

Aggressive Hematopoietic Malignancy Characterized by Biallelic Loss of *SMARCB1*

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CASE PRESENTATION

Here, we report a 14-year-old man with autism spectrum disorder and growth delay who presented with several days of left shoulder pain and new onset of progressive dyspnea. No palpable masses were detected. Imaging demonstrated a large heterogeneous mass in the upper mediastinum resulting in deviation of the trachea (Fig 1). Enlarged lymphadenopathy was seen in the bilateral supraclavicular regions and bilateral neck level IV, III, and Vb regions. No disease was present below the diaphragm. Bone marrow and CSF studies were negative at the time of diagnosis. A malignant lymphoma was suspected, and a biopsy of an involved lymph node was performed.

Histologic sections of the lymph node revealed effacement of the normal architecture by a diffuse atypical mononuclear cell proliferation with irregular nuclear contours and vesicular chromatin. Scattered larger cells were present showing marked pleomorphism and occasional multinucleation. Mitotic figures were numerous including atypical mitoses. There were a few background inflammatory cells (lymphocytes, eosinophils, and histiocytes) and focal necrosis (Figs 2A and 2B).

Immunohistochemical studies showed that the neoplastic cells expressed CD45 (leukocyte common antigen), supporting hematopoietic cell origin. The neoplastic cells also expressed CD2, CD7, CD79a, myeloperoxidase (dim), and CD13, suggesting T/natural killer (NK)-cell, B-cell and myeloid differentiation (Figs 2C-2G). CD3, CD34, and terminal deoxynucleotidyl transferase were negative (not shown). Neoplastic cells did not express cytokeratin or myogenin (Fig 2H) against an epithelial or rhabdoid origin. On the basis of these findings, the diagnosis of malignant hematopoietic neoplasm with myeloid, B-cell, and T/NK-cell differentiation was rendered.

The patient was treated with the non-Hodgkin lymphoma regimen DA-EPOCH, consisting of etoposide, prednisone, vincristine, cyclophosphamide, and doxorubicin. After three cycles of therapy, positron emission tomography (PET) imaging demonstrated a 68% reduction in the size of the mediastinal mass but no notable change in fluorodeoxyglucose avidity (standard uptake value maximum, 11.3 v 10.4 on the initial study). He went on to receive two additional cycles. After the sixth cycle of DA-EPOCH, he exhibited progressive disease, with an interval increase in mass size (approximately 20%) and fluorodeoxyglucose avidity (12.5) on the PET scan.

A repeat biopsy confirmed the persistence of a hematopoietic neoplasm. Given the unique histology and aggressive clinical course of the malignancy, material from his biopsy was sent to Memorial Sloan Kettering Cancer Center for genomic profiling through the Make-an-IMPACT program¹, an initiative that provides genomic profiling for rare tumors using the MSK-IMPACT targeted next-generation sequencing platform (XXXX).² Profiling revealed a biallelic loss of *SMARCB1*, confirmed by loss of *INI1* expression by immunohistochemistry (Figs 3A and 3B). A somatic *TP53* mutation was also identified. The copy number profile was suggestive of broad copy number gains on chromosomes 6, 9, 19, and X. A targeted RNA-based assay (Archer FusionPlex, ArcherDx, Boulder, CO) did not identify any oncogenic gene fusions. To identify rare oncogenic events in genes not included in the targeted DNA and RNA clinical sequencing assays, whole-exome sequencing was performed

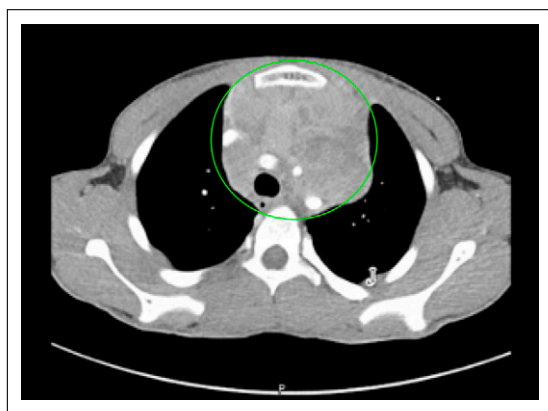


FIG 1. Contrast-enhanced axial computed tomography (CT) imaging of the chest. There is a large, heterogeneous, mildly enhancing mass (circled in green) within the anterior mediastinum. There is mass effect upon the trachea with mild luminal compromise.

ASSOCIATED CONTENT

Data Supplement

Author affiliations and support information (if applicable) appear at the end of this article.

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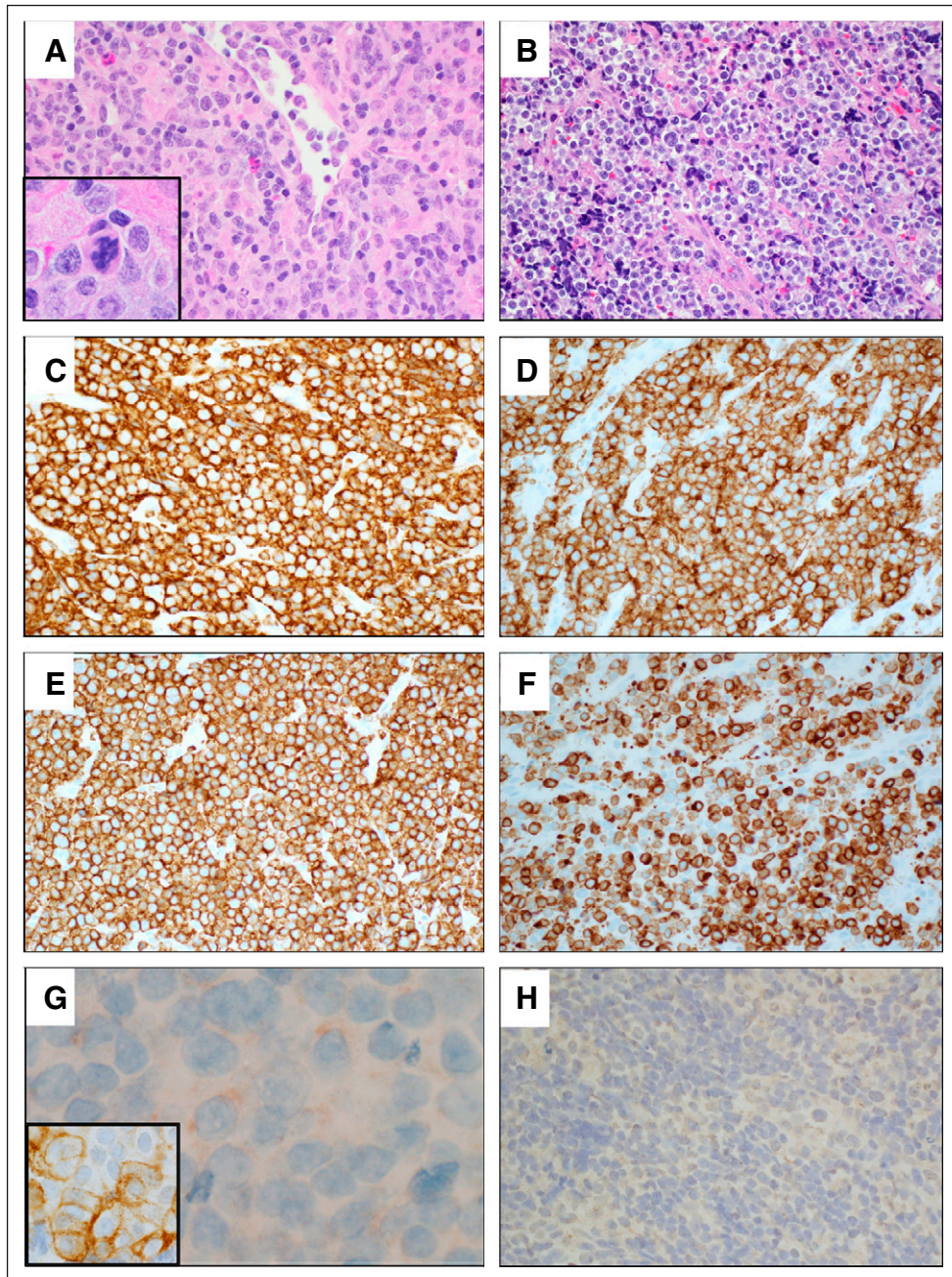


FIG 2. Lymph node and mediastinal involvement by malignant hematopoietic neoplasm with *SMARCB1* deletion showing myeloid, B-cell, and T/natural killer (NK)-cell differentiation. (A) Histologic section of left cervical lymph node revealed diffuse proliferation of atypical mononuclear cells with numerous mitotic figures (inset). There were a few background inflammatory cells (hematoxylin and eosin stain, $\times 400$). (B) Histologic sections of the left anterior mediastinal mass showed atypical mononuclear cells with scattered large cells presenting marked pleomorphism and occasional multinucleation (hematoxylin and eosin stain, $\times 400$). (C) Neoplastic cells showed strong expression of CD45 (leukocyte common antigen; CD45 immunohistochemistry, $\times 400$). (D) CD2 immunohistochemistry showed diffuse and strong expression in neoplastic cells, indicating T/NK-cell differentiation (CD2 immunohistochemistry, $\times 400$). (E) CD7 was diffusely and strongly expressed in neoplastic cells (CD7 immunohistochemistry, $\times 400$). (F) A subset of neoplastic cells shows strong expression of CD79a, indicating B-cell differentiation (CD79a immunohistochemistry, $\times 400$). (G) Weak cytoplasmic expression of myeloperoxidase was seen in a subset of cells, and atypical cells also show immunoreactivity to CD13 immunohistochemistry (inset), supporting myeloid differentiation (myeloperoxidase immunohistochemistry, $\times 1,000$; CD13 immunohistochemistry, $\times 1,000$). (H) Immunohistochemistry for myogenin demonstrated that these cells did not show rhabdoid differentiation (myogenin immunohistochemistry, $\times 400$).

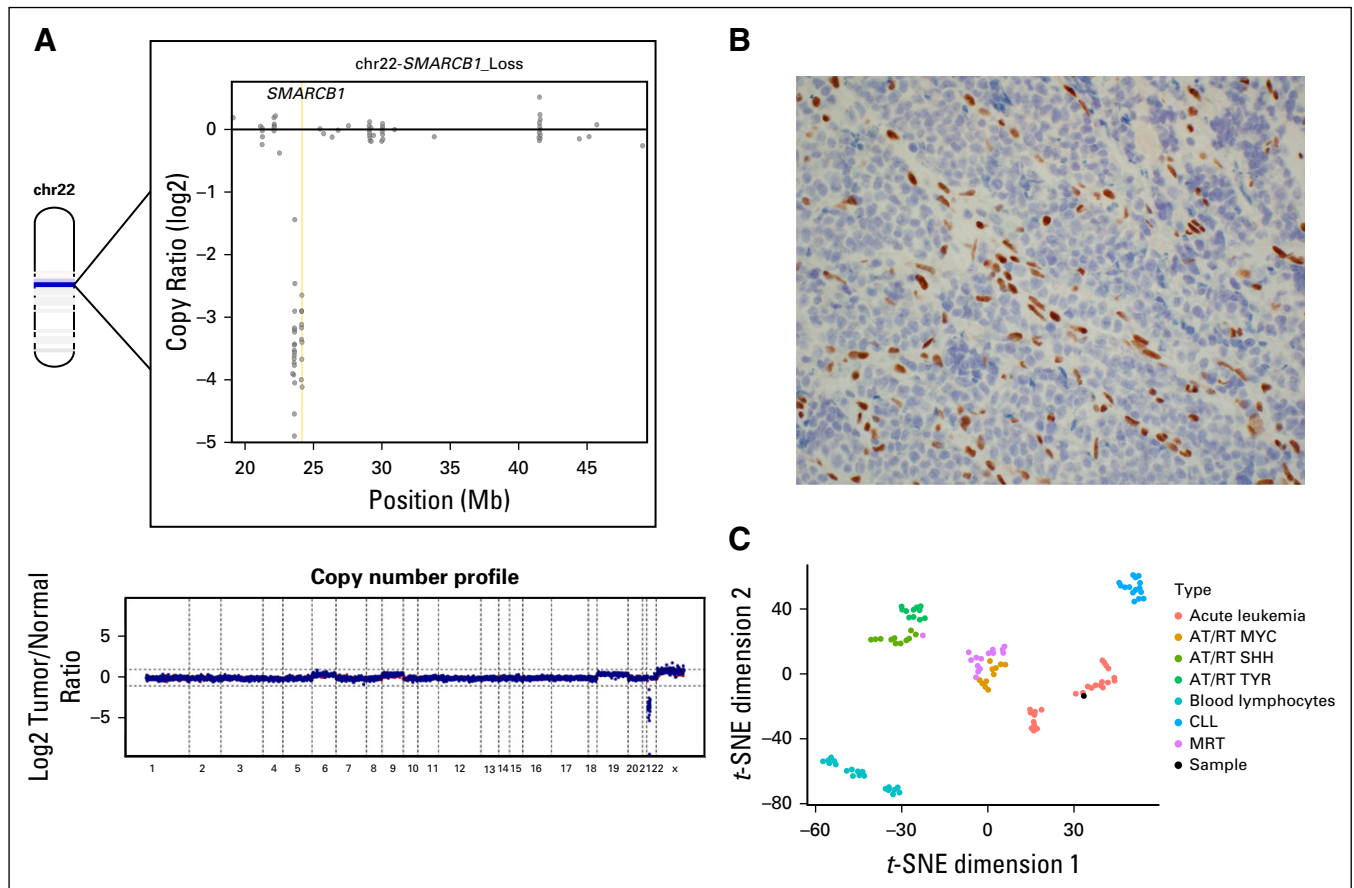


FIG 3. (A) The bottom part of the figure represents the genome wide copy number profile based on log2 copy number coverages at exon locations covered by the MSK IMPACT platform. Chromosome 22 (chr22) is depicted in the upper left part of the figure, with a dark blue band representing the location and loss of *SMARCB1*. The upper right part of the figure demonstrates log2 copy number coverages at exon locations across chr22, demonstrating complete loss of *SMARCB1* at all exon locations. (B) Neoplastic cells showed aberrant loss of *INI1* expression, whereas endothelial cells had normal expression of *INI1* (*INI1*/BAF-47 immunohistochemistry, $\times 400$). (C) *t*-Distributed Stochastic Neighbor Embedding (*t*-SNE) analysis of methylation array data demonstrated localization of the patient's sample within the acute leukemia cluster rather than with other *SMARCB1*-deficient entity clusters. AT/RT, atypical teratoid rhabdoid tumor; CLL, chronic lymphocytic leukemia; MRT, malignant rhabdoid tumor.

but did not identify any additional candidate driver alterations (Data Supplement).

DNA-methylation profiling is an effective modality for the classification of *SMARCB1*-deficient CNS tumors and extracranial malignant rhabdoid tumors.^{3,4} To assess whether this tumor phenotypically clustered with *SMARCB1* CNS or solid tumors, genome-wide methylation profiles were obtained using the Infinium MethylationEPIC/850k platform (Illumina, San Diego, CA) and subjected to a *t*-Distributed Stochastic Neighbor Embedding dimensionality reduction⁵ against *SMARCB1*-deficient entities (atypical teratoid/rhabdoid tumor and malignant rhabdoid tumor) as well as a select cohort of acute and chronic hematopoietic neoplasms (acute leukemia and chronic lymphocytic leukemia) obtained from the University of Heidelberg. Interestingly, the tumor localized to the acute leukemia cluster rather than the *SMARCB1*-deficient solid/CNS tumors and the control hematopoietic tissue (normal blood lymphocytes

and chronic lymphocytic leukemia/small lymphocytic lymphoma samples; Fig 3C).

While MSK-IMPACT profiling was being performed, salvage therapy with brentuximab vedotin, nivolumab, and bendamustine was attempted. Unfortunately, the patient experienced a rapid progression of mediastinal disease and died before the availability of the profiling results. No additional tissue sampling or autopsy was performed. The patient's guardian provided written consent to share his deidentified clinical information in this publication.

DISCUSSION

The SWItch/Sucrose Non-Fermentable (SWI/SNF) chromatin remodeling complex is a large, multiprotein complex, that mobilizes nucleosomes using energy derived from ATP hydrolysis.⁵ Proteins within the complex serve key roles in transcriptional regulation and tumor suppression.⁶⁻⁸

Mutations in genes encoding subunits of SWI/SNF complex have been found in approximately 20% of all tumor genomes sequenced to date, marking it as one of the most commonly mutated chromatin modulators in human cancer.⁹ *SMARCB1* (also known as INI1 or BAF47) is one of the core component proteins in the SWI/SNF chromatin remodeling complex. Homozygous deletions of *SMARCB1* are driver oncogenic events in several solid tumor types, including malignant rhabdoid tumors, renal medullary carcinomas, and a subset of epithelioid sarcomas.¹⁰⁻¹²

To our knowledge, this is the first report of a hematologic malignancy with confirmed somatic biallelic loss of *SMARCB1*. Interestingly, mouse models with inactivating *SMARCB1* mutations are known to develop T-cell lymphomas.¹³ However, genomic profiling studies of T-cell lymphomas have not reported recurrent inactivating *SMARCB1* mutations.^{14,15} Deletions of *SMARCB1* have also been identified in chronic myeloid leukemia samples but only as heterozygous losses, resulting in the potential reduction of gene dosage but not complete inactivation.¹⁶

Our patient had a highly aggressive disease course, a clinical feature shared with *SMARCB1*-deficient solid tumors. Despite intensive multimodal chemotherapy and radiotherapy, he experienced rapid disease progression and death within 6 months of diagnosis. Genomically, the tumor cells exhibited a low tumor mutational burden, which

is congruent with *SMARCB1*-deficient solid tumors.¹⁷ However, the histologic features and DNA-methylation profile identify this tumor as a hematologic malignancy distinct from *SMARCB1*-deficient solid tumors.

In conclusion, we have identified a unique case of an aggressive hematopoietic malignancy characterized by biallelic loss of *SMARCB1*. Several clinical trials have been initiated that are selectively accruing patients with biallelic loss of *SMARCB1*, including ongoing studies of the enhancer of zeste homolog 2 (*EZH2*) inhibitor tazemetostat (ClinicalTrials.gov identifier: [NCT02601937](https://clinicaltrials.gov/ct2/show/study/NCT02601937)). *EZH2* expression is upregulated in the setting of biallelic loss of *SMARCB1*, rendering these tumors potentially vulnerable to target inhibition.¹⁸ Results from phase II studies have led to US Food and Drug Administration approval for patients with metastatic or locally advanced epithelioid sarcoma.¹⁹ Preliminary pediatric data also demonstrate activity in subsets of other *SMARCB1*-deficient tumor histologies.²⁰ Our patient experienced disease progression too rapidly to be enrolled into a therapeutic trial. However, this case highlights the potential clinical utility of early genomic and epigenetic profiling of unusual malignant histologies to ensure the correct cancer type diagnosis and to identify potentially actionable alterations that could serve as targets for therapeutic intervention.

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