

## Research Article

# CG8005 Mediates Transit-Amplifying Spermatogonial Divisions via Oxidative Stress in *Drosophila* Testes

Wanyin Chen, Xiaojin Luan, Yidan Yan, Min Wang, Qianwen Zheng, Xia Chen , Jun Yu , and Jie Fang 

Department of Gynecology, The Affiliated Hospital of Jiangsu University, Jiangsu University, Zhenjiang, Jiangsu 212001, China

Correspondence should be addressed to Xia Chen; [chenxia\\_0511@126.com](mailto:chenxia_0511@126.com), Jun Yu; [yujun9117@126.com](mailto:yujun9117@126.com), and Jie Fang; [fangjie070@163.com](mailto:fangjie070@163.com)

Received 20 July 2020; Revised 19 September 2020; Accepted 30 September 2020; Published 28 October 2020

Academic Editor: Chia-Jung Li

Copyright © 2020 Wanyin Chen et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The generation of reactive oxygen species (ROS) widely occurs in metabolic reactions and affects stem cell activity by participating in stem cell self-renewal. However, the mechanisms of transit-amplifying (TA) spermatogonial divisions mediated by oxidative stress are not fully understood. Through genetic manipulation of *Drosophila* testes, we demonstrated that CG8005 regulated TA spermatogonial divisions and redox homeostasis. Using in vitro approaches, we showed that the knockdown of CG8005 increased ROS levels in S2 cells; the induced ROS generation was inhibited by NAC and exacerbated by H<sub>2</sub>O<sub>2</sub> pretreatments. Furthermore, the silencing of CG8005 increased the mRNA expression of oxidation-promoting factors Keap1, GstD1, and Mal-A6 and decreased the mRNA expression of antioxidant factors cnc, Gclm, maf-S, ND-42, and ND-75. We further investigated the functions of the antioxidant factor cnc, a key factor in the Keap1-cnc signaling pathway, and showed that cnc mimicked the phenotype of CG8005 in both *Drosophila* testes and S2 cells. Our results indicated that CG8005, together with cnc, controlled TA spermatogonial divisions by regulating oxidative stress in *Drosophila*.

## 1. Introduction

In adult *Drosophila* testes, two stem cell lineages, germline stem cells (GSCs) and cyst stem cells (CySCs), wrap around hub cells at the apex of the testis to form the germline stem cell niche [1, 2]. Hub cells mainly maintain the self-renewal and differentiation of these two types of stem cells [3, 4]. CySCs differentiate into Cyst cells, which provide an environment for the growth and differentiation of germ cells [2]. GSCs produce two progeny cells: one which stays in the niche to maintain the characteristics of stem cells and the other which departs from the niche, under the relevant signaling, and undergoes transit-amplifying (TA) spermatogonial divisions prior to terminal differentiation [5, 6].

Reactive oxygen species (ROS) are generated as byproducts of various cell metabolism and homeostasis events [7, 8]. An imbalance in ROS generation leads to apoptosis and tissue damage by triggering the caspase cascade [9, 10]. Oxidative stress can affect stem cell behavior by promoting dif-

ferentiation, proliferation, or apoptosis processes [11]. Evidence shows that mitochondrial dynamics play key roles in regulating early germ cell behavior and are important for regulating the maintenance of early germ cells, in larval testes [12]. By transcriptome analysis of adult testes, 152 genes were found with changes in mRNA expression levels during GSC differentiation induced by ROS accumulation [13]. In the *Drosophila* ovary, a genetic screen for follicle stem cell (FSC) self-renewal is implemented and can identify several genes that are required for FSC maintenance and the regulation of ROS generation. Ectopic expression of catalase or Gpx can eliminate ROS accumulation in mrpL4 and pdsw mutant clones, however, could not rescue defects in FSC maintenance [14].

The Keap1-Nrf2 (Nrf2 also known as cnc in *Drosophila*) signaling transduction pathway has been proven to play highly conserved roles in the regulation of oxidative stress homeostasis [15]. When under a lack of oxidative stress, Nrf2 acts as an antioxidant factor by binding to the

cytoplasmic inhibitor Keap1 through Cul3 ligase and preventing its transport into the nucleus. However, when exposed to oxidative stress, Nrf2 relocates from the cytoplasm to the nucleus and combines with Maf proteins to form a heterodimer, where it can recognize and bind with antioxidant response element (ARE) to promote the expression of antioxidant genes [11].

It has been revealed that defects in spermatogonial differentiation caused disorders in spermatogenesis and resulted in the formation of GSC-like tumors in *Drosophila* [16]. Dpp and Gbb proteins, which are recognized as BMP-like molecules, activate the BMP signaling pathway through membrane receptors, thereby inhibiting the expression of the differentiation factor Bam protein [17–19]. In Bam mutant testes, its absence has been shown to cause the accumulation of undifferentiated germ cells and eventually the formation of GSC-like tumors [20, 21]. However, the relationship between TA spermatogonial divisions and oxidative stress remains unknown. Previously, Yu et al. identified CG8005 as a regulator of stem cell niche homeostasis in *Drosophila* testes [22]. In this study, we investigated the actions of CG8005 during TA spermatogonial divisions and explored its potential involvement in the relationship between TA spermatogonial divisions and oxidative stress.

## 2. Materials and Methods

**2.1. Fly Strains and RNAi Strategies.** All flies were raised on standard corn meal food at 25°C and in a relative humidity of 60%. The transgenic RNAi flies used in this study were ordered from the Tsinghua Fly Center (THFC, Beijing, China). Transgenic strains used were as follows: UAS-CG8005 RNAi (THU4150, THFC), UAS-cnc RNAi (1; THU5248, THFC), and UAS-cnc RNAi (2; THU1052, THFC). The Bam-Gal4; $\Delta$ 86/+ line was a gift from DH Chen (Institute of Zoology, Chinese Academy of Sciences, Beijing, China). The  $W^{1118}$  line was used as the wild-type (WT) fly.

Male flies of Bam-Gal4 were selected to cross with virgin flies of the UAS-RNAi strains. Selected males with the desired genotypes in their offspring were used for further functional analysis.

**2.2. Immunofluorescence and Antibodies.** Fly testes were dissected in 1x phosphate-buffered saline (PBS) and fixed for 30 min in 4% paraformaldehyde. After washing three times in 1x PBS with 0.1% Triton X-100 (PBST) and blocking for 1 h in 5% bovine serum albumin, the testes were incubated with primary antibodies for 1 h at room temperature. Then, the samples were washed three times for 10 min in 0.1% PBST and incubated for 1 h with secondary antibodies at room temperature followed by the final three washes in 0.1% PBST. Testes were then stained with Hoechst 33342 (1.0 mg/mL; Invitrogen, CA, USA) for 5 min before mounting.

The primary antibodies used in this study were as follows: rabbit anti-Vasa (1:200; Santa Cruz Biotechnology, TX, USA), mouse anti-1B1 (1:50; Developmental Studies Hybridoma Bank, IA, USA), rat anti-DE-cadherin (1:20; DSHB), mouse anti-Eya (1:50; DSHB), rat anti-Zfh1

(1:1000; a gift from C Tong; Life Sciences Institute, Zhejiang University, Zhejiang, China), rabbit anti-PH3 (1:400; Cell Signaling Technology, Leiden, Netherlands), and mouse anti-FasIII (1:50; DSHB). Secondary antibodies were conjugated to A488, Cy3, or A647 (Molecular Probes and Jackson Immunological) and used at a dilution of 1:1000.

**2.3. Cell Culture and Transfection.** *Drosophila* Schneider 2 (S2) cells were obtained from the *Drosophila* Genomics Resource Center (IN, USA) and cultured in Schneider *Drosophila* Medium (21720024, Gibco, MA, USA) containing 10% heat-inactivated fetal bovine serum (04-001-1ACS; Biological Industries, Israel) at 28°C. The cells were separated with a supplementary medium at a ratio of 1:4 every 3–4 days. For the knockdown assay, S2 cells were transfected using Lipofectamine 2000 (Lipo2000; 11668019, Invitrogen). Negative control fragments were transfected and used as controls for S2 cells. siRNAs used in this study were designed and synthesized by GenePharma (Suzhou, China) and are listed in Table S1.

**2.4. Quantitative Reverse Transcription-Polymerase Chain Reaction (qRT-PCR).** Total RNA was extracted using TRIzol reagent (9108, Takara, Japan), cDNA was synthesized using the PrimeScript RT Reagent Kit (RR037A, Takara), and qRT-PCR was performed using SYBR Premix Ex Taq (RR420A, Takara), according to the manufacturer's instructions. Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was used as an internal reference. All primers used for qRT-PCR are listed in Table S2.

**2.5. ROS Assay.** S2 cells were isolated and divided into several groups; NAC or H<sub>2</sub>O<sub>2</sub> was added to the corresponding group. Samples were incubated at 28°C for 1 h to alter ROS production and then perform negative control and siRNA transfection. Dihydroethidium (DHE; Biyuntian, Shanghai, China) and 2,7-dichlorofluorescein diacetate (DCFH-DA; Biyuntian) were used to detect ROS generation based on fluorescence intensity. DHE fluorescence was used to detect superoxide production, as previously described [23, 24]. Intracellular ROS can oxidize nonfluorescent DCFH to generate fluorescent dichlorofluorescein (DCF) [25]. After transfection for 48 h, cells were incubated with DHE or DCFH-DA probes for 30 min in the dark. After removing DHE or DCFH-DA probes, the cells were washed with PBS. For *Drosophila*, testes were dissected in PBS and incubated with H<sub>2</sub>O<sub>2</sub> treatment for 1 h at room temperature. After being washed with PBS, the testes were then incubated with DHE working solution in the dark for 5 minutes. After being washed with PBS for three times, the testes were fixed for 30 min in 4% paraformaldehyde and stained with Hoechst 33342 (1.0 mg/mL; Invitrogen, CA, USA) for 5 minutes. Quantification analysis of fluorescence intensity was performed using the ImageJ software.

**2.6. Statistical Analysis.** Each experiment was repeated at least three times, and the data are presented as the mean  $\pm$  standard error of the mean (SEM). Comparisons among groups were determined by one-way analysis of variance

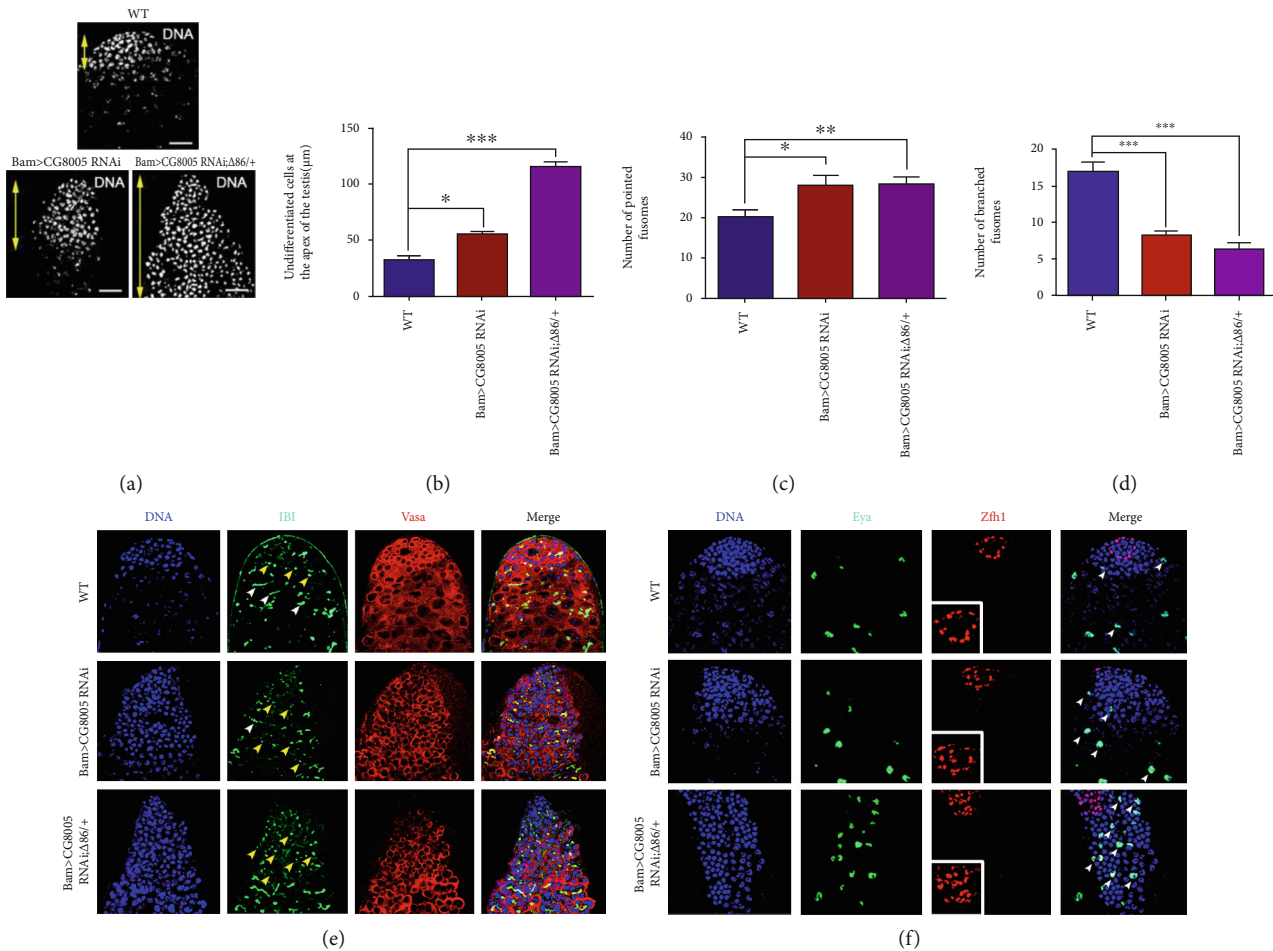


FIGURE 1: Knockdown of CG8005 in spermatogonia resulted in differentiation defects. (a) DNA staining in WT, Bam>CG8005 RNAi, and Bam>CG8005 RNAi;Δ86/+ testes. Yellow double arrows label undifferentiated cells at the apex of the testis. (b) The distance of undifferentiated cells at the apex of the testis. (c) The number of pointed fusomes. (d) The number of branched fusomes. (e, f) Immunostaining of WT, Bam>CG8005 RNAi, and Bam>CG8005 RNAi;Δ86/+ testes. Representative pointed fusomes are indicated with yellow arrowheads, and branched fusomes are indicated with white arrowheads. \* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$ . Scale bar: 20 μm.

(ANOVA) and Student's  $t$ -test using the GraphPad Software (<https://www.graphpad.com/>).

### 3. Results

**3.1. CG8005 Is Required for TA Spermatogonial Divisions.** To examine the physiological function of CG8005 during TA spermatogonial division, we performed a CG8005 RNAi assay in spermatogonia derived from Bam-Gal4 flies. It has been previously reported that the deletion of Bam, a key differentiation factor, could lead to defects in germ cell differentiation [26]. At the apex of the testis, the early stage of germ cells and cyst cells can be stained by DNA dye. Compared to WT flies, undifferentiated cells accumulated in Bam>CG8005 RNAi testes, and the phenotype was enhanced by heterozygous mutation of Bam in Bam>CG8005 RNAi testes (Figures 1(a) and 1(b)). In WT testes, fusomes, which were labeled by IBI, exhibited dynamic changes with punctate distribution in the early stage of germ cells, followed by development and transition into branched shapes in differentiated germ cells [27]. Then, we used immunofluorescence

staining to analyze the apex of testes of both WT and CG8005 RNAi flies. In Bam>CG8005 RNAi testes, the amount of accumulated undifferentiated cells, labeled by Vasa, and pointed fusomes dramatically increased, while the number of branched fusomes significantly decreased (Figures 1(c), 1(d), and 1(e)). Moreover, the differentiation defects of early stage germ cells were clearly exacerbated in Bam>CG8005 RNAi;Δ86/+ testes (Figures 1(c), 1(d), and 1(e)). Phosphorylated histone H3 (phospho-histone H3, PH3) was used as a marker for proliferating cells. We further used immunofluorescence technology to detect whether these accumulated undifferentiated cells obtained proliferation ability. FasIII was used to recognize the hub cells at the apex of the testis. In this study, we found that undifferentiated germ cells obtained their self-renewal ability without normal niche signals (Figure S1). We also stained testes with eyes absent (Eya) and zn finger homeodomain 1 (Zfh1) to distinguish mature cyst cells and early stage cyst cells, respectively. Interestingly, mature cyst cells and early stage cyst cells were present in both Bam>CG8005 RNAi and Bam>CG8005 RNAi;Δ86/+ testes (Figure 1(f)). Taken

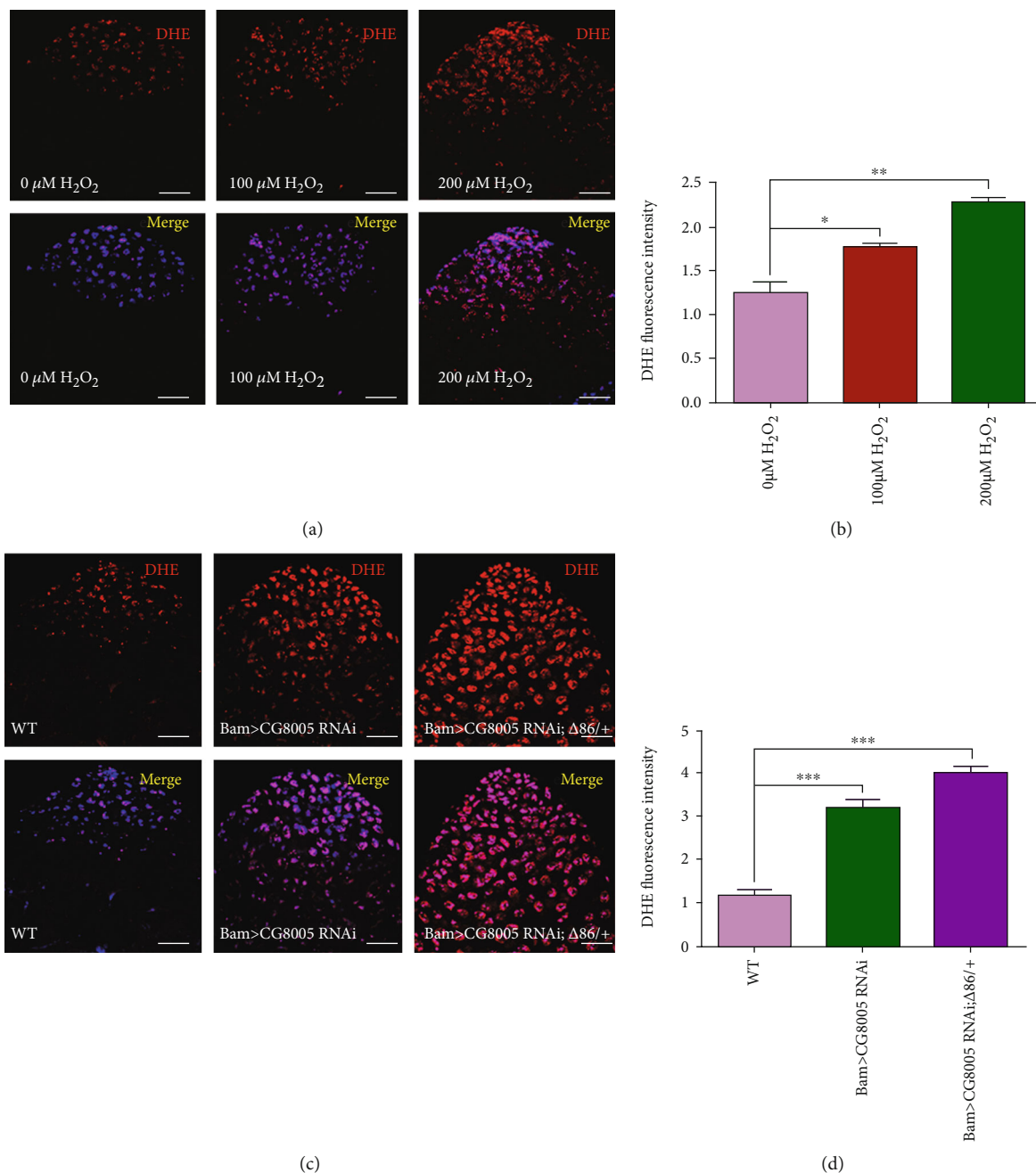


FIGURE 2: ROS generation in CG8005 RNAi testes. (a) DHE staining in testes with H<sub>2</sub>O<sub>2</sub> treatment. (b) DHE fluorescence intensity of testes with H<sub>2</sub>O<sub>2</sub> treatment. (c) DHE staining in WT, Bam>CG8005 RNAi, and Bam>CG8005 RNAi;Δ86/+ testes. (d) DHE fluorescence intensity of WT, Bam>CG8005 RNAi, and Bam>CG8005 RNAi;Δ86/+ testes. \*P < 0.05, \*\*P < 0.01, and \*\*\*P < 0.001. Scale bar: 20 μm.

together, our results indicated that CG8005 mediated germ cell differentiation in spermatogonia.

**3.2. Inhibition of CG8005 Increased ROS Levels in *Drosophila* Testes.** ROS signals are considered important stem cell regulators and affect stem cell homeostasis by facilitating the differentiation and self-renewal of multiple stem cell populations [28]. To further investigate whether dysfunctional CG8005-mediated spermatogonial differentiation defects were involved in oxidative stress, we examined the effect of

the redox state on both WT and CG8005 RNAi testes. Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) is a recognized oxidant, and we determined ROS levels by DHE intensity in H<sub>2</sub>O<sub>2</sub>-treated testes. We found that ROS production increased with H<sub>2</sub>O<sub>2</sub> dosage-dependent effects on *Drosophila* testes (Figures 2(a) and 2(b)). Importantly, we also found that ROS production dramatically increased in both Bam>CG8005 RNAi and Bam>CG8005 RNAi;Δ86/+ testes compared with WT testes (Figures 2(c) and 2(d)), indicating that CG8005 is required for redox homeostasis in *Drosophila* testes.

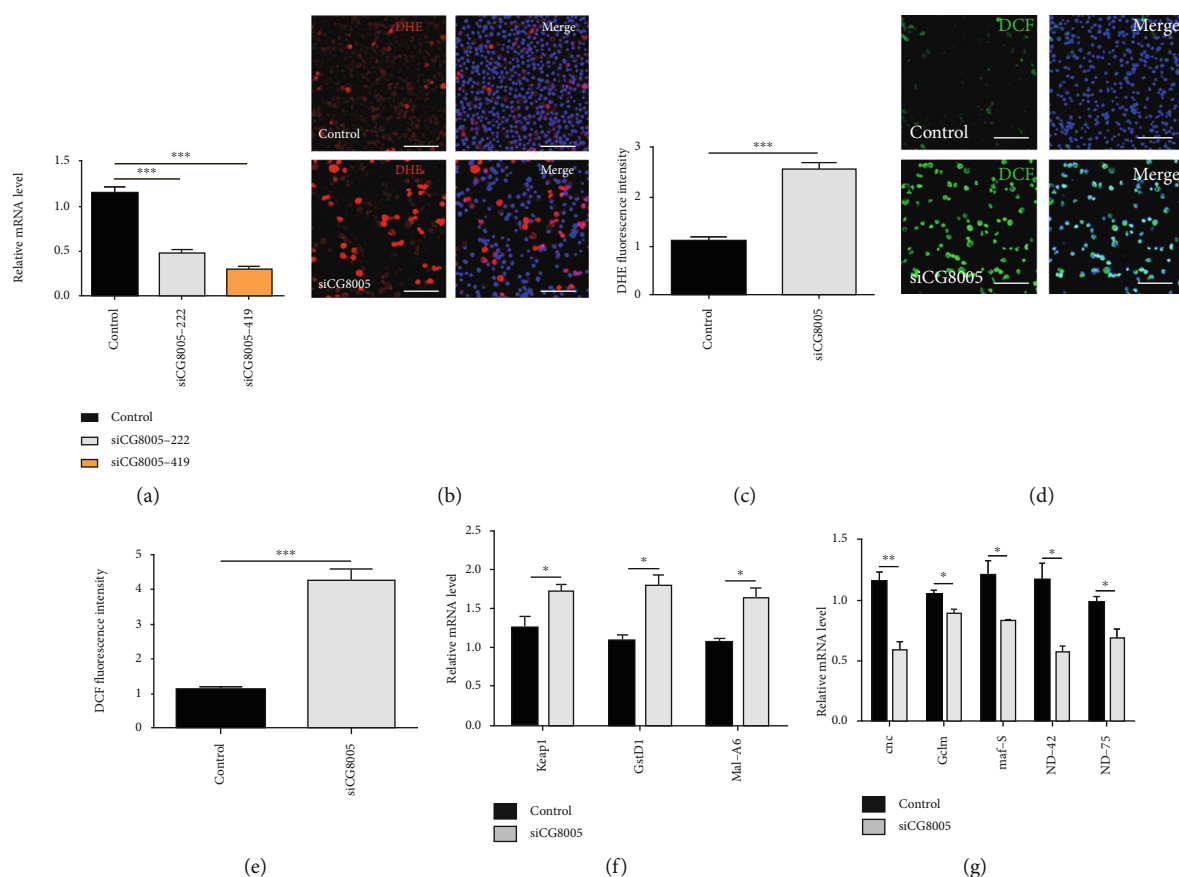


FIGURE 3: Knockdown of CG8005 induced ROS accumulation in S2 cells. (a) RNAi efficiency verification of CG8005 in control and siCG8005 (siCG8005-222 and siCG8005-419) S2 cells. (b) DHE staining in control and siCG8005 S2 cells. (c) DHE fluorescence intensity in control and siCG8005 S2 cells. (d) DCF staining in control and siCG8005 S2 cells. (e) DCF fluorescence intensity in control and siCG8005 S2 cells. (f) Relative mRNA levels of oxidation-promoting factors (Keap1, GstD1, and Mal-A6) in control and siCG8005 S2 cells. (g) Relative mRNA levels of antioxidant factors (cnc, Gclm, maf-S, ND-42, and ND-75) in control and siCG8005 S2 cells. \* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$ . Scale bar: 30  $\mu\text{m}$ .

**3.3. CG8005 Inhibited ROS Levels in S2 Cells.** To further analyze the function of CG8005 by an in vitro approach, we used two small interfering RNAs (siCG8005-222 and siCG8005-419) to silence the CG8005 gene. The relative mRNA expression level of CG8005 was examined in S2 cells, and the results showed that the knockdown efficiency of siCG8005-419 was the highest and most stable at a concentration of 150 nM (Figure 3(a)). Since ROS levels were upregulated in CG8005 RNAi testes, we further tested ROS production by using both DHE and DCF probes, and the results demonstrated that both DHE and DCF fluorescence intensities were dramatically increased in siCG8005-treated S2 cells (Figures 3(b)–3(e)). Furthermore, we detected the relative mRNA levels of oxidative stress-related factors in S2 cells and found a significant increase in the mRNA expression of oxidation-promoting factors Keap1, GstD1, and Mal-A6 and a significant decrease in the mRNA expression of antioxidant factors cnc, Gclm, maf-S, ND-42, and ND-75, as validated by qRT-PCR in control and siCG8005-treated S2 cells (Figures 3(f) and 3(g)). Together, these results supported the notion that CG8005 mediated ROS levels via oxidative stress signals.

**3.4. Antioxidant Treatments with NAC Inhibited siCG8005-Mediated ROS Accumulation in S2 Cells.** NAC is an antioxidant commonly used to inhibit intracellular ROS production. Under the antioxidant treatments with NAC, ROS production, as detected by DHE and DCF probes, was greatly decreased with dosage-dependent effects in S2 cells (Figures 4(a)–4(d)). Next, we investigated the effects of NAC on siCG8005-treated S2 cells. Pretreatment with dose-dependent NAC eliminated the accumulation of ROS induced by the siCG8005-mediated oxidative stress imbalance, and antioxidant treatment with NAC at a final concentration of 2.5 mM was sufficient to recover the ROS levels in siCG8005-treated S2 cells (Figures 4(e)–4(h)). These results indicated that NAC prevented the imbalance of intracellular ROS attributed to CG8005 knockdown in S2 cells.

**3.5. Pretreatment with  $\text{H}_2\text{O}_2$  Aggravated siCG8005-Induced ROS Production in S2 Cells.** We investigated the effects of pretreatment with  $\text{H}_2\text{O}_2$  using DHE and DCF probes in siCG8005-treated S2 cells. First, pretreatment with  $\text{H}_2\text{O}_2$  (at 100  $\mu\text{M}$  and 200  $\mu\text{M}$ ) elevated DHE and DCF fluorescence intensities, reflecting ROS dosage-dependent effects in S2 cells (Figures 5(a)–5(d)). Moreover, pretreatment with

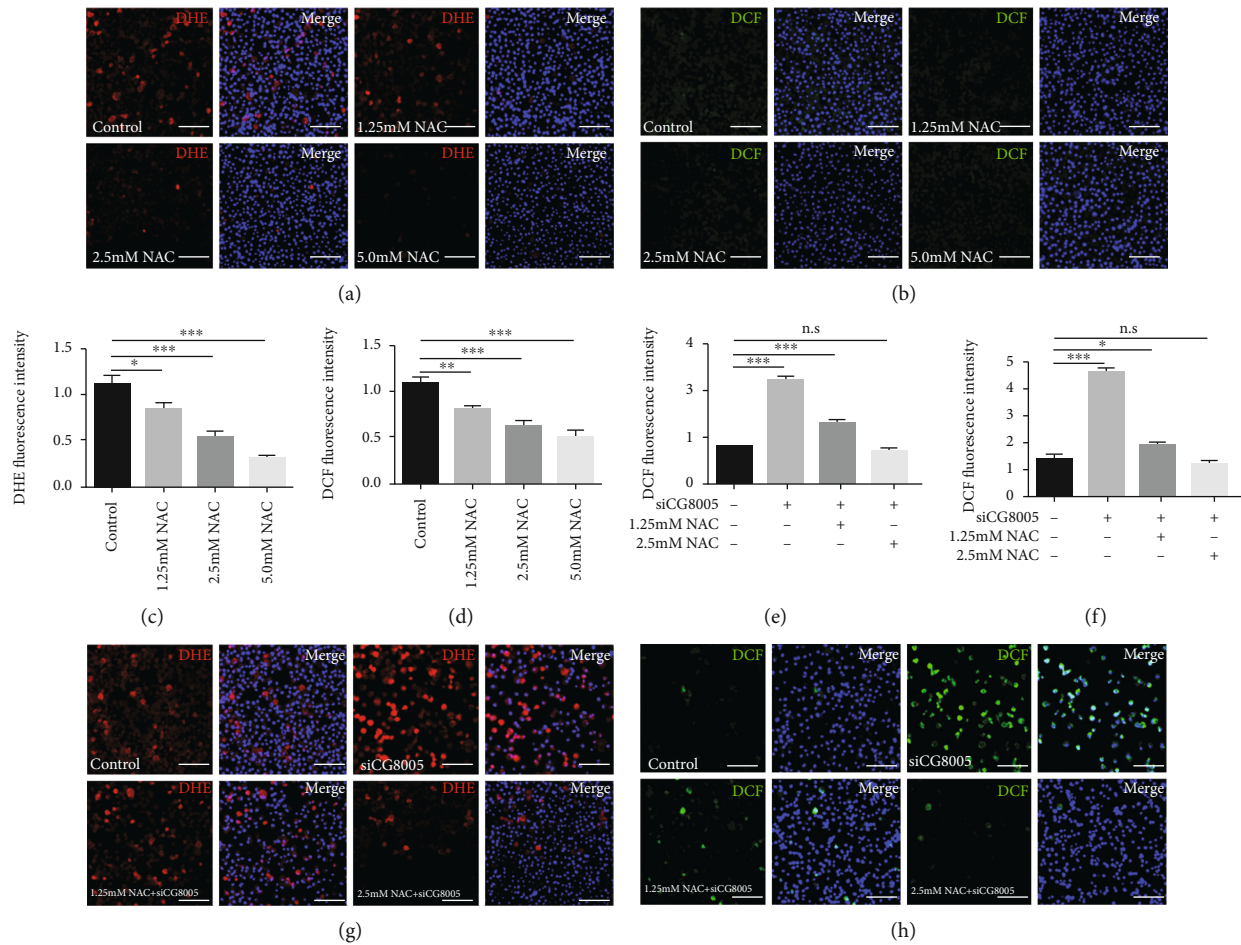


FIGURE 4: Pretreatment with NAC in siCG8005-mediated S2 cells. (a, b) DHE (a) and DCF (b) staining in control, 1.25 mM NAC, 2.5 mM NAC, and 5.0 mM NAC-treated S2 cells. (c, d) DHE (c) and DCF (d) fluorescence intensities in control, 1.25 mM NAC-, 2.5 mM NAC-, and 5.0 mM NAC-treated S2 cells. (e, f) DHE (e) and DCF (f) fluorescence intensities of pretreatment with NAC in siCG8005-treated S2 cells. (g, h) DHE (g) and DCF (h) staining of pretreatment with NAC in siCG8005-treated S2 cells. \* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$ . Scale bar: 30  $\mu\text{m}$ .

dose-dependent  $\text{H}_2\text{O}_2$  increased the accumulation of ROS and significantly elevated the fluorescence intensity of DHE and DCF in siCG8005-treated S2 cells (Figures 5(e)–5(h)). Together, these data suggested that pretreatment of  $\text{H}_2\text{O}_2$  in S2 cells could not reduce and instead further aggravated oxidative stress induced by the silencing of CG8005.

**3.6. Antioxidant Factor *cnc* Is Required for TA Spermatogonial Divisions and ROS Production in *Drosophila* Testes.** The Keap1-*cnc* signaling pathway is highly conserved in a variety of cells and tissues as an antioxidant defense. Previous studies have indicated that *cnc* is an antioxidant factor that interacts with the product of Keap1 to regulate oxidative stress. To determine the role of *cnc*, we used two independent *cnc* RNAi, UAS-*cnc* RNAi (1) and UAS-*cnc* RNAi (2), driven by Bam-Gal4 to knock down *cnc* in spermatogonia. As predicted, the knockdown of *cnc* in spermatogonia did not affect the survival of cyst cells (Figure 6(a)). To explore the effects of *cnc* on spermatogonial differentiation, we stained testes with DNA and Vasa and found that undifferentiated germ cells accumulated in both

Bam>*cnc* RNAi and Bam>*cnc* RNAi; $\Delta 86/+$  testes (Figures 6(b) and 6(c)). Moreover, the degree of differentiation defects was strengthened by heterozygous mutation of Bam ( $\Delta 86/+$ ) in Bam>*cnc* RNAi testes (Figures 6(b) and 6(c)). To further investigate the functions of CG8005 on redox balance, we assessed ROS production using a DHE probe. Interestingly, we found that ROS levels were dramatically upregulated in both Bam>*cnc* RNAi and Bam>*cnc* RNAi; $\Delta 86/+$  testes compared with the WT testes (Figures 6(d) and 6(e)), which mimicked the phenotype of CG8005 in *Drosophila* testes. Taken together, these data indicated that *cnc* contributed to controlling spermatogonial differentiation via ROS in *Drosophila* testes.

**3.7. Antioxidant Factor *cnc* Regulated Oxidative Stress in S2 Cells.** To further analyze the roles of *cnc* in S2 cells, we silenced the *cnc* gene using two siRNAs (sicnc-725 and sicnc-61) and qRT-PCR was used to verify the interference efficiency (Figure 7(a)). Additionally, the mRNA expression of antioxidant factors maf-S, ND-42, and ND-75 was significantly decreased, and the mRNA expression of oxidation-

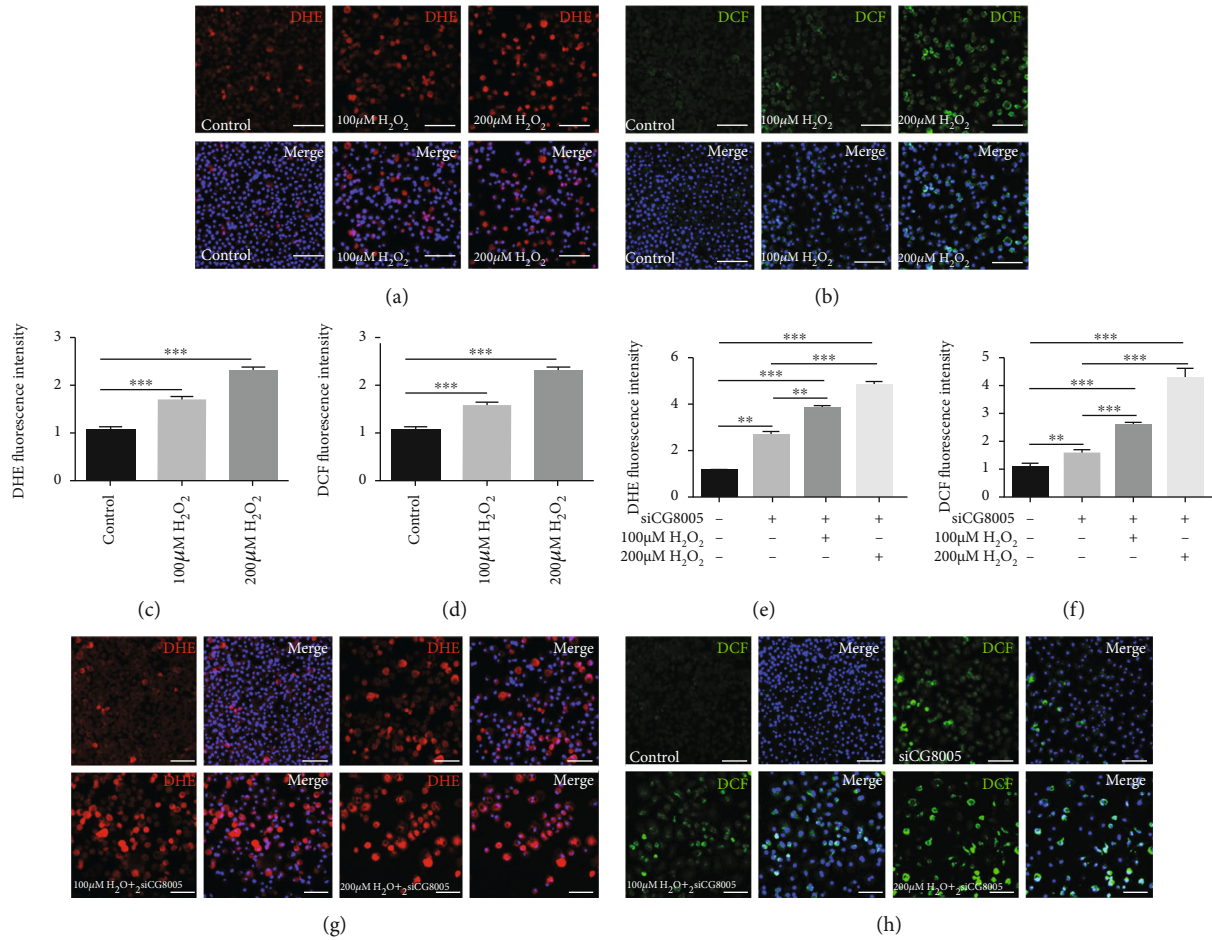


FIGURE 5: Pretreatment with H<sub>2</sub>O<sub>2</sub> in siCG8005-mediated S2 cells. (a, b) DHE (a) and DCF (b) staining in control, 100 μM H<sub>2</sub>O<sub>2</sub>-treated, and 200 μM H<sub>2</sub>O<sub>2</sub>-treated S2 cells. (c, d) DHE (c) and DCF (d) fluorescence intensities in control, 100 μM H<sub>2</sub>O<sub>2</sub>-treated, and 200 μM H<sub>2</sub>O<sub>2</sub>-treated S2 cells. (e, f) DHE (e) and DCF (f) fluorescence intensities of pretreatment with H<sub>2</sub>O<sub>2</sub> in siCG8005-treated S2 cells. (g, h) DHE (g) and DCF (h) staining of pretreatment with H<sub>2</sub>O<sub>2</sub> in siCG8005-treated S2 cells. \*\*P < 0.01 and \*\*\*P < 0.001. Scale bar: 30 μm.

promoting factors Keap1, GstD1, and Mal-A6 was dramatically increased in sicnc-treated S2 cells (Figures 7(b) and 7(c)).

We also used DHE and DCF probes to detect the state of oxidative stress and found that ROS production was increased in sicnc-treated S2 cells (Figures 7(d)–7(g) and Figure S2). Since cnc is essential for spermatogonial differentiation via ROS generation, we evaluated antioxidant and oxidant treatment in sicnc-treated S2 cells. Pretreatment with dose-dependent NAC partially reduced the accumulation of ROS in sicnc-treated S2 cells (Figures 7(d) and 7(e) and Figures S2(a) and S2(b)). Moreover, pretreatment with dose-dependent H<sub>2</sub>O<sub>2</sub> increased ROS levels and significantly elevated the fluorescence intensity of DHE and DCF in sicnc-treated S2 cells (Figures 7(f) and 7(g) and Figures S2(c) and S2(d)). Taken together, these results demonstrated that the antioxidant factor cnc was essential for redox homeostasis.

#### 4. Discussion

In *Drosophila* testes, hub cells maintain the normal behavior of GSCs and CySCs in the niche environment, and GSCs possess

strong self-renewal and differentiation abilities [29]. To produce sufficient differentiated progeny for germline homeostasis, the gonialblast undergoes multiple rounds of TA divisions prior to terminal differentiation [4, 30]. TA spermatogonial divisions are particularly important for normal spermatogenesis, and failure to exit the TA divisions may cause overgrowth during spermatogonial differentiation [6]. A previous report has shown that the mutated CG32364 gene (designated as tut), which encodes a putative RNA-binding protein, acts alongside Bam and Bgcn to form the Tut-Bam-Bgcn complex and represses the translation of mei-P26 mRNA to effectively regulate the balance of proliferation and differentiation in TA cells [31]. CG8005 is identified as a regulator of GSCs in *Drosophila* testes, although its mechanisms have yet to be revealed. In this study, we provided evidence for the significant roles and mechanisms of CG8005 and cnc during TA spermatogonial divisions. Our data suggested that loss of CG8005 and cnc in spermatogonia resulted in the accumulation of undifferentiated germ cells. These differentiation defects were further enhanced by heterozygous mutation of the differentiation factor Bam, indicating that both CG8005 and cnc were involved in regulating spermatogonial TA divisions.

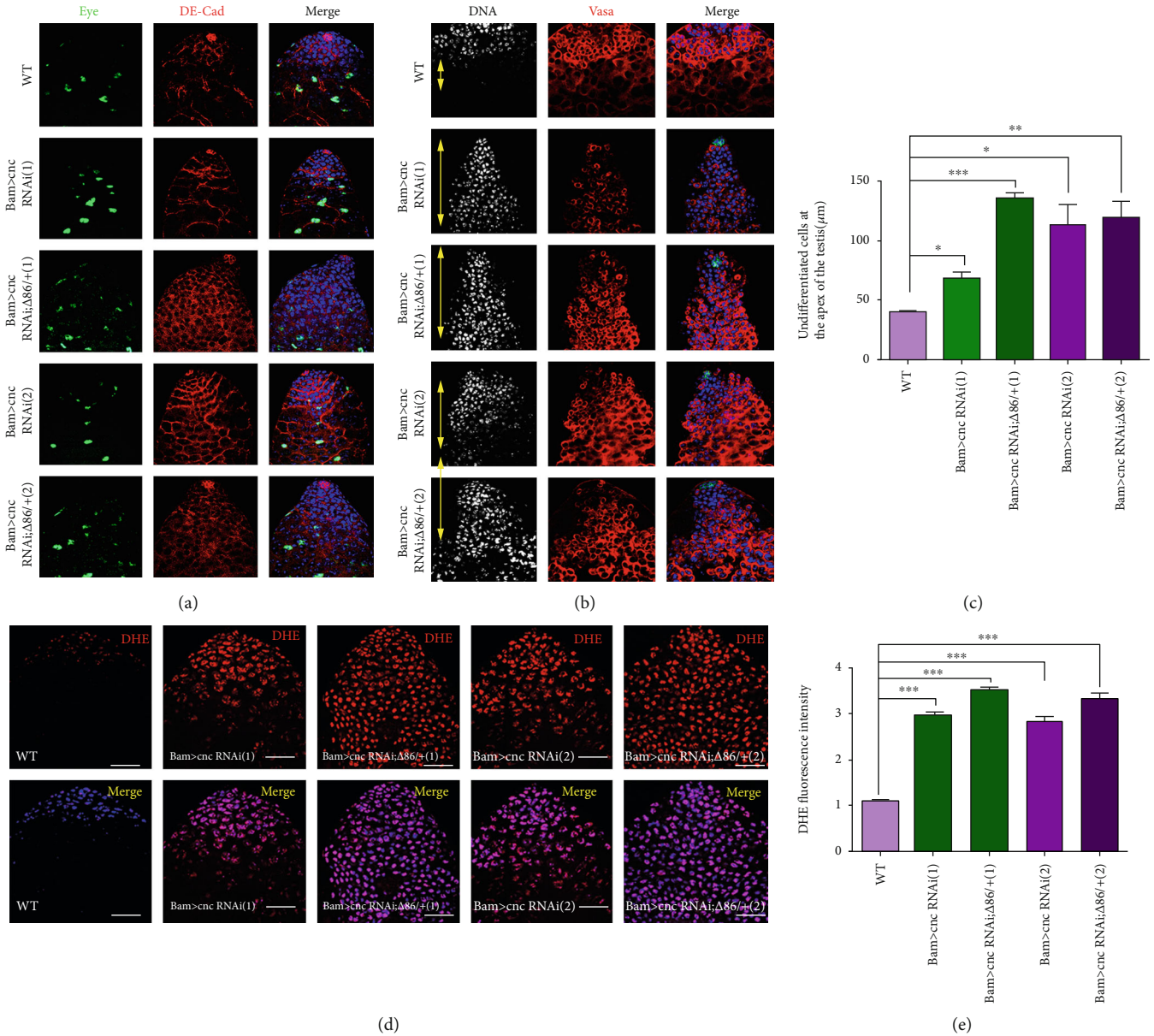


FIGURE 6: Knockdown of *cnc* in spermatogonia led to differentiation defects and ROS accumulation in *Drosophila* testes. (a, b) Immunostaining of WT, Bam>*cnc* RNAi, and Bam>*cnc* RNAi;Δ86/+ testes. Two independent UAS-*cnc* RNAi lines were used for the functional analysis of *cnc*. Yellow double arrows label undifferentiated cells at the apex of the testis. (c) The distance of undifferentiated cells at the apex of the testis. (d) DHE staining in WT, Bam>*cnc* RNAi, and Bam>*cnc* RNAi;Δ86/+ testes. (e) DHE fluorescence intensity of WT, Bam>*cnc* RNAi, and Bam>*cnc* RNAi;Δ86/+ testes. \* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$ . Scale bar: 20 μm.

Oxidative stress is involved in various human diseases such as neurodegenerative diseases, cardiovascular diseases, type II diabetes, and cancer [32–34]. The proliferation, differentiation, and division of various stem cell populations are interfered with by the disruption of redox homeostasis [13, 35]. High levels of ROS have been found in a variety of tissue damage, and oxidative stress has been reported to be necessary for both stem cell maintenance and proliferation [36]. CG8005, a potential deoxythreonate synthase, catalyzed the NAD-dependent oxidative cleavage of spermidine and interacted with eEF5 protein, which is involved in cell cycle progression, mRNA decay, the stress response,

and maintenance of the cell wall integrity (<http://flybase.org>). The potential role of many antioxidants, such as NAC, to improve ROS-related stem cell self-renewal and differentiation disorders has been studied [37, 38].  $H_2O_2$  can exhibit its effect on cells through oxidative stress, which causes cells to gradually lose their ability to proliferate and eventually leads to cell death [39, 40]. In this study, knockdown of CG8005 and *cnc* induced high levels of ROS generation, which were eliminated by pretreatment with NAC or enhanced by pretreatment with  $H_2O_2$ . We also found that the antioxidants CG8005 and *cnc* could regulate the expression of oxidative and antioxidative factors.



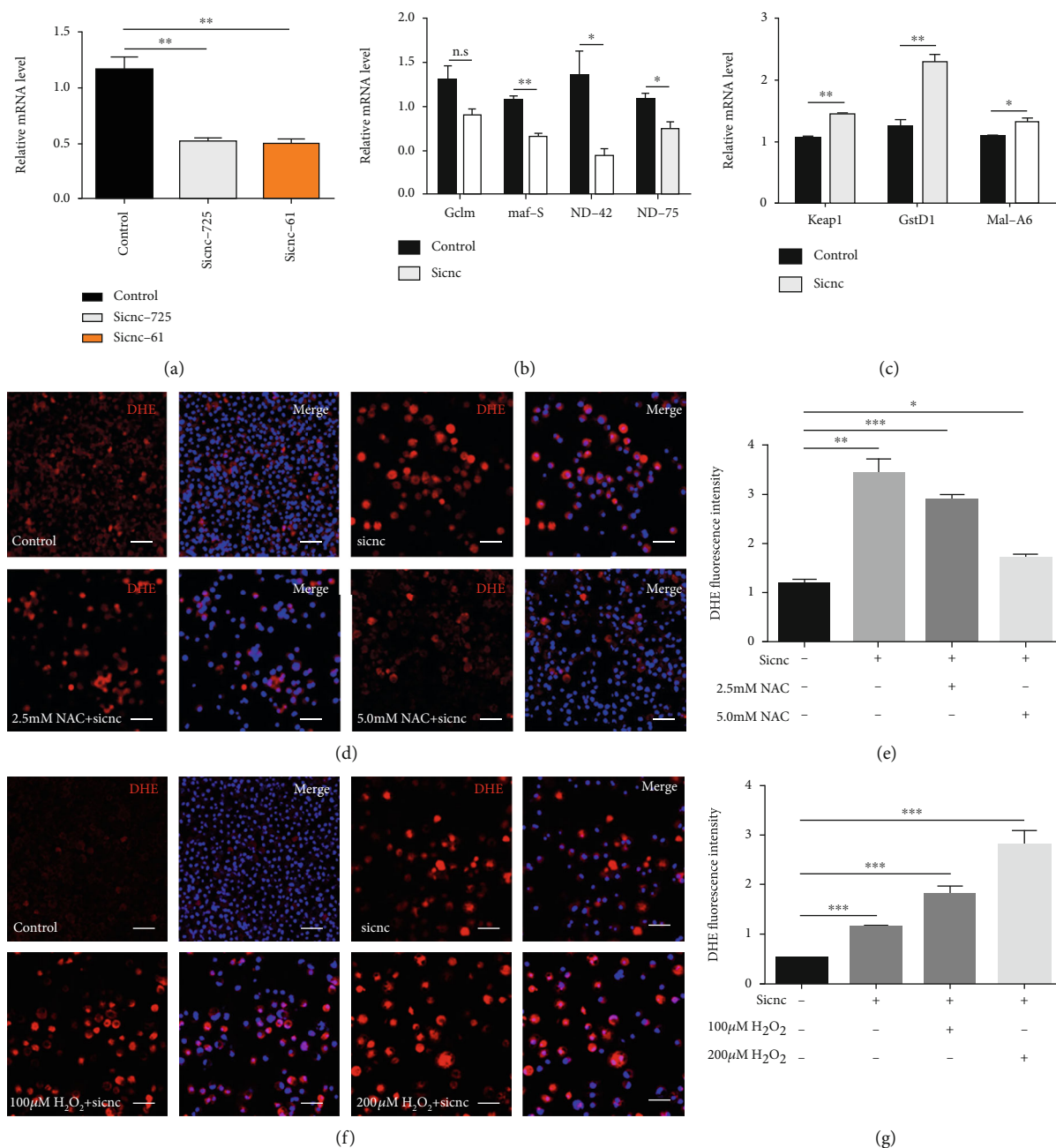


FIGURE 7: Effects of ROS accumulation in sicnc-treated S2 cells. (a) RNAi efficiency verification of cnc in control and cnc-siRNA (sicnc-725 and sicnc-61) S2 cells. (b) Relative mRNA levels of antioxidant factors (Gclm, maf-S, ND-42, and ND-75) in control and sicnc S2 cells. (c) Relative mRNA levels of oxidation-promoting factors (Keap1, GstD1, and Mal-A6). (d, e) DHE staining (d) and DHE fluorescence intensity (e) of pretreatment with NAC in sicnc-treated S2 cells. (f, g) DHE staining (f) and DHE fluorescence intensity (g) of pretreatment with H<sub>2</sub>O<sub>2</sub> in sicnc-treated S2 cells. \**P* < 0.05, \*\**P* < 0.01, and \*\*\**P* < 0.001. Scale bar: 30 μm.

## 5. Conclusion

In summary, the present study showed that CG8005 participated in the switch from self-renewal to differentiation during TA spermatogonial divisions and regulated oxidative stress via the Keap1-cnc signaling pathway. Pretreatment with NAC decreased ROS accumulation in CG8005- and cnc-siRNA-mediated S2 cells, while the opposite effect

occurred in H<sub>2</sub>O<sub>2</sub>-treated S2 cells. The findings of this study provide new insights for understanding the relationships between TA spermatogonial divisions and oxidative stress.

## Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

## Conflicts of Interest

The authors declare that they have no competing interests.

## Authors' Contributions

The manuscript was written through the contributions of all authors. All authors have given approval to the final version of the manuscript.

## Acknowledgments

The authors wish to thank all study participants, research staff, and students who assisted with this work. We would like to thank Chao Tong (Life Sciences Institute, Zhejiang University, Zhejiang, China) and DH Chen (Institute of Zoology, Chinese Academy of Sciences, Beijing, China) for sharing reagents and stocks. This work was supported by the Young Talents Training Program of Jiangsu University (5521470000) and the Key Research Foundation of Zhenjiang Social Development (SH2018065).

## Supplementary Materials

Figure S1: proliferation analysis in CG8005 RNAi testes. Figure S2: effects of pretreatment with NAC and H<sub>2</sub>O<sub>2</sub> in cnc-siRNA-mediated S2 cells. Table S1: siRNA sequences used in this study. Table S2: primer sequences used in this study. (*Supplementary Materials*)

## References

- [1] D. S. Chen, X. X. Zhu, L. J. Zhou et al., "Gilgamesh is required for the maintenance of germline stem cells in *Drosophila* testes," *Scientific Reports*, vol. 7, no. 1, p. 5737, 2017.
- [2] M. Amoyel, J. Anderson, A. Suisse, J. Glasner, and E. A. Bach, "Socs36e controls niche competition by repressing MAPK signaling in the *Drosophila* testis," *PLOS Genetics*, vol. 12, no. 1, article e1005815, 2016.
- [3] Y. C. Chang, H. Tu, J. Y. Chen, C. C. Chang, S. Y. Yang, and H. Pi, "Reproduction disrupts stem cell homeostasis in testes of aged male *Drosophila* via an induced microenvironment," *PLOS Genetics*, vol. 15, no. 7, article e1008062, 2019.
- [4] A. Carbonell, S. Pérez-Montero, P. Climent-Cantó, O. Reina, and F. Azorín, "The germline linker histone dBigH1 and the translational regulator bam form a repressor loop essential for male germ stem cell differentiation," *Cell Reports*, vol. 21, no. 11, pp. 3178–3189, 2017.
- [5] P. Joti, A. Ghosh-Roy, and K. Ray, "Dynein light chain 1 functions in somatic cyst cells regulate spermatogonial divisions in *Drosophila*," *Scientific Reports*, vol. 1, no. 1, 2011.
- [6] M. L. Insko, A. Leon, C. H. Tam, D. M. McKearin, and M. T. Fuller, "Accumulation of a differentiation regulator specifies transit amplifying division number in an adult stem cell lineage," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 52, pp. 22311–22316, 2009.
- [7] A. Glasauer and N. S. Chandel, "Targeting antioxidants for cancer therapy," *Biochemical Pharmacology*, vol. 92, no. 1, pp. 90–101, 2014.
- [8] G. J. Burton and E. Jauniaux, "Oxidative stress," *Best Practice & Research. Clinical Obstetrics & Gynaecology*, vol. 25, no. 3, pp. 287–299, 2011.
- [9] S. Khan, A. Zafar, and I. Naseem, "Redox cycling of copper by coumarin-di(2-picoyl)amine hybrid molecule leads to ROS-mediated modulation of redox scavengers, DNA damage and cell death in diethylnitrosamine induced hepatocellular carcinoma," *Bioorganic Chemistry*, vol. 99, article 103818, 2020.
- [10] S. Rodius, N. de Klein, C. Jeanty et al., "Fisetin protects against cardiac cell death through reduction of ROS production and caspases activity," *Scientific Reports*, vol. 10, no. 1, 2020.
- [11] S. W. S. Tan, Q. Y. Lee, B. S. E. Wong, Y. Cai, and G. H. Baeg, "Redox homeostasis plays important roles in the maintenance of the *Drosophila* testis germline stem cells," *Stem Cell Reports*, vol. 9, no. 1, pp. 342–354, 2017.
- [12] R. Sênos Demarco and D. L. Jones, "Mitochondrial fission regulates germ cell differentiation by suppressing ROS-mediated activation of epidermal growth factor signaling in the *Drosophila* larval testis," *Scientific Reports*, vol. 9, no. 1, 2019.
- [13] S. W. S. Tan, G. W. Yip, T. Suda, and G. H. Baeg, "Small Maf functions in the maintenance of germline stem cells in the *Drosophila* testis," *Redox Biology*, vol. 15, pp. 125–134, 2018.
- [14] Z. A. Wang, J. H. Huang, and D. Kalderon, "*Drosophila* follicle stem cells are regulated by proliferation and niche adhesion as well as mitochondria and ROS," *Nature Communications*, vol. 3, no. 1, 2012.
- [15] P. P. Kushwaha, S. Gupta, A. K. Singh, K. S. Prajapati, M. Shuaib, and S. Kumar, "MicroRNA targeting nicotinamide adenine dinucleotide phosphate oxidases in cancer," *Antioxidants & Redox Signaling*, vol. 32, no. 5, pp. 267–284, 2020.
- [16] N. Dominado, J. E. la Marca, N. A. Siddall et al., "Rbf regulates *Drosophila* spermatogenesis via control of somatic stem and progenitor cell fate in the larval testis," *Stem Cell Reports*, vol. 7, no. 6, pp. 1152–1163, 2016.
- [17] E. Kawase, M. D. Wong, B. C. Ding, and T. Xie, "Gbb/Bmp signaling is essential for maintaining germline stem cells and for repressing bam transcription in the *Drosophila* testis," *Development*, vol. 131, no. 6, pp. 1365–1375, 2004.
- [18] S. M. Bunt and G. R. Hime, "Ectopic activation of dpp signaling in the male *Drosophila* germline inhibits germ cell differentiation," *Genesis*, vol. 39, no. 2, pp. 84–93, 2004.
- [19] A. A. Shivdasani and P. W. Ingham, "Regulation of stem cell maintenance and transit amplifying cell proliferation by TGF- $\beta$  signaling in *Drosophila* spermatogenesis," *Current Biology*, vol. 13, no. 23, pp. 2065–2072, 2003.
- [20] S. H. Eun, P. M. Stoiber, H. J. Wright et al., "MicroRNAs downregulate bag of marbles to ensure proper terminal differentiation in the *Drosophila* male germline," *Development*, vol. 140, no. 1, pp. 23–30, 2012.
- [21] P. Gönczy, E. Matunis, and S. DiNardo, "Bag-of-marbles and benign gonial cell neoplasm act in the germline to restrict proliferation during *Drosophila* spermatogenesis," *Development*, vol. 124, no. 21, pp. 4361–4371, 1997.
- [22] J. Yu, X. Lan, X. Chen et al., "Protein synthesis and degradation are essential to regulate germline stem cell homeostasis in *Drosophila* testes," *Development*, vol. 143, no. 16, pp. 2930–2945, 2016.
- [23] X. Kong, G. D. Wang, M. Z. Ma et al., "Sesamin ameliorates advanced glycation end products-induced pancreatic  $\beta$ -cell dysfunction and apoptosis," *Nutrients*, vol. 7, no. 6, pp. 4689–4704, 2015.

- [24] R. Safi, J. Al-Hage, O. Abbas, A. G. Kibbi, and D. Nassar, "Investigating the presence of neutrophil extracellular traps in cutaneous lesions of different subtypes of lupus erythematosus," *Experimental Dermatology*, vol. 28, no. 11, pp. 1348–1352, 2019.
- [25] X. J. Luan, Y. D. Yan, Q. W. Zheng et al., "Excessive reactive oxygen species induce apoptosis via the APPL1-Nrf2/HO-1 antioxidant signalling pathway in trophoblasts with missed abortion," *Life Sciences*, vol. 254, article 117781, 2020.
- [26] M. Inaba, M. Buszczak, and Y. M. Yamashita, "Nanotubes mediate niche-stem-cell signalling in the *Drosophila* testis," *Nature*, vol. 523, no. 7560, pp. 329–332, 2015.
- [27] X. Chen, X. J. Luan, Q. W. Zheng et al., "Precursor RNA processing 3 is required for male fertility, and germline stem cell self-renewal and differentiation via regulating spliceosome function in *Drosophila* testes," *Scientific Reports*, vol. 9, no. 1, p. 9988, 2019.
- [28] C. L. Bigarella, R. Liang, and S. Ghaffari, "Stem cells and the impact of ROS signaling," *Development*, vol. 141, no. 22, pp. 4206–4218, 2014.
- [29] J. Yu, X. J. Luan, Y. D. Yan et al., "Small ribonucleoprotein particle protein SmD3 governs the homeostasis of germline stem cells and the crosstalk between the spliceosome and ribosome signals in *Drosophila*," *The FASEB Journal*, vol. 33, no. 7, pp. 8125–8137, 2019.
- [30] S. Gupta, B. Varshney, S. Chatterjee, and K. Ray, "somatic ERK activation during transit amplification is essential for maintaining the synchrony of germline divisions in *Drosophila* testes," *Open Biology*, vol. 8, no. 7, article 180033, 2018.
- [31] D. Chen, C. Wu, S. W. Zhao et al., "Three RNA binding proteins form a complex to promote differentiation of germline stem cell lineage in *Drosophila*," *PLOS Genetics*, vol. 10, no. 11, p. e1004797, 2014.
- [32] K. Hakvoort, L. Otto, R. Haeren et al., "Shedding light on human cerebral lipofuscin: an explorative study on identification and quantification," *Journal of Comparative Neurology*, 2020.
- [33] G. Aykutoglu, M. Tartik, E. Darendelioglu, A. Ayna, and G. Baydas, "Melatonin and vitamin E alleviate homocysteine-induced oxidative injury and apoptosis in endothelial cells," *Molecular Biology Reports*, vol. 47, no. 7, pp. 5285–5293, 2020.
- [34] N. Giribabu, K. Karim, E. K. Kilari, S. R. Nelli, and N. Salleh, "Oral administration of *Centella asiatica* (L.) Urb leave aqueous extract ameliorates cerebral oxidative stress, inflammation, and apoptosis in male rats with type-2 diabetes," *Inflammopharmacology*, 2020.
- [35] J. Tower, "Stress and stem cells," *Wiley Interdisciplinary Reviews. Developmental Biology*, vol. 1, no. 6, pp. 789–802, 2012.
- [36] C. W. Xu, J. J. Luo, L. He, C. Montell, and N. Perrimon, "Oxidative stress induces stem cell proliferation via TRPA1/RyR-mediated Ca<sup>2+</sup> signaling in the *Drosophila* midgut," *eLife*, vol. 6, article e22441, 2017.
- [37] K. Livingston, R. A. Schlaak, L. L. Puckett, and C. Bergom, "The role of mitochondrial dysfunction in radiation-induced heart disease: from bench to bedside," *Frontiers in Cardiovascular Medicine*, vol. 7, 2020.
- [38] M. Steyn, K. Zitouni, F. J. Kelly, P. Cook, and K. A. Earle, "Sex differences in glutathione peroxidase activity and central obesity in patients with type 2 diabetes at high risk of cardiovascular disease," *Antioxidants*, vol. 8, no. 12, p. 629, 2019.
- [39] A. Čipak Gašparović, L. Milković, N. Dandachi et al., "Chronic oxidative stress promotes molecular changes associated with epithelial mesenchymal transition, nrf2, and breast cancer stem cell phenotype," *Antioxidants*, vol. 8, no. 12, p. 633, 2019.
- [40] E. Radi, P. Formichi, C. Battisti, and A. Federico, "Apoptosis and oxidative stress in neurodegenerative diseases," *Journal of Alzheimer's Disease*, vol. 42, supplement 3, pp. S125–S152, 2014.