

Published in final edited form as:

J Am Coll Cardiol. 2012 October 16; 60(16): 1546–55. doi:10.1016/j.jacc.2012.05.052.

Quantification of Absolute Myocardial Perfusion in Patients With Coronary Artery Disease: Comparison Between Cardiovascular Magnetic Resonance and Positron Emission Tomography

Geraint Morton, MA, MBBS*, Amedeo Chiribiri, MD, PhD*, Masaki Ishida, MD, PhD*, Shazia T. Hussain, MBChB*, Andreas Schuster, MD, PhD*, Andreas Indermuehle, MD, PhD*, Divaka Perera, MA, MD[†], Juhani Knuuti, MD, PhD[‡], Stacey Baker, BSc*, Erik Hedström, MD, PhD*, Paul Schleyer, BSc*, Michael O'Doherty, MBBS, MD*, Sally Barrington, MBBS, MD*, Eike Nagel, MD, PhD*

London, United Kingdom; and Turku, Finland *King's College London British Heart Foundation Centre of Excellence; National Institute of Health Research (NIHR) Biomedical Research Centre at Guy's and St. Thomas' NHS Foundation Trust; Wellcome Trust and Engineering and Physical Sciences Research Council (EPSRC) Medical Engineering Centre; Division of Imaging Sciences; The Rayne Institute, St. Thomas' Hospital, London, United Kingdom [†]King's College London BHF Centre of Excellence, NIHR Biomedical Research Centre and Department of Cardiology, Guy's and St. Thomas' NHS Foundation Trust, London, United Kingdom [‡]Turku PET Centre, University of Turku and Turku University Hospital, Turku, Finland

Abstract

Objectives—The aim of this study was to compare fully quantitative cardiovascular magnetic resonance (CMR) and positron emission tomography (PET) myocardial perfusion and myocardial perfusion reserve (MPR) measurements in patients with coronary artery disease (CAD).

Background—Absolute quantification of myocardial perfusion and MPR with PET have proven diagnostic and prognostic roles in patients with CAD. Quantitative CMR perfusion imaging has been established more recently and has been validated against PET in normal hearts. However, there are no studies comparing fully quantitative CMR against PET perfusion imaging in patients with CAD.

Methods—Forty-one patients with known or suspected CAD prospectively underwent quantitative ¹³N-ammonia PET and CMR perfusion imaging before coronary angiography.

Results—The CMR-derived MPR (MPR_{CMR}) correlated well with PET-derived measurements (MPR_{PET}) ($r = 0.75$, $p < 0.0001$). MPR_{CMR} and MPR_{PET} for the 2 lowest scoring segments in each coronary territory also correlated strongly ($r = 0.79$, $p < 0.0001$). Absolute CMR perfusion values correlated significantly, but weakly, with PET values both at rest ($r = 0.32$; $p = 0.002$) and during stress ($r = 0.37$; $p < 0.0001$). Area under the receiveroperating characteristic curve for

Correspondence to: Eike Nagel.

Reprint requests and correspondence: Dr. Eike Nagel, Division of Imaging Sciences, The Rayne Institute, Fourth Floor Lambeth Wing, St. Thomas' Hospital, London SE1 7EH, United Kingdom. eike.nagel@kcl.ac.uk.

MPR_{PET} to detect significant CAD was 0.83 (95% confidence interval: 0.73 to 0.94) and for MPR_{CMR} was 0.83 (95% confidence interval: 0.74 to 0.92). An MPR_{PET} 1.44 predicted significant CAD with 82% sensitivity and 87% specificity, and MPR_{CMR} 1.45 predicted significant CAD with 82% sensitivity and 81% specificity.

Conclusions—There is good correlation between MPR_{CMR} and MPR_{PET} . For the detection of significant CAD, MPR_{PET} and MPR_{CMR} seem comparable and very accurate. However, absolute perfusion values from PET and CMR are only weakly correlated; therefore, although quantitative CMR is clinically useful, further refinements are still required. (*J Am Coll Cardiol* 2012;60:1546-55) © 2012 by the American College of Cardiology Foundation

Keywords

coronary disease; ischemia; magnetic resonance imaging; perfusion; positron emission tomography

Noninvasive assessment of myocardial perfusion is clinically important, particularly for the detection and management of patients with coronary artery disease (CAD). Currently, clinical decision making is usually based on visual estimates of relative perfusion. However, this approach relies on the presence of a normally perfused region of myocardium. Quantification of perfusion addresses this limitation, and positron emission tomography (PET) studies have confirmed that it provides incremental value compared with nonquantitative methods (1).

Positron emission tomography can accurately quantify perfusion and myocardial perfusion reserve (MPR) and is currently the noninvasive reference standard. Fully quantitative methods are accurate for the detection of CAD (1) and can result in improved diagnostic accuracy compared with relative perfusion analysis (2). Recent studies have also demonstrated that an abnormal quantitative MPR is an independent predictor of an adverse prognosis (3,4). Furthermore, quantitative data provide unique information about the coronary microcirculation that is not available from nonquantitative methods (5).

Cardiac magnetic resonance (CMR) has, relatively recently, established itself as an accurate and valuable tool for perfusion imaging (6). Currently, visual assessment is standard; however, absolute quantification of perfusion has also been demonstrated to be feasible and accurate (7–10). Quantitative CMR methods have been validated against microspheres in animals (7), coronary sinus flow in patients (9), and PET in healthy volunteers (11–13). However, to date, there is a lack of evidence comparing quantitative CMR with PET in patients with CAD. Patient studies are clinically more relevant, given the inherent differences between patients and healthy volunteers. Therefore, the primary aim of this study was to compare quantitative assessment of myocardial perfusion and MPR with CMR and PET in a cohort of patients with known or suspected CAD.

Methods

Patient population and study design

Patients with a history of stable angina and known or suspected CAD underwent CMR and PET perfusion imaging before planned x-ray coronary angiography. Exclusion criteria: acute coronary syndrome <6 weeks, previous coronary artery bypass graft, previous ST-segment elevation infarction, estimated glomerular filtration rate <30 ml/min, or contraindication to magnetic resonance imaging or adenosine. The local ethics committee approved the study, and the U.K. Administration of Radioactive Substances Advisory Committee licensed radiation exposure. All patients gave written informed consent.

Data acquisition

Patients were instructed to abstain from caffeine and smoking for 24 h before imaging.

CMR imaging

Data were acquired with a 1.5-T scanner (Achieva, Philips, Best, the Netherlands) and a 32-channel coil. Examinations included high-resolution perfusion and functional and scar imaging. Perfusion imaging consisted of 3 short-axis slices acquired every heartbeat covering 16 of the standard myocardial segments (14) (apex excluded). Stress imaging preceded rest by 14 ± 2 min (range 10 to 19 min). Imaging parameters: *k-t* balanced turbo gradient echo sequence, shortest echo time (range 1.35 to 1.54 ms), shortest repetition time (range 2.64 to 3.12 ms), 50° flip angle; 90° prepulse, 100-ms prepulse delay, and typical acquired resolution $1.7 \times 1.9 \times 10$ mm. For stress imaging, 140 $\mu\text{g}/\text{kg}/\text{min}$ of adenosine was administered intravenously for 4 min. Imaging commenced 3 min into the infusion and continued for 1 min. A dual bolus (equal volumes of 0.01 mmol/kg followed by 0.1 mmol/kg after a 20-s pause) of contrast agent (gadobutrol/Gadovist, Berlin-Wedding, Schering, Germany) was injected at 4ml/s by a power injector for perfusion imaging (15).

PET imaging

This was performed with a GE Discovery VCT PET-CT scanner (GE Healthcare, Waukesha, Wisconsin) after administration of ^{13}N ammonia, with 47 transaxial slices reconstructed over an axial field of view of 15 cm. Acquired resolution was $4.8 \times 4.8 \times 4.9$ mm. Computed tomography (CT) scout data determined patient position, and a low-dose CT was used for attenuation correction. Acquisition consisted of dynamic scans from 0 to 6 min (12×10 s, 6×20 s, and 2×60 s) and then a single static scan frame for 20 min. For the rest study, a total of 550 MBq of ^{13}N -ammonia was injected intravenously. Stress imaging was performed approximately 50 min later. Adenosine (140 $\mu\text{g}/\text{kg}/\text{min}$) was administered intravenously for 6 min, with a second equivalent dose of ^{13}N -ammonia administered 2 min into the infusion. Patients remained on the scanner table throughout the entire study; however, attenuation correction was repeated for the stress study.

X-ray coronary angiography

This was performed according to the standard Judkin's technique. Multiple projections of the coronary arteries were acquired, including at least 2 orthogonal views to assess stenosis severity.

Visual analysis

Studies were analyzed by 2 independent experts blinded to all other data. The PET scans were classified as positive for CAD in the presence of a stress-induced perfusion defect involving 2 myocardial segments and CMR scans in the presence of a stress-induced perfusion defect, which was transmural or involved 2 myocardial segments. In the case of disagreement between the observers, the images were reviewed together, and a consensus was reached.

Quantitative analysis

Quantitative analysis was performed by 1 of the experts. Perfusion was quantified in 16 standard American Heart Association segments. Segments were defined separately for the PET and CMR images with the right ventricular insertion points for reference. Segment 17 was excluded. Myocardial border detection was automated and manually corrected where required. Segments unsuitable for analysis due to artifact or poor image quality were excluded. The CMR segments demonstrating late gadolinium enhancement and PET data from patients with >10-mm movement between the CT and PET were considered unreliable and also excluded. Coronary territories were excluded if all relevant segments were unsuitable for analysis.

PET

Original dynamic raw PET scans were used to calculate arterial input function and mean segmental perfusion calculated over the linear portion of the curve from 70 to 210 s, with Quick Cardiac (Hermes Medical Solutions, Stockholm, Sweden) in conjunction with software developed at our institution based on the Patlak method as previously described and validated (16).

CMR

Mean segmental perfusion values were obtained with dedicated ViewForum software (Philips) and the previously validated Fermi deconvolution method (7).

PET AND CMR

Each segment was assigned to the appropriate perfusion territory (14), with segment 15 assigned to the dominant coronary artery (defined by the observer analyzing the angiogram). Myocardial perfusion reserve was defined as stress perfusion divided by rest perfusion and was calculated for each segment and territory. For CAD detection, mean MPR of the 2 lowest scoring segments for each perfusion territory (MPR2) was used for further analysis as described previously (17).

X-RAY ANGIOGRAPHY

All coronary arteries >2 mm in diameter were assessed by an independent cardiologist blinded to all other data. Stenoses <30% and >95% on visual analysis were judged to be nonflow-limiting or flowlimiting, respectively, without further assessment. Quantitative coronary angiography (QCA) was performed for all arteries with a visual stenosis severity of 30% to 95% with a dedicated software program (Medcon UK, Edware, Middlesex, United Kingdom). Mean diameter stenosis from 2 orthogonal views was recorded, and 70% was regarded as significant. Coronary territories subtended by a coronary artery with 70% stenosis were classified as stenotic territories, and all others were classified as remote.

Statistical analysis

The IBM SPSS Statistics (version 19.0, SPSS, Chicago, Illinois) and Medcalc software (Medcalc, Mariakerke, Belgium) were used. Data are presented as mean \pm SD, except where stated. Shapiro-Wilk analysis defined when nonparametric tests were required. Agreement and correlations between PET and CMR were determined with Bland-Altman plots and Spearman's test of correlation with a 2-tailed test of significance, respectively. Analyses were performed on a per-observation basis. Because 3 perfusion territories were analyzed/patient, analyses were repeated within each territory to ensure that any strong correlations did not simply reflect high within-subject correlations. Intraobserver and interobserver reproducibility was determined by a coefficient of variation (CV): SD of the differences divided by the mean. Paired and independent *t* tests were used for comparison of paired and unpaired mean data, respectively. Receiver-operating characteristic (ROC) analysis determined the accuracy of visual analysis, MPR2, and stress perfusion values from PET and CMR for predicting a corresponding coronary artery stenosis of 70% on QCA. Optimal cutoffs were determined by the maximum Youden Index. Logistic regression models were used to calculate the area under the receiver-operating characteristic curve (AUC) when information from both CMR and PET were combined, and this was compared with the ROC curves when CMR or PET was used alone. Sensitivity and specificity were compared with McNemar's test. Significance was determined at <0.05.

Results

Study population

Forty-one patients were recruited. Table 1 shows baseline characteristics. The study protocol was completed in 38 patients (exclusions: 1 atrioventricular block, 1 withdrawal of consent, 1 claustrophobia), and further analysis relates to these patients. Hemodynamic status during imaging is shown in Table 2. Resting heart rate was significantly higher during CMR studies (66 vs. 63 beats/min, respectively). There were no significant differences between rate pressure product, systolic blood pressure, or stress heart rate during CMR and PET studies.

The interval between PET and CMR scans was 3 ± 6 days; 26 (63%) patients underwent both on the same day. The coronary angiogram was 17 ± 19 days and 17 ± 21 days after the CMR and PET, respectively. Twenty-five patients (61%) had 1 stenosis of 70% diameter. Nineteen had single-vessel disease, 5 had 2-vessel disease, and 1 had 3-vessel disease. Left ventricular ejection fraction (CMR) was $63 \pm 12\%$.

Myocardial perfusion

All perfusion studies were suitable for visual analysis. Quantitative data were available for 89%, 84%, and 73% of territories with CMR, PET, and both modalities, respectively. Results are summarized in Table 3. In patients with CAD stress perfusion, MPR and MPR₂ were significantly lower in stenotic territories compared with remote territories with both PET and CMR. Conversely, rest perfusion was not significantly different.

The MPR₂_{PET} was 1.36 ± 0.32 and 1.74 ± 0.32 in stenotic and remote territories, respectively ($p < 0.0001$). The MPR₂_{CMR} values were 1.31 ± 0.30 and 1.70 ± 0.42 ($p < 0.0001$). In patients without significant CAD, MPR₂_{PET} was 1.92 ± 0.39 , and MPR₂_{CMR} was 1.93 ± 0.53 ($p = 0.14$ and $p = 0.16$, respectively, compared with remote territories in patients with CAD).

Agreement between CMR and PET

There was good correlation between MPR_{CMR} and MPR_{PET} ($r = 0.75$; $p < 0.0001$) (Fig. 1A) and between MPR₂_{CMR} and MPR₂_{PET} ($r = 0.79$; $p < 0.0001$) (Fig. 1B). A Bland-Altman plot demonstrates good agreement between MPR_{CMR} and MPR_{PET} (Fig. 2). Absolute CMR perfusion values correlated significantly, but weakly, with PET values both at rest ($r = 0.32$; $p = 0.002$) and during stress ($r = 0.37$; $p < 0.0001$) (Fig. 3). Results were similar when analyses were repeated within each territory. The MPR_{CMR} correlated well with MPR_{PET} in the left anterior descending coronary artery ($r = 0.79$; $p < 0.0001$), circumflex ($r = 0.64$; $p < 0.0001$), and right coronary artery territories ($r = 0.77$; $p < 0.0001$). Bland-Altman limits of agreement for MPR_{CMR} and MPR_{PET} also remained similar in the left anterior descending coronary artery (−0.71 to 0.58), circumflex (−0.7 to 0.8), and right coronary artery territories (−0.62 to 0.48).

Reproducibility

Intraobserver CV for PET stress perfusion, rest perfusion, and MPR were 9%, 12%, and 15%, respectively. Corresponding CV for inter-observer reproducibility were 17%, 13%, and 18%, respectively. The CMR intraobserver CV were 9%, 14%, and 20%, respectively, and interobserver CV were 16%, 18%, and 22%, respectively.

Diagnosis of CAD. VISUAL ANALYSIS

Sensitivity and specificity against QCA were: PET 92% (95% confidence interval [CI]: 72% to 99%) and 69% (95% CI: 41% to 88%); CMR 86% (95% CI: 64% to 96%) and 76% (95% CI: 50% to 92%). The CMR and PET sensitivity and specificity were not significantly different ($p = 0.65$ and 0.71 , respectively). The AUC were: PET 0.81 (95% CI: 0.66 to 0.96); CMR 0.81 (95% CI: 0.65 to 0.96). If only those patients with quantitative data suitable for analysis were considered, sensitivity and specificity changed to 94% (95% CI: 71% to 99%) and 67% (95% CI: 39% to 87%), and 85% (95% CI: 61% to 96%) and 79% (95% CI: 49% to 94%) for PET and CMR, respectively.

Quantitative Analysis

The AUC for MPR_{2PET} to detect significant CAD was 0.83 (95% CI: 0.73 to 0.94) and for MPR_{2CMR} was 0.83 (95% CI: 0.74 to 0.92) ($p = 0.96$). Sensitivity and specificity against QCA were: MPR_{2PET} 1.44: 82% and 87%; and MPR_{2CMR} 1.45: 82% and 81% (Fig. 4). The AUC was not significantly different from AUC for visual analysis ($p = 0.73$ and $p = 0.79$ for PET and CMR, respectively). An example case is shown in Figure 5.

The AUC was 0.85 (95% CI: 0.75 to 0.95) for MPR_{2PET} and MPR_{2CMR} combined. There was no significant difference between AUC when MPR_{2PET} and MPR_{2CMR} were both included in the model and when MPR_{2PET} ($p = 0.406$) or MPR_{2CMR} ($p = 0.4749$) alone were used.

The MPR_{2PET} and MPR_{2CMR} were inversely related to the severity of CAD by QCA. For QCA subgroups <30%, 30% to 49%, 50% to 69%, 70% to 95%, and >95% MPR_{2PET} and MPR_{2CMR} were 1.93 ± 0.39 and 1.89 ± 0.46 , 1.73 ± 0.33 and 1.86 ± 0.67 , 1.58 ± 0.4 and 1.54 ± 0.47 , 1.49 ± 0.33 and 1.28 ± 0.42 , and 1.18 ± 0.20 and 1.26 ± 0.22 , respectively.

The AUC for absolute perfusion values during stress to detect significant CAD were: PET 0.69 (95% CI: 0.56 to 0.81) and CMR 0.72 (95% CI: 0.61 to 0.83). This was significantly lower than MPR₂ AUC with PET ($p = 0.049$) but not CMR ($p = 0.12$). Optimal absolute stress perfusion cutoffs were: PET 1.48 ml/min/g (sensitivity 80%, specificity 53%); and CMR 1.50 ml/min/g (sensitivity 63%, specificity 76%).

Discussion

We have demonstrated that, in patients with known or suspected CAD, there is good correlation between MPR_{PET} and MPR_{CMR}. The MPR_{CMR} also seems to have a similar sensitivity and specificity to MPR_{PET} for the detection of significant coronary artery stenoses. However, the correlation between the absolute perfusion values from PET and CMR is relatively weak.

To our knowledge, there are no previously published studies comparing quantitative CMR myocardial perfusion with PET in patients. Previous studies have been limited to semi quantitative analysis methods (18,19) or have involved healthy volunteers (11–13). Semi-quantitative analysis has been shown to be useful (20) and to correlate well with MPR_{PET} in patients with CAD (18,19). However, unlike fully quantitative analysis, these methods only provide an index of perfusion reserve and not a true MPR and might substantially underestimate perfusion, particularly at higher perfusion values (19).

Studies using fully quantitative CMR methods are limited to small numbers of healthy volunteers. The results of these studies are inconsistent and difficult to compare, because each study employed different PET and CMR methods. Pärkkä et al. (12) studied 18 healthy volunteers with ¹⁵O-water PET and CMR. They used compartmental modeling and found that MPR and stress perfusion values were significantly correlated ($r = 0.48$ and $r = 0.7$, respectively). However, rest perfusion values were not significantly correlated. Fritz-Hansen et al. (11) studied 10 healthy volunteers with ¹³N-ammonia PET with the same CMR

quantification method. They demonstrated, similar to our findings, a significant correlation between global MPR_{PET} and MPR_{CMR} ($r = 0.7$). They found an even stronger correlation for the change in myocardial perfusion between rest and stress ($r = 0.96$). Both of these studies found a tendency for CMR to underestimate MPR compared with PET. In a smaller study of 5 volunteers, Pack et al. (13) found that 3-T CMR and ^{13}N -ammonia PET perfusion and MPR values are well correlated but that MPR might be in better agreement.

Both PET and CMR perfusion and MPR values in our study are physiologically plausible and in the same range as in some previous studies (11,21–25) but lower than those reported by others (26). Patients are known to have lower stress and rest perfusion than normal volunteers even in remote territories (22,23). The mean difference between PET and CMR perfusion values is small; however, the limits of agreement are broad, particularly at higher stress perfusion values. The findings that MPR, a ratio of stress and rest perfusion values, correlates well but that the absolute perfusion values correlate relatively poorly suggests that the errors in quantification had a similar influence on both rest and stress perfusion values and were subsequently cancelled by the calculation of MPR. These errors might be a result of either methodological or physiological factors.

Methodological considerations

Differences in voxel size and acquisition methods (3-dimensional segments were acquired with PET and 10-mm slices through the 16 segments with CMR) and the use of different postprocessing methods mean that, despite the calculation of mean values in standard segments and territories, there is likely to have been a small degree of misregistration between the modalities. Furthermore, although partial volume effects are corrected for with compartmental PET analysis (16), these were not taken into account with CMR.

Positron emission tomography and CMR have both been validated against gold-standard techniques; however, both still have limitations. The PET studies have used different tracers (e.g., ^{13}N -ammonia vs. ^{15}O water), acquisition protocols, and analysis methods (e.g., Patlak, compartmental). In the present study, the method might underestimate absolute perfusion (27)—an effect that would also be cancelled by the calculation of MPR.

Quantitative CMR perfusion studies have used different magnetic field strengths, contrast agents, perfusion sequences, and post-processing algorithms. One of the main challenges faced by CMR quantification is that, at higher concentrations of contrast agent, there is a loss of linearity between contrast agent concentration and signal intensity. It is possible that these signal saturation effects influenced our CMR results. However, we used a dual bolus method to preserve signal linearity in the left ventricular blood pool and allow calculation of the arterial input function. In addition, a previous study has demonstrated that myocardial signal intensity continues to increase at much higher doses of contrast agent (up to at least 0.15 mmol/kg) than those used here (28), although this study was in normal hearts imaged with a hybrid echo planar pulse sequence. Furthermore, a 14-min interval between stress and rest imaging is insufficient for complete dissipation of the contrast agent. We used a published method (29) of baseline correction to account for this; however, this might also have introduced some error into rest perfusion measurements. We did not find, in contrast to

the previous volunteer studies, any evidence of systematic errors for low or high values, indicating that CMR is valid for a wide range of perfusion values.

It is also possible that systematic errors could relate to the fact that PET (30,31) and particularly CMR (32) have only moderate inter-study reproducibility. Regional measurements of perfusion in particular tend to be less reproducible than global measurements. Because the reproducibility of data analysis was relatively high, this might also be partly explained by physiological variation of perfusion.

Physiological considerations

There are also likely to have been real changes in perfusion, even though the interval between the PET and CMR scans was short. Little is known, for example, as to whether diurnal variation or changes in hydration affect myocardial perfusion. It is also possible that such differences are more pronounced in patients than in volunteers—for example, as a result of the use of medications that affect perfusion.

Clinical implications

The novel finding that MPR_{CMR} correlates strongly with MPR_{PET} is important, given the proven utility of MPR_{PET} . The MPR_{PET} correlates inversely with the degree of coronary artery stenosis at angiography (33). Furthermore, recent studies have demonstrated that MPR_{PET} is an independent predictor of outcome and predicts major adverse cardiac events and cardiac death (3) in patients with suspected impaired perfusion and reduced survival in patients with left ventricular impairment (4).

In this study, we identified $MPR2_{CMR}$ and $MPR2_{PET}$ cutoff values that detected CAD with a high sensitivity and specificity. The MPR was superior to absolute stress perfusion values for this purpose. The $MPR2_{PET}$ AUC was significantly higher than absolute stress perfusion values AUC, and there was also a trend toward significance with $MPR2_{CMR}$. This was reflected by optimal cutoff values for absolute stress perfusion values that would be clinically less useful due to modest sensitivity (CMR) or specificity (PET). This is in contrast to some limited recent PET data suggesting that a single absolute stress perfusion cutoff value might be superior to MPR for the detection of CAD (21). In addition, there does not seem to be any incremental value in combining MPR data from both PET and CMR for the diagnosis of CAD.

Quantification of images in general results in more precise, reproducible, and user-independent results. Although in this study there was no incremental benefit over visual analysis, noninvasive quantification of perfusion might be particularly useful in cases where visual assessment is difficult, such as multivessel disease, severe left ventricular impairment, and after coronary artery bypass grafting. Moreover, it might eventually allow the definition of thresholds of perfusion associated with myocardial ischemia (34) and viability, both of which are known to be important for selecting patients for revascularization procedures. This study reinforces the evidence that quantitative CMR perfusion data can be useful for the assessment of patients with CAD, and at present, we can accurately differentiate normal

from abnormal myocardial territories. However, further method refinement is required before the benefits can be fully realized.

Study limitations

The sample size in this study is modest. However, this is the first study in patients and larger than previous volunteer studies. Furthermore, this limitation applies more to the secondary objective of exploring diagnostic accuracy than to the primary aim of comparing quantitative perfusion. Large sample sizes will be required to demonstrate any significant differences in diagnostic accuracy. We compared functional tests against an anatomic reference standard (QCA), despite the well-documented limitations of this approach. It might be better to use fractional flow reserve as a reference standard in future studies. However, x-ray coronary angiography is widely used as the reference standard in both studies and in clinical practice, and again, this does not affect the comparisons made between PET and CMR, which was the main goal of our study.

The patients included in this study had overall good left ventricular function, and it is not clear how well these methods translate into patients with left ventricular systolic dysfunction (35). Finally, the relatively high prevalence of CAD and the exclusion of segments in which image quality precluded quantitative assessment means that these results are likely to represent best possible results with currently available quantitative techniques.

Conclusions

There is a strong correlation between MPR derived from quantitative CMR and PET data, which is important given the proven value of MPR_{PET} for detection of CAD, prognostication, and assessment of the microcirculation. The MPR_{PET} and MPR_{CMR} seem to predict significant CAD equally well and accurately. However, in patients, the absolute perfusion values from PET and CMR are only weakly correlated, suggesting that further refinement of quantitative techniques is required.

Acknowledgment

The authors thank Siobhan Crighton of King's College London for her assistance with the statistical analysis.

This work was supported by a European Union Grant (Grant number 224495 to Drs. Morton and Nagel); the British Heart Foundation (Research Excellence Award RE/08/003 and FS/10/029/28253 to Drs. Schuster, Perera, and Nagel); the Biomedical Research Centre (Grant number BRC-CTF 196 to Drs. Schuster, Perera, and Nagel); the Wellcome Trust and EPSRC (Grant number WT 088641/Z/09/Z to Drs. Chiribiri and Nagel); and the Academy of Finland Centre of Excellence in Molecular Imaging in Cardiovascular and Metabolic Research, Helsinki, Finland (Dr. Knuuti.). Dr. Nagel has received grant support from Philips Healthcare and Bayer Schering Pharma. Dr. Hedstrom has received grant support from Covidien. Dr. Knuuti has received grant support from Gilead Inc.; and is a consultant to Lantheus Inc. All other authors have reported that they have no relationships relevant to the contents of this paper to disclose.

Abbreviations and Acronyms

AUC	area under the receiver-operating characteristic curve
CAD	coronary artery disease

CI	confidence interval
CMR	cardiovascular magnetic resonance
CT	computed tomography
CV	coefficient of variation
MPR	myocardial perfusion reserve
PCI	percutaneous coronary intervention
PET	positron emission tomography
QCA	quantitative coronary angiography
ROC	receiver-operating characteristic

References

1. Knuuti J, Kajander S, Mäki M, Ukkonen H. Quantification of myocardial blood flow will reform the detection of CAD. *J Nucl Cardiol.* 2009; 16:497–506. [PubMed: 19495903]
2. Kajander SA, Joutsiniemi E, Saraste M, et al. Clinical value of absolute quantification of myocardial perfusion with (15)O-water in coronary artery disease. *Circ Cardiovasc Imaging.* 2011; 4:678–84. [PubMed: 21926262]
3. Herzog BA, Husmann L, Valenta I, et al. Long-term prognostic value of 13N-ammonia myocardial perfusion positron emission tomography added value of coronary flow reserve. *J Am Coll Cardiol.* 2009; 54:150–6. [PubMed: 19573732]
4. Tio RA, Dabeshlim A, Siebelink H-MJ, et al. Comparison between the prognostic value of left ventricular function and myocardial perfusion reserve in patients with ischemic heart disease. *J Nucl Med.* 2009; 50:214–9. [PubMed: 19164219]
5. Camici PG, Crea F. Coronary microvascular dysfunction. *N Engl J Med.* 2007; 356:830–40. [PubMed: 17314342]
6. Nandalur KR, Dwamena BA, Choudhri AF, Nandalur MR, Carlos RC. Diagnostic performance of stress cardiac magnetic resonance imaging in the detection of coronary artery disease: a meta-analysis. *J Am Coll Cardiol.* 2007; 50:1343–53. [PubMed: 17903634]
7. Christian TF, Rettmann DW, Aletras AH, et al. Absolute myocardial perfusion in canines measured by using dual-bolus first-pass MR imaging. *Radiology.* 2004; 232:677–84. [PubMed: 15284436]
8. Patel AR, Antkowiak PF, Nandalur KR, et al. Assessment of advanced coronary artery disease: advantages of quantitative cardiac magnetic resonance perfusion analysis. *J Am Coll Cardiol.* 2010; 56:561–9. [PubMed: 20688211]
9. Ichihara T, Ishida M, Kitagawa K, et al. Quantitative analysis of first-pass contrast-enhanced myocardial perfusion MRI using a Patlak plot method and blood saturation correction. *Magn Reson Med.* 2009; 62:373–83. [PubMed: 19353669]
10. Jerosch-Herold M, Wilke N, Stillman AE. Magnetic resonance quantification of the myocardial perfusion reserve with a Fermi function model for constrained deconvolution. *Med Phys.* 1998; 25:73–84. [PubMed: 9472829]
11. Fritz-Hansen T, Hove JD, Kofoed KF, Kelbaek H, Larsson HB. Quantification of MRI measured myocardial perfusion reserve in healthy humans: a comparison with positron emission tomography. *J Magn Reson Imaging.* 2008; 27:818–24. [PubMed: 18383259]
12. Pärkkä JP, Niemi P, Saraste A, et al. Comparison of MRI and positron emission tomography for measuring myocardial perfusion reserve in healthy humans. *Magn Reson Med.* 2006; 55:772–9. [PubMed: 16508915]

13. Pack NA, DiBella EVR, Rust TC, et al. Estimating myocardial perfusion from dynamic contrast-enhanced CMR with a model-independent deconvolution method. *J Cardiovasc Magn Reson.* 2008; 10:52. [PubMed: 19014509]
14. Cerqueira MD, Weissman NJ, Dilsizian V, et al. Standardized myocardial segmentation and nomenclature for tomographic imaging of the heart: a statement for healthcare professionals from the Cardiac Imaging Committee of the Council on Clinical Cardiology of the American Heart Association. *Circulation.* 2002; 105:539–42. [PubMed: 11815441]
15. Ishida M, Schuster A, Morton G, et al. Development of a universal dual-bolus injection scheme for the quantitative assessment of myocardial perfusion cardiovascular magnetic resonance. *J Cardiovasc Magn Reson.* 2011; 13:28. [PubMed: 21609423]
16. Choi Y, Huang SC, Hawkins RA, et al. A simplified method for quantification of myocardial blood flow using nitrogen-13-ammonia and dynamic PET. *J Nucl Med.* 1993; 34:488–97. [PubMed: 8280197]
17. Lockie T, Ishida M, Perera D, et al. High-resolution magnetic resonance myocardial perfusion imaging at 3.0-Tesla to detect hemo-dynamically significant coronary stenoses as determined by fractional flow reserve. *J Am Coll Cardiol.* 2010; 57:70–5.
18. Schwitter J, Nanz D, Kneifel S, et al. Assessment of myocardial perfusion in coronary artery disease by magnetic resonance: a comparison with positron emission tomography and coronary angiography. *Circulation.* 2001; 103:2230–5. [PubMed: 11342469]
19. Ibrahim T, Nekolla SG, Schreiber K, et al. Assessment of coronary flow reserve: comparison between contrast-enhanced magnetic resonance imaging and positron emission tomography. *J Am Coll Cardiol.* 2002; 39:864–70. [PubMed: 11869854]
20. Nagel E, Klein C, Paetsch I, et al. Magnetic resonance perfusion measurements for the noninvasive detection of coronary artery disease. *Circulation.* 2003; 108:432–7. [PubMed: 12860910]
21. Hajjiri MM, Leavitt MB, Zheng H, Spooner AE, Fischman AJ, Gewirtz H. Comparison of positron emission tomography measurement of adenosine-stimulated absolute myocardial blood flow versus relative myocardial tracer content for physiological assessment of coronary artery stenosis severity and location. *J Am Coll Cardiol Img.* 2009; 2:751–8.
22. Iida H, Kanno I, Takahashi A, et al. Measurement of absolute myocardial blood flow with H215O and dynamic positron-emission tomography. Strategy for quantification in relation to the partial-volume effect. *Circulation.* 1988; 78:104–15. [PubMed: 3260151]
23. Karamitsos T, Leccisotti L, Arnold J, et al. Relationship between regional myocardial oxygenation and perfusion in patients with coronary artery disease: insights from cardiovascular magnetic resonance and positron emission tomography. *Circ Cardiovasc Imaging.* 2010; 3:32–40. [PubMed: 19920032]
24. Czernin J, Müller P, Chan S, et al. Influence of age and hemodynamics on myocardial blood flow and flow reserve. *Circulation.* 1993; 88:62–9. [PubMed: 8319357]
25. Selvanayagam J, Cheng A, Jerosch-Herold M, et al. Effect of distal embolization on myocardial perfusion reserve after percutaneous coronary intervention: a quantitative magnetic resonance perfusion study. *Circulation.* 2007; 116:1458–64. [PubMed: 17785626]
26. Nesterov SV, Han C, Mâki M, et al. Myocardial perfusion quantitation with 15O-labelled water PET: high reproducibility of the new cardiac analysis software (Carimas). *Eur J Nucl Med Mol Imaging.* 2009; 36:1594–602. [PubMed: 19408000]
27. Mullani NA. Is the Patlak graphical analysis method applicable to measurement of myocardial blood flow with nitrogen-13-ammonia? *J Nucl Med.* 1993; 34:1831–4. [PubMed: 8410314]
28. Giang TH, Nanz D, Coulden R, et al. Detection of coronary artery disease by magnetic resonance myocardial perfusion imaging with various contrast medium doses: first European multi-centre experience. *Eur Heart J.* 2004; 25:1657–65. [PubMed: 15351166]
29. Jerosch-Herold M, Seethamraju RT, Swingen CM, Wilke NM, Stillman AE. Analysis of myocardial perfusion MRI. *J Magn Reson Imaging.* 2004; 19:758–70. [PubMed: 15170782]
30. Kaufmann PA, Gneccchi-Ruscione T, Yap JT, Rimoldi O, Camici PG. Assessment of the reproducibility of baseline and hyperemic myocardial blood flow measurements with 15O-labeled water and PET. *J Nucl Med.* 1999; 40:1848–56. [PubMed: 10565780]

31. Nagamachi S, Czernin J, Kim AS, et al. Reproducibility of measurements of regional resting and hyperemic myocardial blood flow assessed with PET. *J Nucl Med.* 1996; 37:1626–31. [PubMed: 8862296]
32. Jerosch-Herold M, Vazquez G, Wang L, Jacobs DR, Folsom AR. Variability of myocardial blood flow measurements by magnetic resonance imaging in the Multi-Ethnic Study of Atherosclerosis. *Invest Radiol.* 2008; 43:155–61. [PubMed: 18301311]
33. Uren NG, Melin JA, De Bruyne B, Wijns W, Baudhuin T, Camici PG. Relation between myocardial blood flow and the severity of coronary-artery stenosis. *N Engl J Med.* 1994; 330:1782–8. [PubMed: 8190154]
34. Vogel R, Indermuhle A, Seiler C. Determination of the absolute perfusion threshold preventing myocardial ischaemia in humans. *Heart.* 2007; 93:115–6. [PubMed: 17170349]
35. Schuster A, Morton G, Chiribiri A, Perera D, Vanoverschelde J, Nagel E. Imaging in the management of ischemic cardiomyopathy: special focus on magnetic resonance. *J Am Coll Cardiol.* 2012; 59:359–70. [PubMed: 22261158]

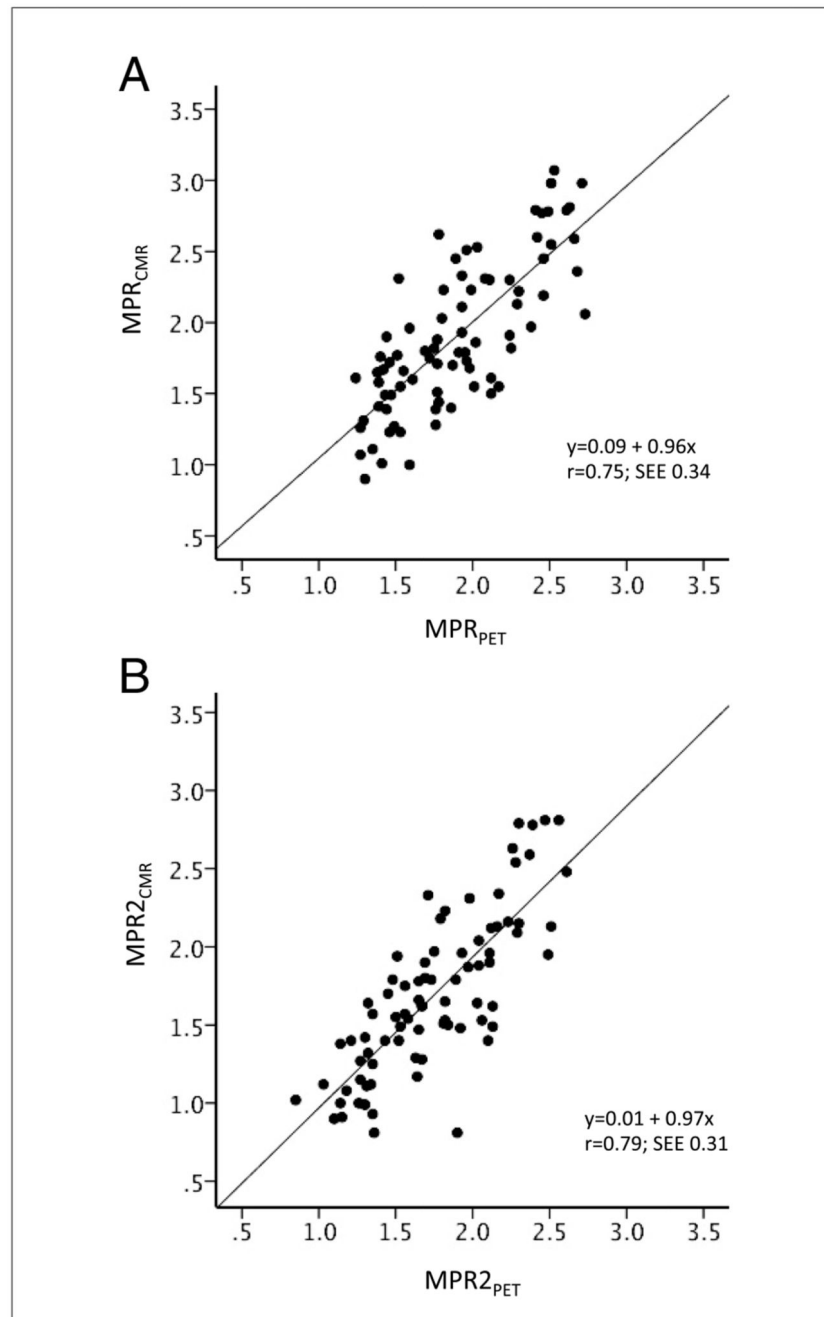


Figure 1. Scatter Plots Comparing CMR- and PET-Derived MPR

Scatter plots with fit lines comparing myocardial perfusion reserve (MPR) values from cardiac magnetic resonance (MPR_{CMR}) and positron emission tomography (MPR_{PET}) for the entire myocardial territory (**A**) and the mean of the lowest 2 segments in each territory (MPR2) (**B**).

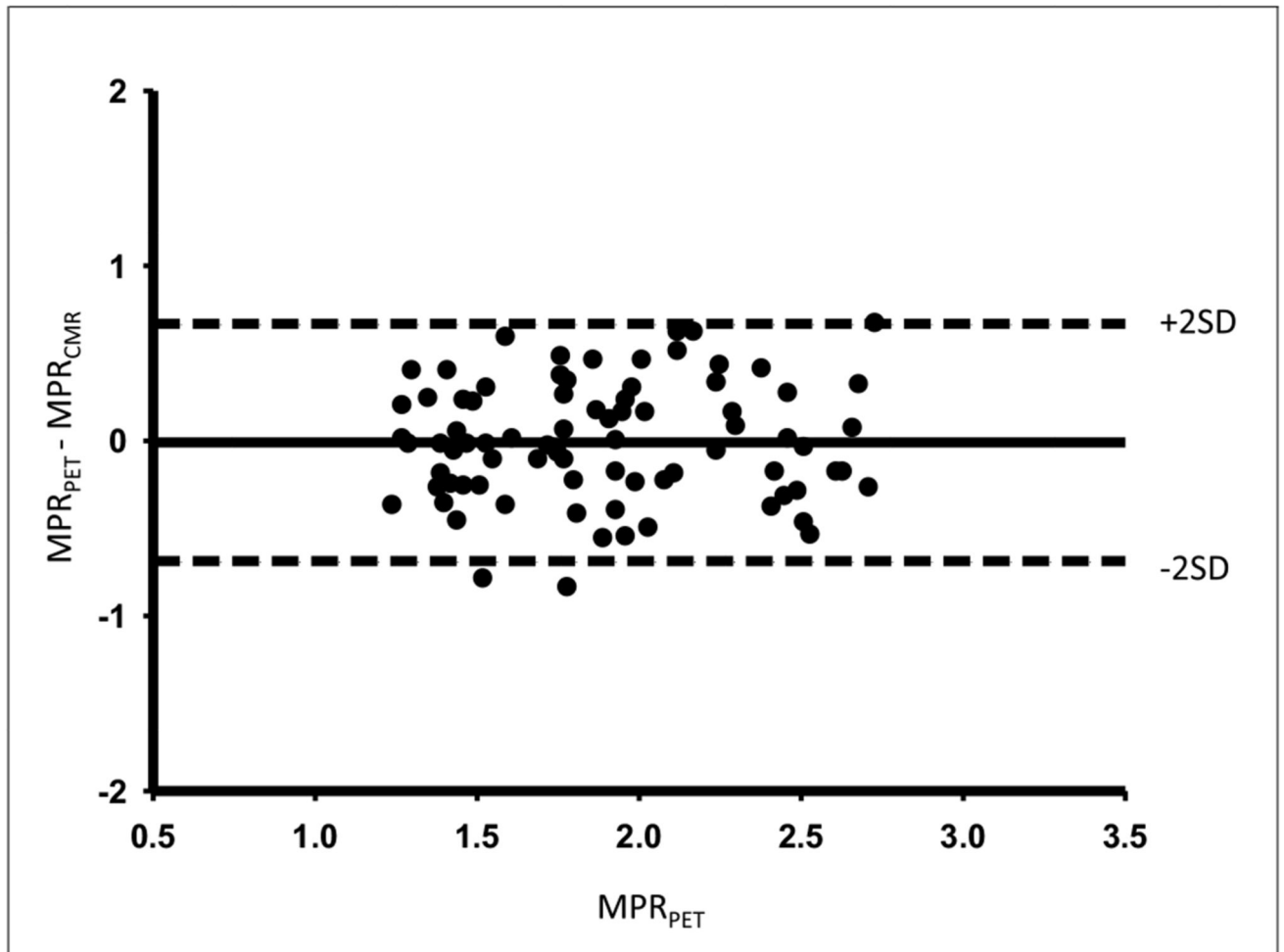


Figure 2. Agreement Between CMR and PET MPR

Bland-Altman plot showing the agreement between CMR- and PET-derived absolute MPR measurements. Abbreviations as in Figure 1.

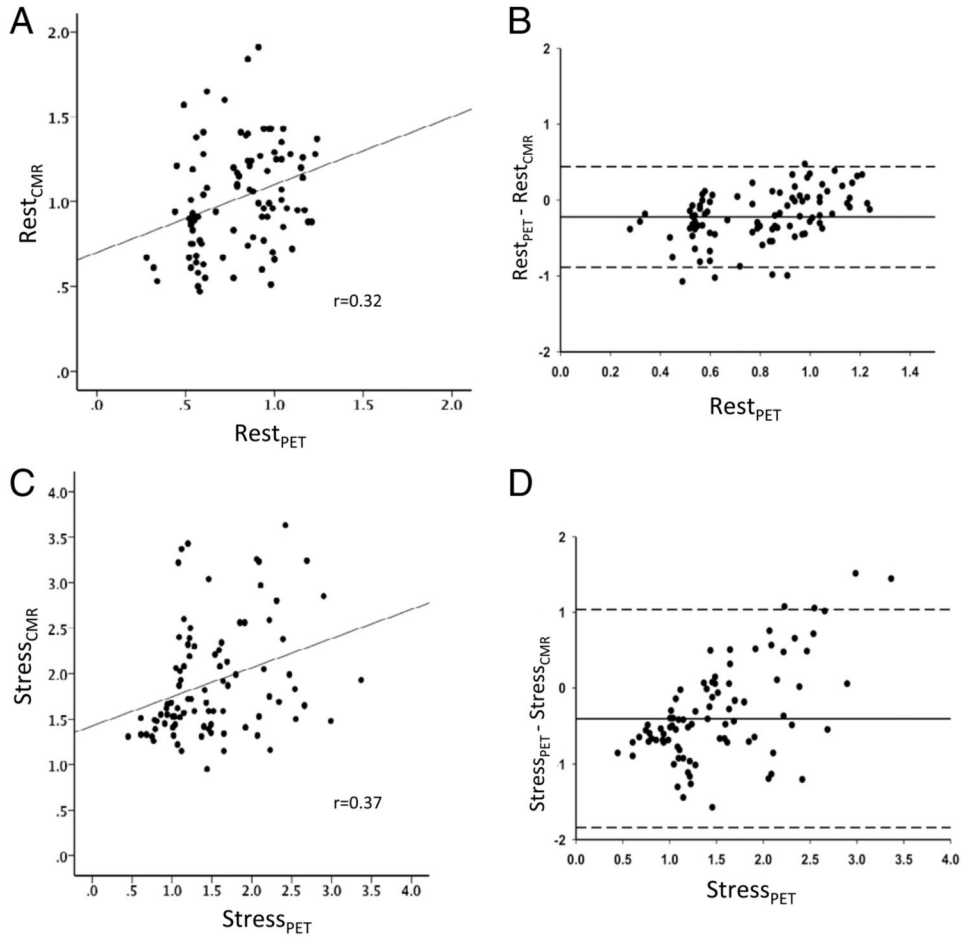


Figure 3. I Correlation and Agreement Between CMR- and PET-Derived Absolute Perfusion Values

Scatter plots illustrating the correlation between absolute measures of myocardial perfusion at rest (A) and during peak stress (C), along with the corresponding Bland-Altman plots (B and D, respectively) with limits of agreement lines (2 SDs). CMR = cardiac magnetic resonance; PET = positron emission tomography.

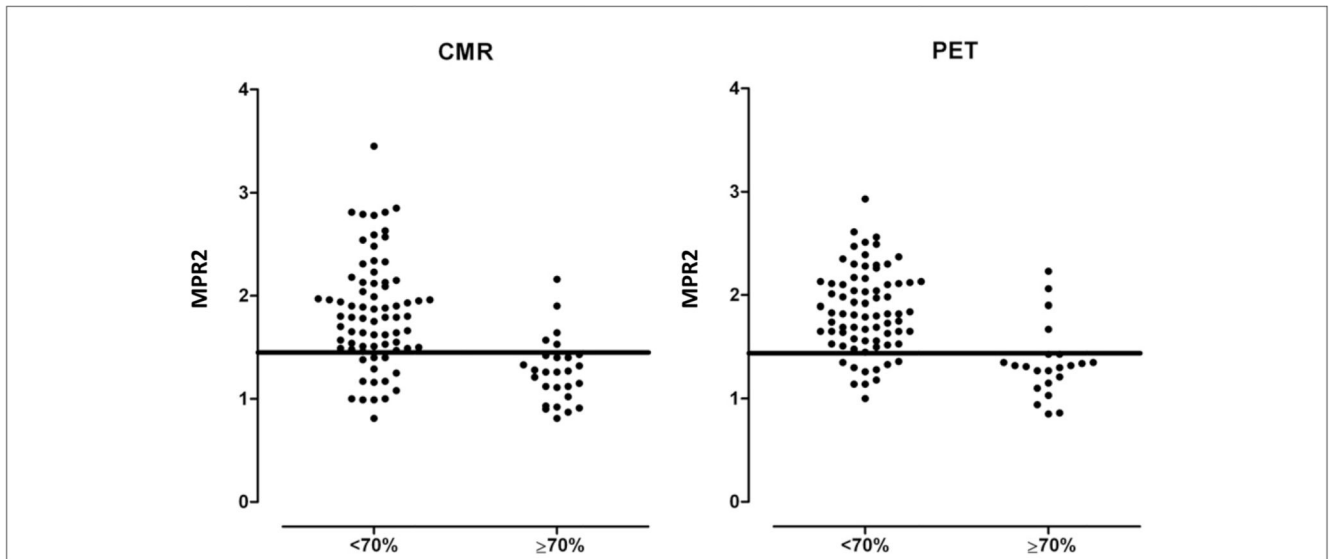


Figure 4. Diagnosis of Significant CAD on Angiography With Quantitative CMR- and PET-Derived MPR

Mean myocardial perfusion reserve (MPR) of the lowest 2 segments (MPR2) in remote (<70%) and stenotic (≥70%) territories. The best cutoff values for the detection of coronary artery disease (CAD) (MPR_{CMR} 1.45 and MPR_{PET} 1.44) are shown. CMR = cardiac magnetic resonance; PET = positron emission tomography.

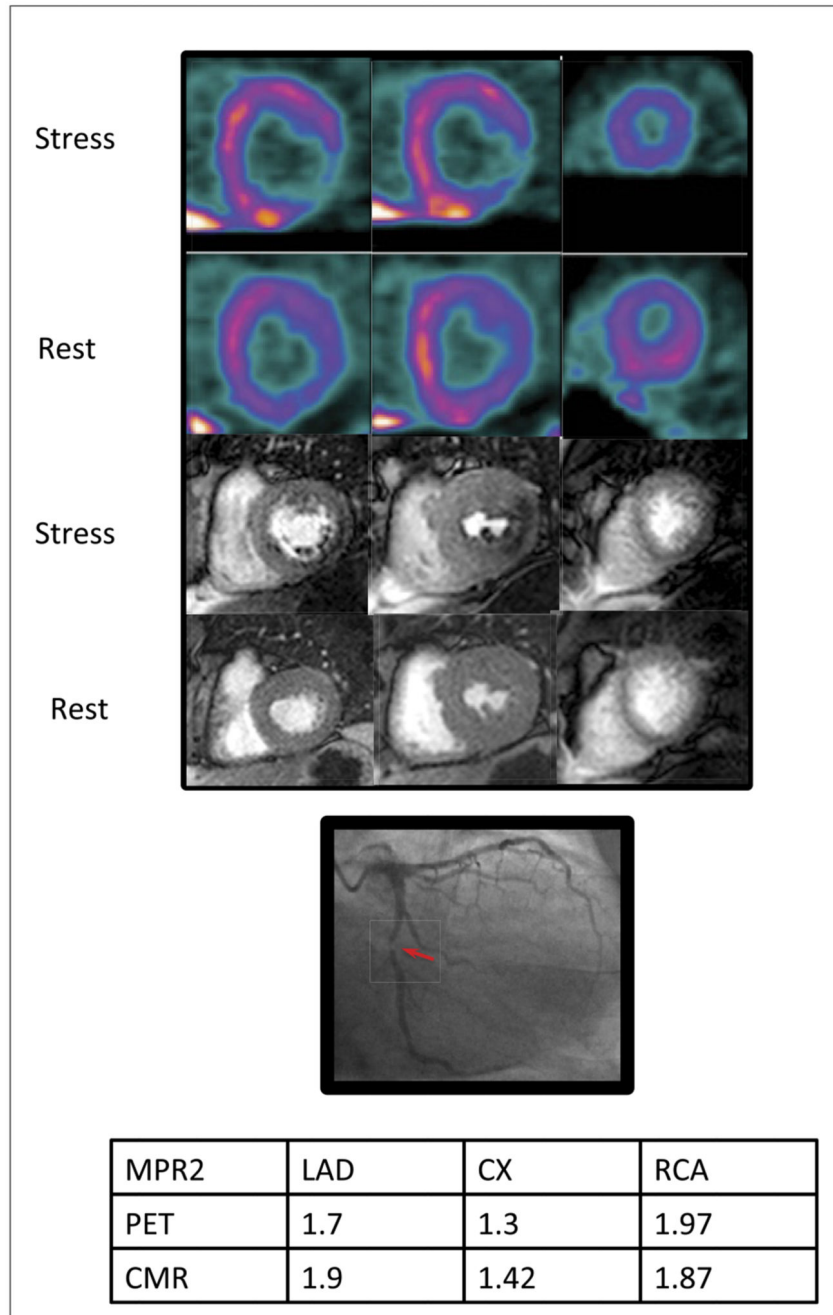


Figure 5. Case Example

Positron emission tomography (PET) (**top**), cardiac magnetic resonance (CMR) (**middle**), and the x-ray angiogram of the left coronary artery of a 54-year-old patient with diabetes and exertional angina. Basal, mid, and apical slices have been taken from the PET study, which approximately correspond to the CMR slices. There is a stress-induced perfusion defect in the infero-lateral region from base to apex visible on both PET and CMR images. There is a corresponding severe (>95%) stenosis of the proximal circumflex artery. There was no other significant angiographic disease. Myocardial perfusion reserve of the lowest 2

segments (MPR2) for each territory are shown in the table. The MPR2 for the circumflex artery (CX) is below the cutoff of 1.44 and 1.45 for both PET and CMR, respectively. LAD = left anterior descending artery; RCA = right coronary artery.

Table 1
Baseline Characteristics of Study Participants

Age (yrs)	63 ± 9
Male	32 (78%)
BMI (kg/m²)	29 ± 5
LVEF (%)	63 ± 12
Angina (CCS)	
Class 1	11 (27%)
Class 2	22 (54%)
Class 3	8 (20%)
Diabetes	13 (32%)
Hypertension	30 (73%)
Smoking	
Current	5 (12%)
Previous	21 (51%)
Family history CAD	19 (46%)
Cerebrovascular disease	4 (10%)
Previous PCI	13 (32%)
Previous myocardial infarct	5 (12%)
Medications	
Aspirin	33 (80%)
Clopidogrel	18 (44%)
Statin	35 (85%)
ACE inhibitor	22 (54%)
Angiotensin receptor blocker	6 (15%)
Beta-blocker	21 (51%)
Calcium channel blocker	13 (32%)
Nitrate	7 (17%)
Coronary disease 70%	
Left anterior descending	7 (17%)
Circumflex	12 (29%)
Right coronary artery	13 (32%)

Values are mean ± SD or n (%). Percentages have been rounded to nearest 1% and therefore might not total 100%. ACE = angiotensin-converting enzyme; BMI = body mass index; CAD = coronary artery disease; CCS = Canadian Cardiovascular Society; LVEF = left ventricular ejection fraction; PCI = percutaneous coronary intervention.

Table 2
Summary of Hemodynamic Data for All Participants During Imaging Studies

	Rest			Stress		
	PET	CMR	p Value	PET	CMR	p Value
HR	63 ± 10	66 ± 11	0.002	84 ± 15	86 ± 15	0.14
SBP	138 ± 18	137 ± 17	0.75	128 ± 21	132 ± 19	0.164
RPP	8,734 ± 336	9,122 ± 329	0.12	10,781 ± 479	11,344 ± 445	0.08

Values are mean ± SD.

CMR = cardiovascular magnetic resonance; HR = heart rate; PET = positron emission tomography; RPP = rate pressure product; SBP = systolic blood pressure.

Table 3
Quantitative Perfusion Values in Patients With and Without Significant CAD

	CAD			No CAD	
	Stenotic Territory	Remote Territory	p Value*	All Territories	p Value [†]
Stress perfusion (ml/min/g)					
PET	1.24 ± 0.49	1.56 ± 0.66	<0.0001	1.72 ± 0.66	0.49
CMR	1.54 ± 0.34	1.94 ± 0.59	0.001	2.03 ± 0.63	0.68
Rest perfusion (ml/min/g)					
PET	0.77 ± 0.24	0.81 ± 0.25	0.08	0.85 ± 0.26	0.71
CMR	1.03 ± 0.30	1.06 ± 0.33	0.28	0.98 ± 0.29	0.47
MPR					
PET	1.57 ± 0.31	1.87 ± 0.36	<0.0001	2.06 ± 0.44	0.20
CMR	1.55 ± 0.36	1.90 ± 0.48	0.001	2.20 ± 0.56	0.13
Mean MPR of 2 lowest segments					
PET	1.36 ± 0.32	1.74 ± 0.32	<0.0001	1.92 ± 0.39	0.14
CMR	1.31 ± 0.30	1.70 ± 0.42	<0.0001	1.93 ± 0.53	0.16

Values are mean ± SD. A stenotic territory is subtended by a coronary artery with ≥70% diameter stenosis, and a remote territory is subtended by coronary arteries with <70% stenosis.

[†]Significance of the difference between stenotic and remote territory values.

*Significance of the difference between the values in patients without coronary artery disease (CAD) and remote territories in patients with CAD.

CMR = cardiovascular magnetic resonance; MPR = myocardial perfusion reserve; PET = positron emission tomography.