

RESEARCH ARTICLE

Continuous administration of a p38 α inhibitor during the subacute phase after transient ischemia-induced stroke in the rat promotes dose-dependent functional recovery accompanied by increase in brain BDNF protein level

John J. Alam^{1*}, Michael Krakovsky², Ursula Germann¹, Aharon Levy²

1 EIP Pharma, Inc., Boston, Massachusetts, United States of America, **2** Pharmaseed Ltd., Ness-Ziona, Israel

* jalam@eippharma.com



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Abstract

There is unmet need for effective stroke therapies. Numerous neuroprotection attempts for acute cerebral ischemia have failed and as a result there is growing interest in developing therapies to promote functional recovery through increasing synaptic plasticity. For this research study, we hypothesized that in addition to its previously reported role in mediating cell death during the acute phase, the alpha isoform of p38 mitogen-activated protein kinase, p38 α , may also contribute to interleukin-1 β -mediated impairment of functional recovery during the subacute phase after acute ischemic stroke. Accordingly, an oral, brain-penetrant, small molecule p38 α inhibitor, neflamapimod, was evaluated as a subacute phase stroke treatment to promote functional recovery. Neflamapimod administration to rats after transient middle cerebral artery occlusion at two dose levels was initiated outside of the previously characterized therapeutic window for neuroprotection of less than 24 hours for p38 α inhibitors. Six-week administration of neflamapimod, starting at 48 hours after reperfusion, significantly improved behavioral outcomes assessed by the modified neurological severity score at Week 4 and at Week 6 post stroke in a dose-dependent manner. Neflamapimod demonstrated beneficial effects on additional measures of sensory and motor function. It also resulted in a dose-related increase in brain-derived neurotrophic factor (BDNF) protein levels, a previously reported potential marker of synaptic plasticity that was measured in brain homogenates at sacrifice. Taken together with literature evidence on the role of p38 α -dependent suppression by interleukin-1 β of BDNF-mediated synaptic plasticity and BDNF production, our findings support a mechanistic model in which inhibition of p38 α promotes functional recovery after ischemic stroke by blocking the deleterious effects of interleukin-1 β on synaptic plasticity. The dose-related *in vivo* efficacy of neflamapimod offers the possibility of having a therapy for stroke that could be initiated outside the short time window for neuroprotection and for improving recovery after a completed stroke.

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Competing interests: I have read the journal's policy and the authors of this manuscript have the following competing interests: John Alam is the scientific founder and CEO of EIP Pharma, Inc., a private company based in Cambridge, Massachusetts, USA, that provided funding for these studies and is developing neflamapimod (VX-745) as a potential treatment for CNS disorders. Michael Krakovsky and Aharon Levy are employees of Pharmaseed Ltd., the pre-clinical laboratory contracted by EIP Pharma, Inc. to conduct the studies in this report. Ursula Germann is scientific advisor to EIP Pharma, Inc. for neflamapimod research studies. This does not alter our adherence to PLOS ONE policies on sharing data and materials.

Introduction

Stroke is a frequent cause of death as well as a leading cause of acquired disability worldwide and is associated with a substantial economic burden due to high costs for treatment and post stroke care [1, 2]. Approximately 80% of strokes are ischemic in nature due to thromboembolic occlusion of a major artery or its branches, leading to a cascade of events that causes irreversible tissue injury [3]. Based on pathological characteristics and their timing, a stroke is classified into three clinical phases, including the acute (i.e. first 48 hours after stroke onset), the subacute (from 48 hours to >6 weeks post stroke) and the chronic phase (starts at 3–6 months post stroke) [4, 5]. The acute phase represents an opportunity to salvage threatened tissue and reduce the extent of injury (i.e. provide neuroprotection), for example via reperfusion or neuroprotection while the subacute phase represents the recovery stage [5, 6]. The subacute phase is characterized by brain repair initiation, so therapeutic strategies include enhancing the underlying spontaneous recovery processes, modifying inflammation, lifting diaschisis, or reducing late neuronal death [5, 6]. Only regenerative approaches would generally be considered to be potentially active in the chronic phase [7].

The only approved pharmacological intervention for acute ischemic stroke is intravenous thrombolysis with recombinant tissue plasminogen activator (TPA), resulting in recanalization of occluded vessels if applied within a short time period (up to 4.5 hours) after stroke [8]. Numerous other attempts at providing neuroprotection during the acute phase of stroke have failed [9–11] and there is an urgent need for alternative, more widely applicable treatment options for ischemic stroke. Such therapeutics might enable treatment of patients who present after the very short time window for thrombolysis, and of patients who are ineligible for intravenous TPA treatment. In particular, there is high interest in the development of novel therapies that are directed at promoting functional recovery from stroke via increasing neuronal and synaptic plasticity during the subacute phase [5, 9, 12]. The main goal is to identify disease-modifying treatments that can be administered after the acute phase of stroke is complete, i.e. treatments that can be administered during the subacute and/or chronic phase [5, 9, 12]. It is expected that the proposed approaches generally target compromised cerebral tissue and/or surrounding intact tissue to promote brain plasticity [5, 12].

For a number of reasons the proinflammatory cytokine interleukin-1beta (IL-1 β) is considered a therapeutic target for treatment of ischemic stroke to promote recovery after stroke. IL-1 β is upregulated after ischemic stroke [13–17] and in subacute/chronic inflammatory conditions, IL-1 β is known to be a key component of the inflammatory response in the brain that mediates neurodegenerative effects of inflammation on cognition and synaptic plasticity [18]. Chronic elevation of IL-1 β , such as IL-1 β elevation in the aging brain, suppresses brain-derived neurotrophic factor (BDNF) production [19, 20], and *in vitro* it has been shown that increased IL-1 β inhibits BDNF effects on neuronal/synaptic plasticity via induction of the intracellular kinase, p38 mitogen-activated-protein-kinase (MAPK) [17, 21]. In the context of recovery after stroke, these effects of IL-1 β on BDNF may be a significant block to recovery as BDNF plays a critical role in neural plasticity and recovery after stroke [22]. P38 MAPK otherwise is a key component of IL-1 β signaling, particularly with respects to its proinflammatory effects. In the brain, IL-1 β signals both through the ubiquitous, but low-affinity proinflammatory IL-1 receptor accessory protein (IL-1RAP) that signals through p38 MAPK, as well through a high-affinity neuron-specific isoform of IL-1RAP that signals via a different kinase, the non-receptor tyrosine kinase Src [23, 24]. IL-1 β signaling via the ubiquitous p38 MAPK-dependent IL-1RAP inhibits long-term potentiation, while signaling via the neuron-specific Src-dependent IL-1RAP facilitates long-term potentiation [25]. These two distinct IL-1 β signaling pathways, a low-affinity p38 MAPK-dependent one and a high-affinity Src-dependent one, may be the

reason why in many systems low basal concentrations of IL-1 β promote synaptic plasticity, while at high concentrations (i.e. those seen under inflammatory conditions) IL-1 β inhibits synaptic plasticity [17, 20, 25]. All the above-described research findings taken together stimulated the idea to investigate whether inhibition of p38 MAPK with a specific small molecule inhibitor may have beneficial efficacy as a subacute phase stroke treatment via promoting functional recovery through blocking the deleterious effects of IL-1 β on BDNF action and production, and with it on synaptic plasticity.

Otherwise, activation of p38 MAPK, particularly the alpha isoform (p38 α), after experimental ischemic stroke in rodents has been demonstrated in neurons, astrocytes and microglia [26–30], and p38 α has been established as a driver of neuroinflammation-mediated cell death in the acute phase of ischemic stroke [31, 32]. Therefore, several inhibitors of p38 MAPK that exhibit different potency and kinase selectivity, all of them most potently blocking p38 α versus other p38 isoforms (p38 β , p38 γ , p38 δ) have been administered during the acute phase in experimental models of cerebral ischemia, and all of them have provided robust neuroprotection [30, 33–37]. Importantly, administration of a p38 MAPK inhibitor was neuroprotective (i.e. reduced infarct size) when administered up to 6- and 12-hours post stroke, but not when administered 24 hours post stroke in a rat transient middle cerebral artery (tMCAO) model [30].

Heretofore, beyond its role during the acute phase, it had been unclear whether p38 α also plays a role in impairing functional recovery during the subacute phase of stroke which embodies brain repair initiation [4–6]. Time course analyses of phospho-p38 (i.e. the activated form of p38) expression in experimental rat stroke models show a biphasic response. In association with the acute inflammatory response, marked increases in phospho-p38, including phospho-p38 α , are seen immediately after cerebral ischemia within neurons and other cell types [27, 29, 30, 36]. The acute increase primarily resolves within the first 24 hours, but then there is progressive elevation to 10 to 14 days, the last time points measured in the two studies that assessed p38 expression beyond the acute phase [36, 38]. A one-week study of a novel p38 α inhibitor, the tetra-substituted thiophene VCP979, evaluated as a treatment of photothrombotic ischemic stroke in streptozocin-induced Type 2 diabetes mellitus (T2DM) mice demonstrated beneficial efficacy with functional outcome seven days post stroke, associated with both neuroprotection (reduced infarct volume) and axonal/white matter remodeling within the motor cortex [37]. Whereas this study offered novel mechanistic insights for a p38 α inhibitor that are relevant for brain repair, it did not address whether the functional recovery was due to neuroprotection or due to effects on recovery mechanisms, as VCP979 administration started at 24 hours post stroke [37], at a time point that is within the acute phase of photothrombotic stroke in mice [39, 40]. Indeed, as VCP979 demonstrated almost an approximate two-thirds reduction in infarct size compared to vehicle administration, the improved functional recovery was likely due to its neuroprotective effects [37].

The pyrimido pyridazine neflamapimod (International Non-proprietary Name for the molecule previously code-named VX-745) is a potent, highly selective, ATP-competitive inhibitor of p38 α that yields higher central nervous system (CNS) than peripheral blood exposure after oral administration [41, 42]. It exhibits efficacy and safety profiles in preclinical species to merit ongoing clinical investigation for CNS disorder indications [42]. Neflamapimod is currently being evaluated in the clinic for its potential to reverse synaptic dysfunction in three different CNS disorders (Alzheimer's disease, Huntington's disease, dementia with Lewy bodies). Phase 2 clinical studies in Alzheimer's disease have been reported [42–44].

Neflamapimod has a potency (IC₅₀) between 10 and 15 ng/mL for IL-1 β -induced production of IL-6 or IL-8 from human peripheral blood mononuclear cells (i.e. for IL-1 β signaling) [45]. Accordingly, neflamapimod was evaluated in 20- to 24-month old Fischer rats with cognitive deficits, which previously have been described to be due to IL-1 β -induced impairment of synaptic

plasticity [46]. Results showed that neflamapimod administered via oral gavage at a dose level of 1.5 mg/kg administered twice daily reversed deficits in spatial learning in the Morris-Water-Maze test, a measure of synaptic plasticity. However, that same dose had no effects on hippocampal IL-1 β levels, while a 3-fold higher dose (4.5 mg/kg administered twice daily) reduced IL-1 β levels in the hippocampus but did not result in pro-cognitive effects [45]. These results suggested that the neflamapimod pro-cognitive effects were mediated through decreasing IL-1 β signaling in hippocampal neuron target cells, rather than through reducing IL-1 β production [45].

In recognition of the unmet need for stroke therapies that enable initiation of therapy at a time point beyond the therapeutic window for neuroprotection and show potential for promoting functional recovery via beneficial effects on synaptic plasticity, the objectives of the present study were (1) to evaluate neflamapimod at two previously reported pharmacologically active and clinically relevant dose levels for its *in vivo* efficacy to promote neurologic recovery as assessed by the modified neurological severity score (mNSS) and additional behavioral tests in a rat tMCAO model; (2) to initiate administration of neflamapimod outside the known neuroprotection window for p38 α inhibitors and to address whether p38 α inhibitor treatment is still effective when given at a later time post stroke; (3) to assess the neurogenic factor BDNF in the brain as a potential biomarker for monitoring neflamapimod effects on synaptic plasticity; and (4) to measure IL-1 β levels as an inflammatory biomarker in the brain after the treatment period to gauge whether chronic inflammation may play a role in this experimental stroke model and whether a neflamapimod effect related to IL-1 β signaling may be detectable.

Materials and methods

To enhance the reproducibility of results presented in this study, a downloadable protocol file has been deposited at <http://dx.doi.org/10.17504/protocols.io.bf69jrh6>.

Animals and general health monitoring

Seventy-six young (3-month old) male Sprague Dawley rats (Harlan Laboratories, Israel) weighing 328 g \pm 20% were included in the study. The protocol for the study was approved by the Israeli Animal Care and Use Committee (approval number IL-15-01-15) and was conducted in accordance with the Israeli guidelines that conform to the United States Public Health Service's Policy on Humane Care and Use of Laboratory Animals. Animal handling was performed according to guidelines of the National Institute of Health (NIH) and the Association for Assessment and Accreditation of Laboratory Animal Care (AAALAC). To assess the health status of the animals throughout the study, the general health status was monitored daily and body weight was determined weekly.

Transient middle cerebral artery occlusion

Transient middle cerebral artery occlusion (tMCAO) in the right brain hemisphere was performed on Day 1 according to a modified method of Longa *et al.* [47] as described by Schmid-Elsaesser *et al.* [48], in which a 4 cm length of 4–0 monofilament nylon suture is inserted under anesthesia through the proximal external carotid artery into the internal carotid artery and thence into the circle of Willis, effectively occluding the middle cerebral artery. A thermostatically regulated heating lamp and pad were used to maintain rat temperature at 37°C during the tMCAO procedure. Anesthesia was induced with 4% isoflurane in a mixture of 70% N₂O and 30% O₂ and maintained with 1.5–2% isoflurane. The surgical wound was subsequently closed, and the animals were returned to their cages to recover from anesthesia. Two hours after occlusion rats were re-anesthetized, monofilament was withdrawn to allow reperfusion, surgical wound was closed, and rats were returned to their cages.

Neurologic scoring and behavioral evaluations

The individual performing neurologic scoring and behavioral assessments was unaware of the group assignments, thus, performed blinded evaluations. Neurologic scoring by mNSS evaluation according to Chen *et al.* [49] was performed at least one day before the Day 1 tMCAO, and on Day 2, at Week 4, and at Week 6 following tMCAO. The mNSS is a composite of motor, sensory, reflex and balance tests and well-defined score values [49]), that were used to assess the effect of neflamapimod compared to vehicle control. Taking into account the different tests and score values, the mNSS was graded on a composite scale of 0 to 18, in which the normal, healthy animal value was 0 and the maximal deficit value after tMCAO was represented by 18 [49] (S1 Table). An overall mNSS value of ≥ 10 was predefined as an inclusion criterion for enrolling a rat with stroke into the neflamapimod treatment study.

Additional behavioral evaluations included stepping test, body swing test, and forelimb placement test that were performed before the tMCAO procedure, and at Week 4 and Week 6 following tMCAO.

A stepping test was utilized to assess forelimb akinesia according to Pharmaseed's internal protocol. Each rat was held with its hind limbs fixed in one hand and the forelimb, not to be monitored, in the other, while the unrestrained forepaw touched the table. The number of adjusting steps of the forelimb to be monitored was counted while the animal was moved sideways along the table surface in the forehand and backhand direction over a distance of 85 cm during approximately five seconds. The stepping test was completed for both the left and right forelimbs at all time points indicated above.

For the body swing test, each rat was held approximately one inch from the base of its tail. It was then elevated to an inch above a surface of the table. The rat was held in the vertical axis, defined as no more than 10° to either the left or the right side. A swing was recorded whenever the rat moved its head out of the vertical axis to either side. Before measuring another swing, the rat must have returned to the vertical position. Twenty (20) total swings were counted. A normal rat typically had an equal number of swings to either side. Adjusted body swing scores were calculated as the difference between leftward and rightward swings (i.e. the number of rightward swings subtracted from leftward swings).

For the forelimb-placing test, the examiner held the rat close to a tabletop and scored the rat's ability to place the forelimb on the tabletop in response to whisker, visual, tactile, or proprioceptive stimulation. Separate sub-scores were obtained for each mode of sensory input and added to give total scores (0 = normal, 12 = maximally impaired). Scores were given in half-point increments: whisker placing (0–2); visual placing (forward (0–2) and sideways (0–2)); tactile placing (dorsal (0–2) and lateral (0–2)); proprioceptive placing (0–2).

Administration of test articles

Test articles, neflamapimod at two different dose levels in 1% (w/v) Pluronic F108 vehicle, or vehicle (1% (w/v) Pluronic F108) were supplied by EIP Pharma, Inc. and administered at a dosing volume of 5 ml/kg by oral gavage twice daily (7 AM and 7 PM) on 5.5 days (i.e. Sunday through Friday at 7 AM and Sunday through Thursday at 7 PM) every week for six weeks (i.e. from Day 3 until Day 42), starting at 48 hours post reperfusion. The dose levels of neflamapimod administered were 1.5 mg/kg and 4.5 mg/kg twice daily.

Sample collection and analysis

On Day 44, two days following the final dosing of test article or vehicle, rats were sacrificed by CO₂ inhalation. Brains were harvested from all animals and samples were divided into left and right hemisphere. Samples were weighed, immediately frozen in liquid nitrogen, and stored at

-80°C. For the IL-1 β and BDNF analyses, tissue samples were defrosted and homogenized in 1 ml/200 mg tissue of 20 mM Tris-HCL pH 7.4 containing a protease inhibitor cocktail (50 μ l/ml). Samples were centrifuged at 10,000 x g for 15 minutes at 4°C. Clear supernatants were aliquoted (150 μ l/aliquot) and stored at -80°C until enzyme linked immunosorbent assay (ELISA) for IL-1 β or BDNF was performed. For IL-1 β analysis, ELISA was performed using the Rat IL-1 beta/IL-1F2 Quantikine ELISA Kit (R&D Systems, Minneapolis, MN) according to manufacturer's instructions. For BDNF analysis, ELISA was performed using the Solid Phase Sandwich Quantikine ELISA Kit (96-well strip plates, R&D Systems, Canada) according to manufacturer's instructions. For IL-1 β and BDNF, standards and samples were tested in duplicates. For IL-1 β assay, all the results below the lower limit of quantification (LLOQ) of 20 pg/mL were assigned half the LLOQ, i.e. 10 pg/mL. For the BDNF assay, all the results were above the LLOQ.

Statistical considerations

Based on previous therapeutic intervention studies at Pharmaseed, a sample size of 15 animals per treatment group was considered to be the minimum number required to provide indicative information regarding potential drug treatment effect. In order to account for anticipated mortality in the first few days associated with transient ischemic stroke, 76 animals (~25 per group) were initially enrolled in the study and underwent surgery to induce stroke.

Except for the analysis of BDNF levels, all *P*-values reported are from statistical analysis performed by one- or two-way analysis of variance (ANOVA) for repeated measures, followed by Bonferroni post hoc test to adjust for the multiple comparisons. The Bonferroni correction was for the six comparisons that were conducted for each parameter: vehicle vs. each of two drug groups (1.5 mg/kg, 4.5 mg/kg), at each of 3 time points (Day 2, Week 4 and Week 6). One-way ANOVA was also used to compare the mNSS values at two different time points (e.g. Week 4 versus Day 2, or Week 6 versus Week 4) within an individual test group. Detailed output of the ANOVA results is provided in the [S1 Appendix](#) in the Supporting information.

Because the BDNF protein level data were not normally distributed, the BDNF statistical analyses were conducted utilizing nonparametric approaches. Specifically, the distribution-free trend test, the Jonckheere-Terpstra test, was utilized to assess for treatment-dependent dose effect across the vehicle, 1.5 mg/kg neflamapimod, and 4.5 mg/kg neflamapimod dose groups, since the result values were not normally distributed. Additionally, the Kruskal-Wallis test with Dunn's post hoc test for multiple comparisons was performed to compare individual neflamapimod dose groups to the vehicle group. The detailed output of the BDNF statistical analysis is provided in the [S2 Appendix](#) in the Supporting information.

Except for the Jonckheere-Terpstra test, all statistical analyses were performed utilizing Prism (GraphPad Software). As there are no nonparametric tests to test dose-response in Prism, the Jonckheere-Terpstra test was programmed and analyzed utilizing S-PLUS (Tibco Software).

Results

Transient ischemia in rats caused significant increase in mNSS value without high incidence of mortality during the acute phase (i.e. 0–48 hours) after stroke

Seventy-six (76) young adult, 3-month old male Sprague-Dawley rats were included in the study and underwent tMCAO. In order to limit the number of animals undergoing surgery and on-study neurologic examinations on any given day, the study was conducted in four

cycles of nineteen animals each that underwent tMCAO on the same day. Nine animals were lost during Day 1 and four additional animals were lost during Day 2 after the procedure, resulting in ~17% mortality during the first 48 hours post tMCAO attributable to the severity of these rat's stroke.

To confirm induction of stroke on Day 2 of each cycle, at 24 hours after reperfusion, surviving animals were subjected to neurological evaluation using the mNSS value (S1 Table [49]) that rates neurological functioning on a scale from 0 (healthy) to 18 (maximum impairment) [50]. The measured mNSS value in individual rats rose from 0 before the Day 1 tMCAO procedure to a mean±standard deviation (SD) value of 14.0±1.4 on Day 2, with individual animal mNSS values ranging from 12 to 17.

On Day 3 of each cycle, at 48 hours post stroke, the 63 surviving animals were allocated based on Day 2 mNSS values and assigned to one of three treatment groups: twice daily 1.5 mg/kg neflamapimod (22 rats initiated treatment), twice daily 4.5 mg/kg neflamapimod (21 rats initiated treatment), or twice daily vehicle (20 control rats initiated treatment) during 5.5 days every week for a total of six weeks.

The first administration of neflamapimod treatment or vehicle control administration occurred on Day 3 at 48 hours post reperfusion. Three additional rats, including one animal from the 1.5 mg/kg neflamapimod dose group and two vehicle control rats, did not survive the first two weeks after initiation of dosing. Since these additional deaths were inversely related to the dose of neflamapimod, they were attributed to the severity of their stroke. The 60 remaining rats ($n = 21$ in the 1.5 mg/kg and $n = 21$ in the 4.5 mg/kg neflamapimod groups, respectively, and $n = 18$ in the vehicle group) completed the planned six weeks of dosing and were included in the neurologic and behavioral evaluations at Week 4 and Week 6.

Observations of animal health preservation including body weight gain after initiation of neflamapimod or vehicle treatment during the subacute phase after stroke

The Day 1 mean±SD body weight data for the animals assigned to each group revealed were well balanced (328.5±11.3 g for the 1.5 mg/kg and 325.3±11.1 g for the 4.5 mg/kg neflamapimod treatment group, respectively, and 328.2±9.6 g for the vehicle control group). Weekly body weight monitoring and two-way ANOVA statistics followed by Bonferroni post hoc comparisons revealed no statistically significant differences in body weight for the three study groups at any time throughout the study. The mean±SD body weight for the 3-month old rats was 327.0±10.9 g on Day 1 and all surviving animals had a mean±SD body weight 418.3±27.6 g at Week 6, and a similar body weight gain of ~27% was observed in all three test groups throughout the study period (S2 Table). These observations together with favorable results for all the daily health assessments point out that the neflamapimod and vehicle treatments were generally well-tolerated, providing no treatment-related adverse clinical signs. Moreover, these findings exclude the possibility that a difference in the general health of the rats contributed to neflamapimod treatment-mediated effects on functional recovery when compared to the effects observed in the vehicle-treated group.

Dose-related improvement in neurologic and behavioral mNSS after initiation of neflamapimod versus vehicle treatment during the subacute phase post stroke

On Day 2, the mean±SD values for mNSS were similar across the groups of stroked rats, indicating that the three randomized experimental groups were balanced a day prior to treatment initiation (Fig 1A). The measured mNSS mean±SD values on Day 2 were 13.9±1.1 in the 1.5

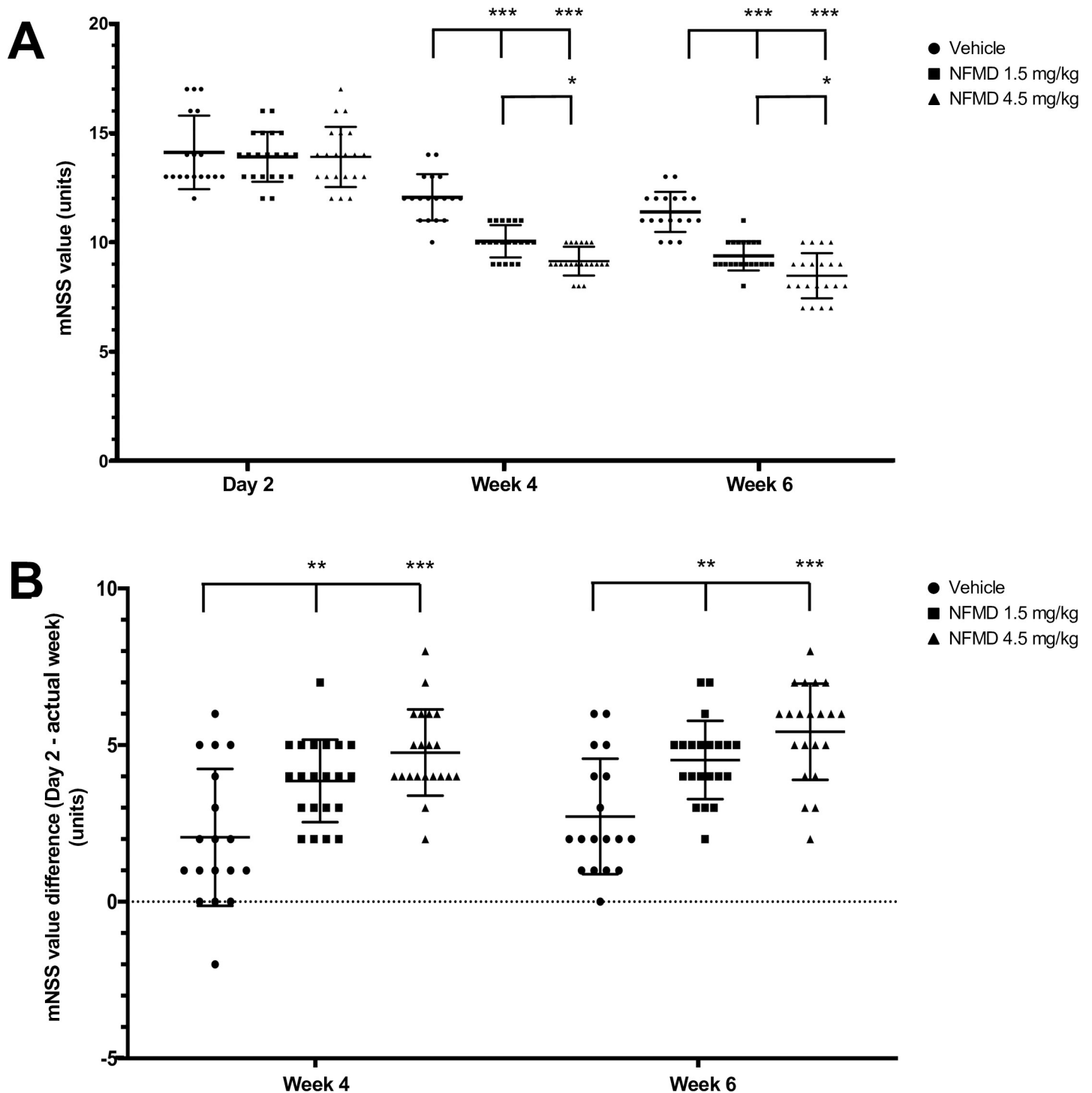


Fig 1. Dose-related neflamapimod (NFMD) subacute treatment effects on mNSS of rats with tMCAO. The study included three groups: vehicle (Pluronic F108; $n = 18$), 1.5 mg/kg neflamapimod ($n = 21$), and 4.5 mg/kg neflamapimod ($n = 21$). (A) Mean \pm SD mNSS value at Day 2, Week 4 and Week 6. Both at Week 4 and Week 6, the mean \pm SD mNSS values were decreased and significantly lower in each neflamapimod dose group when compared to vehicle. In addition, a dose response effect on improvement in neurologic function was indicated by the lower mean \pm SD mNSS value in the 4.5 mg/kg neflamapimod dose group when compared to that of the 1.5 mg/kg dose group. (B) Absolute changes in mNSS \pm SD from Day 2 to Week 4 and Day 2 to Week 6. Significant mNSS differences were observed at both time point comparisons for 1.5 or 4.5 mg/kg neflamapimod dose groups compared to vehicle compared. Significance of between group differences was assessed using a two-way analysis of variance (ANOVA) with a Bonferroni correction ($*P < 0.05$; $**P < 0.01$; $***P < 0.001$; see text for exact P -values > 0.001).

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mg/kg and 13.9 ± 1.4 in the 4.5 mg/kg neflamapimod dose group, and 14.3 ± 1.7 in the vehicle control group. In light of the maximum possible mNSS value of 18 (S1 Table), all animals exhibited severe neurologic deficits prior to neflamapimod treatment initiation.

Importantly, clear differences were demonstrated between the groups treated with neflamapimod compared to the vehicle-treated group. As shown in Fig 1A, the mean \pm SD mNSS values were statistically significantly lower (in the 1.5 mg/kg dose group (10.0 ± 0.7 at Week 4 and 9.4 ± 0.7 at Week 6; ANOVA, Bonferonni post hoc test vs. vehicle at Week 4 $t(37) = 6.049$, $P < 0.001$, and at week 6 $t(37) = 5.761$, $P < 0.001$) and in the 4.5 mg/kg dose group (9.1 ± 0.7 at Week 4 and 8.5 ± 1.0 at Week 6; ANOVA, Bonferonni post hoc test vs. vehicle at Week 4 $t(37) = 8.641$, $P < 0.001$, and at week 6 $t(37) = 8.353$, $P < 0.001$) at both time points when compared to vehicle (see above, 12.1 ± 1.1 at Week 4 and 11.4 ± 0.9 at Week 6). These results document a positive effect of neflamapimod on neurologic repair following induction of ischemic stroke. Furthermore, a dose-related effect on improvement in neurologic function was indicated by the statistically significant lower mean mNSS in the 4.5 mg/kg dose group when compared to the 1.5 mg/kg dose group (ANOVA, Bonferonni post hoc test, $t(40) = 2.698$, $P = 0.03$ at Week 4; $t(40) = 2.698$, $P = 0.03$ at Week 6). Findings were similar when absolute changes in mNSS from Day 2 to Week 4 and Day 2 to Week 6 in the 1.5 mg/kg dose group (3.9 ± 1.3 and 4.5 ± 1.2 , respectively) and the 4.5 mg/kg dose group (4.8 ± 1.4 and 5.4 ± 1.5 , respectively) were compared to the absolute changes in the vehicle group (2.1 ± 2.2 and 2.7 ± 1.8 , respectively), as shown in Fig 1B. Statistically significant differences in absolute change in mNSS were observed at both time point comparisons for the 1.5 mg/kg dose group (ANOVA, Bonferonni post hoc test $t(37) = 3.511$, $P = 0.003$ at Week 4; $t(37) = 3.511$, $P = 0.003$ at Week 6) and the 4.5 mg/kg dose group (ANOVA, Bonferonni post hoc test $t(37) = 5.072$, $P < 0.001$ at Week 4; $t(37) = 5.267$, $P < 0.001$ at Week 6) compared to the vehicle control group.

The vehicle group results for the mean \pm SD mNSS values (12.1 ± 1.1 at Week 4 and 11.4 ± 0.9 at Week 6 versus 14.3 ± 1.7 on Day 2) imply a limited degree of spontaneous recovery of neurologic functions in these study animals from Day 2 to Week 4, as well as from Week 4 to Week 6.

Neflamapimod-mediated improvement in motor and sensory function behavioral tests during the subacute stroke phase

The findings from additional behavioral evaluations of motor and sensory functions (stepping test, body swing and forelimb placement) that were performed to complement the mNSS are presented in Fig 2.

In the left forelimb stepping test, rats in both neflamapimod dose groups had a statistically significant greater mean \pm SD number of steps at Week 4 (9.9 ± 1.5 in 1.5 mg/kg group and 12.6 ± 1.4 in 4.5 mg/kg group; ANOVA, Bonferonni post hoc test vs. vehicle, $t(37) = 5.103$, $P < 0.001$ for 1.5 mg/kg; $t(37) = 9.541$, $P < 0.001$ for 4.5 mg/kg) and Week 6 (13.0 ± 1.2 in 1.5 mg/kg group and 14.4 ± 1.4 in 4.5 mg/kg group; ANOVA, Bonferonni post hoc test vs. vehicle, $t(37) = 11.09$, $P < 0.001$ for 1.5 mg/kg; $t(37) = 12.65$, $P < 0.001$ for 4.5 mg/kg) compared to vehicle-treated animals (7.6 ± 1.3 at Week 4 and 8.7 ± 1.6 at Week 6) as shown in Fig 2A. Supporting that the effects of neflamapimod were specific to neurologic recovery following tMCAO, the mean \pm SD number of steps for the right forelimbs were similar in all groups at Week 4 (19.1 ± 0.4 for 1.5 mg/kg, 19.1 ± 0.4 for 4.5 mg/kg and 18.97 ± 0.2 for vehicle group) and Week 6 (19.1 ± 0.4 for 1.5 mg/kg, 19.2 ± 0.4 for 4.5 mg/kg and 19.1 ± 0.2 for vehicle group), respectively, and comparable to pre-stroke baseline left forelimb (19.8 ± 0.6 for 1.5 mg/kg, 18.8 ± 3.3 for 4.5 mg/kg and 19.7 ± 0.8 for vehicle group) or baseline right forelimb (20.0 ± 0.7 for 1.5 mg/kg, 20.1 ± 0.7 for 4.5 mg/kg and 19.7 ± 0.8 for vehicle group) values.

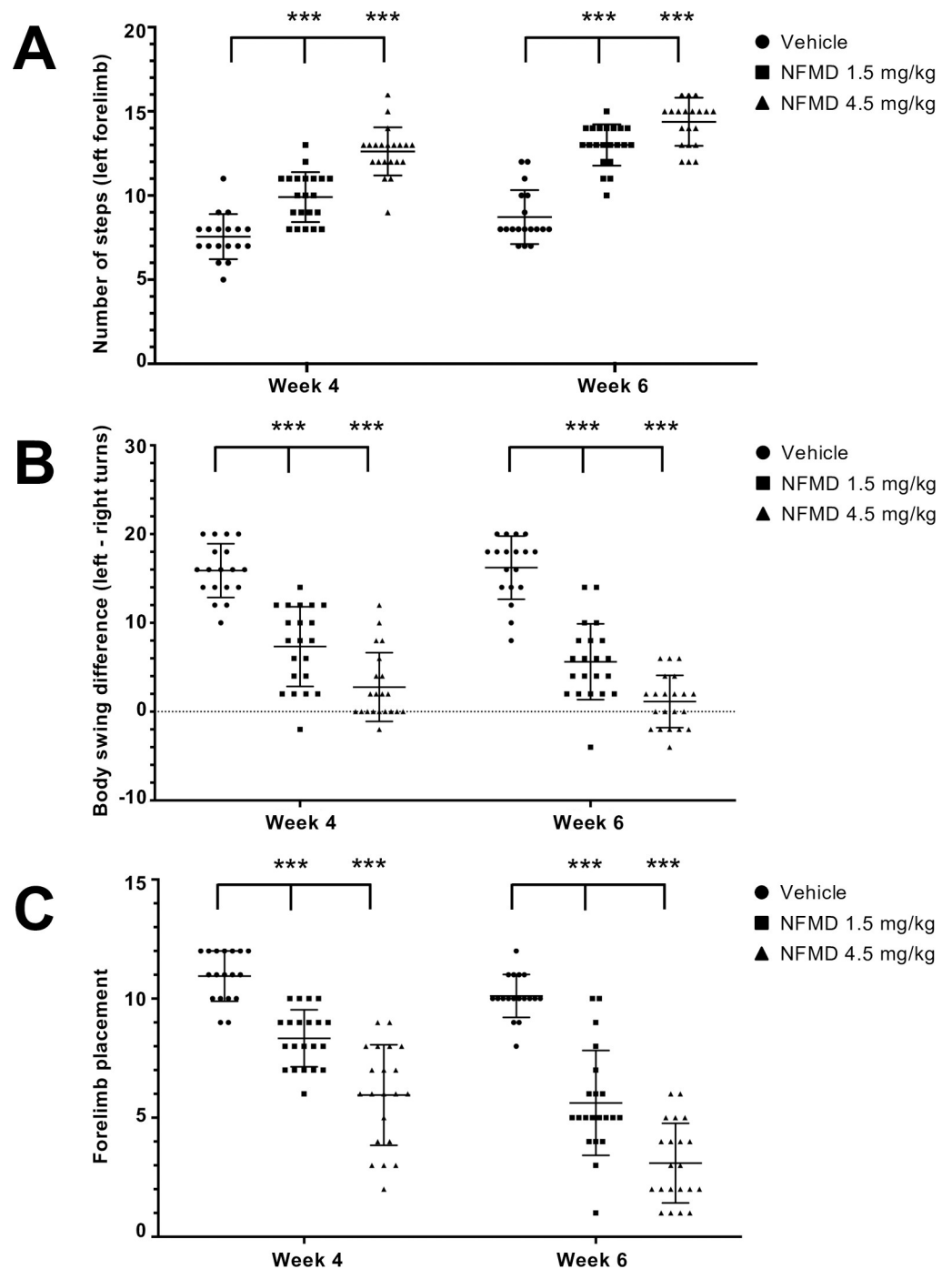


Fig 2. Effects of four- and six-week neflamapimod (NFMD) subacute treatment on stepping, body swing and forelimb placement tests of rats with tMCAO. Treatment groups include vehicle ($n = 18$), 1.5 mg/kg neflamapimod ($n = 21$), and 4.5 mg/kg neflamapimod ($n = 21$). (A) Stepping test. Rats in both neflamapimod dose groups took a significant greater mean \pm SD number of left forelimb steps at Week 4 and Week 6 compared to vehicle-treated animals. (B) Body swing test. Mean \pm SD adjusted body swing score values at Week 4 and Week 6 in the 1.5 mg/kg, 4.5 mg/kg dose groups were significantly lower when compared to vehicle group. (C) Forelimb placement test. Mean \pm SD forelimb placement scores at Week 4 and Week 6 in the 1.5 mg/kg and 4.5 mg/kg dose groups were significantly lower when compared to vehicle group. Significance of between group differences was assessed using ANOVA with a Bonferroni correction (* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; see text for exact P -values > 0.001).

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Additional results from the body swing (BSW) tests indicate a more prominent motor recovery in neflamapimod-treated animals. The mean \pm SD adjusted BSW score in the tests at Week 4 and Week 6 in the 1.5 mg/kg (6.8 \pm 4.6 and 5.6 \pm 4.2, respectively; ANOVA, Bonferonni post hoc test vs. vehicle, $t(37) = 7.511$, $P < 0.001$ at Week 4; $t(37) = 8.75$, $P < 0.001$ at Week 6) and 4.5 mg/kg (2.8 \pm 3.9 and 1.1 \pm 2.9, respectively; ANOVA, Bonferonni post hoc test vs. vehicle, $t(37) = 10.9$, $P < 0.001$ at Week 4; $t(37) = 12.46$, $P < 0.001$ at Week 6) dose groups were all statistically significantly lower when compared to vehicle group (15.9 \pm 3.0 and 16.2 \pm 3.6, respectively) at the same time points, as presented in Fig 2B.

Finally, the mean \pm SD forelimb placement scores in the forelimb placement tests at Week 4 and Week 6 in the 1.5 mg/kg (8.3 \pm 1.2 and 5.5 \pm 2.2, respectively; ANOVA, Bonferonni post hoc test vs. vehicle, $t(37) = 4.941$, $P < 0.001$ at Week 4; $t(37) = 8.742$, $P < 0.001$ at Week 6) and 4.5 mg/kg (6.0 \pm 2.1 and 3.1 \pm 1.7, respectively; ANOVA, Bonferonni post hoc test vs. vehicle, $t(37) = 9.312$, $P < 0.001$ at Week 4; $t(37) = 13.3$; $P < 0.001$ at Week 6) neflamapimod dose groups were also statistically significantly lower when compared to vehicle group (10.9 \pm 1.1 and 10.1 \pm 0.9, respectively), as shown in Fig 2C. The forelimb placement test scored the above-mentioned mean \pm SD forelimb placement scores positive values for the affected left forelimbs post stroke only. All the forelimb placement test scores for the left and right forelimbs at baseline, as well as all the values for the right forelimbs at Week 4 and Week 6 were zero. The lower forelimb placement scores observed in the neflamapimod-treated animals suggest a treatment effect of neflamapimod on recovery of somatosensory function following ischemic stroke.

Study termination biomarker evaluation showing no significant effect of neflamapimod treatment on IL-1 β production, while providing evidence for dose-dependent neflamapimod treatment-associated increase in brain BDNF protein levels

A total of 18 tissue samples for the right and the left brain hemisphere from each animal per group, plus the standard curve, were the maximum number of assays that could be included on the ELISA plate that was utilized to evaluate the effects of neflamapimod on IL-1 β and BDNF biomarkers after study termination. Importantly, all brain tissue samples from the vehicle group were included in this analysis. However, since both the 1.5 mg/kg and the 4.5 mg/kg neflamapimod dose group consisted of 21 animals, the right and left brain tissue samples from three animals each had to be omitted. Selection of the samples to be omitted from the two neflamapimod dose groups was made arbitrarily by the technician in the ELISA lab who was blinded to other data. It was determined in advance, that the right and left brain tissue samples from the 17th, 18th, and 19th animal (by animal number, lowest to highest) in each of the two neflamapimod dose groups would not be analyzed.

The ELISA signal in the IL-1 β assay in the unaffected left brain hemisphere were below the level for noise in the assay (i.e. below LLOQ of 20 pg/mL) in all rats in all groups. However, 9 of 18 animals in the each of the vehicle and 1.5 mg/kg neflamapimod groups and 6 of 18 animals in the 4.5 mg/kg neflamapimod group had quantifiable IL-1 β levels above 20 pg/mL in the injured right brain hemisphere, indicating that despite being six weeks from acute the stroke there was still detectable residual inflammation in a substantial percentage of the animals. Quantifiable IL-1 β levels ranged from 21.3 pg/mL to 203.5 pg/mL, though all but three rats had levels below 100 pg/mL (S3 Table). The mean \pm SD IL-1 β levels in the right hemisphere was 44.3 \pm 55.7 pg/ml in the vehicle group, 30.8 \pm 25.9 pg/ml in the 1.5 mg/kg neflamapimod group and 28.5 \pm 32.4 pg/ml in the 4.5 mg/kg group; with no statistically significant difference between these groups.

BDNF protein was detectable in both brain hemispheres of all animals on Day 44 post stroke as shown in Fig 3, which presents the median and the lower (25th percentile) and upper

(75th percentile) interquartile range (IQR) values for the different groups. Within each brain hemisphere there was a significant dose-related effect of neflamapimod for increasing BDNF protein levels (Jonckheere-Terpstra test: $J = 2.595$, $P = 0.0047$ and $J = 1.673$, $P = 0.047$ for the left (Fig 3B) and right (Fig 3A) hemisphere, respectively). In addition, in the left hemisphere (non-affected side), BDNF levels were statistically significantly higher in the 4.5 mg/kg group (median 2315 pg/ml (IQR: 2016–3463 pg/ml)) than in the vehicle group (median 1884 pg/ml (IQR: 1573–2283 pg/ml)) based on the results of the Kruskal-Wallis test with Dunn's post hoc test for multiple comparisons ($Z = 2.569$; $P = 0.0204$). This is visualized by the horizontal line on top with the two drop downs at its ends, and the star above the 4.5 mg/kg neflamapimod dose group in Fig 3B. The left hemisphere BDNF levels in the 1.5 mg/kg group (median 2085 pg/ml (IQR: 1792–2402 pg/ml)) were intermediate to those in the vehicle and the 4.5 mg/kg group, although not statistically significantly different from the vehicle group when the 1.5 mg/kg and vehicle groups were compared directly.

Discussion

The results for the daily general health assessments, the weekly body weight measurements, the neurologic recovery assessments on Day 2, at Week 4 and at Week 6 post stroke, and the terminal biomarker measurements on Day 44 in this rat tMCAO study taken together led to the following seven key observations for evaluation of delayed and prolonged neflamapimod treatment. (1) Neflamapimod treatment dose-dependently and continually improved neurologic and behavioral outcomes assessed by mNSS and specific measures of sensory and motor function post stroke (Figs 1 and 2). (2) Neflamapimod dose-dependently increased terminal brain BDNF protein as a neurogenic factor biomarker measure for beneficial effects on synaptic plasticity (Fig 3). (3) Neflamapimod and vehicle treatments were generally well-tolerated throughout the study, providing no treatment-related adverse clinical signs. (4) There was an average body weight gain of 27% in all treatment groups throughout the six-week study period (S2 Table) and there was no difference observed in the general health of the rats that would have contributed to any neflamapimod versus vehicle treatment-mediated functional recovery. (5) Slight, albeit statistically significant spontaneous recovery over time was also observed in the vehicle control group (an average ~19% decrease in mNSS value within six weeks; Fig 1) implying that neflamapimod *in vivo* efficacy may enhance a biologic process, like neural or synaptic plasticity, that is already active. (6) Generally, the functional recovery was slow and steady in all neflamapimod- or vehicle-treated animals throughout the prolonged study period (Figs 1 and 2). (7) There were signs for unresolved IL-1 β -mediated chronic inflammation in the injured brain hemisphere of a subset of animals, however, no neflamapimod effect on IL-1 β production was apparent at study termination (S3 Table). Taken together, the results for this research study provide evidence that there is an additional clinical opportunity for p38 α inhibitors in ischemic stroke since neflamapimod treatment initiated outside the neuroprotection time window in a rat tMCAO model promoted recovery.

The most important finding of this study is that the delayed and prolonged treatment with neflamapimod resulted in dose-related neurologic and behavioral improvements versus vehicle control, when this p38 α inhibitor was administered orally to rats with tMCAO in an experimental therapeutic paradigm to enhance synaptic plasticity and functional recovery during the recovery phase after ischemic stroke. To this end, neflamapimod treatment was started at 48 hours after reperfusion at a time when the acute stroke phase was considered complete, since this represents a time point that is outside the previously characterized neuroprotection window of less than 24-hours post tMCAO-induced stroke for a p38 α inhibitor [30]. In the most relevant previously published study, the p38 MAPK inhibitor SB203580 was administered via

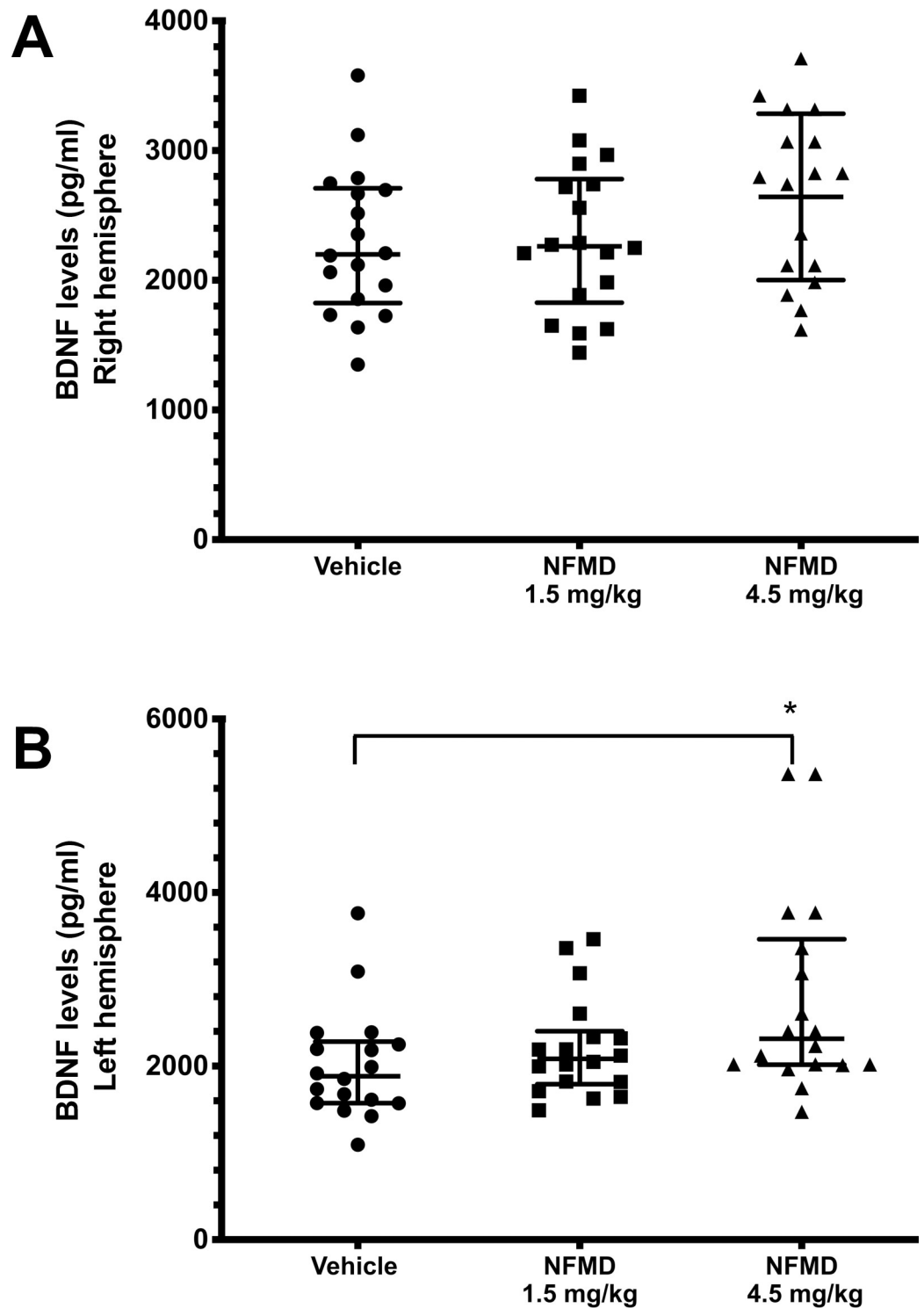


Fig 3. Dose-related neflamapimod (NFMD) subacute six-week treatment effects on rat brain BDNF levels analyzed on Day 44 post tMCAO. Brain homogenate samples from 18 animals in each treatment group (vehicle, 1.5 mg/kg neflamapimod, and 4.5 mg/kg neflamapimod) were prepared two days after the six-week treatment period and analyzed by BDNF ELISA. In the analysis of the right brain hemisphere within the 4.5 mg/kg neflamapimod group, there was an accidental assay failure for one sample, so BDNF ELISA results exist for 17 animals only. The median BDNF value and the 25th and 75th percentile are presented for each different group. (A) Right hemisphere (tMCAO): A significant neflamapimod dose trend by the Jonckheere-Terpstra test for higher BDNF levels was observed ($P < 0.05$) for across the

three groups. (B) Left hemisphere (non-injured side): The Jonckheere-Terpstra test indicated a significant neflamapimod dose trend for higher BDNF levels in the left hemisphere ($P < 0.01$). BDNF levels were also significantly higher in the 4.5 mg/kg neflamapimod group when compared with the vehicle control group ($P < 0.05$). See text for exact P -values > 0.001 .

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intracerebroventricular injection 30 minutes pre-tMCAO, or at 6, 12 or 24 hours post tMCAO to Sprague Dawley rats and demonstrated a decrease in infarct volume that was associated with anti-inflammatory effects in the injured brain and improved recovery from neurologic deficit measured on Day 2 post stroke for all the treatment initiations up to 12 hours post tMCAO, whereas no beneficial effect was observed for the treatment initiation at 24 hours post tMCAO [30]. The observation that a 12-hour time window for the start of p38 α inhibition, but not a 24-hour window allowed for neuroprotective effects in the rat tMCAO-induced stroke model supports the selection of 48-hours as the time point for initiation of subacute phase neflamapimod dosing in the current study, since the results by Piao *et al.* [30] indicated that it is too late a time point for a p38 α inhibitor in this type of model to lead to reduction of neuronal loss as a mechanism for impacting functional outcome.

While infarct size was not measured in this study (the harvested brain was instead utilized to measure protein biomarkers), the acute mortality rate, the high mNSS score at Day 2, and modest spontaneous recovery in the vehicle animals are observations that are consistent with prior validation studies of this model conducted at Pharmaseed, in which the infarct size ranged from 27 to 48% of the injured hemisphere, and reduction in blood flow was between 16 and 31%. Mortality of up to 25% within 24 hours after stroke onset in experimental stroke models is quite common [51–53] and mortality has been reported to increase up to 33% in the following days due to edema [54, 55]. In this tMCAO study, 13 out of 76 animals (~17%) were lost within the 48 hours post stroke, and 3 more during the first two weeks of treatment, resulting in ~21% mortality within the first 16 days after tMCAO. These results are well within the range of the expected mortality results for rodent stroke models in which moderate to large stroke is induced [51–55]. The level of improvement in mNSS scores in the vehicle group (19%) is lower than the range of 30–50% improvement seen in reported studies in control groups over four to six weeks after tMCAO in rats [56–60]. However, the mean mNSS scores at 24 hours after stroke inductions were between 9 and 11 (several excluded animals with mNSS scores of > 12) [56–60] compared to a mean mNSS of > 14 in the current study, again supporting that the animals in the current study had greater infarct volume and were sicker going into the recovery phase than those in the other studies.

The current study is clearly different from all previously reported studies of other small molecule p38 MAPK inhibitors in various experimental stroke models [30, 33–37] both in terms of the delay in treatment initiation to 48 hours post reperfusion and the six-week prolonged dosing duration and overall treatment study focus. All previously reported studies of p38 MAPK inhibitors that most potently inhibit the p38 α isoform, including SB203580 [30], SB239063 [33, 34], RWJ67657 [36], or VCP979 [37], involved compound administration before or soon after the experimental stroke during the acute phase, with treatment initiations at different time points, ranging from as early as 1 hour pre stroke to as late as 24 hours post stroke. Additionally, all prior p38 α inhibitor studies had a focus on demonstrating neuroprotective effects that resulted in significant reduction of infarct volume (assessed as early as 6 hours post stroke) and subsequent improvements of neurological defects assessed at 24 hours [33, 34], 2 days [30], 7 days [37] or 14 days post stroke [36]. The two most recent p38 α inhibitor studies [36, 37] were also the two longest ones prior to the present study. Interestingly, they did demonstrate the validity of mNSS [50] as a readout for functional recovery attributable to neuroprotection via p38 α inhibition, that was measurable during the subacute phase after

photothrombotic ischemic stroke in mice at one and two weeks, respectively [36, 37]. The results of this neflamapimod study extend these positive mNSS effect observations for a p38 α inhibitor to a rat tMCAO model. Evaluation of mNSS [50] and additional functional tests (stepping test, body swing test, forelimb placement test) to assess the sensory motor deficit after stroke together with BDNF protein biomarker effects were deemed sufficient to evaluate and demonstrate the beneficial *in vivo* efficacy of neflamapimod.

Since the present study represents a first both for initiating the dosing of a potent and selective p38 α inhibitor in the subacute phase after rodent tMCAO-induced stroke (i.e. outside the time window for neuroprotection), as well as for dosing this type of a small molecule for a prolonged time period of six weeks, it is a challenge to find directly supportive subacute or chronic stroke phase studies. It is reassuring that the functional recovery demonstrated in this study is consistent with a report by Umezawa *et al.* [61] who demonstrated that selective inhibition of p38 α by SB239063 improved locomotor recovery after SCI in mice. Although in a different disease context, this study may be most relevant since Umezawa *et al.* [61] also utilized a genetic approach (i.e. a heterozygous knockout of MAPK14) to identify the p38 α specificity of the effect. Generally, small molecule treatments in the rodent cerebral ischemia model start before the 48-hour time point after reperfusion, as already discussed for all the previously reported p38 α inhibitor studies [30, 33–37]. Those studies are not inconsistent with this neflamapimod study, only different in that the neurologic recovery effects therein are clearly associated with a primary acute phase neuroprotective effect of the p38 α inhibitors [30, 33–37]. A review of the experimental stroke literature, however, revealed that the results of all these studies are in conflict with several reports that have associated elevated p38 MAPK activity, including p38 α activity, with a neuroprotective role during and after stroke, either in the acute or subacute phase [62–68]. A major caveat is that several of these differing studies used the small molecule SB203580 at a relatively high concentration and did not address the possibility of blocking p38 MAPK isoforms other than p38 α , such as p38 β , or other kinases (e.g. GAK, RIPK2, NLK, JNK3, CSNK1D and others) with this less potent and much less selective p38 α inhibitor tool compound [69]. As an example, SB203850 through inhibiting casein kinase I δ and ϵ blocks WNT-stimulated β -catenin signaling [70, 71], which might account for compound effects on neurogenesis, therefore, render claims for p38 attributed activities invalid [65]. The current results with a selective p38 α inhibitor help to resolve this conflict and indicated that activation of p38 α in the recovery phase is deleterious.

Oral neflamapimod was dosed at 1.5 and 4.5 mg/kg administered in 1% Pluronic F108 vehicle twice daily on 5.5 days per week for six weeks in the present rat tMCAO study. These two neflamapimod dose levels in this vehicle were selected because their twice daily administration for three weeks was previously demonstrated to be pharmacologically active in aged rats with cognitive deficits attributed to chronic inflammation-induced, IL-1 β -mediated impairment of synaptic plasticity [45]. The results obtained in this tMCAO study corroborate that these two neflamapimod dose levels are pharmacologically active in rats. Unlike the aged rat study, in which the 4.5 mg/kg dose did not lead to better cognitive results compared to the 1.5 mg/kg dose [45], the beneficial effects by neflamapimod resulting in neurologic and behavioral improvements after stroke increased in a dose-related manner and enhanced the slight spontaneous recovery observed in the vehicle control group.

The IL-1 β results of the present rat tMCAO study, though limited to one time point, are to our knowledge the first time that such results have been reported this late out after the acute stroke event in this model. The results showed residual elevation of IL-1 β on the stroked side of the brain, but not on the unaffected side. IL-1 β in the injured brain hemisphere was measurable in 50% of animals in the vehicle- and low-dose neflamapimod-treated group as well as in 30% of the animals in the high-dose neflamapimod-treated group at the end of the study. This

together with the observed beneficial neurologic effects by neflamapimod suggests for the first time that chronic inflammation may play a deleterious role in the recovery process in the rat tMCAO stroke model. This is a finding that supplements previously reported IL-1 β results during the acute phase in this type of model [13, 72, 73]. Although this single time point analysis appears to exclude a neflamapimod effect on IL-1 β production, it does not preclude a neflamapimod effect on IL-1 β signaling (see further discussion below).

Terminal measurements of brain BDNF protein demonstrated that six-week neflamapimod treatment resulted in dose-dependent increases of this neurotrophic factor in both the injured and uninjured hemisphere. BDNF is a key regulator of plasticity both in the healthy and injured brain which has been recognized as a key regulator of rehabilitation- and activity-induced functional and motor recovery, respectively, after stroke [74–76]. BDNF is reported to have a critical role in promoting recovery after stroke as a crucial signaling molecule that mediates adaptive brain plasticity [22, 77–80]. Increased BDNF levels in perilesional areas have been observed with interventions that improve functional recovery post stroke [81–83]. Conversely, attenuation of brain BDNF levels or effects following cerebral ischemia results in reduced neuroplastic changes or decreased recovery of function, both in spontaneous and in rehabilitation-induced recovery scenarios [79, 84, 85]. Since regenerative roles have been attributed to BDNF in preclinical models of stroke [79, 86–89], upregulation of BDNF may be a plausible contributor to the neflamapimod-induced functional recovery observed after the ischemic stroke.

How BDNF elevation is linked to p38 α inhibition by neflamapimod in this experimental stroke model will need to be determined in a follow-up mechanistic study. Our favored hypothesis is that the BDNF levels might be interpreted as a marker of a more general effect on IL-1 β signaling [19] that could result from p38 α inhibition [18, 21]. In such a hypothetical mechanistic model, the observation of inhibition of IL-1 β signaling without impacting IL-1 β production would be consistent with previously observed aged rat results demonstrating that neflamapimod is more potent at inhibiting IL-1 β signaling than at inhibiting IL-1 β production [45]. While this stroke study was not intended to address the exact mechanism of action of neflamapimod, the observation that BDNF protein was increased in both brain hemispheres at study termination is nevertheless supporting the underlying hypothesis that functional recovery was associated with enhancing synaptic plasticity.

Taking into account the design of the study that precludes an effect through neuroprotection, the absence of an anti-inflammatory effect, and the possibility of an effect on IL-1 β signaling, combined with the scientific literature regarding p38 MAPK-mediated deleterious effects of IL-1 β on synaptic plasticity [21, 25], the results herein imply a model in which IL-1 β would limit functional recovery after stroke via p38 α -mediated impairment of neural and synaptic plasticity. The results in this neflamapimod-mediated stroke recovery study are also consistent with studies in other disease contexts in which activation of p38 MAPK, particularly the alpha isoform, is associated with impaired synaptic plasticity [90, 91]. Further, the non-clinical to clinical translational potential with neflamapimod as a p38 α inhibitor is that, similar to the observed slight spontaneous recovery observed in the vehicle control group, neural and synaptic plasticity has been argued to be active in humans and at least partially effective in recovery after stroke [92]. Enhancing a biologic process like neural or synaptic plasticity, that is already active, rather than targeting a process that may not represent an intrinsic recovery pathway, such as neurogenesis, should at least theoretically be more likely to have a clinical effect. For this reason, the opportunity of translational success with p38 α inhibition to promote recovery following stroke by enhancing plasticity would be expected to be higher than for neuroprotection.

Potentially the most important consideration for clinical translation when targeting recovery from stroke is the practical consideration of time window for therapeutic intervention. The

narrow time window after onset of ischemia that is required of neuroprotective approaches has posed an insurmountable challenge for clinical development [93]. Stroke patients do not often present within the required first few hours and, even within the initial 24 hours post stroke, the clinical presentation does not allow one to assess the true size and severity of the stroke. Therefore, any clinical study that starts treatment of patients within the first day of the stroke has a highly variable and heterogeneous patient population and thus requires a large sample size to demonstrate clinical effects. These restrictions preclude the ability to demonstrate clinical proof-of-concept in phase 2 clinical testing. In contrast, a time window of 24 to 48 hours after stroke allows patients' clinical course to have stabilized and allows time for a clinical exam and a diffusion-MRI scan to precisely determine the location and extent of the stroke before starting treatment. In addition, a reasonable prognostic indication of the extent of recovery that a patient will attain with or without intervention can be assessed. As a result, starting treatment in a clinical study at 48 hours or later after stroke allows for the inclusion of more homogenous sub-populations of patients and increases statistical power with fewer subjects; therefore, definitive clinical-proof-of-concept could potentially be demonstrated within a phase 2 clinical study.

As immunosuppression with T lymphocyte depletion in the systemic circulation is an intrinsic component of the clinical syndrome during the subacute phase after ischemic stroke [94], a potential concern of targeting IL-1 β and the innate immune system is an increased risk of infection due to further suppressing the immune system systemically. In the case of targeting IL-1 β with selective inhibition of p38 α , this particular concern is low because in transgenic models p38 α is dispensable for T-cell development and activation [95, 96]. Indeed, for T cell development the p38 MAPK isoforms primarily involved in T lymphocyte development are the minor isoforms, p38 γ and p38 δ [97]. Consistent with this notion, in a 3503-patient twelve-week treatment phase 3 trial of the p38 α/β inhibitor losapimod in patients with acute myocardial infection the incidence of infection was 2.7% and 2.4%, and of opportunistic infection 0.3% and 0.4%, in losapimod and placebo treatment groups, respectively [98]. The risk of systemic immunosuppression is expected to be further attenuated with neflamapimod as it preferentially distributes to the CNS, with plasma drug levels in peripheral blood obtained after dosing in humans that have CNS activity being three- to five-fold lower than those required to for pharmacologic activity [42]. Further, neflamapimod has been administered to more than 300 patients and volunteers at doses up to ten-fold-higher than anticipated doses in the stroke setting, and up to six months, and to date there has been no evidence of an increase in infections (EIP Pharma Inc., data on file).

After our studies were completed, additional preclinical and clinical study results have been reported on the potential of therapeutically targeting IL-1 β [99]. In particular, a single-center 80-patient phase 2 placebo-controlled clinical study has been reported on the effects of subcutaneous interleukin-1 receptor antagonist (IL-1Ra anakinra) which blocks both IL-1 α and IL-1 β signaling [100]. IL-1Ra was administered to patients starting within 5 hours of an acute stroke, every 12 hours for six doses. The study met its primary endpoint by reducing serum concentrations of interleukin-6, as well of c-reactive protein levels (i.e. reduced inflammatory markers). However, there was a non-significant trend towards worse functional outcomes at 90 days, as measured by the modified Rankin scale. The authors attribute the potential deleterious effect on functional recovery on a negative interaction with alteplase, as in a previous phase 2 study of IL-1Ra conducted prior to widespread use of thrombolysis they had seen a positive trend on outcomes at 90 days. An additional consideration is that IL-1Ra inhibits all signaling, whether beneficial or deleterious, of both IL-1 α and IL-1 β ; while a later report demonstrated that delayed administration of IL-1 α to mice after stroke induced by filament-based MCAO ameliorated functional deficit and promoted neural repair [101]. In contrast, as

discussed in the introduction, inhibition of p38 α would be expected to only block the deleterious effects of IL-1 on synaptic plasticity that are mediated by the ubiquitous form of IL1-RAP, while preserving signaling through the neuron-specific form of IL1-RAP that enhances synaptic plasticity [23–25].

There are some limitations to this research stroke study, since only one treatment duration was chosen, and functional outcomes were measured at the Week 4 and Week 6 timepoints after the stroke. It cannot be predicted whether longer neflamapimod treatment duration with additional analyses would have led to further mNSS improvement and continued recovery during the chronic phase, and the exact timing of the onset of neflamapimod action on functional recovery was not determined. Neflamapimod treatment was initiated at 48 hours after reperfusion, at a time point that was considered subacute post stroke in light of the prior negative experiences with other p38 α inhibitors and numerous neuroprotective agents when they were administered at 24 hours or more post stroke, which implies an exceedingly low likelihood that neuroprotection plays a role in promoting functional recovery, but without accompanying histologic analyses an effect of neflamapimod on neuronal loss cannot be formally excluded. Further, while we believe that it is highly probable that the functional recovery mediated by neflamapimod was due to enhancement of plasticity mechanisms, in the absence of technologies that can measure synaptic function *in vivo* in real-time, there is no means to directly verify that assumption. Similarly, while the known activity of IL-1 β to impair synaptic plasticity via p38 α implies that inhibition IL-1 β activity is a major contributor to the neflamapimod clinical activity, there is no means to directly confirm that link *in vivo*, or to exclude other potential mechanisms. Finally, in the current study, young (3-months old) male animals were utilized. Although neural and synaptic plasticity recovery functions appear to be active in aged animals and are also at least partially preserved in elderly patients [102], in order to improve clinical translation, a replication and extension of this study to include females, aged animal and animals with co-morbidities may be required since gender, age and reduced health condition (e.g. illnesses, diseases, disorders, health problems) may affect development of ischemic damage and resulting behavioral deficits in patients [103].

Conclusions

Here prolonged, six-week oral administration of the p38 α inhibitor neflamapimod, with treatment initiation starting at 48 hours post reperfusion that is outside the previously characterized neuroprotection window for p38 α inhibitors, resulted in dose-related significant neurologic recovery and improvement of motor and sensory functions measured at four and six weeks post stroke. Additionally, dose-related increases of the neurogenic factor BDNF in the brain as a potential biomarker for neflamapimod effects on synaptic plasticity were observed at termination of the study on Day 44. Thus, neflamapimod use offers the possibility of being effective when given at a later time after a completed stroke by promoting functional recovery.

Supporting information

S1 Table. Modified Neurological Severity Score (mNSS) tests and scoring values.
(PDF)

S2 Table. Body weight monitoring of the three treatment groups, including vehicle control, 1.5 mg/kg neflamapimod (NFMD), and 4.5 mg/kg NFMD throughout the study.
(PDF)

S3 Table. IL-1 β (pg/ml) levels in the injured right brain hemisphere on Day 44 post stroke.
(PDF)

S1 Appendix. Statistical analysis details.

(PDF)

S2 Appendix. BDNF statistical analysis details.

(PDF)

Acknowledgments

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Author Contributions**Conceptualization:** John J. Alam.**Data curation:** Michael Krakovsky.**Formal analysis:** John J. Alam, Michael Krakovsky, Ursula Germann.**Funding acquisition:** John J. Alam.**Investigation:** Michael Krakovsky.**Methodology:** John J. Alam, Michael Krakovsky, Aharon Levy.**Project administration:** John J. Alam, Michael Krakovsky.**Resources:** Michael Krakovsky.**Software:** Michael Krakovsky.**Supervision:** John J. Alam.**Validation:** Michael Krakovsky.**Visualization:** John J. Alam, Ursula Germann.**Writing – original draft:** John J. Alam, Ursula Germann.**Writing – review & editing:** John J. Alam, Michael Krakovsky, Ursula Germann, Aharon Levy.**References**

1. Disease GBD, Injury I, Prevalence C. Global, regional, and national incidence, prevalence, and years lived with disability for 354 diseases and injuries for 195 countries and territories, 1990–2017: a systematic analysis for the Global Burden of Disease Study 2017. *Lancet*. 2018; 392(10159):1789–858. Epub 2018/11/30. [https://doi.org/10.1016/S0140-6736\(18\)32279-7](https://doi.org/10.1016/S0140-6736(18)32279-7) PMID: 30496104; PubMed Central PMCID: PMC6227754.
2. Rajsic S, Gothe H, Borba HH, Sroczynski G, Vujcic J, Toell T, et al. Economic burden of stroke: a systematic review on post-stroke care. *Eur J Health Econ*. 2019; 20(1):107–34. Epub 2018/06/18. <https://doi.org/10.1007/s10198-018-0984-0> PMID: 29909569.
3. Lipton P. Ischemic cell death in brain neurons. *Physiol Rev*. 1999; 79(4):1431–568. Epub 1999/10/03. <https://doi.org/10.1152/physrev.1999.79.4.1431> PMID: 10508238.
4. Zhao LR, Willing A. Enhancing endogenous capacity to repair a stroke-damaged brain: An evolving field for stroke research. *Prog Neurobiol*. 2018; 163–164:5–26. Epub 2018/02/25. <https://doi.org/10.1016/j.pneurobio.2018.01.004> PMID: 29476785; PubMed Central PMCID: PMC6075953.

5. Cramer SC. Treatments to Promote Neural Repair after Stroke. *J Stroke*. 2018; 20(1):57–70. Epub 2018/02/07. <https://doi.org/10.5853/jos.2017.02796> PMID: 29402069; PubMed Central PMCID: PMC5836581.
6. Cassidy JM, Cramer SC. Spontaneous and Therapeutic-Induced Mechanisms of Functional Recovery After Stroke. *Transl Stroke Res*. 2017; 8(1):33–46. Epub 2016/04/26. <https://doi.org/10.1007/s12975-016-0467-5> PMID: 27109642; PubMed Central PMCID: PMC5079852.
7. Savitz SI. Cell therapies: careful translation from animals to patients. *Stroke*. 2013; 44(6 Suppl 1): S107–9. Epub 2013/06/05. PubMed Central PMCID: PMC4218778. <https://doi.org/10.1161/STROKEAHA.112.679605> PMID: 23709699
8. Hacke W, Lichy C. Thrombolysis for acute stroke under antiplatelet therapy: safe enough to be beneficial? *Nat Clin Pract Neurol*. 2008; 4(9):474–5. Epub 2008/07/31. <https://doi.org/10.1038/ncpneuro0867> PMID: 18665145.
9. Grupke S, Hall J, Dobbs M, Bix GJ, Fraser JF. Understanding history, and not repeating it. Neuroprotection for acute ischemic stroke: from review to preview. *Clin Neurol Neurosurg*. 2015; 129:1–9. Epub 2014/12/17. <https://doi.org/10.1016/j.clineuro.2014.11.013> PMID: 25497127.
10. Karsy M, Brock A, Guan J, Tausky P, Kalani MY, Park MS. Neuroprotective strategies and the underlying molecular basis of cerebrovascular stroke. *Neurosurg Focus*. 2017; 42(4):E3. Epub 2017/04/04. <https://doi.org/10.3171/2017.1.FOCUS16522> PMID: 28366066.
11. Xiong XY, Liu L, Yang QW. Refocusing Neuroprotection in Cerebral Reperfusion Era: New Challenges and Strategies. *Front Neurol*. 2018; 9:249. Epub 2018/05/10. <https://doi.org/10.3389/fneur.2018.00249> PMID: 29740385; PubMed Central PMCID: PMC5926527.
12. Venkat P, Shen Y, Chopp M, Chen J. Cell-based and pharmacological neurorestorative therapies for ischemic stroke. *Neuropharmacology*. 2018; 134(Pt B):310–22. Epub 2017/09/05. <https://doi.org/10.1016/j.neuropharm.2017.08.036> PMID: 28867364; PubMed Central PMCID: PMC5832535.
13. Lambertsen KL, Biber K, Finsen B. Inflammatory cytokines in experimental and human stroke. *J Cereb Blood Flow Metab*. 2012; 32(9):1677–98. Epub 2012/06/29. <https://doi.org/10.1038/jcbfm.2012.88> PMID: 22739623; PubMed Central PMCID: PMC3434626.
14. Hewett SJ, Jackman NA, Claycomb RJ. Interleukin-1beta in Central Nervous System Injury and Repair. *Eur J Neurodegener Dis*. 2012; 1(2):195–211. Epub 2012/08/01. PubMed PMID: 26082912; PubMed Central PMCID: PMC4465544.
15. Sobowale OA, Parry-Jones AR, Smith CJ, Tyrrell PJ, Rothwell NJ, Allan SM. Interleukin-1 in Stroke: From Bench to Bedside. *Stroke*. 2016; 47(8):2160–7. Epub 2016/03/05. <https://doi.org/10.1161/STROKEAHA.115.010001> PMID: 26931154.
16. Lambertsen KL, Finsen B, Clausen BH. Post-stroke inflammation—target or tool for therapy? *Acta Neuropathol*. 2019; 137(5):693–714. Epub 2018/11/30. <https://doi.org/10.1007/s00401-018-1930-z> PMID: 30483945; PubMed Central PMCID: PMC6482288.
17. Pozzi D, Menna E, Canzi A, Desiato G, Mantovani C, Matteoli M. The Communication Between the Immune and Nervous Systems: The Role of IL-1beta in Synaptopathies. *Front Mol Neurosci*. 2018; 11:111. Epub 2018/04/21. <https://doi.org/10.3389/fnmol.2018.00111> PMID: 29674955; PubMed Central PMCID: PMC5895746.
18. Song C, Zhang Y, Dong Y. Acute and subacute IL-1beta administrations differentially modulate neuroimmune and neurotrophic systems: possible implications for neuroprotection and neurodegeneration. *J Neuroinflammation*. 2013; 10:59. Epub 2013/05/09. <https://doi.org/10.1186/1742-2094-10-59> PMID: 23651534; PubMed Central PMCID: PMC3656796.
19. Patterson SL. Immune dysregulation and cognitive vulnerability in the aging brain: Interactions of microglia, IL-1beta, BDNF and synaptic plasticity. *Neuropharmacology*. 2015; 96(Pt A):11–8. Epub 2015/01/01. <https://doi.org/10.1016/j.neuropharm.2014.12.020> PMID: 25549562; PubMed Central PMCID: PMC4475415.
20. Tanaka N, Cortese GP, Barrientos RM, Maier SF, Patterson SL. Aging and an Immune Challenge Interact to Produce Prolonged, but Not Permanent, Reductions in Hippocampal L-LTP and mBDNF in a Rodent Model with Features of Delirium. *eNeuro*. 2018; 5(3). Epub 2018/06/19. <https://doi.org/10.1523/ENEURO.0009-18.2018> PMID: 29911174; PubMed Central PMCID: PMC6001264.
21. Tong L, Prieto GA, Kramar EA, Smith ED, Cribbs DH, Lynch G, et al. Brain-derived neurotrophic factor-dependent synaptic plasticity is suppressed by interleukin-1beta via p38 mitogen-activated protein kinase. *J Neurosci*. 2012; 32(49):17714–24. Epub 2012/12/12. <https://doi.org/10.1523/JNEUROSCI.1253-12.2012> PMID: 23223292; PubMed Central PMCID: PMC3687587.
22. Berretta A, Tzeng YC, Clarkson AN. Post-stroke recovery: the role of activity-dependent release of brain-derived neurotrophic factor. *Expert Rev Neurother*. 2014; 14(11):1335–44. Epub 2014/10/17. <https://doi.org/10.1586/14737175.2014.969242> PMID: 25319267.

23. Smith DE, Lipsky BP, Russell C, Ketchum RR, Kirchner J, Hensley K, et al. A central nervous system-restricted isoform of the interleukin-1 receptor accessory protein modulates neuronal responses to interleukin-1. *Immunity*. 2009; 30(6):817–31. Epub 2009/06/02. <https://doi.org/10.1016/j.immuni.2009.03.020> PMID: 19481478; PubMed Central PMCID: PMC4103746.
24. Huang Y, Smith DE, Ibanez-Sandoval O, Sims JE, Friedman WJ. Neuron-specific effects of interleukin-1beta are mediated by a novel isoform of the IL-1 receptor accessory protein. *J Neurosci*. 2011; 31(49):18048–59. Epub 2011/12/14. <https://doi.org/10.1523/JNEUROSCI.4067-11.2011> PMID: 22159118; PubMed Central PMCID: PMC3261076.
25. Prieto GA, Snigdha S, Baglietto-Vargas D, Smith ED, Berchtold NC, Tong L, et al. Synapse-specific IL-1 receptor subunit reconfiguration augments vulnerability to IL-1beta in the aged hippocampus. *Proc Natl Acad Sci U S A*. 2015; 112(36):E5078–87. Epub 2015/08/26. <https://doi.org/10.1073/pnas.1514486112> PMID: 26305968; PubMed Central PMCID: PMC4568670.
26. Ferrer I, Friguls B, Dalfo E, Planas AM. Early modifications in the expression of mitogen-activated protein kinase (MAPK/ERK), stress-activated kinases SAPK/JNK and p38, and their phosphorylated substrates following focal cerebral ischemia. *Acta Neuropathol*. 2003; 105(5):425–37. Epub 2003/04/05. <https://doi.org/10.1007/s00401-002-0661-2> PMID: 12677442.
27. Krupinski J, Slevin M, Marti E, Catena E, Rubio F, Gaffney J. Time-course phosphorylation of the mitogen activated protein (MAP) kinase group of signalling proteins and related molecules following middle cerebral artery occlusion (MCAO) in rats. *Neuropathol Appl Neurobiol*. 2003; 29(2):144–58. Epub 2003/03/29. <https://doi.org/10.1046/j.1365-2990.2003.00454.x> PMID: 12662322.
28. Nozaki K, Nishimura M, Hashimoto N. Mitogen-activated protein kinases and cerebral ischemia. *Mol Neurobiol*. 2001; 23(1):1–19. Epub 2001/10/20. <https://doi.org/10.1385/MN:23:1:01> PMID: 11642541.
29. Piao CS, Che Y, Han PL, Lee JK. Delayed and differential induction of p38 MAPK isoforms in microglia and astrocytes in the brain after transient global ischemia. *Brain Res Mol Brain Res*. 2002; 107(2):137–44. Epub 2002/11/12. [https://doi.org/10.1016/s0169-328x\(02\)00456-4](https://doi.org/10.1016/s0169-328x(02)00456-4) PMID: 12425942.
30. Piao CS, Kim JB, Han PL, Lee JK. Administration of the p38 MAPK inhibitor SB203580 affords brain protection with a wide therapeutic window against focal ischemic insult. *J Neurosci Res*. 2003; 73(4):537–44. Epub 2003/08/05. <https://doi.org/10.1002/jnr.10671> PMID: 12898538.
31. Mao G, Ren P, Wang G, Yan F, Zhang Y. MicroRNA-128-3p Protects Mouse Against Cerebral Ischemia Through Reducing p38alpha Mitogen-Activated Protein Kinase Activity. *J Mol Neurosci*. 2017; 61(2):152–8. Epub 2016/12/03. <https://doi.org/10.1007/s12031-016-0871-z> PMID: 27905005.
32. Han D, Scott EL, Dong Y, Raz L, Wang R, Zhang Q. Attenuation of mitochondrial and nuclear p38alpha signaling: a novel mechanism of estrogen neuroprotection in cerebral ischemia. *Mol Cell Endocrinol*. 2015; 400:21–31. Epub 2014/12/03. <https://doi.org/10.1016/j.mce.2014.11.010> PMID: 25462588.
33. Barone FC, Irving EA, Ray AM, Lee JC, Kassis S, Kumar S, et al. SB 239063, a second-generation p38 mitogen-activated protein kinase inhibitor, reduces brain injury and neurological deficits in cerebral focal ischemia. *J Pharmacol Exp Ther*. 2001; 296(2):312–21. Epub 2001/02/13. PubMed PMID: 11160612.
34. Legos JJ, Erhardt JA, White RF, Lenhard SC, Chandra S, Parsons AA, et al. SB 239063, a novel p38 inhibitor, attenuates early neuronal injury following ischemia. *Brain Res*. 2001; 892(1):70–7. Epub 2001/02/15. [https://doi.org/10.1016/s0006-8993\(00\)03228-5](https://doi.org/10.1016/s0006-8993(00)03228-5) PMID: 11172750.
35. Legos JJ, McLaughlin B, Skaper SD, Strijbos PJ, Parsons AA, Aizenman E, et al. The selective p38 inhibitor SB-239063 protects primary neurons from mild to moderate excitotoxic injury. *Eur J Pharmacol*. 2002; 447(1):37–42. Epub 2002/07/11. [https://doi.org/10.1016/s0014-2999\(02\)01890-3](https://doi.org/10.1016/s0014-2999(02)01890-3) PMID: 12106800.
36. Chang D, Wang YC, Bai YY, Lu CQ, Xu TT, Zhu L, et al. Role of P38 MAPK on MMP Activity in Photothrombotic Stroke Mice as Measured using an Ultrafast MMP Activatable Probe. *Sci Rep*. 2015; 5:16951. Epub 2015/11/20. <https://doi.org/10.1038/srep16951> PMID: 26581247; PubMed Central PMCID: PMC4652271.
37. Cai Y, Lu C, Xu T, Ma Y, Min S, Scammells P, et al. Diffusion Tensor Imaging Evaluation of Axonal/White Matter Remodeling in a Mouse Model of Diabetic Stroke Treated with Novel p38 MAPK Inhibitor, VCP979. *J Biomed Nanotechnol*. 2018; 14(3):585–93. Epub 2018/04/18. <https://doi.org/10.1166/jbn.2018.2522> PMID: 29663930.
38. Roy Choudhury G, Ryou MG, Poteet E, Wen Y, He R, Sun F, et al. Involvement of p38 MAPK in reactive astrogliosis induced by ischemic stroke. *Brain Res*. 2014; 1551:45–58. Epub 2014/01/21. <https://doi.org/10.1016/j.brainres.2014.01.013> PMID: 24440774; PubMed Central PMCID: PMC3987968.
39. Sommer CJ. Ischemic stroke: experimental models and reality. *Acta Neuropathol*. 2017; 133(2):245–61. Epub 2017/01/09. <https://doi.org/10.1007/s00401-017-1667-0> PMID: 28064357; PubMed Central PMCID: PMC5250659.

40. Li H, Zhang N, Lin HY, Yu Y, Cai QY, Ma L, et al. Histological, cellular and behavioral assessments of stroke outcomes after photothrombosis-induced ischemia in adult mice. *BMC Neurosci*. 2014; 15:58. Epub 2014/06/03. <https://doi.org/10.1186/1471-2202-15-58> PMID: 24886391; PubMed Central PMCID: PMC4039545.
41. Duffy JP, Harrington EM, Salituro FG, Cochran JE, Green J, Gao H, et al. The Discovery of VX-745: A Novel and Selective p38alpha Kinase Inhibitor. *ACS Med Chem Lett*. 2011; 2(10):758–63. Epub 2011/10/13. <https://doi.org/10.1021/ml2001455> PMID: 24900264; PubMed Central PMCID: PMC4018046.
42. Alam J, Blackburn K, Patrick D. Neflamapimod: Clinical Phase 2b-Ready Oral Small Molecule Inhibitor of p38alpha to Reverse Synaptic Dysfunction in Early Alzheimer's Disease. *J Prev Alzheimers Dis*. 2017; 4(4):273–8. Epub 2017/11/29. <https://doi.org/10.14283/jpad.2017.41> PMID: 29181493.
43. Scheltens P, Prins N, Lammertsma A, Yaqub M, Gouw A, Wink AM, et al. An exploratory clinical study of p38alpha kinase inhibition in Alzheimer's disease. *Ann Clin Transl Neurol*. 2018; 5(4):464–73. Epub 2018/04/25. <https://doi.org/10.1002/acn3.549> PMID: 29687023; PubMed Central PMCID: PMC5899915.
44. Scheltens P, Alam J, Harrison J, Blackburn K, Prins N. Efficacy and safety results of REVERSE-SD, phase-2b clinical study of the selective p38α kinase inhibitor neflamapimod in early-stage Alzheimer's disease (AD). 12th Clinical Trials in Alzheimer's Disease meeting, Dec 4–7 2019, San Diego, Abstract #OC6. *J Prev Alzheimers Dis*. 2019; 6 (Suppl 1):S9–S10.
45. Alam JJ. Selective Brain-Targeted Antagonism of p38 MAPKalpha Reduces Hippocampal IL-1beta Levels and Improves Morris Water Maze Performance in Aged Rats. *J Alzheimers Dis*. 2015; 48(1):219–27. Epub 2015/09/25. <https://doi.org/10.3233/JAD-150277> PMID: 26401942; PubMed Central PMCID: PMC4923728.
46. Lynch MA. Age-related neuroinflammatory changes negatively impact on neuronal function. *Front Aging Neurosci*. 2010; 1:6. Epub 2010/06/17. <https://doi.org/10.3389/neuro.24.006.2009> PMID: 20552057; PubMed Central PMCID: PMC2874409.
47. Longa EZ, Weinstein PR, Carlson S, Cummins R. Reversible middle cerebral artery occlusion without craniectomy in rats. *Stroke*. 1989; 20(1):84–91. Epub 1989/01/01. <https://doi.org/10.1161/01.str.20.1.84> PMID: 2643202.
48. Schmid-Elsaesser R, Zausinger S, Hungerhuber E, Baethmann A, Reulen HJ. A critical reevaluation of the intraluminal thread model of focal cerebral ischemia: evidence of inadvertent premature reperfusion and subarachnoid hemorrhage in rats by laser-Doppler flowmetry. *Stroke*. 1998; 29(10):2162–70. Epub 1998/10/02. <https://doi.org/10.1161/01.str.29.10.2162> PMID: 9756599.
49. Chen J, Li Y, Wang L, Zhang Z, Lu D, Lu M, et al. Therapeutic benefit of intravenous administration of bone marrow stromal cells after cerebral ischemia in rats. *Stroke*. 2001; 32(4):1005–11. Epub 2001/04/03. <https://doi.org/10.1161/01.str.32.4.1005> PMID: 11283404.
50. Schaar KL, Brenneman MM, Savitz SI. Functional assessments in the rodent stroke model. *Exp Transl Stroke Med*. 2010; 2(1):13. Epub 2010/07/21. <https://doi.org/10.1186/2040-7378-2-13> PMID: 20642841; PubMed Central PMCID: PMC2915950.
51. Kuge Y, Minematsu K, Yamaguchi T, Miyake Y. Nylon monofilament for intraluminal middle cerebral artery occlusion in rats. *Stroke*. 1995; 26(9):1655–7; discussion 8. Epub 1995/09/01. <https://doi.org/10.1161/01.str.26.9.1655> PMID: 7660413.
52. Neumann-Haefelin T, Kastrup A, de Crespigny A, Yenari MA, Ringer T, Sun GH, et al. Serial MRI after transient focal cerebral ischemia in rats: dynamics of tissue injury, blood-brain barrier damage, and edema formation. *Stroke*. 2000; 31(8):1965–72; discussion 72–3. Epub 2000/08/06. <https://doi.org/10.1161/01.str.31.8.1965> PMID: 10926965.
53. Henninger N, Sicard KM, Schmidt KF, Bardutzky J, Fisher M. Comparison of ischemic lesion evolution in embolic versus mechanical middle cerebral artery occlusion in Sprague Dawley rats using diffusion and perfusion imaging. *Stroke*. 2006; 37(5):1283–7. Epub 2006/03/25. <https://doi.org/10.1161/01.STR.0000217223.72193.98> PMID: 16556883.
54. Aspey BS, Cohen S, Patel Y, Terruli M, Harrison MJ. Middle cerebral artery occlusion in the rat: consistent protocol for a model of stroke. *Neuropathol Appl Neurobiol*. 1998; 24(6):487–97. Epub 1999/01/15. <https://doi.org/10.1046/j.1365-2990.1998.00146.x> PMID: 9888159.
55. Modo M, Stroemer RP, Tang E, Veizovic T, Sowniski P, Hodges H. Neurological sequelae and long-term behavioural assessment of rats with transient middle cerebral artery occlusion. *J Neurosci Methods*. 2000; 104(1):99–109. Epub 2001/02/13. [https://doi.org/10.1016/s0165-0270\(00\)00329-0](https://doi.org/10.1016/s0165-0270(00)00329-0) PMID: 11163416.
56. Chen J, Sanberg PR, Li Y, Wang L, Lu M, Willing AE, et al. Intravenous administration of human umbilical cord blood reduces behavioral deficits after stroke in rats. *Stroke*. 2001; 32(11):2682–8. Epub 2001/11/03. <https://doi.org/10.1161/hs1101.098367> PMID: 11692034.

57. Shehadah A, Chen J, Kramer B, Zacharek A, Cui Y, Roberts C, et al. Efficacy of single and multiple injections of human umbilical tissue-derived cells following experimental stroke in rats. *PLoS One*. 2013; 8(1):e54083. Epub 2013/01/24. <https://doi.org/10.1371/journal.pone.0054083> PMID: [23342081](https://pubmed.ncbi.nlm.nih.gov/23342081/); PubMed Central PMCID: PMC3544758.
58. Lee SH, Jin KS, Bang OY, Kim BJ, Park SJ, Lee NH, et al. Differential Migration of Mesenchymal Stem Cells to Ischemic Regions after Middle Cerebral Artery Occlusion in Rats. *PLoS One*. 2015; 10(8):e0134920. Epub 2015/08/05. <https://doi.org/10.1371/journal.pone.0134920> PMID: [26241653](https://pubmed.ncbi.nlm.nih.gov/26241653/); PubMed Central PMCID: PMC4524688.
59. Cho DY, Jeun SS. Combination therapy of human bone marrow-derived mesenchymal stem cells and minocycline improves neuronal function in a rat middle cerebral artery occlusion model. *Stem Cell Res Ther*. 2018; 9(1):309. Epub 2018/11/11. <https://doi.org/10.1186/s13287-018-1011-1> PMID: [30413178](https://pubmed.ncbi.nlm.nih.gov/30413178/); PubMed Central PMCID: PMC6230290.
60. Zhang HL, Xie XF, Xiong YQ, Liu SM, Hu GZ, Cao WF, et al. Comparisons of the therapeutic effects of three different routes of bone marrow mesenchymal stem cell transplantation in cerebral ischemic rats. *Brain Res*. 2018; 1680:143–54. Epub 2017/12/25. <https://doi.org/10.1016/j.brainres.2017.12.017> PMID: [29274877](https://pubmed.ncbi.nlm.nih.gov/29274877/).
61. Umezawa H, Naito Y, Tanaka K, Yoshioka K, Suzuki K, Sudo T, et al. Genetic and Pharmacological Inhibition of p38alpha Improves Locomotor Recovery after Spinal Cord Injury. *Front Pharmacol*. 2017; 8:72. Epub 2017/03/07. <https://doi.org/10.3389/fphar.2017.00072> PMID: [28261102](https://pubmed.ncbi.nlm.nih.gov/28261102/); PubMed Central PMCID: PMC5313485.
62. Lennmyr F, Ericsson A, Gerwins P, Ahlstrom H, Terent A. Increased brain injury and vascular leakage after pretreatment with p38-inhibitor SB203580 in transient ischemia. *Acta Neurol Scand*. 2003; 108(5):339–45. Epub 2003/11/18. <https://doi.org/10.1034/j.1600-0404.2003.00129.x> PMID: [14616304](https://pubmed.ncbi.nlm.nih.gov/14616304/).
63. Pfeilschifter W, Czech B, Hoffmann BP, Sujak M, Kahles T, Steinmetz H, et al. Pyrrolidine dithiocarbamate activates p38 MAPK and protects brain endothelial cells from apoptosis: a mechanism for the protective effect in stroke? *Neurochem Res*. 2010; 35(9):1391–401. Epub 2010/06/02. <https://doi.org/10.1007/s11064-010-0197-0> PMID: [20514517](https://pubmed.ncbi.nlm.nih.gov/20514517/).
64. Cheng CY, Lin JG, Tang NY, Kao ST, Hsieh CL. Electroacupuncture at different frequencies (5Hz and 25Hz) ameliorates cerebral ischemia-reperfusion injury in rats: possible involvement of p38 MAPK-mediated anti-apoptotic signaling pathways. *BMC Complement Altern Med*. 2015; 15:241. Epub 2015/07/19. <https://doi.org/10.1186/s12906-015-0752-y> PMID: [26187498](https://pubmed.ncbi.nlm.nih.gov/26187498/); PubMed Central PMCID: PMC4506591.
65. Lin Y, Zhang JC, Yao CY, Wu Y, Abdelgawad AF, Yao SL, et al. Critical role of astrocytic interleukin-17 A in post-stroke survival and neuronal differentiation of neural precursor cells in adult mice. *Cell Death Dis*. 2016; 7(6):e2273. Epub 2016/06/24. <https://doi.org/10.1038/cddis.2015.284> PMID: [27336717](https://pubmed.ncbi.nlm.nih.gov/27336717/); PubMed Central PMCID: PMC5143370.
66. Cheng CY, Tang NY, Kao ST, Hsieh CL. Ferulic Acid Administered at Various Time Points Protects against Cerebral Infarction by Activating p38 MAPK/p90RSK/CREB/Bcl-2 Anti-Apoptotic Signaling in the Subacute Phase of Cerebral Ischemia-Reperfusion Injury in Rats. *PLoS One*. 2016; 11(5):e0155748. Epub 2016/05/18. <https://doi.org/10.1371/journal.pone.0155748> PMID: [27187745](https://pubmed.ncbi.nlm.nih.gov/27187745/); PubMed Central PMCID: PMC4871485.
67. Cheng CY, Ho TY, Hsiang CY, Tang NY, Hsieh CL, Kao ST, et al. Angelica sinensis Exerts Angiogenic and Anti-apoptotic Effects Against Cerebral Ischemia-Reperfusion Injury by Activating p38MAPK/HIF-1 α /VEGF-A Signaling in Rats. *Am J Chin Med*. 2017; 45(8):1683–708. Epub 2017/11/11. <https://doi.org/10.1142/S0192415X17500914> PMID: [29121798](https://pubmed.ncbi.nlm.nih.gov/29121798/).
68. Cheng CY, Kao ST, Lee YC. Angelica sinensis extract protects against ischemia-reperfusion injury in the hippocampus by activating p38 MAPK-mediated p90RSK/p-Bad and p90RSK/CREB/BDNF signaling after transient global cerebral ischemia in rats. *J Ethnopharmacol*. 2020; 252:112612. Epub 2020/01/29. <https://doi.org/10.1016/j.jep.2020.112612> PMID: [31988015](https://pubmed.ncbi.nlm.nih.gov/31988015/).
69. Davis MI, Hunt JP, Herrgard S, Ciceri P, Wodicka LM, Pallares G, et al. Comprehensive analysis of kinase inhibitor selectivity. *Nat Biotechnol*. 2011; 29(11):1046–51. Epub 2011/11/01. <https://doi.org/10.1038/nbt.1990> PMID: [22037378](https://pubmed.ncbi.nlm.nih.gov/22037378/).
70. Shanware NP, Williams LM, Bowler MJ, Tibbetts RS. Non-specific in vivo inhibition of CK1 by the pyridinyl imidazole p38 inhibitors SB 203580 and SB 202190. *BMB Rep*. 2009; 42(3):142–7. Epub 2009/04/02. <https://doi.org/10.5483/bmbrep.2009.42.3.142> PMID: [19336000](https://pubmed.ncbi.nlm.nih.gov/19336000/); PubMed Central PMCID: PMC4412876.
71. Verkaar F, van der Doelen AA, Smits JF, Blankesteyn WM, Zaman GJ. Inhibition of Wnt/beta-catenin signaling by p38 MAP kinase inhibitors is explained by cross-reactivity with casein kinase I δ /varepsilon. *Chem Biol*. 2011; 18(4):485–94. Epub 2011/04/26. <https://doi.org/10.1016/j.chembiol.2011.01.015> PMID: [21513885](https://pubmed.ncbi.nlm.nih.gov/21513885/).

72. Wang X, Yue TL, Barone FC, White RF, Gagnon RC, Feuerstein GZ. Concomitant cortical expression of TNF-alpha and IL-1 beta mRNAs follows early response gene expression in transient focal ischemia. *Mol Chem Neuropathol*. 1994; 23(2–3):103–14. Epub 1994/10/01. <https://doi.org/10.1007/BF02815404> PMID: 7702701.
73. Berti R, Williams AJ, Moffett JR, Hale SL, Velarde LC, Elliott PJ, et al. Quantitative real-time RT-PCR analysis of inflammatory gene expression associated with ischemia-reperfusion brain injury. *J Cereb Blood Flow Metab*. 2002; 22(9):1068–79. Epub 2002/09/10. <https://doi.org/10.1097/00004647-200209000-00004> PMID: 12218412.
74. Mang CS, Campbell KL, Ross CJ, Boyd LA. Promoting neuroplasticity for motor rehabilitation after stroke: considering the effects of aerobic exercise and genetic variation on brain-derived neurotrophic factor. *Phys Ther*. 2013; 93(12):1707–16. Epub 2013/08/03. <https://doi.org/10.2522/ptj.20130053> PMID: 23907078; PubMed Central PMCID: PMC3870490.
75. Crozier J, Roig M, Eng JJ, MacKay-Lyons M, Fung J, Ploughman M, et al. High-Intensity Interval Training After Stroke: An Opportunity to Promote Functional Recovery, Cardiovascular Health, and Neuroplasticity. *Neurorehabil Neural Repair*. 2018; 32(6–7):543–56. Epub 2018/04/21. <https://doi.org/10.1177/1545968318766663> PMID: 29676956.
76. King M, Kelly LP, Wallack EM, Hasan SMM, Kirkland MC, Curtis ME, et al. Serum levels of insulin-like growth factor-1 and brain-derived neurotrophic factor as potential recovery biomarkers in stroke. *Neurol Res*. 2019; 41(4):354–63. Epub 2019/01/09. <https://doi.org/10.1080/01616412.2018.1564451> PMID: 30620251.
77. Lipsky RH, Marini AM. Brain-derived neurotrophic factor in neuronal survival and behavior-related plasticity. *Ann N Y Acad Sci*. 2007; 1122:130–43. Epub 2007/12/14. <https://doi.org/10.1196/annals.1403.009> PMID: 18077569.
78. Mattson MP. Glutamate and neurotrophic factors in neuronal plasticity and disease. *Ann N Y Acad Sci*. 2008; 1144:97–112. Epub 2008/12/17. <https://doi.org/10.1196/annals.1418.005> PMID: 19076369; PubMed Central PMCID: PMC2614307.
79. Ploughman M, Windle V, MacLellan CL, White N, Dore JJ, Corbett D. Brain-derived neurotrophic factor contributes to recovery of skilled reaching after focal ischemia in rats. *Stroke*. 2009; 40(4):1490–5. Epub 2009/01/24. <https://doi.org/10.1161/STROKEAHA.108.531806> PMID: 19164786.
80. Cowansage KK, LeDoux JE, Monfils MH. Brain-derived neurotrophic factor: a dynamic gatekeeper of neural plasticity. *Curr Mol Pharmacol*. 2010; 3(1):12–29. Epub 2009/12/25. <https://doi.org/10.2174/1874467211003010012> PMID: 20030625.
81. Chen J, Zhang C, Jiang H, Li Y, Zhang L, Robin A, et al. Atorvastatin induction of VEGF and BDNF promotes brain plasticity after stroke in mice. *J Cereb Blood Flow Metab*. 2005; 25(2):281–90. Epub 2005/01/29. <https://doi.org/10.1038/sj.jcbfm.9600034> PMID: 15678129; PubMed Central PMCID: PMC2804085.
82. Kim MW, Bang MS, Han TR, Ko YJ, Yoon BW, Kim JH, et al. Exercise increased BDNF and trkB in the contralateral hemisphere of the ischemic rat brain. *Brain Res*. 2005; 1052(1):16–21. Epub 2005/08/02. <https://doi.org/10.1016/j.brainres.2005.05.070> PMID: 16054599.
83. Vaynman S, Ying Z, Gomez-Pinilla F. Hippocampal BDNF mediates the efficacy of exercise on synaptic plasticity and cognition. *Eur J Neurosci*. 2004; 20(10):2580–90. Epub 2004/11/19. <https://doi.org/10.1111/j.1460-9568.2004.03720.x> PMID: 15548201.
84. Chen J, Zacharek A, Zhang C, Jiang H, Li Y, Roberts C, et al. Endothelial nitric oxide synthase regulates brain-derived neurotrophic factor expression and neurogenesis after stroke in mice. *J Neurosci*. 2005; 25(9):2366–75. Epub 2005/03/05. <https://doi.org/10.1523/JNEUROSCI.5071-04.2005> PMID: 15745963; PubMed Central PMCID: PMC2791344.
85. Madinier A, Bertrand N, Mossiat C, Prigent-Tessier A, Beley A, Marie C, et al. Microglial involvement in neuroplastic changes following focal brain ischemia in rats. *PLoS One*. 2009; 4(12):e8101. Epub 2009/12/04. <https://doi.org/10.1371/journal.pone.0008101> PMID: 19956568; PubMed Central PMCID: PMC2779656.
86. Schabitz WR, Steigleder T, Cooper-Kuhn CM, Schwab S, Sommer C, Schneider A, et al. Intravenous brain-derived neurotrophic factor enhances poststroke sensorimotor recovery and stimulates neurogenesis. *Stroke*. 2007; 38(7):2165–72. Epub 2007/05/19. <https://doi.org/10.1161/STROKEAHA.106.477331> PMID: 17510456.
87. Fritsch B, Reis J, Martinowich K, Schambra HM, Ji Y, Cohen LG, et al. Direct current stimulation promotes BDNF-dependent synaptic plasticity: potential implications for motor learning. *Neuron*. 2010; 66(2):198–204. Epub 2010/05/04. <https://doi.org/10.1016/j.neuron.2010.03.035> PMID: 20434997; PubMed Central PMCID: PMC2864780.
88. Clarkson AN, Overman JJ, Zhong S, Mueller R, Lynch G, Carmichael ST. AMPA receptor-induced local brain-derived neurotrophic factor signaling mediates motor recovery after stroke. *J Neurosci*.

- 2011; 31(10):3766–75. Epub 2011/03/11. PubMed Central PMCID: PMC4698878. <https://doi.org/10.1523/JNEUROSCI.5780-10.2011> PMID: 21389231
89. Cook DJ, Nguyen C, Chun HN, I LL, Chiu AS, Machnicki M, et al. Hydrogel-delivered brain-derived neurotrophic factor promotes tissue repair and recovery after stroke. *J Cereb Blood Flow Metab.* 2017; 37(3):1030–45. Epub 2016/05/14. <https://doi.org/10.1177/0271678X16649964> PMID: 27174996; PubMed Central PMCID: PMC5363479.
90. Correa SA, Eales KL. The Role of p38 MAPK and Its Substrates in Neuronal Plasticity and Neurodegenerative Disease. *J Signal Transduct.* 2012; 2012:649079. Epub 2012/07/14. <https://doi.org/10.1155/2012/649079> PMID: 22792454; PubMed Central PMCID: PMC3389708.
91. Colie S, Sarroca S, Palenzuela R, Garcia I, Matheu A, Corpas R, et al. Neuronal p38alpha mediates synaptic and cognitive dysfunction in an Alzheimer's mouse model by controlling beta-amyloid production. *Sci Rep.* 2017; 7:45306. Epub 2017/04/01. <https://doi.org/10.1038/srep45306> PMID: 28361984; PubMed Central PMCID: PMC5374488.
92. Chollet F. Pharmacologic approaches to cerebral aging and neuroplasticity: insights from the stroke model. *Dialogues Clin Neurosci.* 2013; 15(1):67–76. Epub 2013/04/12. <https://doi.org/10.31887/DCNS.2013.15.1fchollet> PMID: 23576890; PubMed Central PMCID: PMC3622470.
93. George PM, Steinberg GK. Novel Stroke Therapeutics: Unraveling Stroke Pathophysiology and Its Impact on Clinical Treatments. *Neuron.* 2015; 87(2):297–309. Epub 2015/07/17. <https://doi.org/10.1016/j.neuron.2015.05.041> PMID: 26182415; PubMed Central PMCID: PMC4911814.
94. Iadecola C, Buckwalter MS, Anrather J. Immune responses to stroke: mechanisms, modulation, and therapeutic potential. *J Clin Invest.* 2020; 130(6):2777–88. Epub 2020/05/12. PubMed Central PMCID: PMC7260029. <https://doi.org/10.1172/JCI135530> PMID: 32391806
95. Kim JM, White JM, Shaw AS, Sleckman BP. MAPK p38 alpha is dispensable for lymphocyte development and proliferation. *J Immunol.* 2005; 174(3):1239–44. Epub 2005/01/22. <https://doi.org/10.4049/jimmunol.174.3.1239> PMID: 15661878.
96. Hayakawa M, Hayakawa H, Petrova T, Ritprajak P, Sutavani RV, Jimenez-Andrade GY, et al. Loss of Functionally Redundant p38 Isoforms in T Cells Enhances Regulatory T Cell Induction. *J Biol Chem.* 2017; 292(5):1762–72. Epub 2016/12/25. <https://doi.org/10.1074/jbc.M116.764548> PMID: 28011639; PubMed Central PMCID: PMC5290950.
97. Risco A, Martin-Serrano MA, Barber DF, Cuenda A. p38gamma and p38delta Are Involved in T Lymphocyte Development. *Front Immunol.* 2018; 9:65. Epub 2018/02/13. <https://doi.org/10.3389/fimmu.2018.00065> PMID: 29434594; PubMed Central PMCID: PMC5796910.
98. O'Donoghue ML, Glaser R, Cavender MA, Aylward PE, Bonaca MP, Budaj A, et al. Effect of Losmapimod on Cardiovascular Outcomes in Patients Hospitalized With Acute Myocardial Infarction: A Randomized Clinical Trial. *JAMA.* 2016; 315(15):1591–9. Epub 2016/04/05. <https://doi.org/10.1001/jama.2016.3609> PMID: 27043082.
99. Ridker PM. Interleukin-1 inhibition and ischaemic stroke: has the time for a major outcomes trial arrived? *Eur Heart J.* 2018; 39(38):3518–20. Epub 2018/06/30. <https://doi.org/10.1093/eurheartj/ehy360> PMID: 29955817.
100. Smith CJ, Hulme S, Vail A, Heal C, Parry-Jones AR, Scarth S, et al. SCIL-STROKE (Subcutaneous Interleukin-1 Receptor Antagonist in Ischemic Stroke): A Randomized Controlled Phase 2 Trial. *Stroke.* 2018; 49(5):1210–6. Epub 2018/03/24. <https://doi.org/10.1161/STROKEAHA.118.020750> PMID: 29567761.
101. Salmeron KE, Maniskas ME, Edwards DN, Wong R, Rajkovic I, Trout A, et al. Interleukin 1 alpha administration is neuroprotective and neuro-restorative following experimental ischemic stroke. *J Neuroinflammation.* 2019; 16(1):222. Epub 2019/11/16. <https://doi.org/10.1186/s12974-019-1599-9> PMID: 31727174; PubMed Central PMCID: PMC6857151.
102. Small SL, Buccino G, Solodkin A. Brain repair after stroke—a novel neurological model. *Nat Rev Neurol.* 2013; 9(12):698–707. Epub 2013/11/13. <https://doi.org/10.1038/nrneurol.2013.222> PMID: 24217509; PubMed Central PMCID: PMC5549938.
103. Balkaya M, Krober JM, Rex A, Endres M. Assessing post-stroke behavior in mouse models of focal ischemia. *J Cereb Blood Flow Metab.* 2013; 33(3):330–8. Epub 2012/12/13. <https://doi.org/10.1038/jcbfm.2012.185> PMID: 23232947; PubMed Central PMCID: PMC3587814.