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Genome-wide association study implicates *CHRNA2* in cannabis use disorder

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D.D. and A.D.B. conceived the idea, supervised and directed the study. T.E.T., T.T., V.R., J.B.-G., M.B.-H., A.T., E.A., D.M.H., T.W., O.M., P.B.M., M.No., M.J.D., H.S., K.S., A.D.B. provided samples and/or data. D.D., V.M.R., T.E.T., T.D.A., J.G., K.L., D.F.G., J.P., G.W.R., L.M.H., E.A.S. performed the analyses. D.D., V.M.R., T.E.T., T.D.A., J.G., K.L., D.F.G., C.H., J.H.C., H.S., M.Ny, ADB interpreted the results. D.D. wrote the manuscript. D.D., T.E.T., J.G., K.L., D.F.G., H.S., A.D.B. comprised the core revision group. All authors discussed the results and approved the final version of the manuscript.

Competing Interests Statement

T. Werge has been a lecturer and advisor to H. Lundbeck A/S.

T. E. Thorgeirsson, D. F. Gudbjartsson, G. W. Reginsson, H. Stefansson and K. Stefansson are employees of deCODE genetics/ Amgen.

Data availability

Summary statistics with the results from the CUD GWAS is available on the iPSYCH website (<https://ipsych.au.dk/downloads/>). For access to genotypes from the the iPSYCH cohort, interested researchers should contact A.D. Børglum.

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Abstract

Cannabis is the most frequently used illicit psychoactive substance worldwide, and around one in ten users become dependent. The risk for cannabis use disorder (CUD) has a strong genetic component, with twin heritability estimates ranging from 51-70%. Here we performed a genome-wide association study (GWAS) of CUD in 2,387 cases and 48,985 controls, followed by replication in 5,501 cases and 301,041 controls. We report the first genome-wide significant risk locus for CUD ($P=9.31 \times 10^{-12}$) that replicates in an independent population ($P_{\text{replication}}=3.27 \times 10^{-3}$, $P_{\text{metaanalysis}}=9.09 \times 10^{-12}$). The index variant (rs56372821) is a strong eQTL for *CHRNA2* and analyses of the genetically regulated gene expression identified a significant association of *CHRNA2* expression with CUD in brain tissue. At the polygenic level, analyses revealed a significant decrease in the risk of CUD with increased load of variants associated with cognitive performance. The results provide novel biological insights and inform on the genetic architecture of CUD.

Introduction

Cannabis use is increasing in the population¹ and around 9% of users become dependent². Genetic factors explain a large proportion of the risk for cannabis use disorder (CUD) with twin heritability estimates in the range of 51³-70%⁴. The prevalence of diagnosed CUD in the population has been estimated to 1-1.5% among Europeans⁵ and Americans¹. CUD is associated with a range of adverse health problems⁶ including risk of psychosis⁷, bipolar disorder⁸, anxiety disorder⁹ and cognitive impairment with more persistent use associated with greater decline¹⁰. Estimates of the heritability for cannabis use initiation and life-time cannabis use, with respect to the amount of variance explained by common variants (i.e. the single nucleotide polymorphism (SNP) heritability), has been estimated to 0.06¹¹-0.2¹². Four GWASs related to cannabis use initiation or lifetime use have been conducted; three studies without genome-wide significant findings¹¹⁻¹³ and a large GWAS meta-analysis of life-time cannabis use including 184,765 individuals, which identified eight genome-wide significant independent risk loci¹⁴. Four GWASs of problematic use or diagnosed CUD have been conducted; two without genome-wide significant findings^{15,16} and two recent reporting genome-wide significant associations, albeit with negative or ambiguous replication results: a study of DSM-IV cannabis dependence criterion counts in a combined sample of 14,754 European Americans and African Americans¹⁷, which reported three genome-wide significant loci associated with cannabis use severity; and a GWAS of cannabis dependence of 2,080 European cases and 6,435 controls which identified one genome-wide significant locus¹⁸.

Here we present results from a GWAS and subsequent replication based on analyses of the largest cohorts of diagnosed CUD reported so far. Individuals included in the discovery GWAS come from the Danish nation-wide population based cohort collected by the Lundbeck Foundation Initiative for Integrative Psychiatric Research (iPSYCH). The iPSYCH cohort was ascertained to study six major psychiatric disorders (schizophrenia, bipolar disorder, major depressive disorder, attention-deficit hyperactivity disorder (ADHD), anorexia nervosa and autism spectrum disorder) and consists of 79,492 genotyped individuals. The present GWAS included 2,387 individuals with a diagnosis of CUD. In this study CUD was defined by individuals having an ICD10 diagnosis reflecting a problematic and persistent use of cannabis (ICD10 F12.1-12.9; see Supplementary Table 1, 2 and 3 for information on number of cases in each diagnostic subcategory and distribution over diagnostic subcategories in relation to comorbid psychiatric disorders) and 48,985 individuals not diagnosed with CUD, all from the iPSYCH cohort. We identify a genome-wide significant risk locus on chromosome 8, which replicates in an independent cohort from deCODE genetics. The index variant is a strong eQTL for *CHRNA2*, which encodes the neuronal acetylcholine receptor (nAChR) alpha-2 subunit, and in analyses of the genetically regulated gene expression we find reduced expression of *CHRNA2* in brain tissue in individuals with CUD compared to the control group in cerebellum. At the polygenic level, we show that increased load of CUD variants associates with decreased cognitive performance and increased risk of schizophrenia and ADHD.

Results

GWAS of CUD and replication in an independent cohort from Iceland

Data analysis was conducted using the Ricopili pipeline¹⁹, including stringent quality control of genotyped variants and individuals (Methods). Information about non-genotyped markers was obtained by imputation using the 1000 genomes phase 3 as reference panel. GWAS was performed using imputed marker dosages and an additive logistic regression model using the following covariates: relevant principal components to correct for confounders such as population stratification, presence of the major psychiatric disorders studied in iPSYCH and genotyping wave. Only markers with an imputation INFO score >0.7, minor allele frequency (maf) >0.01 and bi-allelic variants were retained, in total 8,971,679 genetic variants were included. We identified 26 genome-wide significant variants ($P < 5 \times 10^{-8}$), located in a single locus on chromosome 8 (Figure 1 and 2). The index variant, rs56372821, showed an odds ratio of 0.73 ($P = 9.31 \times 10^{-12}$) with respect to the minor allele A (Table 1, Figure 1 and 2). We found no signs of contributions from confounding factors like population stratification and cryptic relatedness to the inflation in the distribution of the test statistics ($\lambda = 1.033$; see quantile-quantile plot, Figure 1.B.) using LD score regression, since the intercept was practically one (intercept = 0.996; SE = 0.0079) (Methods). There was no evidence of association of previously identified genome-wide significant cannabis use risk variants with CUD in our analyses^{17,18} (Supplementary Table 5). This might be due to different phenotype definitions among the studies as Sherva et al.¹⁷ analysed association with cannabis use criterion counts, and Agrawal et al.¹⁸ used cannabis exposed (but not dependent) individuals as controls in their study. Additionally, the composition of the cohorts analysed also differ as the previous GWASs were based on cohorts established to

study substance use disorders while the iPSYCH cohort was ascertained for major mental disorders.

The genome-wide significant locus on chromosome 8 was replicated in an independent Icelandic sample consisting of 5,501 cases with CUD and 301,041 population controls (deCODE cohort). The cases were diagnosed with CUD while undergoing inpatient treatment at the SSA National Center of Addiction Medicine, Vogur Hospital in Iceland (www.saa.is). SAA treats around ~80% of individuals with substance abuse in Iceland which makes the deCODE CUD cohort nearly population-wide²⁰. We tested nine variants located in the risk locus in the Icelandic sample; the index variant and eight correlated variants ($0.2 < r^2 < 0.7$) with P-values less than 1×10^{-6} (four genome-wide significant). All variants demonstrated consistent direction of association. The most strongly associated variant (rs56372821) in the discovery GWAS showed a P-value of 3.27×10^{-3} in the deCODE cohort, however with a slightly smaller effect size (Table 1). In the meta-analysis, rs56372821 showed a little stronger association with CUD ($P=9.09 \times 10^{-12}$), and additional two variants became genome-wide significant (Table 1).

We performed polygenic risk score (PRS) analysis in order to evaluate the consistency between the deCODE and the iPSYCH CUD phenotypes. When using the iPSYCH cohort as discovery, we observed a significant association of CUD PRS in the deCODE cohort with CUD (e.g P-value= 2.17×10^{-9} ; P-value threshold < 0.01 in the discovery GWAS) but not with schizophrenia or bipolar disorder (Supplementary Table 4), supporting that the associations in the iPSYCH cohort primarily are driven by CUD and not the comorbid psychiatric disorders.

To assess the proportion of phenotypic variance in the iPSYCH data explained by common variants we applied LD score regression and the GREML method implemented in GCTA, assuming a population prevalence of 1% for cannabis use disorder (Methods). Estimates of the liability-scale SNP heritability were $h^2_{\text{snp}}=0.09$ (SE=0.03) and $h^2_{\text{snp}}=0.042$ (SE=0.014; $P=0.0012$) using LD score regression and GCTA, respectively. This result suggest a smaller contribution from common variants than was previously found for cannabis use¹². The estimate will probably increase with larger sample size as a result of a decrease in the size of the error terms of the SNP effect estimates²¹.

The role of the CUD risk locus in other psychiatric disorders

In the GWAS we corrected for diagnoses of the major psychiatric disorders studied in iPSYCH (see Supplementary Table 1 for distribution of psychiatric disorders among CUD cases and the control group). In addition, we evaluated the impact on the odds ratio for the index variant by leave-one-out analyses excluding psychiatric phenotypes one at a time in the association analysis (Methods). The odds ratio remained stable (Table 2), supporting that the association was independent of a diagnosis of the psychiatric disorders evaluated.

The risk locus on chromosome 8 is also genome-wide significantly associated with schizophrenia¹⁹. In our leave-one-phenotype out analysis the locus remained genome-wide significant when individuals with schizophrenia were removed, excluding any potential confounding from schizophrenia (Table 2). Additionally, we tested the possibility of the

locus being broadly associated with psychiatric disorders by performing a cross disorder GWAS including 35,006 individuals with one of the five major psychiatric disorders and 16,366 controls. In this analysis the index variant (rs56372821) showed a P-value of 0.058 (OR=0.966). When redoing the GWAS while excluding all individuals with CUD, the analysis revealed a cross-disorder P-value=0.33, OR=0.982 for rs56372821 (32,716 cases and 16,269 controls). The close to nominal significant signal observed in the cross-disorder GWAS including all cases disappeared when excluding individuals having a CUD diagnosis. This reinforces that the association signal in the CUD GWAS is caused by CUD, and not by a general association with psychiatric disorders.

The signal observed for the index variant in the GWAS meta-analysis of schizophrenia¹⁹ (rs73229090) is consistent with the direction of association observed in our analysis. Since individuals with schizophrenia often use cannabis (around 13-16%²²) it could be speculated that the significant signal observed in schizophrenia is driven by a sub-group of schizophrenia cases also having CUD. This hypothesis is supported by analysis of the association of rs56372821 with schizophrenia in the iPSYCH sample. We found a nominal significant association of the variant with schizophrenia when including individuals with comorbid CUD (2,281 schizophrenia cases and 23,134 controls; OR=0.9; P=0.036), while after exclusion of individuals with CUD (556 cases and 101 controls excluded) the association signal disappeared (1,727 cases, 23,033 controls; OR=0.97, P=0.63). To further to evaluate the impact of CUD comorbidity on the odds ratio of rs56372821 in schizophrenia a null distribution for the odds ratio was generated by performing 10,000 permutations randomly removing 556 and 101 individuals among the cases and controls, respectively (Supplementary Figure 1). The observed odds ratio for rs56372821, when excluding individuals with comorbid CUD was 0.97, and differed significantly from random removal of the same number of cases and controls ($P_{\text{two-sided}}=0.0027$, $P_{\text{one-sided}}=0.0015$). Thus the permutation test supports the hypothesis that the subgroup diagnosed with CUD among schizophrenia cases drives the nominal association observed for rs56372821 in the iPSYCH schizophrenia sample.

Association of risk locus with age at first CUD diagnosis

We performed a case-only analysis testing all variants in LD with the index variant (19 SNPs; $r^2 > 0.7$) for association with age at first CUD diagnosis. This analysis suggested the risk alleles to be associated with earlier age at first diagnosis (most significant SNP: rs35236974, P=0.020). On average CUD cases homozygous for the protective allele got their diagnosis (mean age: 22.41; st.dev=3.63) one year later than CUD cases having at least one risk allele (mean age: 21.01; st.dev=3.56).

Transcriptome-wide association study implicates *CHRNA2* in CUD

Among the brain tissues analysed in GTEx (<https://gtexportal.org/home/>) the index variant rs56372821 was found to be a strong eQTL for *CHRNA2* in cerebellum (P-value in GTEx=2.1x10⁻⁷), with the risk allele (rs56372821-G) being associated with decreased expression of the gene (Supplementary Figure 2). *CHRNA2* is expressed in the brain and encodes the neuronal acetylcholine receptor (nAChR) alpha-2 subunit, which is incorporated in heteropentameric neuronal nAChRs mainly with the beta-2 or beta-4 subunits²³.

Candidate gene studies of common variants have linked this gene to e.g. substance abuse²⁴ and nicotine dependence²⁵ but no genome-wide significant findings have connected *CHRNA2* to any substance abuse or psychiatric disorder, besides a functional link between schizophrenia risk variants and *CHRNA2* expression identified by Won et al²⁶ (described below).

To further evaluate the potential regulatory impact of the identified locus on chromosome 8 as well as genome-wide gene expression differences between cases and controls, we imputed the genetically regulated gene expression in 11 brain tissues using PrediXcan (Methods). The SNP weights used were derived from models trained on reference transcriptome data sets including 10 brain tissues from GTEx and transcriptome data from dorsolateral prefrontal cortex generated by the CommonMind Consortium. We tested for association of expression of 2,460-10,930 protein coding genes depending on the tissue (Supplementary Table 6) with CUD using logistic regression corrected by the same covariates as in the GWAS. One gene, *CHRNA2*, was significantly differently expressed between cases and controls ($P=2.713 \times 10^{-6}$; $\beta=-0.21$; $SE=0.045$) in the cerebellum. The expression model for *CHRNA2* in cerebellum was based on 47 SNPs including four genome-wide significant SNPs rs59724122, rs73229093, rs7838316 and rs11783093 (Figure 3). *CHRNA2* expression was predicted with a valid model in two other brain tissues showing nominal significant under-expression in cases compared to controls (dorsolateral prefrontal cortex, $P=5.19 \times 10^{-4}$; cerebellar hemisphere $P=5.30 \times 10^{-3}$). That the risk locus for CUD can be linked to *CHRNA2* expression is also supported by Won et al.²⁶ who generated high-resolution 3D maps of chromatin contacts (by Hi-C sequencing approach) to capture functional relationships between regulatory and transcribed elements during human corticogenesis. They applied their map to a set of credible SNPs from the large PGC schizophrenia GWAS¹⁹ and found physical interaction of the genome-wide significant risk locus on chromosome 8 with the regulatory region of *CHRNA2*.

Examination of smoking behavior as a confounder

CHRNA2 is related to other nicotine receptor genes that have been identified as risk loci for nicotine dependence and smoking behavior²⁷. It is therefore relevant to question if our finding is confounded by smoking. In the iPSYCH cohort there is no available information about smoking behavior. As much as 70-90% of cannabis users are reported also to smoke cigarettes²⁸, which impose the risk of confounding. However, smoking is also highly comorbid with psychiatric disorders²⁹, which is prevalent among the control group (66,79% were diagnosed with at least one of the major psychiatric disorders studied in iPSYCH; Supplementary Table 1). Since we expect smoking to be prevalent both among cases and the control group, it is unlikely that smoking alone would generate the high effect size we observe for the risk locus on chromosome 8. However, in the recent GWAS meta-analysis of smoking³⁰ including more than 1.2 mill people the CUD risk locus is genome-wide significantly associated with smoking, with the CUD risk allele increasing risk for smoking initiation, younger age at smoking initiation and continuation of smoking, but it is associated with decreasing number of cigarettes per day (Supplementary Table 7). The strongest association is observed for smoking initiation with an effect size of $OR=0.954$ (rs11783093), which is significantly lower than the effect size observed for CUD in iPSYCH ($OR=0.73$

(rs56372821); $P=7.964873 \times 10^{-09}$; $Z=5.767$) and smaller than in deCODE (OR=0.88; $P=0.061$; $Z=1.87$). This suggests that the locus has a pleiotropic effect on the risk of both smoking and CUD in the general population, with greatest impact on CUD risk. In order to rule out that smoking behavior is not the main factor driving the association signal on chromosome 8 in the present study, we performed several additional analyses.

I) First the results from the CUD GWAS were adjusted by conditioning on the effects of a set of independent associated variants with smoking initiation³⁰ using mtCOJO (Methods). The results, which are robust to confounding by shared genetic and environmental factors with smoking, supports that the CUD association is not strongly confounded by smoking (OR_{corrected}=0.759; $P=4.44 \times 10^{-09}$, Supplementary Table 8).

II) Furthermore, we evaluated by elementary simulation analysis how likely it would be to observe an odds ratio of 0.728 if the signal was primarily driven by tobacco smoking, by assuming a true odds ratio of smoking = 0.954³⁰. We assigned plausible values for the prevalence of tobacco smoking among minor and major allele carriers of rs5637282 as well as CUD among tobacco users and non-users, and empirically approximated the probability of observing an odds ratio of 0.728. The largest P-value encountered was 8.69×10^{-3} (2¹² scenarios were tested), supporting that it is very unlikely that the OR we observe for CUD could be caused by tobacco smoking alone (Methods and Supplementary Table 9).

III) We performed a GWAS of CUD cases against schizophrenia cases in the iPSYCH cohort (2,387 CUD cases and 1,789 individuals with schizophrenia (ICD10 F20) without a CUD diagnosis). Smoking rates among schizophrenia cases is in the range of 60-90%³¹, so comparable with the rates among CUD cases. By comparing the two groups with comparable smoking rates we would expect the association signal to disappear if smoking is the driving factor. The index variant rs56372821 was associated with CUD with $P=1.68 \times 10^{-5}$, OR=0.76. The observed strong association with only a slight increase in OR from 0.73 in the original CUD GWAS to 0.76 supports that the signal is primarily driven by CUD and not confounded by tobacco smoking.

IV) Finally, no strong confounding of tobacco smoking at the polygenic level in the CUD GWAS was observed. This was evaluated using the iPSYCH CUD GWAS as training for generating CUD-PRS in deCODE. The score explained more of the variance and was associated with a higher effect size for CUD risk ($R^2=0.24\%$; OR=1.17) than was found for tobacco smoking ($R^2=0.10\%$, OR=1.07) (Supplementary Table 10).

In summary all four approaches support that the results reflect association with CUD without a strong confounding from smoking, both with respect to the identified CUD risk locus and at the polygenic level.

Genetic overlap with other phenotypes

To evaluate the genetic overlap between CUD and a range of other phenotypes at the polygenic level, we conducted analyses of PRS for 22 phenotypes related to cognition, personality, psychiatric disorders, reproduction and smoking behaviour (Methods; list of references to the GWASs of the 22 phenotypes and results can be found in Supplementary

Table 11). PRS for eight phenotypes (three measures of cognitive performance, age at first birth, life time smoking, ADHD, depressive symptoms and schizophrenia) demonstrated strong association with CUD ($4.33 \times 10^{-4} < P < 7.44 \times 10^{-15}$; Figure 4 and Supplementary Table 11). Strikingly, PRS for measures of educational attainment by the Social Science Genetic Association Consortium (SSGAC) (educational years: $z\text{-score} = -7.78$; $P = 7.44 \times 10^{-15}$; college completion: $z\text{-score} = -5.10$; $P = 3.33 \times 10^{-7}$), were strongly negatively associated with the risk of CUD, a finding which was reinforced by the significant negative association of PRS for human intelligence with CUD ($z\text{-score} = -3.51$; $P = 4.33 \times 10^{-4}$). Our results suggest an overlap in genetic risk factors with a decrease in the odds ratio for CUD with increased number of educational years/cognitive performance (Supplementary Figure 3A-C.). The observed decreased risk of CUD with increased age at first birth ($z\text{-score} = -7.41$; $P = 1.26 \times 10^{-13}$) is supporting the relationship with educational attainment as number of educational years is known to correlate genetically with later birth of the first child³². In order to avoid confounding in the PRS analyses of psychiatric disorders we excluded individuals among CUD cases and the control group with a diagnosis of the psychiatric disorder being analysed. PRS for ADHD ($z\text{-score} = 5.10$; $P = 3.45 \times 10^{-7}$), depressive symptoms ($z\text{-score} = 4.34$; $P = 6.58 \times 10^{-6}$) and schizophrenia ($z\text{-score} = 5.47$; $P = 4.45 \times 10^{-8}$) were all associated with an increased risk for CUD (Figure 4).

Discussion

This is the first study to report a genome-wide significant risk locus, for diagnosed CUD replicated in an independent cohort. The locus is located intergenic on chromosome 8 (index variant rs56372821) and has an odds ratio of 0.73, an effect size in line with findings for other complex disorders^{19,33}. PRS supported a high consistency of the CUD phenotypes analysed in the discovery and replication cohorts from iPSYCH and deCODE, respectively, and supported that the GWAS results reflect association with CUD and not comorbid psychiatric disorders in the iPSYCH cohort. This also holds for the risk locus, where careful dissection with respect to the distribution of effect sizes for the index variant (rs56372821) over co-morbid subgroups revealed no confounding from psychiatric disorders. This included exclusion of individuals with schizophrenia, which still revealed genome-wide significant association of rs56372821 with CUD. This supports the proposed hypothesis that the genome-wide significant signal observed for the CUD risk locus in schizophrenia¹⁹ could be driven by a sub-group of schizophrenia cases having CUD.

Analysis of the imputed genetically regulated gene expression suggested *CHRNA2* underexpression in cerebellum (and probably other brain regions), to be involved in CUD. Cerebellum may play a role in addiction with respect to reward and inhibitory control³⁴, and reward-anticipating responses³⁵. Cerebellum has high density of cannabinoid receptor 1 (CB1), which mediates the effect of delta9-tetrahydrocannabinol (THC), the psychoactive compound in cannabis and has also been found to be affected by cannabis use in neuroanatomic studies^{e.g.}³⁶.

CHRNA2 encodes the neuronal acetylcholine receptor (nAChR) alpha-2 subunit, and there is no reported functional link between cannabis use and alpha-2 subunit containing nAChRs. We hypothesize three potential ways of the involvement of *CHRNA2* in CUD: 1) Substances

in cannabis might interact directly with the alpha-2 subunit containing nAChRs as studies have found cannabidiol, a non-psychoactive component of cannabis, to inhibit the alpha-7 containing nAChRs³⁷. 2) Cannabis could indirectly affect the alpha-2 subunit containing nAChRs. After binding of an agonist (e.g. acetylcholine), the nAChR responds by opening of an ion-conducting channel across the plasma membrane. This causes depolarization of the membrane and can result in presynaptic neurotransmitter release including dopamine³⁸. Since the psychoactive active compound of cannabis THC has been found to affect the release of acetylcholine in various brain regions³⁹ it could be speculated that this, through alpha-2 subunit containing nAChRs, could affect dopamine release, a known neurotransmitter involved in addiction. 3) There could be a strong biological link between expression of *CHRNA2* and the cannabinoid receptor 1 gene (*CNR1*). This hypothesis is based on evaluation of gene-expression correlations from genome-wide microarray gene expression profiles in the Allan Brain Atlas (<http://www.brain-map.org/>). We found that of all genes evaluated (58,692 probes analyzed), *CNR1* demonstrated the strongest negative correlation with *CHRNA2* expression ($r_{\max} = -0.498$; Supplementary Figure 4). The signal was driven by opposite expression patterns in a large number of brain tissues, e.g. in cerebellum where *CNR1* had a relatively high expression in the cerebellar cortex and *CHRNA2* had a relatively low expression and the opposite was observed for cerebellar nuclei (Supplementary Figure 4). This suggests the existence of a currently uncharacterized biological interaction between the endocannabinoid system and alpha-2 subunit containing nAChRs, by which the identified risk locus which is associated with decreased *CHRNA2* expression could be related to increased *CNR1* expression.

The observed association of *CHRNA2* underexpression with CUD, also implies that the alpha-2 subunit can be a potential drug target in the treatment of CUD, by the use of an agonist selective for alpha-2 subunit containing nAChRs. The impact of some compounds with an agonistic effect on the alpha-2 subunit containing nAChRs, like NS9283⁴⁰, ABT-418⁴¹ and TC-1734⁴², have already been studied primarily due to their potential impact on memory and cognitive processes^{e.g. 40,42}.

Since *CHRNA2* and other nicotine receptor genes that have been identified as risk loci for nicotine dependence and smoking behavior^{27,30}, the potential confounding from smoking was evaluated carefully, and all our approaches supported that the results reflect CUD without strong confounding from smoking. Furthermore our results suggested that the identified CUD risk locus, which is also genome-wide significantly associated with smoking³⁰ however with significant lower effect size than observed for CUD, has a pleiotropic effect on both smoking and CUD in the general population, with greatest impact on CUD risk. Additionally a recent GWAS of lung cancer identified a genome-wide significant locus just upstream *CHRNA2*⁴³. This locus is LD distinct from the locus identified in the present study, but the risk allele was also found to be associated with decreased *CHRNA2* expression in cerebellum. In analyses of smoking behavior, the lung cancer risk allele, was reported nominally associated with smoking and age of initiation⁴³. This result is in line with our observation of the CUD risk variant, and thus decreased *CHRNA2* expression, being associated with younger age at first diagnosis of CUD.

Finally it should be mentioned that we cannot exclude other relevant genes in the region like *EPHX2*. This gene encodes soluble epoxide hydrolase involved in xenobiotic metabolism by degrading potentially toxic epoxides. While *EPHX2* is not one of the main metabolizers of THC, some involvement is possible, as soluble epoxide hydrolase may act on at least some endocannabinoid metabolites and metabolites of 8-THC⁴⁴.

At the polygenic level, increased load of variants associated with measures of decreased cognitive performance was associated with CUD. This suggests an underlying genetic component causing the observations of decreased educational performance being related to cannabis use in numerous epidemiological studies^{e.g.}⁴⁵. The direction of causation is unclear and it is unknown if the relationship is caused by cannabis use and subsequent disengagement from education or whether and to what extent the two share overlapping genetic factors⁴⁶.

The overlap in common genetic risk factors for cannabis use disorder with depressive symptoms is in line with recent observations⁴⁷ and the association of CUD with increased load of ADHD risk variants suggests that the epidemiological observations of high comorbidity of ADHD with CUD⁴⁸ are influenced by shared genetic risk components. Likewise, the overlap in common genetic risk variants between CUD and schizophrenia (in line with epidemiological observations^{22,49} and PRS studies⁵⁰). However we cannot rule out whether the overlap involves variants with pleiotropic effects, or if the overlap in general is caused by heterogeneity among schizophrenia cases due to a subgroup comorbid with CUD, in line with the CUD risk locus identified here.

In summary, we identified a genome-wide significant CUD locus, which replicated in an independent sample. An impact of the locus on earlier age of first diagnosis was further suggested. *CHRNA2* was implicated in CUD through analyses of the imputed genetically regulated gene expression, showing decreased *CHRNA2* expression in cerebellum (and other brain regions) of individuals with CUD. An intriguing result, which suggests further studies to evaluate if *CHRNA2* could be a potential therapeutic target for CUD treatment. The identified locus is located in a region previously associated with schizophrenia¹⁹, smoking³⁰ and lung cancer⁴³. Our GWAS result with respect to the identified risk locus did not seem to be confounded by either smoking or psychiatric disorders. Larger GWASs and functional studies are needed in order to further explain the biological mechanisms underlying the phenotypic associations observed for this region.

Methods

Samples

The individuals in this study is a part of the large Danish nation-wide population based sample collected by iPSYCH⁵¹. The iPSYCH sample was originally designed for studying major psychiatric disorders and includes genotypes from 79,492 individuals, including 54,249 cases diagnosed with at least one of six mental disorders (schizophrenia, bipolar disorder, major depressive disorder, ADHD, anorexia and autism spectrum disorder) and 25,243 individuals not having any of the six psychiatric disorders. The iPSYCH sample was selected from a birth cohort comprising individuals born in Denmark between May 1, 1981,

and December 31, 2005, who were residents in Denmark on their first birthday and who have a known mother (N =1,472,762). The iPSYCH cases were identified based on information in the Danish Psychiatric Central Research Register⁵², and 30,000 randomly selected controls were identified from the same nationwide birth cohort in the Danish Civil Registration System⁵³. Subsequently blood spot samples (Guthrie cards) were identified in the in the Danish Newborn Screening Biobank (DNSB) from the included individuals⁵⁴. Processing of DNA, genotyping and genotype calling as well as imputing of genotypes (see below) of the samples were done in 23 waves of approximately 3,500 individuals each.

Individuals with CUD were identified in the iPSYCH sample through information in the Danish Psychiatric Central Research Register⁵² as well as in the Danish National Patient Register⁵⁵ using information up to 2013. All individuals with an ICD10 diagnosis of F12.1-12.9, were included (Supplementary Table 1 and 2). Individuals with acute intoxication (diagnosis code F12.0) were not included among cases (41 individuals had an F12.0 diagnosis; 0.084% of the controls). The F12.0 diagnosis is related to the acute pharmacological effects of cannabis use and resolve with time, and does not necessary reflect long term problematic cannabis use, which is the focus of this study. All individuals in the iPSYCH sample not having a diagnosis of CUD were used as controls and covariates for the major psychiatric disorders studied by iPSYCH were used in the analyses in order to correct for the presence of mental disorders. Only 54 individuals had a diagnosis of CUD and anorexia, and all individuals having a diagnosis of anorexia among cases and the control group were excluded. After QC the study included 2,387 cases and 48,985 in the control group. Supplementary Table 1 gives an overview of the distribution of individuals with a psychiatric disorder among the CUD cases and the control group. None of the comorbid psychiatric disorders deviated with respect to the distribution of individuals over CUD diagnosis sub-categories (Supplementary Table 3).

Processing of samples, genotyping, QC and imputing was done in 23 waves. In this study individuals from the first four waves were excluded as only a few cannabis use disorder cases were present in these waves (range 0-15 cases). The waves represent approximate birth years and the four waves that were excluded represent to a large extend individuals, which in 2013 (the time of extraction of register diagnoses) were children. Mean age of cases is 24,77 years (standard deviation =5.32) and mean age of controls is 22,67 years (standard deviation =3.86). The control group is younger and the age span wider than for the cases. The presence of younger individuals among the controls could reduce power since some controls eventually will become cases, and it also reduces the risk of discovering a false positive association.

The study was approved by the Danish Data Protection Agency and the Scientific Ethics Committee in Denmark.

Statistical analyses

Genotyping, quality control and GWAS—DNA was extracted from dried blood spot samples and whole genome amplified in triplicates as described previously^{56,57}. Genotyping was performed at the Broad Institute of Harvard and MIT (Cambridge, MA, USA) using Illumina's Beadarrays (PsychChip; Illumina, CA, San Diego, USA) according to the

manufacturer's protocols. Genotypes were a result of merging call sets from three different calling algorithms (GenCall, Birdseed and Zcall). GenCall⁵⁸ and Birdseed⁵⁹ were used to call genotypes with minor allele frequency (maf) > 0.01 and zCall⁶⁰ was used to call genotypes with maf < 0.01. The merging was done after pre-QC on individual call sets.

Stringent quality control was applied and only samples with individual call rate (> 0.98) and genotypes with high call rate (> 0.98), no strong deviation from Hardy-Weinberg equilibrium ($P > 1 \times 10^{-6}$ in controls or $P > 1 \times 10^{-10}$ in cases) and low heterozygosity rates ($|F_{\text{het}}| < 0.2$) were included. Genotypes were phased and imputed using the 1000 Genomes Project phase 3 (1KGP3)^{61,62} imputation reference panel and SHAPEIT⁶³ and IMPUTE2⁶⁴. Relatedness and population stratification were evaluated using a set of high quality markers (genotyped autosomal markers with minor allele frequency (maf) > 0.05, HWE $P > 1 \times 10^{-4}$ and SNP call rate > 0.98), which were pruned for linkage disequilibrium (LD) ($r^2 < 0.075$) resulting in a set of 37,425 pruned markers (markers located in long-range LD regions defined by Price et al.⁶⁵ were excluded). Genetic relatedness was estimated using PLINK v1.9^{66,67} to identify first and second-degree relatives ($\hat{\pi} > 0.2$) and one individual was excluded from each related pair (cases preferred kept over controls). Genetic outliers were identified for exclusion based on principal component analysis (PCA) using EIGENSOFT^{68,69}. A genetic homogenous sample was defined based on a subsample of individuals being Danes for three generations (identified based on register information about birth country of the individuals, their parents and grandparents). The subsample of Danes was used to define the center based on the mean values of principal component (PC) 1 and PC2. Subsequently PC1 and PC2 were used to define a genetic homogenous population by excluding individuals outside an ellipsoid with axes greater than six standard deviations from the mean. After outlier exclusion PCA was redone and PCs from this analysis were included in the association analysis (see below).

Association analysis was done using logistic regression and the imputed marker dosages. The following covariates were used: principal component 1-4 and principal components from the PCA associated with case-control status, the 19 data-processing waves and diagnosis of major psychiatric disorders studied by iPSYCH (Supplementary Table 1). Results for 9,729,295 markers were generated, subsequently markers with imputation info score < 0.7 (n=608,367), markers with maf < 0.01 (n=10,220) and multi-allelic markers (n=143,083) were removed. In total after filtering 8,969,939 markers remained for further analysis. All analyses of the iPSYCH sample were performed at the secured national GenomeDK high performance-computing cluster in Denmark (<https://genome.au.dk>).

We evaluated the impact of age on the main identified risk locus on chromosome 8. We corrected for age using age coded as a binary variable with the mean age of cases (24,77 years (standard deviation = 5.32)) as a guideline for the cut-off, grouping all individuals with age < 25 into one group (1,164 cases; 29,818 controls) and age > 24 into another group (1,223 cases; 19,167 controls). Due to the low age-span among cases it did not make sense biologically to divide the cases into smaller age-bins. Correcting for age did not have an impact on the strong association of the risk locus on chromosome 8 (rs5637282; $P = 1.082 \times 10^{-11}$).

Replication—The genome-wide significant locus on chromosome 8 was replicated in an independent European cohort consisting of 5,501 cases with diagnosed CUD and 301,041 population controls collected by deCODE genetics. The characteristics of the SAA treatment sample have been described previously²⁰ and currently diagnoses at Vogur Hospital are made using DSM-V, but most of the diagnoses in this study are based on DSM-IV or DSM-III-R. The genotypes were obtained based on SNP array data and whole genome sequences using long range phasing and two types of imputations⁷⁰. For the replication study, markers were looked up in the results of a GWAS performed by logistic regression treating disease status as the response and genotype counts as covariates. Other available individual characteristics that correlate with disease status were also included in the model as nuisance variables, using previously described methods⁷¹. The resulting P-values were corrected for inflation by the method of genomic controls (correction factor=1.42). Nine genetic variants all representing the same association signal on chromosome 8 with P-values less than 1×10^{-6} were looked up in the deCODE results. The variants in the locus were selected based on LD ($0.2 < r^2 < 0.7$). We included additional markers besides the index SNP in order to be able to evaluate the consistency of direction of association in the replication cohort over a set of variants located in the associated risk locus. The data were meta-analyzed using summary statistics and inverse variance weighted fixed effects model, implemented in the software METAL⁷². All tested variants demonstrated consistent direction of association in the replication cohort.

CUD PRS analysis in deCODE—A PRS was computed from the iPSYCH CUD GWAS summary data using the LD-pred method⁷³. Only markers with a single rs-name from the GWAS were used. Associations between the PRS and CUD (2,257 cases and 145,069 controls), schizophrenia (729 cases and 153,007 controls), bipolar disorder (1,751 cases and 151,923 controls) and smoking (46,941 individuals reporting being a regular smoker in their life and 20,602 controls that has never been regular smokers) was computed using linear regression. Covariates were sex, age and the first 10 principal components. Association P-values were corrected for inflation using LD-score regression.

Heritability—SNP heritability (h^2_{SNP}) was estimated by LD score regression⁷⁴ using summary statistics from the GWAS of CUD and pre-computed LD scores (available on <https://github.com/bulik/ldsc>). The SNP heritability was calculated on the liability scale using a prevalence of CUD of 1% in the population. The h^2_{SNP} estimated by LD score regression was inaccurate as the ratio between the estimated h^2_{SNP} and the standard error was less than four ($h^2_{\text{SNP}}/\text{SE}=2.86$). Therefore, the h^2_{SNP} was also estimated using GCTA⁷⁵. A genomic relationship matrix (GRM) between all pairwise combinations of individuals was estimated for each autosome separately and subsequently merged using the GCTA software. The strict best-guess-genotypes (i.e. 4,299,887 SNPs with info score >0.8 , missing rate <0.01 and MAF >0.01) from imputation with phase 3 of 1000 genomes were used for GRM estimation. Univariate GREML analyses in GCTA was used to estimate h^2_{SNP} on the liability scale, using the combined GRM and the same covariates as in the GWAS.

Leave-one phenotype out GWAS—In the GWAS of CUD we corrected, by covariates, for five major psychiatric disorders analysed in iPSYCH (Supplementary Table 1). However,

in order to further evaluate the potential impact on the association signal of the index variant from the psychiatric phenotypes, sensitivity analyses were performed where one phenotype at a time was excluded from the association analysis. The phenotypes evaluated were schizophrenia, bipolar disorder, major depressive disorder, attention-deficit hyper activity disorder, autism and individuals not having any of the five diagnoses. GWASs were performed as previously described excluding one of the six phenotypes one at a time (sample size for number of cases and controls is shown in Supplementary Table 1).

Cross disorder GWAS for evaluation of the general association of rs56372821 with psychiatric disorders

—In order to investigate the possibility of the risk locus at chromosome 8, being a general risk locus for psychiatric disorders we conducted a cross-disorder GWAS based on the same individuals as included in the CUD GWAS. All individuals with a diagnosis of schizophrenia (ICD10 code F20), bipolar disorder (ICD10 codes F30-F31), attention-deficit hyperactivity disorder (ICD10 code F90.0), autism spectrum disorder (ICD10 codes F84.0, F84.1, F84.5, F84.8, F84.9) and major depressive disorder (ICD10 codes F32-F33) were used as cases, in total 35,006 individuals and 16,366 individuals without any of the listed psychiatric disorders were used as controls. The GWAS was conducted using wave as covariates and PCs from PCA (generated as described for the CUD GWAS).

We re-did the cross-disorder GWAS excluding all individuals with a CUD diagnosis (removing 2,290 individuals from the case group and 97 individuals from the control group), reducing the sample size to 32,716 cases and 16,269 controls. The GWAS was conducted using the PCs from PCA and wave as covariates.

Permutation based evaluation of the odds ratio of rs56372821 for schizophrenia with and without CUD

—We applied a permutation based approach in order to evaluate how the inclusion and exclusion of individuals with CUD affected the association of our index variant rs56372821 with schizophrenia. The analyses were based on the iPSYCH sample consisting of 2,281 schizophrenia cases and 23,134 population based controls (genotyping, QC and imputation were done using the same procedures as explained above). Odds ratios were obtained by the glm() function in R using dosage data from the imputed SNP rs56372821, and including principle components PC1-PC4 and PCs significantly associated with SZ and/or CUD and genotyping wave as covariates.

In the association analysis of schizophrenia the odds ratio of rs56372821 (CI95%) was OR=0.9 (0.81-0.99). When excluding all individuals with CUD, among cases and controls (1,727 cases and 23,033 controls) the odds ratio of rs56372821 (CI95%) was OR=0.97 (0.87-1.09). To evaluate whether this change in the odds ratio is due to removal of individuals with CUD (554 cases and 101 control with CUD removed) or due to reduction in sample size, a null distribution of OR's was generated by randomly removing 554 individuals among schizophrenia cases and 101 individuals among controls (removing a total of 655 individuals). This permutation of sample exclusion was done 9,999 times to generate the null distribution of the odds ratio for rs56372821. To obtain a two-sided P-value, the observed $\ln(\text{OR}_{\text{rs56372821}}) = -0.03$ was mirrored around the mean $\ln(\text{OR}_{\text{rs56372821}})$ of the null distribution, as shown in Supplementary Figure 1. Removal of individuals with CUD (554

SZ cases and 101 controls) changed the OR of rs56372821 significantly, compared to random removal of the same number of schizophrenia cases and controls ($P_{\text{two-sided}}=0.0027$), producing a significant increase of $OR_{rs56372821}$ compared to random removal ($P_{\text{one-sided}}=0.0015$).

mtCOJO conditioning for the effect of smoking—The analysis was performed using summary statistics from the CUD GWAS and summary statistics from GWAS of smoking initiation³⁰ (excluding 23andMe data, 547,219 individuals, 11,733,344 variants). The conditional analysis was run using mtCOJO⁴⁶ (implemented in GCTA⁷⁵) using standard procedures. Variants selected for conditioning were genome-wide significant in the smoking GWAS ($P < 5 \times 10^{-8}$) and not in linkage disequilibrium ($r^2 < 0.05$) with a variant with a lower P-value (61 variants). One variant was removed due to evidence of pleiotropy (HEIDI-outlier threshold of $P = 0.01$).

Simulations analysis to evaluate confounding by tobacco smoking—A simulation analysis was conducted to evaluate how likely it is to observe an odds ratio of 0.728 if the association signal primarily is driven by smoking by assuming a true odds ratio of smoking = 0.954³⁰.

We divided individuals into six strata by assigning them a unique phenotype using the hierarchy: 1) schizophrenia (SZ), 2) bipolar disorder (BP), 3) autism spectrum disorder (ASD), 4) attention deficit hyperactivity disorder (ADHD), 5) major depressive disorder (MDD) and 6) controls (ICON). We assumed a dominant model, where homozygous and heterozygous carriers of the minor allele of rs56372821 were treated equally. See Supplementary Table 9 for distribution of individuals having the major allele or at least one minor allele among the strata.

The null hypothesis was that rs56372821 only affects CUD via its effect on tobacco smoking, which has an odds ratio 0.954 (TOR)³⁰. We assumed a prevalence of CUD among tobacco users of 0.045 (CT)⁷⁶ and a CUD prevalence among tobacco non-users of 0.01 (Ct)⁵, independently of the six groups. Given the prevalence of tobacco smoking (PRE) among one of the six groups, we could calculate the probability of tobacco smoking for minor allele carriers (Tm) and for major allele carriers (TM) among the groups, using the formulae of total probability $PRE = MAF Tm + (1 - MAF)TM$ and odds ratio $TOR = (Tm(1 - TM))/(TM(1 - Tm))$. Then we simply simulated first the tobacco use status of each individual using the probabilities Tm and TM, and further the CUD status using the probabilities CT and Ct, and computed the simulated common Cochran-Mantel-Haenszel odds ratio for CUD. We did not require the total number of simulated CUD cases per stratum to match the observed numbers, because we wanted to keep this approach as elementary as possible. We repeated the simulation 100,000 times and compared the results with the observed odds ratio 0.728 for CUD, obtaining an estimator of the P-value for the null hypothesis with variance less than $1/(4 \times 100000) = 0.000025$. This was repeated for various estimates of PRE among the six groups: a total of 2^{12} combinations from the six sets $PRE_{SZ} \in \{0.6, 0.7, 0.8, 0.9\}$, $PRE_{BP} \in \{0.3, 0.4, 0.5, 0.6\}$, $PRE_{ASD} \in \{0.05, 0.1, 0.15, 0.2\}$, $PRE_{ADHD} \in \{0.1, 0.2, 0.3, 0.4\}$, $PRE_{MDD} \in \{0.2, 0.3, 0.4, 0.5\}$ and $PRE_{ICON} \in \{0.1, 0.15,$

0.2, 0.25}. The largest P-value encountered was 8.69×10^{-3} , making the the null hypothesis very unlikely.

The simulation was also repeated including information about stratification of the six phenotypes (described above) over genotyping waves. The genotyping wave was assumed to have no effect on the various probability parameters of the simulation and this time simulations were repeated only 10,000 times. The largest P-value encountered was 4.0×10^{-4} and the variance of this estimator is less than $1/(4 \times 10000) = 0.000025$, thus rejecting the null hypothesis.

GWAS of CUD cases against schizophrenia cases—This GWAS was based on individuals in the iPSYCH cohort and compared 2,387 CUD cases (ICD10 F12.1-F12.9) against 1,789 individuals diagnosed with schizophrenia (ICD10 F20) without a CUD diagnosis. QC procedures with respect to individual and genotyping QC were the same as described of the CUD GWAS, and association was calculated using logistic regression, corrected for genotyping wave and PCs from PCA.

Association with age at first diagnosis—In order to test for a potential impact of the risk locus on age at first diagnosis, a case only study was performed testing for association of the index variant (rs56372821) and variants in LD with this ($r^2 > 0.7$; 19 variants) with age at first diagnosis. Date of diagnosis was identified from register information in the Danish Psychiatric Central Research Register⁵². Analysis for association with age at first diagnosis (natural logarithm (age at first diagnosis)) was done using linear regression and the same covariates as used in the GWAS (PCs, wave and the psychiatric disorders). A dominant model with respect to the risk allele, was applied.

Analysis of the imputed genetically regulated gene expression—The genetically regulated gene expression was imputed in 11 brain tissues using PrediXcan⁷⁷ (models downloaded from: <https://github.com/hakyimlab/PrediXcan>; version 6 data). PrediXcan was used to impute the transcriptome for cases and the control group using SNP weights derived from models trained on reference transcriptome data sets including 10 brain tissues from GTEx⁷⁸ (<https://www.gtexportal.org/home/>) and transcriptome data from dorsolateral prefrontal cortex generated by the CommonMind Consortium^{79,80}. The models were trained on 1000 genome SNPs and contained only gene expression prediction models with a false discovery rate less than 5%. Gene expression levels in the iPSYCH data were imputed wave-wise and subsequently the imputed data were merged. We tested for association of 2,459 – 10,930 protein coding genes (depending on the tissue; see Supplementary Table 6) with CUD using logistic regression. The analysis was based on the same individuals which were included in the GWAS (thus, genetic outliers and related individuals were removed) and corrected using genotyping wave and psychiatric disorders as covariates as well as the same principal components from PCA which were used in the GWAS. Since gene expression among the different brain tissues is highly correlated, we corrected the P-value for each gene by the total number genes tested in all tissues with a valid model available for the gene, with respect to *CHRNA2* we corrected for 13,166 genes tested in three tissues, thus P-values $< 3.8 \times 10^{-6}$ ($0.05/13,166$) were considered statistical significant for this gene. This is in line

with recent approaches used to correct for multiple testing in transcriptome wide association studies using imputed gene expression⁸¹.

Polygenic risk score analyses—PRS analyses were done using GWAS summary statistics from 22 GWASs (Supplementary Table 11). The summary files were downloaded from public databases and processed using the munge script which is a part of the LDscore regression software⁷⁴. All variants with INFO < 0.9, MAF < 0.01, missing values, out of bounds P-values, ambiguous strand alleles and duplicated rs-ids were removed using the munge script. In addition, multi-allelic variants and insertion and deletion (indels) were removed. The processed summary files were then LD-clumped using Plink, with the following parameter settings: `--clump-p1 1 --clump-p2 1 --clump-r2 0.1 --clump-kb 500`. The clumped file were used as the training dataset. Genetic risk scores were estimated at different P-value thresholds for SNP inclusion: 5×10^{-8} , 1×10^{-6} , 1×10^{-4} , 1×10^{-3} , 0.01, 0.05, 0.1, 0.2, 0.5 and 1.0 for all individuals in the target sample (CUD cases and the control group) from the genotype dosages using Plink's '--score' method, with default arguments. However the PRS scores for ADHD, were generated using the approach described Demontis et al.³³. For each P-value threshold the variance in the phenotype explained by PRS was estimated using Nagelkerke's *R* (R package 'BaylorEdPsych'), and association of PRS with CUD was estimated using logistic regression including the same covariates used in the GWAS analysis (PCs from PCA and the psychiatric disorders listed in Supplementary Table 1). In PRS analyses of psychiatric disorders (ADHD, schizophrenia and depression related phenotypes) individuals with a diagnosis of the disorder being analysed were excluded. The number of individuals excluded with ADHD, schizophrenia and major depressive disorder are listed in Supplementary Table 1.

Reporting Summary.

Further information on research design is available in the Life Sciences Reporting Summary linked to this article.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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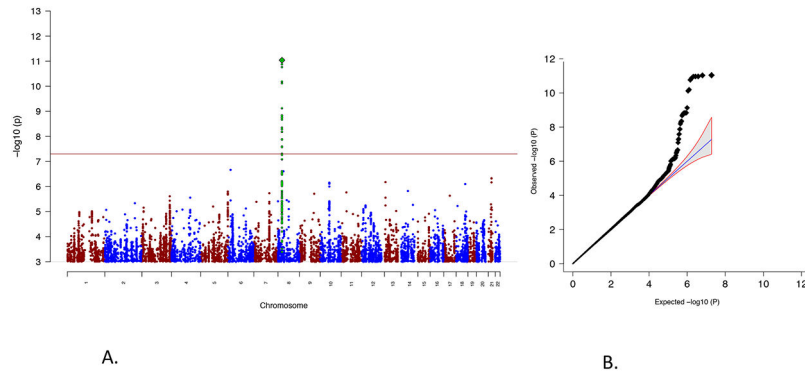


Figure 1. Genome-wide association results from the CUD GWAS results

A) Manhattan plot of the results from the GWAS of CUD (2,387 individuals with CUD and 48,985 controls). Results are from logistic regression and P-values are two-sided. The red horizontal line indicates the threshold for genome-wide significance ($P=5 \times 10^{-8}$). The index variant is highlighted as a green diamond and SNPs in high LD with the index SNP are marked in green. **B)** Quantile-quantile plot of the expected and observed P-values from GWAS of CUD (2,387 individuals with CUD and 48,985 controls). The blue line indicates the distribution under the null hypothesis and the shaded area indicates the 95% confidence band.

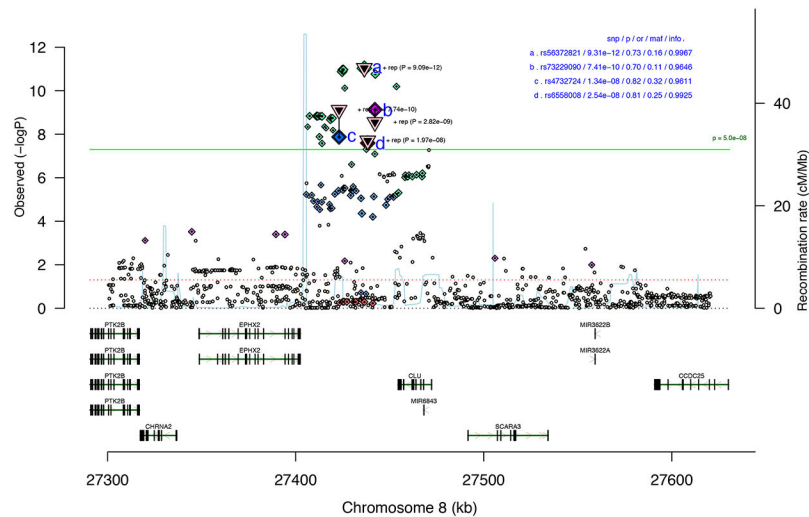


Figure 2. Association results for the genomic region with the CUD risk locus

Regional association plot of the local association results for the risk locus at chromosome 8 based analysis of 2,387 individuals with CUD and 48,985 controls. Results are from logistic regression and P-values are two-sided. The index variant (rs56372821) and additional three correlated genome-wide significant variants (LD with index variant: $0.2 < r^2 < 0.7$) are marked with letters (a-d), the triangle represents the two-sided P-value from meta-analysis (inverse variance weighted fixed effects model) with the replication cohort from deCODE (5,501 individuals with CUD and 301,041 controls). The location and orientation of the genes in the region and the local estimates of recombination rate is shown. The association P-value (p), odds ratio (or), minor allele frequency (maf) and imputation info-score (info) is presented in upper right corner. The horizontal green line represents the threshold for genome-wide significant ($P=5 \times 10^{-8}$).

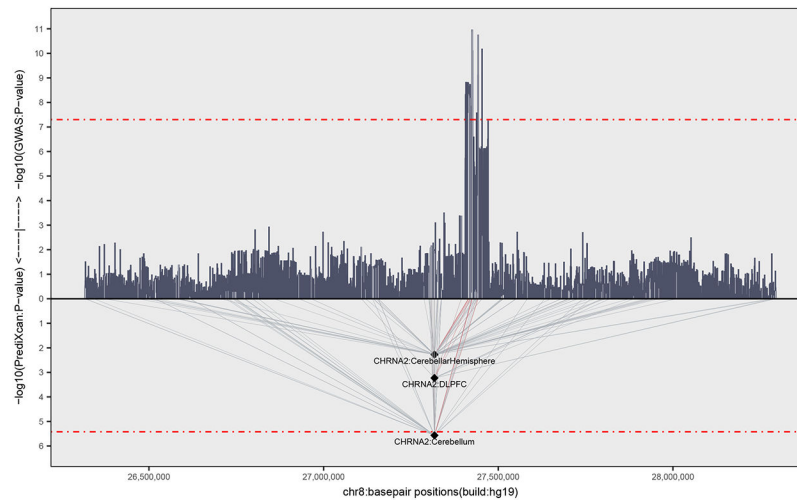


Figure 3. Association of *CHRNA2* expression with CUD

Association of the imputed genetically regulated expression of *CHRNA2* with CUD in three brain tissues with a valid model (cerebellar hemisphere, dorsolateral prefrontal cortex and cerebellum). The two-side P-value from logistic regression for the association of gene expression with CUD ($-\log_{10}(\text{PrediXcan:P-value})$) and two-side P-value from logistic regression from the CUD GWAS ($-\log_{10}(\text{GWAS:P-value})$) is given on the y-axis, with a red dotted line indicating statistical significance. Both analyses include 2,387 individuals with CUD and 48,985 controls. Chromosome position is given on the x-axis and the light grey lines indicate which SNPs that are included in the models used to predict gene expression. The thin red lines indicate genetic predictors that are genome-wide significantly associated with CUD.

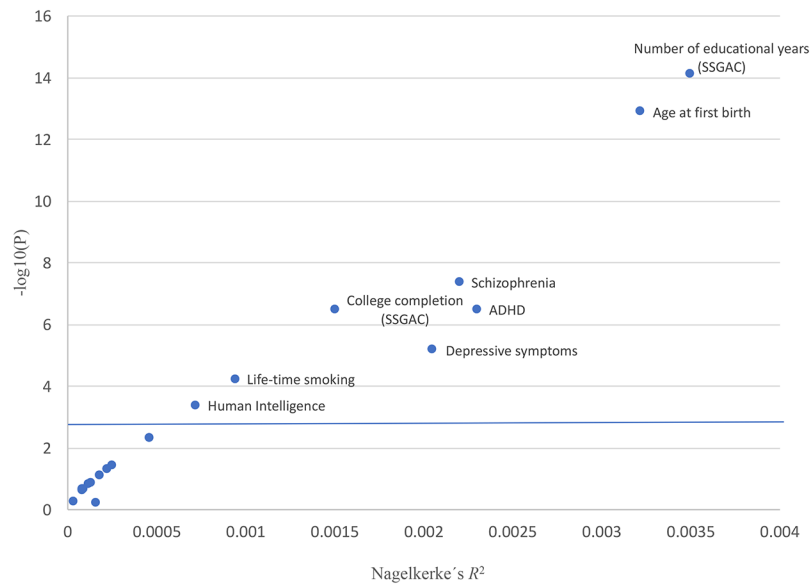


Figure 4. Association of PRS with CUD

PRSs were generated for phenotypes related to cognition, personality, psychiatric disorders, reproduction and smoking behavior based on summary statistics from 22 published GWASs. The variance explained by the scores (Nagelkerke- R^2) is given on the x-axis and the P-value from logistic regression for association of the PRS with CUD on the y-axis (based on analyses of 2,387 individuals with CUD and 48,985 controls). The vertical blue line indicates statistical significance ($P=0.0023$; correcting for 22 tests). In PRS analyses of psychiatric disorders, individuals having a diagnosis of the respective psychiatric disorder being analysed (ADHD, schizophrenia, depressive symptoms and major depressive disorder) were excluded in the CUD target sample.

Genome-wide significant SNPs associated with CUD

Table 1.

Results for variants passing the threshold for genome-wide significance ($P=5 \times 10^{-8}$) in the associated locus at chromosome 8 in the primary GWAS (2,387 individuals with CUD and 48,985 controls) and/or in the meta-analysis with results from an independent cohort collected by deCODE (5,501 cases with CUD and 301,041 controls). Results for the index SNP (Index SNP) is shown together with results from supporting correlated SNPs in the locus ($0.2 < r^2 < 0.7$) (LD SNPs). Alleles for the variants (A1 and A2), frequency of A1 in controls (Frq A1 con), the odds ratio (OR) for the effect of A1, and P-values (P) are given. Two-sided P-values from logistic regression of the iPSYCH and deCODE cohorts and from inverse variance weighted fixed effects meta-analysis. Bolding of P values indicates genome-wide significance

Index SNP	LD SNPs	CHR	BP (hg19)	iPSYCH				deCODE				meta-analysis					
				A1	A2	Frq A1 con	OR	P	A1	A2	Frq A1 con	OR	P	A1	A2	OR	P
rs56372821		8	27436500	A	G	0.163	0.728	9.31x10⁻¹²	A	G	0.144	0.878	3.27 x10 ⁻⁰³	A	G	0.803	9.09 x10⁻¹²
	rs4732724	8	27423062	C	G	0.324	0.820	1.34 x10⁻⁰⁸	C	G	0.367	0.905	1.60 x10 ⁻⁰³	C	G	0.866	7.74 x10⁻¹⁰
	rs6558008	8	27438306	A	C	0.755	1.237	2.54 x10⁻⁰⁸	C	A	0.228	0.915	1.45 x10 ⁻⁰²	A	C	1.159	1.97 x10⁻⁰⁸
	rs73229090	8	27442127	A	C	0.110	0.702	7.41 x10⁻¹⁰	A	C	0.105	0.881	1.30 x10 ⁻⁰²	A	C	0.797	2.82 x10⁻⁰⁹
	rs937220	8	27429730	A	G	0.259	0.8258	2.46 x10 ⁻⁰⁷	A	G	0.269	0.912	7.56 x10 ⁻⁰³	A	G	0.871	4.53 x10⁻⁰⁸
	rs17466060	8	27422740	A	G	0.418	0.8594	3.94 x10 ⁻⁰⁶	A	G	0.430	0.900	5.11 x10 ⁻⁰⁴	A	G	0.881	1.29 x10⁻⁰⁸

Table 2. The effect on the odds ratio of rs56372821 of leave-one-phenotype out association analyses

In the analyses, individuals with a psychiatric diagnosis in the iPSYCH cohort were excluded, one disorder at a time, and an analysis without the controls was also conducted: without ADHD (woADHD), without autism spectrum disorder (woASD), without bipolar disorder (woBP), without the iPSYCH control cohort (woCON), without major depressive disorder (woMDD), without schizophrenia (woSZ). GWASs were done using logistic regression and P-values are two-sided. The P-value for association with CUD (P-value), number of cases (cases), frequency of the minor allele in cases (Freq cases) and number of controls (con), frequency of the minor allele in controls (Freq con), odds ratio (OR) and standard error (SE), are given. For illustration OR and the corresponding standard error.

	P-value	cases	Freq cases con	Freq con	OR	SE	OR
Cannabis							
GWAS	9.31x10 ⁻¹²	2387	0.124	48985	0.163	0.727	0.047
woADHD	2.68x10 ⁻¹⁰	1461	0.118	39284	0.165	0.683	0.060
woASD	7.19x10 ⁻¹⁰	2234	0.126	40287	0.164	0.744	0.048
woBP	5.61x10 ⁻¹⁰	2199	0.126	47809	0.163	0.742	0.048
woCON	2.35x10 ⁻¹¹	2290	0.124	32716	0.162	0.726	0.048
woMDD	2.81x10 ⁻⁰⁷	1253	0.121	32592	0.162	0.711	0.066
woSZ	7.15x10 ⁻¹⁰	1818	0.125	47196	0.163	0.725	0.052