MAJOR ARTICLE







Naturally Acquired Protection Against Upper Respiratory Symptoms Involving Group A *Streptococcus* in a Longitudinal Cohort Study

Joseph A. Lewnard, 1,2,3 Lilith K. Whittles, 4,5,6 Anne-Marie Rick, 7,8 and Judith M. Martin 7,8

¹Division of Epidemiology, School of Public Health, University of California, Berkeley, Berkeley, California, USA, ²Division of Infectious Diseases and Vaccinology, School of Public Health, University of California, Berkeley, Berkeley, California, USA, ³Center for Computational Biology, College of Engineering, University of California, Berkeley, Berkeley, California, USA, ⁴Department of Infectious Disease Epidemiology, School of Public Health, Imperial College London, London, United Kingdom, ⁵Medical Research Council Centre for Global Infectious Disease Analysis, School of Public Health, Imperial College London, London, United Kingdom, ⁶National Institute for Health Research Health Protection Research Unit in Modelling Methodology, School of Public Health, Imperial College London, United Kingdom, ⁷Department of Pediatrics, School of Medicine, University of Pittsburgh, Pennsylvania, USA, and ⁸Department of Pediatrics, University of Pittsburgh Medical Center Children's Hospital of Pittsburgh, University of Pittsburgh, Pennsylvania, USA

Background. Pharyngitis due to group A *Streptococcus* (GAS) represents a major cause of outpatient visits and antibiotic use in the United States. A leading vaccine candidate targets 30 of the >200 *emm* types of GAS. We aimed to assess natural protection conferred by GAS against respiratory symptoms.

Methods. In a 5-year study among school-aged children in Pittsburgh, Pennsylvania, pharyngeal cultures were obtained from children at 2-week intervals, and active surveillance was conducted for respiratory illnesses. We assessed protection via the relative odds of previous detection of homologous strains (defined by field-inversion gel electrophoresis banding pattern), *emm* types, and *emm* clusters at visits where GAS was detected with symptoms, vs visits where GAS was detected without symptoms. We used a cluster bootstrap of children to adjust estimates for repeated sampling.

Results. At visits where previously detected GAS *emm* types were identified, we estimated 81.8% (95% confidence interval [CI], 67.1%–91.7%) protection against typical pharyngitis symptoms among children reacquiring the same strain, and 94.5% (95% CI, 83.5%–98.6%) protection among children acquiring a distinct strain. We estimated 77.1% (95% CI, 33.7%–96.3%) protection against typical symptoms among children acquiring partially heterologous *emm* types belonging to a previously detected *emm* cluster. Protection was evident after both symptomatic and asymptomatic detections of GAS. We did not identify strong evidence of protection against atypical respiratory symptoms.

Conclusions. Within a 5-year longitudinal study, previous detection of GAS *emm* types was associated with protection against typical symptoms when homologous strains were subsequently detected. Naturally acquired protection against partially heterologous types suggests that *emm* type–based vaccines may have broader strain coverage than what has been previously assumed.

Keywords. group A *Streptococcus*; pharyngitis; naturally acquired protection; cohort study.

Group A Streptococcus (GAS; Streptococcus pyogenes) causes a spectrum of clinical manifestations encompassing infections of the skin and upper respiratory tract as well as severe invasive infections, scarlet fever, rheumatic fever, rheumatic heart disease, and poststreptococcal glomerulonephritis. Of these, GAS pharyngitis is the most common illness, causing an estimated 13 cases per 100 children aged 5–12 years annually and substantial antibiotic prescribing in settings where antibiotic treatment of GAS pharyngitis is recommended [1, 2]. Children with GAS pharyngitis who do not receive antibiotics may spread infection and

are at risk for suppurative and nonsuppurative complications including rheumatic heart disease, which remains a prevalent cause of morbidity and mortality in lower-income settings [3].

An effective GAS vaccine would help to reduce GAS disease burden [4, 5]. Among many GAS surface antigens, the M protein is the best characterized and has received the greatest attention as a vaccine target. More than 200 *emm* types of GAS have been defined based on sequences of the hypervariable M protein–encoding gene [6]. Multiple co-circulating strains may encode each *emm* type, in addition to other GAS antigens and virulence factors [7, 8]. Recently, *emm* types have been partitioned into 48 *emm* clusters based on shared molecular properties [9]. A leading vaccine candidate targets 30 *emm* types of GAS across 10 distinct *emm* clusters [10], which collectively account for the greatest share of GAS pharyngitis and invasive disease in Western high-income settings [11].

Evidence that natural exposure to a pathogen confers protection against recurrent infection or disease helps to establish

Clinical Infectious Diseases® 2020;71(8):e244–54

© The Author(s) 2020. Published by Oxford University Press for the Infectious Diseases Society of America. All rights reserved. For permissions, e-mail: journals.permissions@oup.com. DOI: 10.1093/cid/ciaa044

Received 16 October 2019; editorial decision 10 January 2020; accepted 15 January 2020; published online January 19, 2020.

Correspondence: J. A. Lewnard, School of Public Health, University of California, Berkeley, 2121 Berkeley Way, Office 5410, Berkeley, CA 94720 (jlewnard@berkeley.edu).

the feasibility of an efficacious vaccine, and provides a baseline against which the degree of protection conferred by vaccination can be assessed [12]. Historical studies reported reduced likelihood of clinical symptoms among individuals who acquired M serotypes of GAS against which they had preexisting antibodies [13–16]. While this mechanism may account for the lower incidence of GAS pharyngitis among adults than children [17], there have been few modern studies addressing protection and immunity following natural GAS acquisition [18–22].

A previously conducted longitudinal study addressing GAS carriage and pharyngitis among school-aged children [23, 24] presented an opportunity to assess evidence of naturally acquired protection. We revisited data from this study, aiming to estimate protection against symptomatic GAS detections.

MATERIALS AND METHODS

Cohort

We used data from a 5-year school-based longitudinal cohort study of children in Pittsburgh, Pennsylvania, undertaken between October 1998 and May 2003. Study methods have been described previously [23]; in brief, all children (~285 per year) enrolled in a private, tuition-supported elementary school serving students in kindergarten through grade 8 were eligible to participate. As classroom placement within the school is based on readiness rather than age or grade level, significant social mixing occurs across age groups. Study participants came from all classrooms in the school, providing a representative illustration of transmission dynamics.

Throat cultures were performed for each child enrolled in the study approximately every 2 weeks during the school year. In addition, study personnel were "on call" to obtain throat cultures from children within 1 day of onset of any new respiratory illness; if children received care for respiratory illnesses from their personal clinician and if throat cultures were performed by the practitioner, they were retrieved by study personnel. For antibiotic-treated episodes, follow-up cultures were performed 2–4 days after end of treatment; for new GAS detections not treated with antibiotics, because the child did not have respiratory symptoms, follow-up cultures were performed 1 week after the initial positive culture.

Throat specimens were obtained with a rayon-tipped swab (BBL, Becton Dickinson, Sparks, Maryland) and processed the same day by previously described standard culture techniques [24]. In brief, swabs were transported in Amies medium without charcoal and plated within 2 hours on 5% sheep blood agar with bacitracin disks for incubation at 37°C in 5% carbon dioxide, before isolation and subculturing of β -hemolytic colonies. Isolates were typed initially by field-inversion gel electrophoresis (FIGE; a version of pulsed-field gel electrophoresis [25]) using DNA bands within the range of 50–250 kb. Isolates with

identical FIGE banding patterns were considered to represent a single strain. From each FIGE type, 1 or more representative isolates were sent annually for *emm* typing by the Centers for Disease Control and Prevention, as described previously [26].

At each visit, physical examination of the pharynx was performed, and children were questioned about symptoms using a standardized questionnaire. When respiratory illnesses occurred, parents were contacted for additional information regarding symptoms. Children reporting sore throat as a prominent clinical complaint were defined as exhibiting typical symptoms; symptoms were considered atypical if children reported rhinorrhea and/or cough without sore throat. Detection of GAS without accompanying respiratory symptoms was considered colonization.

Design

We aimed to estimate the association of prior GAS detection with protection against typical and atypical symptoms at future visits where the same emm type was detected (homologous), and at visits where a distinct emm type belonging to the same emm cluster was detected (partially heterologous). We estimated protection in a case-control framework. Specifically, we compared the odds of previous detection of homologous or partially heterologous emm types at GAS-positive visits where children experienced typical or atypical symptoms ("case" visits) to odds of previous homologous or partially heterologous emm type detection at GAS-positive visits where children experienced no symptoms ("control" visits). We distinguished recurrent detections of the same emm type according to whether the same FIGE type, or a new FIGE type, was detected. To prevent misclassification of continuous carriage episodes, we defined recurrent detections of FIGE types as visits separated from previous detection of the same type by at least 2 GAS-negative culture results. Visits preceded by detection of the same FIGE type without 2 intervening GAS-negative specimens were excluded from analysis.

Because antibiotic treatment of GAS episodes may prevent seroresponse [15], we also aimed to assess whether receipt of antibiotics influenced the likelihood of protection. For these analyses, we defined independent variables as follows: (1) previous detection ever occurring with an antibiotic prescription (vs no previous detection); or (2) previous detection never occurring with an antibiotic prescription (vs no previous detection).

We further stratified analyses according to whether children had ever experienced typical symptoms, or had never experienced symptoms, at previous detections of an *emm* type or *emm* cluster.

Because the study had an open cohort design with enrollment of new children each year, the risk of missing GAS acquisitions while children were not under surveillance varied among children and over time. We therefore constructed analysis strata within which children were matched on current GAS exposure and the extent of prior surveillance: visits were stratified by semester of occurrence (10 semesters over 5 school years) and the semester children entered the study, resulting in 100 distinct strata.

Statistical Methods

We used the χ^2 test to compare proportions of children with typical symptoms, atypical symptoms, and no symptoms at their first and second detection of FIGE types. For analyses of protection, we estimated Mantel-Haenszel (matched) odds ratios, accounting for strata, via conditional logistic regression. We defined protection estimates as 1 minus the matched odds ratio. Because children contributed multiple visits, we did statistical inference in a cluster-bootstrap framework, resampling study participants at each iteration [27]. We coded the cluster bootstrap de novo and used the *survival* package [28] in R software (version 3.5.3) to fit conditional logistic regression models.

As a sensitivity analysis, we repeated analyses of protection within a subset of visits occurring after children had contributed ≥ 2 years of observations. We expected this subsample would have reduced risk of misclassification resulting from failure to detect GAS acquisitions that occurred outside the study period.

RESULTS

Enrollment and GAS Detection

The study enrolled 145 children, among whom GAS was detected in 110 (Table 1; Supplementary Tables 1–3). Children enrolled during years 1–5 contributed specimens over, on average, 34.0 (range, 8–46), 26.0 (range, 8–34), 20.8 (range, 8–28), 11.9 (range, 3–19), and 8.6 (range, 3–9) distinct months (Supplementary Figure 1). In total, cultures were collected at 7241 visits, and GAS was detected at 1120 visits (15.5%). Typical symptoms among GAS-positive children occurred at 194 visits (82 children), whereas atypical symptoms among GAS-positive children occurred at 82 visits (among 54 children). In addition, GAS was detected without respiratory symptoms at 844 visits (among 78 children).

Dynamics of emm Types

Descriptions of GAS carriage and disease in the cohort have been reported previously [23]. In brief, predominant *emm* types and FIGE types varied by year (Table 2); the majority of isolates belonged to *emm29* and *emm4* in 1998–1999, and to *emm28* and *emm89* in 1999–2000 (Supplementary Figure 2). Detections of *emm6* became prominent in the winter of 2000, and this type became the most prevalent over all subsequent years of the study, accounting for 49.1%, 54.9%, and 30.4% of GAS isolates in 2000–2001, 2001–2002, and 2002–2003, respectively.

Additional prevalent types over these years included *emm*89, *emm*12, *emm*28, *emm*5, and *emm*1.

The contribution of each type to disease and colonization further varied by year (Supplementary Figure 2). For instance, typical symptoms occurred with 22.4% (35/156) of *emm6* detections in 2000–2001, but only 9.2% (12/130) and 4.3% (3/70) of *emm6* detections in 2001–2002 and 2002–2003, respectively. Similarly, typical symptoms occurred with 21.2% (7/33) of *emm8*9 detections in 1999–2000, but only 10.5% (6/58) of *emm8*9 detections in 2000–2001. In 2001–2002, typical symptoms occurred with 31.6% (6/19) and 36.3% (21/130) of *emm1* and *emm*12 detections, respectively, compared with 7.1% (1/14) and 23.2% (14/70) of detections of these same types in 2002–2003.

Recurrent Detections

We identified 280 visits where an FIGE type was newly detected in a child without history of carriage or disease involving the same type (Table 3). Typical and atypical symptoms were noted at 112 (40.0%) and 41 (14.6%) of these visits, respectively; children had no respiratory symptoms at the remaining 127 (45.4%) visits. A second detection of the same FIGE type, separated by ≥2 GAS-negative cultures, occurred in 83 instances. Of these second detections, 17 (20.5%), 8 (9.6%), and 58 (69.9%) presented with typical symptoms, atypical symptoms, and no symptoms, respectively, representing a notable departure from the distribution of symptoms on first detection $(\chi^2_{\rm df=2} = 15.6; P = .004)$. We did not identify differences in symptoms at second detections among children whose first detections involved typical symptoms, atypical symptoms, or no symptoms ($\chi_{df=4}^2 = 3.3; P = .5$). These results suggest that symptomatic and asymptomatic GAS acquisitions conferred similar protection against symptoms at future detections of the same FIGE type.

Protection

We estimated 81.8% (95% confidence interval [CI], 67.1%–91.7%) protection against typical symptoms associated with previous detections of the same FIGE type, separated by ≥ 2 intervening GAS-negative cultures (Table 4). In an analysis limited to visits preceded by ≥ 2 years of surveillance, we estimated 85.9% (95% CI, 47.1%–97.8%) protection. Previous detections of a distinct FIGE type, belonging to the same *emm* type, were associated with 94.5% (95% CI, 83.5%–98.6%) protection against typical symptoms in the full sample and 82.7% (95% CI, 27.3%–97.1%) protection at visits preceded by ≥ 2 years of surveillance. In these analyses, differences in estimated protection against the same and distinct FIGE types were not statistically meaningful at the $P \leq .05$ threshold. We estimated 77.1% (95% CI, 33.7%–96.3%) protection against typical symptoms associated with previous detections of a partially

Table 1. Study Enrollment and Observations

			Academ	ic Year		
Entry Year and Observation	1998–1999	1999–2000	2000–2001	2001–2002	2002–2003	All Years
1998–1999						
Total enrollment	48	45	41	34	26	48
Samples obtained	829	800	736	601	466	3432
Children with GAS detection without symptoms	24	17	16	13	7	30
Children with GAS-positive typical symptoms	12	15	16	8	5	28
Children with GAS-positive atypical symptoms	8	6	5	5	2	18
Visits with GAS detection without symptoms	92	68	111	69	49	389
Visits with GAS-positive typical symptoms	12	21	26	10	8	77
Visits with GAS-positive atypical symptoms	11	7	5	7	2	32
1999–2000						
Total enrollment		30	26	22	18	30
Samples obtained		520	470	410	316	1716
Children with GAS detection without symptoms		15	12	6	3	21
Children with GAS-positive typical symptoms		11	11	10	7	21
Children with GAS-positive atypical symptoms		8	5	1	2	13
Visits with GAS detection without symptoms		89	77	39	23	228
Visits with GAS-positive typical symptoms		15	17	14	8	54
Visits with GAS-positive atypical symptoms		11	6	1	3	21
2000–2001						
Total enrollment			32	26	20	32
Samples obtained			540	478	371	1389
Children with GAS detection without symptoms			10	9	6	12
Children with GAS-positive typical symptoms			14	11	7	23
Children with GAS-positive atypical symptoms			7	5	0	11
Visits with GAS detection without symptoms	•••	•••	58	55	36	149
Visits with GAS-positive typical symptoms		•••	21	15	11	47
Visits with GAS-positive typical symptoms Visits with GAS-positive atypical symptoms	•••	***	7	7	0	14
2001–2002	•••	•••	,	,	U	17
Total enrollment				15	8	15
Samples obtained				216	148	364
Children with GAS detection without symptoms				5	4	8
Children with GAS-positive typical symptoms	•••	•••	•••	4	3	6
Children with GAS-positive atypical symptoms	•••	•••	•••	4	5	9
Visits with GAS detection without symptoms		•••	•••	19	27	46
Visits with GAS-positive typical symptoms	•••	•••	•••	5	6	11
Visits with GAS-positive typical symptoms Visits with GAS-positive atypical symptoms		•••	•••	5	7	12
2002–2003	•••	•••	•••	5	,	12
Total enrollment					20	20
Samples obtained	•••	•••	•••		340	340
Children with GAS detection without symptoms	•••	•••	•••	•••	7	7
	•••	•••	•••	•••		
Children with GAS-positive typical symptoms			***	•••	4	4
Children with GAS-positive atypical symptoms		•••	***	***	3	3
Visits with GAS detection without symptoms	•••	•••	•••	•••	32	32
Visits with GAS-positive typical symptoms	•••		•••		5	5
Visits with GAS-positive atypical symptoms		•••		•••	3	3
Full cohort	40	74	00	07	00	4.45
Total enrollment	48	74	99	97	92	145
Samples obtained	829	1320	1746	1705	1641	7241
Children with GAS detection without symptoms	24	32	38	33	27	78
Children with GAS-positive typical symptoms	12	26	41	33	26	82
Children with GAS-positive atypical symptoms	8	14	17	15	12	54
Visits with GAS detection without symptoms	92	157	246	182	167	844
Visits with GAS-positive typical symptoms	12	36	64	44	38	194
Visits with GAS-positive atypical symptoms	11	18	18	20	15	82

Data are presented as no.

Abbreviation: GAS, group A Streptococcus.

Table 2. Group A Streptococcus Types Observed During Study Period

Charten and					Year		
emm Cluster and emm Type	FIGE Type	1998–1999	1999–2000	2000–2001	2001–2002	2002–2003	All Years
E1							
4	1	35	19	0	0	4	58
E3							
58	2	0	0	0	0	9	9
E4							
22	3	0	0	1	0	0	1
28	4	0	91	33	12	0	136
77	5	0	0	3	0	0	3
89	6	0	34	57	9	36	136
89	7	0	0	0	6	0	6
89	8	0	0	0	0	19	19
E6							
75	9	3	11	7	2	10	33
94	10	0	1	0	0	0	1
A-C3							
1	11	9	6	6	19	5	45
1	12	0	0	0	2	7	9
A-C4							
12	13	0	1	0	12	57	70
12	14	0	0	0	25	0	25
A-C5							
3	15	0	6	16	0	0	22
3	16	0	0	0	18	0	18
5							
5	17	0	0	0	3	0	3
5	18	6	5	25	7	5	48
5	19	7	0	0	0	0	7
6							
6	20	2	20	156	4	0	182
6	21	0	0	0	5	35	40
6	22	0	0	0	126	0	126
6	23	0	0	0	3	27	30
29							
29	24	53	17	11	0	0	81

Cell values indicate the number of isolates, from each year, belonging to each FIGE type, grouped in the left-hand column by the associated emm types and emm clusters. FIGE types are defined by matched banding patterns.

Abbreviation: FIGE, field inversion gel electrophoresis.

heterologous emm type (Table 4). Within the sample of visits preceded by ≥ 2 years of surveillance, data were available from only 16 visits where a partially heterologous emm type was previously detected. We did not identify strong evidence of protection against atypical symptoms associated with previous detection of the same emm type or emm cluster (Table 5).

We estimated 89.6% (95% CI, 66.3%–97.1%) and 42.0% (95% CI, –168.8% to 87.0%) protection against typical and atypical symptoms, respectively, associated with previous detections of the same FIGE type at visits where antibiotics were not prescribed (Table 6). For previous detections resulting in antibiotic prescriptions, we estimated 71.2% (95% CI, 37.8%–89.4%) protection against typical symptoms. We estimated 95.1% (95% CI, 77.3%–99.1%) and 71.1% (95% CI, –120.0% to 95.7%) protection against typical and atypical symptoms, respectively,

associated with previous detections of a distinct FIGE type belonging to the same *emm* type and occurring without an antibiotic prescription. Previous detections of a distinct FIGE type belonging to the same *emm* type, when accompanied by an antibiotic prescription, were associated with 94.1% (95% CI, 71.0%–98.9%) protection against typical symptoms. Previous detections of a partially heterologous *emm* type were associated with 86.7% (95% CI, 38.0%–97.0%) protection against typical symptoms when antibiotics were not prescribed, and with 48.8% (95% CI, -75.4% to 88.4%) protection against typical symptoms when antibiotics were prescribed.

Previous detections of the same *emm* type occurring with typical symptoms and resulting in an antibiotic prescription were associated with 73.2% (95% CI, 36.3%–91.6%) against typical symptoms involving the same FIGE type, and 94.7%

Table 3. Summary of Recurrent Detections of Group A Streptococcus Strains

First Oc	currence of Any Fl	GE Type	Second Occurrence of Same FIGE Type	e, Separated by ≥2 Negative Cultures
				%)
Symptoms	No. (%)	Symptoms	Among Children With First Detection	Among Children With Recurrence
Typical symptoms ^{a,b}	112 (40.0)			
		Typical symptoms	9 (8.0)	9 (27.3)
		Atypical symptoms	3 (2.7)	3 (9.1)
		Without symptoms	21 (18.8)	21 (63.6)
		Any second detection	33 (29.5)	33 (100)
		No second detection	79 (70.5)	
		Total	112 (100)	
Atypical symptoms ^{a,b}	41 (14.6)			
		Typical symptoms	2 (4.9)	2 (20.0)
		Atypical symptoms	2 (4.9)	2 (20.0)
		Without symptoms	6 (14.6)	6 (60.0)
		Any second detection	10 (24.4)	10 (100)
		No second detection	31 (75.6)	***
		Total	41 (100)	
No symptoms ^{a,b}	127 (45.4)			
		Typical symptoms	6 (4.7)	6 (15.0)
		Atypical symptoms	3 (2.4)	3 (7.5)
		Without symptoms	31 (24.4)	31 (77.5)
		Any second detection	40 (31.5)	40 (100)
		No second detection	87 (68.5)	
		Total	127 (100)	***
Any symptoms status ^b	280 (100)			
		Typical symptoms	17 (6.1)	17 (20.5)
		Atypical symptoms	8 (2.9)	8 (9.6)
		Without symptoms	58 (20.7)	58 (69.9)
		Any second detection	83 (29.6)	83 (100)
		No second detection	197 (70.4)	
		Total	280 (100)	

Entries indicate the number of distinct FIGE types detected and redetected after ≥2 intervening group A *Streptococcus* (GAS)–negative cultures, for each child, summed over all children. FIGE types are defined by matched banding patterns.

Abbreviation: FIGE, field inversion gel electrophoresis

(95% CI, 77.2%–98.9%) protection against typical symptoms involving a distinct FIGE type (Table 7). Similarly, we estimated 76.2% (95% CI, 33.6%–94.7%) and 92.8% (95% CI, 66.8%–98.8%) protection against typical symptoms involving the same and distinct FIGE types, respectively, associated with previous asymptomatic detections of the same *emm* type where no antibiotic prescription occurred. Differences in estimated protection against matched and distinct FIGE types were again not statistically meaningful. Previous detection of a partially heterotypic *emm* type was associated with 47.5% (95% CI, –234.0% to 93.8%) protection against typical symptoms when earlier visits occurred with typical symptoms and an antibiotic prescription, and 84.9% (95% CI, 25.9%–96.8%) protection when earlier visits occurred without symptoms or an antibiotic prescription.

DISCUSSION

Evidence of naturally acquired protection against a pathogen strengthens the rationale for vaccine development and provides a benchmark for assessing vaccine efficacy. We identified that prior detection of GAS is associated with protection against typical symptoms at future detections of the same *emm* type or cluster. Our finding of protection after detections of distinct FIGE types belonging to the same *emm* type is consistent with the hypothesis of protective responses to the M protein. These findings support the biological plausibility of preventing symptomatic GAS upper respiratory infections using *emm* type–based vaccines.

Whereas it has previously been thought that GAS carriage does not cause immune responses [21], we find evidence of type-specific protection against typical symptoms after asymptomatic GAS detections. These findings are consistent with

^aWe do not identify strong evidence of a difference in the proportion of children with typical symptoms, atypical symptoms, and no symptoms on their second detection of an FIGE type among those who experienced typical symptoms, atypical symptoms, or no symptoms on their first detection of the same PFGE type ($\chi^2_{df=4} = 3.3$; P = .5).

The distribution of typical symptoms, atypical symptoms, and no symptoms on second detections of an FIGE type differs significantly from the distribution of typical symptoms, atypical symptoms, and no symptoms on first detections ($\chi^2_{df=2} = 15.6$; P = .004).

Table 4. History of Homologous-type Detection at Group A Streptococcus-Positive Visits With Typical Symptoms and No Symptoms

	Previously Dete	ected FIGE Type		ype Belonging to ected <i>emm</i> Type	-	m type Belonging to ected <i>emm</i> Cluster
Outcome	No Previous Detection ^a	Previous Detection	No Previous Detection ^a	Previous Detection	No Previous Detection ^a	Previous Detection
All visits						
GAS-positive without symptoms, No. of visits	93	68	503	192	503	69
GAS-positive typical symptoms, No. of visits	112	18	150	15	150	7
Est. protection against typical symptoms, % (95% CI)	Ref	81.8 (67.1–91.7)	Ref	94.5 (83.5–98.6)	Ref	77.1 (33.7–96.3)
Visits preceded by ≥2 y surveillance						
GAS-positive without symptoms, No. of visits	35	39	202	68	202	11
GAS-positive typical symptoms, No. of visits	44	8	58	4	58	5
Est. protection against typical symptoms, % (95% CI)	Ref	85.9 (47.1–97.8)	Ref	82.7 (27.3–97.1)	Ref	-21.7 (-1519.1 to 85.5)

Estimates of protection are obtained as 1 minus the matched odds ratio, accounting for matching strata. We derive 95% Cls via cluster-bootstrap resampling of children. FIGE types are defined by matched banding patterns.

Abbreviations: CI, confidence interval; Est., estimated; FIGE, field inversion gel electrophoresis; GAS, group A Streptococcus; Ref, reference group.

those of a recent longitudinal study, wherein 70% of new asymptomatic GAS acquisitions resulted in detectable antibody responses [18]. Previous studies have also demonstrated reduced likelihood of seroconversion when GAS episodes are treated with antibiotics [15]. The fact that most antibiotic-treated episodes presented with typical symptoms makes it difficult to disentangle effects of previous symptoms and antibiotic

treatment on children's likelihood of future protection in this study. We obtained lower point estimates of protection against typical symptoms associated with detections occurring with an antibiotic prescription, as compared to detections without an antibiotic prescription. While greater differences in our estimates of protection against atypical symptoms are evident when comparing previous detections that occurred with or without

Table 5. History of Homologous-type Detection at Group A Streptococcus—Positive Visits With Atypical Symptoms and No Symptoms

	Previously Det	tected FIGE Type		Type Belonging to etected <i>emm</i> Type	-	emm Type Belonging to Detected emm Cluster
Outcome	No Previous Detection ^a	Previous Detection	No Previous Detection ^a	Previous Detection	No Previous Detection ^a	Previous Detection
All visits						
GAS-positive without symptoms, no. of visits	93	68	503	192	503	69
GAS-positive atypical symptoms, no. of visits	28	12	42	15	42	7
Est. protection against atypical symptoms, % (95% CI)	Ref	8.3 (-133.0 to 67.9)	Ref	46.5 (-353.2 to 90.00)	Ref	-84.8 (-364.5 to 46.1)
Visits preceded by ≥2 y surveillance						
GAS-positive without symptoms, no. of visits	35	39	202	68	202	11
GAS-positive atypical symptoms, no. of visits	5	5	10	3	10	0
Est. protection against atypical symptoms, % (95% CI)	Ref	13.3 (-∞ to 82.7)	Ref	32.3 (-423.2 to 90.0)	Ref	

Estimates of protection are obtained as 1 minus the matched odds ratio, accounting for matching strata. We derive 95% Cls via cluster-bootstrap resampling of children. FIGE types are defined by matched banding patterns.

Abbreviations: Cl, confidence interval; Est., estimated; FIGE, field inversion gel electrophoresis; GAS, group A Streptococcus; Ref, reference group.

aWe define "no previous detection" as no prior detection of any isolate belonging to the same FIGE type, emm type, or emm cluster. Fewer observations are available for analyses of prior detection of the same FIGE type because we exclude second detections not separated from prior detections by ≥2 GAS-negative swabs.

^aWe define "no previous detection" as no prior detection of any isolate belonging to the same FIGE type, *emm* type, or *emm* cluster. Fewer observations are available for analyses of prior detection of the same FIGE type because we exclude second detections not separated from prior detections by ≥2 GAS-negative swabs.

Table 6. History of Homologous-type Detection and Antibiotic Receipt at Group A Streptococcus-Positive Visits With Typical Symptoms, Atypical Symptoms, and No Symptoms

		Previously Detected FI	FIGEType		Distinct FIGE Type Belonging to Previously Detected <i>emm</i> Type	longing to <i>emm</i> Type	Heter	Heterologous <i>emm</i> Type Belonging to Previously Detected <i>emm</i> Cluster	Belonging to nm Cluster
		Previous	Previous Detection		Previous	Previous Detection		Previous	Previous Detection
Outcome	No Previous Detection ^a	No Antibiotic Prescribed	Antibiotic Prescribed	No Previous Detection ^a	No Antibiotic Prescribed	Antibiotic Prescribed	No Previous Detection ^a	No Antibiotic Prescribed	Antibiotic Prescribed
GAS-positive without symptoms, No. of visits	83	39	29	503	125	67	503	45	24
GAS-positive typical symptoms, No. of visits	112	Ø	12	150	9	ത	150	2	ιΩ
GAS-positive atypical symptoms, No. of visits	28	ന	0	42	9	ത	42	က	4
Est. protection against typical symptoms, from any previous detection, % (95% CI)	Ref	89.6 (66.3–97.1)	71.2 (37.8–89.4)	Ref	95.1 (77.3–99.1)	94.1 (71.0–98.9)	Ref	86.7 (38.0–97.0)	86.7 (38.0–97.0) 48.8 (–75.4 to 88.4)
Est. protection against atypical symptoms, from any previous detection, % (95% CI)	Ref	42.0 (-168.8 to 87.0)	-51.9 (-363.0 to 56.6)	Ref	71.1 (-120.0 to 95.7)	71.1 (-120.0 to 95.7) -20.6 (-1705.0 to 86.1)	Ref	-25.8 (-379.1 to 64.0)	:

VWe define "no previous detection" as no prior detection of any isolate belonging to the same FIGE type, emm type, or emm cluster. Fewer observations are available for analyses of prior detection of the same FIGE type because we exclude second de-Estimates of protection are obtained as 1 minus the matched odds ratio, accounting for matching strata. We derive 95% Cls via cluster-bootstrap resampling of children. FIGE types are defined by matched banding patterns Abbreviations: CI, confidence interval; Est., estimated; FIGE, field inversion gel electrophoresis; GAS, group A Streptococcus; Ref, reference group ections not separated from prior detections by ≥2 GAS-negative swabs an antibiotic prescription, our estimates are underpowered in these strata.

Our findings agree with those of previous studies. In United States Army cohorts, presence of anti-M antibody predicted reduced risk of prolonged colonization and of respiratory symptoms upon reacquisition of homologous GAS M serotypes [13, 14]. In a multiyear study of institutionalized children, GAS pharyngitis epidemics tended to involve M serotypes that had not been detected previously within the population [29]; yearto-year variation in emm types causing pharyngitis and other conditions has also been reported in modern studies [30, 31], and suggests that type-specific protection influences disease dynamics within the community. Age-related increases in the diversity of emm types causing pharyngitis [32], alongside increasing prevalence of antibody against common emm types [33], further suggests that type-specific protection acquired over successive GAS exposures in childhood reduces the incidence of GAS pharyngitis among teens and adults [15, 17, 34]. Contributions of type-specific protection to other features of GAS epidemiology, including differences in the composition and diversity of disease-causing emm types [11, 35], remain important to investigate.

Notably, we identify naturally acquired protection against previously encountered emm types as well as partially heterologous emm types belonging to previously encountered emm clusters. Several lines of evidence support the biological plausibility of this finding. Immune cross-opsonization within emm clusters occurs in animals [10] and humans [19]. In animals, synthetic proteins emulating shared structural components of M protein variants at the level of the emm cluster elicit cross-reactive antibodies and bactericidal activity against partially heterologous emm types [36]. While our study observed only 10 of the 48 emm clusters, the emm types identified account for approximately 70% of GAS isolates in high-income countries [11]. Individual types and clusters may vary in the likelihood of exhibiting cross-protection. Our findings nonetheless suggest the strain coverage of emm type-based vaccines may be greater than what has been previously assumed [11].

Strengths of our study include access to data from children within a single school and community, long-term follow-up with up to 5 years per child to characterize history of GAS detections, and active surveillance of all respiratory symptoms. However, certain limitations should be considered. Children who were GAS culture positive with typical symptoms may not have GAS pharyngitis, as viral coinfections could cause symptoms among GAS culture–positive children. Misclassification may also arise from our inability to account for GAS acquisitions preceding enrollment or occurring during the summers, when sampling did not occur. However, we obtain similar results in analyses restricted to visits preceded by ≥ 2 years of surveillance, suggesting that any resulting bias is small. While FIGE typing enabled us to define unique strains transmitted among children in

Table 7. History of Homologous-type Detection, Symptoms, and Antibiotic Receipt at Group A Streptococcus-Positive Visits With Typical Symptoms, Atypical Symptoms, and No Symptoms

		Previously Detected FIGE	FIGEType	Distinct FIGE Ty	Distinct FIGE Type Belonging to Previously Detected emm Type	viously Detected	Heterolc Previo	Heterologous <i>emm</i> Type Belonging to Previously Detected <i>emm</i> Cluster	nging to Cluster
		Previous Detection	itection		Previous Detection	etection		Previous Detection	Jetection
Exposure and Outcome	No Previous Detection ^a	No Antibiotic Prescribed	Antibiotic Prescribed	No Previous Detection ^a	No Antibiotic Prescribed	Antibiotic Prescribed	No Previous Detection ^a	No Antibiotic Prescribed	Antibiotic Prescribed
Previous detection with typical symptoms									
GAS-positive without symptoms, No. of visits	93	÷	25	503	:	65	503	÷	13
GAS-positive typical symptoms, No. of visits	112	÷	10	150	:	9	150	i	ო
GAS-positive atypical symptoms, No. of visits	28	÷	9	42	:	∞	42	÷	—
Est. protection against typical symptoms, % (95% CI)	Ref	:	73.2 (36.3–91.6)	Ref	:	94.7 (77.2–98.9)	Ref	:	47.5 (-234.0 to 93.8)
Est. protection against atypical symptoms, % (95% CI)	Ref	:	15.5 (-169.7 to 81.1)	Ref	:	38.6 (-238.8 to 90.0)	Ref	÷	÷
Previous detection without symptoms only									
GAS-positive without symptoms, No. of visits	93	18	:	503	88	:	503	37	÷
GAS-positive typical symptoms, No. of visits	112	വ	:	150	9	:	150	2	÷
GAS-positive atypical symptoms, No. of visits	28	က	:	42	ო	:	42	2	÷
Est. protection against typical symptoms, % (95% CI)	Ref	76.2 (33.6–94.7)	:	Ref	92.8 (66.8–98.8)	:	Ref	84.9 (25.9–96.8)	÷
Est. protection against atypical symptoms, % (95% CI)	Ref	-6.3 (-420.5 to 81.9)	:	Ref	77.9 (-105.2 to 95.8)	:	Ref	16.3 (–161.7 to 73.4)	:

Estimates of protection are obtained as 1 minus the matched odds ratio, accounting for matching strata. We derive 95% Cls via cluster-bootstrap resampling of children. FIGE types are defined by matched banding patterns. Abbreviations: CI, confidence interval; Est., estimated; FIGE, field inversion gel electrophoresis; GAS, group A Streptococcus; Ref. reference group.

*We define "no previous detection" as no prior detection of any isolate belonging to the same FIGE type, emm type, or emm cluster. Fewer observations are available for analyses of prior detection of the same FIGE type because we exclude second detections not separated from prior detections by ≥ 2 GAS-negative swabs.

this school-based study, whole-genome sequencing would offer greater resolution for characterizing strains and could enable assessments of protection associated with non-M antigens [7]. We could not fully disentangle differences in protection associated with previous symptoms and antibiotic prescribing. Larger studies, studies undertaken in settings with higher transmission rates, or studies in settings with differing treatment protocols for GAS pharyngitis [37] may better indicate how symptom severity and antibiotics influence naturally acquired protection. Last, our analysis considers only protection against symptoms given new GAS detection, and not protection against colonization. In our study, year-to-year changes in circulating GAS emm types suggest that natural protection may also prevent acquisition of GAS in the upper respiratory tract [38]. Though historical studies presented conflicting evidence of protection against GAS colonization [13, 14, 16], investigational M protein vaccines have conferred type-specific protection against GAS acquisition in the respiratory tract upon challenge [39, 40].

To conclude, we identify that natural GAS acquisition confers strong type-specific protection against future respiratory symptoms when detected a second time during the span of a 5-year longitudinal study. These findings support the plausibility of preventing symptomatic GAS pharyngitis using *emm* type-based vaccines.

Supplementary Data

Supplementary materials are available at *Clinical Infectious Diseases* online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

Notes

Financial support. The original study was supported by the National Institutes of Health (grant number K23-AI0713-02 to J. M. M.). L. K. W. acknowledges joint center funding from the United Kingdom Medical Research Council and Department for International Development (grant MR/R015600/1) and support from the National Institute for Health Research Health Protection Research Unit in Modelling Methodology at Imperial College, London, in partnership with Public Health England (grant number HPRU-2012-10080).

Potential conflicts of interest. The authors: No reported conflicts of interest. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest.

References

- Danchin MH, Rogers S, Kelpie L, et al. Burden of acute sore throat and group A streptococcal pharyngitis in school-aged children and their families in Australia. Pediatrics 2007; 120:950-7.
- Fleming-Dutra KE, Hersh AL, Shapiro DJ, et al. Prevalence of inappropriate antibiotic prescriptions among US ambulatory care visits, 2010–2011. JAMA 2016; 315:1864–73.
- Carapetis JR, Steer AC, Mulholland EK, Weber M. The global burden of group A streptococcal diseases. Lancet Infect Dis 2005; 5:685–94.
- Vekemans J, Gouvea-Reis F, Kim JH, et al. The path to group A Streptococcus vaccines: World Health Organization research and development technology roadmap and preferred product characteristics. Clin Infect Dis 2019; 69:877–83.
- Osowicki J, Vekemans J, Kaslow DC, Friede MH, Kim JH, Steer AC. WHO/IVI global stakeholder consultation on group A Streptococcus vaccine development: report from a meeting held on 12–13 December 2016. Vaccine 2018; 36:3397–405.

- Facklam RF, Martin DR, Marguerite L, et al. Extension of the Lancefield classification for group A streptococci by addition of 22 new M protein gene sequence types from clinical isolates: emm103 to emm124. Clin Infect Dis 2002; 34:28–38.
- Chochua S, Metcalf BJ, Li Z, et al. Population and whole genome sequence based characterization of invasive group A streptococci recovered in the United States during 2015. MBio 2017; 19:e01422–17.
- Kachroo P, Eraso JM, Beres SB, et al. Integrated analysis of population genomics, transcriptomics and virulence provides novel insights into *Streptococcus pyogenes* pathogenesis. Nat Genet 2019; 51:548–59.
- Sanderson-Smith M, De Oliveira DMP, Guglielmini J, et al. A systematic and functional classification of *Streptococcus pyogenes* that serves as a new tool for molecular typing and vaccine development. J Infect Dis 2014; 210:1325–38.
- Dale JB, Penfound TA, Chiang EY, Walton WJ. New 30-valent M protein-based vaccine evokes cross-opsonic antibodies against non-vaccine serotypes of group A streptococci. Vaccine 2011; 29:8175–8.
- Steer AC, Law I, Matatolu L, Beall BW, Carapetis JR. Global *emm* type distribution of group A streptococci: systematic review and implications for vaccine development. Lancet Infect Dis 2009; 9:611–6.
- 12. Lopman B, Kang G. In praise of birth cohorts: norovirus infection, disease, and immunity. Clin Infect Dis **2014**; 58:492–4.
- Wannamaker LW, Denny FW, Perry WD, Siegel AC, Rammelkamp CH Jr. Studies on immunity to streptococcal infections in man. AMA Am J Dis Child 1953; 86:347–8.
- Wannamaker LW, Ferrieri P. Streptococcal infections—updated. Disease-amonth 1975; 21:1–40.
- Lancefield RC. Current knowledge of type-specific M antigens of group A streptococci. J Immunol 1962; 89:307–13.
- Guirguis N, Fraser DW, Facklam RR, El Kholy A, Wannamaker LW. Type-specific immunity and pharyngeal acquisition of group A *Streptococcus*. Am J Epidemiol 1982: 116:933–9.
- Tsoi SK, Smeesters PR, Frost HR, Licciardi P, Steer AC. Correlates of protection for M protein-based vaccines against group A Streptococcus. J Immunol Res 2015; 2015;167089.
- Hysmith ND, Kaplan EL, Cleary PP, Johnson DR, Penfound TA, Dale JB. Prospective longitudinal analysis of immune responses in pediatric subjects after pharyngeal acquisition of group A streptococci. J Pediatric Infect Dis Soc 2017; 6:187-96
- Frost HR, Laho D, Sanderson-Smith ML, et al. Immune cross-opsonization within *emm* clusters following group A *Streptococcus* skin infection: broadening the scope of type-specific immunity. Clin Infect Dis 2017; 65:1523–31.
- Johnson DR, Kurlan R, Leckman J, Kaplan EL. The human immune response to streptococcal extracellular antigens: clinical, diagnostic, and potential pathogenetic implications. Clin Infect Dis 2010; 50:481–90.
- Shulman ST, Tanz RR. Strep: where do we go from here? J Pediatric Infect Dis Soc 2017; 6:197–8.
- Raynes JM, Frost HR, Williamson DA, et al. Serological evidence of immune priming by group A streptococci in patients with acute rheumatic fever. Front Microbiol 2016: 7:1119.
- Martin JM, Green M, Barbadora KA, Wald ER. Group A streptococci among school-aged children: clinical characteristics and the carrier state. Pediatrics 2004; 114:1212–9.
- Martin JM, Green M, Barbadora KA, Wald ER. Erythromycin-resistant group A streptococci in schoolchildren in Pittsburgh. N Engl J Med 2002; 346: 1200-6.
- Carle GF, Frank M, Olson MV. Electrophoretic separations of large DNA molecules by periodic inversion of the electric field. Science 1986; 232:65–8.
- Martin JM, Wald ER, Green M. Field inversion gel electrophoresis as a typing system for group A Streptococcus. J Infect Dis 1998; 177:504–7.
- Cheng G, Yu Z, Huang JZ. The cluster bootstrap consistency in generalized estimating equations. J Multivar Anal 2013; 115:33–47.
- Therneau TM. Package 'survival.' 2019. Available at: https://github.com/therneau/ survival. Accessed 15 October 2019.
- Kuttner AG, Krumwiede E. Observations on the epidemiology of streptococcal pharyngitis and the relation of streptococcal carriers to the occurrence of outbreaks. J Clin Invest 1944; 23:139–50.
- Shulman ST, Tanz RR, Dale JB, et al. Seven-year surveillance of North American pediatric group A streptococcal pharyngitis isolates. Clin Infect Dis 2009; 49:78–84.
- Kaplan EL, Wotton JT, Johnson DR. Dynamic epidemiology of group A streptococcal serotypes associated with pharyngitis. Lancet 2001; 658:1334–7.
- Jaggi P, Tanz RR, Beall B, Shulman ST. Age influences the emm type distribution of pediatric group A streptococcal pharyngeal isolates. Pediatr Infect Dis J 2005; 24:1089–92.

- Jaggi P, Dale JB, Chiang E, Beniwal P, Kabat W, Shulman ST. Age-associated differences in prevalence of group A streptococcal type-specific M antibodies in children. Eur J Pediatr 2009; 168:679–83.
- 34. Lancefield RC. Persistence of type-specific antibodies in man following infection with group A streptococci. J Exp Med 1959; 110:271–92.
- Tartof SY, Reis JN, Andrade AN, Ramos RT, Reis MG, Riley LW. Factors associated with group A Streptococcus emm type diversification in a large urban setting in Brazil: a cross-sectional study. BMC Infect Dis 2010; 10:327.
- Dale JB, Smeesters PR, Courtney HS, et al. Structure-based design of broadly protective group A streptococcal M protein-based vaccines. Vaccine 2017; 35:19–26
- Chiappini E, Regoli M, Bonsignori F, et al. Analysis of different recommendations from international guidelines for the management of acute pharyngitis in adults and children. Clin Ther 2011; 33:48–58.
- Halloran ME, Struchiner CJ, Longini IM Jr. Study designs for evaluating different efficacy and effectiveness aspects of vaccines. Am J Epidemiol 1997; 146:789–803.
- Polly SM, Waldman RH, High P, Wittner MK, Dorfman A, Fox EN. Protective studies with a group A streptococcal M protein vaccine. II. Challenge of volunteers after local immunization in the upper respiratory tract. J Infect Dis 1975; 131:217–24.
- D'Alessandri R, Plotkin G, Waldman RH, et al. Protective studies with group a streptococcal M protein vaccine. III. Challenge of volunteers after systemic or intranasal immunization with type 3 or type 12 group A Streptococcus. J Infect Dis 1978; 138:712–8.