

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active. Contents lists available at ScienceDirect



**Chemico-Biological Interactions** 

journal homepage: www.elsevier.com/locate/chembioint



# 2-Pyridone natural products as inhibitors of SARS-CoV-2 main protease

# Katrina L. Forrestall<sup>a</sup>, Darcy E. Burley<sup>a</sup>, Meghan K. Cash<sup>a</sup>, Ian R. Pottie<sup>b,c</sup>, Sultan Darvesh<sup>a,b,d,\*</sup>

<sup>a</sup> Department of Medical Neuroscience, Faculty of Medicine, Dalhousie University, Halifax, Nova Scotia, B3H 4R2, Canada

<sup>b</sup> Department of Chemistry and Physics, Faculty of Arts and Science, Mount Saint Vincent University, Halifax, Nova Scotia, B3M 2J6, Canada

<sup>c</sup> Department of Chemistry, Faculty of Science, Saint Mary's University, Halifax, Nova Scotia, B3H 3C3, Canada

<sup>d</sup> Department of Medicine (Neurology), Faculty of Medicine, Dalhousie University, Halifax, Nova Scotia, B3H 4R2, Canada

#### ARTICLE INFO

Keywords: Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) COVID-19 Main protease (M<sup>pro</sup>) 2-Pyridone In silico molecular modelling AutoDock

#### ABSTRACT

The disease, COVID-19, is caused by the severe acute respiratory coronavirus 2 (SARS-CoV-2) for which there is currently no treatment. The SARS-CoV-2 main protease (Mpro) is an important enzyme for viral replication. Small molecules that inhibit this protease could lead to an effective COVID-19 treatment. The 2-pyridone scaffold was previously identified as a possible key pharmacophore to inhibit SARS-CoV-2 M<sup>pro</sup>. A search for natural, antimicrobial products with the 2-pyridone moiety was undertaken herein, and their calculated potency as inhibitors of SARS-CoV-2 M<sup>pro</sup> was investigated. Thirty-three natural products containing the 2-pyridone scaffold were identified from the literature. An in silico methodology using AutoDock was employed to predict the binding energies and inhibition constants (Ki values) for each 2-pyridone-containing compound with SARS-CoV-2 M<sup>pro</sup>. This consisted of molecular optimization of the 2-pyridone compound, docking of the compound with a crystal structure of SARS-CoV-2 M<sup>pro</sup>, and evaluation of the predicted interactions and ligand-enzyme conformations. All compounds investigated bound to the active site of SARS-CoV-2 Mpro, close to the catalytic dyad (His-41 and Cys-145). Thirteen molecules had predicted  $K_i$  values <1  $\mu$ M. Glu-166 formed a key hydrogen bond in the majority of the predicted complexes, while Met-165 had some involvement in the complex binding as a close contact to the ligand. Prominent 2-pyridone compounds were further evaluated for their ADMET properties. This work has identified 2-pyridone natural products with calculated potent inhibitory activity against SARS-CoV-2 Mpro and with desirable drug-like properties, which may lead to the rapid discovery of a treatment for COVID-19.

## 1. Introduction

In 2019, a novel human coronavirus, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2; NC\_045512) [1], spread rapidly throughout the global population with suspected human-to-human transmission via respiratory droplets [2,3]. Most SARS-CoV-2-positive individuals developed mild symptoms including upper respiratory tract illness. However, many people with concomitant medical conditions, including pulmonary and cardiovascular disease, hypertension, diabetes mellitus, as well as older adults, have developed severe symptoms or succumbed to the virus [4,5]. The disease, COVID-19, caused by this virus was declared a pandemic by the World Health Organization in March 2020. Over 34.4 million individuals have been infected globally so far, with a case-fatality ratio of 2.98% [6].

SARS-CoV-2 is a 29,903 bp single-stranded RNA coronavirus [7]. It shares 82% nucleotide identity with the related, human severe acute respiratory syndrome coronavirus (SARS-CoV), and both of these viruses belong to clade b of the genus *betacoronavirus* [2,3]. Both SARS-CoV and SARS-CoV-2 have structural spike proteins anchored to the viral envelope, which recognize and bind to the host cell's membrane protease,

https://doi.org/10.1016/j.cbi.2020.109348

Received 1 August 2020; Received in revised form 5 October 2020; Accepted 26 November 2020 Available online 2 December 2020 0009-2797/© 2020 Elsevier B.V. All rights reserved.

*Abbreviations*: ACE2, Angiotensin-converting enzyme 2; ADMET, Absorption, distribution, metabolism, excretion, toxicity; clogD, Calculated distribution coefficient; clogP, Calculated partition coefficient; F(20%), Oral bioavailability score to achieve 20% systemic concentration;  $\Delta G_{calc}$ . Gibbs free energy, calculated; Hbonds, Hydrogen bonds; HBD, Hydrogen bond donors; IC<sub>50</sub>, Half maximal inhibitory concentration;  $K_i$ , Inhibition constant; LD<sub>50</sub>, acute oral toxicity median lethal dose; logP<sub>app</sub>, Calculated Caco-2 permeability; logS, Calculated aqueous solubility; MPO, Multi-parametric optimization;  $M^{pro}$ , Main protease; PDB, Protein databank;  $pK_a$ , Acid dissociation constant; PL<sup>pro</sup>, Papain-like protease; QM, quantum mechanics; R, gas constant; RdRp, RNA-dependent RNA polymerase; RMSD, root mean square deviation; SARS-CoV, Severe acute respiratory syndrome coronavirus; SARS-CoV-2, Severe acute respiratory syndrome coronavirus 2; T, Absolute temperature; TPSA, Total polar surface area.

<sup>\*</sup> Corresponding author. Room 1308, Camp Hill Veterans' Memorial, 5955 Veterans' Memorial Lane, Halifax, Nova Scotia, B3H 2E1, Canada. *E-mail address:* sultan.darvesh@dal.ca (S. Darvesh).

angiotensin-converting enzyme 2 (ACE2) [3,8,9]. ACE2 activates the viral spike protein, allowing the virus to penetrate into the host cell [3, 10,11] and it is expressed on the ciliated epithelial cells of the human lungs and other tissues [12]. The binding affinity of spike-ACE2 was found to be 10-20-fold higher in SARS-CoV-2 than in SARS-CoV [13], which may be attributed to differences in the receptor-binding domain of the S1 region of the spike protein in both viruses [7]. Once inside the host cell, viral RNA is translated by the host ribosome into two polyproteins, which are proteolytically processed by coronavirus main protease (M<sup>pro</sup>) and papain-like protease (PL<sup>pro</sup>) [14,15], for assembly of new viral particles [16]. In order to replicate the RNA genome, the virus encodes an RNA-dependent RNA polymerase (RdRp). Spike, M<sup>pro</sup>, PL<sup>pro</sup> and RdRP are essential proteins for coronaviruses to infect and replicate, thus making them suitable therapeutic targets. As such, M<sup>pro</sup> has been characterized as one of the most attractive targets for the design of coronavirus inhibitors, by using small molecules to block enzyme activity and prevent viral replication [17].

 $M^{pro}$  has a unique cleavage specificity, Leu-Gln↓(Ser,Ala,Gly), that no other human proteases possess, safeguarding potential  $M^{pro}$  inhibitors from being harmful to humans [18]. The crystal structure of SARS-CoV-2  $M^{pro}$  reveals a dimer that shares 96% amino acid identity with SARS-CoV  $M^{pro}$  [18]. SARS-CoV-2  $M^{pro}$  has chymotrypsin- and picoronavirus 3C protease-like domains I and II that are six stranded anti-parallel β-barrels containing the substrate binding site between them [19]. Domain III of  $M^{pro}$  regulates dimerization by a salt-bridge interaction between Glu-290 of one protomer and Arg-4 of the other protomer [20]. Dimerization is important for the catalytic activity as it shapes the pocket of the substrate-binding site [21]. Catalytic efficiency of  $M^{pro}$  was shown to be slightly increased in SARS-CoV-2 than SARS-CoV [18], while the maximal accessible volume of the active site is decreased from SARS-CoV to SARS-CoV-2 [22]. For this reason, inhibitors for SARS-CoV might not be as effective for SARS-CoV-2.

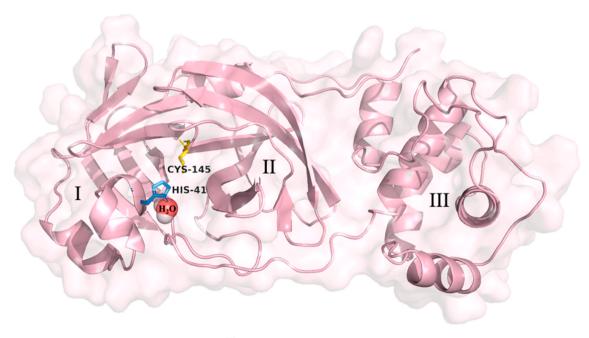
There are several crystal structures that have been determined for SARS-CoV-2 M<sup>pro</sup>. The x-ray crystallographic structure of ligand-free SARS-CoV-2 M<sup>pro</sup> at room temperature (293K) revealed structural plasticity when compared to the x-ray crystallographic structure at low-temperature (100K) [23]. This work also revealed a water molecule deep in the active site, close to His-41, that aids in the catalytic activity

of this enzyme [23]. The crystal structure of SARS-CoV-2 M<sup>pro</sup> (PDB: 6WQF) is depicted in Fig. 1 with its catalytic dyad and the water molecule. This crystal structure has been regarded as the most appropriate model for performing docking studies as it is more representative of physiological conditions [22,23].

Previously,  $\alpha$ -ketoamides have been synthesized to inhibit the  $M^{\text{pro}}$ of beta- and alpha-coronaviruses, although, the half-life and solubility of these compounds in plasma was unsatisfactory [19]. To increase the half-life, solubility and anti-viral activity, α-ketoamides were modified to incorporate a pyridone ring that prevents cleavage from cellular proteases [18]. These new compounds were observed to increase the plasma half-life in mice 3-fold with significantly improved kinetic plasma and thermodynamic solubilities. To identify potential binding sites, the crystal structure of SARS-CoV-2 M<sup>pro</sup> was complexed with the pyridone-containing compound (PDB: 6Y2F) [18]. The 2-pyridone compound was bound to the shallow substrate-binding site of SARS-CoV-2 M<sup>pro</sup> containing the catalytic dyad (His-41 and Cys-145) and inhibited viral activity [18]. Favourable ADMET and pharmacokinetic properties (e.g. lung tropism) and tolerability when administered subcutaneously and/or via inhalation, gave further credence for the development of 2-pyridone compounds as inhibitors of SARS-CoV-2 [18].

Natural products and their analogues have played a significant role in the development of medicines to treat a wide variety of conditions, including infections [24]. The 2-pyridone moiety is observed in a wide variety of simple and complex organic molecules in nature. Many organisms retain the ability to synthesize, degrade, or utilize molecules containing 2-pyridone rings [25,26]. In humans, 2-pyridone compounds have been identified as metabolites of regular cell function [27–29]. A number of 2-pyridone-containing natural products have antimicrobial, -viral, -inflammatory, -proliferative, -diabetic and -cancer properties making them attractive molecules for drug development [30–34]. The chemical properties of 2-pyridones and their presence in many bioactive compounds make them a valuable resource for drug design [33,34]. Furthermore, the 2-pyridone moiety has been shown to restrict degrees of freedom within chemical structures, while mimicking a peptide backbone [18].

The present work was undertaken to identify potential new



**Fig. 1.** The crystal structure of the SARS-CoV-2 main protease (M<sup>pro</sup>) monomer (PDB: 6WQF) is comprised of three domains (I–III). The domains, amino acid residues of the catalytic site, His-41 (yellow), Cys-145 (skyblue) and embedded catalytic water molecule (red/white spheres) as suggested by Kneller et al. [23], are depicted. Figure was developed using PyMOL [43].

Calculated binding energies, inhibition constants ( $K_i$  values) <1  $\mu$ M and all-atom RMSD identified for the interaction between 2-pyridone natural products and SARS-CoV-2 M<sup>pro</sup>.

Compound Name [Reference]	Structure (Compound Number)	SARS-CoV-2 M <sup>pro</sup>	SARS-CoV-2 M <sup>pro</sup>				
		Calculated Binding Energy ( $\Delta G_{calc}$ ; kJ/mol)	Calculated $K_i$ (µM)	RMSD (Å)			
Fusapyridon A-2 [67]		-43.85	0.02086	2.00 (Cluster: 13/50)			
llicicolin H [68]	$1 \\ \stackrel{\text{Ho}}{\underset{\underset{\underset{\underset{\underset{\underset{\underset{\underset{\underset{\underset{\underset{\underset{\underset{\underset{\underset{\underset{\underset{$	-43.39	0.02498	1.98 (Cluster: 11/50)			
Fusapyridon B-2 [67]		-42.72	0.03258	2.00 (Cluster: 12/50)			
Leporin A [69]		-39.58	0.11584	1.02 (Cluster: 29/50)			
Fusapyridon A-1 [67]		-38.53	0.17601	1.69 (Cluster: 11/50)			
Leporin B [70]		-37.91	0.22961	0.26 (Cluster: 40/50)			
(+) Epipyridone [71]		-37.61	0.25692	0.19 (Cluster: 37/50)			
(+) Apiosporamide [72]		-36.65	0.38145	1.12 (Cluster 7/50)			
Torrubiellone E [73]		-36.48	0.40795	1.60 (Cluster: 8/50)			
Farinosone A [74]		-36.28	0.44335	0.85 (Cluster: 5/50)			
Farinosone B [75]	$\begin{array}{c} 10 \\ HO \\ H$	-35.82	0.5296	0.60 (Cluster: 10/50)			
YM-215343 [76]		-34.85	0.78177	1.23 (Cluster: 13/50)			
Cordypyridone D [77]	12 12 12 12 12 12 10 10 10 10 10 10 10 10 10 10	-34.43	0.9259	0.17 (Cluster: 31/50)			
	13						

inhibitors of SARS-CoV-2 M<sup>pro</sup> from the trove of natural products containing a 2-pyridone scaffold. The intention of the present study is to allow for a streamlined and rapid approach in developing therapeutics towards COVID-19.

#### 2. Materials and methods

#### 2.1. Identification of 2-pyridone natural products

The natural products containing a 2-pyridone moiety assessed in the present study were selected through a literature search of 2-pyridone alkaloids discovered from the 1960s to present. The Chemical Abstract Service database, SciFinder (https://scifinder.cas.org), was utilized for conducting structure and keyword ("bioactive 2-pyridones", "2-pyridone alkaloids", "antimicrobial 2-pyridones", "2-pyridone natural products", "antibacterial 2-pyridones" and "antifungal 2-pyridones") searches. An initial investigation provided a review from Jessen and Gademann from which a list of notable, bioactive compounds was developed [35]. Additional compounds, discovered following the publication of this review, were identified through further literature searches on compounds containing a 2-pyridone moiety. The 2-pyridone natural products chosen for the current study were selected based on their putative antimicrobial activity. Natural products with a 2-pyridone scaffold, as well as their putative biological actions, are summarized in Appendix A Supplementary Data (Table A1).

# 2.2. In silico binding studies

Due to the recently identified conformational flexibility of the SARS-CoV-2  $M^{\text{pro}}$  active site and the revelation of a water molecule aiding in catalysis, the room temperature, ligand-free  $M^{\text{pro}}$  structure with an embedded water molecule has been suggested to be the most physiologically relevant crystal structure for performing molecular docking (PDB: 6WQF) [23]. Therefore, this structure was selected for *in silico* binding studies. The calculated binding energies and inhibition constants ( $K_i$  values) for the selected 2-pyridone natural products with SARS-CoV-2  $M^{\text{pro}}$  were obtained using a molecular modelling procedure. This process consisted of optimization of the molecular geometry of each natural product using GaussView 5.0 [36] and docking each minimized molecular structure with a crystal structure of SARS-CoV-2  $M^{\text{pro}}$  (PDB: 6WQF) using AutoDock 4.2.6 software equipped with AutoDock Tools (Scripps Research Institute) [37].

# 2.2.1. Molecular geometry optimization of 2-pyridone natural products

In order to minimize the molecular geometry of compounds assessed, each 2-pyridone-containing natural product was constructed and optimized using a quantum mechanics (QM) calculation (Hartree-Fock using the 3-21G basis set) within GaussView 5.0 [36]. The minimized structure was saved as a .pdb file and was subsequently used for docking procedures.

# 2.2.2. Docking of minimized natural products with crystal structure of SARS-CoV-2 $M^{\rm pro}$

To estimate the potential interaction and conformation of the minimized 2-pyridone natural product and SARS-CoV-2 M<sup>pro</sup> complex, a published molecular modeling procedure using AutoDock 4.2 Software was conducted [38,39]. The published SARS-CoV-2 M<sup>pro</sup> crystal structure (PDB code: 6WQF) was obtained from the protein databank (http://www.rcsb.org/) and uploaded to UCSF Chimera (https://www. cgl.ucsf.edu/chimera/). Water molecules were removed from this structure, except for the oxygen atom located 2.8 Å from His-41, as suggested by Kneller et al. [23]. Hydrogen atoms were added to the oxygen to complete valency of the embedded water, and the structure was saved as a .pdb file. The structure was then uploaded into AutoDock 4.2.6 and each non-polar hydrogen was merged with its bonding partner. Gasteiger partial charges for all atoms within the structure were computed and added to the enzyme. The resulting file was saved as a . pdb file.

The data from the geometry optimization of the 2-pyridone natural product was uploaded into AutoDock 4.2.6. Immediately, all non-polar hydrogen atoms were merged with their bonding partner, aromatic carbon atoms were identified, Gasteiger partial charges for all the atoms within the 2-pyridone natural product were computed and added to the molecule and bonds that could rotate were identified. The data was saved as a .pdbqt file.

Both .pdbqt files were uploaded to AutoDock and the grid maps were produced around the active site of SARS-CoV-2  $M^{pro}$ . The nucleophilic sulfur atom in Cys-145 (X: 16.67 Å, Y: 3.63 Å, Z: 12.50 Å) was located and used as the center of the grid map. The grid map size was 120 grid points  $\times$  120 grid points  $\times$  120 grid points with a grid-point spacing of 0.375 Å. The docking calculation was subsequently performed using the Lamarckian genetic algorithm [38], and each 2-pyridone natural product was evaluated over 50 runs. The initial population size of each run was 150. During each run, the maximum number of generations simulated was 27,000, while the maximum number of energy evaluations to identify the fitness of the 2-pyridone : SARS-CoV-2  $M^{pro}$  interaction was 2,500,000. All predicted binding energies were calculated using a semiempirical free energy force field [38].

The most probable docking location and the lowest binding energy predicted ( $\Delta G_{calc}$ , Table 1) between each 2-pyridone : SARS-CoV-2 M<sup>pro</sup> complex were visualized within AutoDock. In order to quantify the performance of the docking calculation, a known procedure was used wherein each of the 50 docking runs were evaluated for repetition in the location and orientation of the ligand and placed into groups (clusters) within an all-atom root mean square deviation (RMSD) of  $\leq 2$  Å [40,41]. The cluster data produced from each run was placed on a scatter graph to demonstrate the accuracy of the calculation for each 2-pyridone assessed (Appendix A, Supplementary Data, Figure A1). The best cluster, i.e. the cluster containing the highest number of docking runs or the lowest binding energy (where clusters contained the same number of runs) was selected to represent the 2-pyridone : SARS-CoV-2 Mpro complex [42]. The complex with the lowest binding energy from the best cluster was saved as a .pdbqt file and evaluated using the open source visualization software, PyMOL [43]. The amino acid residues that interact with the 2-pyridone natural product, produce hydrogen bonds (H-bonds) [44,45] or surround the docked 2-pyridone compound were identified (Appendix A, Supplementary Data, Table A1). The calculated  $K_i$  values were obtained using  $\Delta G_{calc} = RTlnK_i$ ; where R is the gas constant, 8.314 J  $K^{-1}$ mol<sup>-1</sup>, and T is the absolute temperature (298.15 K), based on the temperature for enzyme crystallography (Tables 1-4) [46,47].

# 2.3. Calculation of ADMET pharmacokinetic properties

To quantify the suitability of the most favourable 2-pyridone inhibitors as drugs for SARS-CoV-2 M<sup>pro</sup>, several absorption, distribution, metabolism, excretion, and toxicity (ADMET) characteristics were calculated. The multi-parameter optimization (MPO) algorithm, was used to score the most favourable 2-pyridone compounds with the lowest calculated  $K_i$  values (1–10) and estimate their theoretical ability to be used as a drug. MPO is based on six pharmacokinetic parameters: calculated partition coefficient (clogP), calculated distribution coefficient (clogD), total polar surface area (TPSA), number of H-bonding donors (HBD), acid dissociation constant  $(pK_a)$ , and molecular weight [48]. These chemical properties were calculated using their corresponding plugin and default settings in MarvinSketch (ChemAxon, https ://www.chemaxon.com) [49]. pKa values were likewise determined using the default settings for macro mode and static acid/base prefix at room-temperature (298 K). Each 2-pyridone compound was scored for a total MPO value from 0.0 to 6.0 (Table 5), where values >3.0 are considered acceptable for compounds possessing drug-like properties [48]. Theoretical aqueous solubility for compounds 1-10 were

Calculated binding energies, inhibition constants ( $K_i$  values) 1–10  $\mu$ M and all-atom RMSD identified for the interaction between 2-pyridone natural products and SARS-CoV-2  $M^{pro}$ .

Compound Name [Reference]	Structure (Compound Number)	SARS-CoV-2 M <sup>pro</sup>				
		Calculated Binding Energy ( $\Delta G_{calc}$ ; kJ/mol)	Calculated $K_i$ (µM)	RMSD (Å)		
Cordypyridone C (N-methoxyPF1140) [77]		-33.05	1.62	0.12 (Cluster: 35/ 50)		
Pyridovericin [78,79]		-31.55	2.99	1.15 (Cluster: 7/50)		
Fusapyridon B-1 [67]		-30.79	4.04	1.99 (Cluster: 5/50)		
Akanthomycin [80]	16 (1) (1) (1) (1) (1) (1) (1) (1)	-30.25	5.03	0.51 (Cluster: 19/ 50)		
Aurodox [81]		∽ -29.87	5.87	0.00 (Cluster: 1/50)		
N-deoxyakanthomycin [82]		-29.29	7.38	1.15 (Cluster: 20/ 50)		
PF1140 [83]	19 $ \begin{array}{c}                                     $	-28.74	9.20	0.25 (Cluster: 41/ 50)		
Pyridoxatin [84]		-28.70	9.40	0.17 (Cluster: 18/ 50)		
	он 21					

calculated using MarvinSketch, where values >10 µg/mL are considered optimal [49,50]. Additional ADMET properties (i.e. oral bioavailability F(20%), acute oral toxicity (LD<sub>50</sub>), and Caco-2 permeability) were generated using the ADMETLab open-source web calculator (https://admet.scbdd.com/) [51,52]. Oral bioavailability is expressed as the fraction of drug required to achieve 20% in systemic circulation (>20% optimal) [53,54]. Toxicity is expressed as mg/kg where values >500 mg/kg are considered optimal [55,56]. Membrane permeability expressed as logP<sub>app</sub> values (m/s) for the human colorectal cell line, Caco-2 (>-5.15 cm/s is optimal) [57]. The theoretical subcellular localization of compounds **1–10**, were also calculated using the admetSAR2.0 open-source web calculator (https://lmmd.educ.cn/adme tsar2.0/) [58,59].

# 2.4. Comparison of top 2-pyridones natural products with known drugs

To display the relevance and drug-suitability of the top 2-pyridonecontaining compounds investigated herein, calculated binding energies and MPO scores of the 2-pyridone compounds ( $K_i$  values  $\leq 1 \mu$ M) were compared with that of a selection of known drugs suggested for repurposing/repositioning against SARS-CoV-2 M<sup>pro</sup> (Table 6). The drugs chosen for comparison were selected based on their consideration in several COVID-19-related *in silico* and drug repurposing studies [60–63]. These known compounds offer a variety of therapeutic classes, from antivirals to neuroleptic drugs.

#### 3. Results

#### 3.1. Search for natural products with the 2-pyridone moiety

From the literature search, a total of 61 natural products containing the 2-pyridone moiety were identified. The putative biological actions of many of these compounds were antimicrobial (anti-bacterial, -viral and -fungal), anti-cancer, and/or anti-proliferative or cytotoxic. Given the objective of the present study to assess potentially efficacious inhibitors of SARS-CoV-2 M<sup>pro</sup> with limited side-effects, only compounds that possessed anti-microbial properties were evaluated herein. For the putative mechanism of action of the selected compounds, see Appendix A, Supplementary Data Table A1.

## 3.2. Evaluation and validation of docking poses

To validate the AutoDock procedure utilized in the present work, a cluster plot was generated summarizing the docking data for each of the 33 compounds assessed (Appendix A, Supplementary Data, Figure A1). The cluster plots reveal that most of the 2-pyridone natural products (30 out of 33 compounds) produced their largest cluster with a similarly low binding energy to that of their most favourable (lowest  $\Delta G$ ) run. Furthermore, 27 of the 2-pyridones evaluated showed RMSD values  $\leq 2$  Å, indicating good correlation within the best cluster result. The data sets for Aurodox **18**, Kirromycin **30**, and Factumycin **31** did not generate any clusters greater than a single run. Within the best cluster for each compound, the conformation showing the lowest binding energy was

Calculated binding energies, inhibition constants ( $K_i$  values) 10–40  $\mu$ M and all-atom RMSD identified for the interaction between 2-pyridone natural products and SARS-CoV-2  $M^{pro}$ .

Compound Name [Reference]	Structure (Compound Number)	SARS-CoV-2 M <sup>pro</sup>				
		Calculated Binding Energy ( $\Delta G_{calc}$ ; kJ/mol)	Calculated $K_i$ (µM)	RMSD (Å)		
Cordypyridone B (8-methylpyridoxatin) [77]		-28.33	10.87	0.53 (Cluster: 28/50)		
Cordypyridone A (8-methylpyridoxatin) [77]		-28.16	11.73	0.63 (Cluster: 27/50)		
Harzianopyridone [85,86]		-28.03	12.25	1.54 (Cluster: 7/50)		
(R) Flavipucine [87]	24	-27.91	12.96	0.15 (Cluster: 32/50)		
Atpenin A4 [88]		-27.45	15.56	1.17 (Cluster: 5/50)		
(S) Flavipucine [87]		-27.45	15.64	0.12 (Cluster: 15/50)		
Torrubiellone A [89]		-27.11	17.90	0.39 (Cluster: 3/50)		
Atpenin A5 [88]	28 HO I I I I I I I I I I I I I I I I I I I	-25.77	30.32	1.60 (Cluster: 4/50)		

#### Table 4

Calculated binding energies, inhibition constants (K<sub>i</sub> values) 40–245 µM and all-atom RMSD identified for the interaction between 2-pyridone natural products and SARS-CoV-2 M<sup>pro</sup>.

Compound Name [Reference]	Structure (Compound Number)	SARS-CoV-2 M <sup>pro</sup>				
		Calculated Binding Energy ( $\Delta G_{calc}$ ; kJ/mol)	Calculated $K_i$ (µM)	RMSD (Å)		
Kirromycin [90]		-24.85	43.91	0.00 (Cluster: 1/50)		
Factumycin [91]	$ \begin{array}{c} \begin{array}{c} \begin{array}{c} \\ \\ \end{array} \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ $	-24.56	49.44	0.00 (Cluster: 1/50)		
Atpenin B [88]		-23.72	69.99	1.23 (Cluster: 5/50)		
Ricinine [34,92]		-20.62	244.7	0.09 (Cluster: 46/50)		
	ї 33					

further analyzed in PyMOL.

# 3.3. Calculated binding energies and K<sub>i</sub> values

A total of 33 natural products containing the 2-pyridone moiety were docked to SARS-CoV-2  $M^{pro}$  using AutoDock [38,39]. Calculated binding energies and  $K_i$  values for each top-ranked 2-pyridone natural product complexed with SARS-CoV-2  $M^{pro}$  are summarized in Tables 1–4. The binding energies ranged from -43.85 to -20.62 kJ/mol and the  $K_i$  values ranged from 0.021  $\mu$ M–244.70  $\mu$ M. Compound 1 had the lowest

binding energy (-43.85 kJ/mol) and calculated  $K_i$  value (0.021  $\mu$ M). Compounds **2–13** (Table 1) also had strong binding energies (-43.39 to -34.43 kJ/mol), with small calculated  $K_i$  values (0.025 to 0.926  $\mu$ M). The stereochemistry of the chiral C-3 methyl substituent in the tetrahydropyran ring of fusapyridon A (**1** and **5**) and fusapyridon B (**3** and **16**) was not indicated in the literature; therefore, both configurations of the methyl substituent (axial and equatorial) were subjected to the docking procedure.

Compounds **14–21** had affinity for SARS-CoV-2 M<sup>pro</sup> with binding energies ranging from -33.05 to -28.70 kJ/mol and the calculated  $K_i$ 

ADMET properties of the ten 2-pyridone natural products with  $K_i$  values,  $<1 \mu$ M when bound to SARS-CoV-2 M<sup>pro</sup>.

Compound	MPO	TPSA (Å)	HBD	pK <sub>a</sub>	clogP	clogD at pH 7.40	logS (µg/mL)	F(20%)	LD <sub>50</sub> (mg/kg)	$logP_{app}$ (m/s)
1	2.00	65.1	0	18.8	5.07	5.07	0.361	0.386	185	-0.0473
2	2.27	86.6	3	8.93	4.84	4.83	0.0119	0.465	342	-0.0529
3	2.00	66.5	0	27.2	5.71	5.71	0.367	0.312	176	-0.0476
4	3.15	38.8	0	23.8	4.53	4.53	0.123	0.619	552	-0.0461
5	2.00	65.1	0	18.8	5.07	5.07	0.361	0.415	161	-0.0473
6	4.70	49.8	1	6.38	4.15	3.11	0.323	0.632	150	-0.0492
7	3.23	58.6	2	9.18	4.28	4.27	0.103	0.602	234	-0.0506
8	3.10	119	4	8.87	1.84	1.83	0.349	0.424	229	-0.0558
9	3.78	118	4	-1.84	1.60	0.39	171	0.473	444	-0.0533
10	2.48	86.6	3	8.86	4.89	4.88	0.00128	0.499	867	-0.0523

# Table 6

Comparison of calculated binding energies and MPO scores from select 2-pyridone natural products with potential drugs targeting SARS-CoV-2 M<sup>pro</sup> suggested in the literature for possible treatment of COVID-19.

2-Pyridone Natural Products	Binding Energy (kJ/mol)	MPO	Repurposed Drugs Against SARS-CoV-2 M <sup>pro</sup>	Binding Energy (kJ/mol)	MPO
1	-43.85	2.00	Darunavir	-48.10 [61]	2.76
2	-43.39	2.27	Lopinavir	-40.58 [62]	2.50
3	-42.72	2.00	Saquinavir	-36.30 [61]	2.90
4	-39.58	3.15	Chlorpromazine	-34.73 [ <mark>63</mark> ]	2.58
5	-38.53	2.00	Festinavir	-29.92 [62]	4.53
6	-37.91	4.70	Oseltamivir	-29.33 [62]	5.48
7	-37.61	3.23	Remdesivir	-28.33 [62]	3.00
8	-36.65	3.10	Ribavirin	-26.36 [60]	4.00
9	-36.48	3.78	Azithromycin	-25.69 [62]	2.38
10	-36.28	2.48	Favipiravir	-20.00 [62]	4.50

values ranging from 1.62 to 9.40  $\mu$ M (Table 2). Compounds **22–29** had binding energies ranging from –28.33 to –25.77 kJ/mol, and larger calculated  $K_i$  values ranging from 10.87 to 30.32  $\mu$ M (Table 3); while compounds **30–33** had the weakest binding energies (–24.85 to –20.62 kJ/mol) and the largest calculated  $K_i$  values (43.91–244.70  $\mu$ M) (Table 4).

#### 3.4. Amino acid interactions

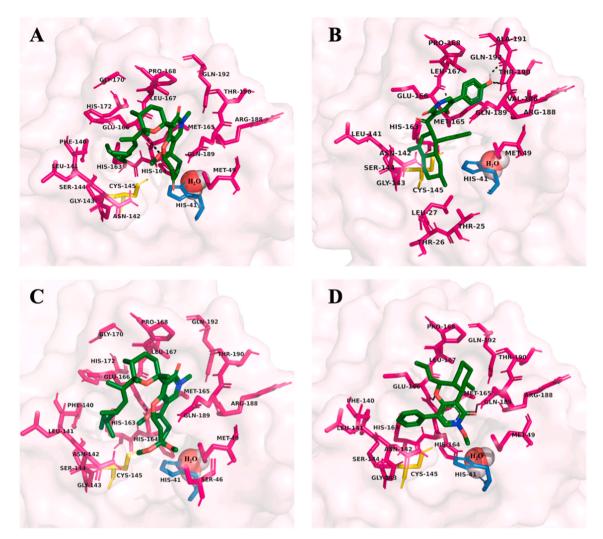
The 2-pyridone natural products investigated in this study exhibited very similar interactions with amino acid residues in the SARS-CoV-2 M<sup>pro</sup> active site. All 33 compounds showed binding inside of the active site gorge. Only compound 9 showed no H-bonding. The recurrent Hbonding amino acid residues were: Glu-166 (22/33), Asn-142 (11/33), Gln-189 (9/33), His-163 (7/33), Gln-192 (7/33), Thr-190 (7/33). The prominent amino acid residues in close contact to the bound 2-pyridone compounds were: Met-165 (33/33), Met-49 (32/33), His-41 (30/33), Cys-145 (29/33), His-164 (28/33), Leu-167 (24/33), Phe-140 (23/33), Gln-189 (22/33), Leu-141 (20/33), Gly-143 (19/33), Pro-168 (19/33), His-163 (18/33), Arg-188 (18/33), Ser-144 (17/33), Asn-142 (16/33), His-172 (13/33), Thr-190 (13/33), Gln-192 (12/33), Ala-191 (10/33), Glu-166 (9/33). The amino acid residues that were most common for both H-bonding and/or as close contacts were: Met-165 (33/33), His-41 (32/33), Met-49 (32/33), His-164 (31/33). Glu-166 (31/33), Gln-189 (31/33). Almost all compounds exhibited bonding or close contact interactions with the catalytic dyad (His-41 and Cys-145). For the compounds with the most favourable binding energies and  $K_i$  values (1–13), residues Gln-189 (12/13), His-164 (12/13), and Arg-188 (10/13) were recurrent. The details regarding amino acid interactions for each of the 2-pyridone compounds are summarized in Appendix A, Supplementary Data, Table A1. The amino acid residues in the SARS-CoV-2 M<sup>pro</sup> active site that interact with the four most potent compounds assessed in this study are depicted in Fig. 2.

The observed H-bonding interactions primarily involved the 2-pyridone ring portion of the natural product molecules. Frequently, Hbonding occurred between the previously highlighted amino acid residues and the 2-pyridone carbonyl (19/33), the C-4 hydroxy/oxygen group (18/33), or the pyridone amine/oxime (8/33). The 13 compounds that exhibited the most favourable calculated  $K_i$  values (<1  $\mu$ M) were mostly ring-based structures containing little or no chains (Tables 1 and A1). These compounds also tended to possess carbonyl, hydroxy, or ether groups at C-3 and a 6 membered-ring functional group off C-5. Interestingly, compound **6** (leporin B) showed no H-bonding despite favourable binding affinity. H-bond interactions are shown in Fig. 2.

# 3.5. Drug suitability

The ten 2-pyridone natural products (1-10) that produced the lowest K<sub>i</sub> values when complexed with SARS-CoV-2 M<sup>pro</sup> were evaluated for their fitness as potential drugs to treat COVID-19. The ADMET properties identified through the MPO score (clogP, clogD, TPSA, HBD, pKa, and molecular weight) as well as theoretical aqueous solubility, oral bioavailability F(20%), LD<sub>50</sub>, subcellular localization, and membrane permeability were calculated and are shown in Table 5. Only compounds 4, 6, 7, 8, and 9 produced MPO scores  $\geq$  3.0. Leporin B (6) had the largest MPO (4.70). All compounds except 9 showed low theoretical aqueous solubility. The oral bioavailability values were all favourably >20%, with Leporin A (4) and B (6) exhibiting the greatest F(20%) values (0.632 and 0.619). All compounds displayed very similar membrane permeability values ranging from -5.578 to -4.614 cm/s, with compounds 1, and 3-7 showing optimal values. Most compounds were indicated to possess mild-toxicity; however, compounds 4 and 10 were in the low-toxicity range. All compounds showed subcellular localization to the mitochondria, except for compound 6 which was suggested to further localize to the plasma membrane.

When compared with drugs currently under repurpose for the inhibition of SARS-CoV-2  $M^{pro}$ , compounds **1–10** exhibited similar, if not, slightly greater binding energies (Table 6). A comparable range of MPO values were identified for the ten 2-pyridone compounds (2.00 to 4.70) and the repurposed drugs targeting SARS-CoV-2  $M^{pro}$  (2.38 to 5.48).



**Fig. 2.** The most potent 2-pyridone natural products, compounds 1–4 (A–D), are docked with SARS-CoV-2 main protease ( $M^{\text{pro}}$ ) (PDB: 6WQF). The structures, binding energies and inhibition constants ( $K_i$  values) for 1–4 are summarized in Table 1, as well as their ADMET properties in Table 5. The H-bonding (dashed black lines) and close contact amino acid residues for 1–4 are depicted in their corresponding panel, as well as the catalytic dyad, His-41 (skyblue), Cys-145 (yellow), and embedded catalytic water (red/white spheres) as suggested by Kneller et al. [23]. Figures were generated using PyMOL [43].

#### 4. Discussion

The novel coronavirus, SARS-CoV-2, has resulted in the global pandemic, COVID-19 [1–3]. This virus is highly contagious [2,3], more so than other coronaviruses [64] and can cause serious and potentially fatal respiratory conditions [4,5]. Currently, there are no vaccines or treatments available to combat SARS-CoV-2. As a result, a global effort has been initiated in search of a cure by repurposing or repositioning existing drugs [65,66].

While a number of unique SARS-CoV-2 proteins are being evaluated for drug development to treat COVID-19, M<sup>pro</sup> has been identified as an appropriate target and a number of inhibitors have been evaluated not only for SARS-CoV-2 but for SARS-CoV as well [17]. In the literature, 2-pyridone molecules have shown promise for the inhibition of SARS-CoV-2 M<sup>pro</sup>. Thus, in the present work, natural products that contain the 2-pyridone scaffold were evaluated through *in silico* studies. These molecules, most of which are of fungal origin, have shown putative antimicrobial properties against bacterial and fungal strains [35]. Given the urgency of discovering a cure for COVID-19, these natural products show promise for further evaluation as therapeutic agents for SARS-CoV-2.

Using the *in silico* AutoDock methodology, selected compounds from the literature were docked and evaluated for their potential inhibitory properties against SARS-CoV-2 M<sup>pro</sup> through calculation of binding energies and  $K_i$  values. To validate the *in silico* binding procedure, each 2-pyridone natural product was docked with SARS-CoV-2 M<sup>pro</sup> to produce a cluster plot. The cluster plots showed convergence of docking poses to the lowest possible binding energy, while the docking conformations within the best cluster had acceptable RMSD values ( $\leq 2$  Å) (Appendix A, Supplementary Data, Figure A1). In this study, 33 2-pyridone molecules were identified from the literature for investigation, 13 of which proved to have calculated  $K_i < 1 \ \mu$ M (compounds 1–13; Table 1), indicating their promise as inhibitors of SARS-CoV-2 M<sup>pro</sup>.

Based on the bonding interactions of the 2-pyridone natural products investigated, it is evident that the 2-pyridone ring is a key scaffold that aids in inhibitor binding to SARS-CoV-2 M<sup>pro</sup>. A previous investigation of  $\alpha$ -ketoamide inhibitors of M<sup>pro</sup> highlighted the potential importance of the 2-pyridone moiety in inhibitor design [18]. In the present work, the 2-pyridone carbonyl and nitrogen were shown to be important atoms in the H-bonding interactions at the active site gorge among all compounds studied. In addition, compounds also exhibited prominent interactions with a hydroxy moiety at the C-4 carbon of the 2-pyridone ring. This suggests that, specifically, 4-hydroxy-2-pyridones may be successful inhibitors of SARS-CoV-2. Moreover, these results suggest that greater affinity for SARS-CoV-2 M<sup>pro</sup> is achieved with the presence of multiple ring structures (e.g. compound 1,  $K_i$  value of 0.021 µM) rather than large portions of carbon chains (e.g. compound **30**,  $K_i$  value of 43.91 µM) (Tables 1 and 4). As well, compounds 1–13 were among the most appropriately sized compounds for binding to and filling the active site gorge of  $M^{\text{pro}}$ . The weakest binding 2-pyridone was ricinine (**33**), which was far too small to fill the active site and to engage with the key amino acids previously identified to be important for enzymatic activity. Similarly, compounds **18**, **30**, and **31** were the largest compounds assessed. Their cluster results indicated poor docking with the SARS-CoV-2  $M^{\text{pro}}$  active site, likely due to the large, bulky carbon chains of these compounds.

It has been suggested that the active site of SARS-CoV-2  $M^{pro}$  contains a catalytic dyad, His-41 (base) and Cys-145 (nucleophile), with an embedded water molecule (Fig. 1) that enables the enzyme to catalyse the hydrolysis of polyproteins and generate the replicase/transcriptase complex for SARS-CoV-2 replication [14-16,23]. The results of the current study show that Glu-166 was present in almost all of the 2-pyridone binding studies conducted, either as a close contact residue or as a participant in H-bonding, as shown previously [18]. This residue may therefore act as a proton donor in hydrolysis, an alternative to the embedded water molecule, to provide increased drug potency. In addition, the frequent occurrence of ligand interactions with Met-49 and Met-165 and for compounds 1-33 and His-164, Arg-188, and Gln-189 for compounds 1-13, also suggests that these residues may be important mediators in inhibitor binding to Mpro. As well, almost all 2-pyridone natural products investigated in this study engaged in interactions with the catalytic dyad as proposed.

The ten top 2-pyridone-containing compounds ( $K_i$  values  $\leq 1.0 \mu$ M) were further evaluated on their suitability for future research as potential drugs to treat COVID-19. ADMET properties including clogD, clogP, HBD, pKa, TPSA, aqueous solubility, oral bioavailability, acute oral toxicity, subcellular localization, and membrane permeability were calculated. The ADMET properties summarize several pharmacokinetic parameters that are considered to provide a basis on the drug-like capacity of a theoretical pharmaceutical [53-59]. Other assessments that include ADMET parameters, such as the MPO calculation have been developed to help theorize the drug-suitability of any given compound [48]. Of the top ten 2-pyridone natural products, compounds 4, 6, 7, 8, and 9 produced favourable MPO values, with leporin B (6) producing the largest MPO value (4.70). Further ADMET analysis showed that all compounds had low, but suitable aqueous solubility [50], favourable oral bioavailability values [53,54], and mild to low toxicity [55,56]. Based on the data for all ADMET parameters (Table 5), only compound (4) was in the optimal range of all parameters, suggesting leporin A (4) may be the most suitable compound for further drug evaluation as an inhibitor of SARS-CoV-2 Mpro.

Additionally, a comparison of the pharmacokinetic properties of the top ten 2-pyridone natural products ( $K_i < 1.0 \mu$ M) with ten similar active compounds (i.e. darunavir, lopinavir, saquinavir, chlorpromazine, festinavir, oseltamivir, remdesivir, ribavirin, azithromycin, and favipiravir) tested against SARS-CoV-2 M<sup>pro</sup> [60–63] showed a similar range of binding energies and MPO scores. This suggests that several of the 2-pyridone-containing compounds highlighted in this study, in addition to favourable calculated inhibitory properties against SARS-CoV-2 M<sup>pro</sup>, also exhibit comparable pharmacokinetic and drug-like profiles to those of approved therapeutics, particularly leporin A (4). These results provide further credence for the potential therapeutic value of the natural products explored herein against COVID-19, and emphasizes the further need for pre-clinical and clinical evaluation of these 2-pyridone natural products.

In summary, 33 natural products containing the 2-pyridone moiety were studied using *in silico* docking techniques against SARS-CoV-2 M<sup>pro</sup>. M<sup>pro</sup> is a key protein involved in the infection and replication processes of SARS-CoV-2, making it a valuable drug target. All 33 natural product 2-pyridones docked to M<sup>pro</sup>, bound to the active site gorge and exhibited promising binding energies and calculated *K*i values in the nanomolar to micromolar range. These 2-pyridones indicated key

interactions with amino acid residues of the SARS-CoV-2 M<sup>pro</sup> active site, such as the catalytic dyad His-41 and Cys-145. Other key residues such as Met-49, His-164, Met-165, Glu-166, and Gln-189 suggest these amino acids may have importance in the inhibitor-SARS-CoV-2 M<sup>pro</sup> binding properties to the active site. These *in silico* findings, and those described previously [18,19], suggest that 2-pyridone natural products may act as potent inhibitors of SARS-CoV-2 M<sup>pro</sup>. Five of these 2-pyridone compounds ( $K_i$  values  $\leq 1.0 \mu$ M) also exhibited pharmacokinetic parameters favourable for drug design, where leporin A (4) showed the most promise for future therapeutic use. This *in silico* study provides a solid foundation for the discovery and development of other effective therapies to combat SARS-CoV-2. Further investigation using *in vivo* and *in vitro* experimentation is required to confirm the effectiveness of 2-pyridone natural products as possible treatments for COVID-19.

#### 5. Conclusion

Natural products are a rich source of compounds that can be utilized as therapeutics. In the context of COVID-19, natural products that are inhibitors of SARS-CoV-2  $M^{pro}$  may provide another avenue for combating the disease. Thirteen 2-pyridone natural products were identified to have favourable predicted inhibitory properties against the virus, five of which also exhibited favourable pharmacokinetic profiles, especially leporin A (4). The present work indicates that natural products containing a 2-pyridone scaffold have the potential to be used as therapeutics against the novel coronavirus SARS-CoV-2 to treat COVID-19.

#### CRediT authorship contribution statement

Katrina L. Forrestall: Formal analysis, Investigation, Methodology, Validation, Visualization, Writing - original draft, Writing - review & editing. Darcy E. Burley: Formal analysis, Investigation, Validation, Visualization, Writing - original draft, Writing - review & editing. Meghan K. Cash: Writing - original draft, Writing - review & editing. Ian R. Pottie: Conceptualization, Formal analysis, Funding acquisition, Investigation, Methodology, Resources, Supervision, Validation, Visualization, Writing - original draft, Writing - review & editing. Sultan Darvesh: Conceptualization, Formal analysis, Funding acquisition, Project administration, Resources, Supervision, Writing - original draft, Writing - review & editing.

# Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

# Acknowledgments

This work was supported in part by the Canadian Institutes of Health Research (PJT – 153319), the Dalhousie Medical Research Foundation (DMRF Chemists and The Durland Breakthrough Fund), the Canadian Foundation for Innovation (37854), and the Dalhousie Medical Research Foundation Irene MacDonald Sobey Endowed Chair in Curative Approaches to Alzheimer's Disease.

# Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.jclepro.2020.125216.

### References

B.S. Gorbalenya, R.S. Baric, R.J. de Groot, C. Drosten, A.A. Gulyaeva, B. L. Haagmans, C. Lauber, A.M. Leontovich, B.W. Neuman, D. Penzar, S. Perlman, L.

L.M. Poon, D.V. Samborskiy, I.A. Sidorov, I. Sola, J. Ziebuhr, The species Severe acute respiratory syndrome-related coronavirus: classifying 2019-nCoV and naming it SARS-CoV-2, Nat. Microbiol. 5 (2020) 536–544, https://doi.org/ 10.1038/s41564-020-0695-z.

- [2] F. Wu, S. Zhao, B. Yu, Y.M. Chen, W. Wang, Z.G. Song, Y. Hu, Z.W. Tao, J.H. Tian, Y.Y. Pei, M.L. Yuan, Y.L. Zhang, F.H. Dai, Y. Liu, Q.M. Wang, J.J. Zheng, L. Xu, E. C. Holmes, Y.Z. Zhang, A new coronavirus associated with human respiratory disease in China, Nature 579 (2020) 265–269, https://doi.org/10.1038/s41586-020-2008-3.
- [3] P. Zhou, X.L. Yang, X.G. Wang, B. Hu, L. Zhang, W. Zhang, H.R. Si, Y. Zhu, B. Li, C. L. Huang, H.D. Chen, J. Chen, Y. Luo, H. Guo, R.D. Jiang, M.Q. Liu, Y. Chen, X. R. Shen, X. Wang, X.S. Zheng, K. Zhao, Q.J. Chen, F. Deng, L.L. Liu, B. Yan, F. X. Zhan, Y.Y. Wang, G.F. Xiao, Z.L. Shi, A pneumonia outbreak associated with a new coronavirus of probable bat origin, Nature 579 (2020) 270–273, https://doi.org/10.1038/s41586-020-2012-7.
- [4] J. Yang, Y. Zheng, X. Gou, K. Pu, Z. Chen, Q. Guo, R. Ji, H. Wang, Y. Wang, Y. Zhou, Prevalence of comorbidities and its effects in coronavirus disease 2019 patients: a systematic review and meta-analysis, Int. J. Infect. Dis. 94 (2020) 91–95, https:// doi.org/10.1016/j.ijid.2020.03.017.
- [5] J.J. Zhang, X. Dong, Y.Y. Cao, Y.D. Yuan, Y.B. Yang, Y.Q. Yan, C.A. Akdis, Y. D. Gao, Clinical characteristics of 140 patients infected with SARS-CoV-2 in Wuhan, China 75, Allergy, 2020, pp. 1730–1741, https://doi.org/10.1111/all.14238.
- [6] Johns Hopkins University, Coronavirus Resource Centre, Johns Hopkins University, United States, 2020.
- [7] A.M. Baig, A. Khaleeq, U. Ali, H. Syeda, Evidence of the COVID-19 virus targeting the CNS: tissue distribution, host-virus interaction, and proposed neurotropic mechanisms, ACS Chem. Neurosci. 11 (2020) 995–998, https://doi.org/10.1021/ acschemneuro.0c00122.
- [8] W. Li, M.J. Moore, N. Vasilieva, J. Sui, S.K. Wong, M.A. Berne, M. Somasundaran, J.L. Sullivan, K. Luzuriaga, T.C. Greenough, H. Choe, M. Farzan, Angiotensinconverting enzyme 2 is a functional receptor for the SARS coronavirus, Nature 426 (2003) 450–454, https://doi.org/10.1038/nature02145.
- [9] J. Netland, D.K. Meyerholz, S. Moore, M. Cassell, S. Perlman, Severe acute respiratory syndrome coronavirus infection causes neuronal death in the absence of encephalitis in mice transgenic for human ACE2, J. Virol. 82 (2008) 7264–7275, https://doi.org/10.1128/jvi.00737-08.
- [10] F. Li, W. Li, M. Farzan, S.C. Harrison, Structure of SARS coronavirus spike receptorbinding domain complexed with receptor, Science 309 (2005) 1864–1868, https:// doi.org/10.1126/science.1116480.
- [11] M. Letko, A. Marzi, V. Munster, Functional assessment of cell entry and receptor usage for SARS-CoV-2 and other lineage B betacoronaviruses, Nat. Microbiol. 5 (2020) 562–569, https://doi.org/10.1038/s41564-020-0688-y.
- [12] O. Palasca, A. Santos, C. Stolte, J. Gorodkin, L.J. Jensen, TISSUES 2.0: an Integrative Web Resource on Mammalian Tissue Expression, Database, Oxford), 2018, 2018 bay003.
- [13] D. Wrapp, N. Wang, K.S. Corbett, J.A. Goldsmith, C.L. Hsieh, O. Abiona, B. S. Graham, J.S. McLellan, Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation, Science 367 (2020) 1260–1263, https://doi.org/10.1126/science.abb2507.
- [14] J. Ziebuhr, E.J. Snijder, A.E. Gorbalenya, Virus-encoded proteinases and proteolytic processing in the Nidovirales, J. Gen. Virol. 81 (2000) 853–879, https://doi.org/10.1099/0022-1317-81-4-853.
- [15] R. Hilgenfeld, From SARS to MERS: crystallographic studies on coronaviral proteases enable antiviral drug design, FEBS J. 281 (2014) 4085–4096, https://doi. org/10.1111/febs.12936.
- [16] P.V. Baranov, C.M. Henderson, C.B. Anderson, R.F. Gesteland, J.F. Atkins, M. T. Howard, Programmed ribosomal frameshifting in decoding the SARS-CoV genome, Virology 332 (2005) 498–510, https://doi.org/10.1016/j. virol.2004.11.038.
- [17] K. Anand, J. Ziebuhr, P. Wadhwani, J.R. Mesters, R. Hilgenfeld, Coronavirus main proteinase (3CLpro) structure: basis for design of anti-SARS drugs, Science 300 (2003) 1763–1767, https://doi.org/10.1126/science.1085658.
- [18] L. Zhang, D. Lin, X. Sun, U. Curth, C. Drosten, L. Sauerhering, S. Becker, K. Rox, R. Hilgenfeld, Crystal structure of SARS-CoV-2 main protease provides a basis for design of improved alpha-ketoamide inhibitors, Science 368 (2020) 409–412, https://doi.org/10.1126/science.abb3405.
- [19] L. Zhang, D. Lin, Y. Kusov, Y. Nian, Q. Ma, J. Wang, A. von Brunn, P. Leyssen, K. Lanko, J. Neyts, A. de Wilde, E.J. Snijder, H. Liu, R. Hilgenfeld, Alphaketoamides as broad-spectrum inhibitors of coronavirus and enterovirus replication: structure-based design, synthesis, and activity assessment, J. Med. Chem. 63 (2020) 4562–4578, https://doi.org/10.1021/acs.jmedchem.9b01828.
  [20] J. Shi, J. Song, The catalysis of the SARS 3C-like protease is under extensive
- [20] J. Shi, J. Song, The catalysis of the SARS 3C-like protease is under extensive regulation by its extra domain, FEBS J. 273 (2006) 1035–1045, https://doi.org/ 10.1111/j.1742-4658.2006.05130.x.
- [21] K. Anand, G.J. Palm, J.R. Mesters, S.G. Siddell, J. Ziebuhr, R. Hilgenfeld, Structure of coronavirus main proteinase reveals combination of a chymotrypsin fold with an extra α-helical domain, EMBO J. 21 (2002) 3213–3224, https://doi.org/10.1093/ emboj/cdf327.
- [22] M. Bzowka, K. Mitusinska, A. Raczynska, A. Samol, J.A. Tuszynski, A. Gora, Structural and evolutionary analysis indicate that the SARS-CoV-2 Mpro is a challenging target for small-molecule inhibitor design, Int. J. Mol. Sci. 21 (2020) 3099, https://doi.org/10.3390/ijms21093099.
- [23] D.W. Kneller, G. Phillips, H.M. O'Neill, R. Jedrzejczak, L. Stols, P. Langan, A. Joachimiak, L. Coates, A. Kovalevsky, Structural plasticity of SARS-CoV-2 3CL

Mpro active site cavity revealed by room temperature X-ray crystallography, Nat. Commun. 11 (2020), https://doi.org/10.1038/s41467-020-16954-7, 3202 (2020).

- [24] D.A. Dias, S. Urban, U. Roessner, A historical overview of natural products in drug discovery, Metabolites 2 (2012) 303–336, https://doi.org/10.3390/ metabo2020303.
- [25] G.K. Sims, L.E. Lee, Degradation of pyridine derivatives in soil, J. Environ. Qual. 14 (1985) 580–584, https://doi.org/10.2134/jeq1985.00472425001400040022x.
- [26] G.Y. Chen, W. Zhong, Z. Zhou, Q. Zhang, Simultaneous determination of tryptophan and its 31 catabolites in mouse tissues by polarity switching UHPLC-SRM-MS, Anal. Chim. Acta 1037 (2018) 200–210, https://doi.org/10.1016/j. aca.2018.02.026.
- [27] R.S. Grant, S.E. Coggan, G.A. Smythe, The physiological action of picolinic Acid in the human brain, Int. J. Tryptophan Res. 2 (2009) 71–79, https://doi.org/ 10.4137/IJTR.S2469.
- [28] A. Lenglet, S. Liabeuf, S. Bodeau, L. Louvet, A. Mary, A. Boullier, A.S. Lemaire-Hurtel, A. Jonet, P. Sonnet, S. Kamel, Z.A. Massy, N-methyl-2-pyridone-5carboxamide (2PY)-Major metabolite of nicotinamide: an update on an old uremic toxin, Toxins (Basel) 8 (2016) 339, https://doi.org/10.3390/toxins8110339.
- [29] S.A. Trammell, M.S. Schmidt, B.J. Weidemann, P. Redpath, F. Jaksch, R. W. Dellinger, Z. Li, E.D. Abel, M.E. Migaud, C. Brenner, Nicotinamide riboside is uniquely and orally bioavailable in mice and humans, Nat. Commun. 7 (2016) 12948, https://doi.org/10.1038/ncomms12948.
- [30] W.S. Hamama, M. Waly, I. El-Hawary, H.H. Zoorob, Developments in the chemistry of 2-pyridone, Synth. Commun. 44 (2014) 1730–1759, https://doi.org/10.1080/ 00397911.2013.862836.
- [31] J.G. Sośnicki, T.J. Idzik, Pyridones powerful precursors for the synthesis of alkaloids, their derivatives, and alkaloid-inspired compounds Synthesis 51 (2019) 3369–3396, https://doi.org/10.1055/s-0037-1611844.
- [32] J. Bao, H. Zhai, K. Zhu, J.H. Yu, Y. Zhang, Y. Wang, C.S. Jiang, X. Zhang, H. Zhang, Bioactive Pyridone Alkaloids from a Deep-Sea-Derived Fungus Arthrinium Sp. UJNMF0008, vol. 16, Mar Drugs, 2018, https://doi.org/10.3390/md16050174.
- [33] L.N. Li, L. Wang, Y.N. Cheng, Z.Q. Cao, X.K. Zhang, X.L. Guo, Discovery and characterization of 4-Hydroxy-2-pyridone derivative sambutoxin as a potent and promising anticancer drug candidate: activity and molecular mechanism, Mol. Pharm. 15 (2018) 4898–4911, https://doi.org/10.1021/acs. moloharmaceut.8b00525.
- [34] Y.-N. Tan, J. Zeng, S.-N. Zhang, R.-J. Ma, Z.-H. Pan, Q.-G. Tan, Pyridone alkaloids from the leaves of Ricinus communis and their inhibitory effect against protein tyrosine phosphatase 1B, Chem. Nat. Compd. 55 (2019) 395–397, https://doi.org/ 10.1007/s10600-019-02702-x.
- [35] H.J. Jessen, K. Gademann, 4-Hydroxy-2-pyridone alkaloids: structures and synthetic approaches, Nat. Prod. Rep. 27 (2010) 1168–1185, https://doi.org/ 10.1039/B911516C.
- [36] M.J. Frisch, G.W. Trucks, H.B. Schlegel, G.E. Scuseria, M.A. Robb, J.R. Cheeseman, G. Scalmani, V. Barone, G.A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. Marenich, J. Bloino, B.G. Janesko, R. Gomperts, B. Mennucci, H.P. Hratchian, J. V. Ortiz, A.F. Izmaylov, J.L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparin, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J.A. Montgomery, J.E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K.N. Kudin, V.N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J.C. Burant, S.S. Iyengar, J. Tomasi, M. Cossi, J.M. Millam, M. Klene, C. Adamo, R. Cammi, J.W. Ochterski, R.L. Martin, K. Morokuma, O. Farkas, J.B. Foresman, D.J. Fox, Gaussian 09, Gaussian, Inc., Wallingford, CT, 2016.
  [37] M.F. Sanper, Puthor: a programming language for software integration and
- [37] M.F. Sanner, Python: a programming language for software integration and development, J. Mol. Graph. Model. 17 (1999) 57–61.
- [38] G.M. Morris, D.S. Goodsell, R.S. Halliday, R. Huey, W.E. Hart, R.K. Belew, A. J. Olson, Automated docking using a Lamarckian genetic algorithm and an empirical binding free energy function, J. Comput. Chem. 19 (1998) 1639–1662, https://doi.org/10.1002/(SICI)1096-987X(19981115)19:14<1639::AID-JJCC10>3.0.CO:2-B.
- [39] G.M. Morris, R. Huey, W. Lindstrom, M.F. Sanner, R.K. Belew, D.S. Goodsell, A. J. Olson, AutoDock4 and AutoDockTools4: automated docking with selective receptor flexibility, J. Comput. Chem. 30 (2009) 2785–2791, https://doi.org/10.1002/jcc.21256.
- [40] M. Vieth, J.D. Hirst, A. Kolinski, C.L. Brooks III, Assessing energy functions for flexible docking, J. Comput. Chem. 19 (1998) 1612–1622, https://doi.org/ 10.1002/(SICI)1096-987X(19981115)19:14<1612::AID-JCC7>3.0.CO;2-M.
- [41] B.D. Bursulaya, M. Totrov, R. Abagyan, C.L. Brooks, Comparative study of several algorithms for flexible ligand docking, J. Comput. Aided Mol. Des. 17 (2003) 755–763, https://doi.org/10.1023/b:jcam.0000017496.76572.6f.
- [42] F. Ballante, G.R. Marshall, An automated strategy for binding-pose selection and docking assessment in structure-based drug design, J. Chem. Inf. Model. 56 (2016) 54–72, https://doi.org/10.1021/acs.jcim.5b00603.
- [43] The PyMOL Molecular Graphics System, Schrödinger, LLC.
- [44] G.A. Jeffrey, An Introduction to Hydrogen Bonding, Oxford University Press, New York and Oxford, 1997.
- [45] M.M. Pinney, A. Natarajan, F. Yabukarski, D.M. Sanchez, F. Liu, R. Liang, T. Doukov, J.P. Schwans, T.J. Martinez, D. Herschlag, Structural coupling throughout the active site hydrogen bond networks of ketosteroid isomerase and photoactive yellow protein, J. Am. Chem. Soc. 140 (2018) 9827–9843, https://doi. org/10.1021/jacs.8b01596.

- [46] N. Phosrithong, J. Ungwitayatorn, Molecular docking study on anticancer activity of plant-derived natural products, Med. Chem. Res. 19 (2010) 817–835, https:// doi.org/10.1007/s00044-009-9233-5.
- [47] K. Thongnum, S. Chanthai, Inhibitory reactivity of capsaicin with α-amylase and α-glucosidase related to antidiabetes using molecular docking and quantum calculation methods, Orient. J. Chem. 34 (2018) 2211–2228, https://doi.org/ 10.13005/oic/340501.
- [48] T.T Wager, X. Hou, P.R. Verhoest, A. Villalobos, Moving beyond rules: the development of a central nervous system multiparameter optimization (CNS MPO) approach to enable alignment of druglike properties, ACS Chem. Neurosci. 1 (2010) 435–449, https://doi.org/10.1021/cn100008c.
- [49] T.J. Hou, K. Xia, W. Zhang, X.J. Xu, ADME Evaluation in Drug Discovery. 4. Prediction of Aqueous Solubility Based on Atom Contribution Approach, J. Chem. Inf. Comput. Sci. 44 (2004) 266–275, https://doi.org/10.1021/ci034184n.
- [50] C.A. Lipinski, F. Lombardo, B.W. Dominy, P.J. Feeney, Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings, Adv. Drug Deliv. Rev. 46 (2001) 3–26, https://doi.org/10.1016/S0169-409X(96)00423-1.
- [51] J. Dong, N.-N. Wang, Z.-J. Yao, L. Zhang, Y. Cheng, D. Ouyang, A.-P. Lu, D.-S. Cao, ADMETlab: a platform for systematic ADMET evaluation based on a comprehensively collected ADMET database, J. Cheminform. 10 (2018), https:// doi.org/10.1186/s13321-018-0283-x.
- [52] L.L.G. Ferreira, A.D. Andricopulo, ADMET modeling approaches in drug discovery 24, Drug Discov. Today, 2019, pp. 1157–1165, https://doi.org/10.1016/j. drudis.2019.03.015.
- [53] S.S.S.J. Ahmed, V. Ramakrishnan, Systems biological approach of molecular descriptors connectivity: optimal descriptors for oral bioavailability prediction 7, PLOS One, 2012, p. e40654, https://doi.org/10.1371/journal.pone.0040654.
- [54] R. Watanabe, T. Esaki, H. Kawashima, Y. Natsume-Kitatani, C. Nagao, R. Ohashi, K. Mizuguchi, Predicting fraction unbound in human plasma from chemical structure: improved accuracy in the low value ranges, Mol. Pharm. 15 (2018) 5302–5311, https://doi.org/10.1021/acs.molpharmaceut.8b00785, 1203.
- [55] H. Zhu, T.M. Martin, L. Ye, A. Sedykh, D.M. Young, A. Tropsha, Quantitative structure-activity relationship modeling of rat acute toxicity by oral exposure, Chem. Res. Toxicol. 22 (2009) 1913–1921, https://doi.org/10.1021/tx900189p.
- [56] T. Lei, Y. Li, Y. Song, D. Li, H. Sun, T. Hou, ADMET evaluation in drug discovery: 15. Accurate prediction of rat oral acute toxicity using relevance vector machine and consensus modeling, J. Cheminform. 8 (2016) 1–19, https://doi.org/10.1186/ s13321-016-0117-7, 6.
- [57] N.-N. Wang, J. Dong, Y.-H. Deng, M.-F. Zhu, M. Wen, Z.-J. Yao, A.-P. Lu, J.-B. Wang, D.-S. Cao, ADME properties evaluation in drug discovery: prediction of Caco-2 cell permeability using a combination of NSGA-II and boosting, J. Chem. Inform. Model. 56 (2016) 763–773, https://doi.org/10.1021/acs.jcim.5b00642.
- [58] H. Yang, C. Lou, L. Sun, J. Li, Y. Cai, Z. Wang, W. Li, G. Liu, Y. Tang, admetSAR 2.0: web-service for prediction and optimization of chemical ADMET properties, Bioinformatics 35 (2018) 1067–1069, https://doi.org/10.1093/bioinformatics/ bty707.
- [59] H. Yang, L. Sun, Z. Wang, W. Li, G. Liu, Y. Tang, ADMETopt: a web server for ADMET optimization in drug design via scaffold hopping, J. Chem. Inform. Model. 58 (2018) 2051–2056, https://doi.org/10.1021/acs.jcim.8b00532.
- [60] R.R. Deshpande, A.P. Tiwari, N. Nyayanit, M. Modak, In silico molecular docking analysis for repurposing therapeutics against multiple proteins from SARS-CoV-2, Eur. J. Pharmacol. 886 (2020) 173430, https://doi.org/10.1016/j. einhar.2020.173430.
- [61] S.A. Khan, K. Zia, S. Ashraf, R. Uddin, Z. Ul-Haq, Identification of chymotrypsinlike protease inhibitors of SARS-CoV-2 via integrated computational approach, J. Biomol. Struct. Dyn. (2020) 1–10, https://doi.org/10.1080/ 07391102 2020 1751298
- [62] C. Shivanika, S. Deepak Kumar, R. Venkataraghavan, T. Pawan, A. Sumitha, P. Brindha Devi, Molecular docking, validation, dynamics simulations, and pharmacokinetic prediction of natural compounds against the SARS-CoV-2 mainprotease, J. Biomol. Struct. Dyn. 78 (2012) 1020–1023, https://doi.org/10.1080/ 07391102.2020.1815584.
- [63] A.-M. Udrea, S. Avram, S. Nistorescu, M.-L. Pascu, M.O. Romanitan, Laser irradiated phenothiazines: New potential treatment for COVID-19 explored by molecular docking, J. Photochem. Photobiol. B 211 (2020) 111997, https://doi. org/10.1016/j.jphotobiol.2020.111997.
- [64] E. de Wit, N. van Doremalen, D. Falzarano, V.J. Munster, SARS and MERS: recent insights into emerging coronaviruses, Nat. Rev. Microbiol. 14 (2016) 523–534, https://doi.org/10.1038/nrmicro.2016.81.
- [65] J.M. Sanders, M.L. Monogue, T.Z. Jodlowski, J.B. Cutrell, Pharmacologic Treatments for Coronavirus Disease 2019 (COVID-19): A Review, JAMA 323 (2020) 1824–1836, https://doi.org/10.1001/jama.2020.6019.
- [66] S. Weston, C.M. Coleman, R. Haupt, J. Logue, K. Matthews, M.B. Frieman, Broad Anti-coronaviral Activity of FDA Approved Drugs against SARS-CoV-2 in Vitro and SARS-CoV in Vivo, bioRixv (2020), https://doi.org/10.1101/2020.03.25.008482.
- [67] M. Tsuchinari, K. Shimanuki, F. Hiramatsu, T. Murayama, T. Koseki, Y. Shioni, Fusapyridons A and B, Novel Pyridone Alkaloids from an Endophytic Fungus, Fusarium sp. YG-45, Z Naturforsch B J Chem Sci 62 (2007) 1203–1207, https:// doi.org/10.1515/znb-2007-0916.
- [68] F.A. Rotsaert, M.G. Ding, B.L. Trumpower, Differential efficacy of inhibition of mitochondrial and bacterial cytochrome bc1 complexes by center N inhibitors antimycin, ilicicolin H and funiculosin, Biochim. Biophys. Acta 1777 (2008) 211–219, https://doi.org/10.1016/j.bbabio.2007.10.011.

- Chemico-Biological Interactions 335 (2021) 109348
- [69] Q. Tao, C. Ding, B.N. Auckloo, B. Wu, Bioactive metabolites from a hydrothermal vent fungus Aspergillus sp. YQ-13, Nat. Prod. Commun. 13 (2018) 571–573, https://doi.org/10.1177/1934578X1801300514.
- [70] C. Zhang, L. Jin, B. Mondie, S.S. Mitchell, A.L. Castelhano, W. Cai, N. Bergenhem, Leporin B., A novel hexokinase II gene inducing agent from an unidentified fungus, Bioorg. Med. Chem. Lett. 13 (2003) 1433–1435, https://doi.org/10.1016/S0960-894X(03)00153-7.
- [71] H.V. Kemami Wangun, C. Hertweck, Epicoccarines A, B and epipyridone: tetramic acids and pyridone alkaloids from an Epicoccum sp. associated with the tree fungus Pholiota squarrosa, Org. Biomol. Chem. 5 (2007) 1702–1705, https://doi.org/ 10.1039/B702378B.
- [72] Y. Zhang, Q. Zhang, J. Bao, J. Huang, H. Zhang, Apiosporamide, A 4-hydroxy-2pyridone alkaloid, induces apoptosis via PI3K/akt signaling pathway in osteosarcoma cells, OncoTargets Ther. 12 (2019) 8611–8620, https://doi.org/ 10.2147/OTT.S218692.
- [73] M. Isaka, R. Haritakun, K. Intereya, D. Thanakitpipattana, N.L. Hywel-Jones, Torrubiellone E., An antimalarial N-hydroxypyridone alkaloid from the spider pathogenic fungus Torrubiella longissima BCC 2022 9, Nat. Prod. Commun, 2014, pp. 627–628, https://doi.org/10.1177/1934578X1400900508.
- [74] Y. Cheng, B. Schneider, U. Riese, B. Schubert, Z. Li, M. Hamburger, Farinosones A.-C., Neurotrophic alkaloidal metabolites from the entomogenous deuteromycete Paecilomyces farinosus, J. Nat. Prod. 67 (2004) 1854–1858, https://doi.org/ 10.1021/np049761w.
- [75] F.M. Talontsi, T.J. Nwemeguela Kenla, B. Dittrich, C. Douanla-Meli, H. Laatsch, Paeciloside A., A new antimicrobial and cytotoxic polyketide from Paecilomyces sp. strain CAFT156, Planta Med. 78 (2012) 379–382, https://doi.org/10.1055/s-0031-1298622.
- [76] M. Shibazaki, M. Taniguchi, T. Yokoi, K. Nagai, M. Watanabe, K. Suzuki, T. Yamamoto, YM-215343, a novel antifungal compound from Phoma sp. QN04621, J. Antibiot. 57 (2004) 379–382, https://doi.org/10.7164/ antibiotics.57.379.
- [77] M. Isaka, M. Tanticharoen, P. Kongsaeree, Y. Thebtaranonth, Structures of cordypyridones A-D, antimalarial N-hydroxy- and N-methoxy-2-pyridones from the insect pathogenic fungus Cordyceps nipponica, J. Org. Chem. 66 (2001) 4803–4808, https://doi.org/10.1021/jo0100906.
- [78] S. Takahashi, N. Kakinuma, K. Uchida, R. Hashimoto, T. Yanagisawa, A. Nakagawa, Pyridovericin and pyridomacrolidin: novel metabolites from entomopathogenic fungi, Beauveria bassiana, J. Antibiot. 51 (1998) 596–598, https://doi.org/10.7164/antibiotics.51.596.
- [79] M. de Souza Santos, W. Jonis Andrioli, M.P. Freire de Morais Del Lama, J. Kenupp Bastos, N.P. Nanayakkara, R.M. Zumstein Georgetto Naal, In vitro anti-allergic activity of the fungal metabolite pyridovericin, Int. Immunopharm. 15 (2013) 532–538, https://doi.org/10.1016/j.intimp.2013.01.017.
- [80] M.M. Wagenaar, D.M. Gibson, J. Clardy, Akanthomycin, a new antibiotic pyridone from the entomopathogenic fungus Akanthomyces gracilis, Org. Lett. 4 (2002) 671–673, https://doi.org/10.1021/ol016737q.
- [81] R.E. McHugh, N. O'Boyle, J.P.R. Connolly, P.A. Hoskisson, A.J. Roe, Characterization of the mode of action of aurodox, a type III secretion system inhibitor from streptomyces goldiniensis, Infect. Immun. 87 (2) (2019) e00595–18, https://doi.org/10.1128/IAI.00595-18.
- [82] E.D. de Silva, A.S. Geiermann, M.I. Mitova, P. Kuegler, J.W. Blunt, A.L. Cole, M. H. Munro, Isolation of 2-pyridone alkaloids from a New Zealand marine-derived penicillium species, J. Nat. Prod. 72 (2009) 477–479, https://doi.org/10.1021/ np800627f.
- [83] Y. Fujita, H. Oguri, H. Oikawa, Biosynthetic studies on the antibiotics PF1140: a novel pathway for a 2-pyridone framework, Tetrahedron Lett. 46 (2005) 5885–5888, https://doi.org/10.1016/j.tetlet.2005.06.115.
- [84] S. Dorsaz, T. Snaka, Q. Favre-Godal, P. Maudens, N. Boulens, P. Furrer, S. N. Ebrahimi, M. Hamburger, E. Allemann, K. Gindro, E.F. Queiroz, H. Riezman, J. L. Wolfender, D. Sanglard, Identification and mode of action of a plant natural product targeting human fungal pathogens, Antimicrob. Agents Chemother. 61 (2017) e00829–17, https://doi.org/10.1128/AAC.00829-17.
- [85] L.P. Ióca, S. Romminger, M.F.C. Santos, K.F. Bandeira, F.T. Rodrigues, M. H. Kossuga, K.J. Nicacio, E.L.F. Ferreira, R.P. Morais-Urano, M.S. Passos, L. K. Kohn, C.W. Arns, L.D. Sette, R.G.S. Berlinck, A strategy for the rapid identification of fungal metabolites and the discovery of the antiviral activity of pyrenocine a and harzianopyridone, Quím. Nova 39 (2016) 720–731, https://doi. org/10.5935/0100-4042.20160092.
- [86] U. Bat-Erdene, D. Kanayama, D. Tan, W.C. Turner, K.N. Houk, M. Ohashi, Y. Tang, Iterative catalysis in the biosynthesis of mitochondrial complex II inhibitors harzianopyridone and atpenin B, J. Am. Chem. Soc. 142 (2020) 8550–8554, https://doi.org/10.1021/jacs.0c03438.
- [87] Y. Kusakabe, S. Mizutani, S. Kamo, T. Yoshimoto, S. Tomoshige, T. Kawasaki, R. Takasawa, K. Tsubaki, K. Kuramochi, Synthesis, antibacterial and cytotoxic evaluation of flavipucine and its derivatives, Bioorg. Med. Chem. Lett. 29 (2019) 1390–1394, https://doi.org/10.1016/j.bmcl.2019.03.034.
- [88] H. Miyadera, K. Shiomi, H. Ui, Y. Yamaguchi, R. Masuma, H. Tomoda, H. Miyoshi, A. Osanai, K. Kita, S. Omura, Atpenins, potent and specific inhibitors of mitochondrial complex II (succinateubiquinone oxidoreductase), Proc. Natl. Acad. Sci. U. S. A. 100 (2003) 473–477, https://doi.org/10.1073/pnas.0237315100.
- [89] M. Isaka, P. Chinthanom, S. Supothina, P. Tobwor, N.L. Hywel-Jones, Pyridone and tetramic acid alkaloids from the spider pathogenic fungus Torrubiella sp. BCC 2165, J. Nat. Prod. 73 (2010) 2057–2060, https://doi.org/10.1021/np100492j.

# K.L. Forrestall et al.

- [90] H. Grubmuller, L.V. Bock, tRNA Dissociation from EF-Tu after GTP Hydrolysis: Primary Steps and Antibiotic Inhibition, Biophys. J. 118 (2020) 151–161, https:// doi.org/10.1016/j.bpj.2019.10.028.
- [91] K. Kimura, M. Iwatsuki, T. Nagai, A. Matsumoto, Y. Takahashi, K. Shiomi, S. Omura, A. Abe, A small-molecule inhibitor of the bacterial type III secretion system protects against in vivo infection with Citrobacter rodentium, J. Antibiot. 64 (2011) 197–203, https://doi.org/10.1038/ja.2010.155.
- [92] A.C. Ferraz, J.A. Anselmo-Franci, S.R. Perosa, E.F. de Castro-Neto, M.I. Bellissimo, B.H. de Oliveira, E.A. Cavalheiro, G. Naffah-Mazzacoratti Mda, C. Da Cunha, Amino acid and monoamine alterations in the cerebral cortex and hippocampus of mice submitted to ricinine-induced seizures, Pharmacol. Biochem. Behav. 72 (2002) 779–786, https://doi.org/10.1016/S0091-3057(02)00750-5.