

MITOGENOME ANNOUNCEMENT

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The complete chloroplast genome of *Rungia pectinata* (Acanthaceae)

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ABSTRACT

Rungia pectinata is an important traditional Chinese herbal medicine from the family Acanthaceae. The complete chloroplast genome (cp genome) of the genus *Rungia* was sequenced for the first time. The cp genome of *R. pectinata* was 149,627 bp in length. It was consisted of a large single copy (LSC) region (81,976 bp) and a small single copy (SSC) region (16,626 bp), which were separated by two inverted repeats (IRs, 25,511 bp). This plastome contained 114 unique genes, including 80 protein-coding genes, 30 tRNA genes, and four rRNA genes. The overall GC content was 38.0%. Phylogenetic analysis of nine species in Acanthaceae was also conducted. This newly sequenced cp genome will be useful to further evolutionary studies, phylogenetic studies, and pharmacognostic identification in the genus *Rungia*.

ARTICLE HISTORY

Received 30 May 2019

Accepted 10 July 2019

KEYWORDS

Rungia pectinata; chloroplast genome; phylogenetic; herbal medicine

Rungia Nees (Acanthaceae) is a genus comprising about 50 species distributed in the tropical and subtropical regions of the Old World (Hu et al. 2011; Mabberley 2017). *Rungia pectinata* (L.) Nees is a widespread species in Asia, the Chinese name of *R. pectinata* is “infant’s herb” as it is effective in treatment of infantile indigestion (Lo 1978). In this study, the complete chloroplast genome of the genus *Rungia* was sequenced for the first time, which will be useful to pharmacognostic identification in medicinal species of *Rungia*.

The complete chloroplast genome of *R. pectinata* was successfully assembled and characterized based on the Illumina pair-end (PE) sequencing data. The silica-gel dried leaves of *R. pectinata* were collected from China, Hainan Province, Baisha Xian, Nankai Xiang, Daoyin Cun (18°59'0.93"N, 109°19'45.88"E, 400 m). The voucher specimen (Z. L. Lin & Q. L. Wang 14022302) was deposited in the South China Botanical Garden Herbarium, Guangzhou, China. Total genomic DNA was isolated with a modified CTAB (Cetyl Trimethyl Ammonium Bromide) method (Doyle and Doyle 1987). Illumina Pair-end (PE) sequencing was performed on the Illumina HiSeq 2500 instruments at BGI-Wuhan. The sequenced clean PE reads were filtered using GetOrganelle pipeline (Jin et al. 2018) to get plastid-like reads. The filtered reads were assembled using SPAdes version 3.11.1 (Bankevich et al. 2012). The genome was automatically annotated using Plastid Genome Annotator (PGA) (Qu et al. 2019) coupled with manual corrections. The final complete plastome was deposited into GenBank (accession number MK946456).

The complete cp genome of *R. pectinata* was 149627 bp in length and presented a typical quadripartite structure including a large single copy (LSC) region (81,979 bp), a small single copy (SSC) region (16,626 bp), and two inverted repeat regions (IRs, 25,511 bp). The overall GC content of *R. pectinata* plastome was 38.0%. The *R. pectinata* plastome contained 114 different genes, including 80 protein-coding genes, 30 tRNA genes, and four rRNA genes. We also detected owing to the presence of internal stop codons, the gene *ycf15* was identified as pseudogene in the plastome of *R. pectinata*.

To reconstruct the phylogeny of Acanthaceae, nine Acanthaceae cp genomes were included in the phylogenetic analysis; plastomes of two relative families were used as out-groups (see Figure 1 for details). The plastomes (each excluding one IR) of all the species were aligned using MAFFT (version 1.3.7) (Katoh and Standley 2013) implemented in Geneious v. 11.0.4 and adjusted manually when necessary. The maximum-likelihood (ML) phylogeny was reconstructed using RAxML version 8.0.0 (Stamatakis 2014).

The present phylogenetic analyses strongly confirmed the monophyly of the family as previous reported (McDade et al. 2008) (Figure 1). Furthermore, all of the four Acanthaceae taxa formed a monophyletic group. Among the nine Acanthaceae species in this work, *R. pectinata* is most close to *Justicia leptostachya*, which all belong to Subtribe Justiciinae. All the nodes received 100% bootstrap support. The genome data in this paper can be subsequently used for phylogenetic studies in the genus *Rungia* and will contribute to further understanding of the phylogeny of Subtribe Justiciinae.

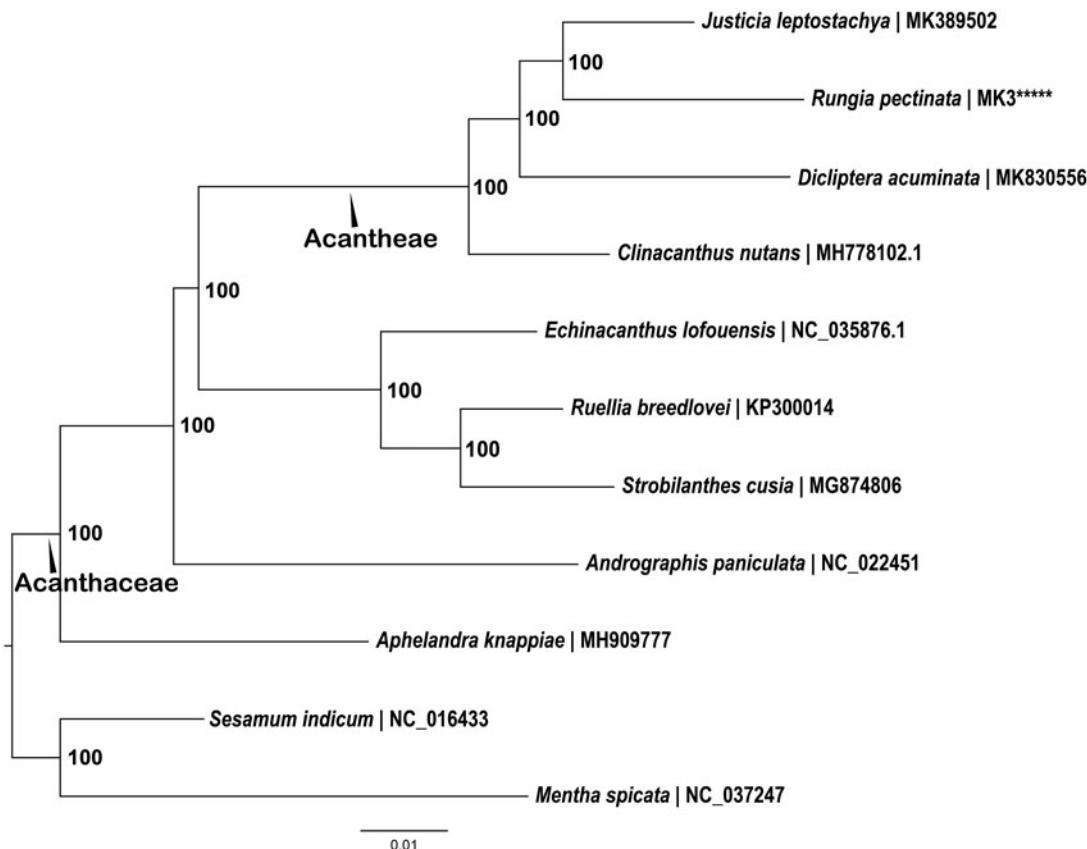


Figure 1. Maximum-likelihood (ML) phylogeny of Acanthaceae based on complete chloroplast genome sequences. Numbers at the right of nodes are bootstrap support values. GenBank accession numbers: *Andrographis paniculata* (NC_022451), *Ruellia breedlovei* (KP300014), *Strobilanthes cusia* (MG874806), *Echinacanthus lofoensis* (NC_035876.1), *Aphelandra knappiae* (MH909777), *Justicia leptostachya* (MK389502), *Clinacanthus nutans* (MH778102.1), *Dicliptera acuminata* (MK830556), *Sesamum indicum* (NC_016433), *Mentha spicata* (NC_037247), and *Rungia pectinata* (MK946456).

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

This work was financially supported by Guangdong Natural Science Foundation [Grant No. 2018A030310390], National Natural Science Foundation of China [Grant No. 31670191], and Southeast Asia Biodiversity Research Institute, Chinese Academy of Sciences [Y4ZK111B01].

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References

- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, et al. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol.* 19:455–477.
- Doyle JJ, Doyle JL. 1987. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochem Bull.* 19:11–15.
- Hu J, Deng Y, John RIW, Thomas FD. 2011. *Rungia* in Flora of China 19. In: Wu Z, Raven PH, Hong D, editors. *Flora of China*. Beijing: Science Press; St. Louis: Missouri Botanical Garden Press; p. 443–477.
- Jin JJ, Yu WB, Yang JB, Song Y, Yi TS, Li DZ. 2018. GetOrganelle: a simple and fast pipeline for de novo assembly of a complete circular chloroplast genome using genome skimming data. *bioRxiv*. 4:256479.
- Katoh K, Standley DM. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol Biol Evol.* 30:772–780.
- Lo HS. 1978. Notes on the genus *Rungia* (Acanthaceae) in China. *Acta Phytotaxon Sin.* 16:91–95.
- Mabberley DJ. (2017) Mabberley's plant-book: a portable dictionary of plants, their classifications and uses. 4th ed. Cambridge: Cambridge University Press.
- McDade LA, Daniel TF, Kiel CA. 2008. Toward a comprehensive understanding of phylogenetic relationships among lineages of Acanthaceae s.l (Lamiales). *Am J Bot.* 95:1136–1152.
- Qu XJ, Moore MJ, Li DZ, Yi TS. 2019. PGA: a software package for rapid, accurate, and flexible batch annotation of plastomes. *Plant Methods.* 15:1–12.
- Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics.* 30:1312–1313.