

Original Article

# Toxicity of humidifier disinfectant polyhexamethylene guanidine hydrochloride by two-week whole body-inhalation exposure in rats

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**Abstract:** The use of polyhexamethylene guanidine hydrochloride (PHMG·HCl) as a humidifier disinfectant caused an outbreak of pulmonary disease, leading to the deaths of pregnant women and children in South Korea. However, limited information is available on the inhalation toxicity of PHMG·HCl. Therefore, this study aimed to characterize the subacute inhalation toxicity of PHMG·HCl by whole-body exposure in rats. F344 rats were exposed to 0 mg/m<sup>3</sup>, 1 mg/m<sup>3</sup>, 5 mg/m<sup>3</sup>, or 25 mg/m<sup>3</sup> of PHMG·HCl for 6 h/day, 5 days/week for two weeks via whole-body inhalation. Emaciation and rale were observed in rats in the 25 mg/m<sup>3</sup> PHMG·HCl group. Significant changes in body weight, hematology, serum chemistry and organ weight were observed in all PHMG·HCl-exposed groups. Gross lesions showed ballooning or red focus in the lungs of rats in the PHMG·HCl-exposed groups. In histopathological examination, most of histological lesions (including degeneration, atrophy, ulcer, inflammatory cell infiltration, inflammation, and fibrosis in nasal cavity, larynx, trachea, and lungs) indicated tissue damage by PHMG·HCl in all PHMG·HCl-exposed groups. Additionally, atrophy of the spleen, thymus, and reproductive organs; immaturity of the testes; and cell debris in the epididymides were affected by the reduction in body weight in PHMG·HCl-exposed groups. In conclusion, two-week repeated whole-body inhalation exposure of rats to PHMG·HCl revealed toxic effects on the respiratory system and secondary effects on other organs. The results of this study indicate that the no observable adverse effect level (NOAEL) for PHMG·HCl is below 1 mg/m<sup>3</sup>. (DOI: 10.1293/tox.2020-0043; J Toxicol Pathol 2020; 33: 265–277)

**Key words:** polyhexamethylene guanidine hydrochloride (PHMG·HCl), subacute inhalation toxicity, humidifier disinfectants

## Introduction

Polyhexamethylene guanidine (PHMG) is a derivative of the polymeric guanidine family and known to be a potent bactericide, virucide, and fungicide<sup>1–3</sup>. It is colorless, odorless, and non-corrosive<sup>4</sup>, and highly soluble in water<sup>5</sup>. PHMG has been widely used in fabric softeners, paints, detergents, and swimming pools and especially, as household humidifier disinfectants in South Korea<sup>3, 5–7</sup>. In 2011, mist from humidifier disinfectants caused an outbreak of pulmonary disease, which leads to the deaths of pregnant women and children in South Korea<sup>7</sup>. The pulmonary injury of the patients was confirmed as acute interstitial pneumonia and fibrosis<sup>8</sup>, reported to be mainly associated with users of humidifier disinfectants containing PHMG-phosphate (PHMG·P), oligo (2-(2-ethoxy) ethoxyethyl guanidinium (PGH), a mixture of chloromethyl-

isothiazolinone and methylisothiazolinone (CMIT/MIT)<sup>7</sup>. However, to a lesser extent, another derivative of PHMG, PHMG hydrochloride (PHMG·HCl) was also involved in the outbreak<sup>8</sup>.

The toxicity of PHMG·HCl has been rarely documented in humans or *in vitro* and *in vivo* tests. PHMG·HCl has been reported to have low toxicity to humans<sup>9, 10</sup>. However, more than 12,500 patients in Russia who drank illegally manufactured vodka with 0.10–0.14% PHMG·HCl were reported to have suffered from acute cholestatic hepatitis<sup>11, 12</sup>. PHMG·HCl induces cellular toxicity through the production of intracellular reactive oxygen species (ROS) and gene expression profile alteration resulting in the progression to cell death and the down-regulation of antioxidants and detoxifying enzymes in human alveolar epithelial A549 cells<sup>13</sup>. Acute oral toxicity studies have shown that the median lethal dose (LD50) of 600 mg/kg is accompanied by signs of neurotoxicity, but a dose of 0.036 mg/kg is not associated with mortality or clinical signs of toxicity even though mid hepatocyte degeneration and tubular hydropic change may be observed<sup>14</sup>.

Only a few studies so far have reported so far on the oral toxicity of PHMG·HCl and the inhalation toxicity of PHMG·HCl has not yet been confirmed in the outbreak of humidifier disinfectant. Therefore, this study aimed to characterize the subacute inhalation toxicity of PHMG·HCl by

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whole-body exposure in rats.

## Materials and Methods

### *Generation, Analysis, and Inhalation Chamber Monitoring*

PHMG-HCl was obtained from Beyond Industry Co., Ltd. (Shanghai, China) through the Ministry of Environment. Filtered tap water was used as vehicle. PHMG-HCl was dissolved in water as 0.2 and 0.5% (w/v).

The PHMG-HCl aerosol was generated using an ultrasonic mist-generator in a whole body chamber (Chamber volume: 1 m<sup>3</sup>, SIS-20RG, Shibata, Saitama, Japan). The phase of PHMG-HCl aerosol was produced as mist in the inhalation chamber. The concentration of PHMG-HCl was measured using a personal sampler (Airchek XR 5000, SKC Inc., Eighty Four, PA, USA) with 25-mm micro glass fiber filters (Pallfex Membrane Filters, Pall Co., Charlotte, NC, USA). The mass median aerodynamic diameter (MMAD) and geometric standard deviation (GSD) were measured using a cascade impactor (nanoMOUDI Impactor, MSP Co., Shoreview, MN, USA). Samples were collected from the middle part of the port at a flow rate of 1 L/min. During the exposure period, the mass concentrations of the aerosols in the chamber were measured at least three times daily. The total airflow for each chamber was set at 20 L/min to achieve 1 L/min flow/rat. Chamber conditions including temperature, relative humidity, pressure, and air flow rate were automatically measured (ICS-20RG).

### *Animal husbandry and maintenance*

Six-week-old specific-pathogen-free F344 rats of both sexes were purchased from Japan SLC, Inc. (Shizuoka, Japan) and acclimated for one week. The room was maintained at a temperature of 22 ± 3°C, relative humidity of 50 ± 20%, 12:12 h light:dark cycle, and fresh air ventilation (10–15 changes per hour). Rats were housed singly in stainless steel wire mesh cages (W 220 mm × L 750 mm × H 180 mm) and had free access to UV-irradiated rodent pellet diet (Teklad Global 18% Protein Rodent Diet, Harlan Laboratories, Inc., Indianapolis, IN, USA) and filtered tap water. The animal protocol was approved by the Institutional Animal Care and Use Committee at Occupational Safety and Health Research Institute (IACUC-1718).

### *Experimental design*

A total of 40 rats (20 males and 20 females) were assigned randomly to one of four groups (5 per sex per group; 0 mg/m<sup>3</sup>, 1 mg/m<sup>3</sup>, 5 mg/m<sup>3</sup>, or 25 mg/m<sup>3</sup>) and exposed for 6 h/day, 5 days/week for two weeks. The PHMG-HCl concentrations used were selected on the basis of an acute toxicity study performed previously (data not shown) using a scale factor of three. Exposures were conducted in accordance with test No. 412 (Subacute Inhalation Toxicity, 2009) by the Organization for Economic Co-operation and Development (OECD)<sup>15</sup>. Inhalation exposures were conducted from 10:00 to 16:00. All rats were euthanized after two weeks of

inhalation.

### *Clinical observations and body weight*

All animals were examined twice daily for mortality and clinical signs, and weighed individually on day 1, 3, 6, and 13 of inhalation exposure.

### *Hematology and serum biochemistry*

In hematology, all animals were fasted overnight before necropsy and blood collection. Blood samples were taken from the abdominal aorta using a syringe with a 24-gauge needle under isoflurane anesthesia (Hana Pharm, Kyonggi-Do, Korea) and collected into vacutainers containing EDTA-2K (Becton Dickinson, Franklin Lakes, NJ, USA). The absolute or relative number in the following parameters were determined in this study: total erythrocyte (RBC), hemoglobin concentration (HGB), hematocrit (HCT), mean cell volume (MCV), mean cell hemoglobin (MCH), mean cell hemoglobin concentration (MCHC), reticulocytes (RET), platelets (PLT), whole leukocytes (WBC), neutrophils (NEU), eosinophils (EOS), basophils (BAS), lymphocytes (LYM) and monocytes (MON). In serum chemistry, blood samples were centrifuged at 1,811×g at 4°C for 10 minutes within 90 minute of collection. The following serum chemistry parameters were evaluated using an automated analyzer (TBA-120FR, Toshiba Medical Systems, Tochigi, Japan): total protein (TP), albumin (ALB), blood urea nitrogen (BUN), creatinine (CREA), alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), glucose (GLU), total cholesterol (T-CHO), and triglycerides (TG).

### *Gross examination and histopathology*

Gross examinations of organs in the cranial, thoracic, and abdominal cavities of the rats were conducted. The absolute and relative (organ-to-body) weights of the brain, lungs, heart, liver, spleen, and kidneys were measured. The following tissues were removed from each animal at necropsy: liver, kidneys, heart, brain, spleen, trachea, tracheo-bronchial lymph node, larynx, lungs and nasal cavity; in males, seminal vesicle, prostate, testes, and epididymides; in female, ovaries, uterus, and vagina. The nasal cavity was sectioned at four levels: 1, posterior to the upper incisors; 2, incisive papilla; 3, second palatine crest; and 4, first molar teeth. The organs were preserved in 10% neutral buffered formalin. All organs were embedded in paraffin, sectioned at 3–4 μm, stained with hematoxylin and eosin (H&E), and examined microscopically at low and high power fields.

### *Data analyses*

Differences among groups in the various parameters were determined using SPSS (ver. 18.0, IBM, Chicago, IL, USA) software. The homogeneity of variance was analyzed by Levene's test, followed by either one-way analysis of variance for samples with homogenous variance or the Kruskal-Wallis test for samples with heterogeneous variance. Scheffe or Dunnett's multiple range test was used to

compare the result of each experimental group with that of the control group if the first statistical result was significant.

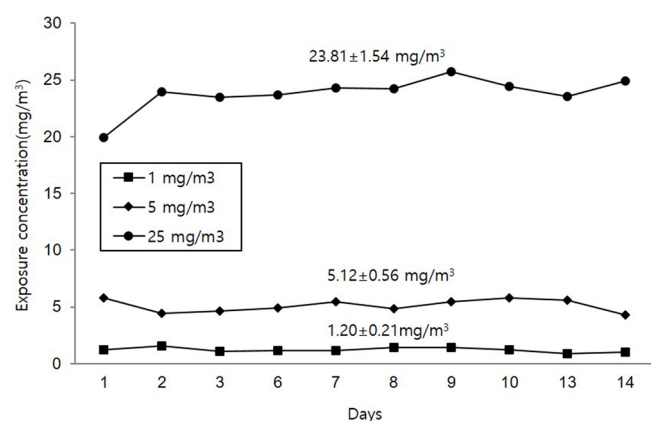
## Results

### Chamber monitoring

The ranges of chamber conditions were 19.6–21.9°C, 49.5–74.8% relative humidity, –66.8––58.1 mmH<sub>2</sub>O pressure, and 220.1–278.2 L/min flow rate. The average concentrations of PHMG·HCl during the study were 1.201 ± 0.21 mg/m<sup>3</sup>, 5.12 ± 0.56 mg/m<sup>3</sup>, and 23.81 ± 1.54 mg/m<sup>3</sup> for 1, 5, and 25 mg/m<sup>3</sup> PHMG·HCl groups, respectively (Fig. 1). The MMAD was 0.602, 0.877, and 1.073 μm, and the GSD 2.42, 1.84, and 1.66 mg/m<sup>3</sup> for 1, 5, and 25 mg/m<sup>3</sup> PHMG·HCl groups, respectively (Table 1).

### Clinical signs

No deaths were observed in any groups. Emaciation and rale were observed in males and females exposed to 25 mg/m<sup>3</sup> (Table 2).



**Fig. 1.** PHMG·HCl concentrations in the inhalation chamber during the study.

### Body weight

Body weights decreased significantly in males and female exposed to 25, 5, and 1 mg/m<sup>3</sup> from day 3, 6, and 13 onward, respectively (Fig. 2).

### Hematology

RBC, HCT, and HGB showed increasing trend or significant increases in males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>. MCHC increased significantly in males and female exposed to 25 mg/m<sup>3</sup>. In contrast, MCH and MCV decreased significantly in males exposed to 5 and 25 mg/m<sup>3</sup> and females exposed to 25 mg/m<sup>3</sup>. Additionally, RET and RET% decreased significantly in males and females exposed to 5 and 25 mg/m<sup>3</sup>. PLT showed decreasing trends or significant decrease in males and females exposed to 25 mg/m<sup>3</sup>. MON, MON%, and NEU increased significantly in males and females exposed to 25 mg/m<sup>3</sup>. NEU% increased significantly in males exposed to 25 mg/m<sup>3</sup> and females exposed to 5 and 25 mg/m<sup>3</sup>. However, LYM% showed decreasing trend or significant decrease in males exposed to 25 mg/m<sup>3</sup> and females exposed to 5 and 25 mg/m<sup>3</sup>. EOS and EOS% decreased significantly in females exposed to 25 mg/m<sup>3</sup> (Table 3).

### Serum biochemistry

ALT showed increasing trends or significant increase in males exposed to 25 mg/m<sup>3</sup> and females exposed to 5 and 25 mg/m<sup>3</sup>. Additionally, AST increased significantly in males and females exposed to 25 mg/m<sup>3</sup>. BUN showed increasing trends or significant increases in males and females

**Table 1.** Particle Size Distribution

Group	Mass median Aerodynamic diameter (μm)	Geometric standard deviation
1 mg/m <sup>3</sup>	0.602	2.42
5 mg/m <sup>3</sup>	0.877	1.84
25 mg/m <sup>3</sup>	1.073	1.66

**Table 2.** Summary of Clinical Signs

Sex	Group	No. of rats	Clinical sign	Days																	
				0	1	2	3	4	5	6	7	8	9	10	11	12	13	14			
Male	0 mg/m <sup>3</sup>	5	No observed	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5		
	1 mg/m <sup>3</sup>	5	No observed	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5		
	5 mg/m <sup>3</sup>	5	No observed	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5		
	25 mg/m <sup>3</sup>	5	No observed	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	0	0	
			Rale	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	4	4
			Emaciation	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	5	5
Female	0 mg/m <sup>3</sup>	5	No observed	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5		
	1 mg/m <sup>3</sup>	5	No observed	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5		
	5 mg/m <sup>3</sup>	5	No observed	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5		
	25 mg/m <sup>3</sup>	5	No observed	5	5	5	5	5	5	5	5	5	5	5	5	0	0	0	0	0	
			Rale	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2	3
			Emaciation	0	0	0	0	0	0	0	0	0	0	0	0	5	5	5	5	5	

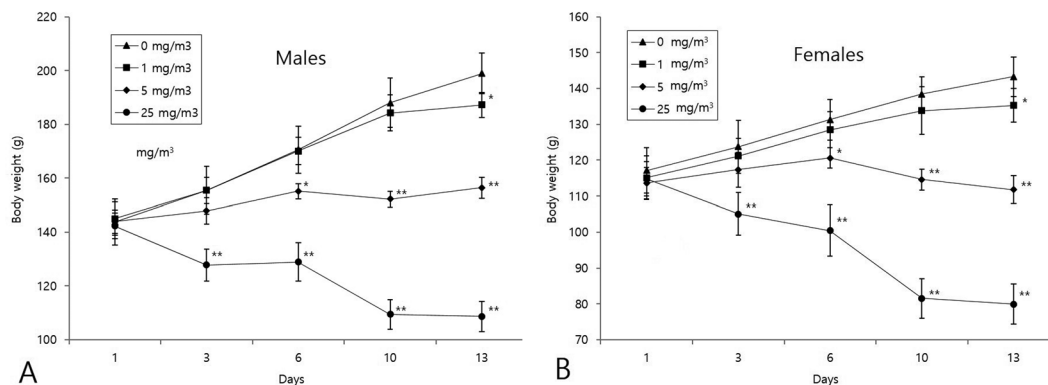


Fig. 2. Changes in body weight during the study. Significant differences compared with the control: \* $p < 0.05$ , \*\* $p < 0.01$ .

Table 3. Summary of Hematological Parameters

Sex	Group	0 mg/m <sup>3</sup>	1 mg/m <sup>3</sup>	5 mg/m <sup>3</sup>	25 mg/m <sup>3</sup>
		No. of rats	5	5	5
Male	RBC ( $\times 10^3/\mu\text{L}$ )	8.42 $\pm$ 0.32	9.18 $\pm$ 0.34**	9.87 $\pm$ 0.28**	10.34 $\pm$ 0.24**
	HGB (g/dL)	15.44 $\pm$ 0.58	16.66 $\pm$ 0.68*	17.56 $\pm$ 0.42**	18.48 $\pm$ 0.33**
	HCT (%)	46.42 $\pm$ 1.96	50.64 $\pm$ 1.72**	52.28 $\pm$ 1.39**	53.3 $\pm$ 0.66**
	MCV (fL)	55.08 $\pm$ 0.36	55.18 $\pm$ 0.48	52.96 $\pm$ 0.42**	51.54 $\pm$ 0.67**
	MCH (pg)	18.32 $\pm$ 0.08	18.18 $\pm$ 0.23	17.82 $\pm$ 0.13**	17.84 $\pm$ 0.05**
	MCHC (g/dL)	33.28 $\pm$ 0.35	32.94 $\pm$ 0.52	33.62 $\pm$ 0.29	34.64 $\pm$ 0.32**
	PLT ( $\times 10^3/\mu\text{L}$ )	921.2 $\pm$ 87.5	919 $\pm$ 49.69	674 $\pm$ 262.78	521 $\pm$ 123.33**
	WBC ( $\times 10^3/\mu\text{L}$ )	3.7 $\pm$ 1.08	4.17 $\pm$ 1.21	4.39 $\pm$ 0.89	3.57 $\pm$ 0.59
	NEU ( $\times 10^3/\mu\text{L}$ )	1.03 $\pm$ 0.21	1.28 $\pm$ 0.25	1.48 $\pm$ 0.33	1.74 $\pm$ 0.31**
	NEU% (%)	28.88 $\pm$ 6.02	31.66 $\pm$ 5.13	33.64 $\pm$ 2.62	49.32 $\pm$ 8.87**
	EOS ( $\times 10^3/\mu\text{L}$ )	0.02 $\pm$ 0.01	0.03 $\pm$ 0.01	0.03 $\pm$ 0.02	0.02 $\pm$ 0.01
	EOS% (%)	0.64 $\pm$ 0.13	0.62 $\pm$ 0.25	0.62 $\pm$ 0.31	0.64 $\pm$ 0.32
	BAS ( $\times 10^3/\mu\text{L}$ )	0.01 $\pm$ 0.01	0.01 $\pm$ 0.01	0.01 $\pm$ 0.01	0.01 $\pm$ 0.01
	BAS% (%)	0.22 $\pm$ 0.11	0.18 $\pm$ 0.08	0.28 $\pm$ 0.13	0.14 $\pm$ 0.09
	MON ( $\times 10^3/\mu\text{L}$ )	0.06 $\pm$ 0.02	0.08 $\pm$ 0.02	0.09 $\pm$ 0.04	0.13 $\pm$ 0.04*
	MON% (%)	1.8 $\pm$ 0.31	1.92 $\pm$ 0.31	1.96 $\pm$ 0.42	3.66 $\pm$ 0.81**
	LYM ( $\times 10^3/\mu\text{L}$ )	2.54 $\pm$ 0.94	2.75 $\pm$ 0.99	2.74 $\pm$ 0.55	1.63 $\pm$ 0.55
LYM% (%)	67.82 $\pm$ 5.85	64.84 $\pm$ 5.37	60.4 $\pm$ 4.09	45.06 $\pm$ 9.08**	
RET ( $\times 10^9/\mu\text{L}$ )	332.14 $\pm$ 40.27	360.48 $\pm$ 56.64	172.98 $\pm$ 38.23**	51.1 $\pm$ 28.16**	
RET% (%)	3.94 $\pm$ 0.42	3.92 $\pm$ 0.55	1.75 $\pm$ 0.39**	0.49 $\pm$ 0.26**	
Female	RBC ( $\times 10^3/\mu\text{L}$ )	8.64 $\pm$ 0.26	9.16 $\pm$ 0.08*	9.94 $\pm$ 0.41**	10.42 $\pm$ 0.19**
	HGB (g/dL)	16 $\pm$ 0.49	16.94 $\pm$ 0.13	18.16 $\pm$ 0.76**	18.9 $\pm$ 0.37**
	HCT (%)	47.22 $\pm$ 1.41	49.9 $\pm$ 0.19	53.16 $\pm$ 2.38*	54.24 $\pm$ 1.19**
	MCV (fL)	54.63 $\pm$ 0.55	54.46 $\pm$ 0.65	53.52 $\pm$ 0.77	52.04 $\pm$ 0.39**
	MCH (pg)	18.54 $\pm$ 0.05	18.5 $\pm$ 0.07	18.32 $\pm$ 0.2	18.16 $\pm$ 0.09**
	MCHC (g/dL)	33.9 $\pm$ 0.34	33.98 $\pm$ 0.37	34.2 $\pm$ 0.22	34.92 $\pm$ 0.19**
	PLT ( $\times 10^3/\mu\text{L}$ )	678 $\pm$ 369.03	874.8 $\pm$ 64.08	745 $\pm$ 89.39	435 $\pm$ 45.14
	WBC ( $\times 10^3/\mu\text{L}$ )	2.77 $\pm$ 0.77	3.21 $\pm$ 0.5	3.81 $\pm$ 1.11	3.87 $\pm$ 1.21
	NEU ( $\times 10^3/\mu\text{L}$ )	0.52 $\pm$ 0.15	0.64 $\pm$ 0.09	1.06 $\pm$ 0.19	2.12 $\pm$ 0.66**
	NEU% (%)	19.38 $\pm$ 4.04	19.88 $\pm$ 1.39	28.64 $\pm$ 4.26**	54.86 $\pm$ 4.02**
	EOS ( $\times 10^3/\mu\text{L}$ )	0.03 $\pm$ 0.01	0.04 $\pm$ 0.01	0.02 $\pm$ 0	0.01 $\pm$ 0.00**
	EOS% (%)	1 $\pm$ 0.19	1.2 $\pm$ 0.26	0.6 $\pm$ 0.10*	0.24 $\pm$ 0.09**
	BAS ( $\times 10^3/\mu\text{L}$ )	0.01 $\pm$ 0.01	0.01 $\pm$ 0	0.01 $\pm$ 0.01	0.01 $\pm$ 0.01
	BAS% (%)	0.2 $\pm$ 0.17	0.18 $\pm$ 0.11	0.32 $\pm$ 0.13	0.24 $\pm$ 0.09
	MON ( $\times 10^3/\mu\text{L}$ )	0.04 $\pm$ 0.01	0.05 $\pm$ 0.01	0.08 $\pm$ 0.03	0.14 $\pm$ 0.05**
	MON% (%)	1.58 $\pm$ 0.29	1.46 $\pm$ 0.54	1.98 $\pm$ 0.48	3.48 $\pm$ 0.72**
	LYM ( $\times 10^3/\mu\text{L}$ )	2.15 $\pm$ 0.66	2.46 $\pm$ 0.41	2.61 $\pm$ 0.90*	1.54 $\pm$ 0.54
LYM% (%)	77.28 $\pm$ 4.28	76.4 $\pm$ 1.42	67.7 $\pm$ 4.14**	39.48 $\pm$ 4.03**	
RET ( $\times 10^9/\mu\text{L}$ )	212.78 $\pm$ 30.33	236.58 $\pm$ 13.38	115.22 $\pm$ 20.93**	44.58 $\pm$ 9.61**	
RET% (%)	2.46 $\pm$ 0.29	2.58 $\pm$ 0.14	1.16 $\pm$ 0.20**	0.43 $\pm$ 0.10**	

All values are expressed as mean  $\pm$  SD. Significant differences compared with the control: \* $p < 0.05$ , \*\* $p < 0.01$ .

**Table 4.** Summary of Serum Chemical Parameters

Sex	Group	0 mg/m <sup>3</sup>	1 mg/m <sup>3</sup>	5 mg/m <sup>3</sup>	25 mg/m <sup>3</sup>
	No. of rats	5	5	5	5
Male	ALT (IU/L)	57.9 ± 6.28	53.18 ± 1.67	57.47 ± 8.81	72.78 ± 15.22
	AST (IU/L)	76.84 ± 4.39	75.5 ± 9.76	92.2 ± 16.29	106.8 ± 9.88**
	ALP (IU/L)	1009.54 ± 49.83	850.9 ± 65.91*	778.63 ± 58.64**	751.05 ± 84.64**
	GLU (mg/dL)	144.9 ± 12.13	134.74 ± 9.45	115.57 ± 9.82	137.9 ± 25.65
	BUN (mg/dL)	18.2 ± 1.92	17.22 ± 0.75	14.97 ± 0.51	21.53 ± 3.11
	CREA (mg/dL)	0.37 ± 0.03	0.36 ± 0.02	0.37 ± 0.02	0.37 ± 0.02
	T-CHO (mg/dL)	60.66 ± 7.44	76.42 ± 8.39	89.67 ± 9.92	79.7 ± 16.48
	TG (mg/dL)	64.22 ± 12.47	47.54 ± 8.91	26.77 ± 3.12**	23.13 ± 5.14**
	TP (g/dL)	5.76 ± 0.09	5.98 ± 0.23	5.73 ± 0.21	5.2 ± 0.25**
ALB (g/dL)	4.2 ± 0.07	4.34 ± 0.11	4.13 ± 0.12	3.78 ± 0.13	
Female	ALT (IU/L)	42.22 ± 2.66	42.36 ± 2.33	63.97 ± 9.60**	65.98 ± 7.33**
	AST (IU/L)	75.42 ± 7.27	83.18 ± 4.37	91.8 ± 11.78	111.54 ± 7.61**
	ALP (IU/L)	656.58 ± 39.43	651.92 ± 62	698 ± 73.72	640.14 ± 71.63
	GLU (mg/dL)	108.76 ± 14.71	97.22 ± 8.18	130.3 ± 26.02	129.5 ± 25.48
	BUN (mg/dL)	14.54 ± 0.81	16.56 ± 0.57	14.27 ± 0.59	23.74 ± 2.90**
	CREA (mg/dL)	0.36 ± 0.01	0.36 ± 0.02	0.36 ± 0.02	0.33 ± 0.02
	T-CHO (mg/dL)	93.54 ± 6.2	94.9 ± 2.58	80.23 ± 9.38	58.54 ± 8.13**
	TG (mg/dL)	40.24 ± 9.24	23.46 ± 3.52**	19.9 ± 4.07**	24.86 ± 5.96*
	TP (g/dL)	5.38 ± 0.18	5.5 ± 0.12	5.23 ± 0.23	4.78 ± 0.15**
	ALB (g/dL)	3.98 ± 0.11	4 ± 0.07	3.83 ± 0.15	3.6 ± 0.12**

All values are expressed as mean ± SD. Significant differences compared with the control: \*p<0.05, \*\*p<0.01.

exposed to 25 mg/m<sup>3</sup>. In contrast, ALP decreased significantly in males exposed to 1, 5, and 25 mg/m<sup>3</sup>. Moreover, TG decreased significantly in males exposed to 5 and 25 mg/m<sup>3</sup> and females exposed to 1, 5, and 25 mg/m<sup>3</sup>. TP and ALB decreased significantly in males and females exposed to 25 mg/m<sup>3</sup>. T-CHO decreased significantly in females exposed to 25 mg/m<sup>3</sup> (Table 4).

#### Organ weight

The absolute and relative organ weights of the lungs increased significantly in males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>. The absolute organ weight of the liver decreased significantly in males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>, as did the relative organ weight of the liver in males exposed to 5 and 25 mg/m<sup>3</sup> and females exposed to 1 mg/m<sup>3</sup>. The absolute organ weight of the spleen decreased significantly in males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>. The absolute organ weight of the kidneys decreased significantly in males exposed to 5 and 25 mg/m<sup>3</sup> and females exposed to 1, 5, and 25 mg/m<sup>3</sup>. However, the relative organ weight of the kidneys increased significantly in males and females exposed to 25 mg/m<sup>3</sup>. The absolute organ weight of the heart decreased significantly in males and females exposed to 5 and 25 mg/m<sup>3</sup>, and the relative organ weight of the heart increased significantly in males and females exposed to 25 mg/m<sup>3</sup>. The absolute organ weight of the brain decreased significantly in males and females exposed to 25 mg/m<sup>3</sup>, and the relative organ weight of the brain increased significantly in males and females exposed to 5 and 25 mg/m<sup>3</sup> (Table 5 and 6).

#### Gross lesion evaluations

Ballooning was observed in the lungs of males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>, and red focus was observed in the lungs of males and females exposed to 25 mg/m<sup>3</sup>. Increased size in the tracheobronchial lymph node was observed in males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>, and red focus and decreased size in the liver, spleen, and thymus were observed in males and females exposed to 25 mg/m<sup>3</sup>. Additionally, decreased size was observed in the testes and epididymides of males exposed to 25 mg/m<sup>3</sup>, in the seminal vesicle and prostate of males exposed to 5 and 25 mg/m<sup>3</sup>, and in the uterus and vagina of females exposed to 1, 5, and 25 mg/m<sup>3</sup> (Table 7).

#### Histopathological examination

##### Nasal cavity

Squamous metaplasia of respiratory and transitional epithelium were observed in males and females exposed to 25 mg/m<sup>3</sup>. Degeneration of respiratory and transitional epithelium were observed in males and females exposed to 5 mg/m<sup>3</sup>. A decrease in number of mucous cells in the respiratory epithelium was observed in males and females exposed to 1 and 5 mg/m<sup>3</sup>. Atrophy of olfactory epithelium was observed in males and females exposed to 5 and 25 mg/m<sup>3</sup>. Ulcer of transitional epithelium was observed in males exposed to 25 mg/m<sup>3</sup> and females exposed to 5 and 25 mg/m<sup>3</sup>. The severity of the nasal cavity lesion increased in a dose-related manner (Table 8 and Fig. 3A–D).

**Table 5.** Summary of Absolute Organ Weight

Group		0 mg/m <sup>3</sup>	1 mg/m <sup>3</sup>	5 mg/m <sup>3</sup>	25 mg/m <sup>3</sup>
No. of rats		5	5	5	5
Male	Body weight (g)	197.264 ± 9.169	178.866 ± 6.031**	145.952 ± 4.916**	100.994 ± 4.790**
	Brain (g)	1.687 ± 0.035	1.692 ± 0.065	1.636 ± 0.027	1.571 ± 0.036**
	Heart (g)	0.656 ± 0.049	0.6 ± 0.023	0.484 ± 0.052**	0.403 ± 0.015**
	Lungs (g)	0.861 ± 0.05	1.387 ± 0.137**	1.306 ± 0.094**	1.407 ± 0.119**
	Liver (g)	6.336 ± 0.336	5.563 ± 0.316**	3.867 ± 0.203**	2.931 ± 0.194**
	Spleen (g)	0.488 ± 0.039	0.418 ± 0.014**	0.318 ± 0.013**	0.22 ± 0.011**
	Kidneys (g)	1.346 ± 0.055	1.271 ± 0.037	1.015 ± 0.035**	0.876 ± 0.068**
Female	Body weight (g)	141.706 ± 5.599	130.036 ± 5.065**	105.012 ± 3.249**	74.874 ± 2.085**
	Brain (g)	1.668 ± 0.029	1.6 ± 0.063	1.59 ± 0.056	1.546 ± 0.030*
	Heart (g)	0.508 ± 0.022	0.468 ± 0.036	0.41 ± 0.015**	0.325 ± 0.019**
	Lungs (g)	0.748 ± 0.088	1.04 ± 0.068**	1.104 ± 0.055**	1.328 ± 0.171**
	Liver (g)	4.02 ± 0.166	3.53 ± 0.151**	2.897 ± 0.178**	2.222 ± 0.122**
	Spleen (g)	0.368 ± 0.02	0.326 ± 0.018*	0.285 ± 0.015**	0.179 ± 0.013**
	Kidneys (g)	1.124 ± 0.068	1.022 ± 0.048*	0.878 ± 0.025**	0.783 ± 0.030**

All values are expressed as mean ± SD. Significant differences compared with the control: \*p<0.05, \*\*p<0.01.

**Table 6.** Summary of Relative Organ Weight

Group		0 mg/m <sup>3</sup>	1 mg/m <sup>3</sup>	5 mg/m <sup>3</sup>	25 mg/m <sup>3</sup>
No. of rats		5	5	5	5
Male	Body weight (g)	197.264 ± 9.169	178.866 ± 6.031**	145.952 ± 4.916**	100.994 ± 4.790**
	Brain (%)	0.857 ± 0.044	0.947 ± 0.054	1.122 ± 0.049**	1.557 ± 0.055**
	Heart (%)	0.333 ± 0.028	0.336 ± 0.016	0.331 ± 0.027	0.4 ± 0.021**
	Lungs (%)	0.436 ± 0.014	0.775 ± 0.070**	0.897 ± 0.087**	1.398 ± 0.163**
	Liver (%)	3.212 ± 0.074	3.109 ± 0.103	2.648 ± 0.054**	2.902 ± 0.131**
	Spleen (%)	0.248 ± 0.024	0.234 ± 0.003	0.218 ± 0.006	0.219 ± 0.014
	Kidneys (%)	0.683 ± 0.024	0.711 ± 0.009	0.696 ± 0.007	0.867 ± 0.051**
Female	Body weight (g)	141.706 ± 5.599	130.036 ± 5.065**	105.012 ± 3.249**	74.874 ± 2.085**
	Brain (%)	1.179 ± 0.057	1.232 ± 0.057	1.516 ± 0.077**	2.066 ± 0.071**
	Heart (%)	0.359 ± 0.013	0.36 ± 0.025	0.39 ± 0.012*	0.435 ± 0.037*
	Lungs (%)	0.527 ± 0.045	0.801 ± 0.075**	1.051 ± 0.044**	1.773 ± 0.216**
	Liver (%)	2.837 ± 0.017	2.715 ± 0.056*	2.756 ± 0.088	2.968 ± 0.149
	Spleen (%)	0.26 ± 0.009	0.251 ± 0.007	0.272 ± 0.01	0.239 ± 0.015
	Kidneys (%)	0.729 ± 0.02	0.786 ± 0.018	0.836 ± 0.024	1.045 ± 0.015**

All values are expressed as mean ± SD. Significant differences compared with the control: \*p<0.05, \*\*p<0.01.

### Larynx

Squamous metaplasia of epithelium and inflammation of the lamina propria were observed in males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>. Ulcer of the epithelium was observed in males exposed to 5 and 25 mg/m<sup>3</sup> and females exposed to 25 mg/m<sup>3</sup>. The severity of the larynx lesion increased in a dose-related manner (Table 8 and Fig. 3E–H).

### Trachea

Degeneration of the epithelium was observed in males exposed to 1, 5, and 25 mg/m<sup>3</sup> and females exposed to 5 and 25 mg/m<sup>3</sup>. Necrosis of the epithelium was observed in males exposed to 25 mg/m<sup>3</sup>. The severity of the trachea lesion increased in a dose-related manner (Table 8 and Fig. 3I–L).

### Lung

Squamous metaplasia of bronchial and bronchiolar epithelium and alveolar emphysema were observed in males and females exposed to 5 and 25 mg/m<sup>3</sup>. Additionally, necrosis with inflammation; alveolar fibrosis; detachment of bronchial and bronchiolar epithelium; and peribronchiolar, perivascular, and alveolar inflammatory cell infiltration were observed in males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>. Alveolar hemorrhage was observed in males exposed to 5 and 25 mg/m<sup>3</sup> and females exposed to 1 and 25 mg/m<sup>3</sup>. Lymphoid hyperplasia of bronchus-associated lymphoid tissue (BALT) was observed in males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>. Alveolar macrophage aggregation was observed in males exposed to 0, 1, 5, and 25 mg/m<sup>3</sup> and females exposed to 1 and 25 mg/m<sup>3</sup>. The severity of the lung lesion increased in a dose-related manner (Table 8 and Fig. 3M–P).

**Table 7.** Summary of Gross Findings

Sex	Organ	Findings	Group	0 mg/m <sup>3</sup>	1 mg/m <sup>3</sup>	5 mg/m <sup>3</sup>	25 mg/m <sup>3</sup>
			No. of rats	5	5	5	5
Male	Unremarkable findings			0	0	0	0
	Lungs	Ballooning		0	5	5	5
		Focus, red		0	0	0	4
	Tracheobronchial lymph node	Increased size		0	4	2	2
		Decreased size		0	0	0	5
	Liver	Decreased size		0	0	0	5
	Epididymides	Decreased size		0	0	0	5
	Prostate	Decreased size		0	0	1	5
	Seminal vesicle	Decreased size		0	0	1	5
	Spleen	Decreased size		0	0	0	5
	Testes	Decreased size		0	0	0	5
Thymus	Decreased size		0	0	0	5	
Female	Unremarkable findings			0	0	0	0
	Lungs	Ballooning		0	5	5	5
		Focus, red		0	0	0	5
	Tracheobronchial lymph node	Increased size		0	3	2	3
		Decreased size		0	0	0	5
	Liver	Decreased size		0	0	0	5
	Spleen	Decreased size		0	0	0	4
	Thymus	Decreased size		0	0	0	4
	Uterus	Decreased size		0	1	1	5
	Vagina	Decreased size		0	1	1	5

**Table 8.** Summary of Histopathology

Sex	Organ	Findings	Group	0 mg/m <sup>3</sup>	1 mg/m <sup>3</sup>	5 mg/m <sup>3</sup>	25 mg/m <sup>3</sup>
			No. of rats	5	5	5	5
Male	Nasal cavity	Squamous metaplasia, respiratory epithelium	+	0	0	0	1
			++	0	0	0	2
			+++	0	0	0	2
		Degeneration, respiratory epithelium	±	0	0	2	0
			+	0	0	1	0
			++	0	0	1	0
			+++	0	0	1	0
			+++	0	0	1	0
		Decreased mucous cell, respiratory epithelium	++	0	3	0	0
			+++	0	2	5	0
		Squamous metaplasia, transitional epithelium	++	0	0	0	3
			+++	0	0	0	2
	Degeneration, transitional epithelium	+	0	0	3	0	
		++	0	0	2	0	
	Ulcer, transitional epithelium	+	0	0	0	2	
		++	0	0	0	1	
	Atrophy, olfactory epithelium	±	0	0	1	0	
		+	0	0	4	0	
		++	0	0	0	3	
		+++	0	0	0	2	
	Larynx	Squamous metaplasia, epithelium	±	0	2	0	0
+			0	2	2	0	
++			0	1	1	3	
+++			0	0	2	2	
Inflammation, lamina propria		±	0	1	3	1	
		+	0	2	0	2	
		++	0	0	2	0	
		+++	0	0	0	1	
Ulcer, epithelium		±	0	0	1	1	
		+++	0	0	0	1	

Table 8. Continued

Sex	Organ	Findings	Group	0 mg/m <sup>3</sup>	1 mg/m <sup>3</sup>	5 mg/m <sup>3</sup>	25 mg/m <sup>3</sup>
			No. of rats	5	5	5	5
	Trachea	Degeneration, epithelium	±	0	3	0	0
			+	0	0	2	0
			++	0	0	3	2
			+++	0	0	0	3
		Necrosis, epithelium	±	0	0	0	1
			+	0	0	0	1
	Lung	Squamous metaplasia, bronchial/ bronchiolar	±	0	0	1	0
			+	0	0	2	0
			++	0	0	2	0
			+++	0	0	0	1
			++++	0	0	0	4
		Necrosis with inflammation, alveolar	++	0	3	2	0
			+++	0	0	1	1
			++++	0	0	0	4
		Fibrosis, alveolar	±	0	5	0	0
			+	0	0	5	0
			++	0	0	0	5
		Detachment, bronchial/bronchiolar epithelium	±	0	3	2	0
			+	0	2	2	0
			++	0	0	1	5
		Inflammatory cell infiltration, peribronchiolar/ perivascular/alveolar	±	0	3	0	0
			+	0	2	5	1
			++	0	0	0	4
		Alveolar emphysema	±	0	0	4	0
			+	0	0	0	2
			++	0	0	0	3
		Hemorrhage, alveolar	±	0	0	1	3
			+	0	0	0	2
		Lymphoid hyperplasia, BALT	±	0	2	3	4
			+	0	1	2	0
		Alveolar macrophage aggregation	±	1	1	2	1
		Lymph node, Tracheobronchial	Lymphoid hyperplasia	±	0	1	3
	+			0	0	1	2
	++			0	2	1	1
	+++			0	2	0	0
	Liver	Atrophy	±	0	0	4	0
			+	0	0	1	2
			++	0	0	0	2
			+++	0	0	0	1
	Spleen	Atrophy	++	0	0	3	0
			+++	0	0	2	5
	Thymus	Atrophy	±	0	0	0	1
			++	0	0	0	2
			+++	0	0	0	2
	Prostate	Atrophy	+	0	0	1	0
			++	0	0	1	2
			+++	0	0	0	3
	Seminal vesicles	Atrophy	+	0	0	3	0
			++	0	0	1	0
			+++	0	0	0	5
	Epididymides	Cell debris, luminal	±	1	1	0	0
			+	1	0	0	3
			++	0	0	0	1
			+++	0	0	0	1
		Reduced sperm, luminal	±	1	0	0	0
			+	0	1	0	0
	Testes	Immaturity	±	0	0	0	1
			++	0	0	0	2
			+++	0	0	0	2



Table 8. Continued

Sex	Organ	Findings	Group	0 mg/m <sup>3</sup>	1 mg/m <sup>3</sup>	5 mg/m <sup>3</sup>	25 mg/m <sup>3</sup>
			No. of rats	5	5	5	5
Female	Nasal cavity	Squamous metaplasia, respiratory epithelium	±	0	0	0	2
			+	0	0	0	1
			++	0	0	0	2
		Degeneration, respiratory epithelium	±	0	0	2	0
			+	0	0	1	0
			++	0	0	2	0
		Decreased mucous cell, respiratory epithelium	+	0	4	0	0
			++	0	1	0	0
			+++	0	0	5	0
		Squamous metaplasia, transitional epithelium	+	0	0	0	2
			++	0	0	0	1
			+++	0	0	0	2
		Degeneration, transitional epithelium	±	0	0	4	0
			+	0	0	1	0
			+++	0	0	3	1
		Ulcer, transitional epithelium	±	0	0	3	0
			+	0	0	2	0
			++	0	0	0	3
	Atrophy, olfactory epithelium	±	0	0	0	2	
		+	0	0	0	0	
		+++	0	0	0	2	
	Larynx	Squamous metaplasia, epithelium	±	0	2	0	0
			+	0	2	2	0
			++	0	1	1	3
			+++	0	0	2	2
		Inflammation, lamina propria	±	0	2	2	1
			+	0	1	2	1
			++	0	0	1	1
		Ulcer, epithelium	+++	0	0	0	1
			++	0	0	0	1
	Trachea	Degeneration, epithelium	±	0	0	3	0
			++	0	0	0	3
			+++	0	0	0	2
	Lung	Squamous metaplasia, bronchial/ bronchiolar	+	0	0	2	0
			++	0	0	3	1
			+++	0	0	0	0
++++			0	0	0	4	
Necrosis with inflammation, alveolar		±	0	1	0	0	
		+	0	2	4	2	
		++	0	0	1	2	
		+++	0	0	0	1	
Fibrosis, alveolar		±	0	5	0	0	
		+	0	0	4	0	
		++	0	0	1	4	
		+++	0	0	0	1	
Detachment, bronchial/bronchiolar epithelium		±	0	5	0	0	
		+	0	0	2	1	
		++	0	0	3	4	
		+++	0	0	0	1	
Inflammatory cell infiltration, peribronchiolar/ perivascular/alveolar		±	0	4	0	0	
		+	0	1	5	0	
		++	0	0	0	4	
		+++	0	0	0	1	
Alveolar emphysema		±	0	0	2	0	
		+	0	0	2	0	
		++	0	0	0	4	
		+++	0	0	0	1	
Hemorrhage, alveolar		±	0	1	0	2	
		+	0	0	0	2	
		++	0	0	0	1	
Lymphoid hyperplasia, BALT		±	0	1	2	5	
		+	0	0	2	0	
		±	0	3	0	2	
Alveolar macrophage aggregation		±	0	3	0	2	

**Table 8.** Continued

Sex	Organ	Findings	Group	0 mg/m <sup>3</sup>	1 mg/m <sup>3</sup>	5 mg/m <sup>3</sup>	25 mg/m <sup>3</sup>
			No. of rats	5	5	5	5
	Lymph node, Tracheobronchial	Lymphoid hyperplasia	±	0	1	1	0
			+	0	2	1	3
			++	0	2	2	2
			+++	0	0	1	0
Liver	Atrophy	±	0	0	2	0	
		+	0	0	1	2	
		++	0	0	0	1	
		+++	0	0	0	2	
Spleen	Atrophy	++	0	0	4	0	
		+++	0	0	1	4	
		++++	0	0	0	1	
Thymus	Atrophy	±	0	0	2	0	
		+++	0	0	0	5	
Ovaries	Atrophy	+	0	0	0	2	
		++	0	0	0	2	
		+++	0	0	0	1	
Uterus	Atrophy	+	0	0	3	0	
		++	0	0	2	3	
		+++	0	0	0	2	
Vagina	Atrophy, epithelial	±	0	5	1	0	
		+	0	0	2	0	
		++	0	0	1	0	
		+++	0	0	1	5	

Grade: ± : minimal, +: mild, ++: moderate, +++: marked, ++++: severe.

### Tracheobronchial lymph node

Lymphoid hyperplasia was observed in males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>. The severity of the tracheobronchial lymph node lesion increased in a dose-related manner (Table 8).

### Other organs

Atrophy of the liver was observed in males and females exposed to 5 and 25 mg/m<sup>3</sup>. Atrophy of the spleen was observed in males and females exposed to 5 and 25 mg/m<sup>3</sup>. Atrophy of the thymus was observed in males exposed to 25 mg/m<sup>3</sup> and females exposed to 5 and 25 mg/m<sup>3</sup>. Atrophy of the prostate and seminal vesicle was observed in males exposed to 5 and 25 mg/m<sup>3</sup>. Cell debris was observed in the epididymides of males exposed to 0, 1, and 25 mg/m<sup>3</sup>, and reduced sperm was observed in males exposed to 0 and 1 mg/m<sup>3</sup>. Immaturity was observed in the testes of males exposed to 25 mg/m<sup>3</sup>. Atrophy of the ovary, uterus, and vagina was observed in females exposed to 5 mg/m<sup>3</sup>; 5 and 25 mg/m<sup>3</sup>; and 1, 5, and 25 mg/m<sup>3</sup>, respectively (Table 8).

## Discussion

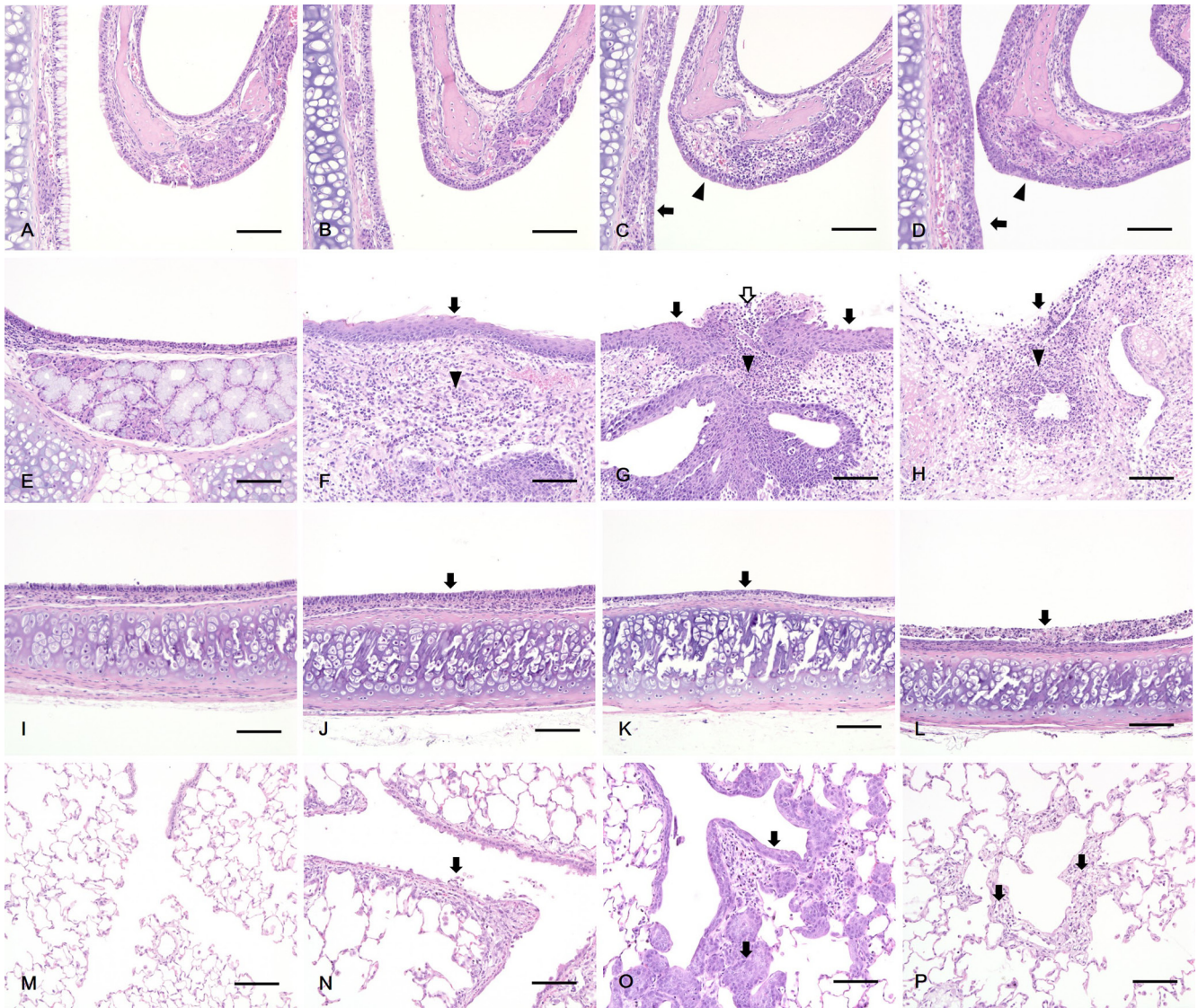
In the present study, the treatment-related effects of PHMG-HCl in rats were observed in clinical signs; body weight gain; hematology and serum biochemistry; organ weight; gross lesions and histopathological lesions in the nasal cavity, larynx, trachea, lungs, tracheobronchial lymph node, liver, spleen, thymus, seminal vesicle, prostate, testes,

epididymides, ovary, uterus, and vagina.

Body weights decreased significantly in males and female exposed to 1, 5 and 25 mg/m<sup>3</sup> until termination of the study. In particular, body weight decreased far less than that in the start of the study in the male and female exposed to 5 and 25 mg/m<sup>3</sup>, which corresponds to aggravated clinical signs including emaciation and rale observed in the males and females exposed to 25 mg/m<sup>3</sup>. This may be attributed to lung inflammation associated with cytokines and stress-induced anorexia even though we did not evaluate food consumption in this study. Inflammatory cytokines and stress are reported to act on the hypothalamus and induce anorexia<sup>16, 17</sup>.

RBC, HCT, HGB, MCHC, MCH, MCV, RET, RET %, and PLT changed significantly in males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>. This may be attributed to decreased hematopoiesis caused by anorexia<sup>18, 19</sup>. In addition, MON%, NEU, NEU%, and LYM% changed significantly in males and females exposed to 5 and 25 mg/m<sup>3</sup>. This may be associated with inflammation lesions including necrosis, inflammatory cell infiltration, and fibrosis<sup>20</sup>. Moreover, decrease in LYM% may be associated with stress responses<sup>21</sup>.

Increase or increasing trends in ALT and AST was observed in males and females exposed to 5 and 25 mg/m<sup>3</sup>. This may be attributed to atrophy of the liver<sup>18</sup>, because integrity of hepatocyte membranes might be disrupted by a decrease in size of the hepatocytes, which causes leakage of enzymes. Increases or increasing trends in BUN was observed in males and females exposed to 25 mg/m<sup>3</sup>. This



**Fig. 3.** Histopathology of rats exposed to PHMG·HCl. In the nasal cavity, (A, B) No abnormal lesion was observed in the control (A) and 1 mg/m<sup>3</sup> PHMG·HCl (B) groups. (C) Degeneration of respiratory epithelium (arrow) and transitional epithelium (arrowhead) in the 5 mg/m<sup>3</sup> PHMG·HCl group. (D) Squamous metaplasia of respiratory epithelium (arrow) and transitional epithelium (arrowhead) in the 25 mg/m<sup>3</sup> PHMG·HCl group. In the larynx, (E) No abnormal lesion was observed in the control group. (F, G) Squamous metaplasia of the epithelium (arrow), inflammation of the lamina propria (arrowhead) in 1 (F) and 5 mg/m<sup>3</sup> (G) PHMG·HCl groups and ulcer of the epithelium (white arrow) in 5 mg/m<sup>3</sup> (G) PHMG·HCl group. (H) Ulcer of the epithelium (arrow) and inflammation of the lamina propria (arrowhead) in the 25 mg/m<sup>3</sup> PHMG·HCl groups. In the trachea, (I) No abnormal lesion was observed in the control group. (J, K) Degeneration of the epithelium (arrow) in 1 (J) and 5 mg/m<sup>3</sup> (K) PHMG·HCl groups. (L) Necrosis of the epithelium (arrow) in the 25 mg/m<sup>3</sup> PHMG·HCl group. In the lung, (M) No abnormal lesion was observed in the control group. (N) Detachment of the bronchiolar epithelium (arrow) in the 1 mg/m<sup>3</sup> PHMG·HCl group. (O) Squamous metaplasia of bronchiolar epithelium (arrow) in the 5 mg/m<sup>3</sup> PHMG·HCl group. (P) Alveolar fibrosis (arrow) in the 25 mg/m<sup>3</sup> PHMG·HCl group. Scale bars=100  $\mu$ m, Magnification:  $\times$ 200, H&E staining.

may be attributed to dehydration or catabolism of protein, because increases or increasing trends in RBC, HCT, HGB, MCH, and MCHC were observed<sup>17</sup>, and clinical signs included emaciation<sup>22, 23</sup>.

Changes in ALP, TG, TP, ALB and T-CHO were observed in males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>. This may be attributed to decreased food consumption<sup>19</sup>, which we speculate from decreased body weights even though we did not measure food consumption. Changes in

the absolute and relative organ weights of the brain, heart, liver, spleen, and kidneys were observed in females exposed to 1, 5, and 25 mg/m<sup>3</sup>. These were associated with the decreased body weights of rats. Increases in the absolute and relative organ weights of the lungs were observed in males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>. These may have been affected by lung inflammation<sup>24, 25</sup>.

Most of histological lesions (including degeneration, atrophy, ulcer, inflammatory cell infiltration, inflamma-

tion, and fibrosis in nasal cavity, larynx, trachea, and lungs) indicated tissue damage by test substance<sup>26</sup>. In particular, lesions in the lungs were observed mainly at the junction of terminal bronchioles and alveolar ducts (the centriacinar region) where the velocity of air flow is decreased and aerosol particulates can be easily deposited and induce damage to cells<sup>25, 26</sup>. Similarly, a centrilobular pattern of lesions was also observed in human patients exposed to humidifier disinfectants<sup>27</sup>. Many studies have reported that the toxicity of PHMG is related to oxidative stress. PHMG·P produces ROS in human alveolar A549 cells, mouse macrophage RAW264.7 cells, or *in vitro* air-liquid interface (ALI) co-culture models and causes fibrosis and inflammation via cellular signals, such as cytokines<sup>13, 28, 29</sup>. In particular, 4-hydroxynonenal (4-HNE), an oxidative stress marker, was confirmed by immunohistochemistry in the macrophages of the fibrotic tissue and the bronchiolar epithelium, mainly in Clara cells in 13-week inhalation study of PHMG·HCl. This indicated that these cells play a critical role in damaging the lung<sup>30</sup>.

Squamous metaplasia is considered to be an adaptive or protective response to irritation than a precursor to neoplastic lesions<sup>31</sup>. Notably, it has been reported that the larynx of rodents is more sensitively affected by inhaled xenobiotics than those of non-rodents. However, these findings lack relevance for humans because of differences in anatomical structures<sup>32</sup>. Even though squamous metaplasia is an adaptive or protective change, it should be considered an adverse effect because of the severity of the lesion and concomitant degenerative/necrotic and/or hyperplastic changes<sup>33</sup>. Interestingly, the grade of squamous metaplasia of lung in our study is found to be more severe than that in 13-week inhalation study of PHMG·HCl (Inhalation concentration: 0 mg/m<sup>3</sup>, 0.13 mg/m<sup>3</sup>, 0.4 mg/m<sup>3</sup>, and 1.20 mg/m<sup>3</sup>)<sup>30</sup>. This indicates that the epithelium of lung, mainly at the junction of terminal bronchioles and alveolar ducts (the centriacinar region) is more severely damaged by PHMG·HCl and rapidly progressed to squamous metaplasia because of concentration of PHMG·HCl much higher than that in 13-week inhalation study period even though squamous metaplasia is generally found in long term study. These lesions of the respiratory system were also observed in a PHMG·P inhalation toxicity study<sup>34</sup>.

Lymphoid hyperplasia of the tracheobronchial lymph node and BALT of the lung were observed in males and females exposed to 1, 5, and 25 mg/m<sup>3</sup>. These are considered to be immune responses to PHMG·HCl<sup>35</sup>. Atrophy of the liver and spleen are considered to be effects of decreased body weight in males and females exposed to 5 and 25 mg/m<sup>3</sup><sup>36, 37</sup>. Atrophy of the thymus is considered to be a stress response affected by aggravated clinical signs including decreased body weight in males and females exposed to 5 and 25 mg/m<sup>3</sup><sup>38</sup>. Atrophy of reproductive organs in both male and female rats indicate alteration of sexual hormones affected by aggravated clinical signs including decreases in body weight<sup>39, 40</sup>.

In conclusion, two-week repeated whole-body inhalation exposure of rats to three different concentrations of PHMG·HCl revealed toxic effects on the respiratory system and secondary effect on other organs. The results of this study indicate that the no observable adverse effect level (NOAEL) for PHMG·HCl is below 1 mg/m<sup>3</sup>. The present study provides useful information regarding inhalation toxicity of PHMG·HCl.

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