# Europe PMC Funders Group Author Manuscript Expert Opin Drug Metab Toxicol. Author manuscript; available in PMC 2023 February 15.

Published in final edited form as: *Expert Opin Drug Metab Toxicol.* 2020 December 01; 16(12): 1187–1198. doi:10.1080/17425255.2020.1828344.

# An update on the use of inositols in preventing Gestational Diabetes Mellitus (GDM) and Neural Tube Defects (NTDs)

"The Experts Group on Inositol in Basic and Clinical Research"

Fabio Facchinetti<sup>1</sup>, Vittorio Unfer<sup>2</sup>, Marialuisa Appetecchia<sup>3</sup>, Cesare Aragona<sup>4</sup>, Matteo Bertelli<sup>5</sup>, Arturo Bevilacqua<sup>6</sup>, Maria Salome Bezerra Espinola<sup>7</sup>, Mariano Bizzarri<sup>8</sup>, Pietro Cavalli<sup>9</sup>, Andrew Copp<sup>10</sup>, Rosario D'Anna<sup>11</sup>, Didier Dewailly<sup>12</sup>, Nicholas Greene<sup>13</sup>, Imelda Hernández Marín<sup>14</sup>, Zdravko A. Kamenov<sup>15</sup>, Eleni Kandaraki<sup>16</sup>, Evanthia Diamanti-Kandarakis<sup>17</sup>, Antonio Simone Laganà<sup>18</sup>, Giovanni Monastra<sup>19</sup>, Mario Montanino Oliva<sup>20</sup>, John E. Nestler<sup>21</sup>, Ali Cenk Ozay<sup>22</sup>, Olga Papalou<sup>23</sup>, Lali Pkhaladze<sup>24</sup>, Giusy Porcaro<sup>25</sup>, Nikos Prapas<sup>26</sup>, Christophe O. Soulage<sup>27</sup>, Annarita Stringaro<sup>28</sup>, Mónica Vazquez-Levin<sup>29</sup>, Artur Wdowiak<sup>30</sup>

<sup>1</sup>Unit of Obstetrics and Gynecology, Mother-Infant Department, University of Modena and Reggio Emilia, Italy

<sup>2</sup>Department of Experimental Medicine, Sapienza University of Rome, Rome, Italy

<sup>3</sup>Oncological Endocrinology Unit - Regina Elena National Cancer Institute -IRCCS, Rome, Italy

<sup>4</sup>Systems Biology Group Lab - Department of Experimental Medicine, Sapienza University of Rome, Rome, Italy

<sup>5</sup>EBTNA-Lab, Via Delle Maioliche, 57/G, 38068, Rovereto, TN, Italy; MAGI's Lab, Via Delle Maioliche, 57/D, 38068, Rovereto, TN, Italy; MAGI

<sup>6</sup>Department of Dynamic and Clinical Psychology, Sapienza University of Rome, Rome, Italy

<sup>7</sup>Systems Biology Group Lab - Department of Experimental Medicine, Sapienza University of Rome, Rome, Italy

<sup>8</sup>Department of Experimental Medicine, Systems Biology Group, University La Sapienza, Rome, Italy

<sup>9</sup>Clinical Genetics, ASST - Bergamo Est, Italy

<sup>10</sup>Newlife Birth Defects Research Centre and Developmental Biology & Cancer Programme, GOS Institute of Child Health, University College London, United Kingdom

<sup>11</sup>Department of Human Pathology, University of Messina, Messina, Italy

**Declaration of interest** 

Vittorio Unfer is employee at Lo. Li. Pharma s.r.l., Rome, Italy. Fabio Facchinetti has been a consultant of the same company. The other authors declare they do not have any potential conflict of interest for the content of this article.

<sup>13</sup>Newlife Birth Defects Research Centre and Developmental Biology & Cancer Programme, Institute of Child Health, University College London, London, United Kingdom

<sup>14</sup>Human Reproduction Department, Hospital Juárez de México, México City - Universidad Nacional Autónoma de México (UNAM), México City, México

<sup>15</sup>Department of Internal Medicine, Medical University of Sofia, Sofia - Bulgaria

<sup>16</sup>Department of Endocrinology & Diabetes, HYGEIA Hospital, Marousi, Athens, Greece

<sup>17</sup>Department of Endocrinology & Diabetes, HYGEIA Hospital, Marousi, Athens, Greece

<sup>18</sup>Department of Obstetrics and Gynecology, "Filippo Del Ponte" Hospital, University of Insubria, Varese, Italy

<sup>19</sup>Systems Biology Group Lab - Department of Experimental Medicine, Sapienza University of Rome, Rome, Italy

<sup>20</sup>Department of Obstetrics and Gynecology Santo Spirito Hospital Rome, Italy

<sup>21</sup>Division of Endocrinology, Diabetes and Metabolism, Department of Internal Medicine, Virginia Commonwealth University, Richmond, VA, USA

<sup>22</sup>Near East University, Faculty of Medicine, Department of Obstetrics and Gynecology, Nicosia Cyprus - Near East University, Research Center of Experimental Health Sciences, Nicosia, Cyprus

<sup>23</sup>Department of Endocrinology & Diabetes, HYGEIA Hospital, Marousi, Athens, Greece

<sup>24</sup>loseb Zhordania Institute of Reproductology, Tbilisi, Georgia

<sup>25</sup>Women's Health Center, USL Umbria 2, Orvieto, Italy

<sup>26</sup>3rd department of OB-GYNAE, Aristotle University of Thessaloniki, Greece - IAKENTRO Fertility Centre, IVF laboratory, Thessaloniki, Greece

<sup>27</sup>University of Lyon, INSERM U1060, CarMeN, INSA de Lyon, Université Claude Bernard Lyon 1, Villeurbanne, France

<sup>28</sup>National Center for Drug Research and Evaluation -Italian National Institute of Health, Rome, Italy

<sup>29</sup>National Council of Scientific and Technical Research - Instituto de Biología y Medicina Experimental (IBYME) - Buenos Aires, Argentina

<sup>30</sup>Diagnostic Techniques Unit, Medical University of Lublin, Poland

## Abstract

**Introduction**—Obstetric history and maternal body composition and lifestyle may be associated with serious complications both for the mother, such as gestational diabetes mellitus (GDM), and for the fetus, including congenital malformations such as neural tube defects (NTDs).

et al.

**Areas covered**—In view of the recent knowledge, changes of nutritional and physical activity habits ameliorate glycemic control during pregnancy and in turn improve maternal and neonatal health outcomes. Recently, a series of small clinical and experimental studies indicated that supplementation with inositols, a family of insulin sensitizers, was associated with beneficial impact for both GDM and NTDs.

**Expert opinion**—Herein, we discuss the most significant scientific evidence supporting myoinositol administration as a prophylaxis for the above-mentioned conditions.

#### Keywords

D-*chiro*-inositol (DCI); gestational diabetes mellitus (GDM); insulin resistance; myo-inositol (MI); neural tube defects (NTDs); prevention

# 1 Introduction

Several factors can be decisive in the onset of unfavorable maternal and fetal outcomes, lifestyle among them [1]. In particular, poor maternal nutritional habits, and especially a lack of glycemic control, may represent leading causes of complications including gestational diabetes mellitus (GDM), large for gestation age babies (LGA) [2, 3] and congenital abnormalities affecting the fetal central nervous system, namely neural tube defects (NTDs) [4].

Recent scientific evidence [5, 6] has shown that the supplementation of inositols is correlated with achievement of a healthy pregnancy. In particular, myo-inositol (MI) may prevent the onset of GDM as well as reduce the risk of recurrence of NTDs. Here, the authors have gathered data from their own and others' studies to provide an overview of up-to-date knowledge on the mechanisms by which MI may prevent GDM and NTDs, and the implications of these findings for future clinical practice.

## 2 The inositols family

## 2.1 Biochemical properties

Inositols are sugar alcohols (polyols) that occur in 9 different stereoisomeric forms. MI, the most common naturally occurring one, can be transformed into numerous derivatives mainly through either epimerization or phosphorylation of one or several of its hydroxyl groups. The conversion to D-*chiro*-, scyllo-, muco-, neo-occurs by means of specific epimerases. In particular, the conversion rate of MI to D-*chiro*-inositol (DCI) ranges from 7% to about 9%, as measured by the analysis of radiolabeled [3H]-MI, whereas the production of other isomers is minimal, not exceeding 0.06% of total radiolabeled MI [7]. The human body can actively synthetize MI: in particular, the kidneys produce up to 4 g/day [8]. MI derives from the isomerization of glucose-6-phosphate (G6P) to inositol-3-phosphate (Ins3P) by the enzyme D-3-myo-inositol-phosphate synthase (inositol synthase, Ino1 or MIPS1) [9]. Then, inositol-3-phosphate is dephosphorylated to free MI by means of inositol-monophosphatase-1 (IMPA-1 or IMPase) [10]. Free MI can also be generated by the recycling of inositol-1,4,5-trisphosphate (InsP3) and inositol-1,4-bisphosphate (InsP2) generated in inositol phosphate signaling [11]. Endogenous production of MI and DCI varies

depending on tissue-specific needs [12], and it is known that certain organs, such as brain, need high MI concentrations (10- to 15-fold higher values than are detected in blood) [9].

Inositol transporters are responsible for uptake and intracellular distribution of inositol. They were classified in two groups according to their transport mechanism. Sodium/myo-inositol cotransporter 1 and 2 (SMIT1 and SMIT2) are coupled with sodium ions while proton/ myo-inositol cotransporter (HMIT), coupled with protons [13]. They have a different tissue distribution. So far, only SMIT2 was detected in the duodenum and jejunum, the two districts of intestine where they were found [14].

## 2.2 Physiological activities

Inside the cells, inositols are not only present as free molecules, but also as components of membrane phosphoinositides (inositol-containing lipids), including phosphatidyl-inositol phosphate and phosphatidyl-inositol bisphosphate (PIP2), both compounds with important physiological roles [15]. Hydrolysis of PIP2 by phospholipase C (PLC) produces inositoltrisphosphate (Ins-1,4,5P3, InsP3), which regulates activities of hormones such as FSH, TSH and insulin as a second messenger [15]. The interaction between InsP3 and the membrane receptors of mitochondria and endoplasmic reticulum stimulates calcium influx into the cytosol. This leads the activation of protein kinase C (PKC) and mediates specific cellular responses.

Inositols are involved in insulin signaling, with both MI and DCI functioning as insulin second messengers, although they mediate different actions of insulin in humans [16, 17, 18, 19]. There is an interplay between MI uptake and cellular glucose uptake, and MI content is elevated in tissues such as brain, heart and ovary that have high glucose utilization and consumption [18, 20]. MI also inhibits adenylate cyclase, thus reducing the release of free fatty acids from adipose tissues [21]. Conversely, DCI levels are elevated in tissues specialized in glycogen storage, including liver, muscle and fat, whereas DCI has low abundance in tissues with high glucose utilization [7].

DCI and MI glycans (IPG-P and IPG-A, respectively) shift glucose metabolism towards glycogen synthesis or glucose catabolism, respectively. In the latter case, MI enhances phosphate-dehydrogenase activity (PDH), thus stimulating pyruvate catabolism. IPG-P seems preferentially produced in metabolic stress, following an increase of insulin release. Indeed, after insulin stimulation, MI is transformed into DCI, through an NAD-NADH-dependent epimerase, to maintain a suitable MI:DCI ratio, as required for tissue metabolism [19, 22, 23].

MI significantly inhibits glucose duodenal absorption and therefore counteracts a rise in blood glucose. This finding can be explained by a mutual interference of intestinal uptake by MI and glucose [19, 24]. In addition to the above-cited effects, MI also acts as one of the FSH second messengers in the ovary, as well as of the TSH in the thyroid, both mechanisms mediated by adenylate cyclase [19, 25].

#### 2.3 Pharmacokinetics

Some patients, identified as "inositol-resistant", exhibit a weaker than normal response to MI treatment, owing to poor absorption by the oral route.

Aiming to improve MI clinical efficacy, Monastra et al planned a pharmacokinetics study [26] in human healthy volunteers to test the effects of  $\alpha$ -LA on MI bioavailability, considering the wide range of  $\alpha$ -LA activities, especially the effects on improved mineral bioavailability [27].

When administered alone, MI average peak plasma concentration at 180 min increased about threefold *versus* the baseline, whereas when associated to α-LA, it augmented fourfold.

After administration of MI and  $\alpha$ -LA, MI plasma concentrations were significantly higher than after administration of MI alone: the increase of Cmax (µmol/l) was 32.4%, while AUC (0-300) increased by 27.5% [26].

Moreover, with the aim to explain the mechanism underlying this effect, the authors carried out an *in vitro* experiment. In detail, they observed that in the presence of digested  $\alpha$ -LA, MI passage across a monolayer of human Caco-2 cells, used to simulate the intestinal barrier, significantly increased, thereby achieving a higher plasma concentration compared with MI administration alone [26].

This combination of MI with  $\alpha$ -LA has proven to be useful for improving the treatment of Polycystic Ovary Syndrome (PCOS) MI-resistant patients [28]. A goal for the future will be to investigate further this combination of MI and  $\alpha$ -LA, for its ability to enhance the potential beneficial effects of MI on complex pathologies, such as GDM and NTDs (described below).

# 3 The safety profile of inositols

Before discussing the efficacy of inositols in some pathological conditions, it is important to consider the available data on inositol's safety as a supplement. As stated by the United States Food and Drug Administration (FDA), MI is included in the list of compounds that are 'generally recognized as safe' (GRAS). This means inositol has proven to be free of side-effects and, importantly, is safe for use in pregnancy [19, 29].

A recent meta-analysis on 965 pregnant women affected by GDM, who were randomized to receive MI, placebo or no treatment, revealed no adverse maternal events, and no congenital malformations in the fetuses or newborns [30]. Moreover, a Cochrane Review of the relationship between inositol and GDM reported no adverse events associated with inositol antenatal supplementation [31].

## 4 MI in management of GDM

## 4.1 GDM diagnosis

GDM, defined as glucose intolerance solely diagnosed during pregnancy, is characterized by increased insulin resistance and hyperglycemia. It represents a worldwide public health problem with a variable prevalence ranging from 2 to 30%, depending on the diagnostic criteria and the population studied [32]. Several clinical risks have been associated with GDM, both for the mother, such as hypertension, cesarean section and possibility of developing type 2 diabetes, as well as for the newborn, such as preterm birth, macrosomia, shoulder dystocia, neonatal hypoglycemia, respiratory distress syndrome and congenital abnormalities [33]. Owing to the strong correlation between GDM and maternal/perinatal/ neonatal complications, an early diagnosis is highly recommended.

In a recent review and meta-analysis, it was shown that risk for perinatal mortality, and neonatal hypoglycemia was greater among early-onset GDM women compared to late-onset GDM women, despite treatment [34].

As a result, screening for GDM should be done as soon as pregnancy is confirmed, especially in high-risk patients, by using fasting plasma glucose cut-off values [35].

The oral glucose tolerance test (OGTT) remains the gold standard for a GDM diagnosis, even though not devoid of limitations, principally associated with patient compliance [36]. One approach, adopted in some countries is a two-step vs one-step testing procedure, although it appears to have similar diagnostic efficiency [37]. Depending on resources, a universal screening of pregnant women has been implemented in some locations while others choose selective screening based on risk factors (age, obesity, obstetric history and ethnicity) [38]. Some serum proteins like insulin, adiponectin, C-reactive protein, sex hormone globulin and glycosylated fibronectin are widely investigated as markers of this pathological condition [39, 40, 41]. Although these results are promising, such predictive biomarkers are yet to achieve clinical applicability.

Advances in molecular biology have recently identified novel biomarkers, which offer the potential to improve GDM risk prediction and, consequently, enable application of intervention protocols. These biomarkers can be readily measured in biological fluids, such as blood, plasma and serum, allowing accurate low-cost diagnosis with improved patient compliance [42]. For example, it was observed that genetic variants, such as single nucleotide polymorphisms (SNPs) in genes involved in specific metabolic pathways may predispose pregnant women to develop GDM [43]. This potentially would allow analysis of a set of SNPs that provide indicators for GDM screening [44].

DNA methylation could also represent an interesting potential parameter correlating with a range of pathophysiological processes, including GDM [45]. Indeed, it was demonstrated that DNA methylation is highly altered in the placenta and cord blood of women with GDM [46]. Micro-RNAs have also proven to be important metabolic regulators during pregnancy, playing a role in GDM onset [47], which would make them ideal candidates for GDM detection. However, establishment of the specificity and sensitivity of potential new

et al.

biomarkers for GDM diagnosis is required – including external validation and population studies – before they can be used in clinical practice.

#### 4.2 Why GDM should be treated

The Hyperglycemia and Adverse Pregnancy Outcome (HAPO) Study evaluated the risk of adverse outcome associated with degrees of glucose intolerance during pregnancy and confirmed the link between maternal glucose and neonatal adiposity and suggested that this relationship is mediated by fetal insulin production influencing fetal growth [48].

Moreover, if not diagnosed and/or untreated, GDM can lead to short-term risks including labor induction, shoulder dystocia, C-section, macrosomia, large for gestational age (LGA) babies, neonatal hypoglycemia, NICU admission [49]. In a lifelong perspective, approximately 40% of all women who are diagnosed with GDM progress to type 2 diabetes within 5 years post-delivery, in addition to their increased risk of GDM in future pregnancies [50, 51].

Noteworthy, infants born to mothers with gestational diabetes are at increased risk of impaired glucose regulation, obesity and diabetes, leading to a vicious cycle of accumulated risks in the next generation [52, 53]. A recent follow-up study reported the longitudinal effects of GDM in the offspring. It included 4.160 children aged 10-14 years, whose mothers had a 75-g OGTT at ~28 weeks of gestation with blinded glucose values and underwent an OGTT. Offspring exposed to untreated GDM in utero are insulin-resistant with limited  $\beta$ -cell compensation compared with offspring born to mothers without GDM. GDM is significantly and independently associated with childhood intolerance to glucose [54].

#### 4.3 Lifestyle changes

With regards to management, before any therapeutic intervention, the mainstay for GDM approach is based on lifestyle changes with adequate diet and exercise, aiming to achieve glycemic targets. A recent individual patient data meta-analysis reported that such an approach was efficacious in containing gestational weight gain while the impact on GDM diagnoses was reported only for selected populations. No other perinatal outcomes were reported as significantly affected by lifestyle changes [55]. This was mainly due to the heterogeneity of approach in the primary trials which included different diets and physical activity programs. Moreover, adherence to lifestyle changes is generally poor and it is inversely correlated with efficacy, namely on GDM onset [56]. Anyway, if glycemic targets are not met, then insulin treatment should be considered without delay, which may also improve the woman's health-related quality of life [57]. Indeed, insulin treatment is associated with less risk of neonatal hypoglycemia and provides a good plasma glucose control [58].

## 4.4 Insulin sensitizers

Earlier observational studies [59, 60, 61] suggested that metformin treatment was associated with a lower risk of GDM, although the designs of these studies appear prone to different sorts of bias. Indeed, the above findings were not confirmed in a trial of 274 pregnancies among 257 women with PCOS who were randomly assigned to receive metformin (2000

mg/day) or placebo from the first trimester until delivery. The patients treated with metformin (17.6 %) and placebo (16.9%) presented a very similar prevalence of GDM. No significant differences were observed in the prevalence of preeclampsia or preterm delivery [62].

Two further trials investigated metformin for the prevention of large for gestational age babies. They included a total of >800 pregnant obese mothers (BMI >30) who received 1500 mg (till 2500 mg) oral metformin daily (or placebo), starting early in the second trimester. No efficacy of intervention was reported on birthweight and the rate of GDM was similar in metformin- and placebo-treated groups [63, 64]. Interestingly, a reduction of preeclampsia was reported with metformin (3.0%) in respect with placebo (11.3%) [63].

On the other hand, several clinical trials demonstrated the significant efficacy of inositols, especially MI, in GDM management (Table 1). Indeed, the concomitant administration of MI and DCI seems to perform less well than MI alone. When the two stereoisomers were directly compared, the non-inferiority analysis demonstrated the largest benefit for women treated with MI alone compared to those who received MI plus DCI or DCI alone [65]. This effect can also be explained referring to the competition between MI and DCI at the transporter site in the intestine. Indeed, SMIT2 in human cells exerts a similar affinity for both compounds with a Km value between 100 and 160  $\mu$ M, depending on the cell model used to determine it [13]. An *in vivo* pharmacokinetic study demonstrated the DCI inhibitory effect on MI bioavailability [66]. Obviously, a significant increase of DCI concentration in the gut, due to exogenous administration, hinders MI absorption with consequences on its bioavailability and, implicitly, its efficacy.

A recent systematic review and meta-analysis supported the prophylactic treatment with MI in the prevention of GDM. Among the included studies, all but one administered MI alone (2 g MI plus 200 µg folic acid, twice daily), while another one used a combination of lower MI dose (1100 mg) plus 27.6 mg DCI/day. The findings demonstrate that the administration of 2 g MI twice daily was associated with improved glycemic homeostasis, reduced GDM and preterm delivery rate, in respect to control groups, all supplemented with folic acid, except one trial [30].

Moreover, a secondary analysis from three trials reported that the same dose of MI supplementation in women at risk for GDM affected birthweight, reducing the rate of both macrosomic newborns and LGA babies [67]. Such neonatal impact agrees with the evidence that dietary glycemic index directly correlates with LGA in newborns [68] and indirectly demonstrate that MI supplementation improves glycemic control in those pregnancies.

Finally, in a condition where insulin-resistance is pathogenetic like PCOS, the beneficial effects of MI supplementation on hormonal and reproductive disturbances [20] were specifically reported also in pregnancy, where GDM rate was reduced [69].

From the experimental point of view, recent animal studies showed that combined inositols treatment (MI/DCI 40:1) in pregnant mouse complicated by metabolic syndrome and obesity improved blood pressure, glucose levels at the glucose tolerance, as well as leptin levels [70]. In addition, the same combination of inositols treatment improved not only maternal

outcomes, but also offspring weight at birth as well as glucose tolerance test, and vascular reactivity in their adult life, thus reducing the vicious cycle of dysmetabolism [71].

Finally, it should be highlighted that inositols administration in pregnancy was well tolerated and found to be safe, unlike metformin which has been associated with several gastrointestinal symptoms [63].

# 5 Prevention of NTDs: the role of MI

## 5.1 Background to NTDs

Neural tube defects (NTDs) are a group of congenital malformations affecting the brain and spinal cord that originate at various times during gestation. Some authors use the term 'NTDs' to denote defects that specifically result from faulty neural tube closure, particularly myelomeningocele (i.e. open spina bifida) and anencephaly. However, NTDs are sometimes considered to also include defects that arise by other embryonic and fetal mechanisms, including encephalocele (a brain herniation defect) and skin-covered low spinal 'dysraphic' conditions (secondary neurulation defects) [81]. Hydrocephalus and Chiari II malformation are closely associated with myelomeningocele and so are frequently present in individuals with NTDs.

Many different factors affect susceptibility to NTDs, including genetic variants and environmental influences, including the antiepileptic drugs valproic acid and carbamazepine [81]. Nutritional status is important, with folate and vitamin B12 levels in maternal blood being independent risk factors for NTDs [82]. When taken together with the findings of the randomized controlled trial of vitamin usage in NTD prevention [83], this evidence has led to the recommendation that all women planning a pregnancy should take folic acid (FA)-containing supplements to minimize the risk of NTDs [84]. Moreover, many countries now mandate addition of FA to staple foods, to counteract folate deficiency on a population-wide basis, and so enhance NTD prevention [85].

#### 5.2 Obesity and diabetes as risk factors for NTDs

Maternal obesity and poorly-controlled diabetes mellitus during pregnancy are established risk factors for NTDs [86, 87], although the complexity of the diabetic milieu has made it difficult to pinpoint the precise mechanism(s) by which the diabetic state enhances NTD risk.

The polyol pathway became of attention in the last century. Aldose reductase, key enzyme in this pathway, converts glucose to sorbitol, which is further processed to fructose. Under normal conditions, this enzyme has a low affinity for glucose, and it processes small amount of substrate. However, in diabetes mellitus, the hyperglycemia in some cells causes a marked production of sorbitol with a concomitant reduction of MI concentration.

The depletion of MI has long been thought to be the underlying defect responsible for decreased nerve conduction velocity in experimental diabetes, leading to increased embryo malformations, such as neural tube defects.

Therefore, the potential of aldose reductase inhibitors has been investigated, with the aim to restore MI tissue content. However, treatment with aldose reductase inhibitors failed to correct MI reduction, did not prevent malformations in the embryos and was found associated with significant side effects [88, 89, 90].

The failure of aldose reductase inhibitors to prevent diabetic malformations suggested that the polyol pathway was not involved; in contrast, supplementation with MI restored MI tissue content and reduced the incidence of neural tube defects, suggesting the involvement of MI in the mechanism of diabetic embryopathy [91, 92].

While hyperglycemia alone is sufficient to cause NTDs in cultured rodent embryos [93], other features of the diabetic environment, including the ketone  $\beta$ -hydroxybutyrate, can also produce NTDs [94].

The effects of the diabetic milieu on the developing rodent neural tube include intracellular oxidative stress and neuroepithelial cell apoptosis, with many studies showing that antioxidant treatment can protect against these effects [95, 96]. Nevertheless, precisely how such effects lead to NTDs is unclear. One possibility is a reduction in cellular expression of genes that are vital for neural tube closure, either as a result of cell death or down-regulation of gene expression. For example, *Pax3* loss of function causes both cranial and spinal NTDs [97], and some studies have identified disruption of *Pax3* expression in mouse embryos of diabetic mothers [98].

In humans, hyperinsulinemia has been suggested as a possible mechanism leading to elevated NTD risk in obese mothers who often have type II diabetes [99]. Even in the absence of diabetes, NTDs have been significantly associated with maternal periconceptional increased intake of simple sugars and a high glycemic index, as well as with features of the 'metabolic syndrome' [100, 101, 102]. Hence, there is plentiful evidence linking dysregulation of glucose metabolism, in both types 1 and 2 diabetes, with NTDs.

### 5.3 Inositols and NTDs: evidence from animal models

Diabetic tissues tend to be inositol-deficient, in conjunction with elevated glucose levels [103], and rat embryos cultured under hyperglycemic conditions exhibit diminished MI levels [90]. At neurulation stages, MI deficiency but not other 'vitamin' deficiencies leads to failure of cranial NTDs in cultured rat embryos [104]. Hence, inositol is essential for the neural tube closure process. In terms of prevention, MI supplementation can diminish the frequency of NTDs resulting from hyperglycemic conditions in rats, both in vivo and in embryo culture [105, 106].

Perhaps of most relevance to human NTDs, inositol can reduce the frequency of NTDs in a mouse genetic NTD model [107]. A multifactorial etiology underlies most human NTDs, with genetic predisposing variants interacting with environmental factors. In the *curly tail* mouse, a hypomorphic allele of the *Grhl3* gene is the major genetic change, with 'modifier' genetic variants and various environmental influences affecting frequency and severity of NTDs [108, 109]. Importantly, NTDs in the *curly tail* strain are FA-resistant, therefore mimicking FA non-responsive human NTDs. Both MI and DCI are preventive in the *curly* 

*tail* model, an effect that is dependent on specific isoforms of protein kinase C [110, 111]. This preventive action of MI in the *curly tail* strain has been replicated in an independent laboratory [112]. Furthermore, in a related strain in which cranial NTDs were inducible by folate deficiency, these defects were prevented by maternal MI supplementation [113]. These findings suggest potential overlap in response to FA and MI for some NTDs.

#### 5.4 Inositols and NTDs: evidence from human studies

Diminished MI concentrations have been detected in the plasma of pregnant Dutch women with NTD-affected fetuses. The odds ratio for NTD risk associated with extremely low maternal MI concentration (10-13.2 microM/L) was 2.6 (95% CI, 1.1-6.0) [114]. Moreover, a genetic connection between inositol metabolism and NTDs was suggested by findings that cranial NTDs arise in mice with loss of function of *Itpk1*, encoding inositol 1,3,4-trisphosphate 5/6-kinase which produces the highly phosphorylated metabolite, inositol hexakisphosphate (IP6) [115]. A subsequent study in a high-risk area of China found an association of a maternal polymorphism of *ITPK1* increased risk of NTD pregnancy [116].

## 5.5 Inositol in the prevention of human NTDs

Population-wide supplementation with FA through food fortification has significantly reduced the prevalence of NTDs in many countries [85]. However, FA is only partially effective, and NTD cases continue to occur worldwide, with or without voluntary or mandatory FA consumption. Hence, additional strategies are needed to achieve further prevention.

Building on the data from animal models, an inositol supplementation study was conducted in Italy, among high-risk women who had experienced 1 or 2 previous NTD-affected pregnancies. In their next pregnancy, the women took 500-1000 mg/day MI plus 5 mg/day FA, from 2 months pre-conception, until 60 days of pregnancy. No NTD recurrences were observed among 29 pregnancies of 27 women, whereas 2-8 recurrent NTDs would be expected, based on typical population recurrence frequencies [117]. Most of the women were likely folate-resistant, having undergone appropriate peri-conceptional folate intake in their previous NTD-affected pregnancies. Hence, these findings support a preventive effect of MI on NTD recurrence in high-risk pregnancies.

A phase I/II double-blind, case-control clinical trial (the PONTI study; EudraCT2006-000157-22) was performed to gain further experience of MI supplementation in human pregnancy [118]. The subjects were UK women with a previous NTD pregnancy who planned to become pregnant again. Of 117 women contacted, 99 proved eligible, and 61 undertook a detailed screening questionnaire of whom 47 (77% of those screened) agreed to be randomized to periconceptional supplementation with MI (1 g/day) plus FA (5 mg/day), or placebo plus FA. Of 33 randomized pregnancies, there was one NTD recurrence in the placebo plus FA group (n= 19) and no recurrences in MI plus FA group (n= 14). Of 52 women who declined randomization, the periconceptional supplementation regime and outcomes of 22 further pregnancies were documented. Two NTDs recurred in women who took only FA in their next pregnancy (n= 3), whereas there were no recurrences in women who took MI plus FA in their next pregnancy (n= 19). Overall, NTDs recurred among 0/33 MI-supplemented pregnancies and 3/22 FA-only pregnancies (p = 0.06, Fisher's Exact test).

Combining data from the Italian and PONTI studies, with post-PONTI experience of MI use, a total of 76 women at high risk of NTDs (52 with one and 24 with two or more previous affected pregnancies) have taken MI plus FA in a subsequent pregnancy, with no recurrences. Assuming a 3% recurrence risk after one NTD pregnancy and 10% after two [119], as used in genetic counselling, this indicates that 4 recurrences would be expected among 76 pregnancies. The observed 0/76 recurrences among MI-supplemented pregnancies are significantly different from this expectation (Single Proportion Fisher's Exact, p= 0.016). These observations, although on small patient numbers are encouraging that MI may have value in increasing NTD prevention beyond that achievable by FA alone.

#### 5.6 Inositol and NTD prevention: next steps

A definitive answer to the question of the efficacy of MI in enhancing NTD prevention is likely to require a fully powered, randomized clinical trial (RCT). However, this will necessitate large numbers of pregnancies, owing to the relatively low recurrence rate for NTDs. Given the current state of knowledge about inositol and NTD prevention, it is important to consider whether a RCT is ethically acceptable, given that the study design would require withholding inositol supplements from a significant number of pregnancies.

Glasziou et al [120] argue that expensive and time-consuming RCTs may be unnecessary in cases where a novel treatment has an obvious effect (i.e. a large 'signal to noise' ratio). However, for NTDs, the effect of inositol at the population level is likely to be small, as NTD frequency is relatively low (0.1% for first occurrence; 2-3% for recurrence), and any effect of inositol will be additional to FA-mediated prevention. On the other hand, for individual women who exhibit FA-resistance, inositol is currently their only option to increase the likelihood of a normal pregnancy.

Freedman [121] suggested that 'clinical equipoise' should be considered to exist when there is genuine uncertainty within the expert medical community about the preferred treatment. Under such circumstances, an RCT would be ethically justified. In the case of NTDs, MI + FA supplements certainly have not superseded FA-only supplements; worldwide, the great majority of low-risk and high-risk women receive FA only (or no supplement at all). To date very few pregnancies have been supplemented with inositol, and this has been used largely where high-risk women have sought advice from the authors of this article.

We conclude that the Glasziou criterion for avoiding an RCT is not met by MI supplementation for NTDs, and that currently the expert field is in a state of clinical equipoise with regard to use of MI in addition to FA. Hence, an RCT appears indicated to provide a definitive answer to the efficacy of MI as an adjunct preventive supplement for NTDs.

## 6 Conclusions

Gestational diabetes mellitus (GDM) and congenital abnormalities of fetal central nervous system, such as neural tube defects (NTDs), are strongly correlated to glycemic status of pregnant women. Before starting a therapeutic intervention to achieve a healthy pregnancy, the principle approach to reach the glycemic targets is based on lifestyle changes by means of an adequate diet and exercise. When glycemic targets are not addressed, an intervention is reasonable to avoid complications both for the mothers and for the fetus.

A supplementation of MI starting at least one month prior to conception and continued until 36<sup>th</sup> week of pregnancy is expected to prevent NTDs as well as GDM onset and its related negative perinatal outcomes. As a side effects free molecule at the usually administered doses, MI therefore becomes a candidate for being one of the first tools to be used in pre-conceptional medicine.

# 7 Expert opinion

The data reported in this brief narrative review are highly encouraging over the use of inositols, namely MI, in pregnant women. To date, the controlled studies available in literature are reassuring about its safety and tolerability. Adverse effects in the short-term period have not reported yet, either for mothers or babies, despite the widespread use of these supplements since several years, in many countries.

Although confirmatory studies will be necessary to elucidate the mechanisms involved [122], there is a general agreement that MI administered early in pregnancy effectively prevents GDM onset (figure 1). Proofs are available for different categories of women at risk, namely overweight, obese, and patients affected by PCOS. In addition, significant reduction of gestational hypertensive disorders, preterm birth and large for gestational age babies suggest a perinatal benefit of MI supplementation. Whether this would apply to low-risk population requires the design of appropriate controlled trials, hopefully comparing different inositols combinations.

To date, although several clinical trials are ongoing, MI supplementation has not yet been inserted in clinical guidelines for GDM prevention/treatment.

For congenital malformations like NTDs, MI can offer a novel means of prevention via peri-conceptional supplementation, particularly in those women whose previous pregnancies have proven resistant to FA supplementation (figure 1).

Thus, given the extensive data documenting the safety of peri-conceptional MI supplementation derived from trials done in Assisted Reproduction, a combination of MI and FA could be recommended for every woman at high risk for NTDs.

Furthermore, it would be interesting to carry out studies to reduce the dose and perhaps improve absorption and bioavailability. Recent research using co-administration of MI and  $\alpha$ -LA [26] is an example of how MI usage may be clinically improved. We believe that

this association in the next years could conquer a relevant position as insulin-sensitizing treatment for GDM and NTDs prevention, due to both efficacy and absence of side effects.

# Funding

This work was not funded

## References

- Rogozinska E, Marlin N, Jackson L, et al. Effects of antenatal diet and physical activity on maternal and fetal outcomes: individual patient data meta-analysis and health economic evaluation. Health Technol Assess. 2017; Aug; 21 (41) 1–158. DOI: 10.3310/hta21410 [PubMed: 28795682]
- McGowan CA, McAuliffe FM. The influence of maternal glycaemia and dietary glycaemic index on pregnancy outcome in healthy mothers. Br J Nutr. 2010; Jul; 104 (2) 153–9. DOI: 10.1017/ S0007114510000425 [PubMed: 20307352]
- Group HSCR. Metzger BE, Lowe LP, et al. Hyperglycemia and adverse pregnancy outcomes. N Engl J Med. 2008; May 8; 358 (19) 1991–2002. DOI: 10.1056/NEJMoa0707943 [PubMed: 18463375]
- Safi J, Joyeux L, Chalouhi GE. Periconceptional folate deficiency and implications in neural tube defects. J Pregnancy. 2012; 2012 295083 doi: 10.1155/2012/295083 [PubMed: 22900183]
- Werner EF, Froehlich RJ. The Potential Role for Myoinositol in the Prevention of Gestational Diabetes Mellitus. Am J Perinatol. 2016; Nov; 33 (13) 1236–1241. DOI: 10.1055/s-0036-1584273 [PubMed: 27213831]
- Noventa M, Vitagliano A, Quaranta M, et al. Preventive and Therapeutic Role of Dietary Inositol Supplementation in Periconceptional Period and During Pregnancy: A Summary of Evidences and Future Applications. Reprod Sci. 2016; Mar; 23 (3) 278–88. DOI: 10.1177/1933719115594018 [PubMed: 26175275]
- 7. Pak Y, Huang LC, Lilley KJ, et al. In vivo conversion of [3H]myoinositol to [3H]chiroinositol in rat tissues. J Biol Chem. 1992; Aug 25; 267 (24) 16904–10. [PubMed: 1387397]
- Beemster P, Groenen P, Steegers-Theunissen R. Involvement of inositol in reproduction. Nutr Rev. 2002; Mar; 60 (3) 80–7. DOI: 10.1301/00296640260042748 [PubMed: 11908744]
- Wong YH, Kalmbach SJ, Hartman BK, et al. Immunohistochemical staining and enzyme activity measurements show myo-inositol-1-phosphate synthase to be localized in the vasculature of brain. J Neurochem. 1987; May; 48 (5) 1434–42. DOI: 10.1111/j.1471-4159.1987.tb05682.x [PubMed: 2435847]
- 10. Loewus MW, Loewus FA, Brillinger GU, et al. Stereochemistry of the myo-inositol-1-phosphate synthase reaction. J Biol Chem. 1980; Dec 25; 255 (24) 11710–2. [PubMed: 7002927]
- Michell RH. Inositol derivatives: evolution and functions. Nat Rev Mol Cell Biol. 2008; Feb; 9 (2) 151–61. DOI: 10.1038/nrm2334 [PubMed: 18216771]
- Monastra G, Unfer V, Harrath AH, et al. Combining treatment with myo-inositol and D-chiroinositol (40:1) is effective in restoring ovary function and metabolic balance in PCOS patients. Gynecol Endocrinol. 2017; Jan; 33 (1) 1–9. DOI: 10.1080/09513590.2016.1247797 [PubMed: 27898267]
- Schneider S. Inositol transport proteins. FEBS Lett. 2015; Apr 28; 589 (10) 1049–58. DOI: 10.1016/j.febslet.2015.03.012 [PubMed: 25819438]
- Aouameur R, Da Cal S, Bissonnette P, et al. SMIT2 mediates all myo-inositol uptake in apical membranes of rat small intestine. Am J Physiol Gastrointest Liver Physiol. 2007; Dec; 293 (6) G1300–7. DOI: 10.1152/ajpgi.00422.2007 [PubMed: 17932225]
- Di Paolo G, De Camilli P. Phosphoinositides in cell regulation and membrane dynamics. Nature. 2006; Oct 12; 443 (7112) 651–7. DOI: 10.1038/nature05185 [PubMed: 17035995]
- Bizzarri M, Fuso A, Dinicola S, et al. Pharmacodynamics and pharmacokinetics of inositol(s) in health and disease. Expert Opin Drug Metab Toxicol. 2016; Oct; 12 (10) 1181–96. DOI: 10.1080/17425255.2016.1206887 [PubMed: 27351907]

- Lagana AS, Garzon S, Casarin J, et al. Inositol in Polycystic Ovary Syndrome: Restoring Fertility through a Pathophysiology-Based Approach. Trends Endocrinol Metab. 2018; Nov; 29 (11) 768– 780. DOI: 10.1016/j.tem.2018.09.001 [PubMed: 30270194]
- Nestler JE, Unfer V. Reflections on inositol(s) for PCOS therapy: steps toward success. Gynecol Endocrinol. 2015; Jul; 31 (7) 501–5. DOI: 10.3109/09513590.2015.1054802 [PubMed: 26177098]
- Facchinetti F, Appetecchia M, Aragona C, et al. Experts' opinion on inositols in treating polycystic ovary syndrome and non-insulin dependent diabetes mellitus: a further help for human reproduction and beyond. Expert Opin Drug Metab Toxicol. 2020; Mar; 16 (3) 255–274. DOI: 10.1080/17425255.2020.1737675 [PubMed: 32129111]
- Unfer V, Nestler JE, Kamenov ZA, et al. Effects of Inositol(s) in Women with PCOS: A Systematic Review of Randomized Controlled Trials. Int J Endocrinol. 2016; 2016 1849162 doi: 10.1155/2016/1849162 [PubMed: 27843451]
- Croze ML, Soulage CO. Potential role and therapeutic interests of myo-inositol in metabolic diseases. Biochimie. 2013; Oct; 95 (10) 1811–27. DOI: 10.1016/j.biochi.2013.05.011 [PubMed: 23764390]
- 22. Sun TH, Heimark DB, Nguygen T, et al. Both myo-inositol to chiro-inositol epimerase activities and chiro-inositol to myo-inositol ratios are decreased in tissues of GK type 2 diabetic rats compared to Wistar controls. Biochem Biophys Res Commun. 2002; May 10; 293 (3) 1092–8. DOI: 10.1016/S0006-291X(02)00313-3 [PubMed: 12051772]
- Heimark D, McAllister J, Larner J. Decreased myo-inositol to chiro-inositol (M/C) ratios and increased M/C epimerase activity in PCOS theca cells demonstrate increased insulin sensitivity compared to controls. Endocr J. 2014; 61 (2) 111–7. DOI: 10.1507/endocrj.ej13-0423 [PubMed: 24189751]
- Chukwuma CI, Ibrahim MA, Islam MS. Myo-inositol inhibits intestinal glucose absorption and promotes muscle glucose uptake: a dual approach study. J Physiol Biochem. 2016; Dec; 72 (4) 791–801. DOI: 10.1007/s13105-016-0517-1 [PubMed: 27601253]
- Milewska EM, Czyzyk A, Meczekalski B, et al. Inositol and human reproduction. From cellular metabolism to clinical use. Gynecol Endocrinol. 2016; Sep; 32 (9) 690–695. DOI: 10.1080/09513590.2016.1188282 [PubMed: 27595157]
- Monastra G, Sambuy Y, Ferruzza S, et al. Alpha-lactalbumin Effect on Myo-inositol Intestinal Absorption: In vivo and In vitro. Curr Drug Deliv. 2018; 15 (9) 1305–1311. DOI: 10.2174/1567201815666180509102641 [PubMed: 29745333]
- Permyakov EA, Berliner LJ. alpha-Lactalbumin: structure and function. FEBS Lett. 2000; May 19; 473 (3) 269–74. DOI: 10.1016/s0014-5793(00)01546-5 [PubMed: 10818224]
- Montanino Oliva M, Buonomo G, Calcagno M, et al. Effects of myo-inositol plus alphalactalbumin in myo-inositol-resistant PCOS women. J Ovarian Res. 2018; May 10. 11 (1) 38. doi: 10.1186/s13048-018-0411-2 [PubMed: 29747700]
- Carlomagno G, Unfer V. Inositol safety: clinical evidences. Eur Rev Med Pharmacol Sci. 2011; Aug; 15 (8) 931–6. [PubMed: 21845803]
- Vitagliano A, Saccone G, Cosmi E, et al. Inositol for the prevention of gestational diabetes: a systematic review and meta-analysis of randomized controlled trials. Arch Gynecol Obstet. 2019; Jan; 299 (1) 55–68. DOI: 10.1007/s00404-018-5005-0 [PubMed: 30564926]
- Crawford TJ, Crowther CA, Alsweiler J, et al. Antenatal dietary supplementation with myoinositol in women during pregnancy for preventing gestational diabetes. Cochrane Database Syst Rev. 2015; Dec. 17 (12) CD011507 doi: 10.1002/14651858.CD011507.pub2 [PubMed: 26678256]
- 32. Zhu Y, Zhang C. Prevalence of Gestational Diabetes and Risk of Progression to Type 2 Diabetes: a Global Perspective. Curr Diab Rep. 2016; Jan. 16 (1) 7. doi: 10.1007/s11892-015-0699-x [PubMed: 26742932]
- Chiefari E, Arcidiacono B, Foti D, et al. Gestational diabetes mellitus: an updated overview. J Endocrinol Invest. 2017; Sep; 40 (9) 899–909. DOI: 10.1007/s40618-016-0607-5 [PubMed: 28283913]
- 34. Immanuel J, Simmons D. Screening and Treatment for Early-Onset Gestational Diabetes Mellitus: a Systematic Review and Meta-analysis. Curr Diab Rep. 2017; Oct 2. 17 (11) 115. doi: 10.1007/ s11892-017-0943-7 [PubMed: 28971305]

- Baz B, Riveline JP, Gautier JF. ENDOCRINOLOGY OF PREGNANCY: Gestational diabetes mellitus: definition, aetiological and clinical aspects. Eur J Endocrinol. 2016; Feb; 174 (2) R43– 51. DOI: 10.1530/EJE-15-0378 [PubMed: 26431552]
- 36. International Association of D, Pregnancy Study Groups Consensus P. Metzger BE, et al. International association of diabetes and pregnancy study groups recommendations on the diagnosis and classification of hyperglycemia in pregnancy. Diabetes Care. 2010; Mar; 33 (3) 676–82. DOI: 10.2337/dc09-1848 [PubMed: 20190296]
- Khalifeh A, Eckler R, Felder L, et al. One-step versus two-step diagnostic testing for gestational diabetes: a randomized controlled trial. J Matern Fetal Neonatal Med. 2020; Feb; 33 (4) 612–617. DOI: 10.1080/14767058.2018.1498480 [PubMed: 29985079]
- Agarwal MM. Gestational diabetes mellitus: An update on the current international diagnostic criteria. World J Diabetes. 2015; Jun 25; 6 (6) 782–91. DOI: 10.4239/wjd.v6.i6.782 [PubMed: 26131321]
- 39. Smirnakis KV, Plati A, Wolf M, et al. Predicting gestational diabetes: choosing the optimal early serum marker. Am J Obstet Gynecol. 2007; Apr; 196 (4) 410 e1–6. DOI: 10.1016/ j.ajog.2006.12.011 [PubMed: 17403439]
- Nanda S, Savvidou M, Syngelaki A, et al. Prediction of gestational diabetes mellitus by maternal factors and biomarkers at 11 to 13 weeks. Prenat Diagn. 2011; Feb; 31 (2) 135–41. DOI: 10.1002/ pd.2636 [PubMed: 21268030]
- Rasanen JP, Snyder CK, Rao PV, et al. Glycosylated fibronectin as a first-trimester biomarker for prediction of gestational diabetes. Obstet Gynecol. 2013; Sep; 122 (3) 586–94. DOI: 10.1097/ AOG.0b013e3182a0c88b [PubMed: 23921871]
- 42. Sahu P, P N, Dubey RD, Paroha S, Chatterjee S, Chatterjee T. Biomarkers: An emerging tool for diagnosis of a disease and drug development. Asian J Res Pharm Sci. 2011; 1: 9–16.
- Wu L, Cui L, Tam WH, et al. Genetic variants associated with gestational diabetes mellitus: a meta-analysis and subgroup analysis. Sci Rep. 2016; Jul 29. 6 30539 doi: 10.1038/srep30539 [PubMed: 27468700]
- 44. Dias S, Pheiffer C, Abrahams Y, et al. Molecular Biomarkers for Gestational Diabetes Mellitus. Int J Mol Sci. 2018; Sep 26. 19 (10) doi: 10.3390/ijms19102926 [PubMed: 30261627]
- 45. Houde AA, Guay SP, Desgagne V, et al. Adaptations of placental and cord blood ABCA1 DNA methylation profile to maternal metabolic status. Epigenetics. 2013; Dec; 8 (12) 1289–302. DOI: 10.4161/epi.26554 [PubMed: 24113149]
- Haertle L, El Hajj N, Dittrich M, et al. Epigenetic signatures of gestational diabetes mellitus on cord blood methylation. Clin Epigenetics. 2017; 9: 28. doi: 10.1186/s13148-017-03293 [PubMed: 28360945]
- 47. Wang K, Yuan Y, Cho JH, et al. Comparing the MicroRNA spectrum between serum and plasma. PLoS One. 2012; 7 (7) e41561 doi: 10.1371/journal.pone.0041561 [PubMed: 22859996]
- Group HSCR. Hyperglycemia and Adverse Pregnancy Outcome (HAPO) Study: associations with neonatal anthropometrics. Diabetes. 2009; Feb; 58 (2) 453–9. DOI: 10.2337/db081112 [PubMed: 19011170]
- 49. Shah BR, Sharifi F. Perinatal outcomes for untreated women with gestational diabetes by IADPSG criteria: a population-based study. BJOG. 2020; Jan; 127 (1) 116–122. DOI: 10.1111/1471-0528.15964 [PubMed: 31553136]
- Bellamy L, Casas JP, Hingorani AD, et al. Type 2 diabetes mellitus after gestational diabetes: a systematic review and meta-analysis. Lancet. 2009; May 23; 373 (9677) 1773–9. DOI: 10.1016/ S0140-6736(09)607315 [PubMed: 19465232]
- Schwartz N, Nachum Z, Green MS. The prevalence of gestational diabetes mellitus recurrenceeffect of ethnicity and parity: a metaanalysis. Am J Obstet Gynecol. 2015; Sep; 213 (3) 310–7. DOI: 10.1016/j.ajog.2015.03.011 [PubMed: 25757637]
- 52. Herring SJ, Oken E. Obesity and diabetes in mothers and their children: can we stop the intergenerational cycle? Curr Diab Rep. 2011; Feb; 11 (1) 20–7. DOI: 10.1007/s11892-010-01569 [PubMed: 20963519]

- 53. Battista MC, Hivert MF, Duval K, et al. Intergenerational cycle of obesity and diabetes: how can we reduce the burdens of these conditions on the health of future generations? Exp Diabetes Res. 2011; 2011 596060 doi: 10.1155/2011/596060 [PubMed: 22110473]
- 54. Lowe WL Jr, Scholtens DM, Kuang A, et al. Hyperglycemia and Adverse Pregnancy Outcome Follow-up Study (HAPO FUS): Maternal Gestational Diabetes Mellitus and Childhood Glucose Metabolism. Diabetes Care. 2019; Mar; 42 (3) 372–380. DOI: 10.2337/dc181646 [PubMed: 30655380]
- 55. International Weight Management in Pregnancy Collaborative G. Effect of diet and physical activity based interventions in pregnancy on gestational weight gain and pregnancy outcomes: meta-analysis of individual participant data from randomised trials. BMJ. 2017; Jul 19. 358 j3119 doi: 10.1136/bmj.j3119 [PubMed: 28724518]
- 56. Bruno R, Petrella E, Bertarini V, et al. Adherence to a lifestyle programme in overweight/obese pregnant women and effect on gestational diabetes mellitus: a randomized controlled trial. Matern Child Nutr. 2017; Jul. 13 (3) doi: 10.1111/mcn.12333 [PubMed: 27647837]
- Crowther CA, Hiller JE, Moss JR, et al. Effect of treatment of gestational diabetes mellitus on pregnancy outcomes. N Engl J Med. 2005; Jun 16; 352 (24) 2477–86. DOI: 10.1056/ NEJMoa042973 [PubMed: 15951574]
- 58. Simeonova-Krstevska S, Bogoev M, Bogoeva K, et al. Maternal and Neonatal Outcomes in Pregnant Women with Gestational Diabetes Mellitus Treated with Diet, Metformin or Insulin. Open Access Maced J Med Sci. 2018; May 20; 6 (5) 803–807. DOI: 10.3889/oamjms.2018.200 [PubMed: 29875849]
- Begum MR, Khanam NN, Quadir E, et al. Prevention of gestational diabetes mellitus by continuing metformin therapy throughout pregnancy in women with polycystic ovary syndrome. J Obstet Gynaecol Res. 2009; Apr; 35 (2) 282–6. DOI: 10.1111/j.1447-0756.2008.00876.x [PubMed: 19708174]
- 60. Glueck CJ, Goldenberg N, Wang P, et al. Metformin during pregnancy reduces insulin,insulin resistance, insulin secretion, weight, testosterone and development of gestational diabetes: prospective longitudinal assessment of women with polycystic ovary syndrome from preconception throughout pregnancy. Hum Reprod. 2004; Mar; 19 (3) 510–21. DOI: 10.1093/ humrep/deh109 [PubMed: 14998944]
- Glueck CJ, Pranikoff J, Aregawi D, et al. Prevention of gestational diabetes by metformin plus diet in patients with polycystic ovary syndrome. Fertil Steril. 2008; Mar; 89 (3) 625–34. DOI: 10.1016/j.fertnstert.2007.03.036 [PubMed: 17678910]
- Vanky E, Stridsklev S, Heimstad R, et al. Metformin versus placebo from first trimester to delivery in polycystic ovary syndrome: a randomized, controlled multicenter study. J Clin Endocrinol Metab. 2010; Dec; 95 (12) E448–55. DOI: 10.1210/jc.20100853 [PubMed: 20926533]
- 63. Syngelaki A, Nicolaides KH, Balani J, et al. Metformin versus Placebo in Obese Pregnant Women without Diabetes Mellitus. N Engl J Med. 2016; Feb 4; 374 (5) 434–43. DOI: 10.1056/ NEJMoa1509819 [PubMed: 26840133]
- 64. Chiswick C, Reynolds RM, Denison F, et al. Effect of metformin on maternal and fetal outcomes in obese pregnant women (EMPOWaR): a randomised, double-blind, placebo-controlled trial. Lancet Diabetes Endocrinol. 2015; Oct; 3 (10) 778–86. DOI: 10.1016/S2213-8587(15)002193 [PubMed: 26165398]
- 65. Celentano C, Matarrelli B, Pavone G, et al. The influence of different inositol stereoisomers supplementation in pregnancy on maternal gestational diabetes mellitus and fetal outcomes in high-risk patients: a randomized controlled trial. J Matern Fetal Neonatal Med. 2020; Mar; 33 (5) 743–751. DOI: 10.1080/14767058.2018.1500545 [PubMed: 30558466]
- 66. Garzon S, Lagana AS, Monastra G. Risk of reduced intestinal absorption of myo-inositol caused by D-chiro-inositol or by glucose transporter inhibitors. Expert Opin Drug Metab Toxicol. 2019; Aug 8. 1–7. DOI: 10.1080/17425255.2019.1651839 [PubMed: 30582386]
- 67. Santamaria A, Alibrandi A, Di Benedetto A, et al. Clinical and metabolic outcomes in pregnant women at risk for gestational diabetes mellitus supplemented with myo-inositol: a secondary analysis from 3 RCTs. Am J Obstet Gynecol. 2018; Sep; 219 (3) 300 e1–300 e6. DOI: 10.1016/ j.ajog.2018.05.018 [PubMed: 29859136]

- 68. Facchinetti F, V V, Petrella Elisabetta, Gambigliani Zoccoli Sofia, Pignatti Lucrezia, Di Cerbo Lidia, Neri Elisabetta. Food glycemic index changes in overweight/obese pregnant women enrolled in a lifestyle program: a randomized controlled trial. AJOG MFM. 2019; 1 (3) 100030 [PubMed: 33345794]
- 69. D'Anna R, Di Benedetto V, Rizzo P, et al. Myo-inositol may prevent gestational diabetes in PCOS women. Gynecol Endocrinol. 2012; Jun; 28 (6) 440–2. DOI: 10.3109/09513590.2011.633665 [PubMed: 22122627]
- 70. Ferrari F, Facchinetti F, Ontiveros AE, et al. The effect of combined inositol supplementation on maternal metabolic profile in pregnancies complicated by metabolic syndrome and obesity. Am J Obstet Gynecol. 2016; Oct; 215 (4) 503 e1–8. DOI: 10.1016/j.ajog.2016.05.038 [PubMed: 27255472]
- Longo M, Alrais M, Tamayo EH, et al. Vascular and metabolic profiles in offspring born to pregnant mice with metabolic syndrome treated with inositols. Am J Obstet Gynecol. 2019; Mar; 220 (3) 279 e1–279 e9. DOI: 10.1016/j.ajog.2018.11.1101 [PubMed: 30521799]
- 72. Matarrelli B, Vitacolonna E, D'Angelo M, et al. Effect of dietary myo-inositol supplementation in pregnancy on the incidence of maternal gestational diabetes mellitus and fetal outcomes: a randomized controlled trial. J Matern Fetal Neonatal Med. 2013; Jul; 26 (10) 967–72. DOI: 10.3109/14767058.2013.766691 [PubMed: 23327487]
- 73. D'Anna R, Scilipoti A, Giordano D, et al. myo-Inositol supplementation and onset of gestational diabetes mellitus in pregnant women with a family history of type 2 diabetes: a prospective, randomized, placebo-controlled study. Diabetes Care. 2013; Apr; 36 (4) 854–7. DOI: 10.2337/ dc121371 [PubMed: 23340885]
- 74. Malvasi A, Casciaro F, Minervini MM, et al. Myo-inositol, D-chiro-inositol, folic acid and manganese in second trimester of pregnancy: a preliminary investigation. Eur Rev Med Pharmacol Sci. 2014; 18 (2) 270–4. [PubMed: 24488919]
- 75. D'Anna R, Di Benedetto A, Scilipoti A, et al. Myo-inositol Supplementation for Prevention of Gestational Diabetes in Obese Pregnant Women: A Randomized Controlled Trial. Obstet Gynecol. 2015; Aug; 126 (2) 310–5. DOI: 10.1097/AOG.00000000000958 [PubMed: 26241420]
- 76. Santamaria A, Di Benedetto A, Petrella E, et al. Myo-inositol may prevent gestational diabetes onset in overweight women: a randomized, controlled trial. J Matern Fetal Neonatal Med. 2016; Oct; 29 (19) 3234–7. DOI: 10.3109/14767058.2015.1121478 [PubMed: 26698911]
- 77. Lubin V, Shojai R, Darmon P, et al. A pilot study of gestational diabetes mellitus not controlled by diet alone: First-line medical treatment with myoinositol may limit the need for insulin. Diabetes Metab. 2016; Jun; 42 (3) 192–5. DOI: 10.1016/j.diabet.2016.01.005 [PubMed: 26948394]
- Farren M, Daly N, McKeating A, et al. The Prevention of Gestational Diabetes Mellitus With Antenatal Oral Inositol Supplementation: A Randomized Controlled Trial. Diabetes Care. 2017; Jun; 40 (6) 759–763. DOI: 10.2337/dc162449 [PubMed: 28325784]
- Fraticelli F, Celentano C, Zecca IA, et al. Effect of inositol stereoisomers at different dosages in gestational diabetes: an open-label, parallel, randomized controlled trial. Acta Diabetol. 2018; Aug; 55 (8) 805–812. DOI: 10.1007/s00592-018-11574 [PubMed: 29774465]
- Pintaudi B, Di Vieste G, Corrado F, et al. Effects of myo-inositol on glucose variability in women with gestational diabetes. Eur Rev Med Pharmacol Sci. 2018; Oct; 22 (19) 6567–6572. DOI: 10.26355/eurrev\_201810\_16073 [PubMed: 30338829]
- Copp AJ, Stanier P, Greene ND. Neural tube defects: recent advances, unsolved questions, and controversies. Lancet Neurol. 2013; Aug; 12 (8) 799–810. DOI: 10.1016/S1474-4422(13)701108 [PubMed: 23790957]
- Kirke PN, Molloy AM, Daly LE, et al. Maternal plasma folate and vitamin B12 are independent risk factors for neural tube defects. Q J Med. 1993; Nov; 86 (11) 703–8. [PubMed: 8265769]
- Prevention of neural tube defects: results of the Medical Research Council Vitamin Study. MRC Vitamin Study Research Group. Lancet. 1991; Jul 20; 338 (8760) 131–7. [PubMed: 1677062]
- 84. Oakley GP Jr, Bell KN, Weber MB. Recommendations for accelerating global action to prevent folic acid-preventable birth defects and other folate-deficiency diseases: meeting of experts on preventing folic acid-preventable neural tube defects. Birth Defects Res A Clin Mol Teratol. 2004; Nov; 70 (11) 835–7. DOI: 10.1002/bdra.20058 [PubMed: 15390317]

- Kancherla V, Wagh K, Johnson Q, et al. A 2017 global update on folic acid-preventable spina bifida and anencephaly. Birth Defects Res. 2018; Aug 15; 110 (14) 1139–1147. DOI: 10.1002/ bdr2.1366 [PubMed: 30070772]
- 86. Rasmussen SA, Chu SY, Kim SY, et al. Maternal obesity and risk of neural tube defects: a metaanalysis. Am J Obstet Gynecol. 2008; Jun; 198 (6) 611–9. DOI: 10.1016/j.ajog.2008.04.021 [PubMed: 18538144]
- Kucera J. Rate and type of congenital anomalies among offspring of diabetic women. J Reprod Med. 1971; Aug; 7 (2) 73–82. [PubMed: 5095696]
- Eriksson UJ, Naeser P, Brolin SE. Increased accumulation of sorbitol in offspring of manifest diabetic rats. Diabetes. 1986; Dec; 35 (12) 1356–63. DOI: 10.2337/diab.35.12.1356 [PubMed: 3770312]
- Eriksson UJ, Brolin SE, Naeser P. Influence of sorbitol accumulation on growth and development of embryos cultured in elevated levels of glucose and fructose. Diabetes Res. 1989; May; 11 (1) 27–32. [PubMed: 2515932]
- 90. Hod M, Star S, Passonneau JV, et al. Effect of hyperglycemia on sorbitol and myo-inositol content of cultured rat conceptus: failure of aldose reductase inhibitors to modify myo-inositol depletion and dysmorphogenesis. Biochem Biophys Res Commun. 1986; Nov 14; 140 (3) 974–80. DOI: 10.1016/0006-291x(86)90731-x [PubMed: 3096334]
- Hod M, Star S, Passonneau J, et al. Glucose-induced dysmorphogenesis in the cultured rat conceptus: prevention by supplementation with myo-inositol. Isr J Med Sci. 1990; Oct; 26 (10) 541–4. [PubMed: 2249926]
- 92. Zabihi S, Loeken MR. Understanding diabetic teratogenesis: where are we now and where are we going? Birth Defects Res A Clin Mol Teratol. 2010; Oct; 88 (10) 779–90. DOI: 10.1002/ bdra.20704 [PubMed: 20706996]
- 93. Cockroft DL, Coppola PT. Teratogenic effects of excess glucose on head-fold rat embryos in culture. Teratology. 1977; Oct; 16 (2) 141–6. DOI: 10.1002/tera.1420160205 [PubMed: 412267]
- 94. Hunter ES 3rd, Sadler TW. D-(-)-beta-hydroxybutyrate-induced effects on mouse embryos in vitro. Teratology. 1987; Oct; 36 (2) 259–64. DOI: 10.1002/tera.1420360214 [PubMed: 3424210]
- 95. Reece EA, Wu YK. Prevention of diabetic embryopathy in offspring of diabetic rats with use of a cocktail of deficient substrates and an antioxidant. Am J Obstet Gynecol. 1997; Apr; 176 (4) 790–7. DOI: 10.1016/s0002-9378(97)706021 [PubMed: 9125602]
- 96. Zhong J, Xu C, Gabbay-Benziv R, et al. Superoxide dismutase 2 overexpression alleviates maternal diabetes-induced neural tube defects, restores mitochondrial function and suppresses cellular stress in diabetic embryopathy. Free Radic Biol Med. 2016; Jul. 96: 234–44. DOI: 10.1016/ j.freeradbiomed.2016.04.030 [PubMed: 27130031]
- 97. Greene ND, Massa V, Copp AJ. Understanding the causes and prevention of neural tube defects: Insights from the splotch mouse model. Birth Defects Res A Clin Mol Teratol. 2009; Apr; 85 (4) 322–30. DOI: 10.1002/bdra.20539 [PubMed: 19180568]
- Fine EL, Horal M, Chang TI, et al. Evidence that elevated glucose causes altered gene expression, apoptosis, and neural tube defects in a mouse model of diabetic pregnancy. Diabetes. 1999; Dec; 48 (12) 2454–62. DOI: 10.2337/diabetes.48.12.2454 [PubMed: 10580436]
- Hendricks KA, Nuno OM, Suarez L, et al. Effects of hyperinsulinemia and obesity on risk of neural tube defects among Mexican Americans. Epidemiology. 2001; Nov; 12 (6) 630–5. DOI: 10.1097/00001648-200111000-00009 [PubMed: 11679789]
- 100. Shaw GM, Quach T, Nelson V, et al. Neural tube defects associated with maternal periconceptional dietary intake of simple sugars and glycemic index. Am J Clin Nutr. 2003; Nov; 78 (5) 972–8. DOI: 10.1093/ajcn/78.5.972 [PubMed: 14594784]
- 101. Yazdy MM, Liu S, Mitchell AA, et al. Maternal dietary glycemic intake and the risk of neural tube defects. Am J Epidemiol. 2010; Feb 15; 171 (4) 407–14. DOI: 10.1093/aje/kwp395 [PubMed: 20042435]
- 102. Ray JG, Thompson MD, Vermeulen MJ, et al. Metabolic syndrome features and risk of neural tube defects. BMC Pregnancy Childbirth. 2007; Sep 19. 7: 21. doi: 10.1186/1471-2393-721 [PubMed: 17880716]

- 103. Greene DA. Sorbitol, myo-inositol and sodium-potassium ATPase in diabetic peripheral nerve. Drugs. 1986; 32 (Suppl 2) 6–14. DOI: 10.2165/00003495-198600322-00004 [PubMed: 3024950]
- 104. Cockroft DL, Brook FA, Copp AJ. Inositol deficiency increases the susceptibility to neural tube defects of genetically predisposed (curly tail) mouse embryos in vitro. Teratology. 1992; Feb; 45 (2) 223–32. DOI: 10.1002/tera.1420450216 [PubMed: 1615432]
- 105. Baker L, Piddington R, Goldman A, et al. Myo-inositol and prostaglandins reverse the glucose inhibition of neural tube fusion in cultured mouse embryos. Diabetologia. 1990; Oct; 33 (10) 593–6. DOI: 10.1007/bf00400202 [PubMed: 2257996]
- 106. Khandelwal M, Reece EA, Wu YK, et al. Dietary myo-inositol therapy in hyperglycemia-induced embryopathy. Teratology. 1998; Feb; 57 (2) 79–84. DOI: 10.1002/ (SICI)1096-9926(199802)57:279::AID-TERA63.0.CO;2-1 [PubMed: 9562680]
- 107. Greene ND, Copp AJ. Inositol prevents folate-resistant neural tube defects in the mouse. Nat Med. 1997; Jan; 3 (1) 60–6. DOI: 10.1038/nm0197-60 [PubMed: 8986742]
- 108. Gustavsson P, Copp AJ, Greene ND. Grainyhead genes and mammalian neural tube closure. Birth Defects Res A Clin Mol Teratol. 2008; Oct; 82 (10) 728–35. DOI: 10.1002/bdra.20494 [PubMed: 18683893]
- 109. De Castro SC, Malhas A, Leung KY, et al. Lamin b1 polymorphism influences morphology of the nuclear envelope, cell cycle progression, and risk of neural tube defects in mice. PLoS Genet. 2012; 8 (11) e1003059 doi: 10.1371/journal.pgen.1003059 [PubMed: 23166514]
- 110. Cogram P, Tesh S, Tesh J, et al. D-chiro-inositol is more effective than myo-inositol in preventing folate-resistant mouse neural tube defects. Hum Reprod. 2002; Sep; 17 (9) 2451–8. DOI: 10.1093/humrep/17.9.2451 [PubMed: 12202440]
- 111. Cogram P, Hynes A, Dunlevy LP, et al. Specific isoforms of protein kinase C are essential for prevention of folate-resistant neural tube defects by inositol. Hum Mol Genet. 2004; Jan 1; 13 (1) 7–14. DOI: 10.1093/hmg/ddh003 [PubMed: 14613966]
- 112. Ting SB, Wilanowski T, Auden A, et al. Inositol-and folate-resistant neural tube defects in mice lacking the epithelial-specific factor Grhl-3. Nat Med. 2003; Dec; 9 (12) 1513–9. DOI: 10.1038/ nm961 [PubMed: 14608380]
- 113. Burren KA, Scott JM, Copp AJ, et al. The genetic background of the curly tail strain confers susceptibility to folate-deficiency-induced exencephaly. Birth Defects Res A Clin Mol Teratol. 2010; Feb; 88 (2) 76–83. DOI: 10.1002/bdra.20632 [PubMed: 19824061]
- 114. Groenen PM, Peer PG, Wevers RA, et al. Maternal myo-inositol, glucose, and zinc status is associated with the risk of offspring with spina bifida. Am J Obstet Gynecol. 2003; Dec; 189 (6) 1713–9. DOI: 10.1016/s0002-9378(03)00807-x [PubMed: 14710103]
- 115. Wilson MP, Hugge C, Bielinska M, et al. Neural tube defects in mice with reduced levels of inositol 1,3,4-trisphosphate 5/6-kinase. Proc Natl Acad Sci U S A. 2009; Jun 16; 106 (24) 9831–5. DOI: 10.1073/pnas.0904172106 [PubMed: 19482943]
- 116. Guan Z, Wang J, Guo J, et al. The maternal ITPK1 gene polymorphism is associated with neural tube defects in a high-risk Chinese population. PLoS One. 2014; 9 (1) e86145 doi: 10.1371/ journal.pone.0086145 [PubMed: 24465924]
- 117. Cavalli P, Ronda E. Myoinositol: The Bridge (PONTI) to Reach a Healthy Pregnancy. Int J Endocrinol. 2017; 2017 5846286 doi: 10.1155/2017/5846286 [PubMed: 28243254]
- 118. Greene ND, Leung KY, Gay V, et al. Inositol for the prevention of neural tube defects: a pilot randomised controlled trial. Br J Nutr. 2016; Mar 28; 115 (6) 974–83. DOI: 10.1017/ S0007114515005322 [PubMed: 26847388]
- 119. Firth, HV, Hurst, JA. Clinical Genetics and Genomics (Oxford Desk Ref). 2 Edition. Oxford University Press; Oxford: 2017.
- 120. Glasziou P, Chalmers I, Rawlins M, et al. When are randomised trials unnecessary? Picking signal from noise. BMJ. 2007; Feb 17; 334 (7589) 349–51. DOI: 10.1136/bmj.39070.527986.68 [PubMed: 17303884]
- 121. Freedman B. Equipoise and the ethics of clinical research. N Engl J Med. 1987; Jul 16; 317 (3) 141–5. DOI: 10.1056/NEJM198707163170304 [PubMed: 3600702]

122. Agha-Jaffar R, Oliver N, Johnston D, et al. Gestational diabetes mellitus: does an effective prevention strategy exist? Nat Rev Endocrinol. 2016; Sep; 12 (9) 533–46. DOI: 10.1038/ nrendo.2016.88 [PubMed: 27339886]

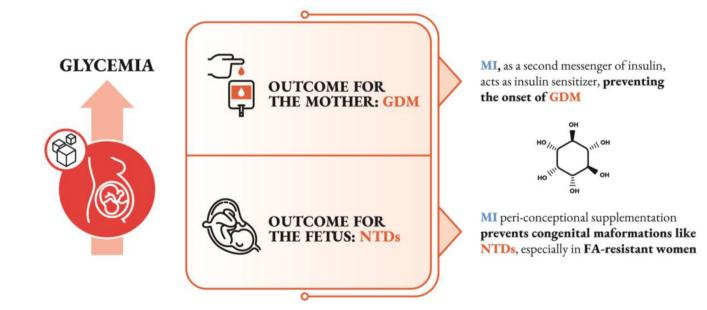


Figure 1. Schematic representation of the effectiveness of MI supplementation in preventing GDM and NTDs.

Table 1
Clinical trials investigating the effects of inositols supplementation in pregnant women.

Study	Population	Active Intervention	Main outcome	Delivery outcomes	Perinatal outcomes
Matarrelli et al 2013 [72]	Non-obese women with an elevated fasting glucose in the first or early second trimester N= 36 (treated) N= 39 (control)	МІ	Lower abnormal OGTT and insulin therapy	Lower rates of Polyhydramnios	Lower neonatal hypoglycemia, birth weight, fetal abdominal circumference, higher GA at delivery
D'Anna et al 2013 [73]	Outpatients with a parent with type 2 diabetes N= 110 each group	MI+FA	Lower GDM rate	No differences in CS and PTB rates	Significant differences between groups on macrosomia rate and birth weight
Malvasi et al 2014 [74]	Non-obese healthy women at 13th -24th week of pregnancy N= 24 each group	MI+DCI+FA+Mn <sup>2+</sup>	Improvement of glycemia and blood parameters (except diastolic blood pressure		
D'Anna et al 2015 [75]	Obese women at 12-13 weeks of gestation N= 110 each group	MI+FA	Lower GDM rate	No differences in CS and PTB rates	No differences in birth weight and macrosomia
Santamaria et al 2016 [76]	Overweight, non-obese women N= 110 each group	MI+FA	Lower GDM rate	No differences in CS and PTB rates	No differences in birth weight and macrosomia
Lubin et al 2016 [77]	Women with GDM uncontrolled by diet N= 32 (treated) N= 28 (control)	MI+FA	Lower need of insulin treatment	Lower labour induction	No differences in birth weight and macrosomia
Farren et al 2017 [78]	Women with a family history of diabetes N= 120 each group	MI+DCI+FA	No changes in GDM rate	No differences in CS and PTB rates	Higher incidence of neonatal hypoglycemia and lower neonatal jaundice
Fraticelli et al 2018 [79]	Women with GDM N= 20 each group	MI+FA DCI+FA MI+DCI+FA	Lower HOMA index and weight gain with MI Lower need for insulin therapy in MI and MI+DCI	No differences in CS, induction and PTB rates	Lower birth weight in inositol groups
Pintaudi et al 2018 [80]	Women with GDM N= 6 each group	MI+FA	Reduction of glucose variability	No differences in CS and PTB rates	No differences in birth weight and macrosomia
Celentano et al 2020 [65]	Non-obese women with elevated fasting glucose in the first or early second trimester N= 105 (treated) N= 52 (control)	MI DCI MI+DCI	Lower abnormal OGTT with MI	No differences in CS and PTB rates	Lower birth weight, fetal abdominal circumference, and higher GA at delivery with MI