

HHS Public Access

J Gastroenterol Hepatol. Author manuscript; available in PMC 2020 October 21.

Published in final edited form as:

Author manuscript

J Gastroenterol Hepatol. 2018 November; 33(11): 1920–1924. doi:10.1111/jgh.14265.

The Association of Circulating Inflammation Proteins and Gallstone Disease

Zhiwei Liu¹, Troy J. Kemp², Yu-Tang Gao³, Amanda Corbel¹, Emma E. McGee¹, Bingsheng Wang⁴, Ming-Chang Shen⁵, Asif Rashid⁶, Ann W. Hsing^{7,8}, Allan Hildesheim¹, Ruth M. Pfeiffer⁹, Ligia A. Pinto², Jill Koshiol¹

¹ Infections and Immunoepidemiology Branch of the Division of Cancer Epidemiology and Genetics, National Cancer Institute, Bethesda, Maryland, USA

² HPV Immunology Laboratory, Frederick National Laboratory for Cancer Research, Leidos, Biomedical Research, Inc, Frederick, MD, USA

³.Department of Epidemiology, Shanghai Cancer Institute, Shanghai, China

⁴ Department of General Surgery, Zhongshan Hospital, School of Medicine, Fudan University, Shanghai, China

^{5.}Department of Pathology, Shanghai Cancer Center, Fudan University, Shanghai, China

⁶Department of Pathology, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

⁷ Stanford Cancer Institute, Palo Alto, CA, USA

⁸ Stanford Prevention Research Center, Department of Medicine, Stanford School of Medicine, Palo Alto, CA, USA

⁹·Biostatistics Branch, Division of Cancer Epidemiology and Genetics, National Cancer Institute, MD, USA

Abstract

Background: Inflammation plays a role in the development of both gallstones and gallbladder cancer; however, few studies have investigated the association of circulating inflammation proteins with risk of gallstones.

Methods: We measured 13 cytokines (including 10 interleukins), that have been associated with cancer in serum samples collected from 150 gallstone patients and 149 population-based controls from Shanghai, China, in 1997–2001. We estimated the associations of each cytokine, categorized into quartiles and coded as a trend, with risk of gallstones using logistic regression models adjusted for potential confounders.

Corresponding author: Zhiwei Liu, Ph.D., Infections and Immunoepidemiology Branch, Division of Cancer Epidemiology and Genetics, National Cancer Institute, 9609 Medical Center Dr, Rockville, MD 20852. Phone: 240-276-6726; Fax: 240-276-7806, zhiwei.liu@nih.gov.

Conflicts of Interest: No authors have any relevant conflicts of interest.

Page 2

Results: Higher levels of interleukin (IL)-6, IL-10, IL-12 (p70), and IL-13 were associated with increased risk of gallstones (i.e., $P_{\text{trend}} < 0.003$, Bonferroni corrected), with odds ratios (ORs) that ranged from OR highest quartile [Q4] vs lowest quartile [Q1] = 3.2 (95% confidence interval [CI]: 1.4, 7.5) for IL-13 to OR _{Q4 vs. Q1}= 5.7 (95% CI: 2.5, 13.5) for IL-12 (p70). In a regression model including all four interleukins, only IL-12 retained statistical significance (P < 0.05).

Conclusions: We found four circulating interleukins that were associated with gallstones. Future studies are needed to validate the findings and evaluate the common pathway or mechanism in the development of gallbladder diseases associated with these cytokine signatures.

Keywords

case-control study; cytokines; gallstones; inflammation; interleukins; risk factor

Introduction

Gallstone disease is prevalent in the general population, affecting 5% to 15% of adults worldwide ¹⁻⁴. The incidence has increased in recent years, particularly in Western populations ^{5–9}. The cholesterol stones comprise the vast majority (over 70%) of gallstones in Western countries, while both cholesterol stones and pigment stones are common in Asian countries ^{9–11}. Gallstones can result in several clinical diseases with severe symptoms, including cholecystitis, cholangitis, and acute pancreatitis. Importantly, gallstones play a key role in the development of gallbladder cancer, as suggested by the fact that approximately 90% or more of patients with gallbladder cancer have a history of gallstones ^{12, 13}, with an odds ratio (OR) of >20¹². Both genetic and environmental factors may contribute to the formation of gallstones, and risk factors that contribute to the formation of stones vary across types ¹⁴. For example, cholesterol stones are more likely to be associated with obesity, which may affect the concentration of the cholesterol level in bile, while pigment stones are more likely to be associated with biliary infections and hemochromatosis ¹⁴. For example, Clonorchis sinensis infection is associated with the development of pigment stones ¹⁵. In addition, inflammation has been proposed to play an essential role in gallstone development, as indicated by the fact that formation of cholesterol gallstones is preceded by histopathologic alterations in the gallbladder wall that indicate inflammation in both animal models and humans ^{16–18}. Some inflammation-related conditions, such as obesity, diabetes, and infections (e.g. Helicobacter pylori), are also associated with risk of cholesterol gallstones¹⁹⁻²¹. However, few studies have investigated the association between inflammation proteins and risk of gallstones ²².

Previously we showed that circulating inflammation proteins are significantly correlated with inflammation proteins measured in bile ²³, suggesting that we can use blood samples to better understand the association between inflammation and gallstone risk. In addition, we found 26 inflammation proteins associated with gallbladder cancer risk when comparing levels of 49 inflammation-related proteins in serum from gallbladder cancer cases to those from patients with gallstones ^{23, 24}. Seven of the markers were associated with risk of early stage gallbladder cancer, suggesting that measurement of inflammation proteins may have clinical utility. This utility may extend to other gallbladder diseases, including gallstones. To better understand the role of inflammation in development of gallstones, we undertook an

analysis of circulating cytokines and gallstone risk in a population-based case-control study in Shanghai, China.

Methods

Study population

The current study is based on biospecimens collected from the Shanghai Biliary Cancer Study, which has been previously described ^{12, 23}. In brief, between 1997 and 2001, a total of 368 patients with gallbladder cancer were enrolled; 774 patients with gallstones (>95% of the eligible gallstone cases) who were undergoing cholecystectomy or medical treatment at the same hospitals as the cancer cases were frequency matched by sex and age (in 5-year category) to the cases; 959 healthy controls (>83% of eligible controls) were also randomly selected from the Shanghai population and frequency matched to cancer cases on age and sex. Although we do not have information on whether the gallstone patients had active cholecystitis, we expect that the presence of severe active cholecystitis is low because cholecystectomy is not recommended for such patients.^{25, 26} Gallstone status was confirmed through self-report of a gallstone history, operative reports and imaging data for cases with gallstones and 87% of the participating controls. In addition to a self-reported gallstone history, transabdominal ultrasound was performed to validate gallstone status and to identify silent gallstones. The Shanghai Cancer Institute and National Cancer Institute institutional review boards approved the study. Written informed consent was obtained for all participants.

We evaluated circulating inflammation proteins in 150 randomly selected gallstone patients who either had serum collected prior to surgery or did not have surgery, and 150 randomly selected healthy controls frequency matched by sex and age (in 5-year category, range: 35–74 years) to the gallstone cases. All gallstones were in gallbladder (N= 126), gallbladder neck (N=13), or both (N=11). A total of 121 gallstones were cholesterol/mixed stones, 16 were pigment/black stones, and 13 had unknown composition. None of the controls had silent gallstones. One control did not have sufficient serum for this assay. We used the Milliplex high-sensitivity (EMD Millipore, Billerica, MA) kit to test for 16 cytokines that have been associated with cancer (interleukin [IL]-1 β , IL-4, IL-5, IL-6, IL-7, IL-8, IL-10, IL-12[p70], IL-13, IL-17 α , IL-21, IL-23, CCL-3 [CC-Chemokine Ligand 3], CCL-20, GM-CSF [Granulocyte macrophage-colony stimulating factor], and CX3CL1 [chemokine (C-X3-C motif) ligand 1])²⁷. Serum samples were incubated with beads in 96-well plates, after which fluorescently labeled detection antibodies were added. A Bio-Plex instrument and Bio-Plex Manager 6.1 software (Bio-Rad, Hercules, CA, USA) were used to analyze the 96-well plates

Details on quality controls have been previously published ^{23, 28, 29}. In brief, all samples were measured in a single batch, and blinded duplicate aliquots from 10 participants were selected to estimate the reproducibility of the assays. We calculated log-adjusted coefficients of variation (CVs) and intraclass correlation coefficients (ICCs) using a log-transformed general linear model. We excluded 3 proteins (CX3CL1, IL-5, and IL-21) with overall CVs > 30%, and/or ICCs <0.75. Thus, the final analysis included 13 evaluable cytokines including 10 interleukins, in 150 patients with gallstones and 149 controls.

Statistical Analyses

All proteins were detectable in 75% of healthy controls and gallstone cases. We created four categories based on quartiles of values above the lower limit of quantitation using the distribution among healthy controls (subjects with undetectable values were included in the lowest quartile). These proteins were modeled both as categorical and ordinal variables (coded as 1, 2, 3, 4) to evaluate linear trend.

We used unconditional logistic regression models to estimate odds ratios (ORs) and 95% confidence intervals (CIs) for associations of each cytokine individually with gallstones. Minimal adjustments for the models included the frequency-matching variables, age group at diagnosis/selection in categories (54, 55–65, or66 years), and sex. Identification of potential confounders was based on prior knowledge. Fully adjusted models additionally included education (none/primary, junior middle, senior middle, some college), ever drinking, ever smoking, self-reported body mass index (underweight, <18.5; normal weight, 18.5-24.9; overweight, 25-29.9; obese, 30 kg/m^2), diabetes, and history of cholecystitis. Two-sided *P* values for trend across protein categories were computed using Wald tests. We tested for modification of cytokine associations by sex (which may differ in the prevalence of some lifestyle factors, such as smoking), BMI, and diabetes (both of which have been shown to be slightly higher among gallstone cases than controls) using likelihood ratio tests for nested models with and without interaction terms.

Statistical analyses were performed using SAS, version 9.4 (SAS Institute, Cary, NC, USA). The statistical tests were two sided with $\alpha = 0.05$. Bonferroni corrected *P* values (0.05/13=0.004) were calculated to account for multiple testing.

Results

Characteristics of the 150 cases with gallstones and the 149 population-based healthy controls, stratified by sex, are shown in Table 1. As expected, gallstone cases were more likely to report a history of cholecystitis than controls (P < 0.001 for both women and men), and women from the case group were more likely to be less educated than women from the control group (P = 0.018). Otherwise, we found no significant differences between the distribution of characteristics of cases and controls, stratified by sex.

Of the 13 cytokines, four were statistically significantly associated with gallstone risk in age- and sex- adjusted models and fully adjusted models (using $P_{\text{trend}} < 0.003$, Bonferroni correction, Table 2). Higher levels of interleukin IL-6, IL-10, IL-12 (p70), and IL-13 were associated with a higher risk of gallstones, with odds rations ranging from OR highest quartile [Q4] vs lowest quartile [Q1] = 3.2 (95% CI: 1.4, 7.5) for IL-13 to OR Q4 vs. Q1= 5.7 (95% CI: 2.5, 13.5) for IL-12 (p70) in fully adjusted models. The association between the remaining nine markers (IL-1 β , IL-4, IL-7, IL-8, IL-17 α , IL-23, CCL3, CCL20, and GM-CSF) and gallstone risk are presented in Supplementary Table 1). Of the 4 proteins significantly associated with gallstone risk after Bonferroni correction, the odds ratios did not differ statistically by sex, BMI, or history of diabetes, although power was limited for these analyses (all P for heterogeneity >0.05, Supplementary Table 2).

Among controls, the correlations between the 4 proteins significantly associated with gallstone risk after Bonferroni correction were strong (spearman correlation coefficient range: 0.314 - 0.490; all *P*<0.0001,). In a regression model including these 4 proteins, only IL-12 retained statistical significance (*P*<0.05).

Discussion

The present study shows associations between gallstones and circulating IL-6, IL-10, IL-12 (p70), and IL-13. In particular, IL-12 (p70) was independently associated with risk of gallstones. Thus, prospective studies of IL-12 (p70) and gallstone risk may be warranted to confirm these findings.

Our results are in line with a previous study indicating that inflammation, as measured by a few cytokines (i.e., IL-1a, IL-6, IL-8, and tumor necrosis factor [TNF]-a), is associated with risk of gallstone diseases ²². However, the magnitude of associations between cytokines with risk of gallstones was much smaller than the associations of cytokines with risk of gallbladder cancer ²³. For example, our previous study found an OR _{Q4 vs. Q1} of IL-8 with gallbladder cancer versus gallstones of 96.8 (95% CI: 11.9, 790.2),²³ whereas no significant association was observed between IL-8 and gallstones in the present study. In addition, the magnitude of associations between gallbladder cancer and gallstones for IL-6 (OR _{Q4 vs. Q1}: = 21.6, 95% CI: 8.0, 58.6) and IL-10 (OR _{Q4 vs. Q1}: = 6.1, 95% CI: 2.6, 14.3) (unpublished data) is much higher than that between gallstones and healthy controls. These findings suggest that gallstones do not cause changes in the circulating inflammation profile that are as marked as those caused by gallbladder cancer. If true, there may be strong potential for inflammation proteins to stratify patients with gallstones based on risk for gallbladder cancer ²⁴.

Although the role of cytokines in the gallbladder and their influence on gallstone formation is not clear, it is well established that the formation of gallstone involves inflammation ^{18, 30}. For example, inflammation may alter the metabolism of several proteins and lipids; these changes may alter cholesterol and bile acid metabolism and increase bile salt levels, thus causing formation of gallstones. In the present study, four cytokines (IL-6, IL-10, IL-12 [p70] and IL-13) were associated with gallstones. These observations are biologically plausible. For example, IL-6 can be expressed by biliary epithelium, and it is known to have proliferative effects on several nonhematopoietic cells ^{17, 31}. IL-6 can also cause infiltration of inflammatory cells and increase gallbladder wall thickness ^{17, 31}. To our knowledge, no previous studies have investigated the role of IL-10, IL-12, and IL-13 on the formation of gallstones. Studies have shown that IL-10 predominantly inhibits lipopolysaccharide (LPS) and bacterial product-mediated induction of pro-inflammatory cytokines, including IL-12³². IL-12 can stimulate the production of TNF-a from T cells and natural killer cells, which directly affects the absorption, secretion, and functions of gallbladder epithelial cells ¹⁶. Interleukin-13 can control IgE synthesis, which causes allergic inflammation and different diseases including asthma ³³; therefore, the association between asthma and gallstones merits further investigation. In an adjusted model including all four proteins, only IL-12 remained statistically significant. Although our results are cross-sectional, this finding suggests that the role of IL-12 in the formation of gallstones merits future investigation. It is

Liu et al.

noteworthy that IL-10, IL-12, and IL-13 were significantly associated gallstones, but not with gallbladder cancer ²⁴, suggesting that the inflammatory processes that drive the development of gallstones may differ from those that drive the development of gallbladder cancer.

The present study is one of the largest studies of cytokines and gallstone to date. Misclassification of disease status was minimized through comprehensive pathology and clinical review, which provided nearly complete confirmation of case status. However, our results should be interpreted in light of several limitations. First, although there is strong biologic plausibility for the association of cytokines with gallstone risk, our observations require replication. Second, reverse causation cannot be ruled out due to the cross-sectional case-control design (i.e., inflammation protein levels may be elevated due to the presence of gallstones or acute cholecystitis among a minority of cases). However, while histologic evidence of cholecystitis was seen among 145 of 147 cases (99%) reviewed, acute cholecystitis is rare. In a previous study, <5% of patients with cholecystitis undergoing cholecystectomy had histologic evidence of acute cholecystitis ³⁴. Finally, a limited number of inflammation markers were assessed in the present study. A more comprehensive assessment of systematic inflammation and gallstone disease development merits investigation in a prospective study, and studies in other populations are needed to replicate our findings.

In conclusion, of the 13 circulating cytokines investigated, four were associated with gallstones, of which three were not associated with gallbladder cancer in a previous study, and the magnitude of the associations generally seemed to be lower than that observed for gallbladder cancer. These differences in the associations of cytokines with gallstones as opposed to gallbladder cancer suggest that the transition from normal to gallstones and from gallstones to cancer may be characterized by processes that vary in terms of the mechanism at play and/or the degree of disease-related changes. Future studies are needed to further evaluate the common pathway or mechanism associated with these cytokine signatures.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

Grant support: Intramural Research Program of the National Institutes of Health, National Cancer Institute, Division of Cancer Epidemiology and Genetics.

Reference

- Everhart JE, Khare M, Hill M, Maurer KR. Prevalence and ethnic differences in gallbladder disease in the United States. Gastroenterology. 1999; 117: 632–9. [PubMed: 10464139]
- [2]. Shaffer EA. Epidemiology and risk factors for gallstone disease: has the paradigm changed in the 21st century? Curr Gastroenterol Rep. 2005; 7: 132–40. [PubMed: 15802102]
- [3]. Shaffer EA. Gallstone disease: Epidemiology of gallbladder stone disease. Best Pract Res Clin Gastroenterol. 2006; 20: 981–96. [PubMed: 17127183]

Liu et al.

- [4]. Chen CH, Huang MH, Yang JC, et al. Prevalence and risk factors of gallstone disease in an adult population of Taiwan: an epidemiological survey. J Gastroenterol Hepatol. 2006; 21: 1737–43. [PubMed: 16984599]
- [5]. Jensen KH, Jorgensen T. Incidence of gallstones in a Danish population. Gastroenterology. 1991; 100: 790–4. [PubMed: 1993501]
- [6]. Lindkvist B, Appelros S, Manjer J, Borgstrom A. Trends in incidence of acute pancreatitis in a Swedish population: is there really an increase? Clin Gastroenterol Hepatol. 2004; 2: 831–7. [PubMed: 15354285]
- [7]. Acalovschi M, Dumitrascu D, Caluser I, Ban A. Comparative prevalence of gallstone disease at 100-year interval in a large Romanian town, a necropsy study. Dig Dis Sci. 1987; 32: 354–7.
 [PubMed: 3829878]
- [8]. Aerts R, Penninckx F. The burden of gallstone disease in Europe. Aliment Pharmacol Ther. 2003; 18 Suppl 3: 49–53. [PubMed: 14531741]
- [9]. Stinton LM, Shaffer EA. Epidemiology of gallbladder disease: cholelithiasis and cancer. Gut Liver. 2012; 6: 172–87. [PubMed: 22570746]
- [10]. Njeze GE. Gallstones. Niger J Surg. 2013; 19: 49–55. [PubMed: 24497751]
- [11]. Qiao T, Ma RH, Luo XB, Yang LQ, Luo ZL, Zheng PM. The systematic classification of gallbladder stones. PLoS One. 2013; 8: e74887.
- [12]. Hsing AW, Gao YT, Han TQ, et al. Gallstones and the risk of biliary tract cancer: a populationbased study in China. Br J Cancer. 2007; 97: 1577–82. [PubMed: 18000509]
- [13]. Lai CH, Lau WY. Gallbladder cancer--a comprehensive review. Surgeon. 2008; 6: 101–10.[PubMed: 18488776]
- [14]. Lammert F, Gurusamy K, Ko CW, et al. Gallstones. Nat Rev Dis Primers. 2016; 2: 16024.[PubMed: 27121416]
- [15]. Qiao T, Ma RH, Luo XB, Luo ZL, Zheng PM. Cholecystolithiasis is associated with Clonorchis sinensis infection. PLoS One. 2012; 7: e42471.
- [16]. Rege RV. Inflammatory cytokines alter human gallbladder epithelial cell absorption/secretion. J Gastrointest Surg. 2000; 4: 185–92. [PubMed: 10675242]
- [17]. van Erpecum KJ, Wang DQ, Moschetta A, et al. Gallbladder histopathology during murine gallstone formation: relation to motility and concentrating function. J Lipid Res. 2006; 47: 32– 41. [PubMed: 16224116]
- [18]. Maurer KJ, Carey MC, Fox JG. Roles of infection, inflammation, and the immune system in cholesterol gallstone formation. Gastroenterology. 2009; 136: 425–40. [PubMed: 19109959]
- [19]. Fremont-Rahl JJ, Ge Z, Umana C, et al. An analysis of the role of the indigenous microbiota in cholesterol gallstone pathogenesis. PLoS One. 2013; 8: e70657.
- [20]. Mark-Christensen A, Brandsborg S, Laurberg S, et al. Increased Risk of Gallstone Disease Following Colectomy for Ulcerative Colitis. Am J Gastroenterol. 2017; 112: 473–8. [PubMed: 28117363]
- [21]. Shabanzadeh DM, Skaaby T, Sorensen LT, Eugen-Olsen J, Jorgensen T. Metabolic biomarkers and gallstone disease - a population-based study. Scand J Gastroenterol. 2017: 1–8.
- [22]. Shengelia M, Intskirveli N, Gogebashvili N. Inflammatory markers of gallstones disease in menopausal women. Georgian Med News. 2012: 52–5.
- [23]. Koshiol J, Castro F, Kemp TJ, et al. Association of inflammatory and other immune markers with gallbladder cancer: Results from two independent case-control studies. Cytokine. 2016; 83: 217– 25. [PubMed: 27173614]
- [24]. Koshiol J, Gao YT, Corbel A, et al. Circulating inflammatory proteins and gallbladder cancer: Potential for risk stratification to improve prioritization for cholecystectomy in high-risk regions. Cancer Epidemiol. 2018; 54: 25–30. [PubMed: 29554539]
- [25]. Borzellino G, Sauerland S, Minicozzi AM, et al. Laparoscopic cholecystectomy for severe acute cholecystitis. A meta-analysis of results. Surg Endosc. 2008; 22: 8–15. [PubMed: 17704863]
- [26]. Yamashita Y, Takada T, Strasberg SM, et al. TG13 surgical management of acute cholecystitis. J Hepatobiliary Pancreat Sci. 2013; 20: 89–96. [PubMed: 23307007]

Liu et al.

- [27]. Elinav E, Nowarski R, Thaiss CA, Hu B, Jin C, Flavell RA. Inflammation-induced cancer: crosstalk between tumours, immune cells and microorganisms. Nat Rev Cancer. 2013; 13: 759– 71. [PubMed: 24154716]
- [28]. Kemp TJ, Castro FA, Gao YT, et al. Application of multiplex arrays for cytokine and chemokine profiling of bile. Cytokine. 2015; 73: 84–90. [PubMed: 25743242]
- [29]. Van Dyke AL, Kemp TJ, Corbel AF, et al. Lipopolysaccharide-pathway proteins are associated with gallbladder cancer among adults in Shanghai, China with mediation by systemic inflammation. Ann Epidemiol. 2016; 26: 704–9. [PubMed: 27793274]
- [30]. Reynoso-Paz S, Coppel RL, Mackay IR, Bass NM, Ansari AA, Gershwin ME. The immunobiology of bile and biliary epithelium. Hepatology. 1999; 30: 351–7. [PubMed: 10421640]
- [31]. Iglesias M, Plowman GD, Woodworth CD. Interleukin-6 and interleukin-6 soluble receptor regulate proliferation of normal, human papillomavirus-immortalized, and carcinoma-derived cervical cells in vitro. Am J Pathol. 1995; 146: 944–52. [PubMed: 7717461]
- [32]. Aste-Amezaga M, Ma X, Sartori A, Trinchieri G. Molecular mechanisms of the induction of IL-12 and its inhibition by IL-10. J Immunol. 1998; 160: 5936–44. [PubMed: 9637507]
- [33]. Rael EL, Lockey RF. Interleukin-13 signaling and its role in asthma. World Allergy Organ J. 2011; 4: 54–64. [PubMed: 23283176]
- [34]. Mittal R, Jesudason MR, Nayak S. Selective histopathology in cholecystectomy for gallstone disease. Indian J Gastroenterol. 2010; 29: 26–30. [PubMed: 20373083]

Table 1.

Characteristics of Gallstone Cases and Population-Based Controls, by Sex, Shanghai, China, 1997 - 2001

	Cases (n=109 Women, n=41 Men)		Controls (n=86 Wo	$\chi 2 P$ value ^a		
Characteristic and Category	Women (%)	Men (%)	Women (%)	Men (%)	Women	Men
Age group, years					0.217	0.08
54	26 (23.8)	4 (9.8)	12 (14.0)	13 (20.6)		
55 - 65	34 (31.2)	11 (26.8)	29 (33.7)	24 (38.1)		
66	49 (45.0)	26 (63.4)	45 (52.3)	26 (41.3)		
Education					0.018	0.207
None/Primary	46 (42.2)	6 (14.6)	45 (52.3)	15 (23.8)		
Jr. Middle	25 (22.9)	8 (19.5)	11 (12.8)	19 (30.1)		
Sr. Middle	29 (26.6)	14 (34.2)	14 (16.3)	18 (28.6)		
Some college	9 (8.3)	13 (31.7)	16 (18.6)	11 (17.5)		
Self-reported body mass index					0.178	0.866
Underweight: <18.5	8 (7.3)	2 (4.9)	11 (12.8)	5 (7.9)		
Normal: 18.5 – 24.9	65 (59.6)	30 (73.2)	56 (65.1)	46 (73.0)		
Overweight: 25 – 29.9	31 (28.4)	9 (22.0)	14 (16.3)	11 (17.5)		
Obese: 30.0	5 (4.6)	0 (0.0)	5 (5.8)	1 (1.6)		
Ever smoker					0.877	0.671
No	102 (93.6)	16 (39.0)	80 (93.0)	22 (34.9)		
Yes	7 (6.4)	25 (61.0)	6 (7.0)	41 (65.1)		
Ever drinker					0.877	0.182
No	102 (93.6)	25 (61.0)	80 (93.0)	30 (47.6)		
Yes	7 (6.4)	16 (39.0)	6 (7.0)	33 (52.4)		
Diabetes					0.091	0.278
No	93 (85.3)	35 (85.4)	80 (93.0)	58 (92.1)		
Yes	16 (14.7)	6 (14.6)	6 (7.0)	5 (7.9)		
History of cholecystitis					< 0.001	< 0.001
No	56 (51.4)	21 (51.2)	77 (89.5)	59 (93.7)		
Yes	53 (48.6)	20 (48.8)	9 (10.5)	4 (6.3)		

 a Fisher's exact $\chi 2$ test was used for age group, self-reported body mass index and history of cholecystitis among men.

Table 2.

Association between Inflammation Proteins Tested from a High Sensitivity Cytokine Panel and Gallstone risk, Shanghai, China, 1997 – 2001.

Inflammation Proteins and Category	No. of Cases (%)	No. of Controls	Age- and Sex- Adjusted	Multivariable- Adjusted	Mutually-Adjusted	
		(%)	ORs (95% CIs) ^a	ORs (95% CIs) ^b	ORs (95% CIs) ^c	
IL-6						
Q1	19 (12.7)	38 (25.5)	reference	reference	reference	
Q2	29 (19.3)	38 (25.5)	1.4 (0.7, 3.0)	1.3 (0.6, 3.2)	1.0 (0.4, 2.7)	
Q3	31 (20.7)	36 (24.2)	1.7 (0.8, 3.6)	1.3 (0.5, 3.1)	0.6 (0.2, 1.8)	
Q4	71 (47.3)	37 (24.8)	3.7 (1.9, 7.5)	3.2 (1.4, 7.2)	1.4 (0.5, 3.6)	
Ptrend d			7.29×10^{-5}	3.26×10^{-3}	0.473	
IL-10						
Q1	24 (16.0)	38 (25.5)	reference	reference	reference	
Q2	24 (16.0)	37 (24.8)	1.0 (0.5, 2.1)	1.0 (0.4, 2.3)	0.5 (0.2, 1.6)	
Q3	28 (18.7)	37 (24.8)	1.1 (0.5, 2.3)	1.3 (0.6, 3.2)	0.6 (0.2, 1.8)	
Q4	74 (49.3)	37 (24.8)	3.4 (1.6, 6.0)	3.1 (1.4, 7.0)	1.4 (0.5, 3.8)	
Ptrend d			2.42×10^{-4}	1.23×10^{-3}	0.267	
IL-12 (p70)						
Q1	21 (14.0)	38 (25.5)	reference	reference	reference	
Q2	14 (9.3)	37 (24.8)	0.6 (0.3, 1.5)	0.8 (0.3, 2.2)	0.9 (0.3, 2.7)	
Q3	38 (25.3)	38 (25.5)	1.7 (0.8, 3.4)	1.9 (0.8, 4.5)	1.8 (0.6, 4.9)	
Q4	77 (51.3)	36 (24.2)	3.6 (1.8, 7.1)	5.7 (2.5, 13.5)	5.5 (1.9, 16.2)	
P trend d			$6.01 imes 10^{-6}$	2.99×10^{-6}	0.002	
IL-13						
Q1	17 (11.3)	38 (25.5)	reference	reference	reference	
Q2	29 (19.3)	38 (25.5)	1.7 (0.8, 3.7)	1.7 (0.7, 4.1)	1.3 (0.5, 3.4)	
Q3	49 (32.7)	36 (24.2)	3.3 (1.6, 7.0)	4.3 (1.9, 10.4)	2.8 (1.0, 7.6)	
Q4	55 (36.7)	37 (24.8)	3.1 (1.5, 6.6)	3.2 (1.4, 7.5)	0.9 (0.3,2.7)	
Ptrend d			7.79×10^{-4}	$2.33 imes 10^{-3}$	0.890	

Abbreviations: CI, confidence interval; IL: interleukin; Q, quartile; OR, odds ratio

^aAdjusted for age and sex.

^bAdjusted for age, sex, body mass index, education level, smoking, alcohol drinking, self-reported history of diabetes, and history of cholecystitis.

^CFour cytokines were mutually adjusted.

 $d_{\text{Two-sided }P\text{ values for trend across protein categories were assessed with the Wald test using categorical values of the proteins with 1 degree of freedom.}$