

# Supramolecular Chirality Synchronization in Thin Films of Plasmonic Nanocomposites

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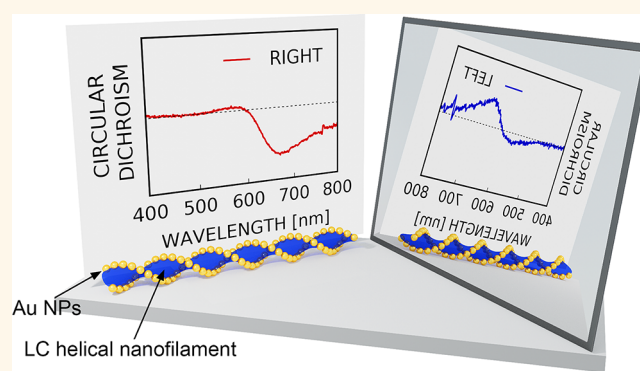
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**ABSTRACT:** Mirror symmetry breaking in materials is a fascinating phenomenon that has practical implications for various optoelectronic technologies. Chiral plasmonic materials are particularly appealing due to their strong and specific interactions with light. In this work we broaden the portfolio of available strategies toward the preparation of chiral plasmonic assemblies, by applying the principles of chirality synchronization—a phenomenon known for small molecules, which results in the formation of chiral domains from transiently chiral molecules. We report the controlled cocrystallization of 23 nm gold nanoparticles and liquid crystal molecules yielding domains made of highly ordered, helical nanofibers, preferentially twisted to the right or to the left within each domain. We confirmed that such micrometer sized domains exhibit strong, far-field circular dichroism (CD) signals, even though the bulk material is racemic. We further highlight the potential of the proposed approach to realize chiral plasmonic thin films by using a mechanical chirality discrimination method. Toward this end, we developed a rapid CD imaging technique based on the use of polarized light optical microscopy (POM), which enabled probing the CD signal with micrometer-scale resolution, despite of linear dichroism and birefringence in the sample. The developed methodology allows us to extend intrinsically local effects of chirality synchronization to the macroscopic scale, thereby broadening the available tools for chirality manipulation in chiral plasmonic systems.

**KEYWORDS:** supramolecular chirality, chirality synchronization, nanostructures, dynamic metamaterials, liquid crystals, circular dichroism imaging, helical assemblies



Developing the ability to efficiently manipulate light at will is foreseen as one of the crucial limitations for the emerging technologies of the 21st century. Inspired by nature, scientists and engineers aim at controllably fabricating nanoarchitectures with sophisticated, hierarchical structures, in which the spatial arrangement of building blocks defines the optical response of the material.<sup>1–6</sup> Development of chiral assemblies made of plasmonic nanoparticles is particularly interesting due to their superior light–matter interactions in comparison to purely organic matter.<sup>3,7–12</sup> Stimulated by recent visionary reports, much interest has been revived toward applying the principles of chiral plasmonics to realize superlenses,<sup>13</sup> chiral catalysts,<sup>14,15</sup> negative refractive index materials,<sup>16</sup> perfect absorbers,<sup>17</sup> broadband circular polarizers,<sup>18</sup> selective reflectors,<sup>19</sup> biosensors,<sup>20</sup> chiral quantum optical devices,<sup>21</sup> chiral emission,<sup>22</sup> and biomanipulation.<sup>23</sup> For this purpose, harnessing chiral plasmonic properties in complex, organic–inorganic nanocomposites is often required. Such composites are usually achieved by either combining intrin-

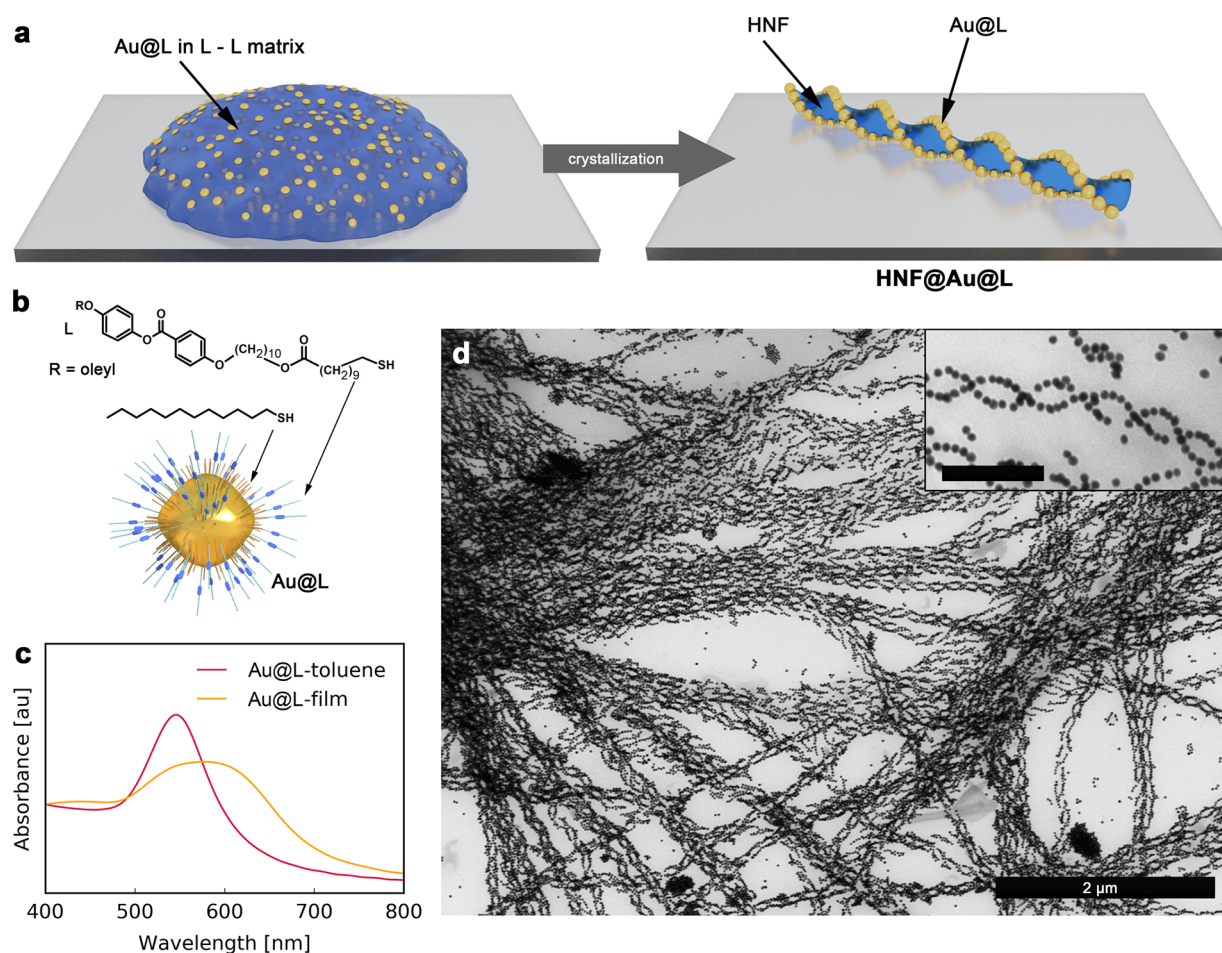
sically chiral NPs<sup>24–27</sup> with achiral materials, through chirality transfer from chiral entities to achiral NPs,<sup>28</sup> or by using other symmetry breaking stimulants, like circularly polarized light<sup>29</sup> or imprinting.<sup>30</sup> The self-assembly driven,<sup>31–34</sup> template-based approach is particularly appealing, as it provides the ability to tailor chiral plasmonic systems. This approach has been realized by tuning the interactions between NPs and chiral DNA,<sup>35,36</sup> peptides,<sup>37,38</sup> proteins,<sup>39</sup> polymers,<sup>40,41</sup> silica,<sup>42</sup> and small molecules<sup>43</sup> used as templates, typically in colloidal or gel systems, and often mimicking the helical geometry of natural materials.<sup>44–46</sup> However, integration of these materials into

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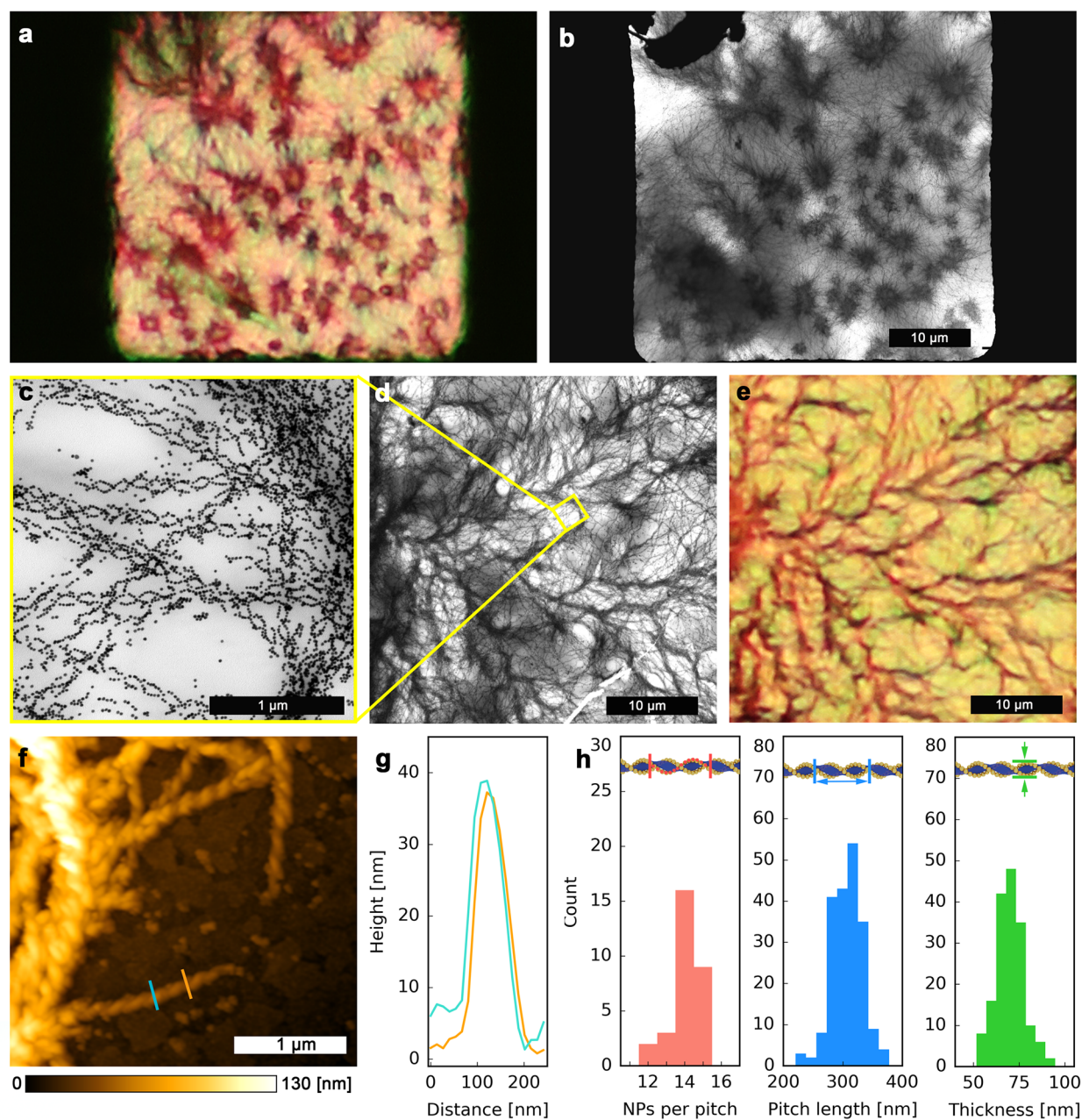


**Figure 1.** Design and preparation of helical assemblies. (a) Scheme of the formation of Au NPs helical assemblies templated by LCs. (b) Model of L-capped Au quasi-spherical nanoparticles (Au@L) and molecular structure of surface ligands (L and DDT). (c) UV-vis spectra of Au@L dispersion in the toluene and HNF@Au@L films. (d) Representative TEM image of HNF@Au@L; the scale bar in the inset corresponds to 250 nm.

functional devices requires accessible and scalable fabrication methods that rely on inexpensive and readily available constituents. One route to achieve this goal comprises the incorporation of metal nanoparticles within chiral liquid crystal (LC) matrices.<sup>47–50</sup> While yielding materials that exhibit interesting optical properties, this method is limited by the requisite that the organic template must be intrinsically chiral. We propose that achiral constituents can also be used to prepare chiral nanocomposites, which may reduce development costs, if only by broadening the scope of available organic building blocks. In this context, the phenomenon of chirality synchronization in liquid crystal thin films, in which achiral organic molecules break the mirror symmetry by cooperative locking at a given chiral state,<sup>51</sup> seems particularly appealing. One of the most prominent cases of chirality synchronization is related to selected bent-core liquid crystalline compounds<sup>52,53</sup> that are able to form helical nanofilament (HNF) phases.<sup>54,55</sup> At a phase transition into the LC state, nucleation points adopt a certain chiral symmetry, which then propagates through the growth of a domain.<sup>56</sup> The final state is a conglomerate structure made of domains with both left and right handedness. Given the possibility of characterizing homochiral domains using far field methods, as well as the versatility and plasticity of soft,<sup>57–60</sup> thermotropic LC thin films, these materials can be of high technological relevance.<sup>61–65</sup> Recently, some of us demon-

strated the incorporation of metal nanoparticles into HNF materials.<sup>66</sup> However, the principles of chirality synchronization and the resulting optical activity properties are still restricted to the organic-only realm,<sup>51,67–69</sup> thus lacking the advantages of strong light–matter interactions. Although the ability to introduce metal nanoparticles in this kind of material should reinforce the optical properties, up to date only structural asymmetry of NP decorated HNFs has been achieved, with no confirmed plasmonic optical activity.

In this contribution we aimed at achieving plasmonic chiral materials through chirality synchronization in nanocomposites. For this purpose, we used a LC mediated method, which results in helical nanofilaments selectively decorated with gold NPs.<sup>66</sup> Following this methodology, quasi-spherical Au NPs were arranged into a double helical structure. Chirality synchronization enabled the formation of homochiral domains, which was evidenced by far field CD spectroscopy. To enable further manipulation of the materials, a simplified optical microscopy-based method was developed toward the rapid identification of chiral domains by their CD signals, thereby addressing the potential interference from linear effects (linear dichroism, birefringence). CD maps were used to guide the selective removal of homochiral domains (chirality discrimination), leading to nanocomposites exhibiting plasmonic chirality at the bulk scale.



**Figure 2.** Structural analysis of HNF@Au@L. (a) Optical microscopy image of HNF@Au@L forming multiple domains. Black areas on the sides are parts of the TEM grid mesh. (b) TEM image of the same sample area. (c, d) TEM images showing HNF@Au@L at high and low magnifications, respectively. (e) Optical microscopy image of the same sample area. (f) AFM 2D elevation profile of an edge of an HNF@Au@L domain on an ITO-covered glass substrate. (g) Height profiles from the AFM image shown in panel (f); the colors of the curves correspond to those of the corresponding lines in (f). (h) Structural analysis of HNF@Au@L based on TEM imaging: NPs population per pitch, as well as pitch length and thickness of the helical nanofibers.

## RESULTS AND DISCUSSION

**Design and Structure of Helical Nanocomposites.** In an attempt to prepare thin films exhibiting plasmonic chirality, we used a solid state crystallization method, recently developed by some of us, leading to helical assemblies of NPs.<sup>66</sup> In this method, organic material (L-L) having the ability to form helical nanofilaments is first codispersed with LC-functionalized NPs in toluene, and then the dispersion is dropcasted onto a substrate and heated to reach the isotropic liquid phase. Hierarchical, helical nanocomposites are formed on subsequent cooling (Figure 1a). The underlying mechanism involves a phase

transition of the organic compound from isotropic liquid into dendritic, helical filaments, while NPs selectively decorate the edges of such filaments as they are expelled from the bulk L-L phase into the air/fiber boundary.<sup>66</sup> To maximize the efficiency of composite formation we followed a few design principles:

- First, as the organic template material we chose a previously reported L-L dimer (Figure S1).<sup>66</sup> Notably, L-L forms HNFs at a relatively low temperature (110 °C), which prevents heat-induced agglomeration of NPs in the course of composite preparation.

- Second, to achieve a strong plasmonic response from the helical composite in the visible region, we prepared helical assemblies of 23 nm diameter quasi-spherical gold NPs (Figure 1d). The NP size was selected as a compromise between the need to ensure strong plasmonic coupling within the assemblies (which requires larger NPs) and the limited width of the edge in L-L helical nanofilaments (~30 nm), which are selectively decorated with NPs.
- Finally, efficient formation of helical composites requires the NPs to be well dispersed in the melted organic matrix; thus, chemical compatibility between NPs and the matrix is necessary. Therefore, we incorporated the L ligand (Figure 1b) within the organic shell on the NPs, which was previously shown to ensure compatibility of NPs with the L-L matrix. Using optical microscopy, it was found that an L:DDT (dodecanethiol) molar ratio of 1:1 during the ligand exchange process (which translated to a 1:2 L:DDT ratio found in the NP shell using thermogravimetric analyses, TGA, Figure S2) provides the highest compatibility and allows a uniform dispersion of NPs during the entire heat-cool cycle, whereas a lower proportion of L (1:3 L:DDT during the exchange process), or the exclusive presence of L, were found to result in irreversible NP aggregation during heating (Figure S3). The latter result might suggest that a dense structure of the L shell over the NPs hinders interactions between the ligand and the matrix molecules and thus a lowered density of L grafting is required for efficient miscibility. This can be achieved by means of a mixed organic coating, comprising L and shorter (alkyl) thiols. In this case, we hypothesize that alkyl ligands serve as a spacer and thus create supramolecular pockets for the penetration of matrix molecules; *i.e.*, penetration of the organic shell becomes more efficient, thereby increasing the stability of NP dispersions. However, there is a lower threshold of L content in the shell, below which chemical compatibility between NPs and L-L is not achieved, as evidenced by poor results for the 1:3 L:DDT sample. Similar observations were previously reported for NP composites with LCs<sup>70–73</sup> and polymers.<sup>74</sup> Based on the above, further experiments were carried out using NPs functionalized using 1:1 L:DDT (referred to as Au@L).

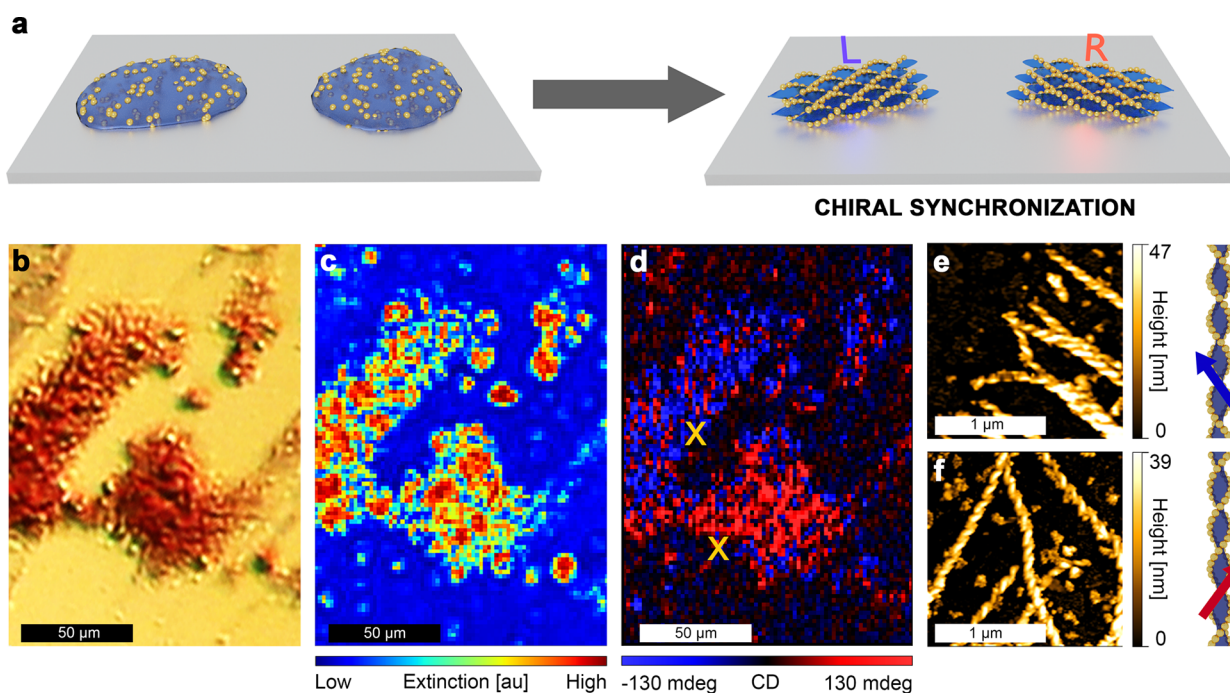
As expected, Au@L exhibited a well-defined localized surface plasmon resonance (LSPR) band centered at ~540 nm in a toluene dispersion (Figure 1c). In a condensed state after the deposition as a thin film, the band was found to broaden and redshift to ~590 nm (Figure 1c) due to plasmon coupling when the distance (center-to-center) between the neighboring NPs was decreased to ~26 nm, which roughly translates to 2–3 nm surface-to-surface distance (Figure S4).

To prepare HNF@Au@L, the drop-casted mixtures were quickly (~40 °C/min) heated up to 140 °C, *i.e.*, ~30 K above the L-L melting point, to ensure an even dispersion of NPs, as confirmed by optical inspection (Figure S5). In comparison to our previous efforts,<sup>66</sup> an increased temperature was required to achieve well-dispersed NPs, which was likely related to their larger size. The sample was then cooled down to room temperature, at a controlled rate of 40 K/min until reaching 120 °C, and then the rate was reduced to 3 K/min. The first, rapid cooling step was designed to minimize the time NPs are exposed to elevated temperatures, which was not required in our previous contribution.<sup>66</sup> Subsequently, a slower cooling rate was

used to stimulate the growth of large domains. Examination by atomic force microscopy (AFM, Figure S6), TEM (Figure 1d, Figure S7), and scanning electron microscopy (SEM, Figure S8) confirmed successful formation of helical nanofilaments decorated with NPs (HNF@Au@L).

One of the key aspects allowing easy and efficient manipulation of HNF@Au@L assemblies is the ability to quickly recognize well-developed helical composites using standard optical microscopy. For this purpose, we compared images obtained by optical and electron microscopies, taken for the same areas of the sample (Figure 2a,b, Figure 2c–e). These images revealed an identical mesoscopic morphology of the sample, confirming the possibility to recognize bundles of HNF@Au@L using an optical microscope. Analysis of TEM images revealed the average pitch of the helical nanostructures to be ~300 nm, lateral thickness to be ~68 nm (measured as the distance between centers of Au NPs on opposite sides of the fibers, which translates into ~44 nm distance between the edges of the organic fibers), and ~14 particles per pitch for a fully decorated fiber (Figure 2f–h). These structural parameters are comparable to those previously reported for helical nanofilaments formed by liquid crystals.<sup>75</sup> Using AFM, we were able to assess the vertical thickness (height) of a single HNF@Au@L fiber, to be ~35 nm (Figure 2f–h, Figure S6). The small discrepancy between the values of the lateral thickness and height can be explained by the soft character of the composite, which leads to fibers flattening on the substrate.

**Chirality Synchronization Probed with Micro-CD Measurements.** Observations of the crystallization process using optical microscopy (Supporting Information Movie 1), as well as optical images of the assemblies (Figure 2a,e, Figure S9, and Figure S10), allowed us to confirm that the growth of HNF@Au@L starts by spontaneous formation of nucleation centers, followed by the dendritic expansion of HNF domains. This growth continues until the isotropic Au@L/L-L melt in contact with the growing domain is completely exhausted. In general, the mechanism of chiral synchronization occurs during the formation of the HNF; *i.e.*, newly added molecules twist in the same direction as the molecules within existing HNF. In our case, Au@L nanoparticles are present within the melt and tend to attach to forming HNFs. This tendency arises from the chemical compatibility (Figure S1) between the L ligand and the L-L matrix. If chiral synchronization occurs independently of the NP deposition on HNFs (in other words, the presence of NPs does not disturb chirality synchronization of L-L), domains with chiral plasmonic response should be formed. Given the achiral environment at the initial stage when nucleation centers are formed, the ratio between left- and right-handed domains is expected to be 1:1, rendering the sample achiral at the bulk scale (racemic). Thus, we decided to perform measurements at the microscale, which should reveal optical activity at the single domain level. For this purpose, we deposited a sample on a glass substrate, allowing for transmission CD measurements with far field, discretely modulated circularly polarized light (DMCPL). In these measurements, one can evaluate directly the difference between extinction ( $\Delta\text{Ext}$ ) of left and right circularly polarized light (LCP and RCP, respectively),<sup>76,77</sup> thereby enabling CD measurements at the domain scale, despite the strong linear dichroism of HNF@Au@L filaments (Note S1 and Figure S11). The DMCPL approach allows us to eliminate such signal interferences. By using optical microscopy (which provides fast HNF@Au@L domain identification, Figure 2c), a sample area with more than one HNF@Au@L domain was selected (Figure



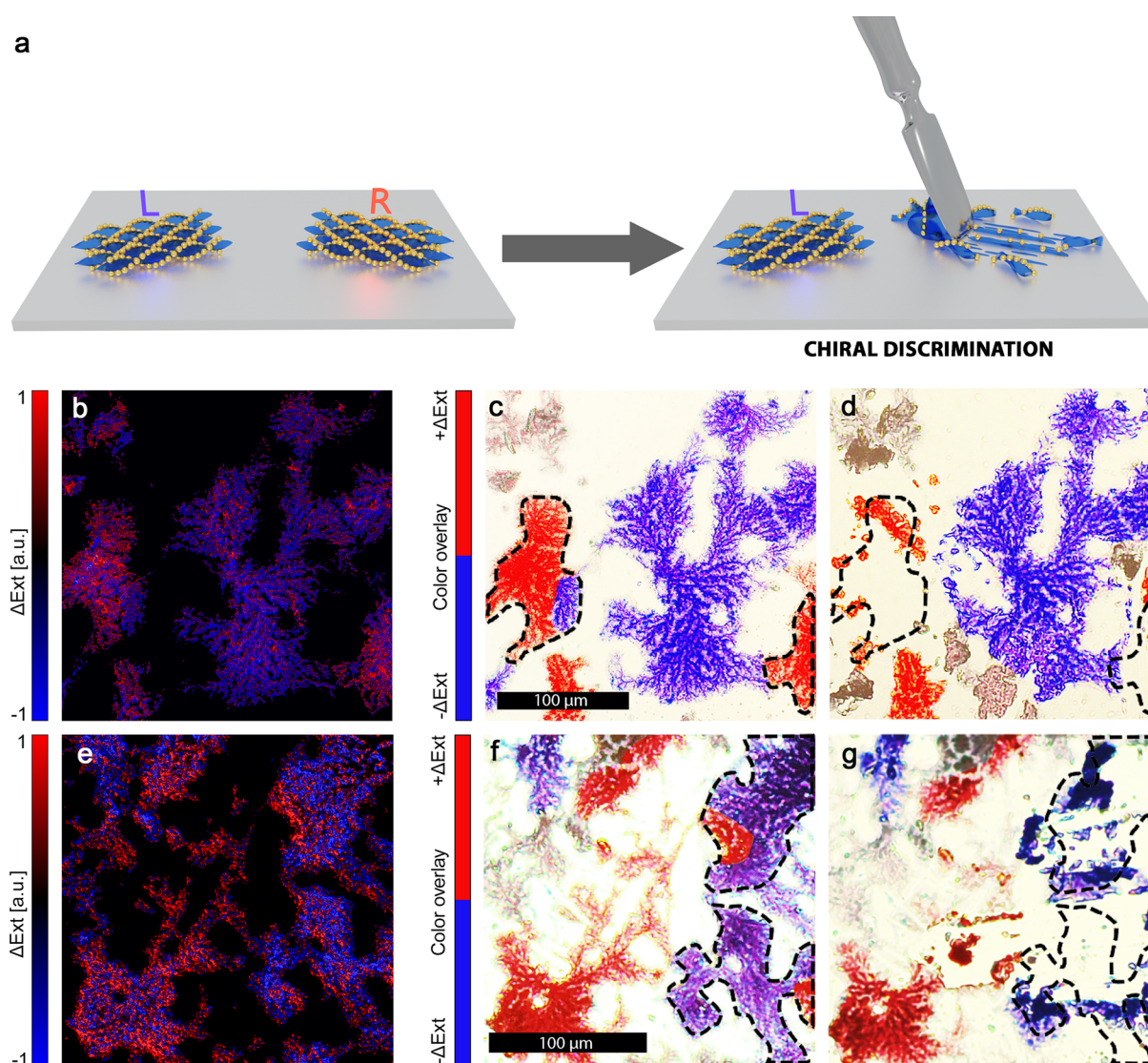
**Figure 3.** Micrometer-scale circular dichroism measurements. (a) Graphical representation of the chiral synchronization process. (b) Optical micrograph, (c) extinction intensity map at 550 nm, and (d) CD map obtained using a CD microscope with DMCPPL at 550 nm from the same region of an HNF@Au@L sample; note that two domains of opposite handedness are revealed. Spatial distribution of CD values is mapped in ellipticity units. (e) AFM height map of a selected region from a left-handed domain (upper X mark in (d) indicates the place of acquisition) and (f) AFM height map of a selected region from a right-handed domain (lower X mark in (d) indicates the place of acquisition). Next to each AFM height map, a graphical model of the helical nanofilament with the corresponding handedness is presented.

3a). First of all, we found that domains can indeed show either negative or positive  $\Delta\text{Ext}$  at 550 nm (Figure 3d), confirming the homochirality of particular domains. The 550 nm light was obtained by filtering white light from a supercontinuum laser (EXR-15, NKT Photonics) with a monochromator. At such a light wavelength we were able to isolate the chiral optical response of the nanoparticles from the optical activity of pure organic HNF, which should arise at the UV part of the spectrum (HNF absorbance peaks are below 300 nm, Figure S12), proving that the registered signal originates from plasmonic chirality. It should be noted that domains with the opposite CD signal were undistinguishable when compared by optical microscopy imaging or extinction intensity (Figure 3b,c, respectively), as should be expected if the only difference is the handedness of the HNF. In order to correlate the optical response of the domains with their structure, we performed AFM measurements and found that negative  $\Delta\text{Ext}$  was recorded from domains consisting mostly of left-handed HNF (Figure 3e), whereas positive  $\Delta\text{Ext}$  originated from domains where right-handed HNFs predominate (Figure 3f). Additional AFM profiles, as well as handedness count analysis, are provided in Figure S13. The observed CD signals are a direct evidence that the helical arrangement, strong plasmonic properties, and relatively short interparticle spacing resulted in chiral plasmonic coupling of NPs. In each domain, small spots (consisting of a few pixels) with opposite handedness were observed. Based on the analysis of the AFM data, most such spots are considered to originate from noise and/or artifacts (Note S2).

**Bulk Homochirality via Chirality Discrimination.** The results presented so far evidence that single HNF@Au@L domains are optically active. However, to achieve a bulk chiral response some kind of chirality discrimination mechanism

should be applied, leading to an enantiomeric excess of domains with a particular handedness. For this purpose, we devised a simplified measurement technique to probe the plasmonic chirality of micrometer-size domains. The technique is based on a conventional polarization optical microscopy (POM) setup,<sup>78</sup> in which the sample is illuminated by LCP or RCP through installation of a quarter-wave plate (we will refer to the technique as POM-CD, Figure S14 and Note S2). By performing analysis of multiple images for each polarization, we were able to obtain a CD signal map despite of strong linear dichroism of the sample, similar to the case of DMCPPL measurements (Figure 3d). The resulting extinction difference map is less accurate and prone to interference (e.g., small changes in the position of the sample during image acquisition would render the results unreliable, Figure S14g,h). However, the time required to obtain data enabling qualitative domain distinction by the CD-signal is much shorter compared to the DMCPPL microscopy that we have at present (Note S2).

We then used the technique described above to aid the preparation of thin film samples exhibiting chiral plasmonic properties at a bulk scale. Two samples with HNF@Au@L domains covering a relatively large area (around  $\sim 9 \text{ mm}^2$ ; see Figure S9a for an optical image of one of the samples) were prepared and analyzed with the POM-CD method, using a green filter to restrict the wavelengths to  $532 \pm 20 \text{ nm}$ . We were able to identify the CD-signal of domains with micrometer scale resolution (false color images in Figure 4c,f). We then reduced the population of the domains based on their CD signal through mechanical removal (as demonstrated on Figure 4a): we targeted domains yielding a positive signal in the first sample (Figure 4b–d) and a negative signal in the second one (Figure 4e–g). This proof-of-principle study was performed using a



**Figure 4.** Controlled domain separation. (a) Graphical representation of the chiral discrimination through manual HNF@Au@L removal. (b)  $\Delta\text{Ext}$  maps obtained through the POM-CD method, of a HNF@Au@L sample fragment at  $532 \pm 20$  nm. (c) Optical microscopy image of the same region shown in (b) with false-color overlay to highlight domain handedness. (d) Optical microscopy image of the same region shown in (c) after removal of right-handed domains. (e, f, g) Images analogous to those shown in panels (b)–(d) for a sample in which left-handed domains were selectively removed.

simple approach based on optical microscope-assisted, needle-point scratching, which trades precision and scalability for a fast and easy setup with little need for specialized equipment. Due to these limitations, only a certain degree of unbalance between CD positive and negative domains can be introduced, as only relatively large domains ( $>100 \mu\text{m}$  diameter) with no opposite-signal neighbors could be targeted.

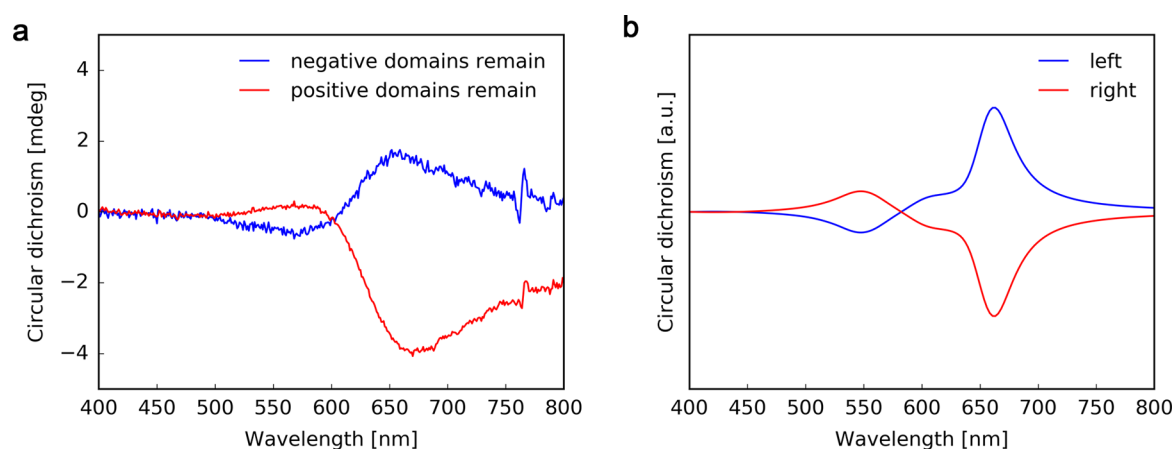
Both samples were then measured using a commercial CD spectrophotometer, yielding CD signals with a clearly visible Cotton effect around the LSPR absorbance peak—an effect that is typical for strongly absorbing chiral materials<sup>79</sup> and characteristic to NP-decorated helical filaments<sup>80</sup> (Figure 5a). Importantly, changing the orientation of the sample by rotating it along the beam direction axis did not affect the signal (Figure S15). The averaged linear dichroism effect of HNF@Au@L is canceled due to lack of orientational order of filaments on the sample level.

To further confirm that the observed response stems from the chiral arrangement of nanoparticles, we conducted finite

difference time-domain (FDTD, Figure 5b) simulations of a helical nanofilament structure made of Au NPs arranged with the average experimentally determined structural parameters (see Materials and Methods for details). The simulated signals are in good agreement with the experimental data, confirming successful preparation of thin film plasmonic structures made of self-assembled helical composites.

## CONCLUSIONS

In conclusion, we described a method to obtain chiral plasmonic materials using LC-based self-assembly of gold NPs into helical aggregates. The intense chiral plasmonic response at the microscale is possible because of (1) the intense LSPR and plasmon coupling of relatively large nanoparticles and (2) chiral synchronization—the ability of HNF@Au@L to retain handedness during domain growth, resulting in homochiral domains. The high Au filling fraction ( $\sim 30\%$  mass Au) of the material allows one to observe strong optical properties even for thin film samples.<sup>81</sup> In order to quickly measure CD at a microscale, a



**Figure 5.** Bulk plasmonic chirality after domain separation. (a) Experimental CD spectra of samples after selective removal of domains with the same handedness. Responses of samples with an excess of right- or left-handed domains are shown in blue and red, respectively. (b) FDTD calculation of the CD spectrum of a single helical nanofilament decorated with 23 nm gold nanoparticles with left- and right-handedness (blue and red, respectively).

simplified, POM-based method for streamlined, qualitative measurements was adapted, which revealed the plasmonic chiral response despite the linear dichroism and birefringence in the thin-film material. This routine allowed us to efficiently map the optical response of large sample areas, which would otherwise be impractical at any larger scale. Being able to quickly determine the CD signal of HNF@Au@L domains, we proposed and successfully demonstrated an efficient approach to obtain plasmonic chirality at a bulk scale, namely, controlled chiral discrimination—a manual removal of either left- or right-handed domains. The approach presented here is a direct evidence that, with the aid of micro-CD measurements, it is possible to translate the chiral synchronization from a local effect at the microscopic scale into a global phenomenon with a bulk-scale optical response, allowing its role in the field of chiral plasmonic materials to be redefined. Utilization of an achiral liquid crystal template ensures low fabrication costs and scalability of the process to maximize the application potential and provides a route toward further reconfigurability.

## MATERIALS AND METHODS

**Materials.** Tetrachloroauric acid ( $\text{HAuCl}_4$ ,  $\geq 99\%$ ), hexadecyltrimethylammonium chloride (CTAC, 25 wt % in water), benzyltrimethylhexadecylammonium chloride (BDAC), citric acid ( $\geq 99.5\%$ ), sodium borohydride ( $\text{NaBH}_4$ ), L-ascorbic acid (AA,  $\geq 99\%$ ), tetrahydrofuran (THF, anhydrous,  $\geq 99.9\%$ ), dodecanethiol (DDT,  $\geq 98\%$ ), and toluene (anhydrous,  $\geq 99.8\%$ ) were purchased from Sigma-Aldrich. All chemicals were used without further purification. Milli-Q water was used in all experiments.

**Synthesis of Au NPs.** Synthesis of Au NPs was carried out according to the seed-mediated growth method reported by Sánchez-Iglesias et al.<sup>82</sup> For the seed preparation, 10 mL of 0.25 mM  $\text{HAuCl}_4$  solution was reduced by 0.25 mL of freshly prepared 25 mM  $\text{NaBH}_4$  in the 50 mM CTAC solution, in the presence of citric acid (5 mM) under vigorous stirring. After a few minutes of stirring, the solution was heated at 90 °C under gentle stirring for 90 min. For Au NPs synthesis, 550  $\mu\text{L}$  of seeds was added under vigorous stirring to the growth solution containing 10 mL of 100 mM BDAC, 0.1 mL of 50 mM  $\text{HAuCl}_4$ , and 75  $\mu\text{L}$  of 100 mM AA. The mixture was left undisturbed for 30 min. Before further modifications Au NPs were centrifuged two times (7000 rpm over 15 min) to remove the excess of reactants.

**Au NPs Functionalization.** The obtained NPs were centrifuged (7000 rpm over 15 min) and redispersed in L-DDT solution in THF in an ultrasonic bath. In a typical process, the molar ratio of  $\text{Au}^0$  to L to DDT was 1:1:1. The mixture was left under mild stirring overnight.

Afterward, the excess of ligand was removed through centrifugation. The precipitate containing modified nanoparticles (Au@L) was dispersed in toluene. In the usual procedure a 1:1:1 molar ratio corresponds to 0.1 mg of Au NPs, 0.4 mg of L, and 0.1 mg of DDT.

**Synthesis of Organic Materials.** L-L and L were synthesized using previously established protocols (Figure S1).<sup>66</sup> Shortly, in the first step, an alkyl spacer unit was synthesized. For this purpose, 11-bromoundecanoic acid was reacted with oxalyl chloride. The formed acid chloride was mixed with 1,12-dodecanediol to obtain an intermediate product (S). Subsequently, ethyl 4-hydroxybenzoate was reacted with an oleyl alcohol in Mitsunobu reaction. Then, a potassium salt of the compound was obtained in a hydrolysis reaction with potassium hydroxide. The salt was then converted to an acid chloride with oxalyl chloride. The acid chloride was subject to an esterification reaction with an excess of hydroquinone. The obtained monoester product was then used as a main substrate for synthesis of L and L-L. To obtain L-L, the Mitsunobu reaction with 1,12-dodecanediol was performed. To obtain L, Mitsunobu reaction with the previously obtained alkyl spacer (S) was conducted. Finally, the bromide atom was substituted with a mercapto group.

**HNF@Au@L Sample Preparation.** In a typical process, 5  $\mu\text{L}$  of functionalized Au@L toluene solution (2 mg/mL) was added to 5  $\mu\text{L}$  of L-L solution (1 mg/mL) and drop casted on a substrate, usually a TEM grid ( $\sim 3 \mu\text{L}$ ) or an ITO-covered glass slide ( $\sim 5 \mu\text{L}$ ). Subsequently, the sample was heated to 140 °C and cooled to 120 °C with a rate  $\sim 40$  °C/min and then to 30 °C with a precisely controlled rate of 3 °C/min.

**FDTD Simulations.** Extinction spectra were simulated for HNF@Au@L under LCP and RCP irradiation. HNF parameters were chosen as follows: 14 NPs per pitch, 290 nm pitch length, and 64 nm thickness (measured from center to center of the NPs), which translates into an interparticle separation of  $\sim 3.0$  nm, in good agreement with the distances measured for the Au@L assemblies. Circular dichroism was calculated as the difference between extinction in RCP and extinction in LCP. The refractive index around the NPs was set to 1.5, as this is a common value for liquid crystal compounds.<sup>83</sup> The perfectly matched layer was set as a boundary condition in all directions. The helical nanofilament direction was perpendicular to incident light. Dielectric functions describing gold were approximated using Johnson and Christy data.<sup>84</sup> All simulations were carried out using Lumerical software.

**Methods.** For structural analysis of the obtained materials, transmission electron microscopy was used: TEM model JEM-1400 (JEOL, Japan), available at the Nencki Institute of Experimental Biology, laboratory of electron microscopy, and TEM model JEM-1011 (JEOL) equipped with a model EDS INCA analyzer (Oxford, U.K.), in the Electron Microscopy Platform, Massakowski Medical Research Centre, Polish Academy of Science, Warsaw. Scanning electron

microscopy analysis was performed using the Quanta 250, FEG, FEI standard Environmental SEM, available at Wielkopolska Centre For Advanced Technologies. Spectroscopy studies in the UV–vis range were performed using GENESYS 50 UV–vis spectrophotometer, available at the University of Warsaw. In order to determine nanoflament handedness, atomic force microscopy was used. The Bruker Dimension Icon microscope (Billerica, U.S.A.), available at the Biological and Chemical Research Centre, University of Warsaw, was used. The chiral discrimination process was controlled using polarized optical microscopy, Zeiss Axio Imager Am.2 microscope, available at University of Warsaw. Bulk CD measurements were performed using a Chirascan Circular Dichroism Spectrometer, available at University of Warsaw. The cooling–heating process of the obtained samples was controlled using the FTIR600 SP Linkam stage with T96 LinkPad system controller, available at the University of Warsaw. Thermogravimetric measurements were performed using a TGA Q50 by TA Instruments.

## ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acsnano.0c03964>.

Additional figures and notes including linear dichroism, CD imaging microscopy, synthetic routes, thermogravimetric analysis, process optimization, histograms, TEM images, optical microscope images, AFM height map, UV–vis spectrum, 2D AFM elevation profiles, POM-CD method, and circular dichroism spectra (PDF)

Timelapse of HNF@Au@L crystallization (AVI)

Raw micro-CD data (ZIP)

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### Author Contributions

W.L. initiated the project; L.M.L.-M. and W.L. coordinated the work; N.K. performed nanoparticles synthesis, M.B. and M.G. performed organic syntheses; P.S. and N.K. obtained HNF@Au@L and performed TEM, SEM, OM, UV–vis, and bulk-scale CD measurements; W.L. and P.S. performed TGA measurements; D.G. performed AFM measurements; T.N., D.G., and H.O. performed DMCPL measurements; P.S. designed and implemented POM-CD and the chiral discrimination procedure, performed FDTD simulations, and analyzed the data; D.P. helped with optical measurements; P.S. and N.K. prepared the figures; W.L. and P.S. wrote the manuscript draft; and all authors discussed and interpreted the results as well as commented on the manuscript.

### Notes

The authors declare no competing financial interest.

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