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AIBL Research Group: <https://aibl.csiro.au/about/aibl-research-team/>

CRedit Statement

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All co-authors meet ICMJE criteria, have seen and approved the contents of the manuscript.

Competing Interest

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3. Previous Publication

This work has not been published in any other form, nor is under consideration at any other publication.

4. Ethics Approvals

The institutional ethics committees of Austin Health, St Vincent's Health, Hollywood Private Hospital, and Edith Cowan University approved the AIBL study, and all volunteers gave written informed consent before participating.

Institutional Review Boards and Research Ethics Boards according to applicable State and Federal requirements for each participating location provided approval for ADNI evaluations, all participants gave informed consent.

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Impact of *APOE-ε4* carriage on the onset and rates of neocortical Aβ-amyloid deposition

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Abstract

Neocortical A β -amyloid deposition, one of the hallmark pathologic features of Alzheimer's disease (AD), begins decades prior to the presence of clinical symptoms. As clinical trials move to secondary and even primary prevention, understanding the rates of neocortical A β -amyloid deposition and the age at which A β -amyloid deposition becomes abnormal is crucial for optimising the timing of these trials. As *APOE- ϵ 4* carriage is thought to modulate the age of clinical onset, it is also important to understand the impact of *APOE- ϵ 4* carriage on the age at which neocortical A β -amyloid deposition becomes abnormal. Here, we show that, for 455 participants with over three years of follow-up, abnormal levels of neocortical A β -amyloid were reached on average at age 72 (66.5-77.1). The *APOE- ϵ 4* carriers reached abnormal levels earlier at age 63 (59.6-70.3), however, non-carriers reached the threshold later at age 78 (76.1-84.4). No differences in rates of deposition were observed between *APOE- ϵ 4* carriers and non-carriers after abnormal A β -amyloid levels had been reached. These results suggest that primary and secondary prevention trials, looking to recruit at the earliest stages of disease, should target *APOE- ϵ 4* carriers between the ages of 60 and 66 and non-carriers between the ages of 76 and 84.

Keywords

A β -amyloid; *APOE*; Alzheimer's disease; Longitudinal; Biomarkers

1. Introduction

Alzheimer's disease (AD), the most common form of dementia, is characterised pathologically by the extracellular accumulation of A β -amyloid and intracellular accumulation of tau in the neocortex (Jack et al., 2018). Neocortical accumulation of A β -amyloid is a key part of the cascade of pathological changes leading to the onset of clinical symptoms in AD (Hardy and Selkoe, 2002; Karran et al., 2011) and is a process that initiates decades prior to clinical manifestation of the disease (Jack et al., 2013a; Villemagne et al., 2013). Increased understanding of onset and rates of neocortical A β -amyloid deposition would provide improved disease staging criteria particularly for pre-clinical AD. This is increasingly important with clinical trials aimed at preventative treatment.

Carriage of an *APOE- ϵ 4* allele is a well-established risk factor for AD (Harold et al., 2009), reported to impact the levels of neocortical A β -amyloid (Liu et al., 2013; Reiman et al., 2009; Rowe et al., 2010; Villemagne et al., 2011), however the nature of this impact is unclear. The literature appears to agree that *APOE- ϵ 4* carriage is associated with deposition of neocortical A β -amyloid at an earlier age (Bilgel et al., 2019; Fleisher et al., 2013; Mishra et al., 2018) as well as an earlier onset of disease (Corder et al., 1995). Some contributions report that *APOE- ϵ 4* carriage is associated with an increased rate of neocortical A β -amyloid deposition (Bilgel et al., 2019; Jack et al., 2013a; Mishra et al., 2018; Toledo et al., 2019), others only report a difference in those with low neocortical A β -amyloid burden (Lim et al., 2017), whilst others report no difference in neocortical A β -amyloid accumulation rates between carriers and non-carriers (Corder et al., 1995; Resnick et al., 2015; Saunders, 2000). Accounting for the temporal relationship between neocortical A β -amyloid deposition and disease stage/progression may provide a clearer understanding of the impact of *APOE- ϵ 4* carriage on neocortical A β -amyloid deposition.

In this study we evaluate the age at which abnormal levels of neocortical A β -amyloid deposition can be detected and test our hypotheses that carriage of an *APOE- ϵ 4* allele would be associated with a) a younger age of onset and b) faster rates of neocortical A β -amyloid deposition. For that purpose, natural history modelling in conjunction with survival analyses is employed to jointly consider onset and rates of neocortical A β -amyloid accumulation in reference to disease stage and progression.

2. Materials & Methods

2.1 AIBL Cohort

The Australian Imaging, Biomarker and Lifestyle (AIBL) cohort study of ageing combines data from neuroimaging, biomarkers, lifestyle, clinical, and neuropsychological assessments. Two study centres in Melbourne, VIC, and Perth, WA, Australia recruit mild cognitively impaired (MCI) individuals and individuals with Alzheimer's disease from primary-care physicians or tertiary Memory Disorders Clinics. Cognitively healthy normal controls (NC) were recruited through advertisement or from spouses of participants in the study. Exclusion criteria were a history of non-Alzheimer's disease dementia, Parkinson's disease, schizophrenia, bipolar disorder, obstructive sleep apnoea, serious head injury, current depression (Geriatric Depression Score >5 out of 15), cancer in the past two years (with the

exception of basal-cell skin carcinoma), symptomatic stroke, uncontrolled diabetes, or current regular alcohol use. Between Nov 3, 2006, and Oct 30, 2008, AIBL recruited 1112 eligible volunteers, who were aged 60 years or older and fluent in English. An enrichment cohort of 86 patients with Alzheimer's disease (AD), 124 MCI and 389 NC were recruited by AIBL between March 30, 2011, and June 29, 2015. At baseline, the AIBL study participants were an average of 72 years of age, consisted of 58% women, and 36% were *APOE-ε4* carriers. The institutional ethics committees of Austin Health, St Vincent's Health, Hollywood Private Hospital, and Edith Cowan University approved the AIBL study, and all volunteers gave written informed consent before participating.

2.1.1 PET Aβ-Amyloid—AIBL Aβ-Amyloid positron emission tomography (PET) studies consisted of a 30-minute acquisition starting 40 minutes after injection of 370 MBq of ¹¹C-Pittsburgh compound-B (¹¹C-PiB). For semi-quantitative analysis, PET images were spatially normalised with CapAIBL[®] using an adaptive atlas (Bougeat et al., 2015). The summed and spatially normalised PET images were then scaled to the recommended reference region, cerebellar cortex, to generate a tissue ratio termed SUV ratio (SUVR), and sampled using a pre-set template of narrow cortical volumes of interest. A global measure of the Aβ-amyloid level was computed using the mean SUVR in the frontal, superior parietal, lateral temporal, lateral occipital, and anterior and posterior cingulate regions. The abnormal threshold for levels of Aβ-amyloid in AIBL participants was set as 1.4 SUVR (Jack et al., 2013b).

2.1.2 Assessment of APOE genotype—*APOE* genotype was determined through TaqMan[®] genotyping assays (Life Technologies) for rs7412 (Assay ID: C____904973_10) and rs429358 (Assay ID: C____3084793_20). TaqMan[®] genotyping assays were performed on a QuantStudio 12K Flex[™] Real-Time-PCR systems (Applied Biosystems, Foster City, CA) using the TaqMan[®] GTXpress[™] Master Mix (Life Technologies) methodology as per manufacturer instructions. *APOE* carrier status was defined by the presence (1 or 2 copies) or absence (0 copies) of the *APOE-ε4* allele.

2.2 ADNI Cohort

Data used in the preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). The ADNI was launched in 2003 as a public-private partnership, led by Principal Investigator Michael W. Weiner, MD. The primary goal of ADNI has been to test whether serial magnetic resonance imaging (MRI), PET, other biological markers, and clinical and neuropsychological assessment can be combined to measure the progression of MCI and early AD participants. Subjects were recruited from 57 sites across the United States and Canada and are followed-up annually. ADNI initially (ADNI 1) recruited 200 NC subjects, 400 MCI subjects and 200 subjects with early AD. In addition, ADNI GO, launched in 2009 included 200 subjects identified as having early mild cognitive impairment (EMCI). In 2011, ADNI 2 [11] recruited 150 NC, 100 EMCI participants, 150 late mild cognitive impairment (LMCI) participants and 150 AD participants. More recently, ADNI 3 was launched (September 2016) to recruit an additional 1,200 volunteers.

2.2.1 PET A β -Amyloid—ADNI A β -Amyloid PET studies consisted of an acquisition of 4 x 5-minute frames commencing 50-70 minutes after injection of 10 mCi of ^{18}F Florbetapir (FBP). In the same manner as the AIBL images, the ADNI PET images were spatially normalised with CapAIBL[®] using an adaptive atlas (Bougeat et al., 2015). The summed and spatially normalised PET images were then scaled to a white matter reference region (a composite of the centrum semiovale and corpus callosum)(Chen et al., 2015) to generate a tissue ratio termed SUV ratio (SUVR), and sampled using the same pre-set template of narrow cortical volumes of interest as for the AIBL cohort. A global measure of the A β -amyloid level was computed using the mean SUVR in the frontal, superior parietal, lateral temporal, lateral occipital, and anterior and posterior cingulate regions. The abnormal threshold for levels of A β -amyloid in ADNI participants was set as 0.61 SUVR (equivalent to 1.4 SUVR for ^{11}C -PiB and 1.10 for FBP whole cerebellum correction (Clark et al., 2012)).

2.2.2 Assessment of APOE genotype—A 3 mL aliquot of blood was taken in ethylenediaminetetraacetic acid (EDTA)-containing vacutainer tubes from ADNI participants, and genomic DNA was extracted at Cogenics (now Beckman Coulter Genomics) using the QIAamp DNA Blood Maxi Kit (Qiagen, Inc, Valencia, CA) following the manufacturer's protocol. The two SNPs (rs429358, rs7412) that define the *APOE* epsilon 2, 3, and 4 alleles were evaluated by polymerase chain reaction amplification, followed by HhaI restriction enzyme digestion, resolution on 4% Metaphor Gel, and visualization by ethidium bromide staining (Potkin et al., 2009; Saykin et al., 2010).

2.3 Statistical Analysis

AIBL (n=209) and ADNI participants (n=246) with at least three years of follow-up evaluations for A β -amyloid with ^{11}C -PiB (AIBL) or ^{18}F Florbetapir PET (ADNI), respectively, who were considered to be accumulating A β -Amyloid (rate of deposition > 0.0 SUVR/year(Villemagne et al., 2013)), and had been genotyped for *APOE* were included in this study. The following analyses were produced in parallel for both the AIBL participants and the ADNI participants. Further, all analyses were again replicated for the NC participants (156 AIBL NC and 106 ADNI NC) in a sensitivity analysis. For comparison purposes, in ADNI, EMCI and LMCI participants were both considered as MCI to align with the classifications in AIBL. All analyses were performed in the R environment (R Development Core Team, 2017).

Demographics: Baseline differences between *APOE*- ϵ 4 carriers and non-carriers were assessed with one-way t tests for continuous data (age), χ^2 testing for categorised data (sex, years of education, disease classification), and Kruskal-Wallis testing for non- normally distributed data (length of follow-up). This was replicated for the individuals excluded from the study as they were not accumulating A β -Amyloid (46 AIBL and 14 ADNI participants).

The differences in rates of A β -amyloid deposition: Each individual's rate of deposition (SUVR/year) was estimated using a linear model regressing their neocortical A β -amyloid levels (SUVR) against time since baseline evaluation (years). Differences in these rates between *APOE*- ϵ 4 carriers and non-carriers, as well as between those above or below the

neocortical A β -amyloid threshold at baseline, were evaluated using one-way t tests, and presented using box and jitter plots. This analysis was also replicated for a combined cohort of those accumulating and not accumulating A β -Amyloid in a sensitivity analysis.

Natural history of deposition: Individual's rates of A β -amyloid deposition, calculated above, were combined to estimate the overall natural history of A β -amyloid deposition using the 4-step procedure described previously (Budgeon et al., 2017; Villemagne et al., 2013) and stratified by *APOE- ϵ 4* carriage. Briefly, the 4-step procedure comprises 1) estimating the mean and slope of each individuals' A β -amyloid using linear models, 2) fitting a polynomial to the estimated means and slopes across all individuals, 3) integrating the reciprocal of the fitted polynomial, 4) inverting the function to obtain the natural history trajectory.

Confidence intervals for the natural history curves were created using the bootstrapping procedure described previously (Budgeon et al., 2017). Note: this analysis was replicated with stratification by sex.

Age of Onset: Cox proportional hazards model of survival, corrected for sex, and years of education were utilised to estimate the age at which participants reached abnormal levels of neocortical A β -amyloid. This analysis was replicated with *APOE- ϵ 4* carriage stratification, to assess the effect of *APOE- ϵ 4* carriage on the age at which participants reached abnormal levels of A β -amyloid. Survival was defined as the time between birth and having a PET scan indicating abnormal levels of A β -Amyloid, withdrawal from the study, or the last completed follow-up examination. The event was classified as having a PET scan indicating abnormal levels of A β -amyloid. For some individuals the date at which their amyloid levels would have become abnormal was imputed, further details on the imputation are provided in supplementary material. The median age at which participants reached abnormal levels of A β -Amyloid, represented by the age at which 50% of the cohort reached abnormal levels of A β -Amyloid, was reported.

2.4 Data Availability

All ADNI and a subset of the AIBL data including images are shared through the LONI Image & Data Archive (<http://adni.loni.usc.edu>), a secure research data repository.

Applications for access to the entirety of the AIBL data can be made via application through the AIBL website (<https://aibl.csiro.au/>).

3. Results

3.1 Demographics

There were a significantly higher proportion of NC participants in the AIBL *APOE- ϵ 4* non-carriers compared to carriers ($p=0.001$), for the ADNI participants this relationship held as a trend ($p=0.057$). Within AIBL, there were significantly more Males among the *APOE- ϵ 4* carriers compared to non-carriers ($p=0.026$), a finding not observed in the ADNI participants ($p=0.683$). The ADNI *APOE- ϵ 4* non-carriers were significantly older than carriers ($p=0.005$), no differences were observed for age between *APOE- ϵ 4* carriers and non-carriers in AIBL ($p=0.196$). No differences were observed between *APOE- ϵ 4* carriers and non-carriers, in either AIBL or ADNI, for Years of Education, or length of follow-up (Table 1).

There were no significant differences between the *APOE-ε4* carriers and non-carriers for these demographic measures in the AIBL participants deemed to be non-accumulators (Supplementary Table 1), caution should be applied to these findings due to the small sample size however. Due to small the sample size comparisons could not be drawn for the ADNI non-accumulators (Supplementary Table 1). There appeared to be more Males and shorter follow-up in the non-accumulators compared to the accumulators, again the small sample size of the excluded non-accumulators should be noted.

3.2 Rates of Aβ-amyloid deposition

A one-way t-test comparison suggested that *APOE-ε4* carriers and non-carriers did not have significantly ($p=0.60$) different rates of deposition in those above the threshold for Aβ-amyloid at baseline (mean rates of deposition of 0.03 ± 0.02 and 0.03 ± 0.02 SUVR/year, respectively; equivalent to 1.7%/year), (Figure 1A). However, prior to reaching the abnormal threshold AIBL *APOE-ε4* carriers appeared to have had significantly ($p=0.005$) faster rates of Aβ-amyloid deposition (0.02 ± 0.02 SUVR/year; 2.1%/year) in comparison to AIBL *APOE-ε4* non-carriers (0.01 ± 0.01 SUVR/year; 1.1%/year), (Figure 1B).

A one-way t-test comparison of individual ADNI participants' rate of deposition of Aβ-amyloid suggest that *APOE-ε4* carriers and non-carriers did not have significantly different rates of deposition either after or prior to reaching the abnormal threshold ($p=0.99$ and $p=0.82$, respectively). The mean rates of Aβ-amyloid deposition for ADNI participants beyond the abnormal threshold were 0.02 ± 0.01 SUVR/year (2.2%/year) for both *APOE-ε4* carriers and non-carriers, (Figure 1C). Prior to reaching the abnormal threshold, ADNI *APOE-ε4* carriers and non-carriers both had rates of Aβ-amyloid deposition of 0.01 ± 0.005 SUVR/year (1.3%/year; Figure 1D).

In a sensitivity analysis considering only the NC participants, the findings were equivalent with the only statistically significant difference ($p=0.001$) being found in the AIBL participants below the threshold (Supplementary Figure 1).

Including the non-accumulators to the full data set resulted in no significant differences between *APOE-ε4* carriers and non-carriers either above or below the threshold, for AIBL or ADNI participants (Supplementary Figure 2).

3.3 Natural history of neocortical Aβ-amyloid deposition

Stratifying the natural history of neocortical Aβ-amyloid deposition by *APOE-ε4* carriage indicated that on average AIBL *APOE-ε4* carriers reached the abnormal threshold 14.9 (0.3-35.2) years prior to AIBL non-carriers, (Figure 2A). Similarly, on average ADNI *APOE-ε4* carriers reached the abnormal threshold 18.9 (CI: 3.5-40.1) years prior to ADNI non-carriers, (Figure 2B). Plots for individuals' longitudinal data (Step 1 in the method) and slope vs mean plots (Step 2) stratified by *APOE-ε4* carriage are provided in Supplementary Figure 3 for AIBL and Supplementary Figure 4 for ADNI. Note: when stratified by sex females appeared to reach the abnormal threshold 2 years prior to males but this was not statistically significant, results not presented.

Replicating this in a sensitivity analysis of the NC, indicated that on average NC AIBL *APOE-ε4* carriers reached the abnormal threshold 11.1 (–3.9–34.6) years prior to CN AIBL non-carriers, (Supplementary Figure 5). Please note that due to the small numbers of CN in the ADNI cohort, specifically *APOE-ε4* carriers (N=37) the models did not converge, and results are not presented.

3.4 Age of onset using survival analysis

Survival analysis indicated that 50% of the AIBL and ADNI participants reached abnormal levels of Aβ-amyloid by ages of 69.3 (66.5–73.5) and 73.6 (CI: 71.2–77.1), respectively, (Figures 3A and B). Stratifying the participants by *APOE-ε4* carriage and replicating the survival analysis indicated 50% of *APOE-ε4* carriers reached abnormal levels of Aβ-amyloid by ages 62.0 (CI: 59.6–66.5) and 65.1 (CI: 62.0–70.3) in AIBL and ADNI, respectively. In contrast, 50% of the *APOE-ε4* non-carriers reached abnormal levels of Aβ-amyloid by ages 77.2 (CI 76.1–NA) and 79.3 (CI 75.9–84.4) in AIBL and ADNI, respectively. These findings suggest that on average *APOE-ε4* carriers reached abnormal levels of Aβ-amyloid 15.2 years prior to *APOE-ε4* non-carriers in AIBL and 14.2 (2.5–20.5) years in ADNI (Figures 3C and D).

In the CN sub-groups, 50% of CN *APOE-ε4* carriers reached abnormal levels of Aβ-amyloid by ages 66.2 (CI: 63.6–76.1) and 66.4 (CI: 63.8–NA) in AIBL and ADNI, respectively. In contrast, 50% of the CN *APOE-ε4* non-carriers reached abnormal levels of Aβ-amyloid by ages 77.6 (CI 71.6–NA) and 79.3 (CI 76.7–NA) in AIBL and ADNI, respectively (Supplementary Figure 6).

4. Discussion

Survival analyses indicated the average age that AIBL and ADNI participants reached abnormal levels of neocortical Aβ-amyloid was seventy years of age, with confidence intervals (CI) ranging from 66 to 77 years of age. Stratifying the survival analyses by *APOE-ε4* carriage suggested that on average *APOE-ε4* carriers reached the abnormal threshold in their early sixties, 15 (CI: 6–24) years earlier than non-carriers who reached the threshold late in their seventies. Further, evaluation of the natural history of deposition of neocortical Aβ-amyloid also suggested that *APOE-ε4* carriers reached the abnormal threshold of neocortical Aβ-amyloid deposition approximately 15–19 (CI: 4–40) years prior to non-carriers, in line with previous findings (Bilgel et al., 2019; Fleisher et al., 2013; Mishra et al., 2018).

When restricting the analysis to only consider the cognitively normal participants, cognitively normal *APOE-ε4* carriers reached the abnormal threshold in their mid-sixties, 12 (CI: 0–24) years earlier than cognitively normal non-carriers who reached the threshold in their midseventies.

It is noted that whilst the age of onset and natural history analyses are not independent, there was exceptional consistency in the findings across the methods as well as across the two cohort studies, despite the use of different Aβ-amyloid tracers. The findings are also consistent with literature looking at the clinical onset of AD which reports *APOE-ε4*

carriage moves the age of clinical onset earlier by 10-20 years in comparison to non-carriers (Bilgel et al., 2016; Corder et al., 1993; Jack et al., 2014; Jansen et al., 2015).

Based on group comparisons, *APOE-ε4* carriers and non-carriers appeared to have similar rates of neocortical Aβ-amyloid deposition, with the only exception being AIBL participants prior to reaching the threshold for neocortical Aβ-amyloid. In this group *APOE-ε4* carriers appeared to have significantly faster rates of deposition than non-carriers.

Overall, the findings presented in this paper suggest that the natural history of neocortical Aβ-amyloid deposition in *APOE-ε4* carriers starts approximately 15 years earlier but has a similar trajectory to that of *APOE-ε4* non-carriers. For the same burden of neocortical Aβ-amyloid, the rate of deposition is similar for both *APOE-ε4* carriers and non-carriers (demonstrated by drawing horizontal lines through Figures 2 A and B). These findings fit with the previous literature that *APOE-ε4* carriage is not associated with the rate of disease progression, only with earlier onset of disease (Corder et al., 1995; Resnick et al., 2015; Saunders, 2000). Further, they go some way to explaining the conflicting reports that *APOE-ε4* carriage is also associated with rate of deposition and/or disease progression (Bilgel et al., 2019; Craft et al., 1998; Hoyt et al., 2005; Jack et al., 2013a; Lim et al., 2017; Mishra et al., 2018; Toledo et al., 2019; Villemagne et al., 2011): if an age matched population was considered (or age corrected modelling used) then the rate of deposition would appear to be higher in *APOE-ε4* carriers versus non-carriers. This would be due to *APOE-ε4* carriers being 15 years further along in disease progression and having higher neocortical Aβ-amyloid burden as well as potentially higher rates of deposition (demonstrated by drawing vertical lines through Figures 2 A and B). Therefore, the difference in rate of deposition between *APOE-ε4* carriers and non-carriers previously reported in the literature may be a function of a difference in disease stage opposed to a difference in *APOE-ε4* carriage. The temporal relationship between onset and rate is an important consideration and previous evaluations considering these as independent factors or not utilising longitudinal data may have limited their ability to draw valid conclusions.

When stratifying by sex, no significant differences between males and females were observed in the natural history evaluations. As the effect of sex was of a much smaller magnitude at 2 years than that of *APOE-ε4* at 15 years, it is possible that this study was not powered to observe a statistically significant difference.

This study has a number of other limitations. Firstly, there were not enough *APOE-ε4* homozygotes to enable evaluations on the dose-effect of *APOE-ε4* genotype to be undertaken. Secondly, a lack of *APOE-ε2* carriers prevented further evaluations to understand the implications of *APOE-ε2* carriage and its interplay with *APOE-ε4* carriage. Thirdly, given the focus on rates of Aβ-amyloid deposition, only accumulators were included in most of this study which may contrast with other reports and might preclude the generalisability of the findings. Analysis of the small number of non-accumulators available resulted in loss of statistical significance of the difference in rates of change between AIBL *APOE-ε4* carriers and non-carriers prior to reaching the threshold, no other differences were found. Fourthly, the analysis is restricted to the longitudinal evaluation of neocortical Aβ-amyloid and it will be necessary to extrapolate this analysis to incorporate peripheral Aβ-

amyloid and large longitudinal tau studies once they become available. The participants were volunteers who were not randomly selected from the community, and were generally well educated, thus these findings might only be valid in similar cohorts and this limitation precludes the generalisation of the findings to the general population. Also, in view of the stringent selection criteria in both AIBL and ADNI, which excluded individuals with cerebrovascular disease or other dementias, the effect of other comorbidities on the trajectories might be underestimated. Lastly, longitudinal A β -amyloid levels were obtained from ^{11}C PiB PET imaging in AIBL and ^{18}F Florbetapir PET imaging in ADNI and while both underwent the same CapAIBL normalisation, differences in PET scanner and tracer kinetics may contribute a somewhat larger variance in the results.

It has been established that rates of neocortical A β -amyloid deposition impact disease progression (Villemagne et al., 2013), earlier onset of A β -amyloid deposition may therefore lead to earlier disease onset. Therefore, understanding the age-related, temporal, deposition of neocortical A β -amyloid as well as the impact of *APOE- ϵ 4* carriage has essential implications for understanding disease mechanisms and informing the timing for therapeutics and diagnostics (Ungar et al., 2014). This is of paramount importance when considering disease staging and/or clinical trial inclusion criteria, for instance clinical trials will potentially need to consider alternative recruitment criteria such as younger age ranges for *APOE- ϵ 4* carriers in comparison to non-carriers. The ability to accurately target individuals at appropriate stages of the disease for inclusion in relevant clinical trials could afford such trials a better chance of success in the quest to delay and prevent AD.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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References

- Bilgel M, An Y, Zhou Y, Wong DF, Prince JL, Ferrucci L, Resnick SM, 2016 Individual estimates of age at detectable amyloid onset for risk factor assessment. *Alzheimer's & Dementia* 12(4), 373–379.
- Bilgel M, Jedynak BM, Initiative, A.s.D.N., 2019 Predicting time to dementia using a quantitative template of disease progression. *Alzheimer's & Dementia: Diagnosis, Assessment & Disease Monitoring* 11, 205–215.
- Bougeat P, Villemagne V, Dore V, Brown B, Macaulay L, Martins R, Masters C, Ames D, Ellis K, Rowe C, Salvado O, Fripp J, 2015 Comparison of MR-less PiB SUVR quantification methods. *Neurobiology of Aging* 36(Supplement 1), 8.
- Budgeon CA, Murray K, Turlach BA, Baker S, Villemagne VL, Burnham SC, 2017 Constructing longitudinal disease progression curves using sparse, short-term individual data with an application to Alzheimer's disease. *Statistics in Medicine*.
- Chen K, Roontiva A, Thiyyagura P, Lee W, Liu X, Ayutyanont N, Protas H, Luo JL, Bauer R, Reschke C, 2015 Improved power for characterizing longitudinal amyloid-b PET changes and evaluating amyloid-modifying treatments with a cerebral white matter reference region. *J Nucl Med* 56(4), 560–566. [PubMed: 25745091]
- Clark CM, Pontecorvo MJ, Beach TG, Bedell BJ, Coleman RE, Doraiswamy PM, Fleisher AS, Reiman EM, Sabbagh MN, Sadowsky CH, 2012 Cerebral PET with florbetapir compared with

- neuropathology at autopsy for detection of neuritic amyloid- β plaques: a prospective cohort study. *The Lancet Neurology* 11(8), 669–678. [PubMed: 22749065]
- Corder E, Saunders A, Strittmatter W, Schmechel D, Gaskell P, Rimmler J, Locke P, Conneally P, Schmechel K, Tanzi R, 1995 Apolipoprotein E, survival in Alzheimer's disease patients, and the competing risks of death and Alzheimer's disease. *Neurology* 45(7), 1323–1328. [PubMed: 7617191]
- Corder E, Saunders A, Strittmatter W, Schmechel D, Gaskell P, Small G.a., Roses A, Haines J, Pericak-Vance MA, 1993 Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 261(5123), 921–923. [PubMed: 8346443]
- Craft S, Peskind E, Schwartz MW, Schellenberg GD, Raskind M, Porte D, 1998 Cerebrospinal fluid and plasma insulin levels in Alzheimer's disease Relationship to severity of dementia and apolipoprotein E genotype. *Neurology* 50(1), 164–168. [PubMed: 9443474]
- Fleisher AS, Chen K, Liu X, Ayutyanont N, Roontiva A, Thiyyagura P, Protas H, Joshi AD, Sabbagh M, Sadowsky CH, 2013 Apolipoprotein E ϵ 4 and age effects on florbetapir positron emission tomography in healthy aging and Alzheimer disease. *Neurobiology of aging* 34(1), 1–12. [PubMed: 22633529]
- Hardy J, Selkoe DJ, 2002 The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *science* 297(5580), 353–356. [PubMed: 12130773]
- Harold D, Abraham R, Hollingworth P, Sims R, Gerrish A, Hamshere ML, Pahwa JS, Moskva V, Dowzell K, Williams A, 2009 Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. *Nature genetics* 41(10), 1088. [PubMed: 19734902]
- Hoyt BD, Massman PJ, Schatschneider C, Cooke N, Doody RS, 2005 Individual Growth Curve Analysis of APOE ϵ 4-Associated Cognitive Decline in Alzheimer Disease. *Archives of neurology* 62(3), 454–459. [PubMed: 15767511]
- Jack CR, Bennett DA, Blennow K, Carrillo MC, Dunn B, Haeberlein SB, Holtzman DM, Jagust W, Jessen F, Karlawish J, 2018 NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease. *Alzheimer's & dementia: the journal of the Alzheimer's Association* 14(4), 535–562.
- Jack CR, Wiste HJ, Lesnick TG, Weigand SD, Knopman DS, Vemuri P, Pankratz VS, Senjem ML, Gunter JL, Mielke MM, 2013a Brain β -amyloid load approaches a plateau. *Neurology* 80(10), 890–896. [PubMed: 23446680]
- Jack CR, Wiste HJ, Weigand SD, Knopman DS, Lowe V, Vemuri P, Mielke MM, Jones DT, Senjem ML, Gunter JL, 2013b Amyloid-first and neurodegeneration-first profiles characterize incident amyloid PET positivity. *Neurology* 81(20), 1732–1740. [PubMed: 24132377]
- Jack CR, Wiste HJ, Weigand SD, Rocca WA, Knopman DS, Mielke MM, Lowe VJ, Senjem ML, Gunter JL, Preboske GM, 2014 Age-specific population frequencies of cerebral β -amyloidosis and neurodegeneration among people with normal cognitive function aged 50–89 years: a cross-sectional study. *The Lancet Neurology* 13(10), 997–1005. [PubMed: 25201514]
- Jansen WJ, Ossenkoppele R, Knol DL, Tijms BM, Scheltens P, Verhey FR, Visser PJ, Aalten P, Aarsland D, Alcolea D, 2015 Prevalence of cerebral amyloid pathology in persons without dementia: a meta-analysis. *Jama* 313(19), 1924–1938. [PubMed: 25988462]
- Karran E, Mercken M, De Strooper B, 2011 The amyloid cascade hypothesis for Alzheimer's disease: an appraisal for the development of therapeutics. *Nature reviews Drug discovery* 10(9), 698. [PubMed: 21852788]
- Lim YY, Mormino EC, initiative, A.s.D.N., 2017 APOE genotype and early β -amyloid accumulation in older adults without dementia. *Neurology* 89(10), 1028–1034. [PubMed: 28794245]
- Liu C-C, Kanekiyo T, Xu H, Bu G, 2013 Apolipoprotein E and Alzheimer disease: risk, mechanisms and therapy. *Nature Reviews Neurology* 9(2), 106–118. [PubMed: 23296339]
- Mishra S, Blazey TM, Holtzman DM, Cruchaga C, Su Y, Morris JC, Benzinger TL, Gordon BA, 2018 Longitudinal brain imaging in preclinical Alzheimer disease: impact of APOE ϵ 4 genotype. *Brain* 141(6), 1828–1839. [PubMed: 29672664]
- Potkin SG, Guffanti G, Lakatos A, Turner JA, Kruggel F, Fallon JH, Saykin AJ, Orro A, Lupoli S, Salvi E, 2009 Hippocampal atrophy as a quantitative trait in a genome-wide association study

identifying novel susceptibility genes for Alzheimer's disease. *PLoS one* 4(8), e6501. [PubMed: 19668339]

- R Development Core Team, 2017 R: A Language and Environment for Statistical Computing. Vienna, Austria.
- Reiman EM, Chen K, Liu X, Bandy D, Yu M, Lee W, Ayutyanont N, Keppler J, Reeder SA, Langbaum JB, 2009 Fibrillar amyloid- β burden in cognitively normal people at 3 levels of genetic risk for Alzheimer's disease. *Proceedings of the National Academy of Sciences* 106(16), 6820–6825.
- Resnick SM, Bilgel M, Moghekar A, An Y, Cai Q, Wang M-C, Thambisetty M, Prince JL, Zhou Y, Soldan A, 2015 Changes in A β biomarkers and associations with APOE genotype in 2 longitudinal cohorts. *Neurobiology of aging* 36(8), 2333–2339. [PubMed: 26004017]
- Rowe CC, Ellis KA, Rimajova M, Bourgeat P, Pike KE, Jones G, Fripp J, Tochon-Danguy H, Morandau L, O'Keefe G, Price R, Raniga P, Robins P, Acosta O, Lenzo N, Szoeka C, Salvado O, Head R, Martins R, Masters CL, Ames D, Villemagne VL, 2010 Amyloid imaging results from the Australian Imaging, Biomarkers and Lifestyle (AIBL) study of aging. *Neurobiology of Aging* 31(8, Sp. Iss. SI), 1275–1283. [PubMed: 20472326]
- Saunders AM, 2000 Apolipoprotein E and Alzheimer disease: an update on genetic and functional analyses. *Journal of Neuropathology & Experimental Neurology* 59(9), 751–758. [PubMed: 11005255]
- Saykin AJ, Shen L, Foroud TM, Potkin SG, Swaminathan S, Kim S, Risacher SL, Nho K, Huentelman MJ, Craig DW, 2010 Alzheimer's Disease Neuroimaging Initiative biomarkers as quantitative phenotypes: genetics core aims, progress, and plans. *Alzheimer's & dementia: the journal of the Alzheimer's Association* 6(3), 265–273.
- Toledo JB, Habes M, Sotiras A, Bjerke M, Fan Y, Weiner MW, Shaw LM, Davatzikos C, Trojanowski JQ, Initiative, A.s.D.N., 2019 APOE Effect on Amyloid- β PET Spatial Distribution, Deposition Rate, and Cut-Points. *Journal of Alzheimer's Disease*(Preprint), 1–11.
- Ungar L, Altmann A, Greicius MD, 2014 Apolipoprotein E, gender, and Alzheimer's disease: An overlooked, but potent and promising interaction. *Brain imaging and behavior* 8(2), 262–273. [PubMed: 24293121]
- Villemagne VL, Burnham S, Bourgeat P, Brown B, Ellis KA, Salvado O, Szoeka C, Macaulay SL, Martins R, Maruff P, 2013 Amyloid β deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study. *The Lancet Neurology* 12(4), 357–367. [PubMed: 23477989]
- Villemagne VL, Pike KE, Chetelat G, Ellis KA, Mulligan RS, Bourgeat P, Ackermann U, Jones G, Szoeka C, Salvado O, 2011 Longitudinal assessment of A β and cognition in aging and Alzheimer disease. *Annals of neurology* 69(1), 181–192. [PubMed: 21280088]

Highlights:

APOE-ε4 carriers reached abnormal levels of Aβ ~15 years earlier than non-carriers

APOE-ε4 carriers and non-carriers had no differences in Aβ deposition rates beyond the threshold

Primary and secondary prevention trials should target *APOE-ε4* carriers aged between 60 and 66

Primary and secondary prevention trials should target *APOE-ε4* non-carriers aged between 76 and 84

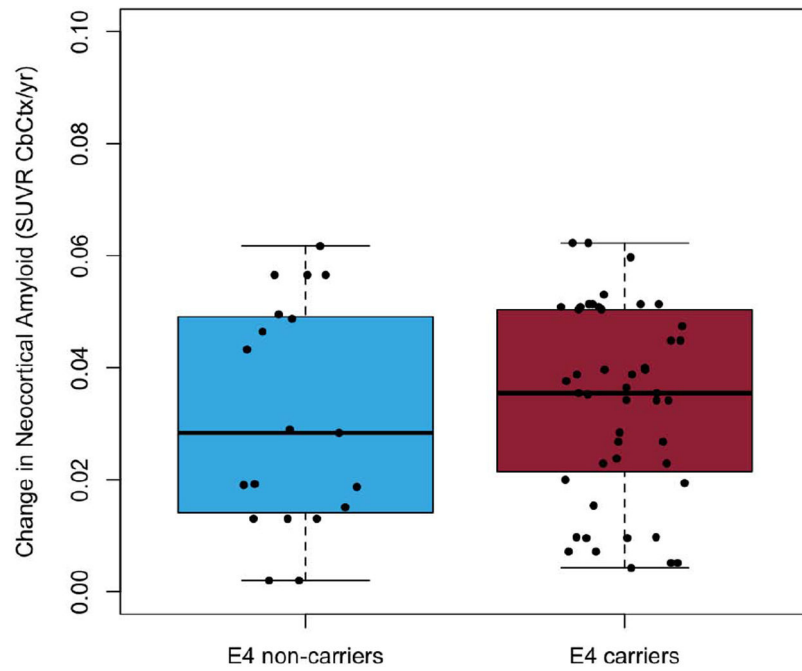


Figure 1A. Boxplots detailing the rates of A β -amyloid deposition for AIBL participants above the abnormal threshold for A β -amyloid at baseline (^{11}C -PiB PET SUVR 1.4) stratified by *APOE*- ϵ 4 carriage

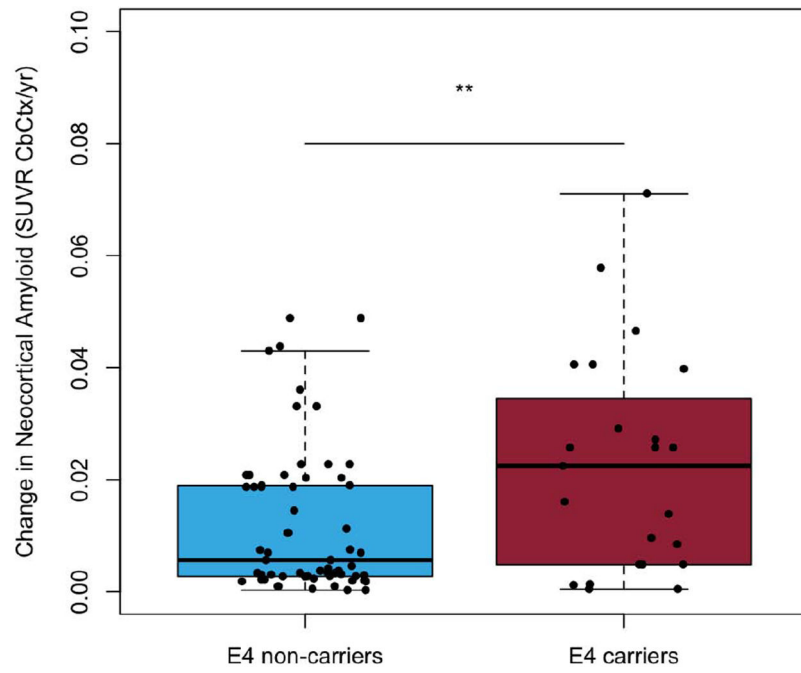


Figure 1B. Boxplots detailing the rates of A β -amyloid deposition for AIBL participants below the abnormal threshold for A β -amyloid at baseline (^{11}C -PiB PET SUVR 1.4) stratified by *APOE*- ϵ 4 carriage

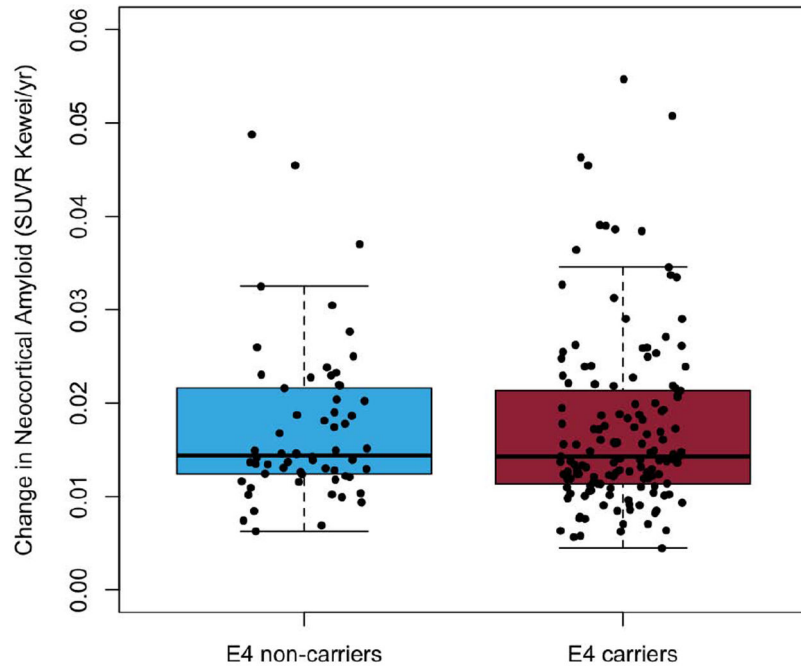


Figure 1C. Boxplots detailing the rates of A β -amyloid deposition for ADNI participants above the abnormal threshold for A β -amyloid at baseline (^{18}F -Florbetapir SUVr 0.61) stratified by *APOE*- ϵ 4 carriage

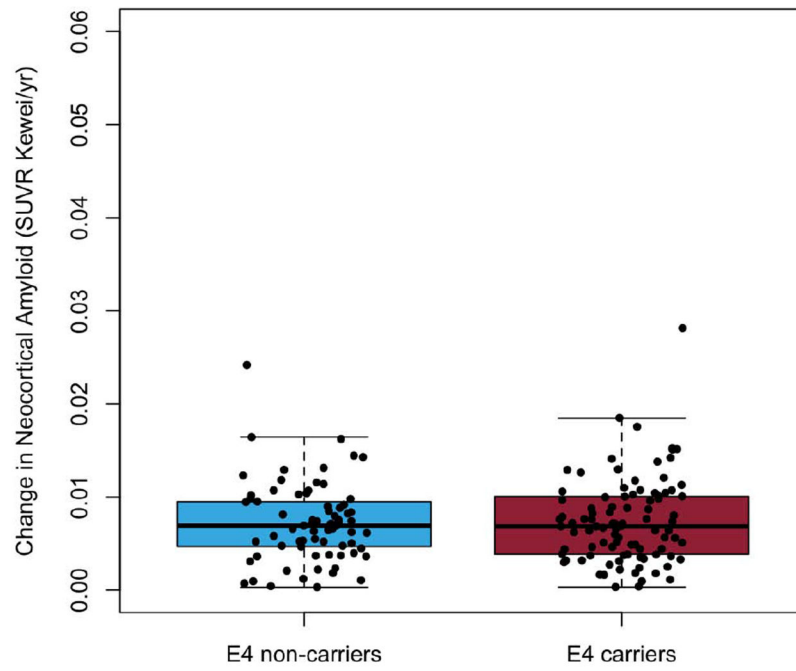


Figure 1D. Boxplots detailing the rates of A β -amyloid deposition for ADNI participants below the abnormal threshold for A β -amyloid at baseline (^{18}F -Florbetapir SUVR 0.61) stratified by *APOE*- ϵ 4 carriage

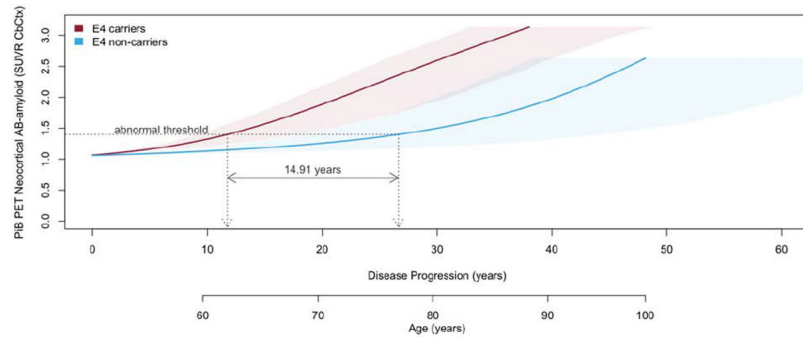


Figure 2A.
The natural history of deposition of neocortical A β -amyloid in AIBL participants stratified by *APOE*- ϵ 4 carriage. Shaded areas indicate 95% confidence intervals.

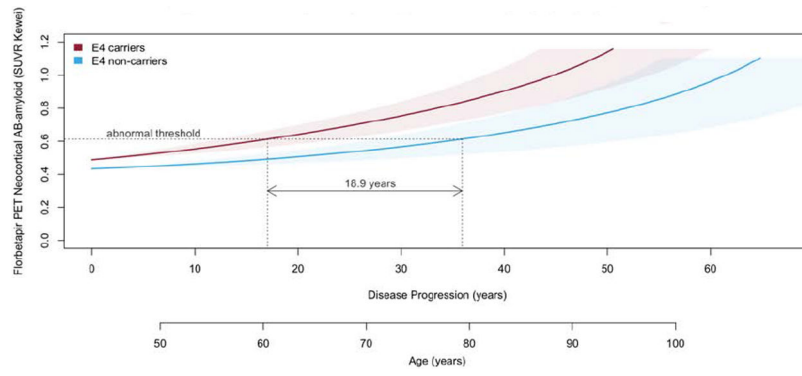


Figure 2B. The natural history of deposition of neocortical A β -amyloid in ADNI participants stratified by *APOE*- ϵ 4 carriage. Shaded areas indicate 95% confidence intervals.

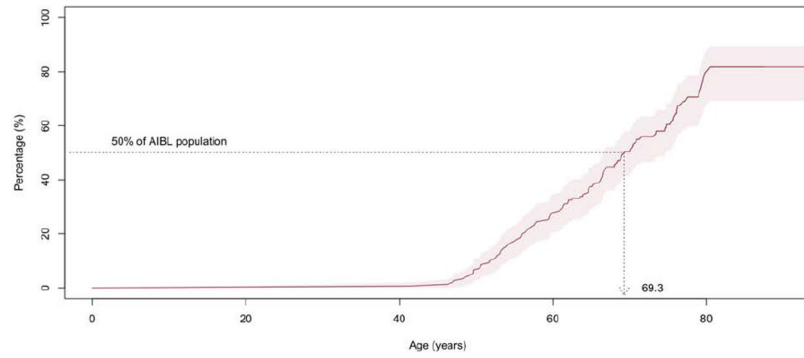


Figure 3A. Kaplan-Meier plot detailing, by age, the prevalence of AIBL participants with high levels of Aβ-amyloid at baseline (^{11}C -PiB PET SUVR ≥ 1.4). Shaded areas indicate 95% confidence intervals.

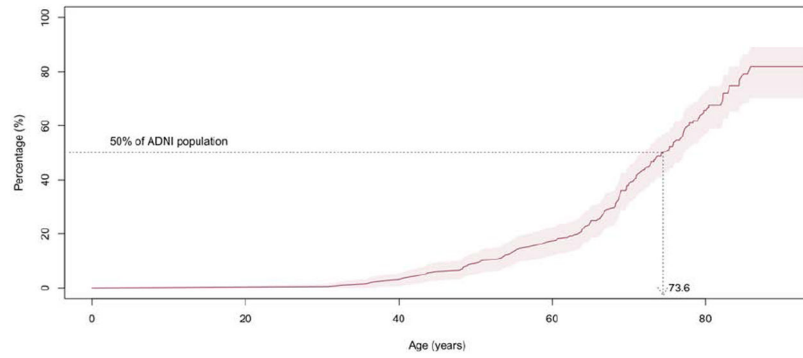


Figure 3B. Kaplan-Meier plot detailing, by age, the prevalence of ADNI participants with high levels of Aβ-amyloid at baseline (^{18}F -Florbetapir SUVR ≥ 0.61). Shaded areas indicate 95% confidence intervals.

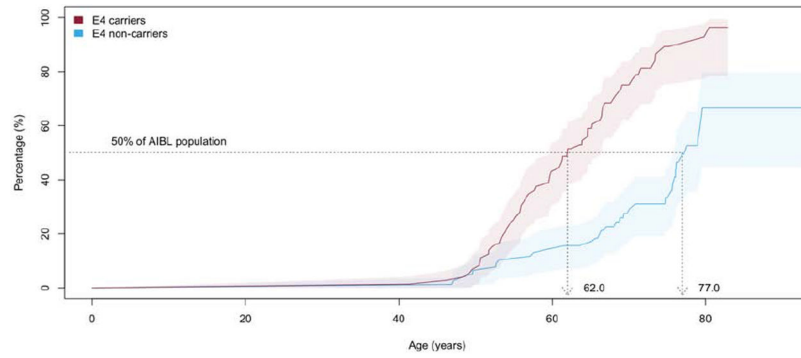


Figure 3C. Kaplan-Meier plot detailing, by age, the prevalence of AIBL participants with high levels of Aβ-amyloid at baseline (^{11}C -PiB PET SUVR ≥ 1.4) stratified by *APOE*- $\epsilon 4$ carriage. Shaded areas indicate 95% confidence intervals.

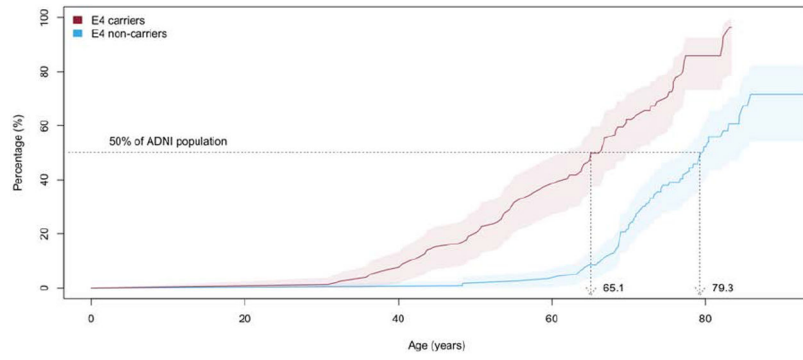


Figure 3D. Kaplan-Meier plot detailing, by age, the prevalence of ADNI participants with high levels of Aβ-amyloid at baseline (^{18}F -Florbetapir SUVR 0.61) stratified by *APOE*-ε4 carriage. Shaded areas indicate 95% confidence intervals.

Table 1.

Demographics table for AIBL and ADNI participants stratified by *APOE-ε4* carriage

| | <i>APOE-ε4</i> carriage in AIBL | | p-value | <i>APOE-ε4</i> carriage in ADNI | | p-value |
|---------------------------------------|---------------------------------|--------------|---------|---------------------------------|--------------|---------|
| | No | Yes | | No | Yes | |
| Number of Participants [N] | 123 | 86 | | 142 | 104 | |
| Clinical Classification NC/MCI/AD [N] | 102/15/6 | 54/16/16 | 0.001 | 69/73/0 | 37/67/0 | 0.057 |
| Gender: Males [N (%)] | 51 (41.46) | 50 (58.14) | 0.026 | 62 (43.66) | 49 (47.12) | 0.683 |
| Age (years) [mean (sd)] | 72.48 (6.99) | 71.18 (7.23) | 0.196 | 72.82 (6.83) | 70.33 (6.87) | 0.005 |
| Years of Education [N (%)] | | | | | | |
| <9 | 8 (6.5) | 5 (5.81) | 0.952 | 0 (0) | 1 (0.96) | 0.620 |
| 9-12 | 50 (40.65) | 38 (44.19) | | 20 (14.09) | 12 (11.54) | |
| 13-15 | 24 (19.51) | 17 (19.77) | | 29 (20.42) | 20 (19.23) | |
| >15 | 41 (33.33) | 26 (30.23) | | 93 (65.49) | 71 (68.27) | |
| Years of Follow-up [Mean (sd)] | 6.93 (1.19) | 6.68 (1.35) | 0.149 | 5.00 (1.20) | 5.09 (1.31) | 0.708 |