomyolipoma. Lancet. 2003;361:1348–9. [PubMed: 12711473]
- [23]. Turner SG, Peters KB, Vredenburgh JJ, Desjardins A, Friedman HS, Reardon DA. Everolimus tablets for patients with subependymal giant cell astrocytoma. Expert Opin Pharmacother. 2011;12:2265–9. [PubMed: 21806479]
- [24]. Bissler JJ, Kingswood JC, Radzikowska E, Zonnenberg BA, Frost M, Belousova E, et al. Everolimus for angiomyolipoma associated with tuberous sclerosis complex or sporadic lymphangioleiomyomatosis (EXIST-2): a multicentre, randomised, double-blind, placebocontrolled trial. Lancet. 2013;381:817–24. [PubMed: 23312829]
- [25]. Bissler JJ, Kingswood JC, Radzikowska E, Zonnenberg BA, Belousova E, Frost MD, et al. Everolimus long-term use in patients with tuberous sclerosis complex: Four-year update of the EXIST-2 study. PLoS One. 2017;12:e0180939. [PubMed: 28792952]
- [26]. French JA, Lawson JA, Yapici Z, Ikeda H, Polster T, Nabbout R, et al. Adjunctive everolimus therapy for treatment-resistant focal-onset seizures associated with tuberous sclerosis (EXIST-3): a phase 3, randomised, double-blind, placebo-controlled study. Lancet. 2016;388:2153–63. [PubMed: 27613521]
- [27]. Byrne M, Mallipeddi R, Pichert G, Whittaker S. Birt-Hogg-Dubé syndrome with a renal angiomyolipoma: further evidence of a relationship between Birt-Hogg-Dubé syndrome and tuberous sclerosis complex. Australas J Dermatol. 2012;53:151–4. [PubMed: 22571569]

- [28]. Tobino K, Seyama K. Birt-Hogg-Dubé syndrome with renal angiomyolipoma. Intern Med. 2012;51:1279–80. [PubMed: 22687807]
- [29]. Dow E, Winship I. Renal angiomyolipoma in Birt-Hogg-Dubé syndrome: A case study supporting overlap with tuberous sclerosis complex. Am J Med Genet A. 2016;170:3323–6. [PubMed: 27643397]
- [30]. Sager RA, Woodford MR, Shapiro O, Mollapour M, Bratslavsky G. Sporadic renal angiomyolipoma in a patient with Birt-Hogg-Dubé: chaperones in pathogenesis. Oncotarget. 2018;9:22220–9. [PubMed: 29774133]
- [31]. Schaffer JV, Gohara MA, McNiff JM, Aasi SZ, Dvoretzky I. Multiple facial angiofibromas: a cutaneous manifestation of Birt-Hogg-Dubé syndrome. J Am Acad Dermatol. 2005;53:S108–11. [PubMed: 16021156]
- [32]. DiCicco B, Johnson W, Allred J, Soldano AC, Ramsdell WM. Koenen's tumor and facial angiofibromas in a case of Birt-Hogg-Dube syndrome: A cutaneous contribution to growing evidence of a relationship with tuberous sclerosis complex. JAAD Case Rep. 2016;2:196–8. [PubMed: 27274535]
- [33]. Misago N, Joh K, Yatsuki H, Soejima H, Narisawa Y. A BHD germline mutation identified in an Asian family with Birt-Hogg-Dubé syndrome. Acta Derm Venereol. 2008;88:423–5. [PubMed: 18709329]
- [34]. Schopf FH, Biebl MM, Buchner J. The HSP90 chaperone machinery. Nat Rev Mol Cell Biol. 2017;18:345–60. [PubMed: 28429788]
- [35]. Miyata Y, Nakamoto H, Neckers L. The therapeutic target Hsp90 and cancer hallmarks. Curr Pharm Des. 2013;19:347–65. [PubMed: 22920906]
- [36]. Woodford MR, Dunn DM, Blanden AR, Capriotti D, Loiselle D, Prodromou C, et al. The FNIP co-chaperones decelerate the Hsp90 chaperone cycle and enhance drug binding. Nat Commun. 2016;7:12037. [PubMed: 27353360]
- [37]. Woodford MR, Sager RA, Marris E, Dunn DM, Blanden AR, Murphy RL, et al. Tumor suppressor Tsc1 is a new Hsp90 co-chaperone that facilitates folding of kinase and non-kinase clients. EMBO J. 2017;36:3650–65. [PubMed: 29127155]
- [38]. Sager RA, Woodford MR, Mollapour M. The mTOR Independent Function of Tsc1 and FNIPs. Trends Biochem Sci. 2018;43:935–7. [PubMed: 30361061]
- [39]. Sager RA, Woodford MR, Backe SJ, Makedon AM, Baker-Williams AJ, DiGregorio BT, et al. Post-translational Regulation of FNIP1 Creates a Rheostat for the Molecular Chaperone Hsp90. Cell Rep. 2019;26:1344–56 e5. [PubMed: 30699359]
- [40]. Woodford MR, Hughes M, Sager RA, Backe SJ, Baker-Williams AJ, Bratslavsky MS, et al. Mutation of the co-chaperone Tsc1 in bladder cancer diminishes Hsp90 acetylation and reduces drug sensitivity and selectivity. Oncotarget. 2019;10:5824–34. [PubMed: 31645902]
- [41]. Baba M, Hong SB, Sharma N, Warren MB, Nickerson ML, Iwamatsu A, et al. Folliculin encoded by the BHD gene interacts with a binding protein, FNIP1, and AMPK, and is involved in AMPK and mTOR signaling. Proc Natl Acad Sci U S A. 2006;103:15552–7. [PubMed: 17028174]
- [42]. Hasumi H, Baba M, Hong SB, Hasumi Y, Huang Y, Yao M, et al. Identification and characterization of a novel folliculin-interacting protein FNIP2. Gene. 2008;415:60–7. [PubMed: 18403135]
- [43]. Nahorski MS, Reiman A, Lim DH, Nookala RK, Seabra L, Lu X, et al. Birt Hogg-Dube syndrome-associated FLCN mutations disrupt protein stability. Hum Mutat. 2011;32:921–9. [PubMed: 21538689]
- [44]. Schmidt LS, Nickerson ML, Warren MB, Glenn GM, Toro JR, Merino MJ, et al. Germline BHDmutation spectrum and phenotype analysis of a large cohort of families with Birt-Hogg-Dubé syndrome. Am J Hum Genet. 2005;76:1023–33. [PubMed: 15852235]
- [45]. Benvenuto G, Li S, Brown SJ, Braverman R, Vass WC, Cheadle JP, et al. The tuberous sclerosis-1 (TSC1) gene product hamartin suppresses cell growth and augments the expression of the TSC2 product tuberin by inhibiting its ubiquitination. Oncogene. 2000;19:6306–16. [PubMed: 11175345]

- [46]. Chong-Kopera H, Inoki K, Li Y, Zhu T, Garcia-Gonzalo FR, Rosa JL, et al. TSC1 stabilizes TSC2 by inhibiting the interaction between TSC2 and the HERC1 ubiquitin ligase. J Biol Chem. 2006;281:8313–6. [PubMed: 16464865]
- [47]. Hoogeveen-Westerveld M, Ekong R, Povey S, Karbassi I, Batish SD, den Dunnen JT, et al. Functional assessment of TSC1 missense variants identified in individuals with tuberous sclerosis complex. Hum Mutat. 2012;33:476–9. [PubMed: 22161988]
- [48]. Toro JR, Wei MH, Glenn GM, Weinreich M, Toure O, Vocke C, et al. BHD mutations, clinical and molecular genetic investigations of Birt-Hogg-Dubé syndrome: a new series of 50 families and a review of published reports. J Med Genet. 2008;45:321–31. [PubMed: 18234728]
- [49]. Centini R, Tsang M, Iwata T, Park H, Delrow J, Margineantu D, et al. Loss of Fnip1 alters kidney developmental transcriptional program and synergizes with TSC1 loss to promote mTORC1 activation and renal cyst formation. PLoS One. 2018;13:e0197973. [PubMed: 29897930]
- [50]. Neckers L, Workman P. Hsp90 molecular chaperone inhibitors: are we there yet? Clin Cancer Res. 2012;18:64–76. [PubMed: 22215907]
- [51]. Sanchez J, Carter TR, Cohen MS, Blagg BS. Old and New Approaches to Target the Hsp90 Chaperone. Curr Cancer Drug Targets. 2019.
- [52]. Chehab M, Caza T, Skotnicki K, Landas S, Bratslavsky G, Mollapour M, et al. Targeting Hsp90 in urothelial carcinoma. Oncotarget. 2015;6:8454–73. [PubMed: 25909217]
- [53]. Palmirotta R, Savonarola A, Ludovici G, Donati P, Cavaliere F, ML DEM, et al. Association between Birt Hogg Dubé syndrome and cancer predisposition. Anticancer Res. 2010;30:751–7. [PubMed: 20392993]



Figure 1.

Clinical manifestations of BHD and TS. Birt-Hogg-Dubé syndrome (BHD) is commonly characterized by cutaneous fibrofolliculomas, spontaneous pneumothorax, and multifocal, bilateral renal tumors of chromophobe histology (blue), while Tuberous Sclerosis (TS) is most often associated with neurological impairment and epilepsy, facial angiofibromas, pulmonary lymphangioleiomyomatosis [17], and renal angiomyolipoma (AML) (red). The identification of angiofibromas and angiomyolipoma in BHD patients and fibrofolliculomas in a TS patient suggest a molecular overlap between these two syndromes. Histology:

Kidney tissue stained with hematoxylin and eosin. Chromophobe (BHD), angiomyolipoma (TS). Scale bar = 50μ M. Imaging: Abdominal CT with contrast. BHD: Asterisks represent bilateral, multifocal renal cell carcinoma. TS: A hash (#) denotes the renal AML. This lesion contains some enhancing components but also areas of macroscopic fat (~-30 HU on non-contrasted scan). Arrowheads denote numerous subcentimeter hypodense lesions through the kidneys bilaterally which likely represent renal cysts.



Figure 2.

Tumor suppressor regulation in renal cancer. The tumor suppressive activity of FLCN and Tsc2 is supported by the Hsp90 chaperone. FNIP1/2 and Tsc1 co-chaperones scaffold these Hsp90 clients, and together these complexes act to inhibit mTOR. Mutations in the Hsp90 clients FLCN and Tsc2 cause Birt-Hogg-Dubé (BHD) and Tuberous Sclerosis (TS) syndromes, respectively. The patient mutation FLCN-L460Qfsx25 cannot bind to FNIP1/2 and is degraded, though its expression is partially rescued by the Tsc1-Hsp90 chaperone complex. This compromises Tsc2 stability, leading to the clinical manifestation of angiomyolipoma in BHD patients.