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OPEN New fossil from mid-Cretaceous **Burmese amber confirms** monophyly of Liadopsyllidae (Hemiptera: Psylloidea)

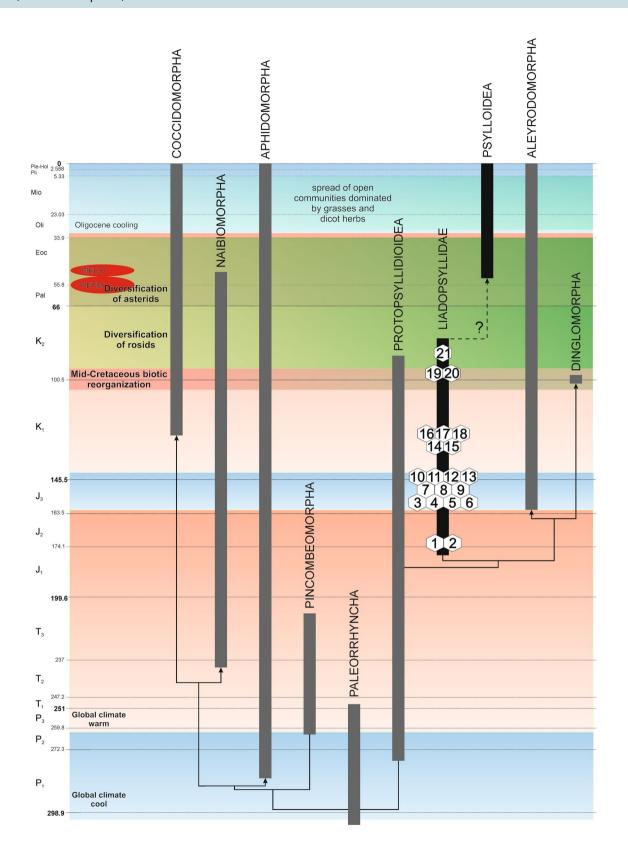
Jowita Drohojowska^{©1™}, Jacek Szwedo^{©2™}, Patrick Müller³ & Daniel Burckhardt^{©4}

Amecephala pusilla gen. et sp. nov. is described and illustrated on the basis of a well-preserved female psyllid (Liadopsyllidae) in a piece of Cretaceous Myanmar amber. The new genus differs from other members of Liadopsyllidae in details of the antennae and forewings. For the first time, the presence of a circumanal ring is documented for Mesozoic psyllids. Based on differences in the length of female terminalia, it is suggested that Liadopsyllidae may have displayed a diversified oviposition biology. As far as known, Liadopsyllidae lack a pulvillus, a putative autapomorphy supporting the monophyly of Liadopsyllidae. An identification key to genera and an annotated checklist of known Liadopsyllidae species are provided. New synonyms and combinations are proposed and the status of the subfamily Miralinae is discussed.

Psyllids or jumping plant-lice are a group of small, generally host-specific plant-sap sucking insects with around 4000 described species¹. A few species are major pests on fruits or vegetables, mostly by transmitting plant pathogens. Others damage forest plantations or ornamental plants by removal of plant-sap, stunting new growth, inducing galls or secreting honeydew and wax, an ideal substrate for sooty mould which reduces photosynthesis² Modern psyllids, defined by the enlarged and immobile metacoxae in adults allowing them to jump, display a wide range of morphological diversity regarding the head, antennae, legs, forewings, terminalia, etc. in adults and body shape, antennal structure and the type of setae or wax pores in immatures. Modern psyllids are documented in the fossil record since the Eocene (Lutetian)³ (Fig. 1). The stem-group of modern psyllids constitutes, according to Burckhardt & Poinar, 2019⁴, the paraphyletic Liadopsyllidae Martynov, 1926⁵ with 17 species and six genera (Liadopsylla Handlirsch, 1925⁶, Gracilinervia Becker-Migdisova, 1985⁷, Malmopsylla Becker-Migdisova, 19857, Mirala Burckhardt & Poinar, 20194, Neopsylloides Becker-Migdisova, 19857 and Pauropsylloides Becker-Migdisova, 1985⁷) from early Jurassic to late Cretaceous^{4,8}. Shcherbakov⁹ added three species from the Lower Cretaceous for one of which he erected the genus Stigmapsylla and for the other two the subgenus Liadopsylla (Basicella). He also transferred two previously described species from Liadopsylla to Cretapsylla Shcherbakov⁹. Further he resurrected the Malmopsyllidae Becker-Migdisova, 1985⁷ splitting it into Malmopsyllinae (for Gracilinervia, Malmopsylla, Neopsylloides and Pauropsylloides) and Miralinae Shcherbakov⁹ (for Mirala). Apart from three species described from amber fossils, all Mesozoic psyllids are poorly preserved impression fossils of which usually only the forewing is preserved. The current classification of Mesozoic psyllids (Liadopsyllidae and Malmopsyllidae) is based almost exclusively upon forewing characters^{7,9}, despite that several phylogenetically significant characters from other body parts have been described from amber inclusions^{4,8}. Judging from the impression fossils, Liadopsyllidae and Malmopsyllidae appear morphologically quite homogeneous but this may be a result of the surprisingly scarce fossil record of psyllids compared to other insect groups. The discoveries of Cretaceous amber fossils radically alter this picture, e.g. the recently described Mirala burmanica Burckhardt & Poinar, 2019 from Myanmar amber⁴.

Here we describe a second taxon of Mesozoic psyllids from Kachin amber, Amecephala pusilla gen. et sp. nov., possessing a series of characters unique within Mesozoic psyllids, discuss the phylogenetic relationships within the group, and provide an updated key to genera as well a checklist of recognised species (Table 1).

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∢Figure 1. Relationships and stratigraphic distribution of Liadopsyllidae and its subunits within Sternorrhyncha according to Drohojowska & Szwedo¹⁰, Hakim et al.¹¹ and Drohojowska et al.¹², modified. Numbers denote described taxa of fossil Liadopsyllidae—1: Liadopsylla geinitzi Handlirsch, 1925—Lower Jurassic, Mecklenburg, Germany, 2: Liadopsylla obtusa Ansorge, 1996—Lower Jurassic, Mecklenburg-Vorpommern, Germany, 3: Liadopsylla asiatica Becker-Migdisova, 1985-Upper Jurassic, Karatau, Kazakhstan, 4: Liadopsylla brevifurcata Becker-Migdisova, 1985—Upper Jurassic, Karatau, Kazakhstan, 5: Liadopsylla grandis Becker-Migdisova, 1985—Upper Jurassic, Karatau, Kazakhstan, 6. Liadopsylla karatavica Becker-Migdisova, 1985—Upper Jurassic, Karatau, Kazakhstan, 7. Liadopsylla longiforceps Becker-Migdisova, 1985—Upper Jurassic, Karatau, Kazakhstan, 8. Liadopsylla tenuicornis Martynov, 1926—Upper Jurassic, Karatau, Kazakhstan, 9. Liadopsylla turkestanica Becker-Migdisova, 1949-Upper Jurassic, Karatau, Kazakhstan, 10. Gracilinervia mastimatoides Becker-Migdisova, 1985—Upper Jurassic, Karatau, Kazakhstan, 11. Malmopsylla karatavica Becker-Migdisova, 1985 – Upper Jurassic, Karatau, Kazakhstan, 12. Neopsylloides turutanovae Becker-Migdisova, 1985—Upper Jurassic, Karatau, Kazakhstan, 13. Pauropsylloides jurassica Becker-Migdisova, 1985—Upper Jurassic, Karatau, Kazakhstan, 14. Liadopsylla mongolica Shcherbakov, 1988—Lower Cretaceous, Bon Tsagaan, Mongolia 15. Liadopsylla apedetica Ouvrard, Burckhardt et Azar, 2010—Lower Cretaceous, Lebanon, 16. Liadopsylla lautereri (Shcherbakov, 2020)—Lower Cretaceous, Buryatia, Russia 17. Liadopsylla loginovae (Shcherbakov, 2020)— Lower Cretaceous, Buryatia, Russia 18. Stigmapsylla klimaszewskii Shcherbakov, 2020—Lower Cretaceous, Burvatia, Russia 19. Mirala burmanica Burckhardt et Poinar, 2019—mid-Cretaceous, Kachin amber, 20. Amecephala pusilla gen. et sp. nov.—mid-Cretaceous, Kachin amber, 21. Liadopsylla hesperia Ouvrard et Burckhardt, 2010-Upper Cretaceous, Raritan amber, U.S.A.

To satisfy a requirement by Article 8.5.3 of the International Code of Zoological Nomenclature this publication has been registered in ZooBank with the LSID: urn:lsid:zoobank.org:act:D3AF7597-47BF-4D6C-9020-982F4C20315E.

Systematic palaeontology. Order Hemiptera Linnaeus, 1758¹⁷
Suborder Sternorrhyncha Amyot et Audinet-Serville, 1843¹⁸
Superfamily Psylloidea Latreille, 1807¹⁹
Family Liadopsyllidae Martynov, 1926⁵

Genus †Amecephala gen. nov. urn:lsid:zoobank.org:act:9DABC236-FFB9-4305-82EC-4E293212849B

Type species. † *Amecephala pusilla* sp. nov., by present designation and monotypy. *Etymology* From ancient Greek ἡ άμε [ē áme] = shovel and ἡ κεφαλή [ē kefalé] = head for its shovel-shaped head. Gender: feminine.

Diagnosis. Vertex rectangular; coronal suture developed in apical half; median ocellus on ventral side of head, situated at the apex of frons which is large, triangular; genae not produced into processes; toruli oval, medium sized, situated in front of eyes below vertex. Eyes hemispheric, relatively small (Fig. 2a,b,e,g). Antenna with pedicel about as long as flagellar segments 1 and 8, longer than remainder of segments. Pronotum ribbon-shaped, relatively long, laterally of equal length as medially. Forewing (Fig. 2a,b,f,g) elongate, widest in the middle, narrowly rounded at apex; pterostigma short and broad, triangular, not delimited at base by a vein thus vein R_1 not developed; veins R and M+Cu subequal in length; vein R relatively short, slightly curved towards fore margin; vein R shorter than its branches which are of subequal length; cell cu_1 low and very long. Female terminalia short, cuneate.

Description. Head weakly inclined from longitudinal body axis; about as wide as pronotum and mesoscutum, dorso-ventrally compressed. Vertex rectangular; anterior margin weakly curved, indented in the middle; posterior margin slightly concavely curved; coronal suture developed in apical half, basal half not visible; lateral ocelli near posterior angles of vertex, hardly raised; median ocellus on ventral side of head, situated at the apex of frons which is large, triangular; genae not produced into processes; preocular sclerites lacking; toruli oval, medium sized, situated in front of eyes below vertex; clypeus partly covered by gas bubble, appearing flattened, pear-shaped. Eyes hemispheric, relatively small (Fig. 2a,b,e,g). Antenna 10-segmented, filiform, moderately long, flagellum 1.6 times as long as head width; pedicel very long, about as long as flagellar segments 1 and 8; rhinaria not visible (Fig. 2a,b). Thorax (ventrally not visible) with pronotum wider than mesopraescutum as wide as mesoscutum, laterally of the same length as medially. Mesothorax large; mesopraescutum triangular, with arcuate anterior margin, almost twice wider than long in the middle; mesopraescutum slightly longer than pronotum in the middle; mesoscutum subtrapezoid with slightly arched anterior margin, about 3.0 times wider than long in the middle; delimitation between mesoscutum and mesoscutellum clearly visible. Metascutellum trapezoid, narrower than mesoscutellum with a submedian longitudinal low ridge on either side. Parapterum and tegula forming small oval structures of about the same size; the former slightly in front of the latter. Forewing (Fig. 2a,b,f) membranous, elongate, narrow at base, widest in the middle, narrowly rounded at apex which lies in cell m₁ near the apex of vein M₃₊₄; vein C+Sc narrow; cell c+sc long, widening toward apex; costal break not visible, perhaps absent; pterostigma short and broad, triangular, not delimited at base by a vein thus vein R₁ not developed; vein R+M+Cu relatively short; veins R and M+Cu subequal in length; vein R₂ relatively short and straight; vein Rs relatively short, slightly curved towards fore margin; vein M shorter than its branches which

Taxon	Locality	Age and formation	Preservation	Sex						
Amecephala gen. nov										
A. pusilla sp. nov.	Myanmar, Kachin State, Hukawng Valley, SW of Maingkhwan, Noije Bum 2001 Summit Site amber mine	Mid-Cretaceous (Aptian/Cenomanian)	Amber inclusion	Female						
Gracilinervia Becker-Migdisova, 1985 ⁷ , p.	Gracilinervia Becker-Migdisova, 1985 ⁷ , p. 78									
<i>G. mastigimatoides</i> Becker-Migdisova, 1985 ⁷ , p.s 79	Kazakhstan, Karatau, Kasharata (Mikhailovka), Aulie	Upper Jurassic (Callovian); Karabastau Fm	Compression	Unknown						
Liadopsylla Handlirsch, 1921 ⁶ , p. 213	Liadopsylla Handlirsch, 1921 ⁶ , p. 213									
<i>L. apedetica</i> Ouvrard, Burckhardt et Azar, 2010 in Ouvrard et al. 2010 ⁸ , p. 173	Lebanon, Mdeyrij-Hammana, Casa Baabda	Lower Cretaceous (late Barremian); Grès du Liban Fm	Amber inclusion	Female						
<i>L. asiatica</i> Becker-Migdisova, 1985 ⁷ , p. 74	Kazakhstan, Karatau, Kasharata (Mikhailovka), Aulie	Upper Jurassic (Callovian); Karabastau Fm	Compression	Unknown						
<i>L. brevifurcata</i> Becker-Migdisova, 1985 ⁷ , p. 69	Kazakhstan, Karatau, Kasharata (Mikhailovka), Aulie	Upper Jurassic (Callovian); Karabastau Fm	Impression	Unknown						
L. geinitzi Handlirsch, 1921 ⁶ , p. 213	Germany, Mecklenburg, Dobbertin	Lower Jurassic (early Toarcian); Har- poceras falciferum ammonoid zone	Impression	Unknown						
<i>L. grandis</i> Becker-Migdisova, 1985 ⁷ , p. 63	isova, 1985 ⁷ , p. 63 Kazakhstan, Karatau, Kasharata (Mikhailovka), Aulie Upper Jurassic (Callovian); Karabastau Fm		Compression	Male						
<i>L. hesperia</i> Ouvrard et Burckhardt, 2010 in Ouvrard et al. 2010 ⁸ , p. 175	U.S.A., New Jersey, Middlesex County, Sayreville	Upper Cretaceous (Turonian); Raritan Fm	Amber inclusion	Female?						
<i>L. karatavica</i> Becker-Migdisova, 1985 ⁷ , p. 73	Kazakhstan, Karatau, Kasharata (Mikhailovka), Aulie	Upper Jurassic (Callovian); Karabastau Fm	Compression	Male						
L. lautereri (Shcherbakov, 2020) 9, p. 130	Russia, SW Buryatia Zakamensk district, Khasurty, 10 km S of Tsakir	Lower Cretaceous (Aptian); Gusinoe Ozero Gr	Impression	Unknown						
L. loginovae (Shcherbakov, 2020) 9, p. 132	Russia, SW Buryatia Zakamensk district, Khasurty, 10 km S of Tsakir	Lower Cretaceous (Aptian); Gusinoe Ozero Gr	Impression	Unknown						
<i>L. longiforceps</i> (Becker-Migdisova, 1985) ⁷ , p. 61	Kazakhstan, Karatau, Kasharata (Mikhailovka), Aulie	Upper Jurassic (Callovian); Karabastau Fm	Compression	Unknown						
L. mongolica Shcherbakov, 1988 ¹³ , p. 61	Mongolia, Bon Tsagaan	Lower Cretaceous (Aptian); Dzun-Bain Fm., Khurilt Mb	Impression	Unknown						
L. obtusa Ansorge, 1996 ¹⁴ , p. 55	Germany, Mecklenburg-Vorpommern, Grimmen, Klein Lehmhagen pit	Lower Jurassic (early Toarcian); Dactyli- oceras tenuicostatum zone	Impression	Unknown						
L. tenuicornis Martynov, 1926 ⁵ , p. 1359	Kazakhstan, Karatau, Kasharata (Mikhailovka), Aulie	Upper Jurassic (Callovian); Karabastau Fm	Compression	Male						
<i>L. turkestanica</i> Becker-Migdisova, 1949 ¹⁵ , p. 42	Kazakhstan, Karatau, Kasharata (Mikhailovka), Aulie	Upper Jurassic (Callovian); Karabastau Fm	Compression	4 Males, 4 females						
${\it Malmopsylla}~Becker-Migdisova, 1985^7, p.$	76									
<i>M. karatavica</i> Becker-Migdisova, 1985 ⁷ : 76	Kazakhstan, Karatau, Kasharata (Mikhailovka), Aulie	Upper Jurassic (Callovian); Karabastau Fm	Impression	Unknown (forewing only)						
Mirala Burckhardt et Poinar, 2019 ⁴ , p. 3				'						
<i>M. burmanica</i> Burckhardt et Poinar, 2019 ⁴ , p. 3	Myanmar, Kachin State, Hukawng Valley, SW of Maingkhwan, Noije Bum 2001 Summit Site amber mine	Mid-Cretaceous (Aptian/Cenomanian)	Amber inclusion	Unknown						
Neopsylloides Becker-Migdisova, 19857, p.	77									
N. turutanovae Becker-Migdisova, 1985 ⁷ , p. 77	Kazakhstan, Karatau, Kasharata (Mikhailovka), Aulie	Upper Jurassic (Callovian); Karabastau Fm	Compression	Unknown						
Pauropsylloides Becker-Migdisova, 1985 ⁷ , p. 79										
<i>P. jurassica</i> Becker-Migdisova, 1985 ⁷ , p. 79	Kazakhstan, Karatau, Kasharata (Mikhailovka), Aulie	Upper Jurassic (Callovian); Karabastau Fm	Compression	Unknown						
Stigmapsylla Shcherbakov, 2020 ⁹ , p. 129										
S. klimaszewskii Shcherbakov, 2020 ⁹ : 130	Russia, SW Buryatia Zakamensk district, Khasurty, 10 km S of Tsakir	Lower Cretaceous (Aptian); Gusinoe Ozero Gr	Impression	Unknown						

Table 1. Annotated checklist of known species of Liadopsyllidae Martynov, 1926⁵. All known specimens are adults. Becker-Migdisova (1985, p. 62)⁷ synonymised *Asientomum* Martynov, 1926⁵, p. 1364 (replacement name for *Lithentomum* Martynov, 1926⁵, p. 1365, *nec* Scudder, 1867¹⁶, p. 206) with *Liadopsylla* and transferred *Lithentomum praecox* Martynov, 1926⁵, p. 1365, to *Liadopsylla*. The venation of the forewing and particularly of the hindwing as described by Martynov⁵ clearly places this species in the Psocodea. Pending an examination of the holotype we follow Martynov⁵ rather than Becker-Migdisova⁷ in this matter.

are of subequal length; vein Cu short, splitting into very long Cu_{1a} and short Cu_{1b} , hence cell cu_1 low and very long; claval suture visible (Fig. 2h,i); anal break near to apex of vein Cu_{1b} (Fig. 2f,i). Hindwing (Fig. 2a) shorter than forewing, more than twice as long as wide, membranous; venation indistinct. Legs similar in shape and size, long, slender (Fig. 2c,d,g); femora slightly enlarged distally, tibiae long and slightly enlarged distally; metatibia lacking genual spine and apical sclerotized spurs, but bearing several apical bristles and, in distal quarter, a row

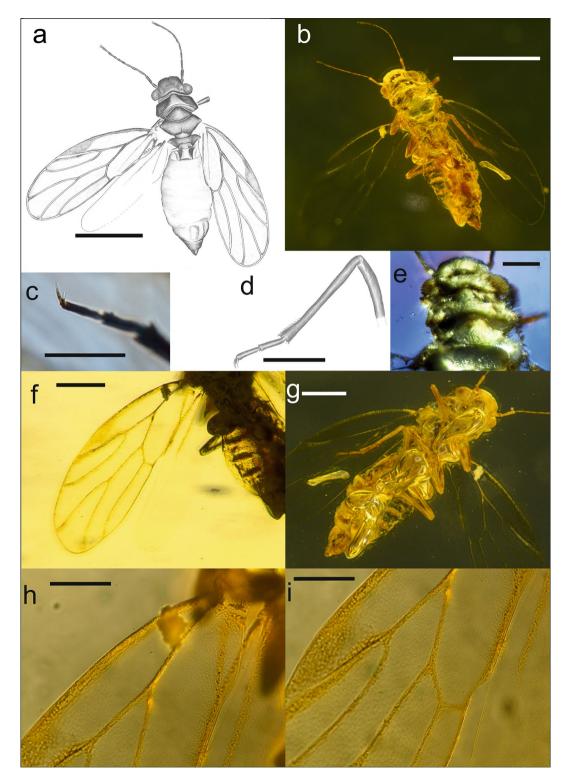


Figure 2. (**a**–**i**) *Amecephala pusilla* **gen. et sp. nov**. imago. Drawing of body in dorsal view (**a**), Body in dorsal view (**b**), Metatarsus (**c**), Drawing of hind leg (**d**), Head in dorsal view (**e**), Forewing (**f**), Body in ventral view (**g**), Basal part of claval suture (**h**), Distal part of claval suture (**i**); Scale bars: 0.5 mm (**a**,**b**); 0.2 mm (**f**,**g**); 0.1 mm (**c**,**d**,**e**,**h**,**i**).

of short bristles (Fig. 2d); tarsi two-segmented, tubular of similar length though basal segment slightly thicker than apical one, claws large, one-segmented, pulvilli absent (Fig. 2c-d). Abdomen appearing flattened, tergites and sternites not clearly visible. Female terminalia short, slightly shorter than head width, cuneate (Fig. 2a,b,g).

Revised key to Mesozoic psylloid genera (after Burckhardt & Poinar⁴, modified).

I.	Forewing lacking pterostigma.
	dopsylla Handlirsch, 1921 (= Cretapsylla Shcherbakov, 2020 syn. nov.; = Basicella Shcherbakov, 2020 syn. nov.)
	-Forewing bearing pterostigma2
2.	Vein Rs in forewing straight, veins Rs and M subparallel; vein M not branched; vein R shorter than M + Cu;
۷.	vein Cu _{1b} almost straight, directed toward wing base
	et Poinar, 2020
	-Combination of characters different. Vein Rs in forewing concavely curved towards fore margin (not
	visible in <i>Stigmapsylla</i>), veins Rs and M from base to apex first converging then diverging; vein M branched;
	vein Cu _{1b} straight or curved, directed toward hind margin or apex of wing
	out out the state of the state
	3
3.	Vein R of forewing distinctly shorter than M+Cu.
	2020
	-Vein R of forewing distinctly longer than M + Cu, or veins R and M + Cu subequal in length4
4.	Vein R of forewing distinctly longer than M+Cu; vein Cu _{1a} almost straight
	Malmopsylla Becker-Migdisova, 1985
	-Veins R and M + Cu of forewing subequal in length; vein Cu _{1a} distinctly curved5
5.	Forewing with cell cu ₁ low and very long, around 6.0 times as long high
	nov.
	-Forewing with cell cu ₁ higher and shorter, less than 2.5 times as long high
	6
5.	Forewing with long pterostigma, vein R ₂ straight
	Becker-Migdisova, 1985
	-Forewing with short pterostigma, vein R ₂ curved7
7.	Vein R+M+Cu of forewing ending at basal quarter of wing
	1985
	-Vein R+M+Cu of forewing ending at basal third of wing.
	1985

†**Amecephala pusilla sp. nov.** urn:lsid:zoobank.org:act:6B20A4F4-57DB-4F06-A43C-5DE3653D76E3 (Fig. 2a–i)

Etymology. From Latin pusillus = tiny, very small—for its small body size.

Holotype. Female, specimen number MAIG 6686; deposited in the Museum of Amber Inclusion, University of Gdańsk, Gdańsk, Poland. Complete and well-preserved (Fig. 2b,g), probably slightly compressed dorso-ventrally; the wings appear slightly detached from thorax and have been probably forced away from the thorax by the compression. Several gas bubbles on the ventral body side obscure parts of the head, thorax, abdomen, legs and the right forewing (Fig. 2g). Syniclusions: Aleyrodidae (part; second part in broken piece).

Locality and stratum. Myanmar, Kachin State, Hukawng Valley, SW of Maingkhwan, former Noije Bum 2001 Summit Site amber mine (closed). Lowermost Cenomanian, Upper Cretaceous.

Species diagnosis. As for the genus.

Description. Female; male unknown. Body minute, 1.20 mm long including forewing when folded over body. Head (ventrally partly covered by gas bubble) 0.28 mm wide, 0.10 mm long; vertex width 0.20 mm wide, 0.09 mm long; microsculpture or setae not visible. Antenna (Fig. 2a,b) with globular scape and cylindrical pedicel, thinner and longer than scape; flagellum 0.40 mm long; 1.6 times as long as head width; flagellar segments slightly more slender than pedicel, relative lengths as 1.0:0.7:0.6:0.6:0.6:0.6:0.7:1.0; flagellar segment 8 bearing two subequal terminal setae shorter that the segment. Clypeus and rostrum not visible, covered by gas bubble. Forewing (Fig. 2a,b,f,g) 0.90 mm long, 0.30 mm wide, 3.0 times as long as wide; membrane transparent, colourless, veins pale; anterior margin curved basally, posterior margin almost straight; vein R+M+Cu ending in basal fifth of wing; vein R slightly shorter that M+Cu; bifurcation of vein R proximal to middle of wing; cell R relatively narrow; vein R distinctly shorter than R relatively short, strongly curved towards fore margin; vein R slightly longer than veins R and R had R had R branching proximal to R branching curved towards fore margin; vein R slightly longer than veins R and R had R branching proximal to R branching crucal towards fore margin; vein R slightly longer than veins R and R had R branching proximal to R branching irregularly oval, about half as long as proctiger.

Character	Liadopsyllidae	Malmopsyllidae	Malmopsyllinae	Miralinae	Modern Psylloidea (NHMB data)	Polarity, apomorphic state
Costal space	Elongate, usually ribbon-shaped	Widening proximally or distally	Not widening distally	Widening distally	Variable, sometimes within a genus	Unknown
Pterostigma	Usually present, but often poorly developed	Distinct, dark	Not mentioned	Not mentioned	Variable, sometimes within a genus	Unknown
R+M+Cu [R+M+CuA] bifurcation	at < 1/5 wing length	at > 1/4 wing length	at 1/4-1/3 wing length	before wing midlength	variable, sometimes within a genus	unknown
R bifurcation	At < 1/3 wing Length	At>0.4 wing length	At 0.4–0.5 wing length	Beyond wing midlength	Variable, often within a genus	Unknown
M	Forked	Not mentioned	Forked	Unforked	Usually unforked	Unforked
M+Cu [M+CuA] bifurcation	Not mentioned	Not mentioned	At 0.35–0.5 wing length	Beyond wing midlength	Variable, sometimes within a genus, inter- grading	Unknown
Cu [CuA] fork	Triangular	Not mentioned	Triangular	Broad subquadrangular	Character poorly defined	Unknown
Cu _{1b} [CuA2]	Not recurrent or very short	Rather long, sometimes recurrent	Long, not recurrent	Recurrent	Variable	Unknown

Table 2. Forewing characters used by Shcherbakov⁹ to define the families and subfamilies. Nomenclature of veins follows Burckhardt & Poinar⁴, terms used by Shcherbakov⁹ are given in brackets.

Discussion

Recent molecular phylogenetic analyses of modern Psylloidea²⁰ support largely the classification by Burckhardt & Ouvrard¹ which is based to a great extent on the morphology of immatures (see also White & Hodkinson²¹) but also on adult characters such as details of the head, legs and terminalia. The venation of the forewing is rarely diagnostic for taxa at or above generic rank due to the high degree of homoplasy¹¹.6. A good example is the presence or absence of a pterostigma, though stable in most genera it varies sometimes, as in *Gyropsylla* Brèthes, 1921²². Shcherbakov⁰ resurrected the Malmopsyllidae, synonymised with Liadopsyllidae by Burckhardt & Poinar⁴, and split it into the two subfamilies Malmopsyllinae and Miralinae using evidence from eight forewing characters (Table 2). A critical review of these characters including in modern Psylloidea shows that they are unsuitable for diagnosing families and subfamilies. Five of the characters are variable within modern genera and one character is poorly defined (Cu [CuA] fork: not clear if it refers to the angle or the shape of cell Cu₂). The remaining two characters constitute autapomorphies defining *Mirala* but leaving Liadopsyllidae sensu Shcherbakov⁰, Malmopsyllidae sensu Shcherbakov⁰ and Malmopsyllinae sensu Shcherbakov⁰ undefined (plesiomorphies!) in a phylogeny based classification²³. They are, therefore rejected here. We propose following formal synonymies: Liadopsyllidae Martynov, 1927⁵; = Malmopsyllidae Becker-Migdisova, 1985⁻, **stat. rev.**; = Miralinae Shcherbakov, 2020⁰, **syn. nov.**

Similarly problematical are the circumscriptions of *Cretapsylla* Shcherbakov, 2020⁹, *Liadopsylla* (subgenus *Basicella* Shcherbakov, 2020⁹) and *Stigmapsylla* Shcherbakov, 2020⁹. The first is separated from *Liadopsylla* by the length ratio of the veins M + Cu and Cu (> 4 versus < 2) and the stronger curved vein M. Both characters vary within genera in modern psyllids and are unsuitable for defining genera. Shcherbakov⁹ provides a putative autapomorphy ("free CuA base") for the monophyly of the subgenus *Basicella* but fails to document the monophyly of the subgenus *Liadopsylla* sensu Shcherbakov⁹. For these reasons we propose following synonymies: *Liadopsylla* Handlirsch, 1921⁶; = *Cretapsylla* Shcherbakov, 2020⁹, **syn. nov.**; = *Basicella* Shcherbakov, 2020⁹), **syn. nov.** and following revised combinations: *Liadopsylla apedetica* Ouvrard, Burckhardt et Azar, 2010⁸, **comb. rev.** and *Liadopsylla hesperia* Ouvrard et Burckhardt, 2010⁸, **comb. rev.** both from *Cretapsylla* Shcherbakov, 2020⁹. The monotypic *Stigmapsylla* Shcherbakov, 2020⁹ is represented by a single, incomplete forewing and represents yet another other poorly defined liadopsyllid genus (along with *Gracilinervia*, *Malmopsylla*, *Neopsylloides and Pauropsylloides*).

Amecephala pusilla **gen. et sp. nov.** differs from the other known taxa of Liadopsyllidae in the very long pedicel of the antenna, the long and narrow forewings (3.0 times as long as wide), that are widest in the middle, the very short vein Rs as well as the very long and low cell cu_1 . It shares with Liadopsylla the absence of vein R_1 and the short vein R+M+Cu ending at basal fifth of wing. Whether these characters reflect a close phylogenetic relationship is difficult to judge as these characters are strongly subjected to homoplasy. Unlike Liadopsylla, Amecephala displays a distinctly pigmented pterostigma as the other Mesozoic Liadopsyllidae.

The antenna of *Amecephala pusilla* shows some remarkable features. In Psylloidea, including Liadopsyllidae, the scape and, to a lesser extent, the pedicel, are in general distinctly wider but much shorter than any of the flagellar segments. In most psyllids, one of the antennal segments 3, 7 or 8 (flagellar segments 1, 5 or 6) constitutes the longest segment. There are a few exceptions such as *Livia* Latreille, 1802²⁴, *Notophyllura* Hodkinson, 1986²⁵, or some species of *Calophya* Löw, 1879²⁶, where the pedicel is longer than the other segments. These taxa have short antennae (usually shorter than head width) and sometimes a reduced number of antennal segments. In *Amecephala pusilla*, the antenna is distinctly longer than the head width and scape and pedicel are almost as slender as the flagellar segments. The long pedicel is an unique feature in Liadopsyllidae and very exceptional in modern psyllids and constitutes probably an apomorphic condition which developed apparently several times independently, in modern psyllids mostly by reduction of the flagellar length. The general head shape of

Amecephala is similar to that of Liadopsylla and Mirala; the compound eyes in Liadopsylla are less protruding than in the other two genera.

The legs of *Amecephala, Liadopsylla* and *Mirala* are of similar build. The hind legs are not modified compared to those in modern psyllids, the tarsal segments are subequal in length and lack pulvilli. Whereas the first two characters are primitive, the last one is derived. Pulvilli or similar structures are present in adults of modern psyllids, in whiteflies, aphids, male scale insects and several groups of Auchenorrhyncha and Heteroptera²⁷. The reduction of pulvilli in Liadopsyllidae constitutes a potential autapomorphy supporting, admittedly weakly, the monophyly of Liadopsyllidae.

Little is known about the terminalia of Liadopsyllidae. In modern psyllids, the terminalia constitute often the most important structure to diagnose species. The male terminalia of following species have been described: Liadopsylla grandis Becker-Migdisova, 19857, Liadopsylla karatavica Becker-Migdisova, 19857, Liadopsylla longiforceps (Becker-Migdisova, 1985)⁷, Liadopsylla tenuicornis Martynov, 1926⁵, and Liadopsylla turkestanica Becker-Migdisova, 1949¹⁵. Of the last species, also the female terminalia have been described. All these species are represented by compression fossils, sometimes difficult to interpret and lacking morphological detail. More details are visible in the amber specimen of Liadopsylla apedetica Ouvrard, Burckhardt et Azar, 20108, a female displaying very long terminalia. The female terminalia of L. turkestanica appear much shorter. In Amecephala pusilla the female terminalia are relatively short and an oval circumanal ring is visible. This structure, always present in modern psyllids²⁸, is documented here for the first time in Mesozoic psyllids. In modern psyllids, the length of the female terminalia is often correlated with the place where the eggs are laid. Short female terminalia are usually present in species that lay their eggs on the surface or in crevices of a twig or at the base of leaf or flower buds, as in many species of Cacopsylla Ossiannilsson, 1970²⁹. Long terminalia are used for depositing the eggs into buds, such as in the Holarctic species of *Psylla* Geoffroy, 1762³⁰, associated with Betulaceae, or into the flower heads of Asteraceae as in species of the predominantly Neotropical Calinda Blanchard, 1852³¹, Burckhardt, pers. obs. This diversity of female terminalia in Liadopsyllidae suggests that the family may have used a range of substrates for oviposition perhaps on different host taxa. According to Burckhardt & Poinar⁴ the Lauraceae could have been among the host families of psyllids from Burmese amber.

Material and methods

The specimen is an inclusion in mid-Cretaceous amber from the Kachin State in northern Myanmar (Burma). The specimen was purchased together with the whole bunch in 2016 from authorised dealer in Bahan, registered by Ministry of Co-operatives in Myanmar. To further prove sample origination, VIS and UV (395 nm) examination of sample was proceeded at Laboratory of Amber, Museum of Amber Inclusions, University of Gdańsk and Fourier Transform Infrared Spectrum with use of Nicolet iS10 in Amber Laboratory of the International Amber Association in Gdańsk. The amber piece was cut and polished for better visibility. For the microscopic examination, we used a Nikon SMZ1500, Nikon SMZ1270, Leica M205C stereoscopic microscopes and a Nikon Microphot-FX equipped with a camera lucida and changeable direct and transmitted light. The photographs were taken using a Nikon Microphot-FX with a Nikon Eclipse E 600 digital camera and Lucia software and edited with Adobe Photoshop Elements 6.0.

Morphological terminology follows mostly Ossiannilsson²⁸ and Hollis³² but the interpretation of veins R_1 and R_2 accords with Becker-Migdisova⁵ and Burckhardt & Poinar⁶.

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References

- $1. \ \, \text{Burckhardt, D. \& Ouvrard, D. A revised classification of the jumping plant-lice (Hemiptera: Psylloidea)}. \ \, \textit{Zootaxa 3509}, 1-34 \ (2012).$
- 2. Burckhardt, D. Psylloid pests of temperate and subtropical crop and ornamental plants (Hemiptera, Psylloidea): A review. *Trends Agric. Sci. Entomol.* 2, 173–186 (1994).
- 3. Ouvrard, D., Burckhardt, D. & Greenwalt, D. The oldest jumping plant-louse (Hemiptera: Sternorrhyncha) with comments on the classification and nomenclature of the Palaeogene Psylloidea. *Acta Mus. Morav. Sci. Biol.* **98**(2), 21–33 (2013).
- 4. Burckhardt, D. & Poinar, G. The first jumping plant-louse from mid-Cretaceous Burmese amber and its impact on the classification of Mesozoic psylloids (Hemiptera: Sternorrhyncha: Psylloidea s.l.). Cret. Res. 106, 104240. https://doi.org/10.1016/j.cretres.2019.104240 (2019).
- Martynov, A. V. Jurassic fossil insects from Turkestan. 6. Homoptera and Psocoptera. Bull. Acad. Sci. U.R.S.S. 1926, 1349–1366 (1926).
- 6. Handlirsch, A. Paläontologie. In Handbuch der Entomologie. Bd. III (ed. Schröder, C.) 117-306 (Gustav Fischer, Jena, 1921).
- Becker-Migdisova, E. E. Iskopaemye nasekomye psillomorfy (Fossil psyllomorphous insects). Trudy Paleontol. Inst. 206, 1–94 (1985) (in Russian).
- 8. Ouvrard, D., Burckhardt, D., Azar, D. & Grimaldi, D. Non-jumping plant-lice in Cretaceous amber (Hemiptera: Sternorrhyncha: Psylloidea). Syst. Entomol. 35(1), 172–180. https://doi.org/10.1111/j.1365-3113.2009.00499.x (2010).
- 9. Shcherbakov, D. E. New Homoptera from the early Cretaceous of Buryatia with notes on the insect fauna of Khasurty. *Russ. Entomol. J.* 29(2), 127–138. https://doi.org/10.15298/rusentj.29.2.02 (2020).
- Drohojowska, J. & Szwedo, J. Early Cretaceous Aleyrodidae (Hemiptera: Sternorrhyncha) from the Lebanese amber. Cret. Res. 52, 368–389. https://doi.org/10.1016/j.cretres.2014.03.015 (2015).
- 11. Hakim, M., Azar, D., Szwedo, J., Brysz, A. M. & Huang, D. Y. New paraneopterans (Protopsyllidioidea, Hemiptera) from the mid-Cretaceous amber of northern Myanmar. Cret. Res. 98, 136–152. https://doi.org/10.1016/j.cretres.2018.12.012 (2019).
- 12. Drohojowska, J., Szwedo, J., Żyła, D., Huang, D.-Y. & Müller, P. Fossils reshape the Sternorrhyncha evolutionary tree (Insecta, Hemiptera). Sci. Rep. 10, 11390. https://doi.org/10.1038/s41598-020-68220-x (2020).
- 13. Shcherbakov, D. E. Novye mezozoïskie ravnokrylye (New Mesozoic Homoptera.) in Novye vidy iskopaemykh bespozvonochnykh Mongolii (New species of fossil invertebrates of Mongolia) (ed. Rozanov, A. Yu.). Sov.-Mongol. Paleontol. Exped. 33, 60–63 (1988) (in Russian).

- Ansorge, J. Insekten aus dem oberen Lias von Grimmen (Vorpommern, Norddeutschland). Neue Paläontol. Abhandl. 2, 1–132 (1996).
- 15. Becker-Migdisova, E. E. Mezozoiskie Homoptera Srednei Azii (Mesozoic Homoptera of Central Asia). *Trudy Paleontol. Inst.* 22, 1–68 (1949) (in Russian).
- 16. Scudder, S. H. Taxonomic names. On some remains of Palaeozoic insects recently discovered in Nova Scotia and New Brunswick. *Can. Nat.* **3**, 202–206 (1867).
- 17. Linnaeus, C. Systema naturae per regna tria naturae, secundum classes, ordines, genera, species, cum characteribus, differentiis, synonymis, locis. Tomus I. Editio decima, reformata. (Laurentius Salvius, Holmia, 1758).
- 18. Amyot, C.J.-B. & Audinet-Serville, J. G. *Deuxième partie. Homoptères. Homoptera Latr. Histoire naturelle des insects. Hemiptères* 1–676 (Librairie encyclopédique de Roret, Paris, 1843).
- 19. Latreille, P. A. Genera crustaceorum et insectorum secundum ordinem naturalem in familias disposita, iconibus exemplisque plurimis explicate 1–280 (A. Koenig, Paris, 1807).
- 20. Percy, D. M. et al. Resolving the psyllid tree of life: Phylogenomic analyses of the superfamily Psylloidea (Hemiptera). Syst. Entomol. 43(4), 762–776. https://doi.org/10.1111/syen.12302 (2018).
- 21. White, I. M. & Hodkinson, I. D. Nymphal taxonomy and systematics of the Psylloidea (Homoptera). *Bull. Brit. Mus. Nat. Hist. Entomol.* **50**, 153–301 (1985).
- Burckhardt, D. & Queiroz, D. L. Systematics of the Neotropical jumping plant-louse genus Limataphalara (Hemiptera: Psylloidea: Aphalaridae) and phylogenetic relationships within the subfamily Aphalarinae. In Studies in Hemiptera in honour of Pavel Lauterer and Jaroslav L. Stehlík (eds Kment, P., Malenovský, I. & Kolibác, J.). Acta Mus. Moraviae. Sci. Biol. 98, 35–56 (2013).
- 23. Hennig, W. Grundzüge einer Theorie der Phylogenetischen Systematik (Deutscher Zentralverlag, Berlin, 1950).
- 24. Latreille, P. A. Histoire naturelle, générale et particulière des crustacés et des insectes: ouvrage faisant suite aux oeuvres de Leclerc de Buffon, et partie du cours complet d'histoire naturelle rédigé par C.S. Sonnini 1–467 (Dufart, F, Paris, 1802).
- 25. Hodkinson, I. D. First records of Euphyllurini (Homoptera: Psylloidea) from Central and South America. *Entomol. Scand.* 17(2), 143–152 (1986).
- 26. Löw, F. Zur Systematik der Psylloden. Verh. d. k. k. Zool.-Bot. Ges. 28, 586-610 (1879).
- 27. Beutel, R. G., Friedrich, F., Ge, S.-Q. & Yang, X.-K. Insect Morphology and Phylogeny 1-531 (De Gruyter, Berlin, 2014).
- 28. Ossiannilsson, F. The Psylloidea (Homoptera) of Fennoscandia and Denmark. Fauna Entomol. Scand. 26, 1-347 (1992).
- 29. Ossiannilsson, F. Contributions to the knowledge of Swedish psyllids (Hem. Psylloidea). Entomol. Scand. 1, 135-144 (1978).
- 30. Geoffroy, E. L. Histoire abrégée des insectes qui se trouvent aux environs de Paris; dans laquelle ces animaux sont rangés suivant un ordre méthodique 1–523 (Durand, Paris, 1762).
- 31. Blanchard, E. Orden VII. Hemípteros; V. Afidídeos; Tribu I.—Silinas. In *Historia física y política de Chile. Zoología* (ed. Gay, C.) 306–316 (Maulde et Renou, Paris, 1852).
- 32. Hollis, D. Australian Psylloidea: Jumping plant lice and lerp insects 1–217 (Australian Biological Resources Study, Canberra, 2004).

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Author contributions

J.D., D.B. and J.S. designed and wrote the paper, J.D. and J.S. prepared and composed illustrations. P.M. found the specimen and brought it to J.S. All authors commented on the final manuscript.

Competing interests

The authors declare no competing interests.

Additional information

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