

BRAIN COMMUNICATIONS

Immune checkpoint inhibitor-associated myopathy: a clinicoseropathologically distinct myopathy

Shahar Shelly,^{1,*} James D. Triplett,^{1,*} Marcus V. Pinto,¹ Margherita Milone,¹ Felix E. Diehn,² Anastasia Zekeridou^{1,3} and Teerin Liewluck¹

*These authors contributed equally to this work.

Immune checkpoint inhibitors have revolutionized the landscape of cancer treatment. Alongside their many advantages, they elicit immune-related adverse events, including myopathy, which potentially result in substantial morbidity if not recognized and treated promptly. Current knowledge of immune checkpoint inhibitor-associated myopathy is limited. We conducted a 5-year retrospective study of patients with immune checkpoint inhibitor-associated myopathy. Clinical features, survival and ancillary test findings were analysed and compared with those of immune-mediated necrotizing myopathy patients without immune checkpoint inhibitor exposure seen during the same time period. We identified 24 patients with immune checkpoint inhibitor-associated myopathy (median age 69 years; range 28–86) and 38 patients with immune-mediated necrotizing myopathy. Ocular involvement occurred in 9/24 patients with immune checkpoint inhibitor exposure, without electrodiagnostic evidence of neuromuscular transmission defect, and in none of the immune-mediated necrotizing myopathy patients ($P < 0.001$). Myocarditis occurred in eight immune checkpoint inhibitor-associated myopathy patients and in none of the immune-mediated necrotizing myopathy patients ($P < 0.001$). Median creatine kinase was 686 IU/l in the immune checkpoint inhibitor cohort (seven with normal creatine kinase) compared to 6456 IU/l in immune-mediated necrotizing myopathy cohort ($P < 0.001$). Lymphopenia was observed in 18 and 7 patients with and without immune checkpoint inhibitor exposure, respectively ($P < 0.001$). Myopathological findings were similar between patients with and without immune checkpoint inhibitor exposure, consisting of necrotic fibres with no or subtle inflammation. Necrotic fibres however arranged in clusters in 10/11 immune checkpoint inhibitor-associated myopathy patients but in none of the immune checkpoint inhibitor-naïve patients ($P < 0.001$). Despite the lower creatine kinase levels in immune checkpoint inhibitor-exposed patients, the number of necrotic fibres was similar in both groups. Immune checkpoint inhibitor-associated myopathy patients had a higher frequency of mitochondrial abnormalities and less number of regenerating fibres than immune-mediated necrotizing myopathy patients ($P < 0.001$). Anti-hydroxy-3-methylglutaryl-CoA reductase or signal recognition particle antibodies were absent in patients with immune checkpoint inhibitor exposure but positive in two-thirds of immune checkpoint inhibitor-naïve patients. Most patients with immune checkpoint inhibitor-associated myopathy responded favourably to immunomodulatory treatments, but four died from myopathy-related complications and one from myocarditis. Intubated patients had significantly shorter survival compared to non-intubated patients (median survival of 22 days; $P = 0.004$). In summary, immune checkpoint inhibitor-associated myopathy is a distinct, treatable immune-mediated myopathy with common ocular involvement, frequent lymphopenia and necrotizing histopathology, which contrary to immune-mediated necrotizing myopathy, is featured by clusters of necrotic fibres and not accompanied by anti-hydroxy-3-methylglutaryl-CoA reductase or signal recognition particle antibodies. Normal or mildly elevated creatine kinase level does not exclude the diagnosis.

1 Department of Neurology, Mayo Clinic, Rochester, MN, USA

2 Department of Radiology, Mayo Clinic, Rochester, MN, USA

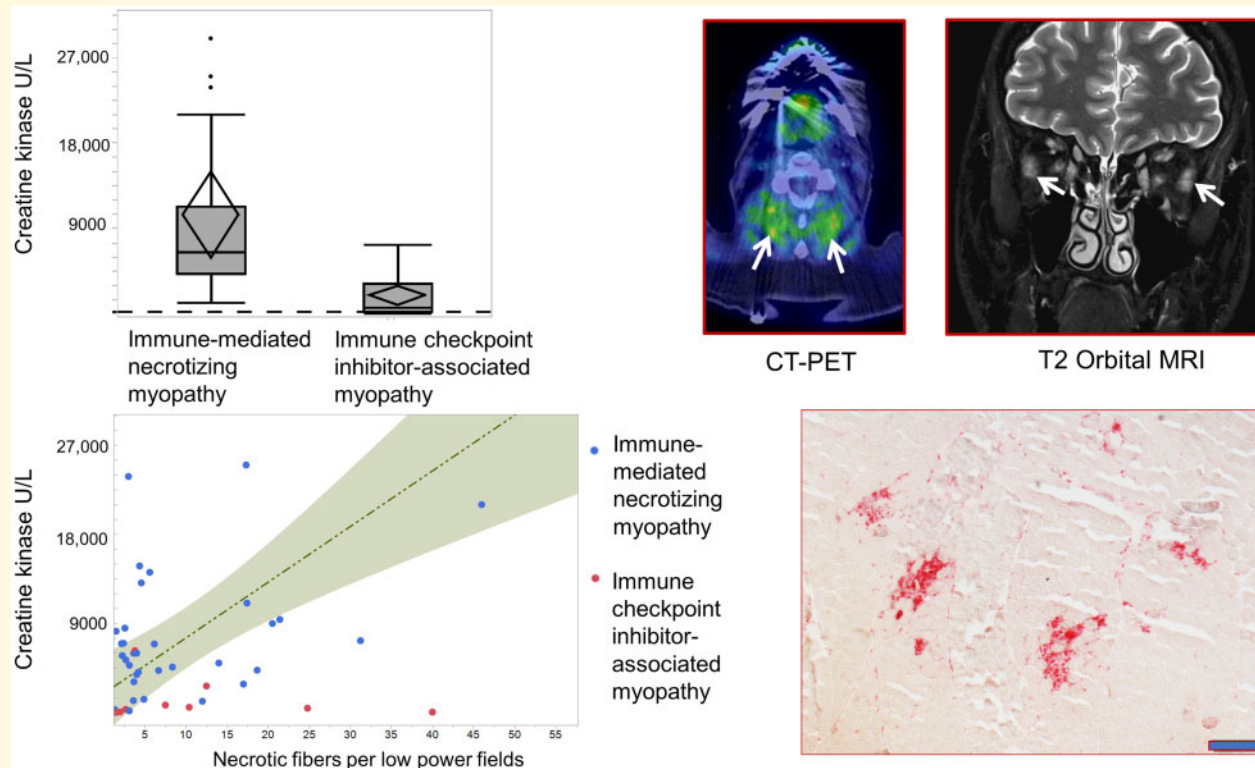
3 Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, MN, USA

Correspondence to: Teerin Liewluck, MD, Department of Neurology, Mayo Clinic, 200 1st St SW, Rochester, MN 55905, USA
E-mail: liewluck.teerin@mayo.edu

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Abbreviations: CK = creatine kinase; CTCAE = Common Terminology Criteria for Adverse Events version; HMGCR = 3-Hydroxy-3-Methylglutaryl-CoA Reductase; ICIAM = immune checkpoint inhibitor-associated myopathy; IMNM = immune-mediated necrotizing myopathy; irAEs = immune-related adverse events; IVMP = intravenous methylprednisolone; PD-1 = programmed cell death-1; RNS = repetitive nerve stimulation; SFEMG = single-fibre EMG; SRP = signal recognition particle

Graphical Abstract



Introduction

The discovery that cancer cells are capable of hijacking immune checkpoints and evade immune attack has revolutionized cancer immunology and led to novel treatments. Immune checkpoint blockade is a novel cancer therapeutic strategy, in which inhibition of immune suppressor molecules promotes immune attack against cancer cells (Ishida *et al.*, 1992; Leach *et al.*, 1996; Sharma and Allison, 2015). These immune checkpoint inhibitors (ICI) include monoclonal IgG antibodies targeting the programmed cell death-1, nivolumab and pembrolizumab, the programmed cell death ligand-1, atezolizumab,

avelumab and durvalumab, and the cytotoxic T-lymphocyte-associated antigen 4, ipilimumab (Bilen *et al.*, 2016).

By unleashing the immune system against cancer, immune checkpoint blockade inevitably causes immune-mediated side effects, or so-called immune-related adverse events (irAEs) (Brahmer *et al.*, 2018). The precise pathophysiology underlying irAEs is not well-understood. Different mechanisms have been postulated, all of which lead to tissue infiltration by highly activated CD4+ or CD8+ T cells (Bilen *et al.*, 2016; Mammen *et al.*, 2019). Although neurological irAEs are not as common as those affecting other organ systems, they can be severe and potentially fatal (Kao *et al.*, 2017). In some cohorts, neuromuscular disorders

account for more than 50% of neurological irAEs, including peripheral nerve diseases, neuromuscular junction disorders and myopathies (Suzuki *et al.*, 2017; Kao *et al.*, 2018; Liewluck *et al.*, 2018; Touat *et al.*, 2018; Dubey *et al.*, 2019). Each neuromuscular complication can occur individually or in combination, such as the frequently reported myasthenia gravis-myopathy overlap syndrome. The hyperCKemia, observed in the majority of the reported ICI-associated myasthenia gravis patients, has been attributed to coexisting myopathy or myocarditis (Kao *et al.*, 2018). However, because orbital myositis can occur in ICI-treated patients (Bitton *et al.*, 2019), it can be difficult to ascertain with confidence the coexistence of myasthenia gravis in the absence of electrodiagnostic evidence of defect of neuromuscular transmission. Indeed, even antiacetylcholine receptor binding antibodies can occasionally be detected in the absence of myasthenia gravis (Maddison *et al.*, 2019).

A wide range of immune-mediated myopathies were reported in ICI-treated patients, including polymyositis, dermatomyositis, granulomatous myositis, necrotizing myopathy and non-specific myopathy (Kao *et al.*, 2018). Ocular involvement and multifocal clusters of necrotic fibres are considered unique features, although not universally present, of immune checkpoint inhibitor-associated myopathies (ICIAM) (Liewluck *et al.*, 2018; Touat *et al.*, 2018). It remains debatable whether ophthalmoparesis in ICIAM patients represents orbital myositis or coexisting myasthenia gravis. The current knowledge of ICIAM is limited and mostly derived from individual case reports or small case series. Herein, we present the largest single-centre cohort of ICIAM patients to date. All biopsy-proven patients displayed necrotizing myopathy-like pathology. We then conducted systematic comparison of their clinical, laboratory, electrophysiological and histopathological findings and survival to a cohort of ICI-naïve immune-mediated necrotizing myopathy (IMNM) patients seen over the same time period.

Materials and methods

Study population

Institutional Review Board approval was granted by Mayo Clinic. Using electronic data search, we identified patients seen at our institution between January 2015 and December 2019 with a new diagnosis of myopathy while on ICI. We included patients who developed subacute or acute onset weakness of Common Terminology Criteria for Adverse Events version 5.0 (CTCAE) Grade III and above (Puzanov *et al.*, 2017). The myopathy was considered Grade III only if caused limitation of self-care activities of daily living, Grade IV if they were life-threatening and needed urgent intervention and Grade V if directly caused death. In patients presenting with ocular symptoms CTCAE Grade III was considered if ophthalmoparesis, causing double vision in lateral gaze but not

in primary gaze, and Grade IV if ophthalmoparesis causing double vision in central gaze or patients required head turning to see beyond central 60°.

A diagnosis of myopathy was further classified as definite, probable and possible. Definite myopathy was designated to patients who had histopathological evidence of myopathy or who fulfilled all three of the following criteria: (i) elevated serum creatine kinase (CK), (ii) electrodiagnostic findings of myopathic motor unit potentials (early recruiting, short-duration, low-amplitude motor unit potentials) and (iii) imaging abnormalities observed in the clinically affected muscle. Probable myopathy was diagnosed if patients fulfilled two of the aforementioned criteria. Patients were categorized as possible myopathy when only one of the three criteria was met. Patients with myopathies from other causes unrelated to ICI were excluded. All ICI-naïve IMNM patients seen during the same time period were identified and served as disease control. Diagnosis of IMNM was based on clinical, serological and pathological findings in keeping with international guidelines (Allenbach *et al.*, 2018b).

Patients' characteristics

Review of clinical features and laboratory data, including electrodiagnostic findings, were undertaken in correlation with cancer immunotherapy. The included patients were classified into two phenotypes: oculobulbar and generalized myopathy. Oculobulbar myopathy was defined by a combination of the symptoms of ptosis, diplopia, dysarthria or dysphagia of at least CTCAE grade III with no or subtle limb symptoms (CTCAE grade I-II) and the evidence of oculobulbar weakness (by neurologic examination or swallowing studies) with no or mild (MRC grade ≥ 4) limb weakness. Generalized myopathy was defined by a combination of the symptoms of limb weakness of at least CTCAE grade III and moderate-to-severe limb weakness (MRC grade < 4) on neurologic examination. Patients with generalized myopathy may or may not have symptoms or signs of ocular or bulbar muscle involvement. Statin exposure was documented if patients were on any statin therapy at the time of symptom onset.

The diagnosis of myasthenia gravis was reserved for patients who had decrement on repetitive nerve stimulation (RNS) or increased jitter on single-fibre EMG (SFEMG), with or without myasthenia gravis antibodies. Diagnosis of myocarditis was made when patients developed new unexplained dyspnoea, chest pain, syncope, arrhythmia or congestive heart failure, accompanied by histopathological evidence of myocardial inflammation or echocardiographic or cardiac PET imaging abnormalities, consistent with myocarditis. Respiratory muscle involvement was considered if patients developed new-onset dyspnoea accompanied by abnormal pulmonary function tests or diaphragmatic ultrasound, consistent with restrictive lung disease or diaphragmatic weakness (Naddaf and Milone, 2017).

The modified Rankin Scale was used to measure functional outcomes in patients with limb muscle weakness. The scale ranges from 0 (asymptomatic) to 6 (death) (Banks and Marotta, 2007). The reduction of modified Rankin Scale scale by at least one point indicated the favourable outcome. Clinical improvement was defined as improvement of muscle strength (most severely affected muscle) at least one grade on MRC scale. Relapse was defined as recurrence of weakness when not attributable to tumour progression or alternative aetiologies.

Ancillary investigations

Electrophysiological testing

Nerve conduction studies (NCS) and electromyography (EMG) were performed according to standard stimulation and recording techniques for the EMG laboratory at Mayo Clinic, Rochester. Two-hertz RNS was utilized to assess a defect of neuromuscular transmission. RNS was deemed abnormal when a physiological pattern of decrement of >10% was seen (Liewluck et al., 2019). SFEMG abnormality was determined by utilizing quality and cut-off guidelines previously published (Sanders et al., 2019).

Antibodies testing

Acetylcholine receptor (AChR) binding and modulating antibodies were detected by radioimmunoprecipitation assay (RIPA) (normal < 0.02 nmol/l) and by monolayer cultures of human muscle cells and quantitated as percentage loss of binding sites for 125 I- α -bungarotoxin (normal \leq 20%), respectively. Striational antibodies were analysed by ELISA immunoassay (normal < 1:120) and diluted to endpoint. Recombinant antigen enzyme-linked immunosorbent assay (QUANTA Lite HMGCR, Inova Diagnostics) detected 3-Hydroxy-3-Methylglutaryl-CoA Reductase (HMGCR-IgG; normal <20 U) (Musset et al., 2014). Myositis-specific [EJ, Jo-1, MDA5, Mi-2, NXP2, OJ, PL-7, PL-12, signal recognition particle (SRP) and TIF-1 γ] and myositis-associated (Ku, PM/Scl-100, SSA, SSB, U1-RNP, U2-RNP and U3-RNP) antibodies were analysed using enzyme immunoassay and RIPA performed by a commercial laboratory (normal <20 U).

Imaging studies

MRI of the head, cervical spine and orbits and whole body [18 F] FDG-PET-CT performed at the time of symptom onset or within three months after symptom onset and following treatment were included when available. All examinations were reviewed by a radiologist (FD) blinded to the patients' clinical state. MRI exams were reviewed for evidence of muscle enlargement, T2 hyperintensity [on any available T2-weighted sequences, included T2 fat-suppressed sequences such as short tau inversion recovery (STIR)], and gadolinium enhancement. Whole body [18 F] FDG-PET-CT scans were reviewed for evidence of increased FDG uptake in skeletal muscles

including the extraocular, masticator, paraspinal and appendicular musculature. In order to be considered positive, unequivocal signal abnormality, enhancement, and/or FDG uptake had to be present, to a degree and in a distribution that could not simply be considered physiologic.

Histopathology

Patients underwent a muscle biopsy for diagnostic purposes. Most samples were processed but all were analyzed in the Mayo Clinic Muscle Laboratory following standard protocol, including haematoxylin & eosin, modified Gomori trichrome, NADH-tetrazolium reductase, succinate dehydrogenase, cytochrome C oxidase (CCO), ATPase, acid phosphatase, myophosphorylase, oil red O, periodic acid-Schiff, non-specific esterase and Congo red stains.

All slides were analysed by two authors (S.S. and T.L.). We quantified each biopsy for the following features per low power fields (LPF): number of necrotic fibres, regenerating fibres, ragged red fibres, ragged-blue fibres and cytochrome c oxidase-negative fibres. We further divided the pattern of necrotic fibres into (i) multifocal clusters [necrotic fibres distributed unevenly among fascicles, with some normal-appearing fascicles and some fascicles containing groups (at least five fibres) of necrotic fibres], or (ii) even distribution (necrotic fibres present diffusely in most fascicles and not clustering).

Statistical analysis

Categorical variables are reported as numbers or percentages and were compared using the Fisher exact test. Quantitative variables are reported as median and ranges and compared using Mann-Whitney two-tail test. The Kaplan–Meier method was used for survival analysis, with the log-rank test used to compare groups. $P < 0.05$ was considered significant. Statistical analysis was performed using JMP software (SAS Institute). For survival analysis data cut-off determined was May 1, 2020, time (months) from ICIAM or IMNM diagnosis to death or last follow-up was collected. The following variables were tested as survival predictive factor in ICIAM: using univariate logistic regression analysis age, gender, CK, time to diagnosis, time from symptom onset to treatment, ptosis, ophthalmoparesis, dysphagia or any bulbar involvement, respiratory involvement, intubation, myocarditis, other irAEs, lymphopenia, ICIAM phenotype (oculobulbar and generalized), striational antibodies and severity of the weakest muscle.

Data availability

Results are available upon reasonable request.

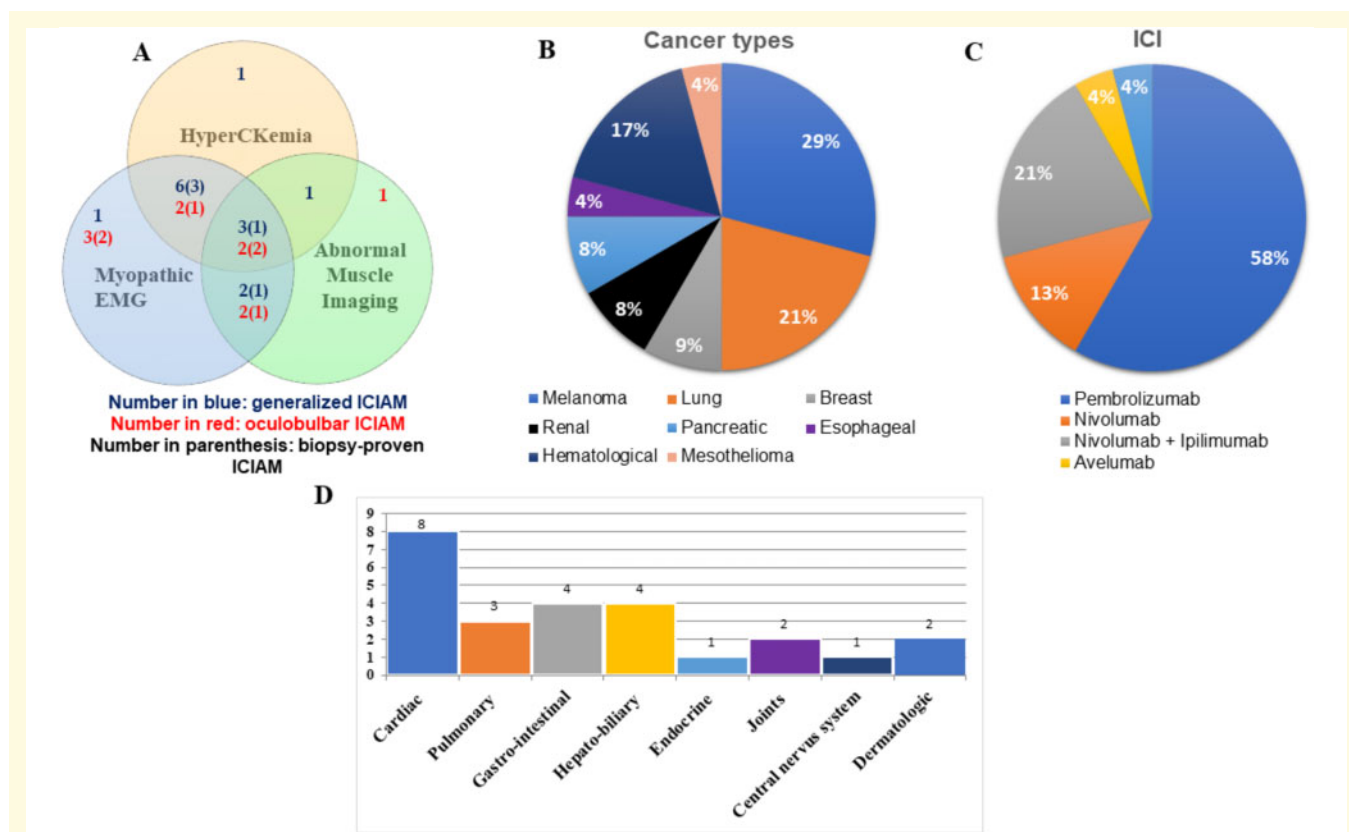


Figure 1 Characteristics of patients with ICIAM. (A) Diagnostic features of ICIAM in the current cohort. Numbers in red and blue indicate numbers of patients with oculobulbar and generalized phenotype, respectively. Numbers in parenthesis indicate numbers of patients who underwent muscle biopsy and had pathologically defined myopathy. There are two patients with possible ICIAM: one oculobulbar patient with abnormal MRI of extraocular muscles (CK and EMG not performed) and one generalized patient with markedly elevated CK level (EMG and MRI not performed). (B) Cancer types in the current cohort. (C) Immune checkpoint inhibitors used in the current cohort. (D) Coexisting immune-related adverse events in the current cohort.

Results

Clinical characteristics

Twenty-four ICIAM (13 definite, 9 probable, 2 possible) and 38 ICI-naïve IMNM patients were included. Figure 1A showed diagnostic features of all ICIAM patients. Among 13 definite ICIAM, 11 patients were myopathologically defined. Three ICIAM patients were previously reported in detail (Liewluck *et al.*, 2018). Patients' demographics, clinical features and statin exposure are shown in Table 1. ICIAM patients with and without muscle biopsy had similar demographics, phenotype and laboratory values (Supplementary Table 1). Underlying malignancies and immune checkpoint regimens in ICIAM patients are outlined in Fig. 1B and C, respectively. In the IMNM group, six patients had active cancers at the time of the myopathy diagnosis. ICIAM patients first noticed symptoms after 1–18 doses (median two doses) of ICI therapy or 1–609 days (median 35 days) after the first dose of ICI treatment. Most ICIAM patients (19/24) developed initial symptoms within the

first three months following ICI initiation. The median time from the last infusion to symptom onset was 8 days (range 0–50 days). The severity of myopathy among 24 ICIAM patients was as follows: CTCAE Grade III ($n=15$), Grade IV ($n=5$) and Grade V ($n=4$). Other coexisting irAEs are shown in Fig. 1D, with myocarditis ($n=8$), being the most common concomitant irAEs.

As shown in Table 1, oculobulbar phenotype was observed in 10 ICIAM patients (four ocular only, three bulbar only and three combined ocular and bulbar weakness) and not in any of the IMNM patients ($P<0.001$). Mild proximal limb or axial weakness was detected in eight oculobulbar ICIAM patients (four proximal only, two axial only and two combined proximal and axial weakness). The remaining ICIAM and all IMNM patients had a generalized weakness, predominantly affecting proximal limb muscles. One generalized ICIAM patient also had ocular and bulbar weakness. Bulbar weakness occurred in both ICIAM and IMNM in a similar proportion, but ocular involvement was only observed in ICIAM group ($P=0.003$). Weakness was non-fatigable in all ICIAM patients and in IMNM patients when tested.

Table 1 Demographics, clinical features and laboratory data of ICIAM versus IMNM patients

	ICIAM	IMNM	P value
Number	24	38	
Age, years (range)	69 (28–86)	55 (18–87)	0.04
Female:male	7:14	19:19	0.5
Follow-up time (days)	396 (11–1506)	731 (74–3602)	0.002
Statin use, n (%)	11 (46)	22 (58)	0.29
Clinical phenotype, n (%)			
1. Oculobulbar	10 (42)	0	<0.001
Ocular involvement	9		
Bulbar involvement	6		
2. Generalized	14 (58)	38 (100)	<0.001
Ocular involvement	1	0	
Bulbar involvement	2	15	
Symptoms, n (%)			
Myalgia	13 (54)	20 (53)	0.26
Dysphagia	9 (38)	12 (31)	0.4
Respiratory insufficiency	10 (42)	10 (26)	0.23
Signs, n (%)			
Ophthalmoparesis	6 (25)	0	0.003
Ptosis	6 (25)	0	0.003
Neck flexor weakness	8 (33)	24 (63)	0.002
Neck extensor weakness	2 (9)	8 (21)	0.19
Hamstrings weaker than quadriceps	6 (25)	15 (40)	0.2
Myocarditis, n (%)	8 (33)	0	<0.001
HyperCKemia, n (%)	15 (68) ^a	37 (100) ^{a,b}	<0.001
CK > 1000 IU/l (%)	10 (45) ^c	37 (100)	0.007
Peak CK, median (range) IU/l	686 (28–7279)	6456 (1151–29 000)	<0.001
Aldolase elevation, n (%)	7/11 (64)	16/16 (100)	0.009
With normal CK, n (%)	2/7 (29) ^d	0	
HMGCR-IgG positive, n (%)	0/13	20/38 (52)	<0.001
SRP-IgG positive, n (%)	0/13	4/38 (10)	<0.001
Other myositis-specific antibodies	0/11	2/29 ^e	0.24
Myositis associated antibodies	3/11	6/29 ^f	0.35
AChR binding antibody positive, n (%)	3/17 (18)	1/3 (33) ^g	0.5
AChR modulating antibody positive, n (%)	2/17 (12)	1/3 (33) ^g	0.4
Striational antibody positive, n (%)	8/17 (47)	1/19 (5) ^g	0.002
Paraneoplastic antibody positive, n (%)	3/17 (18) ^h	0/18	<0.001
Lymphopenia, n (%)	18/24 (75)	7/34 (20)	<0.001
Absolute lymphocyte count, median (range; × 10 ⁹ /l)	0.72 (0.05–3.5)	1.38 (0.56–3.4)	<0.001
Elevated TSH, n (%)	4/24 (19)	12/28 (42)	0.14
AST or ALT elevation, n (%)	15/24 (70)	32/35 (91)	0.003

AChR = acetylcholine receptor; ALT = amino alanine transferase; AST = aspartate amino transferase; CK = creatinine kinase; HMGCR = 3-hydroxy-3-methylglutaryl-coenzyme-A reductase; ICIAM = immune checkpoint inhibitor-associated myositis; IMNM = immune-mediated necrotizing myopathy; NS = not significant; SRP, signal recognition particle; TSH, thyroid stimulating hormone.

All bold values indicate P-value < or = 0.05.

^aCK was not available in 2/24 patients (normal <192 in females and <336 in males).

^bOne patient with normal CK was excluded as it was measured after initiation of immunosuppressive therapy.

^cOculobulbar (n = 2) and generalized (n = 8) phenotypes.

^dBoth patients had normal AST and ALT.

^eOne IMNM patient with strongly positive HMGCR antibodies also had weakly positive MDA-5 antibody; one double seronegative IMNM patient had weakly positive Mi-2 antibody without cutaneous or pulmonary involvement or myopathological hallmark of dermatomyositis.

^fMyositis associated antibodies were positive in three ICIAM (one PM/Sci-100, one SSA and one UI-RNP) and six IMNM (one SSA and five SSB) patients.

^gOne patient with pre-existing myasthenia gravis.

^hOne patient with N type VGCC antibodies (0.08 nmol/l; normal <0.03 nmol/l), one patient with GAD65 antibodies (0.13 nmol/l; normal < 0.02 nmol/l) and one patient with ganglionic nicotinic AChR antibodies (0.04 nmol/l; normal < 0.01).

Ancillary investigations

Laboratory studies

Detailed laboratory findings are shown in [Table 1](#). The CK values were significantly lower ($P < 0.001$) in ICIAM patients (median 686 IU/l compared to IMNM patients (median 6456 IU/l) ([Fig. 2A](#)). The median CK level was not significantly different between pathologically defined ICIAM (493 IU/l) and the remaining ICIAM patients

(1062 IU/l; $P = 0.87$) ([Supplementary Table 1](#)). Seven ICIAM patients had normal CK (five oculobulbar and two generalized). A CK <1000 IU/l was more frequently associated with ICIAM ($P = 0.007$) and even more so with an oculobulbar phenotype ([Fig. 2B](#)). Median CK was 333 IU/l (range 28–3772) in oculobulbar ICIAM and 1284 IU/l (range 30–7303) in generalized ICIAM patients. Aldolase levels correlated with CK values in individual

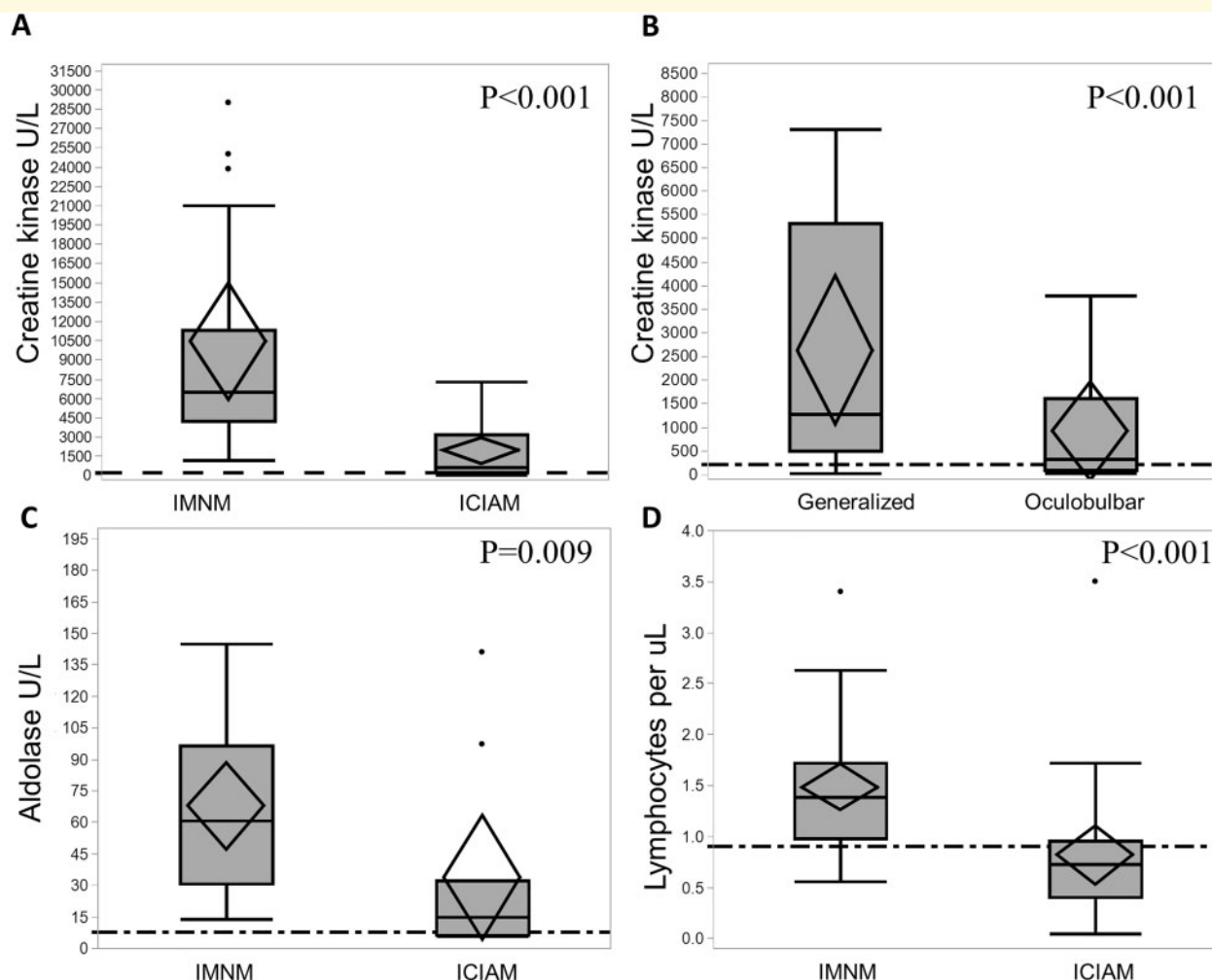


Figure 2 Laboratory data comparison between ICIAM and IMNM patients. **(A)** Patients with ICIAM have lower CK levels compared to those with IMNM. **(B)** Among patients with ICIAM, patients with oculobulbar phenotype have lower CK levels than those with generalized weakness. **(C)** Aldolase levels are significantly higher in the IMNM group compared to the ICIAM group. **(D)** Patients with ICIAM have lower absolute lymphocyte count compared to those with IMNM.

patients (Fig. 2C) except for two ICIAM patients who had elevated aldolase (1.5 and 2 times above upper limit of normal), but normal CK values. Neither of these two ICIAM patients had abnormal liver enzymes. The median time from symptom onset to biopsy was 17 days (range 3–25) in those with normal CK compared to 20 days (range 11–42) in those with hyperCKemia ($P=0.66$). One patient with normal CK received 1-week course of prednisone for unrelated reason 30 days prior to CK measurement, while none of the patients with elevated CK levels received any prior immunomodulatory therapy. Thirteen ICIAM patients were tested for anti-HMGCR and anti-SRP antibodies and all were negative. Amongst IMNM patients, 52% were HMGCR and 10% were SRP antibody positive.

Lymphopenia was more common in ICIAM patients (18/24) compared to IMNM patients (7/34) ($P<0.001$) (Fig. 2D). Pre-ICI lymphocyte count was normal in 10/18

ICIAM patients with lymphopenia at the time of myopathy diagnosis despite 16/18 patients received prior chemotherapy or radiotherapy. After exclusion of eight ICIAM patients with low pre-ICI lymphocyte count, the difference of lymphopenia between ICIAM and IMNM remained significant ($P=0.02$).

Striational antibodies were abnormal in 8/17 ICIAM patients tested (median 1:7680; range 1:3840–1:122880; seven patients with titre $\geq 1:7680$) compared to 1/19 of IMNM patients tested (1:15360) ($P=0.002$). Six ICIAM patients with striational antibodies had oculobulbar phenotype. CK was normal in three of eight striational antibody-positive ICIAM patients. None of the ICIAM patients with striational antibodies had thymoma. Myocarditis was observed in five and two patients with and without striational antibodies, respectively. The calculated specificity of striational antibodies for ICIAM compared to IMNM was 95% with

Table 2 Phenotypic comparison between oculobulbar versus generalized ICIAM patients with acetylcholine receptor antibodies⁶

	Oculobulbar ICIAM (n = 10)		Generalized ICIAM (n = 7)	
	AChR-Ab positive (n = 2) ^a	AChR-Ab negative (n = 8)	AChR-Ab positive (n = 1) ^b	AChR-Ab negative (n = 6)
AChR binding antibody values (nmol/l)	11.9 and 0.1	<0.02	1.43	<0.02
Striational antibody positive, n	2/2	4/8	1/1	1/4
AChR modulating antibody values (%)	93 and 0	<20	67	<20
CK (range, IU/l)	263–444	28–2685 (median 333)	762	611–7307 (median 2924)
EMG	2/2	7/8	1/1	6/6
Myopathic MUP, n	2/2 ^c	7/7 ^c	1/1	6/6
Fibrillation potentials, n	2/2	7/7	1/1	4/6
Abnormal RNS	0/2	0/6	0/1	0/4
Abnormal SFEMG	0/0	0/2	0/0	0/0
Abnormal muscle MRI	1/2 ^d	4/8	0/1 ^e	2/6
Abnormal muscle biopsy	1/1 ^f	5/5	1/1 ^g	3/3
Myocarditis	1/2	3/8	1/1	2/6
Respiratory failure	2/2	3/8	1/1	3/6

⁶The number of patients with positive AChR-Ab is low hence it does not have enough power to make any statistical comparison.

AChR = acetylcholine receptor; CK = creatinine kinase; ICIAM = immune checkpoint inhibitor associated myositis; MUP = motor unit potentials; NA = not applicable; NCS = nerve conduction studies; SFEMG = single-fibre EMG.

^aNormal ranges < 0.02 nmol/l; one patient with ocular weakness and one patient with bulbar weakness.

^bThis patient had axial and proximal weakness without oculobulbar involvement.

^cMyopathic MUP was observed in axial (n = 6), bulbar (n = 6) and limb (n = 8) muscles.

^dMRI abnormalities were observed in bulbar and limb muscles.

^eNormal MRI of brain.

^fSplenius capitis biopsy compatible with ICIAM.

^gDeltoid biopsy compatible with ICIAM.

47% sensitivity when the AUC = 0.7. The cut-off threshold value was 1:3840.

Three ICIAM patients with striational antibodies were also seropositive for anti-AChR antibodies (Table 2). None of these patients had electrophysiological evidence of neuromuscular transmission defect despite severe respiratory insufficiency, but all had short-duration, low-amplitude motor unit potentials with fibrillation potentials, indicating a myopathic process.

Electrophysiological studies

Electrophysiological evaluation was performed in 21 ICIAM patients (9 oculobulbar and 12 generalized) and 37 IMNM patients. Findings are summarized in Supplementary Table 2. Myopathic motor unit potentials were identified in all ICIAM and IMNM patients, when performed. All seven ICIAM patients without limb weakness and seven ICIAM patients with normal CK also had myopathic motor unit potentials in axial or limb muscles. Fibrillation potentials were observed in all, but four ICIAM patients (one oculobulbar and three generalized). Two ICIAM patients, who did not have fibrillation potentials, underwent muscle biopsy which showed necrotic fibres. Myopathic motor unit potentials were accompanied by fibrillation potentials in all, but one IMNM patient who was on immunosuppressive therapy at the time of the EMG. Myotonic discharges were more

common in IMNM (47%) than ICIAM (22%) patients ($P < 0.001$). Six IMNM (16%) and 12 ICIAM (50%) patients had electrodiagnostic evidence of a length-dependent, sensorimotor axonal peripheral neuropathy ($P = 0.01$). One ICIAM patient had axonal polyradiculoneuropathy. The higher frequency of peripheral neuropathy in ICIAM group compared to IMNM group remained significance even after exclusion of eight ICIAM patients with prior chemotherapy ($P = 0.04$). RNS showed no decrement in all 13 (8 oculobulbar and 5 generalized) ICIAM patients performed. Two oculobulbar ICIAM patients underwent SFEMG and both were normal.

Imaging studies

Relevant imaging (MRI or PET) was available in 20/24 ICIAM patients, including 21 MRI and 17 whole body [¹⁸F] FDG PET-CT exams. Intramuscular T2 hyperintensity with or without gadolinium enhancement and/or increased FDG uptake in muscle were seen in 11 ICIAM patients (5 oculobulbar and 6 generalized) (Supplementary Table 3), all of whom had myopathic EMG and 6 had hyperCKemia. Paraspinal muscles were abnormal on imaging studies in all but two patients (one oculobulbar and one generalized), with only one patient with such abnormal paraspinal muscles on imaging having neck extensor weakness on examination. In five

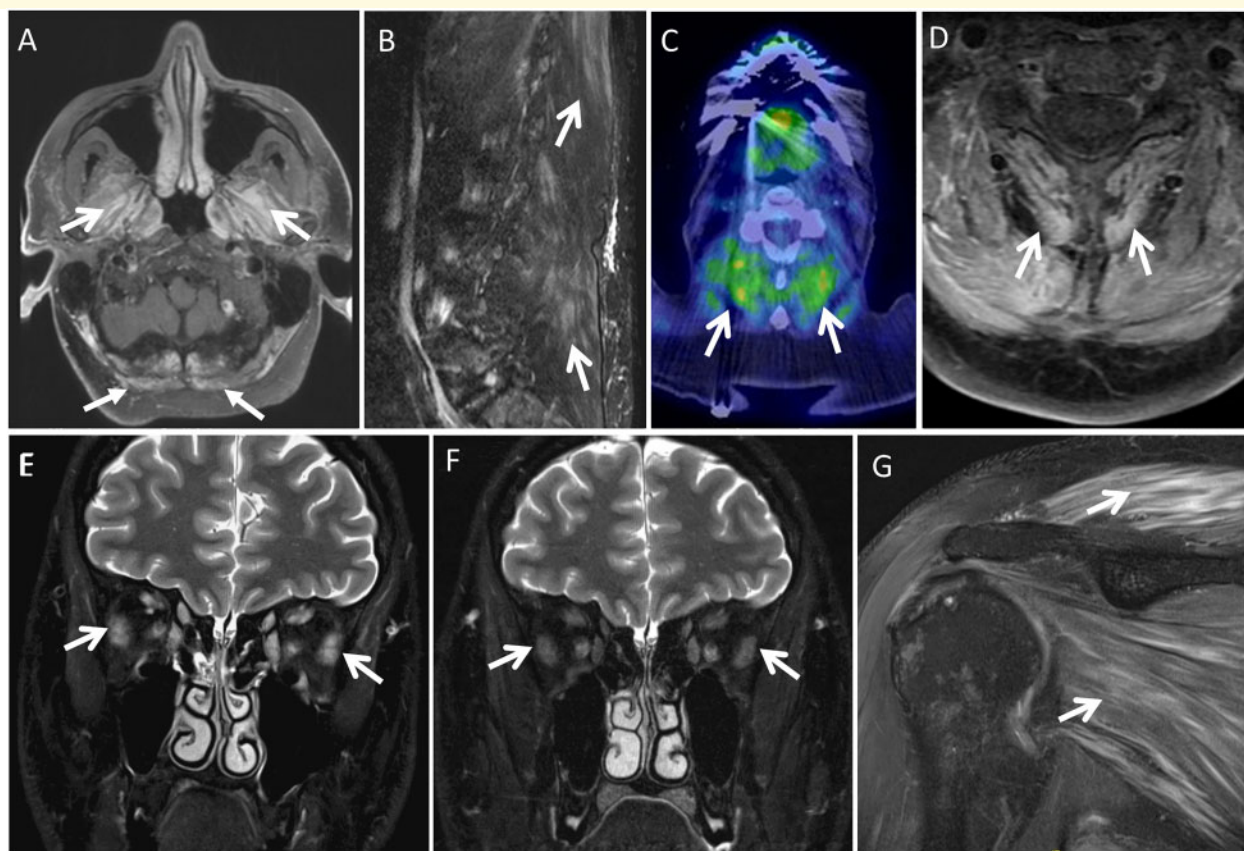


Figure 3 Imaging in ICIAM. Imaging findings of myopathy in five different patients (A–G). (A) Contrast-enhanced axial fat-suppressed T1-weighted brain MRI at the level of foramen magnum reveals contrast enhancement in the muscles of mastication (arrows) and superior posterior neck muscles (block arrows). (B) Off-midline sagittal fat-suppressed T2-weighted lumbar spine MRI demonstrates mild streaky T2-hyperintensity of the lumbar and visualized lower thoracic posterior paraspinal musculature (arrows). (C) Axial 18F-FDG PET CT fused image at C4–5 shows increased FDG uptake in the posterior paraspinal neck musculature (arrows). (D) In the same patient as C, correlative contrast-enhanced axial fat-suppressed T1-weighted cervical spine MRI demonstrates mildly increased enhancement of this posterior paraspinal neck musculature (arrows). (E,F) Coronal fat-suppressed T2-weighted orbit MRI on initial (E) and follow-up (F) initially shows T2 hyperintensity and mild enlargement of the extraocular muscles bilaterally, which resolves on follow-up (arrows on lateral recti). (G) Oblique coronal T2-weighted fat-suppressed shoulder MRI reveals feather like oedema-type T2-hyperintensity enhancement of the shoulder girdle muscles (arrows).

patients with oculobulbar phenotype, muscles of mastication were abnormal in four and extraocular muscles were abnormal in one (Fig. 3). Extraocular muscle abnormality was noted on MRI dedicated to the orbits, while the other four patients with normal radiological findings of extraocular muscles underwent non-dedicated brain MRI. Three patients also underwent MRI and/or PET after immunomodulatory treatment and all three showed a marked decrease of imaging abnormalities (Fig. 3E and F).

Among 10 ICIAM patients who had no radiological abnormalities, imaging studies of clinically affected muscles were not performed in 5 patients (2 oculobulbar and 3 generalized phenotypes). The median time from symptom onset to imaging studies of those radiologically abnormal patients was seven days (range 1–39 days) compared to 16 days (range 0–31 days) in those five patients

with normal imaging studies of clinically affected muscles ($P=0.9$). Myopathic motor unit potentials were observed in all radiologically normal patients. Five patients underwent biopsy of radiologically normal, but electromyographically abnormal muscles, all of which displayed myofibre necrosis.

Muscle pathology

Muscle biopsy slides from 47 patients [11 ICIAM (6 oculobulbar and 5 generalized) and 36 IMNM] were available for review. Several ICIAM patients declined the muscle biopsy because of their terminal illness from cancer itself, severe myopathy or other irAEs requiring ICU care. Data were extracted from biopsy reports of two IMNM patients, whom muscle biopsy slides from other institutes were previously interpreted at our Muscle Lab and slides were no longer available for review. Figure 4 and Table 3

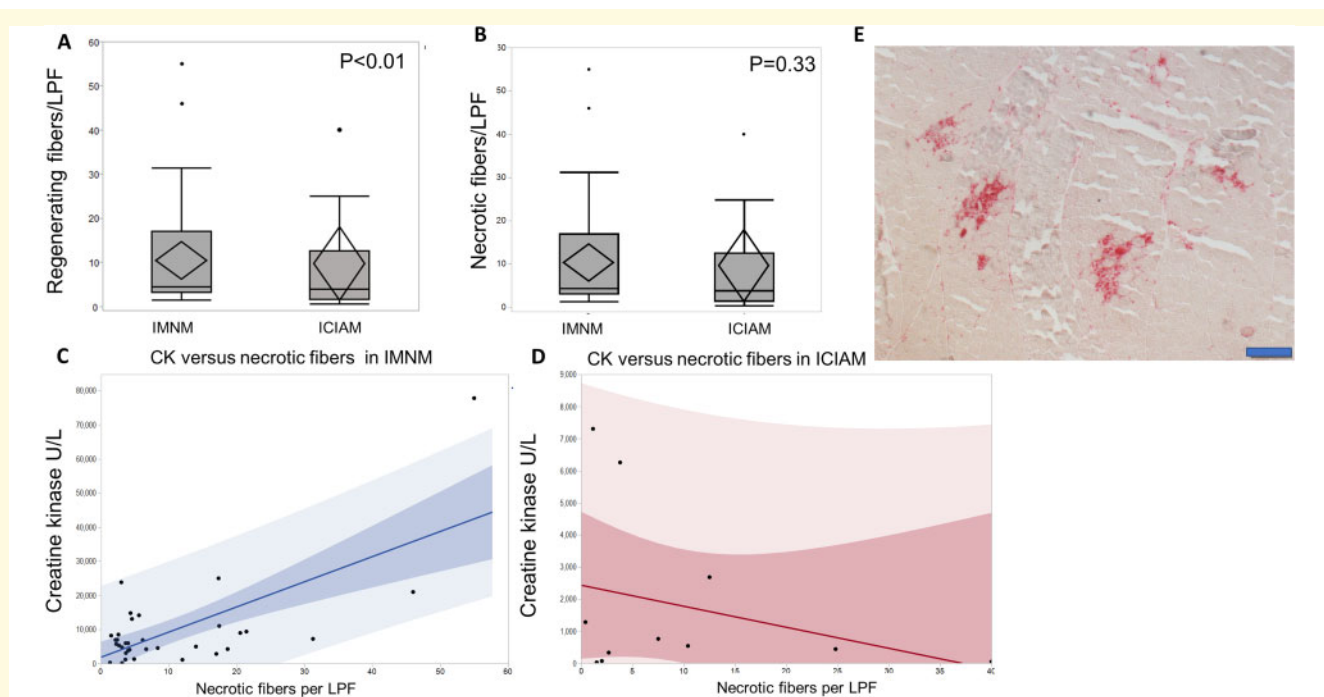


Figure 4 Histopathological comparison between ICIAM and IMNM patients. **(A)** ICIAM patients have lower number of regenerating fiber per low power field compared to IMNM patients. **(B)** ICIAM and IMNM patients have similar number of necrotic fibers per low power field **(B)**. **(C,D)** Correlation between creatinine kinase level and the number of necrotic fibres per low power field demonstrates nonlinear relationship in the ICIAM group and linear relationship in IMNM group. ICIAM patients have relatively similar range of necrotic fibres per low power field but lower CK levels compared to IMNM patients. **(E)** Acid phosphatase stained section shows macrophages (in red) invading necrotic fibres occurring in clusters (bar 200 μm).

summarize the key pathological findings. All muscle biopsies, including four patients with normal CK (three ICIAM and one IMNM), displayed necrotic and regenerating fibres. The IMNM patient with normal CK underwent muscle biopsy prior to immunomodulatory therapy, and the included CK level was measured after treatment.

Necrotic fibres were scattered and evenly distributed throughout the sections in all IMNM patients as opposed to multifocal clusters seen in 10 ICIAM patients ($P < 0.001$, Fig. 4E). A single ICIAM patient with evenly distributed necrotic fibres had dysphagia and mild lower limb weakness for a few months preceding ICI therapy. Multifocal clusters of necrotic fibres had 92% sensitivity and 100% specificity to differentiate ICIAM from IMNM. Few lymphocytes were noted in 3 ICIAM and 12 IMNM patients, but they were too infrequent to diagnose an inflammatory myopathy. ICIAM patients had a significantly lower number of regenerating fibres per LPF and higher number of ragged-blue fibres and cytochrome c oxidase-negative fibres per LPF, while number of necrotic fibres per LPF was not significantly different compared to IMNM patients (Fig. 4A and B). Interstitial histiocytes over-reactive for acid phosphatase in the perimysium were more common in ICIAM ($n = 8$) than IMNM ($n = 3$) patients ($P < 0.001$). CK values directly correlate with the number of necrotic fibres per LPF in

IMNM patients, but not in ICIAM patients (Fig. 4C and D). The number of necrotic fibres/LPF, regenerating fibres/LPF and CK values in oculobulbar and generalized ICIAM patients who underwent muscle biopsy were not significantly different (necrosis; $P = 0.3$, regeneration; $P = 0.7$, CK; $P = 0.4$).

Treatment

ICI was discontinued in 22 patients. ICI was not stopped in two patients (CTCAE Grade III) due to the concern for cancer progression. Both patients received concurrent intravenous methylprednisolone (IVMP) or oral prednisone with normalization of CK and resolution of symptoms. One of them was able to discontinue corticosteroids after 2 months, while the other required ongoing therapy with low dose prednisone for 1.5 years until the last follow-up.

Immunomodulatory therapy was introduced in 22/24 patients, including two patients that ICI was not discontinued. Other two patients declined treatment and went into hospice care. Median time from symptom onset to treatment was 7 days (range 0–95 days). Initial treatments included corticosteroid monotherapy in 16 patients (14 IVMP and 2 oral prednisone) and combined therapy in 6 patients (4 IVMP and plasma exchange, 1 IVMP and IVIG and 1 oral prednisone and IVIG). All patients who received an initial 5-day course of IVMP (1g/day) were

Table 3 Histopathological characteristics of ICIAM versus IMNM patients

	ICIAM	IMNM	P value
Number	11	38	
Mean age at time of biopsy	68	60	0.08
Clinical phenotype:			
Oculobulbar	6 ^a	0	
Generalized	5	38	<0.001
Distribution of necrotic fibres			
Multifocal clusters	10/11	0/36	<0.001
Scattered	1/11	36/36 ^b	
Necrotic fibres per LPF, median (range)	3.78 (0.4–40)	4.7 (1.5–55)	0.7
Regenerating fibres per LPF, median (range)	5 (0–14.6)	8.4 (1–65)	0.003
Inflammation	3/11	13/38 ^{c,d}	0.7
Endomysial	1/3	8/13	<0.001
Perimysial	2/3	9/13	0.4
Invasion of non-necrotic fibre (autoaggression)	0/11	2/38	0.36
Non-rimmed vacuoles, <i>n</i>	2/11	14/38	0.4
Internalized nuclei, <i>n</i>	0/11	8/38	<0.001
Fibre splitting, <i>n</i>	0/11	9/28	<0.001
Fibre type grouping, <i>n</i>	7/11	11/38	0.02
Increased connective tissue	9/11	24/38	0.4
Endomysial	3/11	1/38	
Perimysial	6/11	23/38	
Cytochrome C oxidase-negative fibres, <i>n</i>	9/11	21/38	0.003
Cytochrome C oxidase-negative fibres per LPF, median (range)	1.3(0–5.4)	0.31(0–7.5)	<0.01
Ragged-blue fibres, <i>n</i>	8/11	26/38	<0.01
Ragged-blue fibres per LPF, median (range)	0.67 (0–1.71)	0.14 (0–4)	<0.01
Increased acid phosphatase reactivity in connective tissue, <i>n</i>	8/11	3/38 ^e	<0.001
Sarcoplasmic congophilic deposits, <i>n</i>	1/11	2/38	0.5

LPF = lower power field.

All bold values indicate *P*-value < or = 0.05.

^aMuscle biopsies were taken from triceps (*n* = 3), splenius capitis (*n* = 1), biceps (*n* = 1) and deltoid (*n* = 1). These muscles were not clinically affected but had myopathic changes on needle EMG.

^bMuscle biopsy slides were not available for review in two patients.

^cFour patients had endomysial and perimysial inflammation.

^dOf 13 patient with inflammation, 9 were positive for HMGCR antibodies and the remaining were seronegative.

^eTwo patient were HMGCR antibody positive.

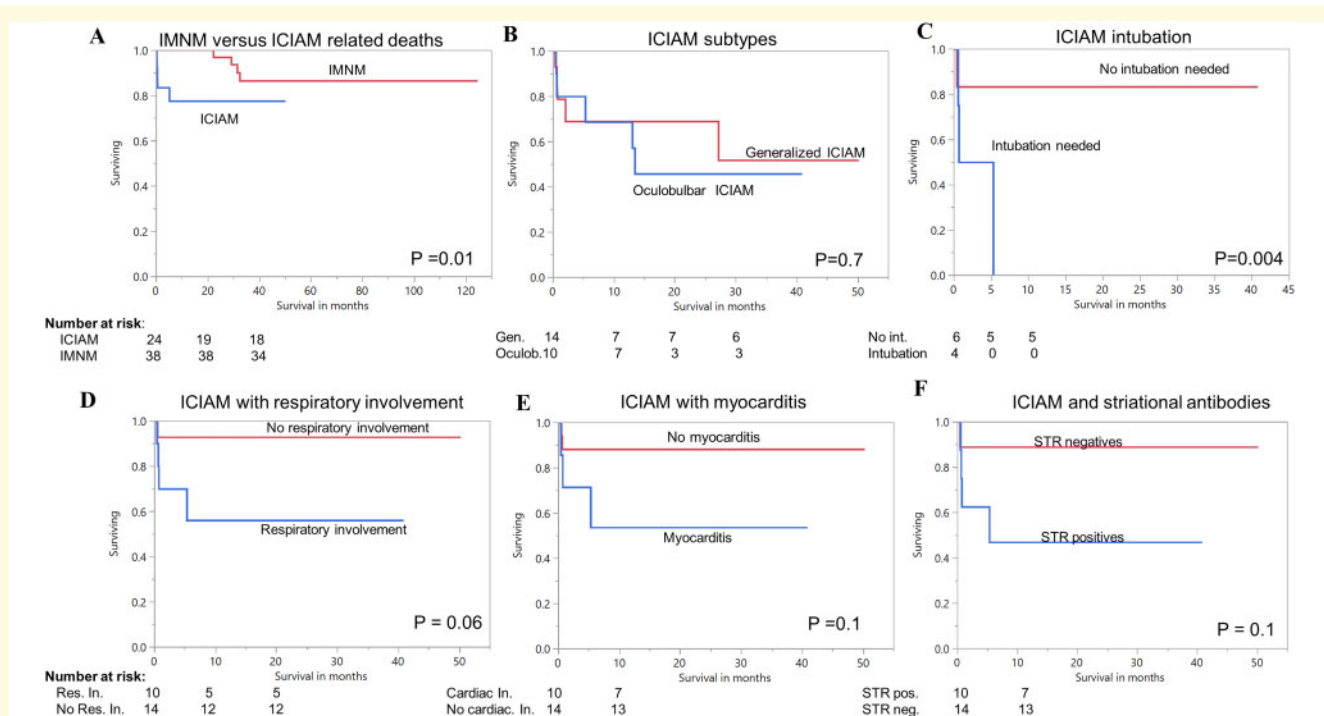
subsequently transitioned to oral prednisone. The median duration of treatment was 9 weeks (range 1–112 weeks). Twenty treated patients showed improvement of muscle strength on the exam; however, one subsequently developed aspiration pneumonia and decided to terminate the treatment for myopathy. Weakness was stabilized in the remaining two patients. The median modified Rankin Scale scores before and after treatment were 3 (range 0–5) and 1 (range 0–6), respectively (*P* = 0.02). Among 14 patients with hyperCKemia, post-treatment CK levels were lower than pre-treatment values in all patients, including eight patients with normalization of CK values. The median time from initiation of immunomodulatory therapy to normal CK was 8 days (range 1–35 days). The patients with persistent hyperCKemia showed a 20–90% reduction (median 37.5%) of CK levels compared to pre-treatment values.

During the follow-up period, eight patients were able to stop immunomodulatory therapy after a median of 5 months of treatment (range 1–24 months) with a median of 23 months of treatment-free (range 0.5–48 months). Sixteen patients required ongoing immunosuppressive therapy with prednisone <20 mg/day (*n* = 13), cyclophosphamide (*n* = 2) or rituximab (*n* = 1). ICI was reinstated in

three patients after the resolution of weakness, one of whom was on low dose prednisone and the others were not on any immunomodulatory therapy at the time of ICI rechallenge. All three patients developed severe irAEs (two recurrent ICIAM and one colitis) after 1–2 doses of ICI.

Survival analysis and outcomes

The overall mortality rate among ICIAM patients was 42% (10/24). Five patients died from ICIAM-related complications (respiratory failure in four CTCAE Grade V patients and myocarditis in one CTCAE Grade III patient) and the remaining five patients died from cancer progression (*n* = 2) or withdrawal/decline of myopathy treatment (*n* = 3). We included only five patients who died from ICIAM or myocarditis in survival analysis. At data cut-offs, 5/24 ICIAM patients died compared to 4/38 IMNM patients (*P* = 0.01). The median overall survival was significantly worse in ICIAM compared to IMNM (Fig. 5A). Probability of survival for ICIAM associated deaths at 0.5, 1 and 2 months was 91%, 83% and 78%, respectively. Probability of survival for IMNM patients at 12 and 36 months was 100% and 82.9%. Overall survival (OS) was similar between



oculobulbar and generalized ICIAM ($P=0.7$; Fig. 5B). Respiratory muscle involvement occurred in 42% ($n=10$) of ICIAM patients with nearly half ($n=4$) requiring intubation. ICIAM patients with intubation ($P=0.004$) had significantly shorter overall survival (median survival of 22 days) but number of patients were too small to draw a definite association. ICIAM patients with striational antibodies, respiratory insufficiency, myocarditis or dysphagia had higher mortality than those without, but none reach statistical significance (Fig. 5D–F).

Discussion

This study reports the clinical, serological and pathological features of the largest cohort of ICIAM patients to date, consisting of 11 patients with pathologically defined myopathy and 13 patients with a diagnosis of myopathy based on combined laboratory, electrophysiological and/or radiological criteria. ICIAM patients with and without muscle biopsies share a similar demographic, clinical and ancillary test profiles. All pathologically defined patients had necrotizing myopathy-like pathology. A systematic comparison between ICIAM and ICI-naïve IMNM

patients discloses several distinctive clinico-sero-pathological characteristics of ICIAM.

Clinical characteristics

Oculobulbar phenotype is a distinctive feature of ICIAM

In most ICIAM patients, myopathy symptoms started shortly after ICI initiation, most commonly after the second dose of ICI. Oculobulbar weakness can be the predominant or sole presentation in nearly half of the ICIAM patients, while the remaining patients had proximal weakness with or without oculobulbar involvement. Bulbar weakness occurred in a similar proportion of ICIAM and IMNM patients, but oculomotor weakness was only observed in ICIAM patients. One-fourth of our ICIAM patients had ophthalmoparesis, which confirm the previous notion of oculomotor involvement as a characteristic feature of ICIAM (Liewluck et al., 2018; Touat et al., 2018; Seki et al., 2019). Ophthalmoparesis is a common manifestation of myasthenia gravis, orbital myositis and certain hereditary myopathies, including mitochondrial myopathies, but is not considered a typical presentation of any other acquired or immune-mediated

myopathies (Kono *et al.*, 2012; Allenbach *et al.*, 2014; Kassardjian *et al.*, 2015; Allenbach *et al.*, 2017; Milone, 2017; McNab, 2020).

CK can be normal in oculobulbar ICIAM, which can mimic myasthenia gravis

Half of our oculobulbar ICIAM patients had normal CK levels in contrast to those previously reported ICIAM patients with oculomotor involvement, who had elevated CK levels (Touat *et al.*, 2018; Seki *et al.*, 2019). The presence of myopathic motor unit potentials and fibrillation potentials in clinically affected or unaffected axial and limb muscles, the absence of decrement on RNS or abnormal jitter on SFEMG, and the observed radiological abnormalities in oculobulbar, axial or appendicular muscles signal the underlying myopathic process. This is further supported by the myopathological evidence of necrotic fibres in three oculobulbar ICIAM patients with normal CK levels. Therefore, it is clinically challenging to distinguish oculobulbar ICIAM from myasthenia gravis when CK level is not elevated.

AChR antibodies in ICIAM: coexisting myasthenia or a marker of autoimmunity?

As mentioned earlier, the myasthenia gravis-myopathy overlap syndrome has been increasingly recognized in ICI-treated patients (Suzuki *et al.*, 2017; Kao *et al.*, 2018). Three of our ICIAM patients were positive for AChR antibodies, all of whom had severe respiratory insufficiency. Myopathic motor unit potentials with associated fibrillation potentials, elevated CK levels and necrotic fibres support a diagnosis of myopathy. It is unclear whether these patients had concomitant myasthenia gravis or the presence of AChR antibodies is simply an indicator of nonspecific autoimmunity (Robbins *et al.*, 2019). Although SFEMG was not performed in any of these three patients, none had decrement on RNS, which is a very sensitive diagnostic tool for a defect of neuromuscular transmission during myasthenic crisis (Oh *et al.*, 2019). The normal RNS in these patients with severe respiratory insufficiency suggests that a defect of neuromuscular transmission, if present, is not a major contributing factor to their symptoms. Some of the reported patients with ICI-associated myasthenia gravis and respiratory insufficiency also had no decrement on RNS, one of whom also had normal SFEMG, suggesting that the myopathy might be the main culprit for the muscle weakness in these reported patients (Loochtan *et al.*, 2015; Sciacca *et al.*, 2016; Chang *et al.*, 2017; Gonzalez *et al.*, 2017; Suzuki *et al.*, 2017; Tan *et al.*, 2017; Fellner *et al.*, 2018).

Ancillary investigations

Few characteristic laboratory findings emerge from the current ICIAM cohort.

ICIAM patients had lower CK levels compared to IMNM patients

CK was not universally elevated in all patients, differing from previous reports (Liewluck *et al.*, 2018; Touat *et al.*, 2018; Seki *et al.*, 2019). Seven ICIAM patients had normal CK level, of which three had muscle biopsies showing necrotic fibres. It is known that the number of necrotic fibres correlates with the CK levels in IMNM (Allenbach *et al.*, 2018a). The number of necrotic fibres in these three patients with normal CK was unexpectedly similar to what observed in ICIAM or IMNM patients with hyperCKemia in our cohort. Therefore, the lower CK levels in some ICIAM patients are not due to the lower number of necrotic fibres in affected muscles but could probably be due to a more focal muscle involvement in these patients.

Two of seven ICIAM patients with normal CK had elevated aldolase levels. Such discrepancy of CK and aldolase levels has been reported in some patients with other immune-mediated myopathies (Carter *et al.*, 2001; Nozaki and Pestronk, 2009; Mathur *et al.*, 2014). A prospective study of aldolase and CK levels in ICIAM patients is crucial to verify the diagnostic utility of aldolase measurement in ICIAM patients with normal CK.

Lymphopenia was more frequent in ICIAM compared to IMNM patients

Lymphopenia encountered at the time of muscle weakness was frequent in ICIAM patients. Although most ICIAM patients received chemotherapy or radiotherapy predated ICI, normal lymphocyte count at the time of ICI initiation suggested that post-ICI lymphopenia was a result of ICI, not other preceding cancer therapies. B cell lymphopenia following ICI administration has been linked to an increased risk of developing irAEs, including ICIAM (Das *et al.*, 2018; Mammen *et al.*, 2019); however, none of our patients underwent lymphocyte subset analysis. Though the mechanism of ICI-induced lymphopenia remains unknown, the routine evaluation of lymphocyte count and subset could be helpful to identify patients potentially at high risk of developing ICIAM or other irAEs.

ICIAM as a seronegative IMNM

While our ICIAM patients had necrotizing myopathy-like pathology, none of the patients tested positive for anti-HMGCR or SRP-antibodies. It is possible that the homogenous phenotype of oculobulbar involvement in ICIAM could be a result of not yet discovered antibody. Striational antibodies were positive in nearly half of our ICIAM patients and were reported in 68% of Japanese programmed cell death-1 inhibitor-associated myopathy cohort (Seki *et al.*, 2019). Six of our eight striational antibody-positive patients had a titre $\geq 1:7680$. Such a high titre of striational antibodies was reported with a high predictive value for underlying cancers (McKeon *et al.*, 2013). Further studies are required to investigate

whether striational antibody positivity can be used as a serologic biomarker for ICIAM.

EMG identified subclinical axial or limb muscle involvement in oculobulbar ICIAM patients

Myopathic motor unit potentials were observed in axial or limb muscles in all ICIAM patients, including the oculobulbar patients with no limb weakness or normal CK. Myopathic changes were accompanied by fibrillation potentials in nearly 85% of patients. EMG can therefore help identify subclinical involvement of axial or limb muscles in oculobulbar ICIAM patients and select the appropriate muscle for biopsy. Several non-myopathic conditions, e.g. deconditioning, malnutrition or myasthenia gravis, may give rise to short-duration, low-amplitude motor unit potentials, mimicking myopathy, but without fibrillation potentials. Muscle biopsy therefore is important to distinguish these pseudomyopathic conditions from myopathy when fibrillation potentials are absent.

Imaging studies identified subclinical cervical paraspinal muscle involvement in ICIAM patients

The systematic assessment of radiological abnormalities in a large cohort of ICIAM patients has not been previously reported. Muscle MRI and PET scans were abnormal in 11/20 patients who had imaging done after symptoms onset. Our result suggests that imaging studies could be utilized as a non-invasive diagnostic tool of ICIAM in conjunction with serologic or electrophysiologic abnormalities, especially when muscle biopsy is not possible. Paraspinal and masticatory muscles were the most common radiologically affected muscles. Radiological abnormalities mirrored the clinical findings in most patients. Interestingly the paraspinal musculature was abnormal on imaging in nearly 50% of patients despite neck extensor weakness identified in only a minority of patients. In comparison to electrophysiological studies, imaging studies are particularly useful in evaluating muscles that are not accessible (e.g. extraocular muscles) or not routinely examined by needle EMG (e.g. muscle of mastication, trapezius or cervical paraspinal muscles) in cases of suspected myopathy. However, needle EMG appears more sensitive than imaging studies when it comes to the evaluation of limb muscles given that all of the radiologically normal ICIAM patients had myopathic changes on needle EMG. Imaging also has sensitivity and specificity limitations, as muscles groups such as extraocular muscles, muscles of mastication and paraspinal muscles can have findings physiologically on imaging. For example, normal extraocular muscles are known to demonstrate physiologic FDG uptake, even to quite prominent degree or robustly enhance on MRI. Thus, as in the example shown in Fig. 3, T2 hyperintensity and/or enlargement of these muscles are more reliable when attempting to identify myopathy. These findings may be seen to better advantage on dedicated orbit MRI than non-dedicated brain MRI.

Multifocal clusters of necrotic fibres are unique to ICIAM

Despite various histopathologically characterized immune-mediated myopathies have been reported in ICIAM patients, all 11 ICIAM patients in our cohort who underwent muscle biopsy had necrotizing myopathy-like pathology. However, by comparing to ICI-naïve IMNM muscle biopsies, we identify multifocal clusters of necrotic fibres as a distinctive pathological feature of ICIAM similar to what previously reported (Seki *et al.*, 2019). Although CK values were significantly lower in ICIAM patients compared to IMNM patients, we observed similar numbers of necrotic fibres between ICIAM and IMNM. Unlike IMNM, the number of necrotic fibres did not correlate with the CK levels in ICIAM. The number of regenerating fibres was significantly lower in ICIAM compared to IMNM, which may suggest an impaired regeneration in ICIAM.

ICIAM muscle biopsies also showed a higher number of cytochrome c oxidase-negative and ragged-blue fibres, which indicate mitochondrial dysfunction, and more frequent interstitial histiocytes compared to IMNM biopsies. The pathological evidence of mitochondrial dysfunction in ICIAM is of uncertain significance. The possibility that the histological signs of mitochondrial dysfunction represent the result of drug toxicity cannot be excluded. Given ophthalmoparesis is a common manifestation of mitochondrial myopathies, whether the mitochondrial changes observed here have any effect on the pathogenesis of extraocular muscle weakness remain to be elucidated. The presence of macrophages in connective tissue elements have been previously reported in ICIAM patients and also in anti-HMGCR antibody-IMNM (Alshehri *et al.*, 2015; Touat *et al.*, 2018; Seki *et al.*, 2019).

Treatment

ICIAM is a steroid-responsive myopathy

All treated patients received steroids, most as a monotherapy. Weakness was improved in most patients and stabilized in a couple of patients after immunomodulatory therapy. In those two patients with CTCAE Grade III, whom ICI was never stopped, symptoms resolved with concurrent immunomodulatory therapy. About one-third of the patients were in remission and was off any immunomodulatory therapy for a median of nearly 2 years, including one patient whom ICI was never discontinued. Therefore, concurrent treatment of ICIAM, while on ICI, may be possible in mild cases, but close follow-up is warranted. The ICI rechallenge in patients with ICIAM has not been widely reported (Delyon *et al.*, 2019). In our cohort, all three patients who were restarted on ICI had serious irAEs necessitating cessation of ICI. Based on our experience, it would be prudent not to reintroduce ICI in patients with ICIAM.

Intubation is a poor prognostic factor in ICIAM

To our knowledge, this study is the first to analyse survival in ICIAM. As expected, ICIAM has an overall worse survival compared to IMNM group. ICIAM patients with severe respiratory failure requiring intubation had a significantly shorter survival. The ICIAM-related mortality rate in our cohort was 20%, which is similar to what reported in World Health Organization (WHO) database of individual safety case reports (Anquetil *et al.*, 2018). However, the WHO database identified coexisting myocarditis as a poor prognostic factor, but authors had only limited access to the detailed medical records of those patients and survival curves or overall survival was not performed. Despite myocarditis occurred in one-third of our patients, myocarditis was not a poor predictor of survival in our study. Two other cohorts of ICIAM estimated mortality rate at 30–50%, but all their patients died from cancer-related causes and no death was associated with ICIAM (Touat *et al.*, 2018; Seki *et al.*, 2019).

Limitations

Patients in this series received non-standardized investigations and treatments, as these were at the discretion of the treating physicians. The exclusion of Grades 1 and 2 ICIAM due to the uncertainty of myopathy diagnosis in those patients could contribute to the bias when compared ICIAM patients to IMNM cohort, especially regarding the disease severity and mortality. Despite these limitations, this study has revealed comprehensive spectrum of the clinical and laboratory features of ICIAM.

Conclusion

ICIAM typically occurs within the first 3 months of ICI initiation and nearly half of the patients had predominant oculobulbar weakness with no or subtle proximal limb involvement. CK levels were normal in one-third of patients, most of which had oculobulbar phenotype. All ICI-treated patients, who develop oculobulbar, axial or limb weakness, should undergo EMG regardless of the CK values. Needle EMG of cervical paraspinals and muscles of mastication should be considered if needle EMG of limb muscles are unrevealing. Positive AChR or striational antibodies should be interpreted cautiously when the patients have severe weakness and no electrodiagnostic evidence of neuromuscular transmission defect. Imaging studies can help evaluate cranial muscles, including for instance extraocular muscles, which are not easily accessible by needle EMG. Concomitant irAEs are not uncommon, especially myocarditis. Intubation is a poor prognostic factor. ICI should be discontinued and immunomodulatory therapy should be commenced promptly. Necrotic fibres arranging in multifocal clusters are distinctive features of ICIAM. Future studies looking into

the regenerating capacity and mitochondrial dysfunction of muscle fibres may shed light on the pathomechanism of ICIAM. Lymphocyte count and subset analysis may help identify patients who are at risk of developing ICIAM or other irAEs.

Supplementary material

Supplementary material is available at *Brain Communications* online.

Competing interest

Dr. Liewluck reports receiving a discretionary fund from the Department of Neurology, Mayo Clinic for unrelated research projects. Dr. Milone reports receiving research funding from a Mayo Clinic benefactor and a discretionary fund from the Department of Neurology, Mayo Clinic for unrelated research projects, and an honorarium to serve as associated editor of *Neurology Genetics*. Dr. Zekeridou reports a patent on PDE10A-IgG as a biomarker of paraneoplastic neurological autoimmunity. Drs. Shelly, Triplett, Pinto and Diehn report no disclosure.

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