



Published in final edited form as:

*Microcirculation*. 2018 October ; 25(7): e12490. doi:10.1111/micc.12490.

## Ovarian Hormones Modulate Endothelin-1 Receptor Responses in Young Women

Kelly N. Sebzda<sup>1</sup>, Andrew V. Kuczmarowski<sup>1</sup>, Ryan T. Pohlig<sup>2</sup>, Shannon L. Lennon<sup>1</sup>, David G. Edwards<sup>1</sup>, Megan M. Wenner<sup>1</sup>

<sup>1</sup>Department of Kinesiology and Applied Physiology, University of Delaware, Newark, DE

<sup>2</sup>Biostatistic Core Facility, College of Health Sciences, University of Delaware, Newark, DE

### Abstract

**Objective:** We recently demonstrated endothelin-B receptors (ETB<sub>R</sub>) mediate vasodilation in young but not postmenopausal women; it is unclear if this is related to age or a decline in ovarian hormones. The purpose of this study was to test the hypothesis that ETB<sub>R</sub> responses are modulated by ovarian hormones.

**Methods:** We measured cutaneous vasodilatory responses in 12 young women (22±1 years, 23±1 kg/m<sup>2</sup>) during the mid-luteal (ML; days 20-25) and early follicular (EF; days 2-5) phases of the menstrual cycle. Cutaneous microdialysis perfusions of lactated Ringer (control), ETB<sub>R</sub> antagonist (BQ-788, 300nM), and ETA<sub>R</sub> antagonist (BQ-123, 500nM) were performed, followed by local heating to 42°C.

**Results:** Serum estradiol (ML: 118±16 vs. EF: 44±9 pg/ml, *P* < 0.05) and progesterone (ML: 8.3±1.0 vs. EF: 0.7±0.2 ng/ml, *P* < 0.05) were higher during ML vs. EF phase. ETB<sub>R</sub> blockade decreased vasodilation during ML (control: 91±2 vs. BQ-788: 83±2 %CVCmax, *P* < 0.05) but not EF (control: 89±2 vs. BQ-788: 89±1 %CVCmax). ETA<sub>R</sub> blockade also decreased vasodilation during ML (control: 91±2 vs. BQ-123: 87±2 %CVCmax, *P* < 0.05) but not EF (control: 89±2 vs. BQ-123: 92±2 %CVCmax).

**Conclusions:** These data suggest that fluctuations in ovarian hormones modulate ETB<sub>R</sub> and ETA<sub>R</sub> responses in young women.

### Keywords

cutaneous microdialysis; endothelin-B receptor; sex hormones

---

**Corresponding Author:** Megan M. Wenner, PhD, 540 South College Ave, STAR Health Sciences Complex Rm 201N, Newark, DE 19713, Phone: 302-831-7343, mwenner@udel.edu.

Research was performed at the University of Delaware

#### AUTHOR CONTRIBUTIONS

KNS, AVK and MMW performed the experiments; KNS, AVK, RTP, and MMW analyzed the data; KNS, RTP, SLL, DGE and MMW interpreted the results of the experiments; AVK and MMW prepared the figures; DGE and MMW contributed to the conception and design of the research; KNS, AVK, and MMW drafted the manuscript; KNS, AVK, RTP, SLL, DGE and MMW edited and revised the manuscript; KNS, AVK, RTP, SLL, DGE and MMW approved the final version of the manuscript.

#### DISCLOSURES

None

## INTRODUCTION

ET-1 is a powerful vasoconstrictor that is released by the endothelium and is involved in regulating vascular tone and function<sup>1,2</sup>. ET-1 binds to two different receptors in the vasculature: ETA<sub>R</sub> and ETB<sub>R</sub><sup>3</sup>. Both receptors are present on vascular smooth muscle cells and elicit vasoconstriction and cell proliferation. Conversely, ETB<sub>R</sub> are also found on the vascular endothelium and facilitate vasodilation through the stimulation of, at least in part, release of NO<sup>4-6</sup>. Given the contrasting actions of these two ET-1 receptors<sup>7</sup>, much attention has been given to the influence of these receptors on cardiovascular function.

Sex differences in cardiovascular function are well recognized, and the mechanisms contributing to sex differences in cardiovascular disease rates have been the focus of numerous investigations in recent years. Sex differences in the ET-1 system are also apparent, and may be a primary contributing factor to the divergent cardiovascular disease rates between men and women throughout the lifespan. There is accumulating evidence demonstrating ETB<sub>R</sub> are important for vascular health in women<sup>5,8-10</sup>. For example, the proportion of ETB<sub>R</sub> to ETA<sub>R</sub> is greater in women (1:1) compared to men (1:3)<sup>4</sup>. In young women, ETB<sub>R</sub> mediate vasodilation<sup>5,8,10</sup>, but elicit vasoconstriction in young men<sup>5</sup>. In postmenopausal women, ET-1 mediated vasoconstriction is lower compared to men of similar age, but blockade of ETB<sub>R</sub> ameliorates this sex difference in ET-1 constriction<sup>9</sup>. We recently demonstrated that ETB<sub>R</sub> mediated vasodilation in young women is lost after menopause<sup>10</sup>. However, it is not clear whether this is due to age-related changes in the ET-1 system, or primarily to the fluctuations in endogenous ovarian hormones.

Fluctuations in circulating ovarian hormones influence the ET-1 system. Studies in animal and cell models indicate that ovarian hormones, specifically E<sub>2</sub>, modulate ET-1 and its receptors. E<sub>2</sub> significantly lowers ET-1 concentration, mRNA expression, and secretion from HUVECs<sup>11,12</sup>. Administration of E<sub>2</sub> up-regulates the expression of ETB<sub>R</sub> in the coronary arteries of hyperlipidemic rabbits<sup>13</sup>, while down-regulating ETB<sub>R</sub> expression on the smooth muscle cells of the ventricular myocardium of rats<sup>14</sup>. In humans, plasma ET-1 is lower in women than in men and is even lower in pregnant women<sup>15</sup>, showing an inverse relationship with E<sub>2</sub> and P<sub>4</sub>. However, the influence of hormonal fluctuations in E<sub>2</sub> and P<sub>4</sub> on ET-1 receptor responses in humans is unclear.

With this background in mind, and given the impact of circulating sex steroids on vascular function, it is important to understand if fluctuations in ovarian hormones modulate ET-1 receptor responses. To our knowledge, no studies have examined their effects in young women. Accordingly, the purpose of this study was to examine ET-1 receptor responses during two distinct phases of the menstrual cycle: the EF phase (when both E<sub>2</sub> and P<sub>4</sub> are low), and the ML phase (when both E<sub>2</sub> and P<sub>4</sub> are elevated). We hypothesized that ETB<sub>R</sub> mediated vasodilation would be greater during ML compared to EF.

## MATERIALS AND METHODS

### Subjects

Twelve young women between the ages of 18-30 years participated in this study. All subjects were non-obese (BMI < 30 kg/m<sup>2</sup>), non-smokers, free from any known cardiovascular disease, and had regular menstrual cycles (1 cycle every approximately 28 days). Women were excluded if they were taking oral contraceptives, had irregular cycles, or if they were pregnant or breast-feeding, all of which affect endogenous sex steroid levels<sup>16</sup>. Women were also excluded if they had any history of cardiovascular or metabolic disease, or any other chronic disease. All experimental procedures and protocols were approved by the Institutional Review Board at the University of Delaware and conformed to the guidelines set forth by the Declaration of Helsinki. All women gave verbal and written consent prior to study participation.

### Screening Visit

Women reported to the Nurse Managed Primary Care Center at the University of Delaware in the morning after an overnight fast for a standard physical exam to ensure eligibility for study participation. A medical history and menstrual history questionnaire were completed and reviewed by a Nurse Practitioner. Height, weight, resting BP (seated), and resting electrocardiogram were measured. A blood sample was taken to assess standard blood chemistry such as cholesterol, glucose, hemoglobin, and hematocrit.

### Experimental Visits

Women participated in two identical experimental visits, which were conducted during two different phases of their menstrual cycle (EF and ML). Consistent with previous studies, the EF phase visit occurred during days 2-5 of the menstrual cycle and the ML phase visit occurred during days 20-25 of the menstrual cycle<sup>17-20</sup>. Cycle phase was confirmed by serum sex hormones; a serum P<sub>4</sub> level of >5ng was used to confirm ovulation<sup>21</sup>. Women refrained from exercise for 24 hours, alcohol and caffeine for 12 hours, and food for at least 4 hours prior to testing. Upon arrival, women provided a small urine sample for an over-the-counter pregnancy test and were weighed. After lying supine for at least 20 minutes, a blood sample was taken to assess serum E<sub>2</sub> and P<sub>4</sub>, as well as plasma ET-1. Women were instrumented for measures of BP (Dinamap DASH 2000, GE Medical, Chicago, IL) and intradermal microdialysis coupled with LDF.

### Microvascular Vasodilatory Function Assessment

We measured vasodilatory function in the cutaneous microcirculation using LDF coupled with cutaneous microdialysis. This approach allows for the measurement of SkBF while simultaneously administering pharmacological substances to garner insight into the specific mechanisms regulating vascular function<sup>22,23</sup>. Importantly, the cutaneous circulation has been used as an accessible and representative vascular bed to examine the mechanisms of peripheral microvascular vasodilatory function in humans<sup>10,23-30</sup>. Three microdialysis fibers (CMA 31 Linear, Harvard Apparatus, Holliston, MA) were inserted intradermally on the dorsal side of the right forearm as previously described<sup>10,27-30</sup>. The forearm was cleaned

with betadine and alcohol, and ice was applied for 10 minutes to the skin surface in order to provide a short-term local anesthetic. A 23-gauge needle was then inserted through the intradermal space to serve as a guide cannula. The entry and exit points of each needle were 2 cm apart, with each site separated by at least 1 inch. After insertion of all three needles, microdialysis fibers were threaded through the lumen of the needle and the needles were removed, leaving only the semi-permeable portion of the fiber under the dermis. All fibers were taped down and connected to a syringe pump (Bioanalytical Systems, West Lafayette, IN), and perfused with lactated Ringers solution (B. Braun Medical, Bethlehem, PA) at a rate of 2  $\mu\text{l}/\text{min}$  for 60-90 minutes to allow for recovery of local SkBF following fiber insertion. SkBF was measured as cutaneous red blood cell flux from 1.5  $\text{mm}^2$  of skin using a multi-fiber laser Doppler flow probe (Moor Instruments, Devon, UK) placed in a local heater (Moor Instruments), and secured on the skin with tape directly above each microdialysis site.

### Experimental Protocol

Local skin temperature was maintained at 32°C. The microdialysis sites were randomly perfused with either the ETB<sub>R</sub> antagonist BQ-788 (300 nM, Sigma-Aldrich, St. Louis, MO), the ETA<sub>R</sub> antagonist BQ-123 (500 nM, Sigma-Aldrich), or lactated Ringers solution to serve as the control site<sup>10,27,28</sup>; all sites were perfused for 45 minutes at a rate of 5  $\mu\text{l}/\text{min}$ . After the ET-1 receptor blocker perfusion, the temperature of the local heaters was increased to 42°C for approximately 30 minutes, or until a stable plateau in SkBF was reached; this plateau is largely mediated by NO<sup>31-33</sup>. Finally, all three syringes were perfused with SNP (28 mM, Marathon Pharmaceuticals, Northbrook, IL) at a rate of 5  $\mu\text{l}/\text{min}$  for approximately 15-20 minutes with the temperature of the local heaters set to 43°C to elicit maximal vasodilation<sup>27</sup>.

### Blood Analysis

Venous blood samples were collected in separate tubes without anticoagulants for the analysis of E<sub>2</sub> and P<sub>4</sub>. A separate blood sample was collected in an EDTA tube for the analysis of plasma ET-1. Tubes were centrifuged, the serum or plasma pipetted off, and frozen at -80°C until analysis. Serum E<sub>2</sub> and P<sub>4</sub> were measured using competitive ELISA (Alpco, Salem, NH, USA). The range for the E<sub>2</sub> assay was 20 - 3200 pg/ml with a sensitivity of 10 pg/ml. Intra-assay and inter-assay coefficients of variation for the E<sub>2</sub> assay were 2.71% and 1.42%, respectively. The range for the P<sub>4</sub> assay was 0.3 - 60 ng/ml with a sensitivity of 0.1 ng/ml. Intra-assay and inter-assay coefficients of variation for the P<sub>4</sub> assay were 3.52% and 1.42%, respectively. Plasma ET-1 was analyzed using an ELISA (R&D systems, Minneapolis, MN, USA). Intra- and inter-assay coefficients of variation were less than 2.5%. All samples were measured at a wavelength of 450nm on an Infinite F200 Pro microplate reader and data was analyzed with Magellan IQ software (Tecan Group Ltd., Männedorf, CH).

### Data Analysis

LDF data was recorded at 1000 Hz using PowerLab (ADInstruments, Bella Vista, New South Wales, Australia). We analyzed a 5-minute segment after the local heating plateau was reached (42°C), and a 2-minute segment during SNP + local heating to 43°C (maximum

dilation) for each site. CVC was calculated as  $SkBF/MAP$ . CVC was expressed as a percent of maximum dilation to account for site-to-site variations in  $SkBF$  and was used as an indicator of vasodilatory function<sup>10,28</sup>.

### Statistical Analyses

We compared vasodilatory responses to local heating in the ET-1 receptor-blocked sites (BQ-788 and BQ-123) and control site (lactated Ringers) during the EF and ML phases of the menstrual cycle using a 2x3 repeated measures ANOVA (cycle phase x  $SkBF$  site). Paired t-tests were used to compare serum  $E_2$  and  $P_4$ , along with weight and MAP between EF and ML visits. Significance was set at  $\alpha < 0.05$ . Results are expressed as mean  $\pm$  SEM.

## RESULTS

### Subject Characteristics

Subject characteristics from the screening visit are presented in Table 1. All women were normotensive, with standard blood chemistry levels within normal clinical limits. The average cycle length was  $29 \pm 1$  days. The racial demographics of our participants were as follows: 7 women were Caucasian, 2 Asian, 2 African American, and 1 woman was Hispanic. Subject characteristics from the two experimental visits (EF and ML phases of the menstrual cycle) are presented in Table 2. Blood analyses of ovarian hormones confirmed that women were in the appropriate phase (EF vs. ML) for the experimental visits. Serum  $E_2$  and  $P_4$  levels were significantly higher in women during the ML compared to the EF phase ( $P < 0.05$ , Table 2). Weight and BP were similar between EF and ML phases, as well as plasma ET-1 (Table 2).

### Vasodilatory Responses to ET-1 Receptor Blockade

During the ML phase, vasodilatory responses to  $ETB_R$  blockade were attenuated compared to control conditions (control:  $91 \pm 2\%$  vs BQ-788:  $83 \pm 2\%$ ,  $P < 0.05$ ; Figure 1). There was no change in %CVCmax with  $ETB_R$  blockade during EF (control:  $89 \pm 2\%$  vs BQ-788:  $89 \pm 1\%$ ,  $P > 0.05$ ; Figure 1). However, when comparing  $ETB_R$  blockade responses between phases, vasodilation was attenuated during ML compare to EF (ML:  $83 \pm 2\%$  vs EF:  $89 \pm 1\%$ ,  $P < 0.05$ ; Figure 1). Cutaneous vasodilatory responses to  $ETA_R$  blockade were also lower during ML (control:  $91 \pm 2\%$  vs BQ-123:  $87 \pm 2\%$ ,  $P < 0.05$ ; Figure 2), whereas there were no differences during EF (control:  $89 \pm 2\%$  vs BQ-123:  $92 \pm 2\%$ ,  $P > 0.05$ ; Figure 2). When comparing  $ETA_R$  blockade responses between phases, vasodilation was attenuated during ML compare to EF (ML:  $87 \pm 2\%$  vs EF:  $92 \pm 2\%$ ,  $P < 0.05$ ; Figure 2). Vasodilatory function at control sites were not different between ML and EF phases ( $91 \pm 2$  vs  $89 \pm 2\%$ ,  $P > 0.05$ ).

## DISCUSSION

The novel finding of the current investigation is that endogenous fluctuations in ovarian hormones during the menstrual cycle modulate ET-1 receptor responses in women. Specifically,  $ETB_R$  mediate dilation during the ML phase, when endogenous  $E_2$  and  $P_4$  are elevated. To our knowledge, this is the first study to examine *in vivo* ET-1 receptor responses to changing endogenous ovarian hormone levels in young women. We previously examined

ET-1 receptor responses in young and postmenopausal women, and demonstrated that ETB<sub>R</sub> mediated dilation is lost after menopause<sup>10</sup>. Results from the current study extend our previous findings by suggesting that changes in sex steroid levels alter ETB<sub>R</sub> and ETA<sub>R</sub> responses in women. Taken together, these data are an important first step in demonstrating the interactions between ovarian hormones and ET-1 receptor responses in women.

Although the importance of ET-1 and its involvement with vascular tone and function are well known, less is understood regarding regulation of ET-1 and its receptors by ovarian hormones. Evidence in animal models suggest a direct relation between alterations in ovarian hormones and ET-1 receptor expression. For example, ovariectomy caused an increase in ETB<sub>R</sub> mRNA and protein expression in the myocardium<sup>14</sup> and kidney<sup>34</sup> of animal models, which was reversed with administration of either E<sub>2</sub><sup>14</sup> or E<sub>2</sub> and P<sub>4</sub><sup>34</sup>. Although we did not measure changes in receptor expression in the current study, it is possible that the ETB<sub>R</sub>-mediated dilation during the ML phase was related to an upregulation of endothelial ETB<sub>R</sub> or downregulation of vascular smooth muscle ETB<sub>R</sub>. However, while E<sub>2</sub> treatment increased ETB<sub>R</sub> expression and elicited vessel relaxation in rabbits, the addition of P<sub>4</sub> negated these changes in receptor expression and abolished the E<sub>2</sub> vasorelaxation<sup>13</sup>. With this in mind, and given that both E<sub>2</sub> and P<sub>4</sub> were elevated during the ML phase in our study, changes in receptor expression may not solely explain our findings. Stimulation of the ETB<sub>R</sub> can also result in increased NO and prostacyclin<sup>35</sup>, which may be in part related to the results of the current study. It is well appreciated that E<sub>2</sub> increases NO release to elicit dilation. Furthermore, both E<sub>2</sub> and P<sub>4</sub> increase prostacyclin production in animal and cell culture models<sup>36-38</sup>. Thus, our findings may also be related to pathways downstream of ETB<sub>R</sub> and/or other signaling mechanisms related to changes in ovarian hormones.

Our data also suggest that endogenous hormone production impacts ETA<sub>R</sub> responses, as we observed a reduction in vasodilation with blockade of ETA<sub>R</sub> during the ML phase. Since ETA<sub>R</sub> mediate vasoconstriction, we interpret these findings to suggest that elevations in endogenous ovarian hormones attenuate the vasoconstrictor capacity of ETA<sub>R</sub>. In this manner, these findings are consistent with data in animal models where ETA<sub>R</sub> expression on vascular smooth muscle cells was reduced with E<sub>2</sub> administration<sup>39</sup>. However, ETA<sub>R</sub> and ETB<sub>R</sub> can form heterodimers and thus functionally interact<sup>40,41</sup>, whereby stimulation of one receptor could impact the function of the other. It is possible that this receptor interaction occurred in our sample and may have impacted our results. In other words, if ETB<sub>R</sub> expression was greater during the ML phase, then stimulation of these receptors may impact the ETA<sub>R</sub> and partially explain the reduction in vasodilation with blockade of ETA<sub>R</sub> during the ML phase. Alternatively, there may also be biased ligands for ETA<sub>R</sub> such that blockade of this receptor may elicit differential signaling<sup>41-43</sup>, contributing to the reduction in vasodilation with blockade of ETA<sub>R</sub>. Finally, it is also possible that changes in estrogen receptors interact with ET-1 signaling and have impacted our data. Expression of the ERα receptor changes during the menstrual cycle<sup>44</sup>, and an over-expression of ERα receptors in cell models inhibits ET-1 release<sup>45</sup>. Further, activation of estrogen receptors such as ERα or GPER using selective estrogen receptor modulators reduce ET-1 mediated constriction<sup>46-48</sup>. Thus, ET-1 signaling and receptor activity may be influenced directly by fluctuations in ovarian hormones or also through changes in gonadal steroid receptors.

In the current study, we demonstrate that ETB<sub>R</sub> mediate dilation during the ML phase of the menstrual cycle, and observed no impact of blocking ETB<sub>R</sub> during the EF phase. This was in contrast to our recent publication where we demonstrated ETB<sub>R</sub> mediate dilation in young women tested during the EF phase<sup>10</sup>. However, our prior publication included women using hormonal contraceptives, and were tested during the ‘low hormone’ or ‘placebo’ week of the pill. Although we were not powered to examine differences in ETB<sub>R</sub> responses between those who were using hormonal contraceptives and those who were not, it is possible that the chronic exposure from exogenous hormones impacted ETB<sub>R</sub> responses, and may partially explain in the current study (with no women using exogenous hormones) why we saw a minimal effect of blocking ETB<sub>R</sub> during the EF phase. Alternatively, while often termed the ‘low hormone’ phase of the pill (placebo week), women may experience an endogenous surge in hormone production during the placebo week<sup>16,49-52</sup>; based on the finding from the current study, it is likely that endogenous hormone production impacts ETB<sub>R</sub> responses. Our findings may also help to explain prior reports of sex differences in ET-1 receptor responses on vascular function, as we demonstrate that female reproductive hormones have a strong influence on ET-1 receptor responses. The receptor density of ETA<sub>R</sub> to ETB<sub>R</sub> is greater in men compared to women, and predominately mediate vasoconstriction. Based on current literature, we speculate that the greatest difference in ET-1 receptor responses would be between men and women tested during the ML phase. Further study is needed to characterize the impact of sex vs. sex hormones on the ET-1 system.

We did not observe statistical differences in the vasodilatory responses at the control site between the two phases of the menstrual cycle. We utilized a standard heating protocol that has been well established to elicit NO-mediated dilation<sup>31-33</sup> and mirrors the response to acetylcholine perfusions via microdialysis<sup>53</sup>. However, the amount of dilation achieved via local heating is proportional to the temperature used<sup>54,55</sup>, and therefore we may have achieved maximal dilation in both phases. To this end, a recent paper by Choi and colleagues<sup>56</sup> demonstrated that a rapid heating protocol to 39°C may offer an improved approach to isolate NO-depend dilation in the microcirculation. Using a ramping protocol for local skin heating (a slower and graded increase), vasodilatory responses were shown to be greater during the high hormone phase of oral contraceptive use compared to the placebo / low hormone phase<sup>57</sup>. To our knowledge, ours is the first study to examine changes in cutaneous vasodilation during *endogenous* hormonal fluctuations. Interestingly, our findings in the cutaneous microcirculation also mirror those of conduit artery endothelial function; although flow-mediated dilation of the brachial artery increases during ovulation, it declines during the luteal phase such that there are no differences between EF and ML phases<sup>58</sup>. Based on findings using a controlled hormone intervention to isolate the independent effects of E<sub>2</sub>, the increase in vasodilation is primarily related to E<sub>2</sub>, and these effects are negated by the addition of P<sub>4</sub><sup>59</sup>. Taken together, we speculate that chronic exposure to exogenous sex steroids alters vasodilatory function in women, and further supports that exogenous and endogenous hormones may have differentiating effects on vascular function. Indeed, there are many formulations of hormonal contraceptives which can impact tissues differently, in particular the vascular endothelium<sup>60-62</sup>. Further study is needed to fully understand the

impact of endogenous versus exogenous hormones on vascular function, especially given the number of women using hormonal contraceptives.

### Limitations

Several vasoactive substances are involved in regulating vascular function, such as NO, prostaglandins, angiotensin II, and norepinephrine<sup>63-65</sup>, all of which are influenced by E<sub>2</sub><sup>66</sup> and interact with ET-1<sup>67</sup>. It is possible that the effects we observed in our study are related to the influence of E<sub>2</sub> on one or more of these substances; we are not able to determine which additional mechanisms may be involved from our data. Unfortunately, we did not have additional blood samples to also measure NO, angiotensin II, prostaglandins, or norepinephrine to gain insight into these other pathways. Additional study is needed to further characterize the interactions among ovarian hormones and the mechanisms regulating vascular function. Also, since E<sub>2</sub> has been shown to reduce circulating ET-1<sup>11,12</sup>, it is also possible that our findings are related to a reduced production of ET-1. However, we did not observe differences in plasma ET-1 between ML and EF. Vasoconstrictor responses to ET-1, independent of maximum vasoconstrictor capability, do not differ between men and women, despite showing sex differences in ETA<sub>R</sub> and ETB<sub>R</sub> responses<sup>9</sup>, suggesting that the receptors – and not ET-1 itself – play a crucial role in determining the vascular response. Nevertheless, our data indicate that ovarian hormonal fluctuations influence both ETB<sub>R</sub> and ETA<sub>R</sub> *in vivo*.

### Conclusions

In conclusion, we demonstrate that ETB<sub>R</sub> blockade reduced vasodilation in young women during the ML phase of the menstrual cycle. These data suggest that ETB<sub>R</sub> mediate vasodilation when endogenous E<sub>2</sub> and P<sub>4</sub> levels are high. This extends previous findings of ETB<sub>R</sub> mediating vasodilation in young women<sup>5,8,10</sup> by demonstrating fluctuations in hormones play a primary role in modulating these responses. In addition, we saw a reduction in vasodilation during ETA<sub>R</sub> blockade, suggesting that high levels of circulating ovarian hormones impact ETA<sub>R</sub> responses as well. These findings are an important first step in understanding the mechanisms by which changes in endogenous hormone exposure may impact vascular function in women.

### Perspectives

Our data demonstrate that fluctuations in endogenous ovarian hormones that occur during the menstrual cycle impact ETB<sub>R</sub> and ETA<sub>R</sub> in healthy women. Alterations in endogenous hormone production that occur with menopause<sup>10</sup> or reproductive disorders such as polycystic ovary syndrome<sup>27</sup> display a dysregulation of ETB<sub>R</sub> responses in the vasculature. These data further support the link between sex hormones and the ET-1 system on cardiovascular function.

### ACKNOWLEDGMENTS

The authors wish to thank Stephanie Mraz and Kenneth Kirschner for assistance with data collection/analysis, and the participants for their time.



**Funding Support:** This research was supported in part by the DE-CTR Accel Program (U54-GM104941), the DE-INBRE Program (5 P20 GM103446-13), the University of Delaware Research Foundation, and P20 GM113125 (Center of Biomedical Research Excellence).

**Funding Support:** U54-GM104941, 5 P20 GM103446-13, P20 GM113125, and the University of Delaware Research Foundation

## List of Abbreviations

<b>BP</b>	blood pressure
<b>BMI</b>	body mass index
<b>CVC</b>	cutaneous vascular conductance
<b>EF</b>	early follicular
<b>E<sub>2</sub></b>	estradiol
<b>EDTA</b>	ethylenediaminetetraacetic acid
<b>ELISA</b>	enzyme-linked immunosorbent assays
<b>ET-1</b>	Endothelin-1
<b>ETA<sub>R</sub></b>	endothelin A receptor
<b>ETB<sub>R</sub></b>	endothelin B receptor
<b>HUVECs</b>	human umbilical vein endothelial cells
<b>LDF</b>	laser Doppler flowmetry
<b>MAP</b>	mean arterial pressure
<b>ML</b>	mid-luteal
<b>NO</b>	nitric oxide
<b>P<sub>4</sub></b>	progesterone
<b>SkBF</b>	skin blood flow
<b>SNP</b>	sodium nitroprusside

## REFERENCES

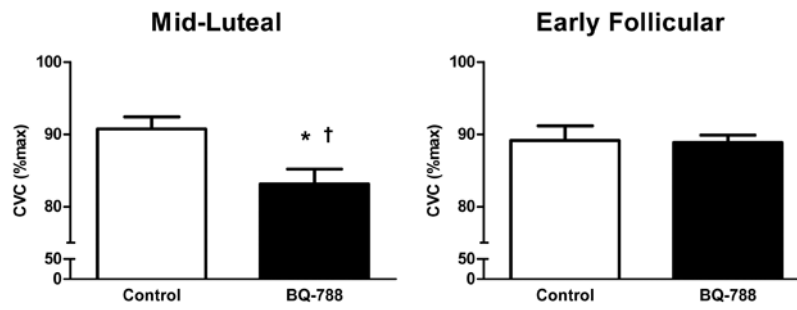
1. Brain SD, Crossman DC, Buckley TL, Williams TJ. Endothelin-1: demonstration of potent effects on the microcirculation of humans and other species. *J Cardiovasc Pharmacol.* 1989; 13 Suppl 5:S147–149; discussion S150. [PubMed: 2473291]
2. Yanagisawa M, Kurihara H, Kimura S, et al. A novel potent vasoconstrictor peptide produced by vascular endothelial cells. *Nature.* 1988;332(6163):411–415. [PubMed: 2451132]
3. Bull HA, Bunker CB, Terenghi G, et al. Endothelin-1 in human skin: immunolocalization, receptor binding, mRNA expression, and effects on cutaneous microvascular endothelial cells. *J Invest Dermatol.* 1991;97(4):618–623. [PubMed: 1658154]

4. Ergul A, Shoemaker K, Puett D, Tackett RL. Gender differences in the expression of endothelin receptors in human saphenous veins in vitro. *J Pharmacol Exp Ther*. 1998;285(2):511–517. [PubMed: 9580591]
5. Kellogg DL Jr., Liu Y, Pergola PE. Selected contribution: Gender differences in the endothelin-B receptor contribution to basal cutaneous vascular tone in humans. *J Appl Physiol*. 2001;91(5):2407–2411; discussion 2389–2490. [PubMed: 11641388]
6. Kohan DE, Rossi NF, Inscho EW, Pollock DM. Regulation of blood pressure and salt homeostasis by endothelin. *Physiol Rev*. 2011;91(1):1–77. [PubMed: 21248162]
7. Schneider MP, Boesen EI, Pollock DM. Contrasting actions of endothelin ET(A) and ET(B) receptors in cardiovascular disease. *Annu Rev Pharmacol Toxicol*. 2007;47:731–759. [PubMed: 17002597]
8. Stanhewicz AE, Jandu S, Santhanam L, Alexander LM. Alterations in endothelin type B receptor contribute to microvascular dysfunction in women who have had preeclampsia. *Clin Sci (Lond)*. 2017;131(23):2777–2789. [PubMed: 29042489]
9. Stauffer BL, Westby CM, Greiner JJ, Van Guilder GP, Desouza CA. Sex differences in endothelin-1-mediated vasoconstrictor tone in middle-aged and older adults. *Am J Physiol Regul Integr Comp Physiol*. 2010;298(2):R261–265. [PubMed: 19939973]
10. Wenner MM, Sebzda KN, Kuczmarski AV, Pohlig RT, Edwards DG. ETB receptor contribution to vascular dysfunction in postmenopausal women. *Am J Physiol Regul Integr Comp Physiol*. 2017;313(1):R51–R57. [PubMed: 28438762]
11. Bilsel AS, Moini H, Tetik E, Aksungar F, Kaynak B, Ozer A. 17Beta-estradiol modulates endothelin-1 expression and release in human endothelial cells. *Cardiovasc Res*. 2000;46(3):579–584. [PubMed: 10912468]
12. Juan SH, Chen JJ, Chen CH, et al. 17beta-estradiol inhibits cyclic strain-induced endothelin-1 gene expression within vascular endothelial cells. *Am J Physiol Heart Circ Physiol*. 2004;287(3):H1254–1261. [PubMed: 15130882]
13. Pedersen SH, Nielsen LB, Mortensen A, Nilas L, Ottesen B. Progestins oppose the effects of estradiol on the endothelin-1 receptor type B in coronary arteries from ovariectomized hyperlipidemic rabbits. *Menopause*. 2008;15(3):503–510. [PubMed: 18188139]
14. Nuedling S, van Eickels M, Allera A, et al. 17 Beta-estradiol regulates the expression of endothelin receptor type B in the heart. *Br J Pharmacol*. 2003;140(1):195–201. [PubMed: 12967949]
15. Polderman KH, Stehouwer CD, van Kamp GJ, Dekker GA, Verheugt FW, Gooren LJ. Influence of sex hormones on plasma endothelin levels. *Ann Intern Med*. 1993;118(6):429–432. [PubMed: 8439117]
16. Stachenfeld NS, Taylor HS. Challenges and methodology for testing young healthy women in physiological studies. *Am J Physiol Endocrinol Metab*. 2014;306(8):E849–853. [PubMed: 24569589]
17. Carter JR, Lawrence JE, Klein JC. Menstrual cycle alters sympathetic neural responses to orthostatic stress in young, eumenorrheic women. *Am J Physiol Endocrinol Metab*. 2009;297(1):E85–91. [PubMed: 19401460]
18. Fu Q, Okazaki K, Shibata S, et al. Menstrual cycle effects on sympathetic neural responses to upright tilt. *J Physiol*. 2009;587(Pt 9):2019–2031. [PubMed: 19237424]
19. Lawrence JE, Ray CA, Carter JR. Vestibulosympathetic reflex during the early follicular and midluteal phases of the menstrual cycle. *Am J Physiol Endocrinol Metab*. 2008;294(6):E1046–1050. [PubMed: 18398013]
20. Minson CT, Halliwill JR, Young TM, Joyner MJ. Influence of the menstrual cycle on sympathetic activity, baroreflex sensitivity, and vascular transduction in young women. *Circulation*. 2000;101(8):862–868. [PubMed: 10694525]
21. Leiva R, Bouchard T, Boehringer H, Abulla S, Ecochard R. Random serum progesterone threshold to confirm ovulation. *Steroids*. 2015;101:125–129. [PubMed: 26111590]
22. Cracowski JL, Minson CT, Salvat-Melis M, Halliwill JR. Methodological issues in the assessment of skin microvascular endothelial function in humans. *Trends Pharmacol Sci*. 2006;27(9):503–508. [PubMed: 16876881]

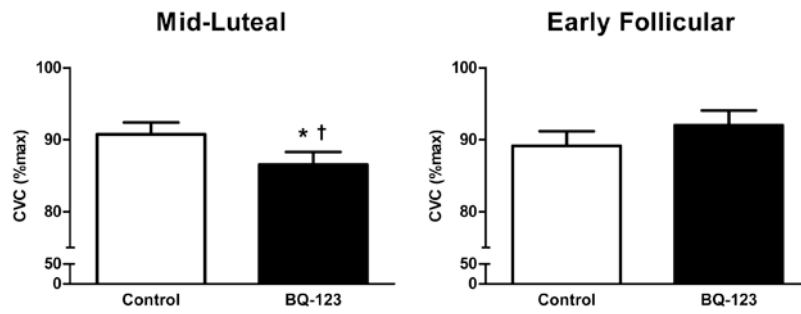
23. Holowatz LA, Thompson-Torgerson CS, Kenney WL. The human cutaneous circulation as a model of generalized microvascular function. *J Appl Physiol.* 2008;105(1):370–372. [PubMed: 17932300]
24. Abularrage CJ, Sidawy AN, Aidinian G, Singh N, Weiswasser JM, Arora S. Evaluation of the microcirculation in vascular disease. *J Vasc Surg.* 2005;42(3):574–581. [PubMed: 16171612]
25. I Jzerman R, de Jongh RT, Beijk MA, et al. Individuals at increased coronary heart disease risk are characterized by an impaired microvascular function in skin. *Eur J Clin Invest.* 2003;33(7):536–542. [PubMed: 12814388]
26. Stewart J, Kohen A, Brouder D, et al. Noninvasive interrogation of microvasculature for signs of endothelial dysfunction in patients with chronic renal failure. *Am J Physiol Heart Circ Physiol.* 2004;287(6):H2687–2696. [PubMed: 15297253]
27. Wenner MM, Taylor HS, Stachenfeld NS. Androgens influence microvascular dilation in PCOS through ET-A and ET-B receptors. *Am J Physiol Endocrinol Metab.* 2013;305(7):E818–825. [PubMed: 23921139]
28. Wenner MM, Taylor HS, Stachenfeld NS. Endothelin B receptor contribution to peripheral microvascular function in women with polycystic ovary syndrome. *J Physiol.* 2011;589(Pt 19):4671–4679. [PubMed: 21825025]
29. Wenner MM, Taylor HS, Stachenfeld NS. Peripheral Microvascular Vasodilatory Response to Estradiol and Genistein in Women with Insulin Resistance. *Microcirculation.* 2015;22(5):391–399. [PubMed: 25996650]
30. Wenner MM, Taylor HS, Stachenfeld NS. Progesterone enhances adrenergic control of skin blood flow in women with high but not low orthostatic tolerance. *J Physiol.* 2011;589(4):975–986. [PubMed: 21173076]
31. Bruning RS, Santhanam L, Stanhewicz AE, et al. Endothelial nitric oxide synthase mediates cutaneous vasodilation during local heating and is attenuated in middle-aged human skin. *J Appl Physiol.* 2012;112(12):2019–2026. [PubMed: 22500004]
32. Kellogg DL Jr., Liu Y, Kosiba IF, O'Donnell D. Role of nitric oxide in the vascular effects of local warming of the skin in humans. *J Appl Physiol.* 1999;86(4):1185–1190. [PubMed: 10194201]
33. Minson CT, Berry LT, Joyner MJ. Nitric oxide and neurally mediated regulation of skin blood flow during local heating. *J Appl Physiol.* 2001;91(4):1619–1626. [PubMed: 11568143]
34. Gohar EY, Yusuf C, Pollock DM. Ovarian hormones modulate endothelin A and B receptor expression. *Life Sci.* 2016;159:148–152. [PubMed: 26776836]
35. Gohar EY, Giachini FR, Pollock DM, Tostes RC. Role of the endothelin system in sexual dimorphism in cardiovascular and renal diseases. *Life Sci.* 2016;159:20–29. [PubMed: 26939577]
36. Geary GG, Krause DN, Duckles SP. Estrogen reduces mouse cerebral artery tone through endothelial NOS- and cyclooxygenase-dependent mechanisms. *Am J Physiol Heart Circ Physiol.* 2000;279(2):H511–519. [PubMed: 10924048]
37. Hermenegildo C, Oviedo PJ, Garcia-Martinez MC, Garcia-Perez MA, Tarin JJ, Cano A. Progestogens stimulate prostacyclin production by human endothelial cells. *Hum Reprod.* 2005;20(6):1554–1561. [PubMed: 15734756]
38. Sobrino A, Oviedo PJ, Novella S, et al. Estradiol selectively stimulates endothelial prostacyclin production through estrogen receptor- $\alpha$ . *J Mol Endocrinol.* 2010;44(4):237–246. [PubMed: 20110403]
39. Wang TH, Tan Z, Liu PQ, Lu W, Yang D, Pan JY. [Down-regulation of ETA receptor of vascular smooth muscle cells by 17 beta-estradiol]. *Sheng Li Xue Bao.* 2001;53(5):380–384. [PubMed: 11833423]
40. Kapsokalyvas D, Schiffers PM, Maij N, et al. Imaging evidence for endothelin ETA/ETB receptor heterodimers in isolated rat mesenteric resistance arteries. *Life Sci.* 2014; 111(1-2):36–41. [PubMed: 25066928]
41. Watts SW. Endothelin receptors: what's new and what do we need to know? *Am J Physiol Regul Integr Comp Physiol.* 2010;298(2):R254–260. [PubMed: 19907001]
42. De Mey JG, Compeer MG, Lemkens P, Meens MJ. ETA-receptor antagonists or allosteric modulators? *Trends Pharmacol Sci.* 2011;32(6):345–351. [PubMed: 21481481]

43. Maguire JJ. Evidence for biased agonists and antagonists at the endothelin receptors. *Life Sci.* 2016;159:30–33. [PubMed: 26898124]
44. Gavin KM, Seals DR, Silver AE, Moreau KL. Vascular endothelial estrogen receptor alpha is modulated by estrogen status and related to endothelial function and endothelial nitric oxide synthase in healthy women. *J Clin Endocrinol Metab.* 2009;94(9):3513–3520. [PubMed: 19509105]
45. Ali SH, O'Donnell AL, Mohamed S, Mousa S, Dandona P. Stable over-expression of estrogen receptor-alpha in ECV304 cells inhibits proliferation and levels of secreted endothelin-1 and vascular endothelial growth factor. *Mol Cell Endocrinol.* 1999;152(1-2):1–9. [PubMed: 10432218]
46. Barton M, Meyer MR. Postmenopausal hypertension: mechanisms and therapy. *Hypertension.* 2009;54(1):11–18. [PubMed: 19470884]
47. Meyer MR, Field AS, Kanagy NL, Barton M, Prossnitz ER. GPER regulates endothelin-dependent vascular tone and intracellular calcium. *Life Sci.* 2012;91(13-14):623–627. [PubMed: 22326502]
48. Meyer MR, Prossnitz ER, Barton M. The G protein-coupled estrogen receptor GPER/GPR30 as a regulator of cardiovascular function. *Vascul Pharmacol.* 2011;55(1-3):17–25. [PubMed: 21742056]
49. Creinin MD, Lippman JS, Eder SE, Godwin AJ, Olson W. The effect of extending the pill-free interval on follicular activity: triphasic norgestimate/35 micro g ethinyl estradiol versus monophasic levonorgestrel/20 micro g ethinyl estradiol. *Contraception.* 2002;66(3):147–152. [PubMed: 12384201]
50. Schlaff WD, Lynch AM, Hughes HD, Cedars MI, Smith DL. Manipulation of the pill-free interval in oral contraceptive pill users: the effect on follicular suppression. *Am J Obstet Gynecol.* 2004;190(4):943–951. [PubMed: 15118618]
51. van Heusden AM, Fauser BC. Activity of the pituitary-ovarian axis in the pill-free interval during use of low-dose combined oral contraceptives. *Contraception.* 1999;59(4):237–243. [PubMed: 10457868]
52. Wenner MM, Stachenfeld NS. Blood pressure and water regulation: understanding sex hormone effects within and between men and women. *J Physiol.* 2012;590(Pt 23):5949–5961. [PubMed: 23027816]
53. Stanhewicz AE, Jandu S, Santhanam L, Alexander LM. Increased Angiotensin II Sensitivity Contributes to Microvascular Dysfunction in Women Who Have Had Preeclampsia. *Hypertension.* 2017;70(2):382–389. [PubMed: 28652473]
54. Charkoudian N. Skin blood flow in adult human thermoregulation: how it works, when it does not, and why. *Mayo Clin Proc.* 2003;78(5):603–612. [PubMed: 12744548]
55. Pergola PE, Kellogg DL Jr., Johnson JM, Kosiba WA, Solomon DE. Role of sympathetic nerves in the vascular effects of local temperature in human forearm skin. *Am J Physiol.* 1993;265(3 Pt 2):H785–792. [PubMed: 8214111]
56. Choi PJ, Brunt VE, Fujii N, Minson CT. New approach to measure cutaneous microvascular function: an improved test of NO-mediated vasodilation by thermal hyperemia. *J Appl Physiol (1985).* 2014;117(3):277–283. [PubMed: 24903917]
57. Charkoudian N, Stephens DP, Pirkle KC, Kosiba WA, Johnson JM. Influence of female reproductive hormones on local thermal control of skin blood flow. *J Appl Physiol.* 1999;87(5):1719–1723. [PubMed: 10562614]
58. Williams MR, Westerman RA, Kingwell BA, et al. Variations in endothelial function and arterial compliance during the menstrual cycle. *J Clin Endocrinol Metab.* 2001;86(11):5389–5395. [PubMed: 11701712]
59. Miner JA, Martini ER, Smith MM, et al. Short-term oral progesterone administration antagonizes the effect of transdermal estradiol on endothelium-dependent vasodilation in young healthy women. *Am J Physiol Heart Circ Physiol.* 2011;301(4):H1716–1722. [PubMed: 21856917]
60. Meendering JR, Torgimson BN, Miller NP, Kaplan PF, Minson CT. A combined oral contraceptive containing 30 mcg ethinyl estradiol and 3.0 mg drospirenone does not impair endothelium-dependent vasodilation. *Contraception.* 2010;82(4):366–372. [PubMed: 20851231]
61. Meendering JR, Torgimson BN, Miller NP, Kaplan PF, Minson CT. Ethinyl estradiol-to-desogestrel ratio impacts endothelial function in young women. *Contraception.* 2009;79(1):41–49. [PubMed: 19041440]

62. Torgrimson BN, Meendering JR, Kaplan PF, Minson CT. Endothelial Function Across An Oral Contraceptive Cycle In Women Using Levonorgestrel And Ethinyl Estradiol. *Am J Physiol Heart Circ Physiol*. 2007.
63. Ignarro LJ, Buga GM, Wood KS, Byrns RE, Chaudhuri G. Endothelium-derived relaxing factor produced and released from artery and vein is nitric oxide. *Proc Natl Acad Sci U S A*. 1987;84(24):9265–9269. [PubMed: 2827174]
64. Moncada S, Vane JR. Pharmacology and endogenous roles of prostaglandin endoperoxides, thromboxane A2, and prostacyclin. *Pharmacol Rev*. 1978;30(3):293–331. [PubMed: 116251]
65. Palmer RM, Ferrige AG, Moncada S. Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor. *Nature*. 1987;327(6122):524–526. [PubMed: 3495737]
66. Tostes RC, Nigro D, Fortes ZB, Carvalho MH. Effects of estrogen on the vascular system. *Braz J Med Biol Res*. 2003;36(9):1143–1158. [PubMed: 12937779]
67. Rubanyi GM, Botelho LH. Endothelins. *FASEB J*. 1991;5(12):2713–2720. [PubMed: 1916094]



**Figure 1:** Cutaneous vasodilatory responses expressed as cutaneous vascular conductance (CVC %max) during microdialysis perfusion of lactated Ringer (Control) and ETBR blockade (BQ-788) during the Mid-Luteal Phase (left) and Early Follicular Phase (right) of the menstrual cycle. Data are presented as means  $\pm$  SEM. \*  $P < 0.05$  vs control. †  $P < 0.05$  vs. Early Follicular Phase.



**Figure 2:** Cutaneous vasodilatory responses expressed as cutaneous vascular conductance (CVC %max) during microdialysis perfusion of lactated Ringer (Control) and  $ETA_R$  blockade (BQ-123) during the Mid-Luteal Phase (left) and Early Follicular Phase (right) of the menstrual cycle. Data are presented as means  $\pm$  SEM. \*  $P < 0.05$  vs control. †  $P < 0.05$  vs. Early Follicular Phase.

**Table 1:**

## Demographic and Screening Characteristics

Variable	n=12
<b><i>Demographic Information</i></b>	
Age (years)	22 ± 1
Height (cm)	165 ± 2
Weight (kg)	62 ± 3
BMI (kg/m <sup>2</sup> )	23 ± 1
<b><i>Hemodynamic Measurements</i></b>	
Heart rate (beats/min)	61 ± 4
Systolic BP (mmHg)	111 ± 3
Diastolic BP (mmHg)	72 ± 2
MAP (mmHg)	85 ± 2
<b><i>Blood Chemistry</i></b>	
Total cholesterol (mg/dl)	154 ± 6
High-density lipoprotein (mg/dl)	62 ± 4
Low-density lipoprotein (mg/dl)	76 ± 7
Triglycerides (mg/dl)	76 ± 14
Hemoglobin (mg/dl)	12.9 ± 0.2
Hematocrit (%)	39.8 ± 0.6
Glucose (mg/dl)	74 ± 2

Body mass index (BMI), blood pressure (BP), mean arterial pressure (MAP).

Data are presented as mean ± SEM.



**Table 2:**

Subject Characteristics during Experimental Visits.

Variable	Early Follicular Phase	Mid-Luteal Phase
Weight (kg)	62 ± 3	62 ± 3
Systolic BP (mmHg)	105 ± 3	101 ± 2
Diastolic BP (mmHg)	65 ± 2	61 ± 1
MAP (mmHg)	78 ± 2	74 ± 2
Day of Cycle Tested	4 ± 1	23 ± 1 *
Plasma ET-1, pg/mL	1.3 ± 0.1	1.4 ± 0.1
Estradiol, serum (pg/ml)	44 ± 9	118 ± 15 *
Progesterone, serum (ng/ml)	0.7 ± 0.2	8.3 ± 0.8 *

Blood pressure (BP), endothelin-1 (ET-1), mean arterial pressure (MAP).

Data are presented as mean ± SEM.

\*  $P < 0.05$  vs. Early Follicular Phase.