

# REVIEW ARTICLE OPEN



# Insights into the use of mesenchymal stem cells in COVID-19 mediated acute respiratory failure

Nisha Durand (o¹), Jorge Mallea (o²) and Abba C. Zubair (o¹) ™

The emergence of severe acute respiratory syndrome corona virus 2 (SARS-CoV-2) at the end of 2019 in Hubei province China, is now the cause of a global pandemic present in over 150 countries. COVID-19 is a respiratory illness with most subjects presenting with fever, cough and shortness of breath. In a subset of patients, COVID-19 progresses to hypoxic respiratory failure and acute respiratory distress syndrome (ARDS), both of which are mediated by widespread inflammation and a dysregulated immune response. Mesenchymal stem cells (MSCs), multipotent stromal cells that mediate immunomodulation and regeneration, could be of potential benefit to a subset of COVID-19 subjects with acute respiratory failure. In this review, we discuss key features of the current COVID-19 outbreak, and the rationale for MSC-based therapy in this setting, as well as the limitations associated with this therapeutic approach.

npj Regenerative Medicine (2020)5:17; https://doi.org/10.1038/s41536-020-00105-z

#### **INTRODUCTION**

The severe acute respiratory syndrome (SARS) corona virus 2 (SARS-CoV-2) disease (COVID-19) emerged in December 2019 in Wuhan, Hubei Province, People's Republic of China when a cohort of patients with pneumonia of unknown etiology presented to local hospitals<sup>1</sup>. Since the initial outbreak of COVID-19 in Wuhan, the disease has since spread to over 150 countries with migratory epicenters that include Iran, Italy, Spain, and New York City, USA<sup>2</sup>. Given the widespread incidence of COVID-19, the World Health Organization (WHO) declared the current outbreak as a Public Health Emergency of International Concern (PHEIC) on January 30th 2020, and subsequently elevated the outbreak to pandemic status on March 11th 2020<sup>3</sup>. As of July 1, 2020, globally, there are about 10 million confirmed cases of COVID-19 and just over 500,000 fatalities have been reported<sup>4</sup>.

The causative agent of COVID-19, SARS-CoV-2, is an enveloped, positive-sense RNA coronavirus. Coronaviruses primarily cause respiratory and intestinal infections, and infect several species from bats to humans. Coronaviruses can have serious implications for public health: in addition to the current COVID-19 pandemic, they have also been responsible for SARS and Middle East respiratory syndrome, significant epidemics in the last two decades<sup>5</sup>. For the current outbreak of COVID-19, the initial patients who presented with the illness had all visited a local fish and wild animal market in Wuhan, Hubei Province, China which is indicative of a possible zoonotic transmission, but the exact mechanism which resulted in the first-in-person infection remains elusive. Propagation of the virus among humans is thought to occur primarily through three main transmission mechanisms: (I) contact transmission, (II) droplet transmission and (III) aerosol transmission<sup>6</sup>. The full genome sequence of SARS-CoV-2 has been elucidated, and it shares 79.6% sequence identity with SARS-CoV, and at the whole-genome level is 96% identical to a bat coronavirus. Entry into and subsequent infection of human alveolar epithelial cells by SARS-CoV-2 is mediated by the angiotensin-converting enzyme II (ACE2) receptor<sup>7</sup>.

For most patients with COVID-19 infections, initial symptoms include, but are not limited to fever, shortness of breath, cough and general malaise<sup>8,9</sup>. In the majority of patients, COVID-19 manifests as a mild-to-moderate disease and with conservative medical management, full recovery from the ailment is expected<sup>10,11</sup>. However, in a subset of high-risk patients; the elderly, and those with underlying medical conditions such as hypertension and diabetes, the disease can become critical, and is characterized by severe pneumonia, acute respiratory distress syndrome (ARDS), and multiple organ dysfunction<sup>9,12–14</sup>. Widespread inflammation, cytokine storm syndrome and a dysregulated immune response mediates the severe clinical manifestations seen in high-risk individuals with COVID-19<sup>15–17</sup>. In this cohort, disease- associated mortality is especially significant<sup>9,10</sup>.

There is no known cure for COVID-19, but given the prevalence of the disease, and the associated morbidity and mortality, various therapeutic modalities that might be effective against COVID-19 have been evaluated in clinical settings. Therapeutic agents assessed in COVID-19 patients thus far include, convalescent plasma<sup>18</sup>, antimalarials like Hydroxychloroquine<sup>19</sup> and antivirals<sup>20</sup>. Consideration has also been given to the use of immunoregulatory agents in subjects with severe COVID-19<sup>21-23</sup>, since dysregulated and hyper-activated inflammatory responses tends to be a main driver of COVID-19-induced mortality<sup>10</sup>.

One such immunoregulatory agent with potential for COVID-19 subjects with critical disease is the mesenchymal stem cell (MSC). MSCs are multipotent stromal cells that induce immunomodulation and regeneration, and undergo tri-lineage differentiation when stimulated in vitro<sup>24,25</sup>. Beyond their immunomodulatory and regenerative capacity, MSCs also exhibit antimicrobial activity<sup>26</sup> and an 'immune privileged' phenotype, which facilitates allogeneic transplantation in the absence of rejection<sup>27</sup>. MSCs have been isolated from multiple tissue types including bone marrow, adipose tissue and umbilical cord, and their functional effects are primarily mediated by paracrine signaling via secretion of cytokines and chemokines<sup>28–30</sup>.

<sup>&</sup>lt;sup>1</sup>Laboratory Medicine and Pathology and Center for Regenerative Medicine, Mayo Clinic, Jacksonville, FL 32224, USA. <sup>2</sup>Department of Medicine, Division of Allergy, Pulmonary and Sleep Medicine, Mayo Clinic, Jacksonville, FL 32224, USA. <sup>⊠</sup>email: zubair.abba@mayo.edu





Table 1. Phase and	d cell source for MS	C therapy for COVID-19 related	conditions (Clir	nicaltrials.gov).			
Clinical trial phase	Total number (%)	MSC source per phase					
		Umbilical cord/Wharton's jelly	Bone marrow	Adipose tissue	Dental pulp	Olfactory mucosa	Unknown
1	8 (20.5)	2	3	1	1	0	1
1/2	12 (30.8)	8	1	1	1	1	0
2	16 (41.0)	7	2	5	0	0	2
2/3	1 (2.6)	0	0	0	0	0	1
3	1 (2.6)	0	1	0	0	0	0
Unknown	1 (2.6)	1	0	0	0	0	0
Total # of MSC sou	urce (%)	18 (46.2)	7 (17.9)	7 (17.9)	2 (5.1)	1 (2.6)	4 (10.3)

In clinical settings, while the safety profile of MSCs has been well-established, often times, the efficacy data obtained in pre-clinical models have not been recapitulated in most cases when MSCs have been administered to human cohorts. MSC therapeutic effectiveness is likely limited to a niche of immunological disorders and immune-mediated illnesses such as graft-versus-host-disease, for which MSCs have demonstrated varying levels of efficacy in Phase 1/2 clinical studies<sup>31,32</sup>. In multiple pre-clinical studies, MSCs have been shown to be effective against ARDS and acute lung injury (ALI)<sup>33,34</sup>, both of which are severe clinical manifestations of COVID-19, and while, administration of MSCs to subjects with ARDS was well tolerated<sup>35,36</sup>, efficacy data at the clinical level is not as compelling. To date, there are about 40 clinical trials registered on *clinicaltrials.gov* for evaluating the use of MSCs in subjects with COVID-19 (Table 1)<sup>37</sup>.

In this review, we critically examine the mechanisms that render MSCs potentially beneficial for a subset of COVID-19 subjects with acute respiratory failure; the limitations associated with this therapeutic modality and the current status of COVID-19-relavant MSC clinical studies.

#### COVID-19

Pathogenesis of COVID-19 mediated acute respiratory failure and its clinical manifestation

COVID-19 subjects usually present with fever, cough, fatigue and myalgias. Some patients develop gastrointestinal symptoms like diarrhea and nausea. Shortness of breath is present in 80% of the patients with COVID-19 that require hospitalization<sup>38</sup>. For most subjects with a non-severe manifestation of the disease, a full recovery is expected. However, a subset of patients with COVID-19 develop serious complications in the course of their disease. These complications commonly require admission to the intensive care unit (ICU), and include ARDS, cardiomyopathy, acute kidney injury, acute liver injury and shock<sup>17,39,40</sup>. It is estimated that 5% of patients with COVID-19 will need admission to the ICU, with the majority of them requiring mechanical ventilation<sup>41</sup>. ARDS associated with COVID-19 usually appears 1–2 weeks after the onset of symptoms, and is associated with a higher mortality. The mortality from COVID-19 is currently estimated at about 5%<sup>4</sup>.

### Immune response to COVID-19 infection

A well-orchestrated innate immune response is the initial defense mechanism against viral pathogens, but under some circumstances hyper-inflammatory responses can occur during viral infections<sup>5</sup>. In line with this notion of hyper-inflammatory immune responses, the most dire complications that arise from COVID-19 such as ARDS and acute cardiac injury are associated with widespread inflammation. In a study of 1099 COVID-19 patients, most patients had elevated levels of C-reactive protein (CRP), an inflammatory biomarker<sup>39</sup>. When compared to patients who had recovered from COVID-19,

those who succumbed to the disease had much higher levels of CRP in their blood<sup>9</sup>. Key inflammatory chemokines and cytokines such as interleukin (IL)-6, IL-8, tumor necrosis factor alpha (TNF-α), monocyte chemoattractant protein-1 (MCP-1) and macrophage inflammatory protein 1 alpha (MIP-1α) are elevated in COVID-19 subjects, and this is typically more pronounced in patients with severe manifestations of the disease<sup>9,15,17,42</sup>. In addition to the changes in the levels of inflammatory markers, alterations in immune cell quantity were also observed in COVID-19 patients. Lymphocyte counts are decreased in COVID-19 subjects and this is more pronounced in the subset of patients with severe disease<sup>15</sup> On the other hand, in critically ill COVID-19 patients, elevated neutrophil counts have been reported 15,17. Increased quantities of pro-inflammatory chemokines and cytokines, in conjunction with accompanying shifts in immune cell numbers is indicative of a dysregulated immune response in COVID-19 subjects, especially for those with critical illness. This dysregulated and hyper-activated immune response has been an area of focus for targeting COVID-19.

#### Therapeutic interventions

Treatments used to combat COVID-19 thus far include intravenous (IV) antibiotics, systemic glucocorticoids, IV immunoglobulins, convalescent plasma and antiviral therapy 9,39,43. The antimalarial drug chloroguine has also received much attention in the fight against COVID-19 and has already been evaluated in clinical trials<sup>1</sup> Both chloroquine and the antiviral drug Remdesivir are effective in controlling SARS-CoV-2 infection in vitro<sup>44</sup>. However, the utility of agents like hydroxychloroquine for COVID-19 remains inconclusive as there have been reports of viral clearance associated with use of the drug<sup>19</sup> but also data from multiple reports which indicate otherwise<sup>45,46</sup>. While drugs like chloroquine<sup>44</sup> and immunoglobulins<sup>47</sup> target the viral life cycle, a subset of drugs (glucocorticoids) used in COVID-19 subjects target inflammation. This is particularly important since ARDS, the most life-threatening complication associated with COVID-19, is primarily an inflammatory based condition. Targeting this aspect of the disease is crucial, and therapeutic agents like MSCs, which can counteract aggravated immune responses, should be considered in efforts geared towards the mitigation of COVID-19, particularly in those subjects with critical disease. Beyond immunomodulation, the regenerative potential and antimicrobial properties of MSCs would also be of practical importance in the setting of COVID-19.

# RATIONALE FOR MSC THERAPY TO MANAGE COVID-19 MEDIATED ACUTE RESPIRATORY FAILURE: EVIDENCE FROM PRE-CLINICAL STUDIES

Immunomodulation

Seminal studies by Meisel et al.<sup>48</sup> defined an anti-inflammatory role for MSCs when they showed that MSCs derived from bone

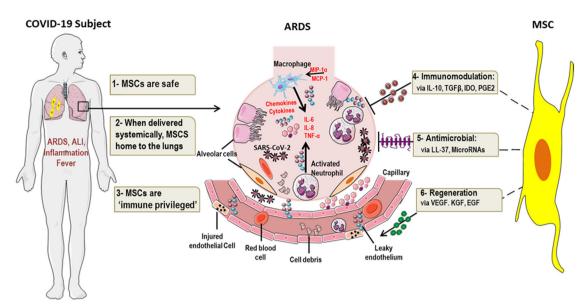


Fig. 1 Rationale for MSC therapy in COVID-19 subjects with critical illness. Subjects with critical COVID-19 can develop serious inflammatory-mediated complications like ARDS. Immune cells like macrophages and neutrophils are activated, pro- inflammatory cytokines (IL-6, TNF- $\alpha$ ) are released, and endothelial cells are injured in the lungs of these subjects. In this setting, MSCs could be of potential benefit since they are (I) Safe, (II) home to the lungs, (III) 'immune privileged', (IV) immunomodulatory, (V) antimicrobial and (VI) regenerative. Created using elements of Servier Medical Art by Servier, licensed under CC BY 3.0 (https://smart.servier.com/).

marrow could inhibit T cell responses in a manner which was dependent on indoleamine 2, 3-dioxygenase. Since then, countless other studies have demonstrated the anti-inflammatory properties of MSCs. MSCs secrete anti-inflammatory chemokines and cytokines such as IL-10, transforming growth factor beta and Prostaglandin E2 (PGE2) (Fig. 1)<sup>49–51</sup>. Transcriptionally, MSCs regulate inflammation via *Heme Oxygenase-1 (HO-1)*<sup>52</sup> and Inducible T Cell Costimulator Ligand (ICOSL)<sup>53</sup>. In COVID-19relevant animal models, administration of MSCs have resulted in amelioration of disease symptoms. In liposaccharide (LPS)-induced ARDS in mice, treatment with MSCs resulted in attenuation of neutrophil infiltration, collagen deposition and fibrosis<sup>54</sup>. In a similar model, administration of MSCs protected mice from LPSinduced ALI by inhibiting macrophages in a manner that was dependent on the secretion of PGE2, granulocyte-macrophage colony-stimulating factor (GM-CSF) and IL-13<sup>51</sup>. In addition to secreting anti-inflammatory molecules themselves, MSCs can also antagonize the release of certain pro-inflammatory cytokines. This was demonstrated by Xu et al.; in a rodent model of oleic acidinduced ALI, IV transplantation of MSCs reduced the concentration of pro-inflammatory TNF-α in blood plasma and lung tissue extracts. Conversely, a concomitant increase in the level of antiinflammatory IL-10 in these animals following MSC treatment was observed<sup>55</sup>. MSCs also govern the specific types of immune cells that are recruited to sites of injury; biased towards the infiltration of tolerogenic cell types like regulatory T cells. In a recent study by Wang et al.<sup>56</sup>, an improvement in LPS-induced ALI mediated by MSCs was dependent on the regulation of the balance between regulatory T cells and Th17 cells. Increased regulatory T cells were also observed in a mouse model of lung contusion following IV MSC administration<sup>57</sup>. The effectiveness of MSCs in mediating anti-inflammatory responses in the pulmonary space has also been recapitulated in large animal models. The severity of ALI as shown by improved oxygenation, was reduced in a sheep model of bacterial pneumonia following IV delivery of MSCs<sup>58</sup>.

COVID-19 infection has been shown to trigger a proinflammatory state. Huang et al. reported that initial plasma analysis of 41 patients infected with the novel coronavirus in Wuhan, China, shows that IL-1 beta, IL-1 receptor antagonist, IL-7, IL-8, IL-9, IL-10, basic fibroblast growth factor (FGF), granulocyte-colony stimulating factor (G-CSF), GM-CSF, interferon gamma (IFN-γ), interferon-inducible protein 10 (IP10), MCP-1, MIP-1 $\alpha$ , MIP-1 $\beta$ , platelet-derived growth factor (PDGF), TNF- $\alpha$ , and vascular endothelial growth factor (VEGF) were elevated in hospitalized patients when compared with healthy controls. Levels of IL-2, IL-7, IL-10, G-CSF, IP10, MCP-1, MIP-1 $\alpha$ , and TNF- $\alpha$  were higher in patients requiring ICU care<sup>59</sup>. Ruan et al. reported higher levels of CRP and IL-6 in the patients that died with COVID-19 when compared to the patients that were discharged from the hospital. They concluded that the COVID-19 associated mortality may likely be due to a virus-activated "cytokine storm syndrome"<sup>60</sup>.

Though it is well-established that MSCs attenuate inflammatory responses, we and others have shown that MSCs secrete various inflammatory cytokines, particularly IL-6. We have actually shown that IL-6 plays a critical role in MSC functioning<sup>61</sup>. IL-6 function is dependent on how its signal is transmitted to target cells. IL-6 acts via its classical signaling pathway by binding to its membrane bound receptor (IL-6R), which then dimerizes with glycoprotein 130 (gp130). Activation of the classical IL-6 signaling was shown to induce anti-inflammation and epithelial cell regeneration. In addition, IL-6 can act via its soluble receptor (sIL-6R) coupled with gp130 protein to act on non-IL-6R expressing target cells. This is referred to as the trans-signaling pathway. It has been suggested that IL-6 trans-signaling is pro-inflammatory<sup>62</sup>.

### Regeneration and repair

While the immunomodulatory effects of MSCs are critical for counteracting the aggravated immune response, repair to damage and injured cells and tissues would also be required in the setting of ARDS. MSCs secrete multiple factors implicated in regeneration and modulation of tissue injury (Fig. 1); epidermal growth factor (EGF), PDGF, FGF, hepatocyte growth factor (HGF), VEGF and insulin-like growth factor<sup>29,63–65</sup>. Inhibition of key regenerative mediators in MSCs abrogated their ability to attenuate lung injury in vivo, as demonstrated by Yang et al. who used a sophisticated series of experiments to make this determination. They showed that while MSC transplantation in



mice reduced lung permeability, protected lung epithelium from apoptosis, facilitated vascular endothelium (VE)-cadherin recovery and reduced lung injury, these beneficial effects were significantly diminished when VEGF expression was inhibited in the transplanted MSCs<sup>66</sup>. MSCs can also induce the expression of regenerative mediators in the setting of ALI. In a similar study, reduced HGF expression in MSCs impaired their ability to protect against lung fibrosis in vivo, in a mouse model of Bleomycininduced pulmonary fibrosis. Furthermore, it was shown that inhibition of pulmonary cell apoptosis mediated by MSC conditioned medium (CM) was HGF dependent<sup>67</sup>. In addition to secreting these regenerative molecules themselves, MSCs can also induce the expression of these factors in specific cells and tissues. IV administration of MSCs in a mouse model of LPSinduced ALI resulted in upregulated expression of keratinocyte growth factor<sup>68</sup>, an important mediator of alveolar repair<sup>69</sup>. In a rodent model of radiation-induced pulmonary fibrosis, administration of MSCs preserved the architecture of irradiated lungs, and this was partly mediated by endogenous expression of HGF<sup>7</sup> Repair of the pulmonary blood-air barrier in a rodent model of phosgene-induced lung injury has also been shown to be mediated by MSCs<sup>71</sup>.

#### Antimicrobial properties

There is a growing body of evidence which implicates an antimicrobial role for MSCs (Fig. 1); a characteristic that would be applicable for combating COVID-19. Multiple of the studies which have evaluated the nature of MSC antimicrobial activity have revealed a role for antimicrobial peptides (AMPs) in the process. AMPs are a diverse class of low molecular weight proteins produced as a first line of defense against pathogens such as bacteria, fungi and viruses<sup>72</sup>. In revolutionary work by Krasnodembskaya et al. it was shown that the antibacterial effect of MSCs was partly mediated by LL-37, an AMP. Using a series of welldesigned experiments, they showed that in vitro, MSCs as well as MSC-CM inhibited the growth of both Gram-negative and Grampositive bacteria. In addition, in a mouse model of E. coli-induced pneumonia, administration of MSCs reduced bacterial growth in bronchoalveolar lavage (BAL) fluid and lung homogenate, both of which were attenuated when a neutralizing antibody to LL-37 was introduced<sup>26</sup>. In a similar study of E. coli-induced pneumonia, MSC therapy decreased lung bacterial burden and this was accompanied by elevated alveolar concentrations of LL-37<sup>73</sup>. MSCinduced bacterial clearance mediated by LL-37 has also been demonstrated in mouse models of cystic fibrosis<sup>74</sup>. In a rodent model of E. coli-induced ARDS, MSC administration was associated with reduced formation of *E. coli* colony forming units in BAL<sup>75</sup>. Beta-defensin 2 (BD2), is another AMP secreted by MSCs; in a mouse model of E. coli-induced ALI, MSC administration resulted in enhanced bacterial clearance in a manner consistent with augmented BD2 secretion<sup>76</sup>. In contrast to the multiple studies focusing on the antibacterial activity of MSCs, reports of specific antiviral activity as it relates to MSCs have been less forthcoming, nevertheless, MSC-derived exosomal microRNAs have been shown to inhibit Hepatitis C virus infection in vitro<sup>77</sup>. MSCs also inhibit inflammasome activation in the presence of Coxsackievirus B3<sup>78</sup>. Finally, while studies of influenza virus-induced ALI in mice have demonstrated that MSC administration reduced lung injury and pulmonary inflammation, and restored alveolar fluid clearance<sup>79–81</sup> the actual ability of MSCs to inhibit viral replication in relevant animal models has not been demonstrated.

Paradoxically, while multiple studies have shown a definite antimicrobial role for MSCs, there are reports which indicate otherwise. In a study published by Schwartz et al., they demonstrated that MSC treatment in mice infected with *Mycobacterium bovis* promoted bacterial growth in the spleen. Suppression of bacterial growth was only achieved after the MSCs

were preconditioned with TLR-3 ligand<sup>82</sup>. This data is telling, but not entirely unexpected, as MSCs are actively involved in immunomodulation. This however raises important questions relating to MSC function in inflammatory-mediated viral illnesses such as COVID-19; would the abatement of the cytokine storm syndrome be achieved at the cost of increased viral load? The answers to these critical questions remain unknown.

#### First pass entrapment

As described above, MSCs exhibit several characteristics in vitro and in vivo, which if mirrored in human cohorts could be of potential therapeutic benefit for a subset of patients with severe manifestations of COVID-19. However, with all drug therapeutics, the mode of drug delivery and targeting strategy needs to be given serious consideration. Dependent on the indication, MSCs have been delivered via multiple routes of administration, from direct injection to IV delivery83. In the context of COVID-19associated ARDS, MSCs need to exert their therapeutic effects in the lungs. In pioneering studies by Fisher et al.<sup>84</sup> it was shown that when MSCs are delivered intravenously, the majority of cells remained trapped in the lungs, with limited quantities reaching other major organs like the heart, kidneys, and liver. This pulmonary first-pass effect has been investigated extensively; in a rodent model of silicosis fibrosis, when fluorescently labeled MSCs were delivered by IV, fluorescence intensity in the lungs peaked 6 h post injection; and 15 days post injection, fluorescence signals could still be detected in the lungs, albeit at much lower levels<sup>85</sup>. While this pulmonary entrapment of MSCs may be a barrier to certain types of MSC-based therapies, in the context of COVID-19-related ARDS, it is ideal (Fig. 1).

# Outcome of clinical studies using MSCs

Most of the data demonstrating the beneficial effects mediated by MSCs have been obtained from studies conducted in vitro and in vivo; however, in clinical settings the therapeutic efficacy of MSC-based therapy has not been readily demonstrated. Although multiple MSC-based clinical studies have been completed in the pulmonary space for conditions like chronic obstructive pulmonary disease (COPD), ARDS, emphysema and obstructive chronic lung allograft dysfunction 36,86-90, these studies are primarily Phase 1 trials which evaluate safety and feasibility. Data from these earlystage clinical trials demonstrate that administration of MSCs was safe and well-tolerated with no serious adverse events being reported. Nonetheless, in some of these early stage-clinical studies, positive indicators of MSC therapy have been reported. In subjects with ARDS, when MSCs were administered, 5 days post infusion, serum levels of surfactant protein D, an ALI biomarker, were significantly lowered as compared to baseline levels preinfusion. However, in this same study, no significant differences between the treatment and control groups in the PaO<sub>2</sub>/FiO<sub>2</sub> ratio were observed<sup>91</sup>. Administration of MSCs to subjects with severe emphysema resulted in increased CD31 expression, and CD3<sup>+</sup> and CD4<sup>+</sup> T cells, but the significance of these changes in the context of emphysema pathogenesis remains elusive. More importantly, in this study, no macroscopic or molecular evidence for repair of emphysematous lesions were detected following MSC infusion<sup>88</sup>. In a comprehensive study of two patients with severe ARDS for which MSCs were administered on a compassionate basis, several beneficial outcomes were reported: respiratory, hemodynamic and multi-organ failure were resolved following MSC treatment. Moreover, there were reduced levels of systemic and pulmonary markers of inflammation including epithelial apoptosis, alveolarcapillary fluid leakage, and pro-inflammatory chemokines, micro-RNAs and cytokines<sup>92</sup>. While the aforementioned studies are primarily Phase 1 studies which were not necessarily designed to evaluate efficacy, the efficacy data from these studies is weak.



Delivery method	Total number (%)	Number of	doses to be adn	ninistered			
		1	2	3	4	5	Unknown
Intramuscular injection	1 (2.6)	1*	0	0	0	0	0
Intravenous	36 (92.3)	5	6	10	5	2	8
Unknown	2 (5.1)	1	1	0	0	0	0
Total # of doses (%)		7 (17.9)	7 (17.9)	10 (25.6)	5 (12.8)	2 (5.1)	8 (20.5)

Convincing efficacy data from larger scale clinical studies for lung-related illnesses using MSCs are limited. In a randomized double-blinded study of 62 COPD subjects, systemic delivery of MSCs resulted in a significant decrease in levels of circulating CRP in patients who had elevated CRP levels at initiation of the study<sup>90</sup>. As noted above, COVID-19 subjects with critical illness have elevated CRP levels in comparison to their counterparts with a non-severe form of the disease<sup>9,39</sup>. While modulation of inflammatory markers in response to MSC treatment might be a beneficial outcome in the setting of systemic inflammation, MSC treatment did not alter important clinical parameters such as COPD exacerbations, pulmonary function tests (PFT) or quality of life indicators. In a larger scale, Phase 2, multi-centered clinical trial for patients with moderate to severe ARDS, while post-hoc analyses revealed a trend towards improvement in oxygenation index as well as a reduction in endothelial injury as determined by reduced plasma concentrations of angiopoietin-2, in the MSCtreatment group compared to the control group, more relevant clinical outcomes such as a ventilator-free days did not significantly vary between the two groups<sup>35</sup>. The positive indicators associated with MSC administration reported in the aforementioned studies are mostly related to auxiliary measurements such as modulation of specific biomarker levels, instead of objective clinical indicators of responsiveness such as mortality, PFT, days hospitalized or ventilator-free days.

In yet another larger scale, Phase 1/2 study in patients with Bronchiolitis obliterans syndrome, positive clinical outcomes in response to MSC treatment were reported. For this specific study, 81 patients were enrolled, among which 49 received MSCs in addition to their standard therapy while 32 did not. After 3 months of treatment, 71% of patients in the MSC group had achieved response (PFT improvement and steroid sparing) as compared to 44% in the non-MSC group (p=0.013). Clinical improvement was also accompanied by an increase in IL-10 producing CD5<sup>+</sup> B cells<sup>93</sup>. The varied clinical outcomes reported in the larger scale clinical studies suggest that the beneficial effects of MSCs may be limited to specific pulmonary ailments and not all variations of lung-mediated illnesses, and that results from these studies can be influenced by multiple variables such as MSC dosing and drug preparation regimens.

MSCs have already been delivered to subjects with COVID-19, and the number of MSC-based clinical trials in these subjects is on the rise. To date, there are almost 40 clinical trials (Tables 1 and 2), across multiple phases for evaluating MSC therapy in COVID-19 subjects listed on *clinicaltrials.gov*<sup>37</sup>. The majority of these trials have not been completed and as such results from these studies have not been published. According to one case report, a three-time IV administration of umbilical cord MSCs to a critically ill COVID-19 subject was well tolerated and resulted in reduced serum CRP, normalization of white blood cell counts, and alleviation of pneumonia<sup>94</sup>. In another clinical study of ten subjects with COVID-19 pneumonia, 7 were administered MSCs, while the remaining 3 served as a placebo control. MSC

transplantation was safe with no reports of infusion-related reactions or delayed hypersensitivity reported. In terms of efficacy, for one critically ill patient with severe pneumonia, MSC administration led to a significant and robust decrease in plasma CRP and an increase in oxygen saturation. Compared to the placebo control, subjects in the MSC group experienced normalization of immune cell populations, reduced serum TNF-α and increased IL-10. Of note, the investigators also went on to show that the transplanted MSCs lack ACE2 expression indicating that they are free of COVID-19 infection<sup>95</sup>. According to data from a recently published study, administration of Wharton's Jellyderived MSCs to a single subject with severe COVID-19 pneumonia resulted in resolution of fever and shortness of breath within two days of MSC transplantation and significant reductions in ground-glass opacity and pneumonia infiltration after six days. In this subject, improved clinical function was associated with increased T cell numbers, and reduced inflammatory mediators such as CRP, IL-6 and TNF-  $\alpha^{96}$ . Collectively, the data (Table 3) from these initial reports suggest that administration of MSCs to subjects with severe manifestations of COVID-19 appears to be beneficial in the resolution of major disease symptoms.

# **BARRIERS TO MSC THERAPY**

As described above, MSCs exhibit multiple beneficial effects and data from countless studies conducted in vivo in relevant animal models have shown that MSCs are effective against conditions such as ARDS and *E. coli*-induced pneumonia. However, translation of these positive findings in pre-clinical studies has been difficult to replicate in clinical trials with human subjects, with many MSC trials failing to progress beyond Phase 1. There are many challenges associated with MSC therapy including low in vivo survival rates, dosing, cell isolation and growth strategies, and donor variability issues. (Refer to ref. <sup>97</sup> for a review focused on the challenges associated with MSC therapy).

As is relates to survivability, data obtained from pre-clinical studies indicate that when MSCs are administered in vivo, they do not survive long term<sup>98</sup>. Evaluation of MSC viability 24 h post transplantation in the infarcted myocardium of rats showed that almost 70% of the transplanted MSCs were dead<sup>99</sup>. Interestingly, the short half-life of MSCs does not seem to be directly related to their efficacy; Yang et al. 100 showed that MSCs rescued hematopoiesis in mice despite rapid clearance after infusion. This apparent conundrum between MSC survivability and efficacy is one of the many unresolved issues in the MSC field. Similarly, while there are countless studies<sup>28,29,63,66</sup> which demonstrate that MSC function is largely mediated by paracrine signaling, some reports have indicated otherwise. In addition to paracrine signaling, MSCs also rely on cell-cell contact and direct mitochondrial transfer to elicit their function 101. This was shown by Rosado et al.<sup>102</sup> who demonstrated that MSC immunosuppressive activity required for inhibition of B-cell proliferation was dependent on cell-cell contact between MSCs and T cells. This



Table 3.         Published clinical studies for MSC use in COVID-19 related	es for MSC use in	COVID-19 relate	d conditions.						
Study title	Source of MSCs Trial phase (sample siz	Trial phase (sample size)	Autologous or allogeneic	utologous or COVID-19 condition Location Dose delivery logeneic	Location	Dose delivery	Outcome measures	Results	Ref.
Clinical remission of a critically Umbilical cord ill COVID-19 patient treated by human umbilical cord mesenchymal stem cells		N/A (1)	Allogeneic	Critically ill type COVID-19	China	3 doses of $5 \times 10^7$ cells, IV administration	3 doses of $5 \times 10^7$ Tolerability, Clinical cells, IV improvement administration	Well tolerated, reduced CRP, pneumonia alleviation	4
Transplantation of ACE2-Mesenchymal Stem Cells Improves the Outcome of Patients with COVID-19 Pneumonia	Unknown	N/A (10)	Allogeneic	Covid-19 pneumonia China (common, severe, critically severe)	China	1×10 <sup>6</sup> cells/kg, IV administration	1×10 <sup>6</sup> cells/kg, IV Adverse effects, clinical No adverse events, administration outcomes, change in reduced CRP and T inflammatory and increased IL-10, immune function level normalization of irr cell populations	No adverse events, reduced CRP and TNF- $\alpha$ , increased IL-10, normalization of immune cell populations	56
Intravenous infusion of human Wharton's Jelly N/A (1) umbilical cord Wharton's jelly-derived mesenchymal stem cells as a potential treatment for patients with COVID-19 pneumonia	Wharton's Jelly	N/A (1)	Allogeneic	Covid-19 pneumonia, critically severe	China	1×10 <sup>6</sup> cells/kg, IV Safety, clinical administration improvement	Safety, clinical improvement	No adverse events, resolution of fever and shortness of breath, decreased inflammatory factors (TNF-α and IL-6)	8

specific knowledge gap in MSC function and biology, likely contributes to the lack of effectiveness at the clinical level.

The use of inadequate animal models in a well-controlled setting which does not sufficiently mimic the natural course of human disease is another major contributing factor to the poor outcomes in MSC clinical trials. For example, many of the pre-clinical studies of pulmonary fibrosis in mice have relied on the use of bleomycin to induce an acute phase of the aliment<sup>67</sup>, however, for actual patients it is unlikely that their disease developed in such a manner. As such, there would be differences in the pathophysiology of the disease, and how it responds to treatment in these respective settings. While it is true that there are inherent limitations associated with the use of any animal model used for mimicking human disease, even with *perfect* animal models, there are other discrepancies which need to be addressed before successful translation of MSC therapy can be achieved.

The source of the MSCs, as well as the donor (allogeneic setting) from which they are derived can be a source of huge variability in MSC phenotype and function. We have previously shown that MSCs derived from different donors exhibit significant variation as it relates to cell growth and cytokine secretion profile<sup>30</sup>. In addition, MSCs derived from different tissue sources from the same donor also exhibit variability, as demonstrated by Wegmeyer et al. 103 who showed that MSCs derived from umbilical cord and amniotic membrane from the same donor have different growth characteristics and morphology, and distinctive cytokine and growth factor secretion patterns. For clinical studies, selection of the ideal cell source and optimal donor for generation of the most potent cell product is particularly challenging. Beyond the variability among cell source and donors, variability in cell expansion protocols and cell preparation techniques, can also adversely affect the results of clinical studies. MSCs expanded in cell culture flatware have different growth kinetics compared to their counterparts expanded in automated 3Dbioreactor systems<sup>104</sup>. Clinical outcomes in MSCs trials are also influenced by cell preparation techniques. In the Phase 2 study for ARDS patients discussed above<sup>35</sup>, the investigators reported that there were substantial differences in MSC viability after preparation for infusion across the different sites participating in the trial. They also reported a significant viability-dependent effect on angiopoietin-2 concentrations in plasma 6 h post MSC administration.

Successful translation of MSC-based therapies in clinical settings continues to face significant challenges, with very few studies progressing to Phase 3. Though convincing data relating to MSC efficacy in larger cohort studies are limited, they do exist. According to data from a completed Phase 3 trial in which MSCs derived from Adipose Tissue were administered to subjects with Crohn's disease, 50% of subjects in the MSC cohort achieved remission while only 34% in the control group did (p=0.024)<sup>105</sup>. This study demonstrates that positive clinical outcomes in response to MSC treatment in large scale, Phase 3 studies are feasible, if thorough and detailed consideration is given to key parameters such as cell source and preparation techniques, in the design and execution of the study.

# **UNIQUE CHALLENGES OF MSC THERAPY FOR COVID-19**

We have discussed at length the beneficial effects of MSCs as it relates to their immunomodulatory, regenerative and antimicrobial nature, however, it is important to note that the majority of these findings were delineated in studies conducted in vitro, and in vivo models. As highlighted above, recapitulation of efficacy data obtained in animal models in human subjects remains a major barrier to successful translation of MSC therapy in clinical settings. This is even more relevant in the setting of COVID-19, as there is no pre-clinical efficacy data for the use of MSC in animal models of COVID-19 pneumonia. These pre-clinical studies are crucial, as they can provide valuable information on the consequences, both beneficial and deleterious of MSC



administration specifically in the context of COVID-19 pneumonia. For example, while multiple studies have shown that MSC antimicrobial effects are mediated by LL-37, this molecule has also been associated with the pathogenesis of certain autoimmune disorders such as psoriatic arthritis<sup>106</sup>. The implication of this finding in the context of inflammatory mediated conditions like COVID-19 remains unknown.

Even in the absence of pre-clinical data specific to MSCs and COVID-19, MSCs have already been administered to COVID-19 subjects and there are multiple ongoing trials for use of MSCs in these patients (Tables 1 and 2). The data from the initial studies suggest that MSC administration to subjects with COVID-19 pneumonia is associated with alleviation of disease symptoms <sup>94–96</sup>. As depicted in Tables 1 and 2, for the ongoing and proposed clinical studies, there is considerable variation is terms of cell source and dosing regimen. Though administration of MSCs have already been associated with positive outcomes in COVID-19 subjects, best practices as it relates to formulation, cell source, dosing regimen and timing has not been established. These clinical studies (Tables 1 and 2) evaluating MSCs in COVID-19 subjects could resolve some of these key issues. Pre-clinical studies in relevant animal models of COVID-19 pneumonia could also assist in answering these questions.

# **SUMMARY AND PERSPECTIVES**

COVID-19-associated mortality is increasing daily and as such there remains a critical and unmet need for effective therapeutic options for COVID-19 subjects with severe illness. In patients with critical manifestations of COVID-19, life-threatening complications such as ARDS and multiple organ dysfunction are mediated by extensive inflammation, cytokine storm syndrome, and dysregulated immune responses. MSCs because of their immunomodulatory, regenerative, and antimicrobial properties (Fig. 1) could render several therapeutic effects in the setting of COVID-19. While data from countless clinical studies have established that MSCs are safe, the data with regards to MSC efficacy hasn't been as conclusive.

The safety of MSCs, the extensive pre-clinical efficacy data relating to their use in conditions with pathophysiologies analogous to that of COVID-19 pneumonia, and the initial reports of positive clinical indicators in response to MSC treatment in human cohorts with COVID-19, leads us to believe that MSCs are a potentially beneficial therapeutic option for COVID-19 subjects with critical manifestations of the disease which is governed by a hyper-inflammatory state. While we believe that MSC therapy may be potentially beneficial in this setting, we want to emphasize that MSCs cannot and should not be viewed as a panacea for COVID-19 and that administration of MSCs should not occur in the absence of grave disease complications such as hypoxic respiratory failure and ARDS, and only in the context of an approved clinical trial at no cost to the subjects.

Received: 16 April 2020; Accepted: 6 October 2020; Published online: 26 October 2020

#### **REFERENCES**

- Lu, H., Stratton, C. W. & Tang, Y. W. Outbreak of pneumonia of unknown etiology in Wuhan, China: the mystery and the miracle. J. Med. Virol. 92, 401–402 (2020).
- Prevention, C. f. D. C. a. Locations With Confirmed COVID-19 Cases, by WHO Region. https://www.cdc.gov/coronavirus/2019-ncov/cases-updates/world-map. html (2020)
- Organization, W. H. Rolling Updates on Coronavirus Disease (COVID-19). https:// www.who.int/emergencies/diseases/novel-coronavirus-2019/events-as-theyhappen (2020).
- 4. University, J. H. Coronavirus COVID-19 Global Cases by the Center for Systems Science and Engineering (CSSE) at Johns Hopkins University. https://coronavirus.jhu.edu/map.html (2020).

- Channappanavar, R. & Perlman, S. Pathogenic human coronavirus infections: causes and consequences of cytokine storm and immunopathology. Semin. Immunopathol. 39, 529–539 (2017).
- Adhikari, S. P. et al. Epidemiology, causes, clinical manifestation and diagnosis, prevention and control of coronavirus disease (COVID-19) during the early outbreak period: a scoping review. *Infect. Dis. Poverty* 9, 29 (2020).
- Zhou, P. et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. Nature 579, 270–273 (2020).
- Chen, G. et al. Clinical and immunologic features in severe and moderate Coronavirus Disease 2019. J. Clin. Investig. https://doi.org/10.1172/JCI137244 (2020).
- Chen, T. et al. Clinical characteristics of 113 deceased patients with coronavirus disease 2019: retrospective study. Br. Med. J. 368, m1091 (2020).
- Deng, Y. et al. Clinical characteristics of fatal and recovered cases of coronavirus disease 2019 (COVID-19) in Wuhan, China: a retrospective study. Chin. Med. J. (Engl.) https://doi.org/10.1097/CM9.000000000000824 (2020).
- Lian, J. et al. Analysis of epidemiological and clinical features in older patients with Corona Virus Disease 2019 (COVID-19) out of Wuhan. Clin. Infect. Dis. https://doi.org/10.1093/cid/ciaa242 (2020).
- Chen, N. et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet* 395, 507–513 (2020).
- Shi, S. et al. Association of cardiac injury with mortality in hospitalized patients with COVID-19 in Wuhan, China. *JAMA Cardiol*. https://doi.org/10.1001/jamacardio.2020.0950 (2020).
- Zhou, B., She, J., Wang, Y. & Ma, X. The clinical characteristics of myocardial injury 1 in severe and very severe patients with 2019 novel coronavirus disease. J. Infect. https://doi.org/10.1016/j.jinf.2020.03.021 (2020).
- 15. Qin, C. et al. Dysregulation of immune response in patients with COVID-19 in Wuhan, China. Clin. Infect. Dis. https://doi.org/10.1093/cid/ciaa248 (2020).
- Sun, D. et al. Clinical features of severe pediatric patients with coronavirus disease 2019 in Wuhan: a single center's observational study. World J Pediatr. https://doi.org/10.1007/s12519-020-00354-4 (2020).
- Wu, C. et al. Risk factors associated with acute respiratory distress syndrome and death in patients with coronavirus disease 2019 pneumonia in Wuhan, China. JAMA Intern. Med. https://doi.org/10.1001/jamainternmed.2020.0994 (2020).
- Shen, C. et al. Treatment of 5 critically ill patients with COVID-19 with convalescent plasma. J. Am. Med. Assoc. https://doi.org/10.1001/jama.2020.4783 (2020)
- Gautret, P. et al. Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial. *Int. J. Antimicrob. Agents* https://doi.org/10.1016/j.ijantimicag.2020.105949 (2020).
- Wu, J. et al. Early antiviral treatment contributes to alleviate the severity and improve the prognosis of patients with novel coronavirus disease (COVID-19). J. Intern. Med. https://doi.org/10.1111/joim.13063 (2020).
- Conti, P. et al. Induction of pro-inflammatory cytokines (IL-1 and IL-6) and lung inflammation by Coronavirus-19 (COVI-19 or SARS-CoV-2): anti-inflammatory strategies. J. Biol. Regul. Homeost. Agents https://doi.org/10.23812/CONTI-E (2020).
- Shang, L., Zhao, J., Hu, Y., Du, R. & Cao, B. On the use of corticosteroids for 2019nCoV pneumonia. *Lancet* 395, 683–684 (2020).
- Zhang, W. et al. The use of anti-inflammatory drugs in the treatment of people with severe coronavirus disease 2019 (COVID-19): the experience of clinical immunologists from China. Clin Immunol. https://doi.org/10.1016/j. clim.2020.108393 (2020).
- Dominici, M. et al. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. Cytotherapy 8, 315–317 (2006).
- 25. Gao, F. et al. Mesenchymal stem cells and immunomodulation: current status and future prospects. *Cell Death Dis.* **7**, e2062 (2016).
- Krasnodembskaya, A. et al. Antibacterial effect of human mesenchymal stem cells is mediated in part from secretion of the antimicrobial peptide LL-37. Stem Cells 28, 2229–2238 (2010).
- 27. Ryan, J. M., Barry, F. P., Murphy, J. M. & Mahon, B. P. Mesenchymal stem cells avoid allogeneic rejection. *J. Inflamm.* **2**, 8 (2005).
- Davies, L. C., Heldring, N., Kadri, N. & Le Blanc, K. Mesenchymal stromal cell secretion of programmed death-1 ligands regulates T cell mediated immunosuppression. Stem Cells 35, 766–776 (2017).
- Durand, N., Russell, A. & Zubair, A. C. Effect of comedications and endotoxins on mesenchymal stem cell secretomes, migratory and immunomodulatory capacity. J. Clin. Med. https://doi.org/10.3390/jcm8040497 (2019).
- Russell, A. L., Lefavor, R., Durand, N., Glover, L. & Zubair, A. C. Modifiers of mesenchymal stem cell quantity and quality. *Transfusion* 58, 1434–1440 (2018)
- 31. Gao, L. et al. Phase II multicenter, randomized, double-blind controlled study of efficacy and safety of umbilical cord-derived mesenchymal stromal cells in the

- **n**pj
- prophylaxis of chronic graft-versus-host disease after HLA-haploidentical stem-cell transplantation. J. Clin. Oncol. **34**, 2843–2850 (2016).
- Zhao, K. et al. Immunomodulation effects of mesenchymal stromal cells on acute graft-versus-host disease after hematopoietic stem cell transplantation. *Biol. Blood Marrow Transplant.* 21, 97–104 (2015).
- Cardenes, N. et al. Cell therapy for ARDS: efficacy of endobronchial versus intravenous administration and biodistribution of MAPCs in a large animal model. BMJ Open Respir. Res. 6, e000308 (2019).
- Taki, T. et al. Fetal mesenchymal stem cells ameliorate acute lung injury in a rat cardiopulmonary bypass model. J. Thorac. Cardiovasc. Surg. 153, 726–734 (2017).
- Matthay, M. A. et al. Treatment with allogeneic mesenchymal stromal cells for moderate to severe acute respiratory distress syndrome (START study): a randomised phase 2a safety trial. *Lancet Respir. Med.* 7, 154–162 (2019).
- Wilson, J. G. et al. Mesenchymal stem (stromal) cells for treatment of ARDS: a phase 1 clinical trial. *Lancet Respir. Med.* 3, 24–32 (2015).
- clinicaltrials.gov. Mesenchymal Stem Cells | COVID-19, https://clinicaltrials.gov/ct2/ results?cond=COVID-19&term=Mesenchymal+Stem+Cells&cntry=&state= &citv=&dist (2020).
- Shikha Garg, M. et al. Hospitalization rates and characteristics of patients hospitalized with laboratory-confirmed coronavirus disease 2019—COVID-NET, 14 States, March 1–30, 2020. MMWR Morb. Mortal. Wkly. Rep. 69, 458–464 (2020).
- Guan, W. J. et al. Clinical characteristics of coronavirus disease 2019 in China. N. Engl. J. Med. https://doi.org/10.1056/NEJMoa2002032 (2020).
- Han, H. et al. Analysis of heart injury laboratory parameters in 273 COVID-19 patients in one hospital in Wuhan, China. J. Med. Virol. https://doi.org/10.1002/jmv.25809 (2020).
- Wu, Z. & McGoogan, J. M. Characteristics of and important lessons from the coronavirus disease 2019 (COVID-19) outbreak in China: summary of a report of 72314 cases from the chinese center for disease control and prevention. *J. Am. Med. Assoc.* https://doi.org/10.1001/jama.2020.2648 (2020).
- Xiong, Y. et al. Transcriptomic characteristics of bronchoalveolar lavage fluid and peripheral blood mononuclear cells in COVID-19 patients. *Emerg. Microbes Infect.* 9, 761–770 (2020).
- Li, L. et al. Effect of convalescent plasma therapy on time to clinical improvement in patients with severe and life-threatening COVID-19: a randomized clinical trial. J. Am. Med. Assoc. https://doi.org/10.1001/jama.2020.10044 (2020).
- Wang, M. et al. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. Cell Res. 30, 269–271 (2020).
- Molina, J. M. et al. No evidence of rapid antiviral clearance or clinical benefit with the combination of hydroxychloroquine and azithromycin in patients with severe COVID-19 infection. *Med. Mal. Infect.* 50, 384 (2020).
- Tang, W. et al. Hydroxychloroquine in patients with mainly mild to moderate coronavirus disease 2019: open label, randomised controlled trial. Br. Med. J. 369, m1849 (2020).
- Sun, Z. et al. Potent neutralization of SARS-CoV-2 by human antibody heavychain variable domains isolated from a large library with a new stable scaffold. MAbs 12, 1778435 (2020).
- Meisel, R. et al. Human bone marrow stromal cells inhibit allogeneic T-cell responses by indoleamine 2,3-dioxygenase-mediated tryptophan degradation. *Blood* 103, 4619–4621 (2004).
- Hong, J. W. et al. Immune tolerance of human dental pulp-derived mesenchymal stem cells mediated by CD4(<sup>+</sup>)CD25(<sup>+</sup>)FoxP3(<sup>+</sup>) regulatory T-Cells and induced by TGF-beta1 and IL-10. Yonsei Med. J. 58, 1031–1039 (2017).
- Lu, Y. et al. Human gingiva-derived mesenchymal stem cells alleviate inflammatory bowel disease via IL-10 signalling-dependent modulation of immune cells. Scand. J. Immunol. 90, e12751 (2019).
- 51. Zhu, H. et al. Therapeutic effects of human umbilical cord-derived mesenchymal stem cells in acute lung injury mice. *Sci. Rep.* **7**, 39889 (2017).
- Zhang, Z. H. et al. Mesenchymal stem cells increase expression of heme oxygenase-1 leading to anti-inflammatory activity in treatment of acute liver failure. Stem Cell Res. Ther. 8, 70 (2017).
- Lee, H. J., Kim, S. N., Jeon, M. S., Yi, T. & Song, S. U. ICOSL expression in human bone marrow-derived mesenchymal stem cells promotes induction of regulatory T cells. Sci. Rep. 7, 44486 (2017).
- Jung, Y. J., Park, Y. Y., Huh, J. W. & Hong, S. B. The effect of human adiposederived stem cells on lipopolysaccharide-induced acute respiratory distress syndrome in mice. *Ann. Transl. Med.* 7, 674 (2019).
- Xu, Y. L. et al. Intravenous transplantation of mesenchymal stem cells attenuates oleic acid induced acute lung injury in rats. Chin. Med. J. 125, 2012–2018 (2012).
- Wang, L. et al. Lung-resident mesenchymal stem cells promote repair of LPSinduced acute lung injury via regulating the balance of regulatory T cells and Th17 cells. *Inflammation* 42, 199–210 (2019).
- Gore, A. V. et al. Mesenchymal stem cells increase T-regulatory cells and improve healing following trauma and hemorrhagic shock. J. Trauma Acute Care Surg. 79, 48–52 (2015). discussion 52.

- Asmussen, S. et al. Human mesenchymal stem cells reduce the severity of acute lung injury in a sheep model of bacterial pneumonia. *Thorax* 69, 819–825 (2014).
- 59. Huang, C. et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* **395**, 497–506 (2020).
- Ruan, Q., Yang, K., Wang, W., Jiang, L. & Song, J. Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan, China. *Intensive Care Med.* https://doi.org/10.1007/s00134-020-05991-x (2020).
- Huang, P. et al. Mechanism of mesenchymal stem cell-induced neuron recovery and anti-inflammation. Cytotherapy 16, 1336–1344 (2014).
- Rose-John, S. IL-6 trans-signaling via the soluble IL-6 receptor: importance for the pro-inflammatory activities of IL-6. Int. J. Biol. Sci. 8, 1237–1247 (2012).
- Ding, C. et al. Human amniotic mesenchymal stem cells improve ovarian function in natural aging through secreting hepatocyte growth factor and epidermal growth factor. Stem Cell Res. Ther. 9, 55 (2018).
- Ge, Q. et al. VEGF secreted by mesenchymal stem cells mediates the differentiation of endothelial progenitor cells into endothelial cells via paracrine mechanisms. *Mol. Med. Rep.* 17, 1667–1675 (2018).
- Kamprom, W. et al. Effects of mesenchymal stem cell-derived cytokines on the functional properties of endothelial progenitor cells. Eur. J. Cell Biol. 95, 153–163 (2016).
- Yang, Y. et al. The vascular endothelial growth factors-expressing character of mesenchymal stem cells plays a positive role in treatment of acute lung injury in vivo. Mediators Inflamm. 2016, 2347938 (2016).
- Cahill, E. F., Kennelly, H., Carty, F., Mahon, B. P. & English, K. Hepatocyte growth factor is required for mesenchymal stromal cell protection against bleomycininduced pulmonary fibrosis. Stem Cells Transl. Med. 5, 1307–1318 (2016).
- Xiang, B. et al. Transplantation of menstrual blood-derived mesenchymal stem cells promotes the repair of LPS-induced acute lung injury. *Int. J. Mol. Sci.* https://doi.org/10.3390/ijms18040689 (2017).
- Stern, J. B., Jaffre, S., Dehoux, M. & Crestani, B. Keratinocyte growth factor and Hepatocyte growth factor: their roles in alveolar epithelial repair. *Rev. Mal. Respir.* 20, 896–903 (2003).
- Dong, L. H. et al. The anti-fibrotic effects of mesenchymal stem cells on irradiated lungs via stimulating endogenous secretion of HGF and PGE2. Sci. Rep. 5, 8713 (2015).
- Chen, J. et al. Bone marrow-derived mesenchymal stem cells attenuate phosgene-induced acute lung injury in rats. *Inhal. Toxicol.* 27, 254–261 (2015).
- Boparai, J. K. & Sharma, P. K. Mini review on antimicrobial peptides, sources, mechanism and recent applications. *Protein Pept. Lett.* 27, 4–16 (2020).
- Devaney, J. et al. Human mesenchymal stromal cells decrease the severity of acute lung injury induced by E. coli in the rat. Thorax 70, 625–635 (2015).
- Sutton, M. T. et al. Antimicrobial properties of mesenchymal stem cells: therapeutic potential for cystic fibrosis infection, and treatment. Stem Cells Int. 2016, 5303048 (2016).
- Curley, G. F. et al. Cryopreserved, xeno-free human umbilical cord mesenchymal stromal cells reduce lung injury severity and bacterial burden in rodent Escherichia coli-induced acute respiratory distress syndrome. Crit. Care Med. 45, e202–e212 (2017).
- Sung, D. K. et al. Antibacterial effect of mesenchymal stem cells against *Escherichia coli* is mediated by secretion of beta- defensin- 2 via toll- like receptor 4 signalling. *Cell Microbiol.* 18, 424–436 (2016).
- Qian, X. et al. Exosomal microRNAs derived from umbilical mesenchymal stem cells inhibit hepatitis C virus infection. Stem Cells Transl. Med. 5, 1190–1203 (2016).
- Miteva, K. et al. Mesenchymal stromal cells inhibit NLRP3 inflammasome activation in a model of coxsackievirus B3-induced inflammatory cardiomyopathy. Sci. Rep. 8, 2820 (2018).
- Chan, M. C. et al. Human mesenchymal stromal cells reduce influenza A H5N1associated acute lung injury in vitro and in vivo. *Proc. Natl Acad. Sci. USA* 113, 3621–3626 (2016).
- 80. Li, Y. et al. Mesenchymal stromal cell treatment prevents H9N2 avian influenza virus-induced acute lung injury in mice. Stem Cell Res. Ther. 7, 159 (2016).
- Loy, H. et al. Therapeutic implications of human umbilical cord mesenchymal stromal cells in attenuating influenza A(H5N1) virus-associated acute lung injury. J. Infect. Dis. 219, 186–196 (2019).
- Schwartz, Y. S. et al. BCG infection in mice is promoted by naive mesenchymal stromal cells (MSC) and suppressed by poly(A:U)-conditioned MSC. *Tuberculosis* 101, 130–136 (2016).
- Kanazawa, M. et al. The influence of systemically or locally administered mesenchymal stem cells on tissue repair in a rat oral implantation model. *Int. J. Implant Dent.* 4, 2 (2018).
- Fischer, U. M. et al. Pulmonary passage is a major obstacle for intravenous stem cell delivery: the pulmonary first-pass effect. Stem Cells Dev. 18, 683–692 (2009).

**n**pj

- Li, X. et al. Targeted migration of bone marrow mesenchymal stem cells inhibits silica-induced pulmonary fibrosis in rats. Stem Cell Res. Ther. 9, 335 (2018).
- Chambers, D. C. et al. Mesenchymal stromal cell therapy for chronic lung allograft dysfunction: results of a first-in-man study. Stem Cells Transl. Med. 6, 1152–1157 (2017).
- Keller, C. A. et al. Feasibility, safety, and tolerance of mesenchymal stem cell therapy for obstructive chronic lung allograft dysfunction. Stem Cells Transl. Med. 7, 161–167 (2018).
- 88. Stolk, J. et al. A phase I study for intravenous autologous mesenchymal stromal cell administration to patients with severe emphysema. *QJM* **109**, 331–336 (2016)
- 89. Tzouvelekis, A. et al. A prospective, non-randomized, no placebo-controlled, phase lb clinical trial to study the safety of the adipose derived stromal cells-stromal vascular fraction in idiopathic pulmonary fibrosis. *J. Transl. Med.* 11, 171 (2013).
- Weiss, D. J., Casaburi, R., Flannery, R., LeRoux-Williams, M. & Tashkin, D. P. A placebo-controlled, randomized trial of mesenchymal stem cells in COPD. *Chest* 143, 1590–1598 (2013)
- Zheng, G. et al. Treatment of acute respiratory distress syndrome with allogeneic adipose-derived mesenchymal stem cells: a randomized, placebocontrolled pilot study. Respir. Res. 15, 39 (2014).
- Simonson, O. E. et al. In vivo effects of mesenchymal stromal cells in two patients with severe acute respiratory distress syndrome. Stem Cells Transl. Med. 4. 1199–1213 (2015).
- Chen, S. et al. The efficacy of mesenchymal stem cells in bronchiolitis obliterans syndrome after allogeneic HSCT: a multicenter prospective cohort study. EBio-Medicine 49, 213–222 (2019).
- Liang, B. et al. Clinical remission of a critically ill COVID-19 patient treated by human umbilical cord mesenchymal stem cells: A case report. Medicine (Baltimore) 99. e21429. (2020).
- 95. Leng, Z. et al. Transplantation of ACE2(-) mesenchymal stem cells improves the outcome of patients with COVID-19 pneumonia. *Aging Dis.* **11**, 216–228 (2020).
- Zhang, Y. et al. Intravenous infusion of human umbilical cord Wharton's jellyderived mesenchymal stem cells as a potential treatment for patients with COVID-19 pneumonia. Stem Cell Res. Ther. 11, 207 (2020).
- 97. Galipeau, J. & Sensebe, L. Mesenchymal stromal cells: clinical challenges and therapeutic opportunities. *Cell Stem Cell* 22, 824–833 (2018).
- 98. Eggenhofer, E. et al. Mesenchymal stem cells are short-lived and do not migrate beyond the lungs after intravenous infusion. *Front. Immunol.* **3**, 297 (2012).
- Liu, X. B. et al. Angiopoietin-1 preconditioning enhances survival and functional recovery of mesenchymal stem cell transplantation. J. Zhejiang Univ. Sci. B 13, 616–623 (2012).
- 100. Yang, X., Balakrishnan, I., Torok-Storb, B. & Pillai, M. M. Marrow stromal cell infusion rescues hematopoiesis in lethally irradiated mice despite rapid clearance after infusion. Adv. Hematol. 2012, 142530 (2012).
- Silva, L. H. A. et al. Strategies to improve the therapeutic effects of mesenchymal stromal cells in respiratory diseases. Stem Cell Res. Ther. 9, 45 (2018).
- Rosado, M. M. et al. Inhibition of B-cell proliferation and antibody production by mesenchymal stromal cells is mediated by T cells. Stem Cells Dev. 24, 93–103 (2015).
- 103. Wegmeyer, H. et al. Mesenchymal stromal cell characteristics vary depending on their origin. *Stem Cells Dev.* **22**, 2606–2618 (2013).

- 104. Haack-Sorensen, M. et al. Culture expansion of adipose derived stromal cells. A closed automated quantum cell expansion system compared with manual flaskbased culture. J. Transl. Med. 14, 319 (2016).
- Panes, J. et al. Expanded allogeneic adipose-derived mesenchymal stem cells (Cx601) for complex perianal fistulas in Crohn's disease: a phase 3 randomised, double-blind controlled trial. *Lancet* 388, 1281–1290 (2016).
- 106. Yuan, Y. et al. Identification of novel autoantibodies associated with psoriatic arthritis. *Arthritis Rheumatol.* **71**, 941–951 (2019).

#### **ACKNOWLEDGEMENTS**

We would like to thank the Mayo Clinic Center for Regenerative Medicine for funding support.

#### **AUTHOR CONTRIBUTIONS**

N.D. designed, wrote and revised the paper. J.M. wrote and revised the paper. ACZ designed, revised, and supervised the writing of this paper.

#### COMPETING INTERESTS

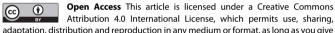
The authors declare no competing interests.

#### ADDITIONAL INFORMATION

Correspondence and requests for materials should be addressed to A.C.Z.

Reprints and permission information is available at http://www.nature.com/reprints

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <a href="https://creativecommons.org/licenses/by/4.0/">https://creativecommons.org/licenses/by/4.0/</a>.

© The Author(s) 2020