Published in final edited form as: *Neuroimage*. 2021 February 15; 227: 117622. doi:10.1016/j.neuroimage.2020.117622.

CIVET-Macaque: An automated pipeline for MRI-based cortical surface generation and cortical thickness in macaques

Claude Lepage^a, Konrad Wagstyl^{a,b}, Benjamin Jung^{c,d}, Jakob Seidlitz^{e,f}, Caleb Sponheim^g, Leslie Ungerleider^c, Xindi Wang^a, Alan C. Evans^a, Adam Messinger^{c,*} ^aMontreal Neurological Institute (MNI), McGill University, Montreal, Canada

^bWellcome Centre for Human Neuroimaging, University College London, London, UK

^cLaboratory of Brain and Cognition, National Institute of Mental Health, Bethesda, MD, USA

^dDepartment of Neuroscience, Brown University, Providence, RI, USA

^eDepartment of Child and Adolescent Psychiatry and Behavioral Science, Children's Hospital of Philadelphia, Philadelphia, PA, USA

^fDepartment of Psychiatry, University of Pennsylvania, Philadelphia, PA, USA

^gDepartment of Organismal Biology and Anatomy, University of Chicago, Chicago, IL, USA

Abstract

The MNI CIVET pipeline for automated extraction of cortical surfaces and evaluation of cortical thickness from *in-vivo* human MRI has been extended for processing macaque brains. Processing is performed based on the NIMH Macaque Template (NMT), as the reference template, with the anatomical parcellation of the surface following the D99 and CHARM atlases. The modifications needed to adapt CIVET to the macaque brain are detailed. Results have been obtained using CIVET-macaque to process the anatomical scans of the 31 macaques used to generate the NMT and another 95 macaques from the PRIME-DE initiative. It is anticipated that the open usage of CIVET-macaque will promote collaborative efforts in data collection and processing, sharing, and automated analyses from which the non-human primate brain imaging field will advance.

Keywords

Surface-based morphometry; Surface registration; Rhesus monkey; Template; Atlas; Primate

*Corresponding author. messinga@mail.nih.gov (A. Messinger).

Declaration of Competing Interest

The authors report no competing interest.

This is an open access article under the CC BY-NC-ND license (https://creativecommons.org/licenses/by-nc-nd/4.0/)

Credit authorship contribution statement

Claude Lepage: Conceptualization, Software, Formal analysis, Writing - review & editing. Konrad Wagstyl: Conceptualization, Writing - review & editing. Benjamin Jung: Resources, Writing - review & editing, Software. Jakob Seidlitz: Conceptualization, Resources, Writing - review & editing. Caleb Sponheim: Conceptualization, Resources, Writing - review & editing. Leslie Ungerleider: Supervision, Funding acquisition. Xindi Wang: Software, Data curation. Alan C. Evans: Supervision, Funding acquisition. Adam Messinger: Conceptualization, Resources, Investigation, Writing - review & editing, Validation, Visualization, Project administration, Supervision.

1 Introduction

The cerebral cortex has a two-dimensional folded topology, tangentially subdivided into cortical areas and radially into layers. Human neuroimaging studies of cortical structure and function have been enriched through the development of automated surface reconstruction techniques (Fischl, 2012; Kim et al., 2005). These techniques have facilitated intersubject and intermodal coregistration, compression and analysis of large datasets, and helped reveal areal (Glasser et al., 2016) and laminar organization (Wagstyl et al., 2020) in a common surface-based coordinate system.

Cortical meshes are amenable to quantification of morphometry: per-vertex cortical thickness, curvature, and surface area can all be readily measured from mesh representations. The spherical topology of the modeled cortical surfaces allows for intersubject coregistration (Fischl et al., 1999; Lerch and Evans, 2005; Robinson et al., 2014). Surfaces can be aligned to a common average template for group-comparison corticometric studies and surface-based parcellations, such as atlases, can be mapped to individual surface meshes for regional analyses, with per-vertex correspondence between the white matter and pial surfaces. Furthermore, within this framework, coregistering other imaging modalities to the same space facilitates quantitative sampling of any number of *in vivo* modalities: fMRI, qMRI, PET, EEG, MEG (Bonaiuto et al., 2018; Greve et al., 2014; Weiskopf et al., 2013; Yeo et al., 2011), as well as *post mortem* histological (Wagstyl et al., 2020) and genetic data (Hawrylycz et al., 2012). Once sampled to the mesh, and coregistered to a common template, vertex-based representations allow for intermodal analysis and application of more complex analytical techniques.

Such techniques have been used to advance human neuroimaging, and have the potential to similarly advance NHP neuroimaging. For instance, in humans, surface analysis has been used to parcellate the cortex according to multimodal *in vivo* data from task and resting state fMRI and structural images in the same subjects (Glasser et al., 2016). Radial sampling between gray and white cortical surfaces across the cortex can produce depth-dependent intensity profiles that, with sufficient resolution and contrast, can be used to segment cortical layers (Wagstyl et al., 2020). Surface-based descriptions have also been used to characterize human semantic organization (Huth et al., 2016) and pathology (Lerch et al., 2005). In NHP studies, surfaces can be used to identify similar regions across species and facilitate interspecies comparison (Donahue et al., 2018; Miranda-Dominguez et al., 2014; Xu et al., 2019). Thus, cortical surface reconstruction techniques provide a powerful framework for neuroimaging that is complementary to volumetric analysis.

Several cortical surface reconstructions for macaques have been presented (Autio et al., 2020; Donahue et al., 2018; Koo et al., 2012; Oguz et al., 2015; Wang et al., 2018). In addition, a few new pipelines for macaque cortical surface generation (e.g. NHP-Freesurfer; Precon_all; PREEMACS, see (Garcia-Saldivar et al., 2020) have recently been shared on the PRIME-Resource Exchange (https://prime-re.github.io/) (Messinger et al., 2020). These efforts have made surface generation less labor intensive. A surface-free approach to evaluate cortical thickness, using the DiReCT volume registration-based method in ANTS (Tustison et al., 2014), has also been extended to the macaque (Calabrese et al., 2015).

However, most of the procedures described require both T1 and T2-weighted anatomical scans of sufficient quality, multiple software packages, various manual interventions, and several end-arounds to get human software tools to process data. Furthermore, these packages are limited to exemplary scans or small datasets, some without means for surface-based registration for group analyses of large cohorts. The work presented here addresses the need for an open-access dedicated pipeline for non-human primate cortical surface reconstruction, with capabilities comparable to human pipelines.

The CIVET pipeline (Lepage et al., 2017), which was developed at the Montreal Neurological Institute (MNI), is a comprehensive automated pipeline for corticometric and morphometric analyses of human MR images. The pipeline provides high resolution cortical surface reconstruction from T1-weighted MR images, featuring intersubject surface-based coregistration with per-vertex correspondence between gray and white matter surfaces. In this work, CIVET has been customized for the processing of non-human primate MR images. The present work covers the extension to the macaque monkey and is based on the NIMH Macaque Template (NMT) (Seidlitz et al., 2018).

The availability of a new automated pipeline for processing non-human primate anatomical scans opens new opportunities for analyzing large datasets with their wealth of structural, functional, diffusion and post mortem data. For example, the UNC-Wisconsin Rhesus Macaque Neurodevelopmental Database provides a rich dataset of longitudinal structural and diffusion MRI during early postnatal development in 34 rhesus macaques (Young et al., 2017). For post mortem data, the Allen Brain Institute provides gene expression and histological data throughout macaque developmental stages, with reference MRI (Bakken et al., 2016). Others have published post mortem diffusion tensor imaging datasets (Calabrese et al., 2015). Recent initiatives such as the PRIMatE Data Exchange (PRIME-DE) (Milham et al., 2018) have helped to harmonize these data, generating large scale neuroimaging datasets that require robust, automated analytical pipelines. Furthermore, adaptation of statistical map repositories like Neurovault (Gorgolewski et al., 2015) have made it easier to share standardized volumetric results in NHPs. Analysis of such data in terms of anatomical areas and morphometry would be enhanced by surface analysis capabilities (Calabrese et al., 2015). Even existing surface-based studies of NHPs may benefit from improved techniques and automation (Koo et al., 2012).

2. Material and methods

2.1 Subject information

This study uses anatomical MR scans from 3 sources: the *ex-vivo* scan of the single-subject D99 template (Reveley et al., 2017), 31 macaques from the Central Animal Facility at the National Institute of Mental Health (NIMH, USA), and *in-vivo* T1-weighted (T1w) anatomical scans of 95 macaques obtained from the INDI PRIME-DE consortium (Milham et al., 2018), http://fcon_1000.projects.nitrc.org/indi/indiPRIME.html. All animal procedures were conducted in compliance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals or, in the case of the PRIME-DE scans, with the animal care and use policies of the institution where the data were collected (Table 1).

Lepage et al.

The rhesus macaques (*Macaca mulatta*) scans from the NIMH, referred to hereafter as the NMT cohort, have been previously described (Seidlitz et al., 2018). The 25 male and 6 female animals were, at the time of the scans, juveniles and adults $(5.2 \pm 1.9 \text{ years})$ with an average weight of 6.3 ± 1.7 kg. The animals did not have surgery or procedures within the intracranial cavity prior to the scans. However, 14 subjects had a non-metallic post surgically implanted on top of their skull that made the dorsal portions of the skull appear thin or absent in some scans. T1w MR volumes were acquired using the modified driven equilibrium Fourier transform (MDEFT) sequence (Deichmann et al., 2004; Lee et al., 1995) on a 4.7T horizontal scanner (Bruker Biospec 47/40). On the day of scanning, the animals were placed under food restriction, anesthetized with isoflurane, and placed in the scanner in the sphinx position. Each whole-brain MDEFT scan was acquired over a 40–60 min session using either a 14 or 16.5 cm diameter single loop circular head coil (FOV = $96 \times 96 \times 70$ mm³; 0.5 mm isotropic voxels). For most subjects, multiple MDEFT scans were collected consecutively during the session (mean = 2.5 MDEFT scans/subject, range = 1-7). In these cases, subsequent MDEFT scans were linearly (6 parameter rigid-body) registered to the

The scans for the INDI PRIME-DE consortium consisted of *Macaca mulatta* (N = 90) and *Macaca fascicularis* (N = 5) monkeys and were collected at various sites with different scanner types and field strengths. For this study, we selected all T1w scans collected in the sphinx position from animals for which age and sex information was available (see Table 1).

first, and then averaged to create a single volume for each subject.

2.2 Overview

The CIVET pipeline for extraction of cortical surfaces from in-vivo MR images of the human brain has been extended for processing macaque brains, and as such the processing stages for macaques parallel those for the human. Extensions of CIVET to the macaque use the NIMH Macaque Template (NMT) as the standardized space for extracting and characterizing the surfaces of individual monkeys (Seidlitz et al., 2018). The macaque option for CIVET, herein referred to as CIVET-macaque, is invoked by the simple specification of a command line parameter for the choice of the model for the standardized space. CIVET takes as input a T1w scan and registers if to standardized space, herein the NMT (Seidlitz et al., 2018), using an affine transformation. The image is resampled into the NMT space, corrected for non-uniformities, masked, then classified into cerebrospinal fluid (CSF), cortical and subcortical gray matter (GM), and white matter (WM). A white matter surface, per hemisphere, is fitted at the GM-WM boundary using a marching-cubes algorithm, then adjusted to the maximum T1w intensity gradient position, before being expanded to the pial surface. Cortical thickness is calculated as the distance between the two cortical surfaces. Surface-based registration is performed and the cortical thickness maps are blurred and resampled at the common vertex locations of the template for group comparisons and analyses.

The original CIVET pipeline is based on the *minc* storage format developed at the MNI (Vincent et al., 2016) and this format has been maintained as the processing unit for the customization of CIVET-macaque. The *minc-2* format is based on the HDF5 library with automatic internal file compression for efficient data representation of large volumes

Lepage et al.

(The HDF Group, 2016). Cortical surfaces are represented using the *MNI-obj* format (http:// www.bic.mni.mcgill.ca/Services/HowToWorkWithObjectFiles). Many tools to manipulate and visualize volumes and surfaces are available in the openly available *minc toolkit* (http:// bic-mni.github.io/#MINC-Toolkit). Full compatibility exists to read input T1w scans in NIfTI format and to export the outputs in the commonly used NIfTI/GIfTI formats, enabling integration with a wide range of other analysis tools. The CIVET pipeline is available on the CBRAIN platform (Sherif et al., 2014) for collaborative studies and sources and binaries for Linux platforms are available via the CIVET project page (https://github.com/ aces/CIVET_Full_Project).

While the processing flow in CIVET is the same for humans and macaques, the main implementation issues in the extension of CIVET to macaques are the elaboration of species-specific volumetric and surface templates for registration as well as the scaling of all length-related parameters. Numerous modules of the pipeline have been modified to include a linear length scale parameter to account for the smaller brain size of the macaque compared to the human. A length scale of 0.4 was derived from the cubic root of the macaque-to-human volume ratio for the whole brain (0.43 using 92.8cc for the NMT vs. ~1200cc for the human; see section 3.1; (Seidlitz et al. 2018; Jung et al. 2020). Similar values were obtained using ratios of gray matter (0.41) or white matter volumes (0.37)(Donahue et al., 2018). Length scales appear in image blurring (full width at half maximum, fwhm), registration (search distances), masking (search radius), surface blurring of cortical thickness (fwhm), morphological operations such as voxel erosion and dilation (distance), non-uniformity corrections (spline distance), and voxel size representation proportional to brain structures. Other algorithms such as tissue classification (histogram-based) and cortical surface extraction (topology based) are unaffected. These changes are summarized in Table 2. Further adaptations of the pipeline to other non-human primate brains should be straightforward by introducing the appropriate length scales and supplying species specific templates.

2.3 Volumetric template

The first step in the extension of CIVET is the selection of a volumetric template for stereotaxic registration and the preparation of its associated atlases. The NIMH Macaque Template (NMT, https://afni.nimh.nih.gov/NMT) version 1.2 (asymmetric model), built from the non-linear average of the 31 scans from the NMT cohort, was selected as this registration target (Seidlitz et al., 2018). At the inception of this project, the NMT was the template based on the largest number of scans, with complete head coverage, and with individual scans being available. It is noted, while it is not used in CIVET, that the NMT has been recently updated and now includes a symmetric macaque template in Horsley-Clarke stereotaxic coordinates (Horsley and Clarke, 1908; Saleem and Logothetis, 2012).

The NMT (version 1.2) was created by iterative symmetric diffeomorphic registration methods using the ANTs software package (Avants et al., 2010). First, whole-head single subject images were initially aligned to a standard coordinate space (D99-SL from (Reveley et al., 2017)), and resampled from 0.5 to 0.25 mm isotropic resolution. Then, following a voxelwise N4 bias field correction (Avants et al., 2011) applied on each subject's aligned

image, a spatially-unbiased population average was created using the symmetric group-wise normalization (SyGN) (Avants et al., 2010; Love et al., 2016). The resultant NMT was placed in the neurological orientation, rotated so that, like the D99 atlas, the horizontal plane contained both the superior extent of the anterior commissure and the inferior extent of the posterior commissure (Horsley and Clarke, 1908; Saleem and Logothetis, 2012), and translated to place its origin (AP 0, SI 0, ML 0) on the midline at the center of the anterior commissure. Intensity values in the NMT were capped to account for the extreme values of the blood vessels, and a final N4 bias field correction was performed.

Atlases on the NMT (version 1.2) are created for its use in CIVET. The template is segmented to label the cerebellum, brainstem, ventricles, amygdala, hippocampus, and subcortical region. This segmentation is used in CIVET to exclude the brainstem and cerebellum during tissue classification and to mask out the ventricles and subcortical gray areas prior to the extraction of the white matter surface. A brain mask of the template is obtained using a tailored version of FSL *bet* for macaques (Smith, 2002). Finally, tissue priors are defined on the NMT (500 + points for each of CSF, GM, and WM). Priors are also defined in the background (BG) outside the skull. These priors are later coregistered to each subject and used for training the tissue classifier. The preparation steps of the template are summarized in Fig. 1.

2.4 Registration to standardized template space

Registration to the NMT uses *minctracc* (Collins et al., 1994). Linear registration follows a hierarchical multi-scale approach from coarse to fine image blurring and sampling rates. For the macaque, all length scales in the registration procedure are scaled by 0.4 to account for brain size relative to that of the human. That is, the *fwhm* for image blurring and sampling intervals are scaled proportionally by 0.4. With this scaling, the registration procedure for the human brain achieves optimal performance on the macaque brain. The length scales in the non-linear registration procedure are adjusted in the same way.

Linear and non-linear registration to the macaque template requires the use of a brain mask since head size varies significantly between males and females, unlike humans, resulting in variable scan field-of-views. For instance, the male macaque typically has larger temporalis muscles than the female, resulting in thicker "scalp" (muscle) and head size. In some cases, brains have been pre-processed and de-skulled. Registration, using *minctracc*, proceeds by specifying the brain mask on the target image only, herein the NMT volume with its associated brain mask, and evaluating the source image, herein the subject's scan, at the sampling points within the target mask, without the need to have a brain mask explicitly defined on the subject. This registration approach has proven to be robust in overcoming the variability in scans due to head size and field-of-view. In the rare case when registration fails, a custom brain mask in native space can be supplied. This custom brain mask can be obtained from automated tools within the CIVET environment or by other external tools. The accuracy of such custom masks should be assessed by the user prior to processing.

The macaque brain is resampled in linear standardized space at the processing voxel resolution of 0.25 mm or 0.50 mm, which is of comparable resolution relative to the human brain (processing resolution at 0.50 mm or 1.0 mm).

2.5 Non-uniformity correction

The image, resampled into standardized space, is corrected for non-uniformities using N3 bias field correction (Sled et al., 1998). The sampling region for N3 is restricted by the brain mask of the volumetric template, as it is for humans, since a reliable custom mask cannot be obtained prior to N3 correction. The parametric spline distance must be entered by the user. This value can be determined empirically such that the intensities for gray matter and white matter appear mostly uniform across the brain. This distance varies with the field strength of the scanner and, to some extent, the species' head size. Finally, denoising is performed on the input image to smooth out voxels with local maxima of intensity (100 iterations), thereby avoiding the inclusion of blood vessels in the calculation of the bias field.

2.6 Brain-masking

Brain-masking is based on extensions of FSL bet version 1 (Smith, 2002) and is applied in standardized space. The original version of this software blindly sampled intensities over the full head to determine the masking thresholds. Given the differences in total head size and scalp musculature between male and female macaques, it was found critical to exclude the scalp when calculating the histogram of intensities used to establish the masking thresholds. To discount the macaque scalp and skull, the intensities were sampled within the brain mask of the volumetric template. Although the brain mask of the volumetric template is an approximation of the scan's true brain mask, this approach provides accurate thresholds for reliable brain masking. Brain extraction a la FSL bet thus provides a refinement of the template's brain mask. For some scanning protocols, the blood vessels have very large signal intensities that interfere with convergence and result in an incomplete mask. To prevent this, denoising is performed on the image to smooth out voxels with local maxima of intensity (100 iterations). Moreover, the brain-masking procedure can incorporate information from complementary MR scans, such as a T2-weighted image, for improving the T1w brain mask. The procedure proved to be very robust at extracting accurate masks on scans of individual subjects (see Fig. 2).

2.7 Tissue classification

The tissue classification procedure is identical for both human and macaque scans and relies on the T1w scan, with the possibility of a multimodal classification should T2-weighted and/or proton density scans be available. The brain is classified into cerebrospinal fluid, gray matter, and white matter using an artificial neural network (ANN) trained on the intensities of the tissue priors (BG, CSF, GM, WM) mapped non-linearly from the NMT volume to the subject (Zijdenbos et al., 1998). The subcortical region labelled on the NMT allows for the discrimination of cortical from non-cortical gray matter in the specific region. Partial volume estimates are obtained for the tissue classes (Tohka et al., 2004). Partial volumes for CSF are combined with curvatures (eigenvalues of the Hessian matrix of intensities) in the reconstruction of the CSF skeleton delineating buried sulci which ensures full sulcal penetration of the pial surface during its creation.

2.8 Masking blood vessels

Prior to the extraction of the white matter surface, it is critical to identify T1w hyperintensities, commonly representing major blood vessels and erroneously segmented as white matter by the classifier, to prevent these spurious white matter voxels from interfering with the process of extracting the white matter surface. The basic assumptions in the detection algorithm are that blood vessels are represented by T1w hyperintensities with a line morphology propagating in CSF within sulci. Sulcal lines are identified using a watershed algorithm in non-white voxels, then dilated partly through gray matter and, consequently, through blood vessels, but not white matter. The trace of the Hessian matrix of second derivatives (i.e., the sum of the Hessian's three eigenvalues) is used as an approximation of the local curvatures to detect one-dimensional line features. Thresholds for the expected signal intensity in white matter and for the trace of the Hessian are used to discriminate lines structures in sulci from blood vessels. Supplementary Figure 1 illustrates blood vessels detected on a single scan.

2.9 Cortical surface extraction

A white matter mask is created from the tissue classification, masked for the brain. Using the segmented regional masks on the NMT volume non-linearly transformed to the subject, the ventricles and subcortical gray matter are filled in as white matter, the hippocampus and the amygdala are masked out, and the brainstem and cerebellum are excluded for the purpose of extracting the white matter surface. The brain, in NMT space, is then split into the left and right hemispheres, at the mid-plane wall, and the white matter surfaces are extracted using a marching-cubes algorithm (Lepage et al., 2017). Spherical topology of the cortical mantle is preserved by advancing the surface, in voxel space, outside-in from a sphere enclosing the hemisphere, hence the requirement for masking off the blood vessels obstructing the progression of the surface. Mesh adaptation operations (vertex smoothing, edge collapsing and swapping) are used to efficiently reduce the surface mesh to a manageable size (around 140,000 vertices) for further processing. The decimated white matter surface is inflated to a sphere and resampled at a fixed number of vertices on a canonical sphere obtained by refinement of the icosahedron (40,962 vertices). In the case of a right-oriented surface, the icosahedral sphere is mirrored in the x-axis, thus establishing a direct correspondence in how vertices are numbered between mirror-symmetric positions in the two hemispheres. This correspondence facilitates comparison between like positions in the two hemispheres, without altering the morphology of either hemisphere. The resampled surface is then registered to the average white matter surface template (see subsection below) for mapping the surface region on which the intensity gradient correction is disabled, either because it is meaningless (cut through the corpus callosum and brainstem on the medial wall) or misleading (e.g., cavity of the hippocampus and amygdala, insula region with low gray-white contrast in proximity of the claustrum). The white matter surface is then adjusted to the maximum intensity gradient (magnitude of first derivative of T1w intensity normal to the surface).

The pial surface is obtained by expanding the white matter surface radially outwards up to the pial boundary using the CLASP algorithm (Kim et al., 2005). Consequently, a per-vertex

correspondence exists linking the vertices of the white matter and pial surfaces, from which the mid-cortical surface is obtained by averaging the position of the linked vertices.

No special adjustments were made for macaque surface extraction except for scaling (by 0.4) the search distance for the gradient correction and the *fwhm* for blurring volumes. The surfaces can also be extracted at high-resolution (163,842 vertices), but such resolution is excessive given the morphology of the macaque brain.

2.10 Surface-based registration

Surface-based registration is used to find the correspondence between the cortical surface of an individual subject to a surface template (Robbins, 2003). The registration operates on a spherical representation of the surfaces and the mesh alignment is driven by a *depth potential* term, akin to sulcal depth (Boucher et al., 2009). Thus sulci are aligned to sulci and gyri are aligned to gyri. Following registration, any map defined on the subject's surface can be interpolated at the sampling vertices of the reference surface template. Resampling of surface maps, for instance cortical thickness, enables vertex-wise group comparisons across subjects of a population. Furthermore, regional analyses can be performed based on parcellations defined on the template. The generalization from human to macaque required accounting for head size by scaling the fwhm for blurring the sulcal depth term on the surface.

Surface-based registration uses a white matter surface template for the gradient correction adjustment of the white matter surface and a mid-cortical surface template for resampling surface maps such as cortical thickness, curvature, etc. The next subsections describe the creation of these average white and mid-cortical surface templates as the targets for surface-based registration and the application of the D99 and CHARM parcellation on these surface templates.

2.11 Average surface template

The generation of a CIVET-specific average surface was motivated by: 1) having a population average surface based on the same subjects used to create the population average volumetric template; 2) requiring having both corresponding white matter and mid-cortical average surfaces for surface-based registration; and, 3) establishing, within the CIVET suite, a common framework for translational studies involving other primate species, including the human. It is noted that surface-based registration can be used to map surface fields (e.g. cortical thickness, curvature) and atlases from the CIVET average surface to other surface templates, such as the Yerkes19 average surface (Donahue et al., 2018).

The average surface template, generated from the NMT dataset (N = 29, 2 subjects were rejected – one with a lesion, the second one due to abnormal ventricles), is the target for surface-based registration. Like for the human brain, the average surface is constructed by averaging the surfaces of the individual subjects, as opposed to using the cortical surfaces extracted on the average volume of all subjects. But, unlike the human brain, the average surface is generated in the non-linear standardized space, since subject-to-subject sulcal variability is low for the macaque. That is, the non-linear transformation from the subject's volume to the NMT template is applied to the subject's surfaces before averaging them, thus

reducing geometrical variance. This low-frequency non-linear transformation essentially corrects for gross distortion, like global twist, while retaining micro shape differences describing biological variability (see Figure 3). Non-linear warping was also applied in the construction of the NMT volume (Seidlitz et al., 2018).

The creation of the average surface is cyclic: CIVET requires the existence of an average surface as a template for surface-based registration, but surfaces must exist prior to averaging them. An iterative process is followed by first averaging unregistered surfaces and then using this average as the target for registration on the next iteration, and so on. The initial average is made possible since all surfaces have the same number of vertices, via sphere-to-sphere interpolation of the inflated surface to a canonical sphere, and that coordinates are aligned in stereotaxic space (Lyttelton et al., 2007).

The generation of the average white matter surface using the marching-cubes algorithm is computationally intensive since the algorithm requires repeated processing steps at each averaging iteration, namely, extraction, mesh decimation, inflation to sphere, resampling, and surface-based registration in order to apply the mask for the gradient correction. This mask must be defined on an intermediate average surface used as the registration target, which itself evolves as the average surface is iteratively converged. As such, the convergence of the individual white matter surfaces depends on the average surface at each iteration and these white matter surfaces must therefore be recomputed in full at each iteration, making the overall process lengthy. To accelerate the convergence process and avoid having to repeatedly manually paint the gradient mask on transient average surfaces, the intensity gradient correction is disabled for the first 5 iterations of the creation of the average white matter surface. The gradient mask was manually defined once on this intermediate average white matter surface and then frozen through 5 more global iterations of surface averaging, with the intensity gradient correction enabled. Full convergence is rapidly achieved - after the first five iterations, without the gradient correction, and through the last five iterations, with the gradient correction (see Fig. 4).

The creation of the average mid-cortical surface is much simplified since the extraction of the cortical surfaces does not need to be repeated at each iteration. The mid-cortical surfaces are initially registered to the average white matter surface to align the vertices between the two surfaces. Ten global iterations are used in the process, but convergence is achieved after only a few iterations (see Fig. 4).

For the symmetric averages, the right surfaces are mirrored as left surfaces before being averaged. For both white matter and mid-cortical average surfaces, the vertices are equidistributed on the surface to obtain a surface with nearly equal-size triangles, with the goal to have uniform sampling over the entire cortical sheet (see Fig. 4). The area equalization process is performed at the end of each averaging iteration and consists of moving each vertex to the centroid of its neighboring vertices, weighted by the area of their connected triangles.

The average white matter surface is shown in Fig. 5. The variability in shape (mean vertex displacement) across the N = 29 subjects is highest in the parieto-occipital area

and the medial and ventral occipital lobe. The average mid-cortical surface exhibits the same properties as the average white matter surface.

2.12 Cortical morphometry

The cortical surfaces, extracted in standardized space, are transformed back to native space and cortical thickness is evaluated as the distance between the white matter and pial surfaces. Three cortical thickness metrics are available in CIVET:

- 1 *tlink*: distance between linked vertices between the white matter and pial surfaces (sensitive to mesh distortion during expansion of the pial surface, which gives rise to an overestimation of cortical thickness);
- 2 *tlaplace*: distance along radial lines defined by a Laplacian field between the white matter and pial surfaces (not sensitive to mesh distortion) (Lerch and Evans, 2005);
- 3 *tfs*: symmetric average of projected distance from white matter-to-pial and pial-to-white matter surfaces (like FreeSurfer).

Other metrics are also produced such as vertex-wise surface areas and volumes, and mean and Gaussian curvatures. These measurements are blurred isotropically on the mid-cortical surface in native space (Boucher et al., 2009). Default values for surface blurring (*fwhm*) are proportionally less for the macaque than for the human (e.g. fwhm of 8mm). All measurements are resampled on the average NMT symmetric surface template as a basis for group comparison analyses.

2.13 CHARM surface atlas

The D99 subject was processed with CIVET-macaque to obtain the D99 surface parcellation (Reveley et al., 2017). This post mortem brain scan has no skull, a T1w-like contrast, and has been pre-processed: non-uniformities have been corrected and the scan has been symmetrized and transformed to a standardized space. A value of 0 was used as the spline parametric distance to disable N3 and the blood-vessel algorithm was disabled as well (because the scan was acquired ex vivo). Surface-based registration was used to register the D99 mid-cortical surfaces to the NMT symmetric surface template to ensure a direct vertexto-vertex correspondence. The resampled D99 mid-cortical surfaces were then intersected with the D99 volumetric segmentation (Reveley et al., 2017) to obtain parcellation labels, which mapped directly to the NMT symmetric surface template (see Fig. 6). A few minor morphological operations on the extracted labels were necessary to enforce symmetry and to correct for mis-labeled and unassigned vertices arising when intersecting the surfaces with the labelled volume. Using the final D99 labels, each vertex was assigned 6 additional anatomical labels; one for each level of the Cortical Hierarchical Atlas of the Rhesus Macaque (CHARM) (Jung et al., 2020). The 6 anatomical levels of CHARM range from a lobular parcellation at the coarsest level (4 ROIs) to nearly the full set of D99 cortical areas at the finest level (139 ROIs). Fig. 7 illustrates the 6 parcellation levels of CHARM. The CHARM parcellation can be used for regional analyses of cortical morphometry at multiple spatial scales.

2.14 Quality control

CIVET produces a series of images to monitor the main processing stages of each subject. These images provide an efficient means for visual inspection of volumetric registration, tissue classification, surface extraction and convergence, surface-based registration, and cortical thickness. Moreover, several metrics are extracted (e.g. brain volume, mean cortical thickness, tissue percentages, etc.) and tabulated into a global csv file for detection of outliers within the population. Failed runs can be systematically identified at this level of quality control.

3 Results

3.1 CIVET preprocessing

The reliability of CIVET-macaque pipeline's volumetric registration and brain-masking was tested on the T1w scans of the 31 macaques in the NMT cohort (Seidlitz et al., 2018) and 95 additional T1w scans of macaques obtained from the INDI PRIME-DE consortium (Milham et al., 2018). CIVET was run with the *without-surface* option, terminating after the stereotaxic registration, brain masking, and tissue classification stages. After some exploration, a value of 50 was used as the common spline distance for N3 for all PRIME-DE scans, regardless of the field strength of the scanner, and a value of 75 was used for the NMT scans. A head height of 50 mm, measured from the top of the head downwards, was used to crop the image to remove the neck for the purpose of registration. A voxel resolution of 0.25 mm was chosen for resampling in standardized space.

Upon visual quality control, all brains were accurately registered to the NMT, without exception. The brain masking stage was successful for all subjects, except those from Site 10 for which the mask leaked into the scalp. For those 3 scans, the bone marrow inside the skull revealed signal intensities similar to GM and WM and was fused with cortex, without a visible inner bone border, causing the mask leakage. (The use of the T2w scan to correct the mask was preempted since the temporal lobes were truncated by the scanner field of view.) In a few other cases, from other sites, the mask touched the edge of the orbits or the bottom half of the cerebellum was truncated from the field of view of the native scan, but such small defects do not have any impact on cortical surface extraction. Fig. 8 reports the volume of the brain masks in standardized space. Agreement of the individual brain mask volumes with that of the NMT (red line) confirms successful registration to the NMT and brain extraction. The few outliers from site 10 are clearly visible in Fig. 8. The mean stereotaxic brain mask volume for the PRIME-DE subjects, excluding the 3 outliers from site 10, is 92.9 ± 3.0 cc, in agreement with the volume of the NMT brain mask of 92.8cc. Fig. 9 shows the scan acquisition voxel size within and across sites. Scans for which all voxel dimensions were large (sites 12-14) generally had larger brain mask volumes and greater inter-subject variability. These failure rates are comparable to those observed in studies of human scans.

3.2 Cortical surface generation

Cortical surfaces were obtained for 29 of the 31 subjects in the NMT cohort (same two exclusions as for the creation of the average surface templates). The surfaces were produced

Lepage et al.

at 40,962 vertices per hemisphere. The *tlaplace* metric was used for the calculation of cortical thickness, blurred isotropically at a fwhm of 8 mm. The disk usage per processed subject is about 325Mb (for short input data type scans) to 550Mb (for float input data type scans). (The T1w image input data type was preserved during processing.) Total processing time ranged from 15–18 hours per subject, which was comparable to equivalent surface-based techniques for human processing. (For pre-processing only, without surface extraction, the processing time is about 1.5 h per subject.)

Fig. 10 shows an overview of the CIVET stages and results for 4 NMT subjects. The results display the same level of accuracy for all subjects. Surface contours superimposed on the volume (top right panel for each subject) generally showed faithful correspondence to the gray and white borders. For some NMT subjects, the gray matter surface did not fully extend to the CSF on the orbital frontal cortex (OFC) surface, especially around the gyrus rectus (not shown), leading to artifactually smaller OFC thickness in these cases. These errors are driven by segmentation errors due to scanning artifacts present for some of the NMT subjects and may not be an issue for other datasets. In many cases, the WM surface along the insula was slightly internal to the WM/GM border, lying along the lateral surface of the putamen, especially dorsally where the claustrum is very thin (see Fig. 10). These shifts in the position of the WM surface are caused by a relaxation of the WM surface intensity gradient correction adjacent to the claustrum. Cortical thickness values for the insula are likely slightly inflated in these cases. The WM surface in this region was, however, smooth and did not exhibit the 'claustrum invagination problem' (Autio et al., 2020). Nor were there issues with segmentation in such problem areas as the ventral and anterior claustrum or the temporal stem. Additionally, surface generation was accurate in scans with head post implants, as demonstrated in the right panels of Fig. 10.

Sulcal variability across the 4 subjects can be observed on both the WM and pial surfaces (Fig. 10, bottom row). Cortical thickness also exhibited some variability across subjects. The 3 subjects shown in (Koo et al., 2012) show similar sulcal and thickness variability. In general, thickness increased from posterior to anterior.

3.2.1 Cortical thickness—The average cortical thickness map across the 29 subjects and its coefficient of variation (COV=standard deviation/mean) are shown in Fig. 11. The cortical thickness map patterns follow closely those obtained from human MRIs processed with CIVET, with cortical thickness increasing from posterior to anterior, as well as published macaque cortical thickness maps (Autio et al., 2020; Calabrese et al., 2015; Donahue et al., 2018; Koo et al., 2012). The COV was low in most regions. Upon thorough examination of the cortical surfaces, the higher COV in some regions likely reflects a mixture of intersubject variability and limitations in the surface extraction process, namely the frontal pole (scanning artifacts due to nasal cavity), the temporal pole (presence of the middle cerebral artery), and the occipital lobe (lower gray-white contrast, owing to the high myelin content of this region (Donahue et al., 2018)).

3.2.2 Regional analysis—Regional cortical thicknesses based on the CHARM atlas are reported in Table 3. Thickness is reported for hierarchical atlas levels 1, 2, and 3 (with 4,17, and 36 regions, respectively) in the left and right hemispheres. Regional thickness was

highly symmetric between the hemispheres. Mean cortical thickness, within the brain and averaged across the 29 subjects, was 1.99 ± 0.43 mm. The frontal lobe was thickest and most varied in its thickness across vertices $(2.33 \pm 0.43 \text{ mm})$, followed by the temporal $(2.12 \pm 0.41 \text{ mm})$, parietal $(1.93 \pm 0.31 \text{ mm})$, and occipital lobes $(1.67 \pm 0.25 \text{ mm})$. At level 2, particularly thick regions include the motor cortex $(2.62 \pm 0.36 \text{ mm})$, including primary motor cortex, premotor cortex, and the dorsal aspect of the cingulate gyrus, and the temporal pole $(2.49 \pm 0.49 \text{ mm})$. Particularly thin regions include the somatosensory cortex $(1.89 \pm 0.30 \text{ mm})$, the core and belt regions of the auditory cortex $(1.79 \pm 0.36 \text{ mm})$, and primary visual cortex $(1.66 \pm 0.22 \text{ mm})$, especially within the parieto-occipital sulcus and on the ventral surface in the inferior occipital sulcus. Fig. 12 shows the histogram for the distribution of cortical thickness for several individuals and for the averaged cortical thickness across all 29 subjects (without blurring).

3.2.3 Surface area—The total area of the white matter and pial surfaces is displayed in Fig. 13 for the NMT subjects (N = 29). This total area includes the mid-plane wall cutting through the corpus callosum and the brainstem. Despite the inter-subject variability, areas of the left and right hemispheres are in close agreement within each subject for both cortical surfaces. The mean area per hemisphere was 10,098 mm² for the white matter surface and 13,441 mm² for the pial surface. These values are larger than those reported for the macaque (Autio et al., 2020; Donahue et al., 2018; Koo et al., 2012), even when accounting for the mid-plane wall region. The larger values obtained here could be an indication of deeper sulcal penetration of the pial surface and increased morphological details captured by the extracted surfaces.

4. Discussion

We have introduced CIVET-macaque, a pipeline for automated reconstruction of cortical surfaces of macaque brains. The pipeline requires only a T1-weighted MRI image to generate high resolution cortical surfaces, capable of intersubject surface-based coregistration with per-vertex correspondence between gray and white matter surfaces. Pre-processing and surface reconstruction were customized for macaques to address species related issues such as variations in head size and brain morphology.

The pipeline has been developed primarily on 29 scans out of the 31 subjects in the NMT cohort. Further tests were conducted on a larger heterogeneous collection of 95 scans from the INDI PRIME-DE consortium. The success rate depended, inevitably, on the quality of the imaging data. Input voxel resolution, field inhomogeneities, the presence of visible blood vessels, and scanning artifacts can all potentially cause various defects in the cortical surfaces. Supplementary Figure 2 shows the typical variability amongst scans and some common artifacts and their repercussion on the outcome of the pipeline. Some scans have a desirable high GM-WM contrast, but suffer from scanning artifacts anterior to the frontal and temporal poles. These artifacts interfered with surface placement, in particular for the NMT cohort, causing the COV for cortical thickness in these regions to peak. Some other scans were mostly free of scanning artifacts, but had low GM-WM contrast, which proved problematic during the extraction of the white matter surface (intensity gradient correction). Still other scans were of acceptable quality and produced expected results. It is hoped that

scanning protocols better suited to corticometry analyses will be employed in the future for additional structural data acquisition in monkeys.

Overall, the pre-processing phases of CIVET-macaque proved to be highly reliable and robust with regard to registration and brain-masking, independently of the quality of the scans. Cortical surface extraction was also successful although more challenging, being very sensitive to the quality of the data and, in particular, to the presence of high intensities representing blood vessels and other acquisition artifacts. The availability of a T2w scan could be used, in the future, to improve the detection of such blood vessels. Finally, sufficient scan resolution (0.5 mm voxel size or smaller) is necessary to delineate the fine structures of the macaque brain morphology, unlike the human brain for which a scan resolution at 1 mm voxel size has been employed for many years.

At present, to circumvent possible shortcomings in the white matter surface extraction, a brain mask with special annotations can be supplied with the input T1w scan, in native space. This initial custom brain mask is obtained from a first run of the pipeline and annotated in problematic areas where errors occurred during the white matter surface extraction. Label=1 represents the underlying brain mask. Label=2 represents offending misclassified voxels that should not be white matter (e.g. blood vessels, meninges/dura, gray matter with high signal intensities). Label=3 represents missing white matter voxels (e.g. reconstructed thin parahippocampal gyrus or incompletely filled ventricles for the purpose of white matter surface extraction). Basically, the annotated mask allows construction of a valid white matter mask for the white matter surface extraction by retaining classified WM within the cerebrum (label = 1), while removing voxels incorrectly labeled as WM (label = 2), and adding missing WM voxels to preserve the spherical topology of the cortical mantle (label = 3). Hollow ventricles can also be closed completely (label = 3) to avoid leakage of the white matter surface into the ventricles. CIVET is restarted with the new annotated custom mask and new surfaces are obtained.

All 31 NMT T1w scans revealed high-intensity voxels representing blood vessels or other scanning artifacts causing various white matter surface extraction errors of different degrees. An annotated mask, in native space, was thus needed for each of the 29 NMT scans included in this study. The task was fairly quick (at most 1h per subject) and amenable for this small dataset. After the masks were annotated, the pipeline was restarted in a fully automated way. The ensuing white matter surfaces were of high quality, as specifically needed for the generation of the average white matter and mid-cortical surfaces with minimal variability due to algorithmic noise.

The generated maps of cortical thickness for the NMT cohort (Fig. 11 and Table 3) revealed expected morphological trends. These include thinner somatosensory, visual and auditory areas, thicker temporal pole and motor cortex, with the remaining homotypic isocortex exhibiting thicknesses in between these extremes. This pattern is consistent with existing studies of macaque MRI morphology (Wagstyl et al., 2015) and similar to patterns seen in human histological studies of cortical thickness in the human BigBrain (Amunts et al., 2013; Wagstyl et al., 2020). Direct *post mortem* histological validation of macaque cortical thickness requires the creation of an equivalent 3D histological atlas of the macaque brain.

CIVET-macaque is an essential tool in carrying out automated morphological analysis of these atlases and facilitates comparisons between *in vivo* MRI and histology.

The thickness map in Fig. 11 is qualitatively similar to that of published maps of individual subjects (see Koo et al., 2012; Oguz et al., 2015, Fig. 4 of Autio et al., 2020) and group averages (see Fig. 4 of Autio et al., 2020, Fig. 3 of Koo et al., 2012, Fig. S2 of Seidlitz et al., 2018, n = 31, in particular that in Fig. 6 of Calabrese et al., 2015, n = 10, and in Fig. S4 of Donahue et al., 2018, n = 19). The mean cortical thickness (*tlaplace*, fwhm = 8 mm) reported here is 1.99 ± 0.17 mm. The mean thickness of the NMT cohort agrees with that reported by Donahue et al., 2018 ($2.0 \pm 0.1 \text{ mm}$, n = 19) and Autio et al., 2020, ($2.1 \pm 0.1 \text{ mm}$, n = 4), but is smaller than the value reported by Koo et al., 2012, $(2.37 \pm 0.19 \text{ mm}, \text{n} = 18)$. The pipeline employed by Koo et al., 2012 is a variant from an older version of CIVET. Their surfaces included on the order of 5000 vertices and the *tlink* metric was used for cortical thickness, which leads to an overestimation due to mesh distortion during the expansion of the pial surface. Note that the standard deviation, here, is the vertex-wise standard deviation across the n subjects, relative to the local vertex-wise mean of cortical thickness for the n subjects, averaged across the number of vertices on the surface (by hemisphere). It reflects how much cortical thickness varies locally - at the vertex level - across the subjects, relative to the local mean.

The variance in cortical thickness across the NMT cohort was low in most places. Greater COV (see Fig. 11) in the frontal pole, superior part of the occipital lobe and temporal pole appears in part to be driven by poor scan quality and algorithmic shortcomings. The frontal pole suffers from scanning artifacts due to the nasal cavity. In those subjects with the thinnest occipital lobe, the gray-white contrast was poor in that region, causing the true gray-white intensity transition to be missed and the white matter surface to be displaced towards the pial boundary. Finally, blood vessels (namely, the middle cerebral artery) in the vicinity of the temporal pole appear to be the source of the defects in both the white and gray matter surfaces in that region. Overall, the regions of high COV in this cohort are conjectured to be driven by processing errors and to be a weak indicator of true biological variance. Caution should be used when reporting results of biological relevance in these regions.

5 Conclusions

CIVET-macaque is the first true fully automated pipeline for processing macaque brains for corticometric studies. The pipeline, based on the imaging minc data format, has been developed in PERL and C/C++ for Linux systems. The open-source project (https://github.com/aces/CIVET_Full_Project) is also found on the PRIME-Resource Exchange portal (https://prime-re.github.io/). Inputs can be in either minc (HDF5-based) or NIfTI format and outputs are provided in minc (HDF5-based) format and, optionally, in NIfTI/ GIfTI formats.

This new software enables the field of NHP imaging to further explore questions of structure and function across large populations and datasets in a new way. It is anticipated that the open usage of CIVET-macaque will promote collaborative efforts in data collection,

sharing, and automated analyses from which the non-human primate brain-image field will further advance (Autio et al., 2020). For example, corticometry would provide a natural and complementary insight on longitudinal developmental studies in the macaque monkey (Scott et al., 2016; Wang et al., 2018), as well as facilitate cross-modal integration of *in vivo* and *post mortem* data. Finally, it is our intent to share complete CIVET outputs for the INDI PRIME-DE on the PRIME-RE portal (https://prime-re.github.io/) (Messinger et al., 2020).

CIVET has the potential to be expanded to other non-human primate species, with possible extensions to the marmoset and the chimpanzee, using repositories of previously collected and, if possible, publically available anatomical scans. The ability to process marmoset, macaque, chimpanzee, and human data with CIVET will allow for comparative studies of surface-based morphometry.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

This research was supported (in part) by the Intramural Research Program of the NIMH and includes the relevant Annual Report number in the following format (ZICMH002899). Parts of this work have received support from Healthy Brains for Healthy Lives. KW is supported by the Wellcome Trust (215901/Z/19/Z). The authors report no conflicts of interest.

Data accessibility and availability

Scans used for the development of CIVET-macaque are already publicly accessible. These include the D99 *post mortem* brain and atlas (https://afni.nimh.nih.gov/Macaque), the NMT volume (version 1.2; https://afni.nimh.nih.gov/NMT), and single subject scans from the INDI PRIME-DE repository (http://fcon_1000.projects.nitrc.org/indi/indiPRIME.html/). CIVET-macaque will be versioned and added to the global open-source CIVET project (https://github.com/acces/CIVET_Full_Project). Binaries for common Linux platforms will be provided for convenience. CIVET-macaque, the NMT, the CHARM (Jung et al., 2020), and updates to these resources can be centrally accessed from the PRIME-Resource Exchange portal (https://prime-re.github.io/) (Messinger et al., 2020).

References

- Amunts K, Lepage C, Borgeat L, Mohlberg H, Dickscheid T, Rousseau M-É, Bludau S, Bazin P-L, Lewis LB, Oros-Peusquens A-M, Shah NJ, et al. BigBrain: an ultrahigh-resolution 3D human brain model. Science. 2013; 340: 1472–1475. [PubMed: 23788795]
- Autio JA, Glasser MF, Ose T, Donahue CJ, Bastiani M, Ohno M, Kawabata Y, Urushibata Y, Murata K, Nishigori K, Yamaguchi M, et al. Towards HCP-Style Macaque Connectomes: 24-Channel 3T Multi-Array Coil, MRI sequences and preprocessing. Neuroimage. 2020. 116800 [PubMed: 32276072]
- Avants BB, Tustison NJ, Song G, Cook PA, Klein A, Gee JC. A reproducible evaluation of ANTs similarity metric performance in brain image registration. Neuroimage. 2011; 54: 2033–2044. [PubMed: 20851191]

- Avants BB, Yushkevich P, Pluta J, Minkoff D, Korczykowski M, Detre J, Gee JC. The optimal template effect in hippocampus studies of diseased populations. Neuroimage. 2010; 49: 2457–2466. [PubMed: 19818860]
- Bakken TE, Miller JA, Ding S-L, Sunkin SM, Smith KA, Ng L, Szafer A, Dalley RA, Royall JJ, Lemon T, Shapouri S, et al. A comprehensive transcriptional map of primate brain development. Nature. 2016; 535: 367–375. [PubMed: 27409810]
- Bonaiuto JJ, Rossiter HE, Meyer SS, Adams N, Little S, Callaghan MF, Dick F, Bestmann S, Barnes GR. Non-invasive laminar inference with MEG: comparison of methods and source inversion algorithms. Neuroimage. 2018; 167: 372–383. [PubMed: 29203456]
- Boucher M, Whitesides S, Evans A. Depth potential function for folding pattern representation, registration and analysis. Med Image Anal. 2009; 13: 203–214. [PubMed: 18996043]
- Calabrese E, Badea A, Coe CL, Lubach GR, Shi Y, Styner MA, Allan Johnson G. A diffusion tensor MRI atlas of the postmortem rhesus macaque brain. NeuroImage. 2015; doi: 10.1016/ j.neuroimage.2015.05.072
- Collins DL, Neelin P, Peters TM, Evans AC. Automatic 3D intersubject registration of MR volumetric data in standardized Talairach space. J Comput Assist Tomogr. 1994; 18: 192–205. [PubMed: 8126267]
- Deichmann R, Schwarzbauer C, Turner R. Optimisation of the 3D MDEFT sequence for anatomical brain imaging: technical implications at 1.5 and 3 T. Neuroimage. 2004; 21: 757–767. [PubMed: 14980579]
- Donahue CJ, Glasser MF, Preuss TM, Rilling JK, Van Essen DC. Quantitative assessment of prefrontal cortex in humans relative to nonhuman primates. Proc Natl Acad Sci U S A. 2018; 115: E5183– E5192. [PubMed: 29739891]
- Fischl B. FreeSurfer. Neuroimage. 2012; 62: 774-781. [PubMed: 22248573]
- Fischl B, Sereno MI, Tootell RB, Dale AM. High-resolution intersubject averaging and a coordinate system for the cortical surface. Hum Brain Mapp. 1999; 8: 272–284. [PubMed: 10619420]
- Garcia-Saldivar P, Garimella A, Garza-Villarreal EA, Mendez FA, Concha L, Merchant H. PREEMACS: Pipeline for Preprocessing and Extraction of the Macaque Brain Surface. NeuroImage.
- Glasser MF, Coalson TS, Robinson EC, Hacker CD, Harwell J, Yacoub E, Ugurbil K, Andersson J, Beckmann CF, Jenkinson M, Smith SM, et al. A multi-modal parcellation of human cerebral cortex. Nature. 2016; 536: 171–178. [PubMed: 27437579]
- Gorgolewski KJ, Varoquaux G, Rivera G, Schwarz Y, Ghosh SS, Maumet C, Sochat VV, Nichols TE, Poldrack RA, Poline J-B, Yarkoni T, et al. NeuroVault.org: a web-based repository for collecting and sharing unthresholded statistical maps of the human brain. Front Neuroinform. 2015; 9: 8. [PubMed: 25914639]
- Greve DN, Svarer C, Fisher PM, Feng L, Hansen AE, Baare W, Rosen B, Fischl B, Knudsen GM. Cortical surface-based analysis reduces bias and variance in kinetic modeling of brain PET data. Neuroimage. 2014; 92: 225–236. [PubMed: 24361666]
- Hawrylycz MJ, Lein ES, Guillozet-Bongaarts AL, Shen EH, Ng L, Miller JA, van de Lagemaat LN, Smith KA, Ebbert A, Riley ZL, Abajian C, et al. An anatomically comprehensive atlas of the adult human brain transcriptome. Nature. 2012; 489: 391–399. [PubMed: 22996553]
- Horsley V, Clarke RH. The structure and functions of the cerebellum examined by a new method. Brain. 1908; doi: 10.1093/brain/31.1.45
- Huth AG, de Heer WA, Griffiths TL, Theunissen FE, Gallant JL. Natural speech reveals the semantic maps that tile human cerebral cortex. Nature. 2016; 532: 453–458. [PubMed: 27121839]
- Jung B, Taylor PA, Seidlitz J, Sponheim C, Perkins P, Ungerleider LG, Glen D, Messinger A. A comprehensive macaque fMRI pipeline and hierarchical atlas. NeuroImage.
- Kim JS, Singh V, Lee JK, Lerch J, Ad-Dab'bagh Y, MacDonald D, Lee JM, Kim SI, Evans AC. Automated 3-D extraction and evaluation of the inner and outer cortical surfaces using a Laplacian map and partial volume effect classification. Neuroimage. 2005; 27: 210–221. [PubMed: 15896981]

Lepage et al.

- Koo B-B, Schettler SP, Murray DE, Lee J-M, Killiany RJ, Rosene DL, Kim D-S, Ronen I. Age-related effects on cortical thickness patterns of the Rhesus monkey brain. Neurobiol Aging. 2012; 33: 200 e23–31.
- Lee J-H, Garwood M, Menon R, Adriany G, Andersen P, Truwit CL, Uğurbil K. High contrast and fast three-dimensional magnetic resonance imaging at high fields. Magn Reson Med. 1995; 34: 308–312. [PubMed: 7500867]
- Lepage C, Lewis L, Jeun S, Bermudez P, Khalili-Mahani N, Omidyegaheh M, Zijdenbos A, Vincent RD, Adalat R, Evans AC. Human MR Evaluation of Cortical Thickness Using CIVET v2.1. Organ Hum Brain Map. 2017.
- Lerch JP, Evans AC. Cortical thickness analysis examined through power analysis and a population simulation. Neuroimage. 2005; 24: 163–173. [PubMed: 15588607]
- Lerch JP, Pruessner JC, Zijdenbos A, Hampel H, Teipel SJ, Evans AC. Focal decline of cortical thickness in alzheimer's disease identified by computational neuroanatomy. Cereb Cortex. 2005; 15: 995–1001. [PubMed: 15537673]
- Love SA, Marie D, Roth M, Lacoste R, Nazarian B, Bertello A, Coulon O, Anton J-L, Meguerditchian A. The average baboon brain: MRI templates and tissue probability maps from 89 individuals. Neuroimage. 2016; 132: 526–533. [PubMed: 26975558]
- Lyttelton O, Boucher M, Robbins S, Evans A. An unbiased iterative group registration template for cortical surface analysis. Neuroimage. 2007; 34: 1535–1544. [PubMed: 17188895]
- Messinger A, Sirmpilatze N, Heuer K, Loh KK, Mars R, Sein J, Xu T, Glen D, Jung B, Seidlitz J, Taylor P, et al. An open resource for non-human primate neuroimaging data analysis. NeuroImage.
- Milham MP, Ai L, Koo B, Xu T, Amiez C, Balezeau F, Baxter MG, Blezer ELA, Brochier T, Chen A, Croxson PL, et al. An open resource for non-human primate imaging. Neuron. 2018; 100: 61–74. e2 [PubMed: 30269990]
- Miranda-Dominguez O, Mills BD, Grayson D, Woodall A, Grant KA, Kroenke CD, Fair DA. Bridging the gap between the human and macaque connectome: a quantitative comparison of global interspecies structure-function relationships and network topology. J Neurosci. 2014; 34: 5552– 5563. [PubMed: 24741045]
- Oguz I, Styner M, Sanchez M, Shi Y, Sonka M. LOGISMOS-B for primates: primate cortical surface reconstruction and thickness measurement. Proc SPIE Int Soc Opt Eng. 2015; 9413. doi: 10.1117/12.2082327
- Reveley C, Gruslys A, Ye FQ, Glen D, Samaha JE, Russ B, Saad ZK, Seth A, Leopold DA, Saleem KS. Three-dimensional digital template atlas of the macaque brain. Cereb Cortex. 2017; 27: 4463– 4477. [PubMed: 27566980]
- Robbins, S. Anatomical standardization of the human brain in euclidean 3-space and on the cortical 2-manifold. [PhD thesis]. School of Computer Science, McGill University; Montreal, Quebec, Canada: 2003.
- Robinson EC, Jbabdi S, Glasser MF, Andersson J, Burgess GC, Harms MP, Smith SM, Van Essen DC, Jenkinson M. MSM: a new flexible framework for Multimodal Surface Matching. Neuroimage. 2014; 100: 414–426. [PubMed: 24939340]
- Saleem, KS, Logothetis, NK. A Combined MRI and Histology Atlas of the Rhesus Monkey Brain in Stereotaxic Coordinates. Academic Press; 2012.
- Scott JA, Grayson D, Fletcher E, Lee A, Bauman MD, Schumann CM, Buonocore MH, Amaral DG. Longitudinal analysis of the developing rhesus monkey brain using magnetic resonance imaging: birth to adulthood. Brain Struct Funct. 2016; doi: 10.1007/s00429-015-1076-x
- Seidlitz J, Sponheim C, Glen D, Ye FQ, Saleem KS, Leopold DA, Ungerleider L, Messinger A. A population MRI brain template and analysis tools for the macaque. Neuroimage. 2018; 170: 121– 131. [PubMed: 28461058]
- Sherif T, Rioux P, Rousseau M-E, Kassis N, Beck N, Adalat R, Das S, Glatard T, Evans AC. CBRAIN: a web-based, distributed computing platform for collaborative neuroimaging research. Front Neuroinform. 2014; 8: 54. [PubMed: 24904400]
- Sled JG, Zijdenbos AP, Evans AC. A nonparametric method for automatic correction of intensity nonuniformity in MRI data. IEEE Trans Med Imaging. 1998; 17: 87–97. [PubMed: 9617910]

Lepage et al.

- Smith SM. Fast robust automated brain extraction. Hum Brain Mapp. 2002; 17: 143–155. [PubMed: 12391568]
- The HDF Group. Hierarchical Data Format, version 5. 2016. 1997–2016. URL: http:// www.hdfgroup.org/HDF5
- Tohka J, Zijdenbos A, Evans A. Fast and robust parameter estimation for statistical partial volume models in brain MRI. Neuroimage. 2004; 23: 84–97. [PubMed: 15325355]
- Tustison NJ, Cook PA, Klein A, Song G, Das SR, Duda JT, Kandel BM, van Strien N, Stone JR, Gee JC, Avants BB. Large-scale evaluation of ANTs and FreeSurfer cortical thickness measurements. Neuroimage. 2014; 99: 166–179. [PubMed: 24879923]
- Vincent RD, Neelin P, Khalili-Mahani N, Janke AL, Fonov VS, Robbins SM, Baghdadi L, Lerch J, Sled JG, Adalat R, MacDonald D, Zijdenbos AP, Collins DL, Evans AC. MINC 2.0: A flexible format for multi-modal images. Front Neuroinform. 2016; 10: 35. [PubMed: 27563289]
- Wagstyl K, Larocque S, Cucurull G, Lepage C, Cohen JP, Bludau S, Palomero-Gallagher N, Lewis LB, Funck T, Spitzer H, Dickscheid T, Fletcher PC, Romero A, Zilles K, Amunts K, Bengio Y, Evans AC. BigBrain 3D atlas of cortical layers: Cortical and laminar thickness gradients diverge in sensory and motor cortices. PLoS Biol. 2020; 18 e3000678 [PubMed: 32243449]
- Wagstyl K, Ronan L, Goodyer IM, Fletcher PC. Cortical thickness gradients in structural hierarchies. Neuroimage. 2015; 111: 241–250. [PubMed: 25725468]
- Wang, F; Lian, C; Xia, J; Wu, Z; Duan, D; Wang, L; Shen, D; Li, G. Construction of spatiotemporal infant cortical surface atlas of rhesus macaque; 2018 IEEE 15th International Symposium on Biomedical Imaging (ISBI 2018); 2018. 704–707.
- Weiskopf N, Suckling J, Williams G, Correia MM, Inkster B, Tait R, Ooi C, Bullmore ET, Lutti A. Quantitative multi-parameter mapping of R1, PD(*), MT, and R2(*) at 3T: a multi-center validation. Front Neurosci. 2013; 7: 95. [PubMed: 23772204]
- Xu T, Nenning K-H, Schwartz E, Hong S-J, Vogelstein JT, Fair DA, Schroeder CE, Margulies DS, Smallwood J, Milham MP, Langs G. Cross-species functional alignment reveals evolutionary hierarchy within the connectome. bioRxiv. 2019; doi: 10.1101/692616
- Yeo BTT, Krienen FM, Sepulcre J, Sabuncu MR, Lashkari D, Hollinshead M, Roffman JL, Smoller JW, Zöllei L, Polimeni JR, Fischl B, et al. The organization of the human cerebral cortex estimated by intrinsic functional connectivity. J Neurophysiol. 2011; 106: 1125–1165. [PubMed: 21653723]
- Young JT, Shi Y, Niethammer M, Grauer M, Coe CL, Lubach GR, Davis B, Budin F, Knickmeyer RC, Alexander AL, Styner MA. The UNC-Wisconsin Rhesus Macaque Neurodevelopment Database: A Structural MRI and DTI database of early postnatal development. Front Neurosci. 2017; 11: 29. [PubMed: 28210206]
- Zijdenbos, A, Forghani, R, Evans, A. Medical Image Computing and Computer-Assisted Intervention —MICCAI'98, Lecture Notes in Computer Science. Wells, WM, Colchester, A, Delp, S, editors. Springer Berlin Heidelberg; Berlin, Heidelberg: 1998. 439–448.

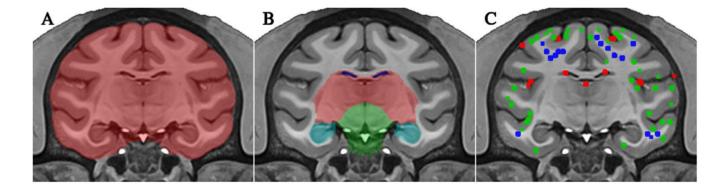


Fig. 1. Preparation of NMT

A) brain mask (red); B) regional masks for ventricles (dark blue), subcortical region (red), brainstem (green), cerebellum (not visible in this plane), hippocampus (cyan), and amygdala (not visible in this plane); C) subset of the points used to define priors for the three tissue classes: CSF (red), GM (green), WM (blue).

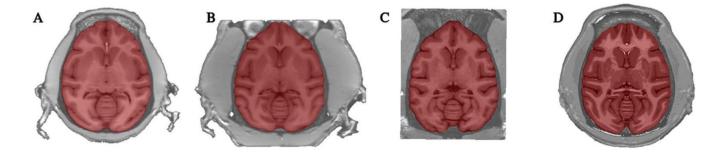


Fig. 2. Automated extraction of brain mask in NMT standardized space on sample scans of the NMT cohort, highlighting head size differences in male and female macaques and inter-scan field-of-view variability

A) female macaque; B) male macaque; C) male macaque with cropped field of view; and D) the NMT volume itself (25 males, 6 females). All masks are perfectly extracted regardless of the field of view of the original scan or the animal's head size.

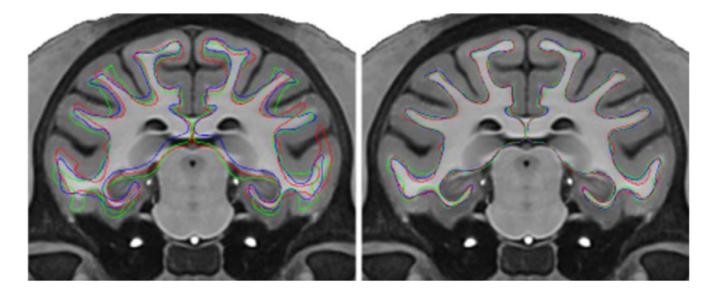


Fig. 3.

View of white matter surfaces for 3 NMT subjects in linear (affine) space on the left and in non-linear space on the right. Averaging in non-linear space eliminates shape differences while retaining the full cortical morphology of the macaque brain during the creation of the average white matter surface. The surfaces are overlaid onto the T1w NMT. The colored lines correspond to the three subjects.

Lepage et al.

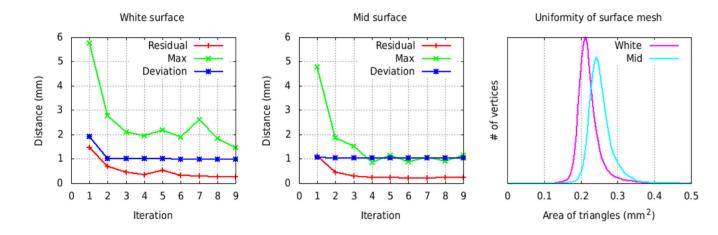


Fig. 4.

Convergence of the average symmetric white matter (left panel) and mid-cortical (middle panel) surfaces for the NMT cohort (N = 29). The residual represents the vertex-wise mean displacement in the average surface from one iteration to the next and should tend to zero as the vertices are no longer moving. Max represents the largest vertex-wise displacement from one iteration to the next. Deviation represents the mean vertex-wise displacement for the N = 29 subjects (58 surfaces) and is a measurement of inter-subject shape variability. The rightmost panel shows the histogram of the area of the triangles on the final average white matter and mid-cortical surfaces, indicating a nearly uniform point sampling over the surface.

Lepage et al.

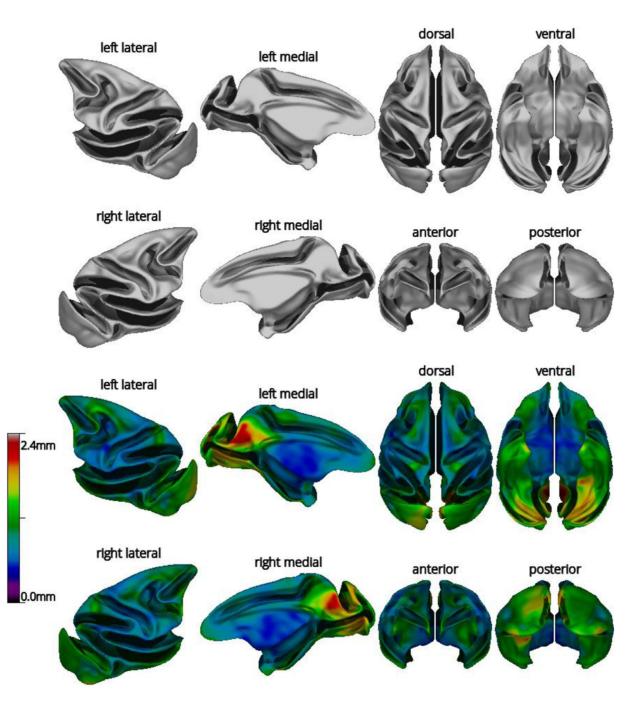


Fig. 5.

Views of the average white symmetric surface (top) and color map of mean vertex displacement for the N = 29 subjects of the NMT dataset (bottom). Displacements were larger in the parieto-occipital area (visual areas V6/V6A) and the occipital lobes, regions known to exhibit substantial inter-subject shape variability in macaques. For the top and bottom views, anterior is on top. For other views, dorsal is towards the top.

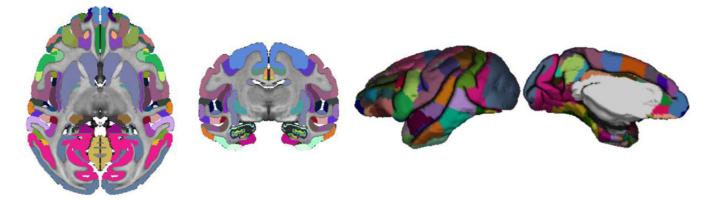


Fig. 6. D99 volumetric segmentation defined on the anatomical scan of the D99 subject, transposed to the D99 registered pial surface.

Lepage et al.

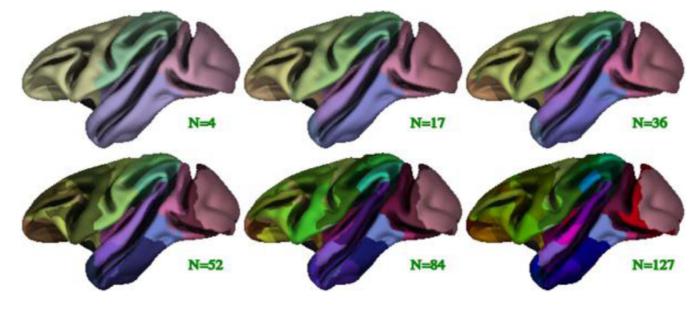


Fig. 7.

Illustration of the 6 levels of the symmetric CHARM parcellation viewed on the average symmetric mid-cortical surface for the NMT dataset. The CHARM parcellation is used in regional analyses by means of surface-based registration of individual subjects to the NMT surface template.

Lepage et al.

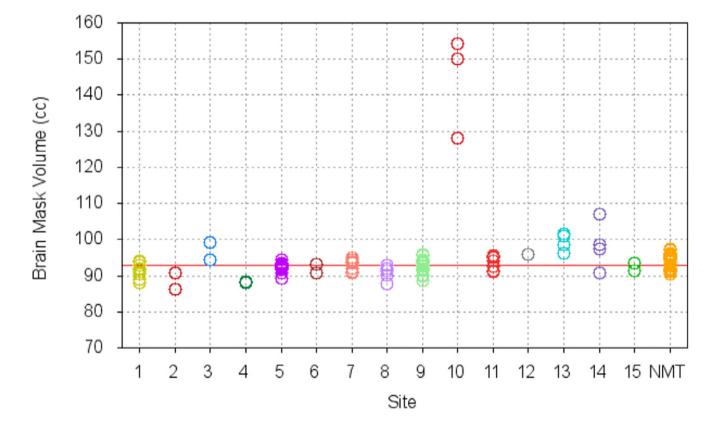


Fig. 8.

Brain mask volume in NMT standardized space for N = 95 scans from the INDI PRIME-DE consortium and N = 31 scans from the NMT cohort. The brain mask volume for the NMT volume is 92.8cc (red line). The small standard deviation for the volume of the brain mask in standardized space attests to the robustness of the brain extraction tool and the reliability of the volumetric registration to the NMT. The few obvious outliers show the processing failures.

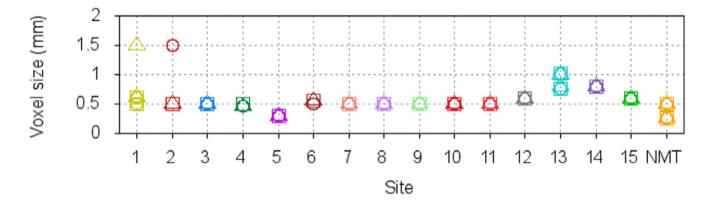


Fig. 9.

Acquisition voxel size for the N = 95 scans from the INDI PRIME-DE consortium and N = 31 scans from the NMT cohort. The three symbols (square, circle, triangle) refer to the voxel size in each of the 3 planes. By comparison to Fig. 8, scans with a voxel size above 0.5 mm tend to show higher deviation and variability from the volume of the brain mask of the NMT.

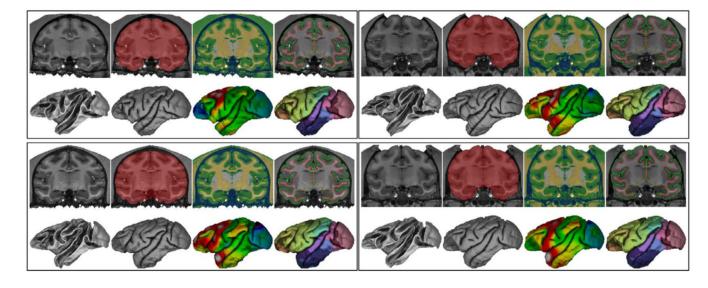


Fig. 10.

CIVET-macaque results for 4 NMT subjects. Within each set of images for a single subject, the top row, from left to right: coronal slice of the T1w image, brain mask, tissue classification (CSF in blue, cortical GM in green, portions of non-cortical GM in white, WM and other non-cortical GM in yellow), and cortical surfaces overlaid on the T1w image (WM surface in red, pial surface in green) bottom row, from left to right: white and pial surfaces; cortical thickness (range 1.0 mm light blue to 3.0 mm red) and the Level-4 CHARM parcellation viewed on the pial surface (52 ROIs).

Lepage et al.

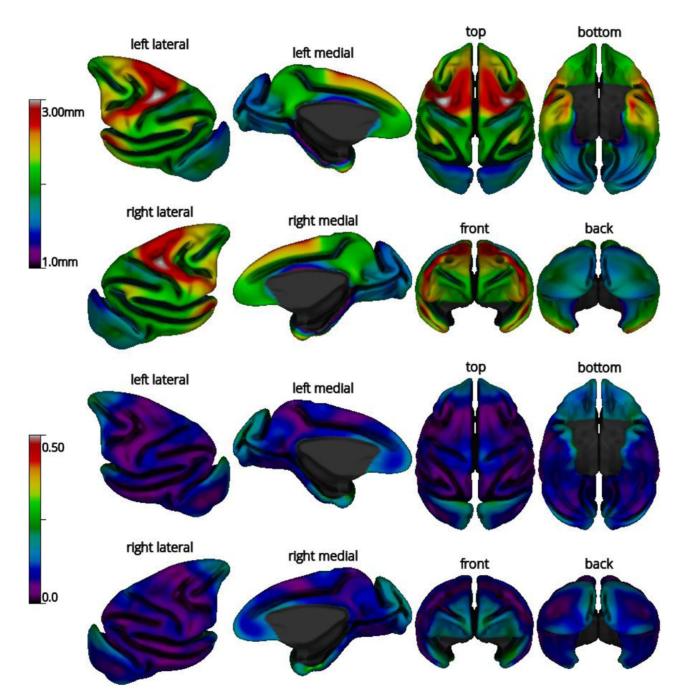


Fig. 11.

Mean cortical thickness (top) and coefficient of variation (COV) of cortical thickness (bottom) for the subjects of the NMT dataset (N = 29), viewed on the NMT average midcortical surface. Cortical thickness was computed from the tlaplace metric and smoothed over the surface using a fwhm of 8mm. For the top and bottom views, anterior is on top. For other views, dorsal is towards the top.

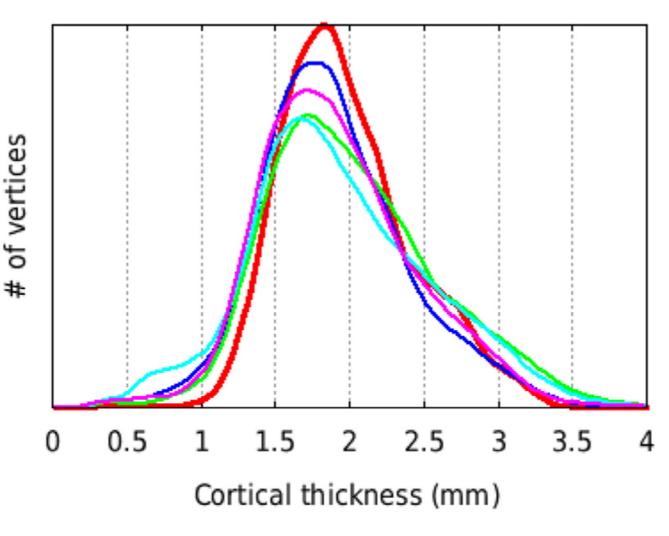


Fig. 12.

Histogram of mean cortical thickness for N = 29 subjects of the NMT dataset (red thick line) and for 4 single NMT subjects (thin lines), using the tlaplace metric with no spatial smoothing (fwhm = 0 mm). All 5 curves show the same total number of vertices (40,962 × 2 hemispheres) and area under the curve. The distribution for the mean curve is narrower (i.e., more central) than for most individual subjects. Consequently, the mean also has a higher peak than the other distributions.

Lepage et al.

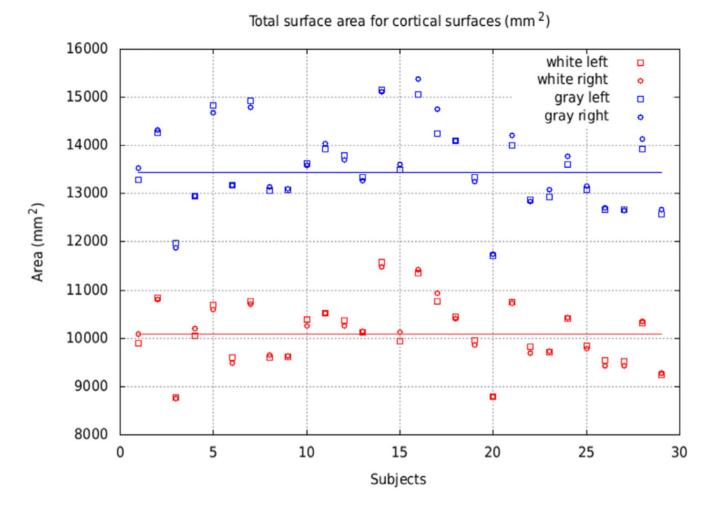


Fig. 13.

Total area for the white matter (red) and pial (blue) surfaces for the NMT cohort (N = 29), for left and right hemispheres. The area per hemisphere, averaged across subjects and hemispheres, is 10,098 mm² for the white matter surface (red line) and 13,441 mm² for the pial surface (blue line). Left and right surface areas agreed for all subjects, independently of inter-subject variability.

Table 1

Scanning information for 95 subjects with T1w scans obtained from the INDI PRIME-DE consortium. For horizontal scanners, acquisition was in the sphinx position and all scans included complete published information about age and sex.

| Site | N (M/F) | Age range (yr) | Scanner Type |
|---|-----------|----------------|-----------------------|
| 1-Newcastle University | 12 (10/2) | 5–13 | Bruker Vertical, 4.7T |
| 2-NIMH (Leopold, Russ) | 2 (1/1) | 6–7 | Bruker Vertical, 4.7T |
| 3-Princeton University | 2 (2/0) | 3 | Siemens Prisma, 3T |
| 4-University of Minnesota | 2 (0/2) | 10+ | Siemens Syngo, 7T |
| 5-UC Davis | 19 (0/19) | 18-22 | Siemens Skyra, 3T |
| 6-NKI | 2 (1/1) | 6–7 | Siemens TIM Trio, 3T |
| 7-Mount Sinai | 9 (8/1) | 3–8 | Philips Achieva, 3T |
| 8-Mount Sinai | 5 (5/0) | 5–6 | Siemens Skyra, 3T |
| 9-Oxford University | 20 (20/0) | 2–6 | n/a, 3T |
| 10-Western Ontario | 3 (3/0) | 4–7 | Siemens Magnetom, 7T |
| 11-Institute of Neuroscience | 8 (7/1) | 3–5 | Siemens TIM Trio, 3T |
| 12-McGill University | 1 (0/1) | 12 | Siemens TIM Trio, 3T |
| 13-East China Normal University-Kwok | 4 (4/0) | 2–3 | Siemens TIM Trio, 3T |
| 14-Aix-Marseille Universite | 4 (3/1) | 7–8 | Siemens Prisma, 3T |
| 15-Netherlands Institute for Neuroscience | 2 (2/0) | 5 | Philips Ingenia, 3T |

Table 2

Differences in processing steps between the CIVET-human and CIVET-macaque configurations. The length scale is defined as a linear factor relative to the human brain size. The fwhm on surfaces is scaled to account for the gyral patterns in macaques.

| Processing Step | Human Processing | Macaque Processing |
|---|----------------------------|----------------------------|
| Stereotaxic template | MNI-ICBM, ADNI, Colin27 | NMT |
| Stereotaxic voxel sampling | 0.50 or 1.0 mm | 0.25 or 0.50 mm |
| Volumetric registration (fwhm, simplex, sampling) | length scale 1.0 | length scale 0.4 |
| Neck cropping (headheight) | 175 mm | 50 mm |
| Masking (search distance) | length scale 1.0 | length scale 0.4 |
| Non-uniformity corrections | spline distance 125 (3T) | spline distance 50 (4.7T) |
| Classification | priors on human template | priors on NMT |
| Segmented masks (brainstem, ventricles, cerebellum, etc.) | on human template | on NMT |
| Surface extraction (fwhm, gradient, dilation/erosion) | length scale 1.0 | length scale 0.4 |
| Surface resolution | 40,962 or 163,842 vertices | 40,962 or 163,842 vertices |
| Cortical thickness method | tlink, tlaplace, tfs | tlink, tlaplace, tfs |
| Cortical thickness blurring | 30 mm fwhm | 8mm fwhm |
| Surface registration (fwhm) | length scale 1.0 | length scale 0.4 |
| Surface parcellation | AAL, DKT, lobes | CHARM, D99, lobes |

Table 3

Regional mean cortical thickness of the NMT cohort (N = 29) for the 3 broadest levels of the CHARM anatomical atlas, sorted by ordinal label number. Region names and atlas number labels are listed in different fonts and with different degrees of indentation depending on the CHARM level(s) in which they appear. Levels 1, 2, and 3, which contain 4,17, and 36 regions, respectively, are included. For each subject, the mean and standard deviation of the thickness (tlaplace metric with fwhm of 0mm) was computed across the vertices within each cortical region and hemisphere. Each region's mean and standard deviation was then averaged across subjects. The overall standard deviation is higher than the standard deviation of individual specialized regions, indicating higher inter-regional variability than intra-regional variability. Thinner areas include somatosensory, auditory and visual cortices, while temporal pole and motor cortex were the thickest areas. The thickness of each region was nearly identical between the left and right hemispheres. The smaller cortical thickness values in the parahippocampal cortex (region 148) reflect its proximity to the hippocampus, which is not modelled. This region should be excluded in analyses.

| Level Label Description Left Right 1 1 Frontal Lobe 2.33 ± 0.43 2.32 ± 0.43 2 2 anterior cingulate gyrus 1.99 ± 0.27 2.02 ± 0.30 3 3 anterior cingulate cortex 2.07 ± 0.29 2.13. ± 0.33 3 11 mid-cingulate cortex 1.86 ± 0.14 1.85 ± 0.13 2 16 orbital frontal cortex 2.10 ± 0.33 2.10 ± 0.33 2.10 ± 0.33 2.10 ± 0.35 3 17 medial orbital frontal cortex 2.10 ± 0.26 2.11 ± 0.27 3 25 lateral orbital frontal cortex 2.19 ± 0.51 2.13 ± 0.58 2 50 lateral prefrontal cortex 2.19 ± 0.51 2.13 ± 0.58 3 51 periarcuate area 8A (Frontal Eye Fields) 2.32 ± 0.19 2.31 ± 0.20 3 64 ventrolateral prefrontal cortex 2.65 ± 0.37 2.63 ± 0.36 3 18 lateral motor cortex 2.65 ± 0.37 2.63 ± 0.36 3 90 parietal Lobe 1.94 ± | CHARM parcellation Cortical thick | | | | |
|--|-----------------------------------|-------|--|-----------------------------------|------------------|
| 22anterior cingulate gyrus 1.99 ± 0.27 2.02 ± 0.30 33anterior cingulate cortex 2.07 ± 0.29 2.13 ± 0.33 311mid-cingulate cortex 1.86 ± 0.14 1.85 ± 0.13 216orbital frontal cortex 2.10 ± 0.33 2.10 ± 0.33 317medial orbital frontal cortex 2.03 ± 0.27 2.07 ± 0.30 325lateral orbital frontal cortex 2.10 ± 0.26 2.11 ± 0.27 337caudal orbital frontal cortex 2.19 ± 0.51 2.13 ± 0.58 250lateral prefrontal cortex 2.22 ± 0.36 2.20 ± 0.35 351periarcuate area 8A (Frontal Eye Fields) 2.32 ± 0.19 2.31 ± 0.20 354dorsolateral prefrontal cortex 2.14 ± 0.38 2.10 ± 0.40 277motor cortex 2.66 ± 0.37 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.66 ± 0.37 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.66 ± 0.37 2.63 ± 0.36 2.61 ± 0.37 390Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.88 ± 0.30 1.88 ± 0.30 392primary somatosensory cortex 1.87 ± 0.32 1.85 ± 0.32 395secondary somatosensory cortex 1.98 ± 0.17 1.90 ± 0.23 397visual areas V6 and V6A 2.08 ± 0.17 2.08 ± 0.17 3102area 5 <th>Level</th> <th>Label</th> <th>Description</th> <th>Left</th> <th>Right</th> | Level | Label | Description | Left | Right |
| 33anterior cingulate cortex 2.07 ± 0.29 2.13 ± 0.33 311mid-cingulate cortex 1.86 ± 0.14 1.85 ± 0.13 216orbital frontal cortex 2.10 ± 0.33 2.10 ± 0.35 317medial orbital frontal cortex 2.03 ± 0.27 2.07 ± 0.30 325lateral orbital frontal cortex 2.03 ± 0.27 2.07 ± 0.30 325lateral orbital frontal cortex 2.10 ± 0.26 2.11 ± 0.27 337caudal orbital frontal cortex 2.19 ± 0.51 2.13 ± 0.58 250lateral prefrontal cortex 2.22 ± 0.36 2.20 ± 0.35 351periarcuate area 8A (Frontal Eye Fields) 2.32 ± 0.19 2.31 ± 0.20 354dorsolateral prefrontal cortex 2.27 ± 0.34 2.27 ± 0.33 364ventrolateral prefrontal cortex 2.66 ± 0.37 2.63 ± 0.36 277motor cortex 2.66 ± 0.37 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.66 ± 0.37 2.63 ± 0.30 1.88 ± 0.30 390Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.89 ± 0.30 1.88 ± 0.30 392primary somatosensory cortex 1.89 ± 0.17 1.96 ± 0.18 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.23 397visual areas V6 and V6A 2.08 ± 0.17 2.08 ± 0.17 3102area 5 2.0 | 1 | 1 | Frontal Lobe | 2.33 ± 0.43 | 2.32 ± 0.43 |
| 311mid-cingulate cortex 1.86 ± 0.14 1.85 ± 0.13 216orbital frontal cortex 2.10 ± 0.33 2.10 ± 0.35 317medial orbital frontal cortex 2.03 ± 0.27 2.07 ± 0.30 325lateral orbital frontal cortex 2.10 ± 0.26 2.11 ± 0.27 337caudal orbital frontal cortex 2.19 ± 0.51 2.13 ± 0.58 250lateral prefrontal cortex 2.22 ± 0.36 2.20 ± 0.35 351periarcuate area 8A (Frontal Eye Fields) 2.32 ± 0.19 2.31 ± 0.20 354dorsolateral prefrontal cortex 2.27 ± 0.34 2.27 ± 0.33 364ventrolateral prefrontal cortex 2.66 ± 0.37 2.63 ± 0.36 277motor cortex 2.66 ± 0.37 2.63 ± 0.37 378lateral motor cortex 2.66 ± 0.37 2.63 ± 0.31 387medial supplementary motor areas 2.45 ± 0.22 2.40 ± 0.27 190Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.89 ± 0.30 1.88 ± 0.30 392primary somatosensory cortex 1.94 ± 0.31 1.92 ± 0.31 391somatosensory cortex 1.94 ± 0.31 1.92 ± 0.31 392primary somatosensory cortex 1.89 ± 0.17 1.96 ± 0.18 392primary somatosensory cortex 1.94 ± 0.31 1.92 ± 0.31 3102area 5 2.01 ± 0.20 1.99 ± 0.20 < | 2 | 2 | anterior cingulate gyrus | 1.99 ± 0.27 | 2.02 ± 0.30 |
| 216orbital frontal cortex 2.10 ± 0.33 2.10 ± 0.35 317medial orbital frontal cortex 2.03 ± 0.27 2.07 ± 0.30 325lateral orbital frontal cortex 2.10 ± 0.26 2.11 ± 0.27 337caudal orbital frontal cortex 2.19 ± 0.51 2.13 ± 0.58 250lateral prefrontal cortex 2.22 ± 0.36 2.20 ± 0.35 351periarcuate area 8A (Frontal Eye Fields) 2.32 ± 0.19 2.31 ± 0.20 354dorsolateral prefrontal cortex 2.27 ± 0.34 2.27 ± 0.33 364ventrolateral prefrontal cortex 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.45 ± 0.22 2.40 ± 0.27 190Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.87 ± 0.32 1.85 ± 0.32 392primary somatosensory cortex 1.87 ± 0.32 1.85 ± 0.32 393secondary somatosensory cortex 1.98 ± 0.17 1.90 ± 0.23 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3101ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.15 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial supprior temporal area 1.72 ± 0.15 1.72 ± 0.15 | 3 | 3 | anterior cingulate cortex | 2.07 ± 0.29 | $2.13. \pm 0.33$ |
| 317medial orbital frontal cortex 2.03 ± 0.27 2.07 ± 0.30 325lateral orbital frontal cortex 2.10 ± 0.26 2.11 ± 0.27 337caudal orbital frontal cortex 2.19 ± 0.51 2.13 ± 0.58 250lateral prefrontal cortex 2.22 ± 0.36 2.20 ± 0.35 351periarcuate area 8A (Frontal Eye Fields) 2.32 ± 0.19 2.31 ± 0.20 354dorsolateral prefrontal cortex 2.27 ± 0.34 2.27 ± 0.33 364ventrolateral prefrontal cortex 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.66 ± 0.37 2.63 ± 0.36 387medial supplementary motor areas 2.45 ± 0.22 2.40 ± 0.27 190Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.89 ± 0.30 1.88 ± 0.30 392primary somatosensory cortex 1.98 ± 0.17 1.96 ± 0.18 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.23 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.15 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial superior temporal area 1.72 ± 0.15 1.72 ± 0.15 | 3 | 11 | mid-cingulate cortex | 1.86 ± 0.14 | 1.85 ± 0.13 |
| 325lateral orbital frontal cortex 2.10 ± 0.26 2.11 ± 0.27 337caudal orbital frontal cortex 2.19 ± 0.51 2.13 ± 0.58 250lateral prefrontal cortex 2.22 ± 0.36 2.20 ± 0.35 351periarcuate area 8A (Frontal Eye Fields) 2.32 ± 0.19 2.31 ± 0.20 354dorsolateral prefrontal cortex 2.27 ± 0.34 2.27 ± 0.33 364ventrolateral prefrontal cortex 2.14 ± 0.38 2.10 ± 0.40 277motor cortex 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.66 ± 0.37 2.63 ± 0.37 387medial supplementary motor areas 2.45 ± 0.22 2.40 ± 0.27 190Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.87 ± 0.32 1.88 ± 0.30 392primary somatosensory cortex 1.98 ± 0.17 1.96 ± 0.18 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.23 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial superior temporal area 1.72 ± 0.15 1.72 ± 0.15 | 2 | 16 | orbital frontal cortex | 2.10 ± 0.33 | 2.10 ± 0.35 |
| 337caudal orbital frontal cortex 2.19 ± 0.51 2.13 ± 0.58 250lateral prefrontal cortex 2.22 ± 0.36 2.20 ± 0.35 351periarcuate area 8A (Frontal Eye Fields) 2.32 ± 0.19 2.31 ± 0.20 354dorsolateral prefrontal cortex 2.27 ± 0.34 2.27 ± 0.33 364ventrolateral prefrontal cortex 2.14 ± 0.38 2.10 ± 0.40 277motor cortex 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.66 ± 0.37 2.63 ± 0.37 387medial supplementary motor areas 2.45 ± 0.22 2.40 ± 0.27 190Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.89 ± 0.30 1.88 ± 0.30 392primary somatosensory cortex 1.87 ± 0.32 1.85 ± 0.32 395secondary somatosensory cortex 1.98 ± 0.17 1.96 ± 0.18 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.23 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 2112inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3113lateral intraparietal sulcus 1.72 ± 0.15 1.72 ± 0.15 | 3 | 17 | medial orbital frontal cortex | 2.03 ± 0.27 | 2.07 ± 0.30 |
| 250lateral prefrontal cortex 2.22 ± 0.36 2.20 ± 0.35 351periarcuate area 8A (Frontal Eye Fields) 2.32 ± 0.19 2.31 ± 0.20 354dorsolateral prefrontal cortex 2.27 ± 0.34 2.27 ± 0.33 364ventrolateral prefrontal cortex 2.14 ± 0.38 2.10 ± 0.40 277motor cortex 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.66 ± 0.37 2.63 ± 0.36 387medial supplementary motor areas 2.45 ± 0.22 2.40 ± 0.27 190Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.87 ± 0.32 1.88 ± 0.30 392primary somatosensory cortex 1.87 ± 0.32 1.85 ± 0.32 395secondary somatosensory cortex 1.98 ± 0.17 1.96 ± 0.18 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.20 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3102area 5 2.01 ± 0.36 1.98 ± 0.16 3113lateral intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial superior temporal area 1.72 ± 0.15 1.72 ± 0.15 | 3 | 25 | lateral orbital frontal cortex | 2.10 ± 0.26 | 2.11 ± 0.27 |
| 351periarcuate area 8A (Frontal Eye Fields) 2.32 ± 0.19 2.31 ± 0.20 354dorsolateral prefrontal cortex 2.27 ± 0.34 2.27 ± 0.33 364ventrolateral prefrontal cortex 2.14 ± 0.38 2.10 ± 0.40 277motor cortex 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.66 ± 0.37 2.63 ± 0.36 387medial supplementary motor areas 2.45 ± 0.22 2.40 ± 0.27 190Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.89 ± 0.30 1.88 ± 0.30 392primary somatosensory cortex 1.87 ± 0.32 1.85 ± 0.32 395secondary somatosensory cortex 1.98 ± 0.17 1.90 ± 0.23 397visual areas V6 and V6A 2.08 ± 0.17 2.08 ± 0.17 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3113lateral intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial superior temporal area 1.72 ± 0.15 1.72 ± 0.15 | 3 | 37 | caudal orbital frontal cortex | 2.19 ± 0.51 | 2.13 ± 0.58 |
| 354dorsolateral prefrontal cortex 2.27 ± 0.34 2.27 ± 0.33 364ventrolateral prefrontal cortex 2.14 ± 0.38 2.10 ± 0.40 277motor cortex 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.66 ± 0.37 2.63 ± 0.36 387medial supplementary motor areas 2.45 ± 0.22 2.40 ± 0.27 190Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.89 ± 0.30 1.88 ± 0.30 392primary somatosensory cortex 1.87 ± 0.32 1.85 ± 0.32 395secondary somatosensory cortex 1.98 ± 0.17 1.96 ± 0.18 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.23 307visual areas V6 and V6A 2.08 ± 0.17 2.08 ± 0.17 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 2112inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial superior temporal area 1.72 ± 0.15 1.72 ± 0.15 | 2 | 50 | lateral prefrontal cortex | 2.22 ± 0.36 | 2.20 ± 0.35 |
| 364ventrolateral prefrontal cortex 2.14 ± 0.38 2.10 ± 0.40 277motor cortex 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.66 ± 0.37 2.63 ± 0.37 387medial supplementary motor areas 2.45 ± 0.22 2.40 ± 0.27 190Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.89 ± 0.30 1.88 ± 0.30 392primary somatosensory cortex 1.87 ± 0.32 1.85 ± 0.32 395secondary somatosensory cortex 1.98 ± 0.17 1.96 ± 0.18 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.23 397visual areas V6 and V6A 2.08 ± 0.17 2.08 ± 0.17 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 2112inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3113lateral intraparietal sulcus 1.72 ± 0.15 1.72 ± 0.15 | 3 | 51 | periarcuate area 8A (Frontal Eye Fields) | 2.32 ± 0.19 | 2.31 ± 0.20 |
| 277motor cortex 2.63 ± 0.36 2.61 ± 0.37 378lateral motor cortex 2.66 ± 0.37 2.63 ± 0.37 387medial supplementary motor areas 2.45 ± 0.22 2.40 ± 0.27 190Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.89 ± 0.30 1.88 ± 0.30 392primary somatosensory cortex 1.87 ± 0.32 1.85 ± 0.32 395secondary somatosensory cortex 1.98 ± 0.17 1.96 ± 0.18 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.23 397visual areas V6 and V6A 2.08 ± 0.17 2.08 ± 0.17 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 2112inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3113lateral intraparietal sulcus 1.72 ± 0.15 1.72 ± 0.15 | 3 | 54 | dorsolateral prefrontal cortex | 2.27 ± 0.34 | 2.27 ± 0.33 |
| 378lateral motor cortex2.66 \pm 0.372.63 \pm 0.37387medial supplementary motor areas2.45 \pm 0.222.40 \pm 0.27190Parietal Lobe1.94 \pm 0.311.92 \pm 0.31291somatosensory cortex1.89 \pm 0.301.88 \pm 0.30392primary somatosensory cortex1.87 \pm 0.321.85 \pm 0.32395secondary somatosensory cortex1.98 \pm 0.171.96 \pm 0.18296superior parietal lobule1.93 \pm 0.231.90 \pm 0.23397visual areas V6 and V6A2.08 \pm 0.172.08 \pm 0.173102area 52.01 \pm 0.201.99 \pm 0.203107ventromedial intraparietal sulcus1.76 \pm 0.181.72 \pm 0.162112inferior parietal lobule2.01 \pm 0.361.98 \pm 0.363113lateral intraparietal sulcus1.89 \pm 0.191.85 \pm 0.183119medial superior temporal area1.72 \pm 0.151.72 \pm 0.13 | 3 | 64 | ventrolateral prefrontal cortex | 2.14 ± 0.38 | 2.10 ± 0.40 |
| 387medial supplementary motor areas 2.45 ± 0.22 2.40 ± 0.27 190Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.89 ± 0.30 1.88 ± 0.30 392primary somatosensory cortex 1.87 ± 0.32 1.85 ± 0.32 395secondary somatosensory cortex 1.98 ± 0.17 1.96 ± 0.18 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.23 397visual areas V6 and V6A 2.08 ± 0.17 2.08 ± 0.17 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 2112inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial superior temporal area 1.72 ± 0.15 1.72 ± 0.15 | 2 | 77 | motor cortex | 2.63 ± 0.36 | 2.61 ± 0.37 |
| 190Parietal Lobe 1.94 ± 0.31 1.92 ± 0.31 291somatosensory cortex 1.89 ± 0.30 1.88 ± 0.30 392primary somatosensory cortex 1.87 ± 0.32 1.85 ± 0.32 395secondary somatosensory cortex 1.98 ± 0.17 1.96 ± 0.18 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.23 397visual areas V6 and V6A 2.08 ± 0.17 2.08 ± 0.17 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 2112inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial superior temporal area 1.72 ± 0.15 1.72 ± 0.13 | 3 | 78 | lateral motor cortex | 2.66 ± 0.37 | 2.63 ± 0.37 |
| 291somatosensory cortex 1.89 ± 0.30 1.88 ± 0.30 392primary somatosensory cortex 1.87 ± 0.32 1.85 ± 0.32 395secondary somatosensory cortex 1.98 ± 0.17 1.96 ± 0.18 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.23 397visual areas V6 and V6A 2.08 ± 0.17 2.08 ± 0.17 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 2112inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial superior temporal area 1.72 ± 0.15 1.72 ± 0.13 | 3 | 87 | medial supplementary motor areas | 2.45 ± 0.22 | 2.40 ± 0.27 |
| 392primary somatosensory cortex 1.87 ± 0.32 1.85 ± 0.32 395secondary somatosensory cortex 1.98 ± 0.17 1.96 ± 0.18 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.23 397visual areas V6 and V6A 2.08 ± 0.17 2.08 ± 0.17 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 2112inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial superior temporal area 1.72 ± 0.15 1.72 ± 0.13 | 1 | 90 | Parietal Lobe | $\textbf{1.94} \pm \textbf{0.31}$ | 1.92 ± 0.31 |
| 395secondary somatosensory cortex 1.98 ± 0.17 1.96 ± 0.18 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.23 397visual areas V6 and V6A 2.08 ± 0.17 2.08 ± 0.17 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 2112inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial superior temporal area 1.72 ± 0.15 1.72 ± 0.13 | 2 | 91 | somatosensory cortex | 1.89 ± 0.30 | 1.88 ± 0.30 |
| 296superior parietal lobule 1.93 ± 0.23 1.90 ± 0.23 397visual areas V6 and V6A 2.08 ± 0.17 2.08 ± 0.17 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 2112inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial superior temporal area 1.72 ± 0.15 1.72 ± 0.13 | 3 | 92 | primary somatosensory cortex | 1.87 ± 0.32 | 1.85 ± 0.32 |
| 397visual areas V6 and V6A 2.08 ± 0.17 2.08 ± 0.17 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 2112inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial superior temporal area 1.72 ± 0.15 1.72 ± 0.13 | 3 | 95 | secondary somatosensory cortex | 1.98 ± 0.17 | 1.96 ± 0.18 |
| 3102area 5 2.01 ± 0.20 1.99 ± 0.20 3107ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 2112inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3113lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3119medial superior temporal area 1.72 ± 0.15 1.72 ± 0.13 | 2 | 96 | superior parietal lobule | 1.93 ± 0.23 | 1.90 ± 0.23 |
| 3 107 ventromedial intraparietal sulcus 1.76 ± 0.18 1.72 ± 0.16 2 112 inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3 113 lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3 119 medial superior temporal area 1.72 ± 0.15 1.72 ± 0.13 | 3 | 97 | visual areas V6 and V6A | 2.08 ± 0.17 | 2.08 ± 0.17 |
| 2 112 inferior parietal lobule 2.01 ± 0.36 1.98 ± 0.36 3 113 lateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3 119 medial superior temporal area 1.72 ± 0.15 1.72 ± 0.13 | 3 | 102 | area 5 | 2.01 ± 0.20 | 1.99 ± 0.20 |
| 3 113 Iateral intraparietal sulcus 1.89 ± 0.19 1.85 ± 0.18 3 119 medial superior temporal area 1.72 ± 0.15 1.72 ± 0.13 | 3 | 107 | ventromedial intraparietal sulcus | 1.76 ± 0.18 | 1.72 ± 0.16 |
| 3 119 medial superior temporal area 1.72 ± 0.15 1.72 ± 0.13 | 2 | 112 | inferior parietal lobule | 2.01 ± 0.36 | 1.98 ± 0.36 |
| | 3 | 113 | lateral intraparietal sulcus | 1.89 ± 0.19 | 1.85 ± 0.18 |
| 3 120 area 7 in inferior parietal lobule $2.18 \pm 0.40 2.15 \pm 0.41$ | 3 | 119 | medial superior temporal area | 1.72 ± 0.15 | 1.72 ± 0.13 |
| | 3 | 120 | area 7 in inferior parietal lobule | 2.18 ± 0.40 | 2.15 ± 0.41 |

| CHARM parcellation Cortical thickness (mm | | | | |
|---|-------|--|-----------------------------------|-----------------|
| Level | Label | Description | Left | Right |
| 2 | 125 | posterior medial cortex | 1.97 ± 0.29 | 1.98 ± 0.29 |
| 3 | 126 | area 7 (PGm) on medial wall | 2.03 ± 0.20 | 2.05 ± 0.19 |
| 3 | 127 | posterior cingulate gyrus | 1.94 ± 0.33 | 1.95 ± 0.33 |
| 1 | 138 | <u>Temporal Lobe</u> | $\textbf{2.13} \pm \textbf{0.40}$ | 2.11 ± 0.42 |
| 2 | 139 | medial temporal lobe | 2.10 ± 0.66 | 1.99 ± 0.70 |
| 3 | 148 | parahippocampal cortex | 1.64 ± 0.25 | 1.52 ± 0.25 |
| 3 | 153 | rhinal cortex | 2.64 ± 0.57 | 2.56 ± 0.64 |
| 2/3 | 165 | temporal pole | 2.47 ± 0.51 | 2.51 ± 0.48 |
| 2 | 175 | inferior temporal cortex | 2.13 ± 0.23 | 2.13 ± 0.23 |
| 3 | 176 | area TEO | 2.09 ± 0.12 | 2.10 ± 0.11 |
| 3 | 177 | area TE | 2.25 ± 0.21 | 2.24 ± 0.22 |
| 3 | 188 | fundus of superior temporal sulcus | 1.92 ± 0.12 | 1.92 ± 0.12 |
| 2 | 193 | superior temporal region | 2.30 ± 0.33 | 2.28 ± 0.34 |
| 3 | 194 | rostral superior temporal region | 2.33 ± 0.36 | 2.32 ± 0.36 |
| 3 | 199 | caudal superior temporal gyrus | 2.23 ± 0.25 | 2.20 ± 0.26 |
| 2 | 204 | core and belt areas of auditory cortex | 1.80 ± 0.36 | 1.78 ± 0.36 |
| 3 | 205 | belt areas of auditory cortex | 1.78 ± 0.37 | 1.75 ± 0.34 |
| 3 | 218 | polar rostrotemporal cortex | 2.54 ± 0.45 | 2.64 ± 0.46 |
| 3 | 219 | core areas of auditory cortex | 1.74 ± 0.21 | 1.74 ± 0.23 |
| 2/3 | 224 | floor of the lateral sulcus | 2.04 ± 0.47 | 1.99 ± 0.53 |
| 1 | 231 | Occipital Lobe | 1.67 ± 0.25 | 1.66 ± 0.25 |
| 2/3 | 232 | middle temporal area | 1.72 ± 0.09 | 1.71 ± 0.08 |
| 2 | 233 | extrastriate visual areas 2-4 | 1.67 ± 0.27 | 1.66 ± 0.27 |
| 3 | 234 | visual area 4 | 1.91 ± 0.22 | 1.91 ± 0.21 |
| 3 | 237 | preoccipital visual areas 2-3 | 1.61 ± 0.24 | 1.60 ± 0.25 |
| 2/3 | 246 | primary visual cortex | 1.67 ± 0.23 | 1.65 ± 0.21 |
| | | Total | 1.99 ± 0.43 | 1.98 ± 0.44 |

Page 37