



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



## Full Length Article

## Laboratory biomarkers associated with COVID-19 severity and management

S. Keddie<sup>a,1,\*</sup>, O. Ziff<sup>a,1</sup>, M.K.L. Chou<sup>a</sup>, R.L. Taylor<sup>a</sup>, A. Heslegrave<sup>b</sup>, E. Garr<sup>a</sup>, N. Lakdawala<sup>a</sup>, A. Church<sup>a</sup>, D. Ludwig<sup>c</sup>, J. Manson<sup>c</sup>, M. Scully<sup>d</sup>, E. Nastouli<sup>e</sup>, M.D. Chapman<sup>a</sup>, M. Hart<sup>a</sup>, M. P. Lunn<sup>a</sup>

<sup>a</sup> Neuroimmunology and CSF laboratory, University College London Hospitals NHS Trust National Hospital of Neurology and Neurosurgery, Queen Square, London, UK

<sup>b</sup> UK Dementia Research Institute, University College London, London, UK

<sup>c</sup> Department of Rheumatology, University College London Hospitals NHS Trust, London, UK

<sup>d</sup> Department of Haematology, University College London Hospitals NHS Foundation Trust and Cardiometabolic Programme-NIHR UCLH/UC BRC, London, UK

<sup>e</sup> Infection control department, University College London Hospitals NHS Trust, London, UK



## ARTICLE INFO

## Keywords:

COVID-19  
Cytokines  
Biomarkers  
Intensive care  
IL-6  
CRP  
IL-10  
LDH

## ABSTRACT

The heterogeneous disease course of COVID-19 is unpredictable, ranging from mild self-limiting symptoms to cytokine storms, acute respiratory distress syndrome (ARDS), multi-organ failure and death. Identification of high-risk cases will enable appropriate intervention and escalation. This study investigates the routine laboratory tests and cytokines implicated in COVID-19 for their potential application as biomarkers of disease severity, respiratory failure and need of higher-level care.

From analysis of 203 samples, CRP, IL-6, IL-10 and LDH were most strongly correlated with the WHO ordinal scale of illness severity, the fraction of inspired oxygen delivery, radiological evidence of ARDS and level of respiratory support ( $p \leq 0.001$ ). IL-6 levels of  $\geq 3.27$  pg/ml provide a sensitivity of 0.87 and specificity of 0.64 for a requirement of ventilation, and a CRP of  $\geq 37$  mg/l of 0.91 and 0.66.

Reliable stratification of high-risk cases has significant implications on patient triage, resource management and potentially the initiation of novel therapies in severe patients.

### 1. Introduction

International efforts to ‘flatten the curve’ of Coronavirus Disease-2019 (COVID-19) through social isolation restrictions have allowed most health systems to cope with huge demands on healthcare facilities, in particular providing time to acquire ventilators and intensive care beds. As governments loosen lockdown processes and with the unknown influences of seasonal variation and acquired immunity, front line services need to prepare for a second wave of infection. Lessons learnt regarding COVID-19 disease biology, transmission, risk factors, complications and treatments will be used to adapt and improve clinical services and improve outcomes.

The heterogeneous disease course of COVID-19 is unpredictable with most patients experiencing mild self-limiting symptoms. However up to 30% require hospitalisation, and up to 17% of these require intensive care support for acute respiratory distress syndrome (ARDS),

hyperinflammation and multiorgan failure [1–3]. A cytokine storm in patients with severe disease was identified in the early reports of Wuhan patients and is intrinsic to disease pathology. In this cohort, elevated plasma interleukin (IL)-2, IL-7, IL-10, Granulocyte colony stimulating factor (GCSF), interferon  $\gamma$ -induced protein 10 (IP10), monocyte chemoattractant protein-1 (MCP1), macrophage inflammatory protein 1-alpha (MIP1A) and tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ) levels in ICU patients were identified [4]. Subsequent studies have implicated IL-6 as a valuable predictor of adverse clinical outcome and a potential therapeutic target [5,6]. One or more clinical and wet biomarkers may enable early identification of high-risk cases, assisting disease stratification and effective use of limited specialist resources.

This study comprehensively explored clinical disease features and routine laboratory tests against specialist cytokine biomarkers associated with COVID-19 disease and its complications, to address their association with disease severity, respiratory intervention and outcome.

\* Corresponding author.

E-mail address: [stephen.keddie@nhs.net](mailto:stephen.keddie@nhs.net) (S. Keddie).

<sup>1</sup> Authors contributed equally.

## 2. Methods

One hundred consecutive hospital in-patients with COVID-19 infection, whose sera were sent for cytokine testing, were investigated as part of standard care and routine practice at University College London Hospital NHS Trust. All cases were confirmed as COVID-19 by polymerase chain reaction (PCR) analysis (86%) or deemed highly likely based on internationally recognised diagnostic criteria including recent exposure, clinical, radiological and laboratory features [7]. Laboratory biomarkers including cytokines were collated with baseline demographics, risk factors, disease features, treatments and outcomes. Level of clinical care (ward based, high-dependency or intensive treatment), oxygen requirements and vital signs were associated with the biomarkers at the time of sample collection, at peak illness and on most recent assessment. Serial samples were analysed to establish whether biomarkers correlated with, or could predict disease course. Severity of COVID-19 illness was determined utilising the World Health Organisation (WHO) COVID-19 ordinal severity scale which broadly measures level of respiratory compromise and requirement of intervention, with a score of 1 defined by no limitation of activities, to 7 requiring ventilation

and additional organ support [8]. Other features of organ system compromise such as neurological involvement, renal failure and requirement for dialysis were analysed for association with laboratory biomarkers.

Bloods for cytokine analysis were centrifuged within 4 h of collection, separated and sera frozen at -80 °C for up to 24 h before being analysed. Immunoassays were performed according to manufacturer's instructions and reads were fully automated. Routine laboratory data from automated analysers including C-reactive protein (CRP), D-dimer, lactate dehydrogenase (LDH), lymphocyte count, ferritin, fibrinogen and platelets were collated with the clinical and cytokine data.

A four-plex cytokine enzyme linked immunosorbent assay (ELISA)-based electrochemiluminescent immunoassay (Meso Scale Discovery, Rockville, Maryland) was introduced on the 17th April 2020 for clinical use in COVID-19 cases. The immunoassay measures IL-6, IL-1 $\beta$ , IL-10 and TNF- $\alpha$  to levels  $\leq 1$  pg/ml, with the choice of the four analytes determined by preliminary published reports, known disease pathogenesis and treatment considerations. The acquired data were part of assay validation and verification to provide normal and disease ranges and clinical interpretation.

**Table 1**

Demographic characteristics, features of COVID-19 infection and biomarkers stratified against the WHO COVID-19 ordinal severity scale.

	All patients	WHO 3 Hospitalised	WHO 4 & 5 Oxygen $\pm$ NIV	WHO 6 & 7 Ventilated	P-value
Number of patients	100	24	23	53	
<b>Demographics</b>					
Age (years)	59 (20–92)	59 (20–90)	64 (24–92)	57 (33–71)	$\leq 0.0001$
Male	74 (75%)	17 (71%)	16 (70%)	41 (77%)	0.715
<b>Comorbidities</b>					
Diabetes	25	9	1	14	$\leq 0.005$
Hypertension	37	9	8	19	<b>0.005</b>
Cardiovascular disease	12	4	5	2	0.53
Cerebrovascular disease	8	4	3	0	0.16
Malignancy	10	3	3	2	0.88
Current smoker	13	4	3	6	0.632
<b>Clinical features</b>					
Fever on admission	84	19	18	47	$\leq 0.0001$
Rigors	21	2	7	12	<b>0.03</b>
Cough	77	17	17	43	$\leq 0.0001$
Breathlessness	73	10	20	43	$\leq 0.0001$
Sputum	15	1	7	7	0.091
Fatigue	32	5	7	20	$\leq 0.01$
Myalgia	16	5	2	9	0.099
Anosmia	2	1	1	0	0.61
Sore throat	10	3	1	6	0.15
Headache	12	6	2	4	0.37
Diarrhoea	15	5	1	9	$\leq 0.05$
Nausea	15	4	2	9	0.074
Vomiting	9	3	0	6	$\leq 0.050$
Normal chest x-ray	18	16	2	0	$\leq 0.0001$
Focal consolidation	8	4	4	0	0.13
Bilateral consolidation	74	4	17	53	$\leq 0.0001$
<b>Treatment</b>					
Dialysis	17	0	1	16	$\leq 0.0001$
Antibiotics	75	7	21	47	$\leq 0.0001$
Antivirals	7	1	2	4	0.37
Steroids	28	3	4	21	$\leq 0.0001$
<b>Biomarkers</b>					
IL-1 $\beta$ pg/ml	0.4	0.7	0.2	0.4	$\leq 0.05$
IL-6 pg/ml	50.8	5.9	16.1	85.4	$\leq 0.0001$
TNF $\alpha$ pg/ml	5.2	4.1	4.4	6.1	0.10
IL-10 pg/ml	3.4	1.5	3.2	4.4	$\leq 0.0001$
Lymphocyte $\times 10^9/l$	1.4	1.4	1.4	1.4	0.22
CRP mg/l	102.7	49.4	73.3	135.7	$\leq 0.0001$
D-dimer ug/l	7131	4007	6490	8732	$\leq 0.0001$
Ferritin ug/l	1470	945	2129	1436	$\leq 0.001$
LDH IU/l	406.5	311.0	378.4	451.7	$\leq 0.001$
Fibrinogen g/l	17.6	4.2	5.0	26.0	$\leq 0.001$
Platelets $\times 10^9/l$	295.4	274.9	274.5	314.0	0.24

Bold indicates the significance of P value  $\leq 0.05$ .

## 2.1. Statistical analysis

Chi-square was used for the comparison of categorical variables. Biomarker levels comparing dichotomous variables were evaluated using Welch's unpaired *t*-test. Comparison of multiple group outcomes was performed using one-way ANOVA. Biomarker and continuous clinical variables were correlated using Pearson correlation coefficient. The predictive values of biomarkers were calculated by identifying Youden's index on receiver operator characteristic curves. *P*-value  $\leq 0.05$  was considered statistically significant. All statistical analyses were made using R statistical package 3.6.2.

## 3. Results

Samples from 100 patients were collected from the 6th April to the 18th May from patients hospitalised at University College London Hospital (UCLH) sites, 78 from the main hospital and 22 from the National Hospital for Neurology and Neurosurgery (including neuro-intensive care). Eighty-six patients were COVID-19 PCR positive and 14 deemed highly likely based on diagnostic criteria [7]. Seventy-four cases were male, with a median age of 59 years (20–92). One hundred and forty samples were collected from ward environments versus 63 on intensive care. The WHO COVID severity score ranged from 3 (hospitalised, no oxygen therapy) to 7 (severe disease, ventilation and additional organ support) with a median of 6 (severe disease, intubated and ventilated). Background characteristics associated with more severe infection and intervention included older age ( $p \leq 0.001$ ), diabetes ( $p \leq 0.05$ ) and hypertension ( $p \leq 0.05$ ). Clinical features and grouped biomarkers are displayed in Table 1.

IL-1 $\beta$  was the only biomarker which significantly differentiated by gender, with higher levels in males ( $p \leq 0.01$ ) who have a higher mortality [9]. Biomarkers of COVID-19 severity did not differ by number of comorbidities or smoking status.

All the biomarkers in Fig. 1 were analysed for their correlations with WHO COVID-19 severity scores and arranged into a heat map (See Fig. 1). Biomarkers that significantly correlated with the WHO severity

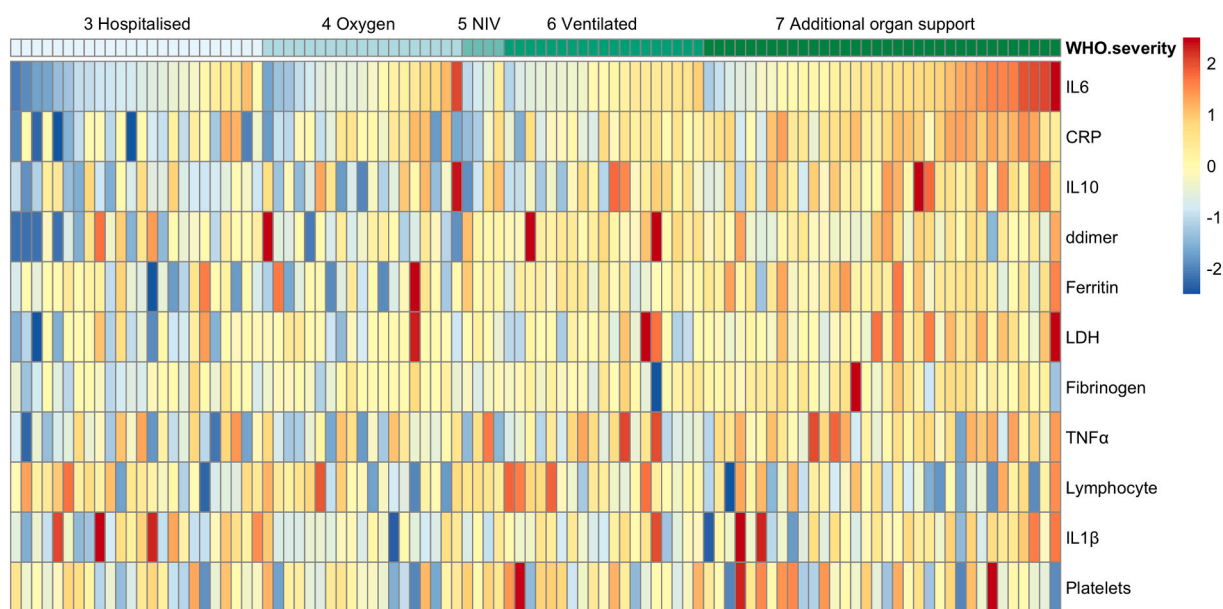
score included CRP ( $R = 0.53, p = 2.6e^{-12}$ ), IL-6 ( $R = 0.49, p = 1.3e^{-13}$ ), LDH ( $R = 0.38, p < 2.1e^{-5}$ ) and IL-10 ( $R = 0.35, p = 2.5e^{-7}$ ). IL-6 and CRP were most significantly indicative of the level of respiratory support (Fig. 2a and b) with IL-6 being marginally better at differentiating respiratory requirement from none. IL-6 and CRP were both closely associated with increased fraction of inspired oxygen delivery (FiO<sub>2</sub>) requirements ( $R = 0.54$  and  $0.58$  respectively) and radiological evidence of acute respiratory distress syndrome (ARDS) (IL-6  $\leq 0.00001$ , CRP  $\leq 0.005$ ). A cut-off of 3.27 pg/ml for IL-6 in this assay gives a sensitivity 0.87 and specificity 0.64 for requirement of intubation. Similarly, a 37 mg/l cut-off for CRP has a sensitivity of 0.91 and specificity of 0.66. TNF- $\alpha$  also correlated with WHO severity ( $p \leq 0.05$ ) and level of respiratory support ( $p \leq 0.01$ ) but to a lesser degree than IL-6 and CRP.

Renal failure leading to a requirement for renal dialysis was determined by higher levels of TNF- $\alpha$  (mean 5.87 vs 4.7 pg/ml,  $p < 0.01$ ), CRP (162 vs 82 mg/l,  $p < 0.01$ ), IL-6 (1.1 vs 0.6 pg/ml,  $p < 0.01$ ) and IL-10 (3.9 vs 2.44 pg/ml,  $p < 0.05$ ). IL-1 $\beta$  did not correlate with disease severity measures, presence of ARDS or level of respiratory support.

Neurological presentations with COVID-19 were also common, with cases presenting with delirium (12), ischemic stroke (5), intracerebral haemorrhage (2), seizures (3), encephalopathy (3), encephalitis (3), and acute disseminated encephalomyelitis (ADEM) in 2. Neurological presentations were inversely associated with the other disease markers of severity with IL-6, TNF $\alpha$ , IL-10, CRP, d-dimer and LDH which were all significantly lower in patients with neurological diseases.

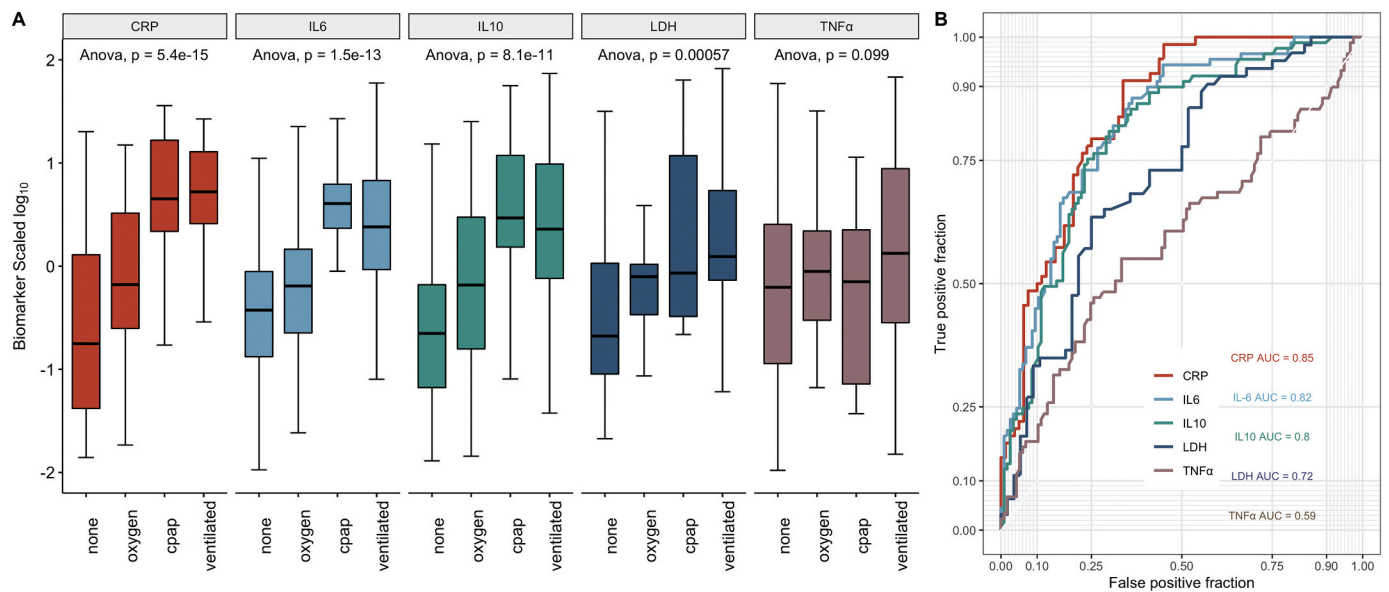
The strongest correlation between two biomarkers was unsurprisingly CRP and IL-6 ( $R = 0.65, p \leq 0.00001$ ) as CRP production is driven directly by IL-6, however all 4 cytokines analysed correlated significantly with one another (data not shown). D-dimer, ferritin and lymphocytes (inversely) correlated with all cytokine levels, indicative of severe COVID-19 resulting in a frank proinflammatory cytokine storm.

Longitudinal serial serum samples were also collected in 56 cases. Although not statistically significant, IL-6 and CRP levels tended to be lower in patients located in ward environments compared to those in intensive therapy unit (ITU) where levels remained persistently high, and increased in those who subsequently died (data not shown). Serial



**Fig. 1.** Heatmap of biochemical markers and their association with COVID-19 illness severity.

Heatmap shows biochemical signatures for each patient, arranged by WHO COVID-19 severity score. Visualisation was performed using the *heatmap* R package. Biomarkers values are log<sub>10</sub> transformed, centred and scaled. No patients had WHO severity scores 0, 1 or 2 as these are non-hospitalised patients. A score of 3 is hospitalised patients requiring no oxygen therapy; a score of 4 requires oxygen therapy; 5 requires non-invasive ventilation (NIV); of 6 requires intubation and mechanical ventilation; and 7 requires ventilation and additional organ support including vasopressors, renal replacement therapy and ECMO. A score of 8 is death. Higher levels of CRP, IL-6, IL-10 LDH and TNF- $\alpha$  are associated with higher WHO COVID-19 severity scores.



**Fig. 2.** CRP, IL-6, IL-10, LDH and TNF- $\alpha$  levels predict the requirement for mechanical ventilation and intensive care.

A) Split by four categories of respiratory support (none; supplemental oxygen; continuous positive airway pressure (CPAP); and mechanical ventilation). One way ANOVA shows these four categories are significantly different for CRP, IL-6, IL-10, LDH and TNF- $\alpha$ .

B) Receiver operator characteristic curves of CRP, IL-6, IL-10, LDH and TNF- $\alpha$  demonstrating the area under the curve (AUC) for predicting the requirement for intensive care.

levels of other biomarkers did not significantly correlate with clinical indicators of improvement i.e. as patients' oxygenation improved or were stepped down from ITU.

#### 4. Discussion

With 37.8 million COVID-19 confirmed worldwide cases and over 1 million deaths by the 13th October 2020 [10], understanding of COVID-19 disease pathogenesis, stratification and management has become a global priority. The identification of patients' ventilatory requirements has been paramount to healthcare structuring and resource management. Clinical assessment of COVID-19 cases remains key to decision making, but laboratory biomarkers enable finesse of this information assisting decision making and provide data of underlying biological processes.

This study has comprehensively analysed the range of COVID-19 biomarkers for their usefulness in COVID-19 disease stratification. We have demonstrated that IL-6, CRP, IL-10, LDH and TNF- $\alpha$  are indicative of different aspects of COVID-19 severity; oxygen requirement, presence of ARDS and requirements of intensive care support including dialysis and ventilation. IL-6 is a potent proinflammatory cytokine which stimulates CRP and has been identified as a driver of the COVID-19 cytokine storm associated with poor outcome [5,6,11–13]. Herold et al have recently found an IL-6 cut-off value of 80 pg/ml in their assay is highly indicative of respiratory failure, misclassifying only one patient not requiring intubation [5]. Such a clear definition has not been replicated in this study, which could be due to differences in assays or sample populations. Regardless, both studies demonstrate the significant utility of IL-6 as a key biomarker of disease in COVID-19. This supports the rationale behind the trial use of anti-IL-6 monoclonals as one of the therapies for COVID-19 [14,15]. Pre-selecting patients with the worst prognosis and most severe disease on the basis of highly elevated IL-6 may justify anti-interleukin immune therapy and avoid unnecessary complications by avoiding treatment in others.

IL-10 is a macrophage activated immunoregulatory cytokine which inhibits the expression of Th1 cytokines and enhances B cell stimulation. Elevated levels are therefore likely in response to the proinflammatory cytokine milieu, inherent to severe disease and reflect a functioning

immune system [16]. The binding of COVID-19 to the Toll Like Receptor (TLR) causes release of pro-IL-1 $\beta$  which is cleaved by caspase-1, followed by inflammasome activation and production of active IL-1 $\beta$  which is a mediator of lung inflammation, fever and fibrosis. IL-1 $\beta$  has been shown to be raised in COVID-19 infection compared to healthy controls by some [4]. This study however does not provide any indication that IL-1 $\beta$  levels reflect severity of disease [4,17]. Despite this, initial trials of the IL-1 $\beta$  antagonist Anakinra appear to reduce the need for mechanical ventilation and mortality in severe cases [18]. However no studies have studied the influence of the drug on pre- and post-treatment levels of IL-1 $\beta$ , or stratified entry to Anakinra on IL-1 $\beta$  levels. Our study would indicate that non-IL-1 $\beta$  biomarkers might be of more practical utility.

The association of TNF- $\alpha$ , CRP and IL-10 with dialysis requirement without any association with IL-6 possibly supports a role for sepsis driving renal failure. In post-mortem studies microthrombi are seen in both the kidney and alveolar blood vessels, but the primary insult on those two tissues may be different.

Neurological cases had lower levels of COVID-19 disease severity markers, which may be accounted for in part by selection bias, where patients were hospitalised due to neurological disease with concurrent, less severe COVID-19 infection compared to those admitted primarily due to COVID-19 respiratory failure. The primary targets for SARS-CoV-2 are respiratory epithelial cells and alveolar macrophages. Following infection of respiratory cells, rapid production and release of cytokines and chemokines occur. Macrophages are in turn activated, as are other key components of the innate immune system, such as dendritic cells, which leads to a more extensive immune response initiating the cytokine storm [19]. It is therefore conceivable from our findings that only severe pulmonary infection and infiltration results in cytokine activation and proliferation, and neurological features are either a consequence or coincidence of COVID-19 infection in this cohort. However more research is required to understand the neurological manifestations of COVID-19 and the associated cytokine response to more definitively explain such findings.

C-reactive protein is unsurprisingly very strongly correlated with levels of IL-6, as IL-6 drives the production of CRP primarily in the liver. CRP may potentially suffice in isolation to predict high risk cases requiring more aggressive intervention, negating the requirement of



more specialist cytokine tests. Similarly, LDH, released from multiple tissues on cell death and activated through T-cell proliferation may be a practical and easily measured surrogate biomarker although its utility here is lower than the CRP. Levels of ferritin were raised across the cohort likely reflecting heightened acute phase response to infection as opposed to supra-high levels indicative of sHLH. Absolute lymphocyte count and subsets have been shown to inversely correlate with severity in COVID-19 [16,20,21], although not demonstrated in this study. Lymphocyte counts may be a less reliable marker due to the variety of factors which cause lymphocytosis such as sepsis from concomitant infections and the stress response. Although select routine laboratory markers correlate with disease severity, cytokine analysis provides additional predictive support for prognosis and likely interventions in the COVID-19 cytokine storm as well as reassurance that when low, little escalation of intervention may be required. Note that a limitation of the study is that 16 cases were deemed not appropriate for intensive care, therefore escalation of respiratory support would be precluded regardless of clinical severity. We conducted a sensitivity analysis and inclusion/exclusion of these cases had no effect on biomarker correlations with severity.

Biomarker analysis of CRP, LDH and the cytokines IL-6, IL-10 and TNF $\alpha$ , alongside thorough clinical assessment of COVID-19 patients, enables more accurate stratification of high from low risk cases and the need for intensive care support. Such stratification enhances management not only for individual patients but has significant implications on resource management, hospital flow and staffing, and potentially the initiation of novel therapies in the most severe patients.

## Funding

Dr. Keddie is funded by the Association of British Neurologists and the Guarantors of Brain. Professor Mike Lunn is supported by the National Institute of Health Research, University College London Hospitals, Biomedical Research Centre.

## Declaration of Competing Interest

None.

## References

- [1] D. McGonagle, K. Sharif, A. O'Regan, C. Bridgewood, The role of cytokines including interleukin-6 in COVID-19 induced pneumonia and macrophage activation syndrome-like disease, *Autoimmun. Rev.* 19 (6) (2020) 102537.

- [2] ICNARC, ICNARC Report on COVID-19 in Critical Care. London, 2020.
- [3] European center for disease prevention and control, Rapid Risk Assessment - Coronavirus Disease 2019 COVID-19 in the EU/EEA and the UK – Eighth Update, 2020.
- [4] C. Huang, Y. Wang, X. Li, et al., Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China, *Lancet* 395 (10223) (2020) 497–506.
- [5] T. Herold, V. Jurinovic, C. Arnreich, et al., Elevated levels of IL-6 and CRP predict the need for mechanical ventilation in COVID-19, *J Allergy Clin Immunol* 146 (1) (Jul 2020) 128–136, <https://doi.org/10.1016/j.jaci.2020.05.008> (2020.04.01.20047381).
- [6] E.A. Coomes, H. Haghbayan, Interleukin-6 in COVID-19: a systematic review and meta-analysis, *Rev Med Virol.* 26 (e2141) (Aug 2020) (2020.03.30.20048058).
- [7] European center for disease prevention and control, Case Definition for Coronavirus Disease 2019 (COVID-19), as of 29 May 2020, 2020.
- [8] World Health Organisation, Blueprint Novel Coronavirus COVID-19 Therapeutic Trial Synopsis, 2020.
- [9] E.J. Williamson, A.J. Walker, K. Bhaskaran, et al., Factors associated with COVID-19-related death using OpenSAFELY, *Nature* 584 (7281) (Aug 2020) 430–436, <https://doi.org/10.1038/s41586-020-2521-4>.
- [10] World Health Organisation, Coronavirus Disease (COVID-19) Situation Reports, 2020.
- [11] F. Liu, L. Li, M. Xu, et al., Prognostic value of interleukin-6, C-reactive protein, and procalcitonin in patients with COVID-19, *J. Clin. Virol.* 127 (2020), 104370.
- [12] X. Chen, B. Zhao, Y. Qu, et al., Detectable serum SARS-CoV-2 viral load (RNAemia) is closely correlated with drastically elevated interleukin 6 (IL-6) level in critically ill COVID-19 patients, *Clin. Infect. Dis.* 17 (449) (Apr 2020), <https://doi.org/10.1093/cid/ciaa449>.
- [13] F.A. Lagunas-Rangel, V. Chávez-Valencia, High IL-6/IFN- $\gamma$  ratio could be associated with severe disease in COVID-19 patients, *J. Med. Virol.* 16 (10) (April 2020), <https://doi.org/10.1002/jmv.25900>.
- [14] B. Maes, C. Bosteels, E. De Leeuw, et al., Treatment of severely ill COVID-19 patients with anti-interleukin drugs (COV-AID): a structured summary of a study protocol for a randomised controlled trial, *Trials* 21 (1) (2020) 468.
- [15] J. Rilinger, W.V. Kern, D. Duerschmied, et al., A prospective, randomised, double blind placebo-controlled trial to evaluate the efficacy and safety of tocilizumab in patients with severe COVID-19 pneumonia (TOC-COVID): a structured summary of a study protocol for a randomised controlled trial, *Trials* 21 (1) (2020) 470.
- [16] S. Wan, Q. Yi, S. Fan, et al., Relationships among lymphocyte subsets, cytokines, and the pulmonary inflammation index in coronavirus (COVID-19) infected patients, *Br. J. Haematol.* 189 (3) (2020).
- [17] C. L. L. HG, L. W, et al., Analysis of clinical features of 29 patients with 2019 novel coronavirus pneumonia, *Chin. J. Tuberc Respir. Dis.* 43 (0) (2020).
- [18] T. Huet, H. Beaussier, O. Voisin, et al., Anakinra for severe forms of COVID-19: a cohort study, *Lancet Rheumatol.* 2 (7) (Jul 2020) e393–e400.
- [19] Y.H. Balcioglu, U.H. Yesilkaya, H. Gokcay, S.S. Kirlioglu, May the central nervous system be fogged by the cytokine storm in COVID-19? an appraisal, *J. NeuroImmune Pharmacol.* 15 (3) (2020) 343–344.
- [20] W. Huang, J. Berube, M. McNamara, et al., Lymphocyte subset counts in COVID-19 patients: a meta-analysis, *Cytom Part A* 97 (8) (2020) 772–776.
- [21] R. He, Z. Lu, L. Zhang, et al., The clinical course and its correlated immune status in COVID-19 pneumonia, *J. Clin. Virol.* 127 (2020), 104361.