

RESEARCH PAPER

Elevated CO₂ has concurrent effects on leaf and grain metabolism but minimal effects on yield in wheat

Guillaume Tcherkez^{1,2}, Sinda Ben Mariem³, Luis Larraya⁴, Jose M. García-Mina⁵, Angel M. Zamarreño⁵, Alberto Paradela⁶, Jing Cui¹, Franz-Werner Badeck⁷, Diego Meza⁸, Fulvia Rizza⁷, James Bunce⁹, Xue Han¹⁰, Sabine Tausz-Posch¹¹, Luigi Cattivelli⁷, Andreas Fangmeier⁸ and Iker Aranjuelo^{3,*}

¹ Research School of Biology, ANU Joint College of Sciences, Australian National University, Canberra, ACT 2601, Australia

² Institut de Recherche en Horticulture et Semences, INRA d'Angers, Université d'Angers, Structure Fédérative de Recherche QUASAV, 42 rue Georges Morel, 49071 Beaucouzé, France

³ AgroBiotechnology Institute (IdAB), CSIC-Government of Navarre, Av. Pamplona 123, 31006 Mutilva, Spain

⁴ Institute for Multidisciplinary Applied Biology, Departamento de Agronomía, Biotecnología y Alimentación, Universidad Pública de Navarra, Campus Arrosadia, 31006 Pamplona, Spain

⁵ Facultades de Ciencias y Farmacia y Nutrición, Grupo de Biología y Química Agrícola (Departamento de Biología Ambiental), Universidad de Navarra, C/ Irunlarrea 1, 31008 Pamplona, Spain

⁶ Centro Nacional de Biotecnología-CSIC, C/ Darwin 3, 28049 Madrid, Spain

⁷ CREA Research Centre for Genomics and Bioinformatics, via San Protaso 302, 29017 Fiorenzuola d'Arda, Italy

⁸ Institute of Landscape and Plant Ecology, University of Hohenheim, August-von-Hartmann-Str. 3, 70599 Stuttgart, Germany

⁹ Adaptive Cropping Systems Lab, Beltsville Agricultural Research Center, Agricultural Research Service, US Department of Agriculture, Beltsville, MD 20705, USA

¹⁰ Institute of Environment and sustainable Development in Agriculture, Chinese Academy of Agricultural Sciences (IEDA, CAAS), 12 Zhongguancun South Street, Haidian District, Beijing 100081, China

¹¹ Department of Agriculture, Science and the Environment, School of Health, Medical and Applied Sciences, CQUniversity Australia, Kawana, QLD 4701, Australia

* Correspondence: iker.aranjuelo@csic.es

Received 15 April 2020; Editorial decision 9 July 2020; Accepted 14 July 2020

Editor: Robert Sharwood, Australian National University, Australia

Abstract

While the general effect of CO₂ enrichment on photosynthesis, stomatal conductance, N content, and yield has been documented, there is still some uncertainty as to whether there are interactive effects between CO₂ enrichment and other factors, such as temperature, geographical location, water availability, and cultivar. In addition, the metabolic coordination between leaves and grains, which is crucial for crop responsiveness to elevated CO₂, has never been examined closely. Here, we address these two aspects by multi-level analyses of data from several free-air CO₂ enrichment experiments conducted in five different countries. There was little effect of elevated CO₂ on yield (except in the USA), likely due to photosynthetic capacity acclimation, as reflected by protein profiles. In addition, there was a significant decrease in leaf amino acids (threonine) and macroelements (e.g. K) at elevated CO₂, while other elements, such as Mg or S, increased. Despite the non-significant effect of CO₂ enrichment on yield, grains appeared to be significantly depleted in N (as expected), but also in threonine, the S-containing amino acid methionine, and Mg. Overall, our results suggest a strong detrimental effect of CO₂ enrichment on nutrient availability and remobilization from leaves to grains.

Keywords: Climate change, free-air CO₂ enrichment (FACE), multiple locations, N/C metabolism, physiology, varieties, wheat.

Introduction

Wheat represents 27% of global grain production (FAOSTAT, 2018: <http://www.fao.org/faostat/en/#home>) and, along with corn and rice, is a major component of the human diet because of its high nutritional value: it is a source of carbohydrates, macroelements and micro (trace) elements, protein, and free amino acids. Bread wheat (*Triticum aestivum* L.) constitutes up to 95% of global wheat production, while durum wheat (*Triticum turgidum* ssp. *durum*) makes up the remainder (He *et al.*, 2013). While bread wheat is cultivated in all parts of the world (Reynolds *et al.*, 2012), durum wheat is mostly grown in the Mediterranean region (Oliveira *et al.*, 2012). Like other crops, wheat cultivation will face serious challenges in the coming decades, with a need to increase production despite challenging environmental conditions caused by climate change. Doubling production by 2050 to meet the anticipated demand is expected to be difficult, since yield would have to increase by 2.4% per year globally whereas the actual rate of increase is only 1.3% per year and, furthermore, ~39% of wheat-cultivating areas have shown no increase in yield in a decade (Ray *et al.*, 2012). Among important questions that must be answered to understand how wheat yield responds to changing climatic conditions is the potential impact of rising atmospheric CO₂.

Atmospheric CO₂ has progressively increased since the beginning of the industrial revolution, and the CO₂ mole fraction is anticipated to double (the predicted mole fraction ranges between 730 and 1020 ppm) by 2100 (Meehl *et al.*, 2007). Since photosynthetic activity in C₃ plants is CO₂-limited, it was predicted that the increase in atmospheric CO₂ would stimulate photosynthesis and thus plant growth (Bowes, 1993). However, crop response to elevated CO₂ varies widely, depending on growth (cultivation) conditions and inter- and intra-specific variability. While a CO₂-driven increase in yield is generally observed, there is substantial variation, with sometimes no yield gain at all (Ainsworth and Long, 2005; Högy and Fangmeier, 2008). This effect mostly comes from photosynthetic acclimation to high CO₂, which in turn disfavors grain production. In wheat, cultivation under free-air atmospheric CO₂ enrichment (FACE) leads, on average, to a yield enhancement of only 10% (Kimball, 2010, 2016). Photosynthesis acclimates to elevated CO₂ via lower average stomatal conductance (g_c), lower photosynthetic capacity (maximum carboxylation rate V_{cmax}), and/or lower ribulose 1,5-bisphosphate regeneration ability (Miglietta *et al.*, 1996; Adam *et al.*, 2000; Rogers and Humphries, 2000; Wechsung *et al.*, 2000; Zhang *et al.*, 2009). Of course, wheat response to elevated CO₂ also depends on water and N supply. In general, elevated CO₂ leads to a higher relative increase in wheat grain yield under water-restricted conditions (up to 20%) compared with rain-fed plants (up to 10%), although yield absolute values are, as expected, lower under water restriction (Kimball *et al.*, 1995; Hunsaker *et al.*, 1996; Tubiello *et al.*, 1999). In addition, the extent to which yield is stimulated by elevated CO₂ under low water input depends on the sowing date and whether additional irrigation is implemented under control (well-watered) conditions (O'Leary *et al.*, 2015). Unsurprisingly, minimal effects of elevated CO₂ on photosynthesis and yield are observed under N-limited conditions (Li

et al., 2000; Grant *et al.*, 2001; Kimball *et al.*, 2001a; Pacholski *et al.*, 2015; Walker *et al.*, 2017; Manderscheid *et al.*, 2018). Furthermore, elevated CO₂ triggers developmental changes that are detrimental to yield. In particular, wheat cultivated under FACE has a lower shoot-to-root ratio (Wechsung *et al.*, 1995; Pacholski *et al.*, 2015) and perhaps an increased root exudation rate (thereby increasing organic matter deposition in soil and thus soil respiration) (Pendall *et al.*, 2001) and accelerated flag leaf senescence (Zhu *et al.*, 2009). By contrast, elevated CO₂ causes an increase in tiller number (Yang *et al.*, 2007a, b) and has a marginal effect on transpiration, making plants slightly more water-efficient, especially under high N supply (Hunsaker *et al.*, 1996; Erbs *et al.*, 2009; O'Leary *et al.*, 2015; Kimball, 2016; Manderscheid *et al.*, 2018).

In addition to such quantitative changes in crop production and photosynthesis, elevated CO₂ leads to modifications in leaf biochemical composition and grain quality. Under elevated CO₂, wheat leaves invest more C in cellulose and flavonoids (Akin *et al.*, 1995; Peñuelas *et al.*, 2000) and are generally less N-rich, with a higher C/N ratio (Zhu *et al.*, 2009; Aranjuelo *et al.*, 2015) and lower amounts of proteins (or transcripts encoding them) involved in photosynthesis (Calvin cycle enzymes) (Nie *et al.*, 1995; Adam *et al.*, 2000; Zhang *et al.*, 2009; Pandey *et al.*, 2017). Proteomics conducted on leaves sampled during anthesis in wheat cultivated in a CO₂-enriched greenhouse (at 700 $\mu\text{mol mol}^{-1}$ CO₂) have shown that the Rubisco content did not increase, while there was a slightly higher content of Rubisco activase (Aranjuelo *et al.*, 2015). When grown under FACE, leaves have been found to contain less Rubisco and more mitochondrial malate dehydrogenase (NAD-dependent), suggesting a change in respiratory metabolism (respiratory CO₂ loss) (Pandey *et al.*, 2017). The detrimental effect of CO₂ on the photosynthetic machinery of flag leaves is accompanied by an increase of ear photosynthesis and a faster decline in flag leaf N due to earlier senescence (Sicher and Bunce, 1998; Zhu *et al.*, 2009). Taken as a whole, elevated CO₂ has a major effect on leaf N metabolism, which in turn impacts on remobilization and thus grain filling during maturation.

Unsurprisingly therefore, wheat grains (and flour produced from them) produced at elevated CO₂ have a generally lower nutritional quality, with lower N (and S) content, less protein, and more starch and fibres (Kimball *et al.*, 2001a, b; Högy *et al.*, 2013; Wroblewitz *et al.*, 2013). We have recently shown that when wheat plants are grown in the greenhouse at elevated CO₂ under ample N supply, grains are less N-rich and do not have the same kinetics of gliadin and glutenin accumulation (Soba *et al.*, 2019). In this case, it has been suggested that this effect did not come from deficient N remobilization in leaves but rather a N dilution effect, whereby the quantity (or size) of grains increased while the total available N for remobilization did not change. In addition, under FACE conditions, an increase in the glutenin-to-gliadin ratio has been found (Pandey *et al.*, 2017), as well as a general decrease in all storage proteins except for globulins and albumins (Wieser *et al.*, 2008, but see Verrillo *et al.*, 2017, where globulins also decreased). Metabolic analyses

have demonstrated that at elevated CO₂, grains contain more secondary metabolites (shikimate and quinate) and free hexoses (fructose and glucose) but less sucrose and alanine, regardless of developmental stage. At maturity, grains are generally depleted in free amino acids, except for the branched-chain amino acids valine, leucine, and isoleucine, which are significantly more abundant (Wroblewitz *et al.*, 2013; Soba *et al.*, 2019). Furthermore, it has been suggested that the metabolism of lysine recycling changes and favours the saccharopine pathway over pipecolate (Soba *et al.*, 2019). It is also worth noting that elevated CO₂ causes a decline in the content of several microelements in wheat grains, such as Fe and Zn (Wroblewitz *et al.*, 2013; Pandey *et al.*, 2017; Beleggia *et al.*, 2018), further impacting on nutritional quality.

Nevertheless, the effect of elevated CO₂ likely depends not only on the cultivation conditions (water and N supply, time of sowing, and temperature), but also on soil properties and the species and cultivar of wheat. For example, wheat lines of different ploidy do not respond similarly to elevated CO₂ in FACE experiments, with hexaploid bread wheat being the most responsive in terms of yield (but the least in terms of photosynthesis) (Uprety *et al.*, 2009). There is also substantial variability between cultivars, from no response at all to significant response to CO₂, with little correlation between yield and photosynthesis, but significant correlation with N content and specific leaf area (Thilakarathne *et al.*, 2013, 2015) or transpiration efficiency (Tausz-Posch *et al.*, 2013b; but see also Bourgault *et al.*, 2013). The impact of elevated CO₂ on both yield and elemental contents has been found to depend on the cultivar (Fernando *et al.*, 2014; Fares *et al.*, 2016; Houshmandfar *et al.*, 2016; Beleggia *et al.*, 2018). Variability in the response to elevated CO₂ between wheat cultivars likely results from differences in N and chlorophyll content, transpiration efficiency, or tillering capacity (Tausz-Posch *et al.*, 2013a; Veres *et al.*, 2017). Modelling has also suggested that variation in the response of wheat lines to elevated CO₂ can result from differences in development and allocation (sink/source relationships) (Wolf *et al.*, 2002). In addition, it is probable that the response to elevated CO₂ may vary when soil properties change. This aspect is less documented but, in principle, the spatial variability of wheat yield in the field under ambient CO₂ has been shown to be related to soil P or organic C content, soil structure, or the proportion of carbonates (Miller *et al.*, 1988; Bhatti *et al.*, 1991; Moulin *et al.*, 1994). Since in wheat there is a relationship between root deposition stimulated by elevated CO₂, root-driven acidification, and cation mobility (Cheng *et al.*, 2010), the effect of elevated CO₂ could be modulated by soil Na, Ca, and K content and pH.

Taken as a whole, whenever there are differences among growth conditions and cultivars, it is difficult to understand the physiological mechanisms underlying the response to elevated CO₂. In fact, it could well be that FACE experiments conducted separately in different countries with different cultivars are not comparable, not only because soil and climate conditions vary but also because the response to CO₂ is cultivar-specific. To tackle this issue, we conducted a targeted metabolic analysis of samples collected during FACE experiments (at

550 μmol mol⁻¹ CO₂) conducted in different countries (USA, Australia, Germany, China, and Italy) with different species (*T. aestivum* and *T. turgidum* ssp. *durum*) and cultivars (Table 1; Supplementary Fig. S1). We carried out elemental analyses (macroelements and some microelements), quantitation of amino acids and sugars, and targeted protein analysis in leaves and grains. Our objective was to assess whether a significant effect of elevated CO₂ could be observed in both leaves and grains across countries and wheat lines and, by doing so, to look at the potential effect of leaf-to-grain nutrient remobilization on CO₂ responsiveness. Our data could also be exploited further by exploring the potential relationship between yield (treated as a quantitative response variable) and grain biochemical composition. Our working hypothesis was that (i) detrimental effects of elevated CO₂ on photosynthesis and N assimilation were associated with significant changes in protein and amino acid composition in leaves, and thus (ii) the amino acid composition of grains was affected, reflecting alteration of both the provision and recycling of nitrogenous compounds; and (iii) there might be a relationship between grain metabolic features and yield.

Materials and methods

Plant material and experimental design

The experiment was conducted with durum wheat (cultivars Miradoux, Duramant, Claudio, and Simeto) and bread wheat (cultivars Janz, Kite, Norin, Triumph, Milan, and PRL). The study involved five FACE facilities located in Stuttgart-Hohenheim (Germany), Fiorenzuola (Italy), Changping-Beijing (China), Horsham (Australia), and Beltsville (USA), in which plants were exposed to ambient (400 ppm) and elevated (550 ppm) CO₂ conditions. FACE facilities included three to five rings (for each CO₂ concentration condition) with a diameter of 12–14 m each or (in Stuttgart-Hohenheim) 2×2 m square plots. In all cases, CO₂ enrichment was performed from sunrise to sunset (in Australia and Italy) or 24 h a day (in China, Germany, and the USA) and throughout the entire growing season. The principle of operation and the performance of the FACE systems used here have been described previously (Mollah *et al.*, 2009; Fangmeier *et al.*, 2016; Fares *et al.*, 2016). When plants reached the stem elongation (Z31) and anthesis half-way (Z65) stages (BBCH), sampling and gas-exchange analyses were carried out in the last expanded leaves (Z31) or flag leaves (Z65). Collected leaf samples were immediately frozen in liquid N₂ and stored at -80 °C for later analyses. Finally, at maturity stage (Z90), grain samples were collected for agronomic and metabolic analyses.

Gas exchange determinations

Analyses were carried out in healthy expanded flag leaves grown under ambient or elevated CO₂ conditions. The light-saturated rate of CO₂ assimilation (A_{sat}), stomatal conductance (g_s), and intercellular CO₂ concentration (c_i) were estimated at a photosynthetic photon flux density of 1500–1600 μmol m⁻² s⁻¹ using equations developed by von Caemmerer and Farquhar (1981). The vapour pressure deficit was 1.5 kPa. CO₂ response curves were constructed by a series of measurements whereby photosynthesis was determined first under standard conditions (400 μmol mol⁻¹), then at low CO₂, then back to standard conditions, and then at high CO₂; overall, the sequence of CO₂ concentrations in the reference channel was 400, 200, 100, 50, 200, 400, 600, 750, and 950 μmol mol⁻¹. Estimations of the maximum carboxylation velocity of Rubisco (V_{cmax}) and the maximum electron transport rate contributing to ribulose 1,5-bisphosphate regeneration (J_{max}) (measurements made in Germany) were made using the method of Harley *et al.* (1992).

Mineral composition

C and N concentration (%) analyses were determined using an elemental analyzer (EA, Carlo Erba Strumentazione, Milan, Italy). Micronutrient and macronutrient concentrations were determined by inductively coupled plasma/optical emission spectrometry (ICP/OES, iCAP 6500 Duo, Thermo Fisher Scientific, Waltham, MA, USA).

C isotope discrimination

Carbon isotope composition was determined using an elemental analyzer (EA1108; Carlo Erba Strumentazione, Milan, Italy) coupled to an isotope ratio mass spectrometer (Delta C; Finnigan MAT, Bremen, Germany). Values were expressed as delta values $\delta^{13}\text{C} = (R_{\text{sample}}/R_{\text{standard}}) - 1$ and expressed in ‰, where R_{standard} is the ^{13}C -to- ^{12}C ratio of the international standard (V-PDB). The carbon isotope discrimination (Δ) was calculated as follows:

$$\Delta (\text{‰}) = (\delta^{13}\text{C}_{\text{air}} - \delta^{13}\text{C}_{\text{sample}}) / (1 + \delta^{13}\text{C}_{\text{sample}}) \quad (1)$$

where $\delta^{13}\text{C}_{\text{air}}$ (‰) is the isotope ratio in atmospheric CO₂. The addition of (industrial) CO₂ in the course of the FACE experiments was such that $\delta^{13}\text{C}_{\text{air}}$ was lower than the natural ^{13}C abundance in the atmosphere (near -8‰). The $\delta^{13}\text{C}$ value of the added CO₂ was known (for the experiment in Germany) or calculated from mass balance, taking advantage of the known contribution of added CO₂ to total CO₂: $c = (550 - 400) / 550 = 27.2\%$. In fact, if the isotope fractionation under ambient CO₂ and under FACE conditions was the same, then the proportion of C coming from the added CO₂ could be calculated as $c = (\delta_{\text{OM,amb}} - \delta_{\text{OM,FACE}}) / (\delta_{\text{CO}_2,\text{amb}} - \delta_{\text{CO}_2,\text{added}})$, where OM refers to organic matter and amb refers to ambient conditions, and $\delta^{13}\text{C}$ values are abbreviated to δ to simplify the notation. $\delta_{\text{OM,FACE}}$ is the $\delta^{13}\text{C}$ value of organic matter under FACE conditions if the fractionation had not changed compared with ambient conditions. In practice, the isotope fractionation could have varied, and thus (neglecting denominators): $\delta_{\text{OM,FACE}} = \delta_{\text{OM,FACE}} + \Delta_{\text{FACE}} - \Delta_{\text{amb}}$, where Δ_{FACE} and Δ_{amb} are the net photosynthetic fractionation under FACE and ambient conditions, respectively. These equations can be combined to give $c = (\delta_{\text{OM,amb}} - \delta_{\text{OM,FACE}}) / (\delta_{\text{CO}_2,\text{amb}} - \delta_{\text{CO}_2,\text{added}}) - (\Delta_{\text{FACE}} - \Delta_{\text{amb}}) / (\delta_{\text{CO}_2,\text{amb}} - \delta_{\text{CO}_2,\text{added}})$. The right term is relatively small since added industrial CO₂ was naturally ^{13}C depleted and thus $\delta_{\text{CO}_2,\text{amb}} - \delta_{\text{CO}_2,\text{added}}$ is much larger than $\Delta_{\text{FACE}} - \Delta_{\text{amb}}$. That is, by neglecting the right term, $\delta_{\text{CO}_2,\text{added}}$ can be estimated (and therefore Δ_{FACE} with equation 1) by imposing $c = 0.272$. Note that such an approximation was not critical, since even under the assumption $(\Delta_{\text{FACE}} - \Delta_{\text{amb}}) / (\delta_{\text{CO}_2,\text{amb}} - \delta_{\text{CO}_2,\text{added}})$ represented 0.10 (i.e. 40% error in c); this would ultimately cause an error of only 1‰ in Δ_{FACE} .

Abscisic acid content

Extraction, purification, and quantification of abscisic acid (ABA) were carried out as described by Torres *et al.* (2018), using a high-resolution mass spectrometry (HPLC-ESI-HRMS) system, with some modifications: freeze-dried material (15 mg) was used instead of frozen powdered material (0.1 g), and the residue obtained after final evaporation (SpeedVac) was redissolved in 0.25 ml methanol instead of 0.5 ml.

Soluble sugar and starch content

Sucrose, glucose, and fructose content were determined using a Beckman P/ACE5500 capillary electrophoresis system (Beckman Instruments, Fullerton, CA, USA), following the method of Cabrerizo *et al.* (2001). Starch content in the pellet was determined according to Ethier and Livingston (2004).

Amino acid content

Amino acids were derivatized at 22–25 °C for 12–16 h with 1 mM fluorescein isothiocyanate dissolved in 20 mM acetone/borate, pH 10. The content of single amino acids was determined by capillary electrophoresis in a Beckman-Coulter PA-800 system.

Protein content

Samples were previously quantified by microBCA analysis (Pierce) and similar amounts (5 µg per sample) were individually dissolved in 8 M urea, 25 mM ammonium bicarbonate, reduced with DTT, and alkylated with iodoacetamide, according to a method described by López-Ferrer *et al.* (2004). Digested samples were diluted with 0.2% trifluoroacetic acid in water and subjected to multiple reaction monitoring analysis using a 1D Plus nanoLC Ultra system (Eksigent, Dublin, CA, USA) interfaced to a Sciex 5500 QTRAP triple quadrupole mass spectrometer (Sciex, Framingham, MA, USA) equipped with a nano-electrospray ionization source and controlled by Analyst v.1.5.2. software (ABSciex). Trypsin-digested samples were loaded online on a C18 PepMap 300 µm internal diameter × 5 mm trapping column (5 µm, 100 Å, Thermo Scientific) and separated using a BioSphere C18 75 µm internal diameter × 150 mm capillary column (3 µm, 120 Å, Nanoseparations). A list of 84 transitions (usually 3–4 per peptide, with a preference toward higher-mass y series ions), corresponding to 21 unique peptides selected for 10 different proteins, was monitored. Skyline software determined automatically the collision energy values for the candidate peptides according to MacLean *et al.* (2010).

Statistical analysis

To explore the effect of CO₂, univariate statistics were conducted with a two-way ANOVA, with one factor representing ‘conditions’ (time of sampling, country, and species/cultivar) while the second factor was CO₂. It was not possible to carry out a three-way analysis because not all species/cultivars were represented in all countries. Thus, here, the two-way ANOVA was constructed with the factor ‘conditions’ having 12 possible qualitative values (combinations time–country–cultivar) and the factor ‘CO₂’ having two values (ambient or elevated). Unless otherwise stated, statistical significance was accepted when $P < 0.05$. Multivariate statistics were carried out using orthogonal projection on latent structure (OPLS; with Simca®, Umetrics), using CO₂ as the predicted Y variable (while ‘conditions’ are embedded into orthogonal dimensions), and metabolic features (elemental contents, metabolites, and proteins) as predicting X variables. Before running the OPLS, a principal component analysis was conducted to check the presence of outliers (samples outside the Hotelling’s ellipse). The performance of the OPLS was assessed using the correlation coefficient between predicted and observed Y (R^2), the cross-validated correlation coefficient (Q^2), the Q^2 intercept of the permutation test (which was checked to be negative), and the P -value of testing the OPLS model against a random-error model (i.e. average \pm error) via a χ^2 test (this P -value is referred to as $P_{\text{CV-ANOVA}}$). Univariate and multivariate analyses were combined using $-\log(P\text{-value})$ (univariate) plotted against the OPLS loading (p_{corr}) in a volcano plot. To explore the relationship between yield and grain composition, we conducted (i) an OPLS using yield as the predicted Y variable (with the same parameters of performance as described above) and (ii) a univariate analysis by linear regression (done in R). Linear regression was done without and with variable elimination. Without variable elimination, each variable was correlated with yield separately and the associated P -value is reported. To carry out variable elimination, we used a multiple linear regression. It was carried out via sampling subsets (regsubsets) of eight variables (304 iterations) and a correlation plot was generated to select the eight most correlated variables overall. Then, a multiple linear regression model (lm) comprising only these eight variables was generated and the P -value for each variable was calculated.

Results

Photosynthetic parameters

The photosynthetic activity of flag leaves was assessed using measurements of gas exchange at various times of the development cycle (shown as BBCH stage numbers), at the CO₂ mole fraction (c_a) used during growth (Fig. 1). Overall, there

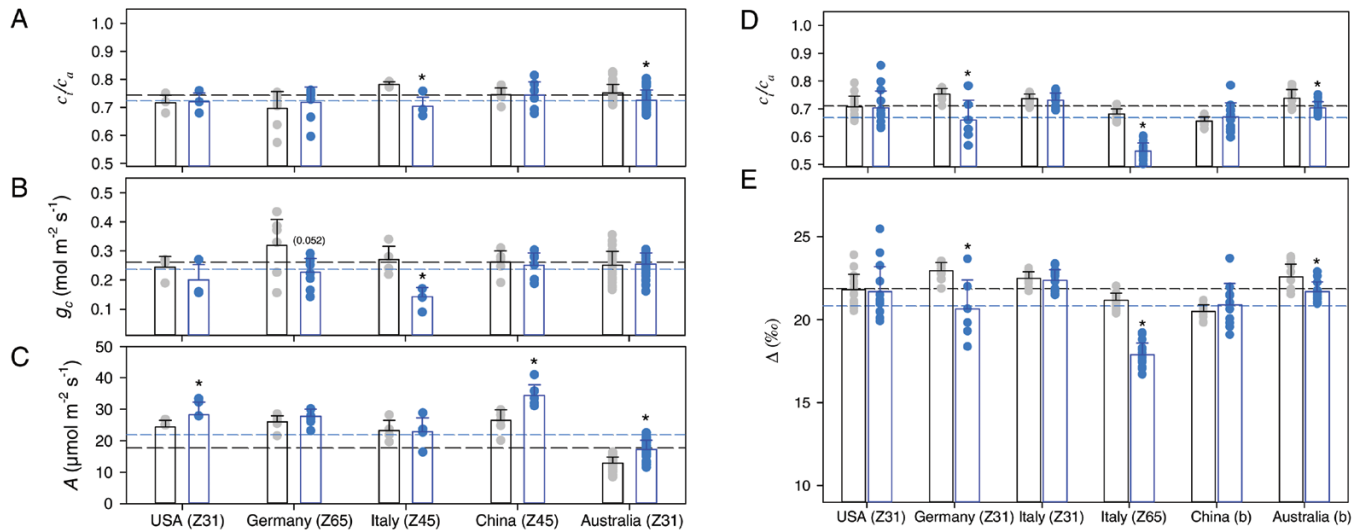


Fig. 1. Photosynthetic parameters in wheat grown under ambient (grey) or elevated ($550 \mu\text{mol mol}^{-1}$, blue) CO_2 mole fraction. (A–C) Leaf gas exchange properties: intercellular-to-atmospheric CO_2 ratio (c_i/c_a) (A), stomatal conductance for CO_2 (B), and net photosynthesis (C). (D, E) $^{12}\text{C}/^{13}\text{C}$ carbon isotope fractionation (Δ) measured using leaf total organic matter $\delta^{13}\text{C}$ (E) and calculated average c_i/c_a values (E). Asterisks indicate significant differences ($P < 0.05$) between ambient and elevated CO_2 . P -values very close to statistical significance are given in parentheses. Dashed lines show average values across all countries. The developmental stage (BBCH) for plants in each country is given in parentheses; for China and Australia, values obtained at both BBCH stages 31 and 65 were pooled together (indicated by b). ND, not determined. (This figure is available in colour at *JXB* online.)

was little variation in the photosynthetic rate, which was $\sim 22 \mu\text{mol m}^{-2} \text{s}^{-1}$ regardless of CO_2 , with the following properties (Fig. 1C): (i) in China, the USA, and Australia, there was a significantly higher photosynthetic rate at elevated CO_2 , and (ii) in Australia, plants generally had lower photosynthetic rates compared with other countries. These effects were unrelated to stomatal conductance (Fig. 1B), which was roughly constant at $0.25 \text{ mol m}^{-2} \text{ s}^{-1}$; however, CO_2 had a significant depressing effect in plants grown in Italy. As a result, there was little variation in the intercellular-to-external CO_2 ratio (c_i/c_a) (Fig. 1A), which was always ~ 0.75 , except in Italy ($c_i/c_a < 0.7$) and in Australia ($c_i/c_a \approx 0.7$) at elevated CO_2 . The natural carbon isotope abundance ($\delta^{13}\text{C}$) in raw leaf matter was measured, and ‘average’ c_i/c_a (i.e. across the leaf lifespan) was estimated using the simplified equation of Farquhar *et al.* (1982) (Fig. 1D, E). The $^{12}\text{C}/^{13}\text{C}$ fractionation (raw matter versus source CO_2) was within 20–22‰, showing that average c_i/c_a was always ~ 0.7 . However, the isotope fractionation was significantly lower (and therefore so was average c_i/c_a) in Germany, Italy, and Australia at elevated CO_2 , suggesting a long-term decrease in stomatal (or internal) conductance and/or an increase in photosynthetic capacity at elevated CO_2 . In addition, there were no significant changes either between CO_2 conditions or among countries in ABA content (data not shown).

Leaf metabolism

The contents of amino acids, sugars, macro- and microelements, and some target proteins were analyzed in flag leaves (contents were expressed relative to dry weight). A statistical analysis combining multivariate (OPLS) and univariate (two-way ANOVA) statistics was performed and results showing the effect of CO_2 are represented as a volcano plot in Fig. 2A. In both the OPLS and ANOVA, two factors were considered: CO_2 and

‘conditions’ (comprising country, species/cultivar, and growth stage). Growth at elevated CO_2 led to an increased content of several elements (Mo, Mg, Ca, Cr, Ti, Fe, and S) and a decrease in K. Several enzymes [superoxide dismutase, Rubisco (large subunit), carbonic anhydrase, pyruvate dehydrogenase, and nitrate reductase] had a lower content at elevated CO_2 , and so had threonine. There was a tendency to have more fructose and less sucrose at elevated CO_2 (significant changes in ANOVA) but OPLS loadings were rather small, showing that the change in sugar composition was not a strong marker of growth at elevated CO_2 . When expressed as a percentage of total amino acids and proteins, glycine and glycine decarboxylase (GDH subunit h) were lower ($P < 0.001$) under elevated CO_2 (Fig. 2B), likely reflecting lower photorespiration activity. When examined separately for each country, the fructose-to-sucrose ratio appeared to be significantly different in Germany and Italy (Fig. 2C), and threonine content appeared to be significantly different in Italy and China (Fig. 2D), showing that the effect of elevated CO_2 was country-specific (i.e. it depended on environmental local conditions). This phenomenon was clearly visible in the results of the two-way ANOVA, where several features appeared to be significant for the $\text{CO}_2 \times$ conditions effect (Supplementary Fig. S2). For example, under elevated CO_2 , there was a notably lower content of amino acids in Australia, nearly opposite effects on starch content in the USA and Australia, and a strong decrease in the content of several microelements in China. Nevertheless, the patterns of mineral elements were the same across all countries and CO_2 conditions. In fact, there was as expected an antagonism between major cations, with $\text{Ca} + \text{Na}$ or Mg being negatively correlated with K (Fig. 2E). However, there was little difference in this correlation between ambient and elevated CO_2 . In other words, the significant decrease in K and increase in Ca and Mg at elevated CO_2 (Fig. 2A) followed the same cation balance relationship as under ambient CO_2 .

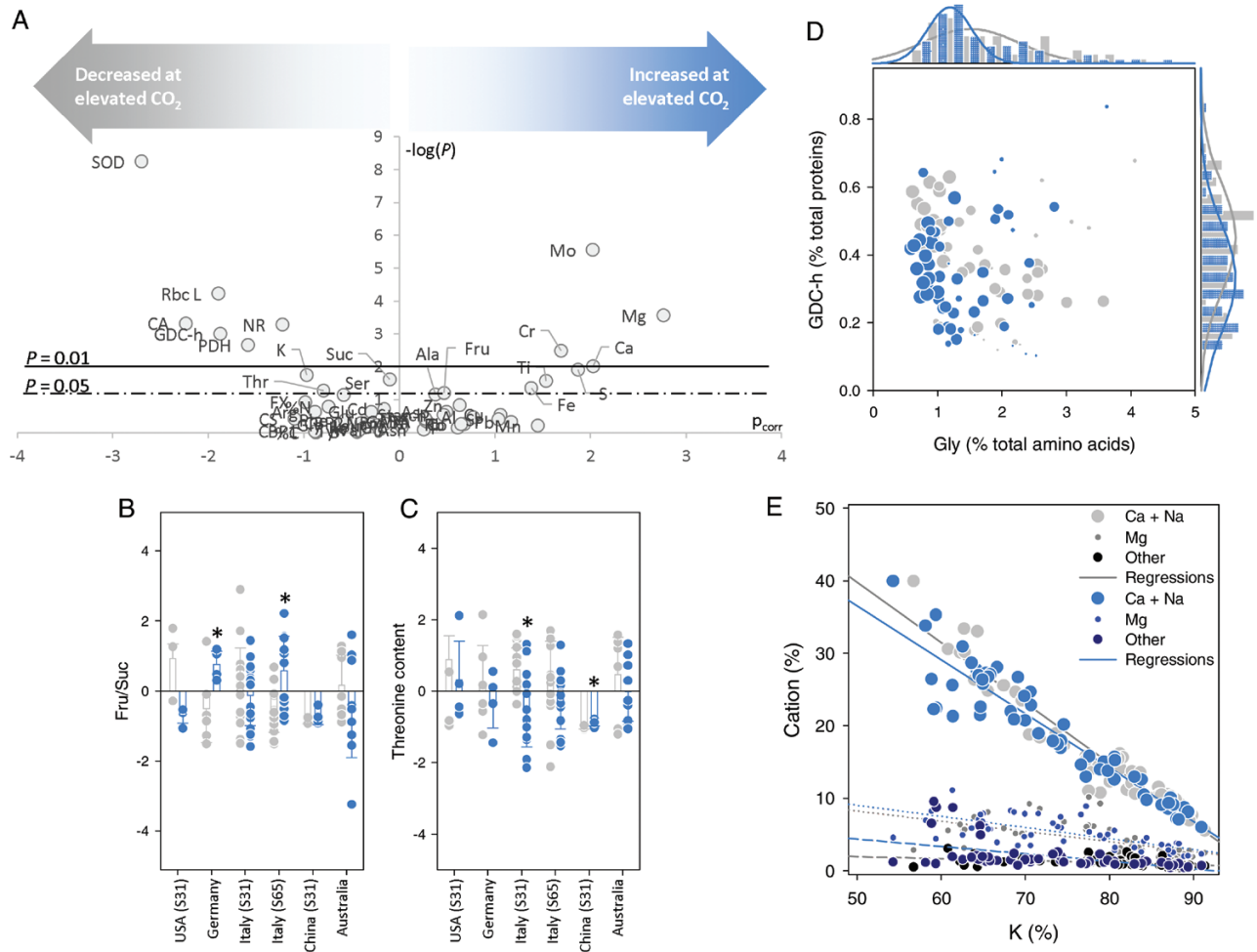


Fig. 2. Metabolism of wheat leaves in plants grown under ambient (grey) or elevated ($550 \mu\text{mol mol}^{-1}$, blue) CO₂ mole fraction. (A) Volcano plot [$-\log(P)$ -value] from ANOVA versus the loading p_{corr} from OPLS] showing the best discriminating components (metabolites, proteins, and elements) associated with the effect of CO₂ enrichment. Threshold P -values (0.01 and 0.05) are shown with horizontal solid and dashed lines. (B, C) Relative fructose-to-sucrose ratio (B) and relative threonine content (C). In these panels, cultivars are pooled together for each country. Asterisks indicate significant differences ($P < 0.05$) between the different average values. (D) Distribution of data points in the bi-plot showing the relative content of glycine decarboxylase h subunit (GDC-h) and glycine (Gly) (as a percentage of total proteins and total amino acids, respectively), with frequency plots on each axis. Countries are represented by different symbol sizes. (E) Relationship between K content and other cations under elevated and ambient CO₂, with linear regressions (all significant, $P < 0.05$). (This figure is available in colour at JXB online.)

Grain metabolism

The contents of free amino acids, macro- and microelements, and total proteins and starch were quantified in mature grains (relative to dry weight). As in leaves, an analysis combining univariate (ANOVA) and multivariate (OPLS) statistics was performed and presented as a volcano plot (Fig. 3A). Interestingly, no component appeared to be significantly increased at elevated CO₂. By contrast, there were significant decreases in several elements (N, Mg, and Pb), amino acids (methionine, threonine, alanine, and tyrosine), and the glutamate derivative γ -aminobutyrate (GABA). Although nitrogen content (%N) was found to be lower, there was no significant change in protein content. Such a decrease in N content could be explained by changes in the concentration of many N-containing compounds (e.g. free amino acids, polyamines, or insoluble proteins not accounted for in our assay), which in total sum up to a larger and significant change in overall %N. When plotted together, %N and carbon content (%C) showed considerable scattering

across countries (Fig. 3B), with a difference greater than 1.5% in %N between samples. The effect of elevated CO₂ on alanine and threonine appeared to be condition-specific, with significant differences between conditions found in the USA and Italy (Fig. 3C, D). More generally, the two-way ANOVA showed that amino acids, %N, and Pb were significant for the CO₂ \times conditions effect (Supplementary Fig. S3). Elevated CO₂ did not lead to significant changes in major cations (K, Ca, Na, and Mg) (Fig. 3E). In fact, Ca + Na and Mg were negatively correlated with K, reflecting cation balance, and the correlation was not changed by elevated CO₂; that is, the lower Mg content at elevated CO₂ (Fig. 3A) was not associated with a change in the Mg/K balance in grains.

Relationships with yield

There were considerable differences in grain yield between countries, with low values in the USA ($\approx 200 \text{ g grain m}^{-2}$) and

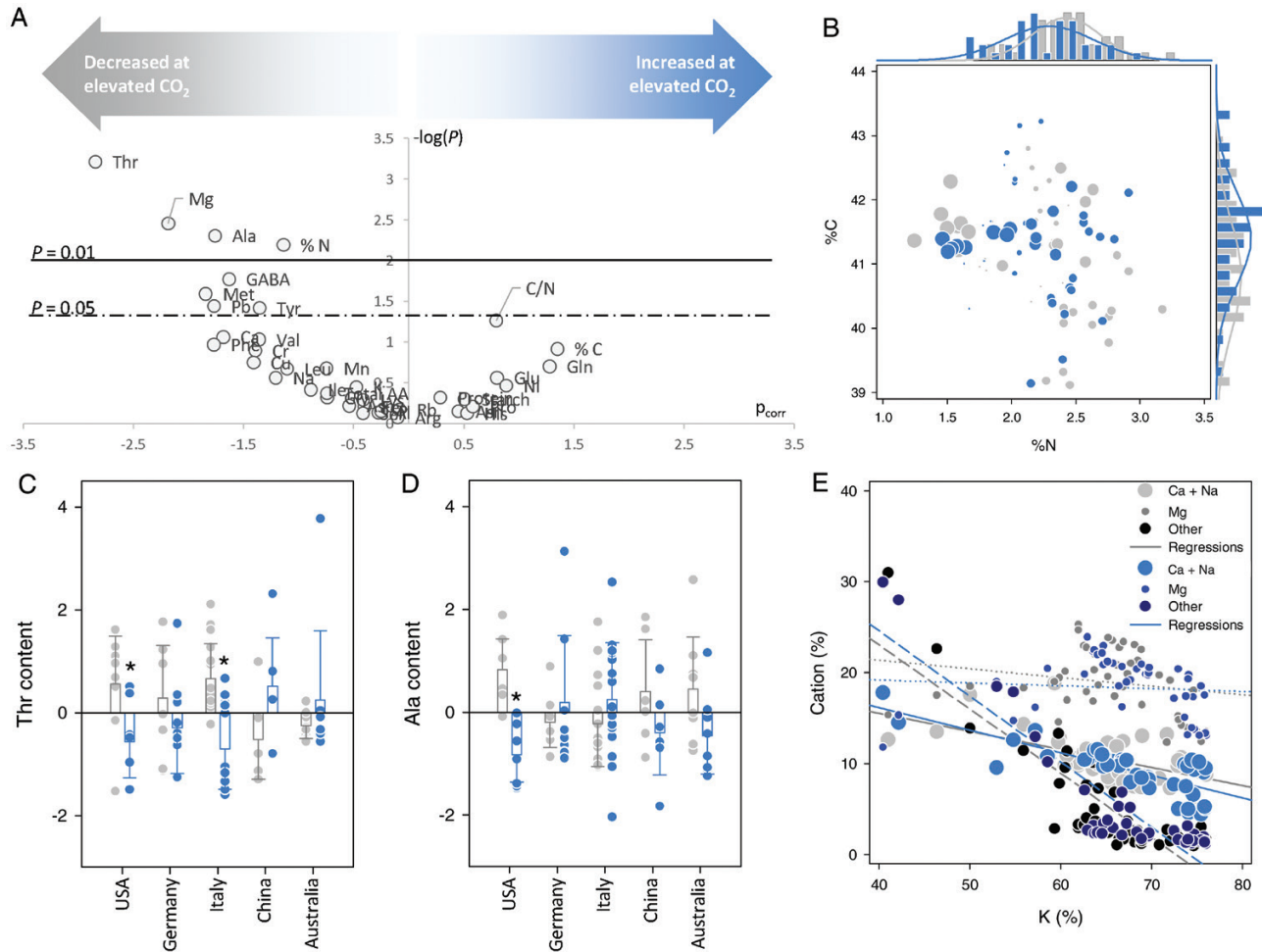


Fig. 3. Metabolism of wheat grains in plants grown under ambient (grey) or elevated ($550 \mu\text{mol mol}^{-1}$, blue) CO_2 mole fraction. (A) Volcano plot [$-\log(P)$ -value) from ANOVA versus the loading p_{corr} from OPLS] showing the best discriminating components (metabolites and elements) associated with the effect of CO_2 enrichment. Threshold P -values (0.01 and 0.05) are shown with horizontal solid and dashed lines. (B) Distribution of data points in the bi-plot showing elemental C and N content, with frequency plots on each axis. Countries are represented by different symbol sizes. (C, D) Relative threonine (C) and alanine (D) content. In these panels, cultivars are pooled together for each country. Asterisks indicate significant differences ($P < 0.05$) between control and elevated CO_2 . (E) Relationship between K content and other cations under elevated and ambient CO_2 , with linear regressions (all significant, $P < 0.05$, except for Mg). (This figure is available in colour at JXB online.)

high values in Australia and Italy ($\approx 600 \text{ g grain m}^{-2}$) (Fig. 4A). The difference in yield between countries was unsurprising and resulted from seasonal differences, as well as differences in climatic conditions, fertilization (Table 1), and soil composition (Supplementary Fig. S1), or sowing density (which ranged within $120\text{--}350 \text{ m}^{-2}$). To gain insight into the metabolic determinants of yield, we conducted both a multivariate OPLS analysis using yield as a predicted quantitative Y variable, and a univariate analysis by linear regression. The OPLS analysis generated a good statistical model ($R^2=0.75$) with good robustness ($Q^2=0.67$) and very high significance ($P_{\text{CV-ANOVA}}=10^{-16}$). When observed and predicted yield were plotted together, it was clear that the relationship was driven partly by differences between countries (Fig. 4B) and not at all by the CO_2 treatment (Fig. 4C). Accordingly, there were no significant CO_2 effects, except in the USA (Fig. 4A). The best drivers of yield were investigated using volcano plots. Two volcano plots are shown here: the first (Fig. 4D) is associated with an analysis that disregards ‘conditions’ (countries and species/cultivars), while the second (Fig. 4E) incorporates ‘conditions’ as a variable. In Fig. 4D,

P -values are associated with a variable-by-variable linear regression or with a multiple linear regression including variable elimination (where variables that are not best correlated with yield are discarded) (Supplementary Fig. S4). Regardless of country and cultivar, the yield appeared to be significantly and positively related to micro- and macroelements (Fe, Al, K, Mg, Cr, and N) and negatively related to starch, threonine, Rb, and %C. After variable elimination upon multiple linear regression, only six features were significantly related to yield: Mg, glutamine, and asparagine (positively related), and Rb, valine, and %C (negatively related). Among these features, some appeared to be significant because of the confounding factor of country of origin (Fig. 4B, C). When this effect was removed by incorporating ‘conditions’ as a variable, the two most significant drivers were glutamine (increased) and valine (decreased) (Fig. 4E). Tyrosine and isoleucine were found to be significant ($P < 0.05$) but their loading value was close to zero, showing that their impact (in multivariate analysis) was numerically very small. When plotted separately, there was a positive relationship between yield and the glutamine-to-valine ratio in grains (Fig. 4F).

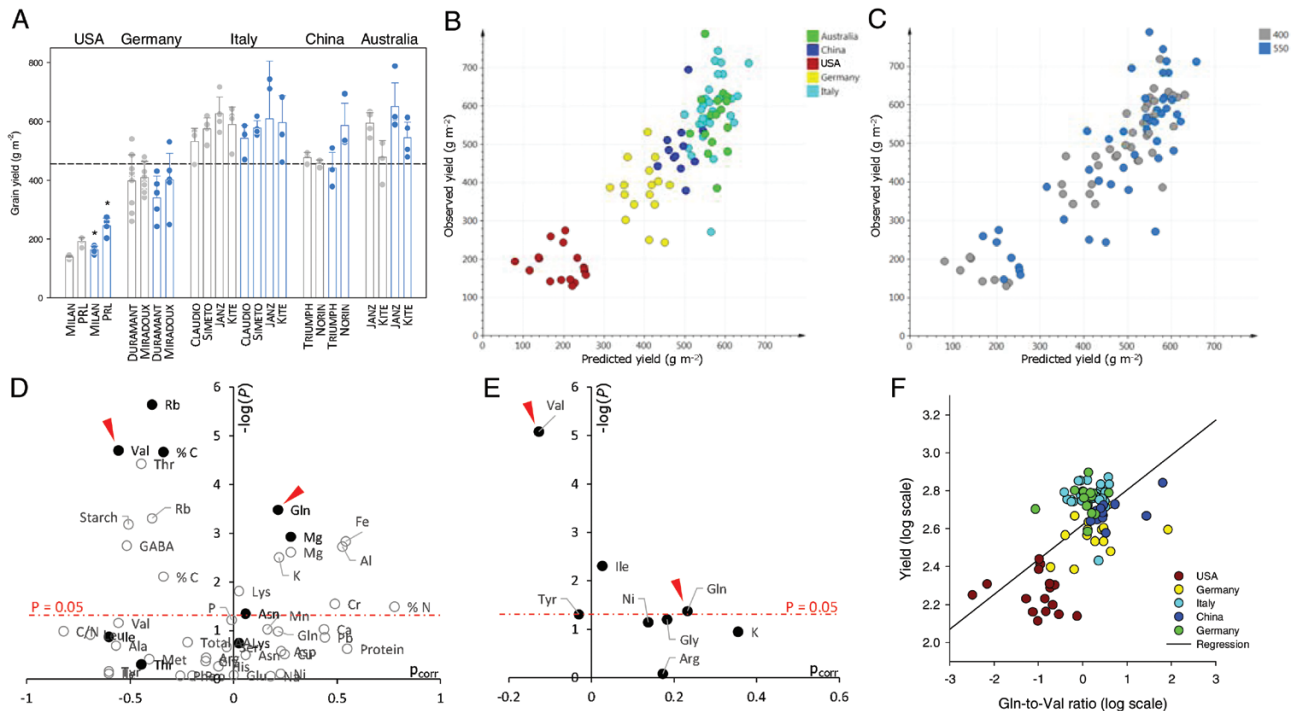


Fig. 4. Yield analysis. (A) Yield (in g grains m⁻²) of the different cultivars and countries. Asterisks indicate significant differences ($P < 0.05$) between control (grey) and elevated (blue) CO₂. (B, C) Relationship between observed yield and yield predicted using the OPLS model (comprising country+cultivar as a qualitative X variable), differentiating countries (B) or CO₂ levels (C) ($R^2 = 0.75$). (D) Volcano plot [$-\log(P)$ -value] versus OPLS loading p_{corr} showing the importance of variables for statistical analysis: P -values from separate linear regressions (each variable taken separately) (white circles), or P -values from variable elimination and linear model (black circles) against OPLS loadings (both univariate and OPLS models without country+cultivar as a qualitative X variable). (E) P -values from variable elimination and linear model against OPLS loadings (both univariate and OPLS models with country+cultivar as a qualitative X variable). In D and E, the horizontal dashed line indicates the P -value threshold of 0.05, and arrowheads indicate valine (Val) and glutamine (Gln). (F) Relationship between observed yield and the glutamine-to-valine ratio (log scales). The solid line represents the linear regression ($R^2 = 0.37$). (This figure is available in colour at *JXB* online.)

Discussion

Here, we used five FACE sites in different countries, with different wheat cultivars/species, to investigate the metabolic effects of elevated CO₂ in leaves and grains. Our results show that elevated CO₂ (i) had a limited effect on photosynthesis rate and grain yield, reflecting photosynthetic acclimation; (ii) caused a decline in proteins involved in photosynthesis, photorespiration, or N assimilation in leaves; and (iii) altered grain quality, with lower contents of amino acids and mineral nutrients. In addition, a quantitative analysis of yield suggested that high yield values correlate with higher glutamine, K, and Mg contents and lower valine content.

Growth at elevated CO₂ had a limited impact on crop yield

Crop yield and photosynthetic responsiveness to elevated CO₂ depend considerably on surrounding (local) environmental conditions. Here, we found that with the exception of the FACE site in the USA, grain yield was not significantly affected by elevated CO₂. In practice, changes in wheat yield in response to high CO₂ are determined by the cultivar(s), fertilization protocols, and interactions with other environmental factors such as water availability and temperature. The absence of an effect of CO₂ was probably linked to the lack of significant changes in assimilation (mechanisms are further

discussed below). Accordingly, several previous studies have shown that the initial stimulation by CO₂ can be compensated for by acclimation (reviewed in Ainsworth and Rogers, 2007). The meta-analyses carried out by Galmés *et al.* (2013) showed that under elevated CO₂ conditions, Rubisco content is the primary driver in the regulation of Rubisco activity and, consequently, photosynthetic activity. Within this context, the absence of an effect of elevated CO₂ on crop yield could be linked to limitations in N assimilation and altered leaf C sink/source balance (Ainsworth and Rogers, 2007). Photosynthetic performance is believed to be affected by two key factors: the CO₂ concentration in the chloroplast (c_c) and the carboxylation capacity, which is linked to leaf N content. Here, the absence of significant differences in stomatal conductance (accompanied by the lack of an effect on leaf ABA content) suggests that stomatal limitation of CO₂ diffusion was a minor component of acclimation under our conditions. The lower content of nitrate reductase (and other proteins) suggests an inhibitory effect of elevated CO₂ on N metabolism, as previously observed in wheat leaves (Jauregui *et al.*, 2015). The inhibition of nitrate assimilation under elevated CO₂ in wheat has been suggested to reflect lower electron allocation to nitrite reductase in the chloroplast and/or the inhibition of the GS/GOGAT cycle driven by photorespiration (Bloom *et al.*, 2014). Here, the lower contents of ferredoxin, chlorophyll binding proteins, and glycine decarboxylase under elevated CO₂ would be consistent with a down-regulation of N assimilation.

Table 1. Growth conditions associated with the CO₂ enrichment experiments in this study

Site general information			Climatic conditions during growth				Fertilization			
Country	Location	Cultivation time window	Species and cultivars	Sowing month	P _{tot} (mm)	T _{av,min} (°C)	T _{av,max} (°C)	Cumulated d·°C	Nitrogen (kg N ha ⁻¹)	Elemental N-P ₂ O ₅ -K ₂ O
USA	Beltsville	16/10/15–7/6/16	<i>T. durum</i> Milan, PRL	October	141	4.2	25.9	981	250	10-10-10
	Fiorenzuola	9/11/15–11/7/16	<i>T. aestivum</i> Janz, Kite;	November	347	10.3	27.8	1774	183	15-15-15
Germany	Hohenheim	5/4/16–1/8/16	<i>T. durum</i> Claudio, Simeto	April	271	2.6	24.8	1679	202	10-5-5
China	Beijing	6/10/15–20/6/16	<i>T. durum</i> Miradoux, Duramant	October	307	10.9	30.8	1843	200	20-16-9
Australia	Horsham	1/06/16–21/12/16	<i>T. aestivum</i> Janz, Kite	June	334	9.0	23.1	2394	50	10-1-10

Soil mineral conditions are further documented in [Supplementary Fig. S1](#). Cumulated d·°C, cumulated temperature during cultivation period (in days·°C); P_{tot}, total precipitation; T_{av,min}, average minimum daily temperature; T_{av,max}, average maximum daily temperature.

Besides the effect of CO₂ (or lack thereof), our study shows important differences in yield between locations and cultivars. The lowest values were found in the USA, where sowing density (120 m⁻²), precipitation, and cumulated day·°C values were low ([Table 1](#)), and seedling mortality was pronounced in the year of the FACE experiment due to bad weather. The high yield values observed in China were probably explained by higher water availability. We note that the FACE site in the USA, where elevated CO₂ had a significant effect on yield, had the highest N fertilization rate (250 kg ha⁻¹) despite low plant sowing density and low cumulated days·°C; this suggests that perhaps the two most important factors for CO₂ responsiveness were available N and intercepted light (minimal shading) under our conditions. Differences between locations could also have originated from the contrasting behaviour of the two species of wheat studied, with bread wheat exhibiting generally higher yield values than durum wheat. This species effect comes from the fact that durum wheat is more water conservative and thus more suitable to grow in stressful environments, while bread wheat is believed to have a higher yield potential ([Slafer et al., 1996](#); [Marti and Slafer, 2014](#)).

Photosynthetic effects of elevated CO₂

Growth at elevated CO₂ generally leads to a down-regulation of some photosynthetic parameters, such as carboxylation efficiency, maximal carboxylation velocity, and CO₂ conductance (see Introduction). Here, photosynthesis increased by a modest but significant amount at elevated CO₂ (in the USA, China, and Australia) or did not increase at all (in Italy and Germany). In Italy, the lack of a stimulating effect of CO₂ mole fraction was associated with a decrease in stomatal conductance ([Fig. 1B](#)). It is also possible that internal conductance decreased under elevated CO₂, and this could contribute to explaining why the ¹²C/¹³C isotope fractionation did not increase much (with apparent c_i/c_a staying at ~0.7 despite the increase in CO₂ mole fraction) ([Fig. 1E](#)). In addition, there was probably a decrease in carboxylation capacity, as suggested by the significantly lower Rubisco content in leaves ([Fig. 2](#)), and A/c_i response curves for wheat grown in Germany suggest a lower V_{cmax} (non-significant) and J_{max} ($P < 0.05$) ([Supplementary Fig. S5](#)). More generally, elevated CO₂ led to a down-regulation of the photosynthetic and photorespiratory machinery, with less carbonic anhydrase, glycine decarboxylase, and superoxide dismutase, an enzyme of redox metabolism (as reported previously in [Aranjuelo et al., 2015](#)). As a result, the lower content of major enzymes such as Rubisco was accompanied by a decline in %N in some countries (the USA, China, and Australia) but not others. The origin of this effect of elevated CO₂ could be lower biosynthesis of proteins or an increase in protein degradation, for example, earlier remobilization of proteins to facilitate export of N from leaves to developing grains ([Sicher and Bunce, 1998](#); [Zhu et al., 2009](#)). Earlier remobilization seems unlikely, since leaves were sampled at stage Z31 (stem elongation with first node above tillering node) or Z65 (anthesis, 50% of anthers mature), that is, before the onset of remobilization. Therefore, it is probable that elevated CO₂ down-regulated N metabolism, causing a general decrease in the biosynthesis of

major proteins and thus potentially photosynthesis. Despite the change in Rubisco content, the net effect on photosynthesis also depends on other enzyme activities, because net assimilation is not highly sensitive to Rubisco content. For example, other enzymes, such as Rubisco activase, are crucial for carboxylation efficiency. We have previously shown using proteomics that elevated CO₂ leads to a decline in Rubisco activase content (Aranjuelo *et al.*, 2015). Here, our analysis focused on a number of specific proteins, which did not include Rubisco activase. Photosynthetic products (sugars) were affected by CO₂ mole fraction, with significantly less sucrose and generally more fructose ($P=0.06$); as a result, the fructose-to-sucrose ratio was larger at elevated CO₂ in two countries (Germany and Italy) (Fig. 2B). In other words, elevated CO₂ was associated with a reconfiguration of sugar metabolism, likely including a larger allocation of carbon to starch and a futile sucrose synthesis–degradation cycle.

Effect of CO₂ on grain composition

We found that elevated CO₂ had a clear effect on grain composition, leading to lower contents of N, amino acids, and microelements (Fig. 3). The general effect of elevated CO₂ on N content (and proteins) in grains has been documented before, and has been suggested to be linked to perturbations in both leaf-to-grain N transport (e.g. glutamine) and amino acid metabolism in grains (e.g. in the lysine degradation pathway; see the Introduction and Soba *et al.*, 2019). Here, it is remarkable that Mg was found to be more abundant in leaves but less abundant in grains, suggesting that elevated CO₂ compromised microelement remobilization (the same occurred with Ca but this was not significant in grains). Nutrient quantitation during grain filling in different cultivars of spring wheat has suggested that Mg is remobilized from the lower stem and leaves to grains, with little effect of cultivar (Tiryakioglu *et al.*, 2014). Growth at elevated CO₂ has been found to affect Mg content in wheat (Sanchez de la Puente *et al.*, 2000; Högy *et al.*, 2013; Aranjuelo *et al.*, 2015; Beleggia *et al.*, 2018). Mg remobilization involves chlorophylls and Rubisco degradation in leaves, and Mg circulation via the phloem. This raises the question of whether phloem transport was affected by elevated CO₂ in the current study. On the one hand, glutamine and glutamate were found to be more abundant in grains at elevated CO₂ (Fig. 3), suggesting that phloem N export from leaves was not affected. On the other hand, leaves contained significantly less K, which is the cornerstone of phloem ion movement. Therefore, our data do not provide a firm answer to this question. Nevertheless, we recognize that the higher glutamine (glutamate) content in grains could also have originated from a lower efficiency of glutamine utilization and thus a lower incorporation of N to anabolism. Grains were specifically depleted in GABA, methionine, tyrosine, alanine, and threonine. Interestingly, these amino acids belong to distinct pathways: aspartate metabolism (methionine and threonine), glutamate degradation/GABA shunt (GABA and alanine), and aromatics (tyrosine). The common branching point is pyruvate, since aspartate can come from anaplerotic phosphoenolpyruvate (PEP) carboxylase fixation (which forms oxaloacetate), alanine comes from pyruvate,

and the biosynthesis of aromatics requires PEP. Therefore, it is possible that the decrease in amino acids was caused by a specific effect of elevated CO₂ on (phospho)enolpyruvate synthesis via glycolysis. For example, pyruvate Pi dikinase, which resynthesizes PEP from pyruvate, has been shown to be essential for starch accumulation at the late grain-filling stage (Prioul *et al.*, 2012).

Importantly, these metabolic effects depended on ‘conditions’, that is, country and species/cultivar (Supplementary Figs S2, S3). It was not possible here to separate the specific contributions of country and species/cultivar, since not all plant lines were cultivated in each country. There was nevertheless a strong effect of location, with the effect of elevated CO₂ on some micro- and macro-elements being country specific. For example, the effect of elevated CO₂ on grain Pb content was largest in Australia, where Pb was the least abundant at the field location (Supplementary Fig. S1). Similarly, there was a very limited effect of CO₂ on grain N content (with generally low values) in Australia (Supplementary Fig. S3), where N fertilization was the lowest (Table 1), while the effect on leaf N content was strong (Supplementary Fig. S2). There was also a strong effect of CO₂ on grain free amino acid content in both Australia and the USA, despite the fact that the sites in the two countries used different fertilization levels and wheat species. Therefore, our results show the importance of growth conditions (i.e. location) to delineate the effects of elevated CO₂; in particular, soil composition (and not only N fertilization) seems to be important. Yet, the decrease in threonine and Mg content in grains was independent of ‘conditions’, that is, it was not associated with a CO₂ × conditions interaction effect.

Potential metabolic markers of yield

We used the dataset to examine possible relationships between yield and grain metabolic properties, using multivariate and univariate statistics (Fig. 4). Because the country of origin (and associated conditions such as climate, fertilization, and sowing density) and species/cultivar (i.e. ‘conditions’) represented a confounding factor, we carried out two types of analyses: (i) independent of and (ii) accounting for ‘conditions’. In doing so, we assumed that the variable ‘conditions’ can be a driver of yield regardless of other variables (e.g. contents of metabolites or micro-/macro-elements). Growth at elevated CO₂ was not a confounding factor, since there was no significant effect of CO₂ on yield overall (Fig. 4A, C). Only two significant and strong drivers of yield remained after the elimination of the ‘conditions’ effect: glutamine and valine (Fig. 4E, F).

This result highlights the well-known role of glutamine in grain protein synthesis: glutamate is a major amino acid in wheat stem phloem (Hayashi and Chino, 1986) used as a primary source of N for conversion to glutamine and, furthermore, glutamine is the most represented residue in accumulated proteins. It has been shown that glutamine accounts for ~40% of free amino acids not only in phloem sap from the spikelet peduncle but also in endosperm cavity sap, showing its crucial role as a N source during grain development (Fisher and Macnicol, 1986). In addition, glutamine has the highest

supply rate from the vascular bundle to the developing endosperm, at $\sim 0.5 \mu\text{mol grain}^{-1} \text{d}^{-1}$ (Ugalde and Jenner, 1990).

Valine was negatively related to yield (Fig. 4E), suggesting that increased valine degradation rather than valine synthesis is beneficial to grain production. Proteomics analyses have shown that acetolactate synthase, which catalyses the first step of valine and leucine biosynthesis, is most abundant at the beginning of grain development (pre-filling stage) and then declines up to maturity (Tasleem-Tahir *et al.*, 2012), while methylmalonate semialdehyde dehydrogenase, which is involved in branched-chain amino acid degradation, appears at the last stage of grain filling (Vensel *et al.*, 2005) and its content increases with ABA (Zhang *et al.*, 2012). Accordingly, metabolic profiling has shown that branched-chain amino acids (valine, leucine and isoleucine) tend to increase during grain filling and then sharply decline (Shewry *et al.*, 2012). Valine metabolism can have several roles. First, valine degradation generates acetyl-CoA (via methylmalonate semialdehyde), which can in turn be used for respiration or lipid synthesis. Second, valine can be converted to leucine via 2-oxoisovalerate. Labelling with ^{14}C -valine in developing wheat spikes has shown up to 30% conversion to leucine (Kolderup, 1978), and during grain development, the valine content correlates with that of leucine (Martín Del Molino *et al.*, 1988). Interestingly, a quantitative trait locus analysis has shown that 2-methylmalate, an intermediate of leucine synthesis, correlates positively with the number of tillers and thermal time to heading, and negatively with yield (Hill *et al.*, 2015).

Conclusions

Taken as a whole, our results show that growth at elevated CO_2 in FACE experiments had different effects on leaf and grain metabolism, but no significant effect on yield. Elevated CO_2 appeared to be detrimental to photosynthesis and leaf proteins (e.g. Rubisco large subunit) and to alter grain composition, in particular %N, amino acids (e.g. threonine and alanine), and minerals (Mg). The yield was related to metabolic features of grains but this relationship was not influenced by elevated CO_2 .

The lack of a positive effect of CO_2 fertilization can be explained not only by the down-regulation of carbon fixation (i.e. of several photosynthetic parameters) but also by an effect on grain metabolism itself. In particular, elevated CO_2 appeared to be detrimental to N metabolism, with a decrease in several amino acids and, accordingly, a positive effect of elevated CO_2 on yield was observed only in the USA, where N fertilization was the highest.

Elevated CO_2 also had an effect on Mg redistribution between source leaves and grains. The detrimental effect of CO_2 on the content of microelements has been found elsewhere for Zn (Myers *et al.*, 2014). A meta-analysis has recently highlighted the decrease in the content of many elements (including macroelements such as S) in crops cultivated at elevated CO_2 (Loladze, 2014). Interestingly, recent experiments in which Mg availability was varied have shown that low Mg causes a significant decline in grain starch

content and yield (Ceylan *et al.*, 2016). Here, despite significantly lower Mg content in grains at elevated CO_2 , starch content and yield were not significantly affected, suggesting that the decrease in Mg associated with elevated CO_2 was too small (<10%) to affect yield. In addition, the relationship between Mg content in grains and yield is not driven by genotype (Oury *et al.*, 2006). We nevertheless recognize that further work is required to determine the reason for the decreased Mg content in grains found here. For example, isotopic labelling with ^{25}Mg (or ^{26}Mg) would be helpful to examine the role played by phloem Mg transport during remobilization from leaves at elevated CO_2 .

The overall effect on N, K, and Mg nutrition found here raises the question of whether specific fertilization management strategies could compensate for the lack of effect of elevated CO_2 observed at most of the FACE sites. Previous studies compared ammonium-based and nitrate-based fertilizers and found that with nitrate, plants tended to show higher photosynthetic acclimation (Bowler and Press, 1996; Geiger *et al.*, 1999; Cruz *et al.*, 2014), which in turn limits the response of production or yield to elevated CO_2 . In addition, high nitrate availability tends to exaggerate photosynthetic acclimation, due to down-regulation of the expression of photosynthetic genes (Vicente *et al.*, 2017). Therefore, in an effort to find optimal fertilization strategies adapted to local field conditions, both the quantity and the quality of N fertilizer seems to be important. The fact that nutrients other than N are also affected by elevated CO_2 further indicates that the nutrient balance itself is also of importance. A specific multifactorial experiment (nutrient compositions $\times \text{CO}_2$) would be necessary to determine the best solutions to improve yield responsiveness to CO_2 . In addition, the selection of varieties with a high harvest index, which is associated with optimal allocation and efficient nutrient remobilization capacity, might be desirable to obtain better grain yield and quality at elevated CO_2 .

Finally, we found that wheat grain yield was related to glutamine and valine grain content, regardless of country and cultivar, showing the importance of amino acid metabolism for grain maturation. However, while the role of glutamine is clear (as a N source and utilization for storage proteins, which are glutamine rich), understanding the metabolic role(s) of valine requires further work, such as tracing with ^{13}C -valine and isotope-assisted metabolomics. This will be addressed in a future study.

Supplementary data

The following supplementary data are available at JXB online.

Fig. S1. Elemental composition of soil (top layer) at the different sites used in this study.

Fig. S2. Leaf features significant for a $\text{CO}_2 \times$ conditions (country, cultivar, and time) effect.

Fig. S3. Grain features significant for a $\text{CO}_2 \times$ conditions (country, cultivar, and time) effect.

Fig. S4. Correlation plot showing the best variables obtained by subset sampling in linear models of yield.

Fig. S5. Photosynthetic response curve for wheat cultivated in Germany.

Acknowledgements

This work was supported by the Department of Industry, Energy and Innovation of the Government of Navarre (PI040 TRIGOCCLIM). The technical support given by Inés Urretavizcaya, Petra Högy, and Jürgen Franzaring in harvesting and sample management is acknowledged. JC was supported by an Australia Awards PhD Scholarship. GT was supported by a Connect Talent Award from the Region Pays de la Loire – Angers Loire Metropole (France). Research at the Australian Grains Free Air CO₂ Enrichment (AGFACE) facility was jointly run by the University of Melbourne and Agriculture Victoria with funding from the Grains Research and Development Corporation (under contract no. DAV00137) and the Australian Commonwealth Department of Agriculture and Water Resources (under contract no. FtRG 1193982–41). CAAS-FACE was supported by the National Key Research and Development Project (under contracts 2016YFD0300401 and 2019YFA0607403). The FACE experiment in Italy was supported by the AGER project ‘Durum wheat adaptation to global change: effect of elevated CO₂ on yield and quality traits’ and by the collaboration CREA-CNR. Finally, the authors also acknowledge support of the publication fee by the CSIC Open Access Publication Support Initiative through its Unit of Information Resources for Research (URICI).

References

- Adam NR, Wall GW, Kimball BA, *et al.* 2000. Acclimation response of spring wheat in a free-air CO₂ enrichment (FACE) atmosphere with variable soil nitrogen regimes. 1. Leaf position and phenology determine acclimation response. *Photosynthesis Research* **66**, 65–77.
- Ainsworth EA, Long SP. 2005. What have we learned from 15 years of free-air CO₂ enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy properties and plant production to rising CO₂. *New Phytologist* **165**, 351–371.
- Ainsworth EA, Rogers A. 2007. The response of photosynthesis and stomatal conductance to rising [CO₂]: mechanisms and environmental interactions. *Plant, Cell & Environment* **30**, 258–270.
- Akin DE, Kimball BA, Windham WR, Pinter PJ, Wall GW, Garcia RL, LaMorte RL, Morrison WH. 1995. Effect of free-air CO₂ enrichment (FACE) on forage quality of wheat. *Animal Feed Science and Technology* **53**, 29–43.
- Aranjuelo I, Erice G, Sanz-Sáez A, *et al.* 2015. Differential CO₂ effect on primary carbon metabolism of flag leaves in durum wheat (*Triticum durum* Desf.). *Plant, Cell & Environment* **38**, 2780–2794.
- Beleggia R, Fragasso M, Miglietta F, Cattivelli L, Menga V, Nigro F, Pecchioni N, Fares C. 2018. Mineral composition of durum wheat grain and pasta under increasing atmospheric CO₂ concentrations. *Food Chemistry* **242**, 53–61.
- Bhatti AU, Mulla DJ, Frazier BE. 1991. Estimation of soil properties and wheat yields on complex eroded hills using geostatistics and thematic mapper images. *Remote Sensing of Environment* **37**, 181–191.
- Bloom AJ, Burger M, Kimball BA, Pinter Jr PJ. 2014. Nitrate assimilation is inhibited by elevated CO₂ in field-grown wheat. *Nature Climate Change* **4**, 477–480.
- Bourgault M, Dreccher MF, James AT, Chapman SC. 2013. Genotypic variability in the response to elevated CO₂ of wheat lines differing in adaptive traits. *Functional Plant Biology* **40**, 172–184.
- Bowes G. 1993. Facing the inevitable: plants and increasing atmospheric CO₂. *Annual Review of Plant Biology* **44**, 309–332.
- Bowler JM, Press MC. 1996. Effects of elevated CO₂, nitrogen form and concentration on growth and photosynthesis of a fast- and slow-growing grass. *New Phytologist* **132**, 391–401.
- Cabrerizo PM, González EM, Aparicio-Tejo PM, Arrese-Igor C. 2001. Continuous CO₂ enrichment leads to increased nodule biomass, carbon availability to nodules and activity of carbon-metabolising enzymes but does not enhance specific nitrogen fixation in pea. *Physiologia Plantarum* **113**, 33–40.
- Ceylan Y, Kutman UB, Mengutay M, Cakmak I. 2016. Magnesium applications to growth medium and foliage affect the starch distribution, increase the grain size and improve the seed germination in wheat. *Plant and Soil* **406**, 145–156.
- Cheng L, Zhu J, Chen G, Zheng X, Oh NH, Ruffy TW, Richter DD, Hu S. 2010. Atmospheric CO₂ enrichment facilitates cation release from soil. *Ecology Letters* **13**, 284–291.
- Cruz JL, Alves AA, LeCain DR, Ellis DD, Morgan JA. 2014. Effect of elevated CO₂ concentration and nitrate: ammonium ratios on gas exchange and growth of cassava (*Manihot esculenta* Crantz). *Plant and Soil* **374**, 33–43.
- Erbs M, Franzaring J, Högy P, Fangmeier A. 2009. Free-air CO₂ enrichment in a wheat-weed assembly – effects on water relations. *Basic and Applied Ecology* **10**, 358–367.
- Ethier GJ, Livingston NJ. 2004. On the need to incorporate sensitivity to CO₂ transfer conductance into the Farquhar–von Caemmerer–Berry leaf photosynthesis model. *Plant, Cell and Environment* **27**, 137–153.
- Fangmeier A, Torres-Toledo V, Franzaring J, Damsohn W. 2016. Design and performance of a new FACE (free air carbon dioxide enrichment) system for crop and short vegetation exposure. *Environmental and Experimental Botany* **130**, 151–161.
- Fares C, Menga V, Badeck F, Rizza F, Miglietta F, Zaldei A, Codianni P, Iannucci A, Cattivelli L. 2016. Increasing atmospheric CO₂ modifies durum wheat grain quality and pasta cooking quality. *Journal of Cereal Science* **69**, 245–251.
- Farquhar GD, O’Leary MH, Berry JA. 1982. On the relationship between carbon isotope discrimination and the intercellular carbon dioxide concentration in leaves. *Functional Plant Biology* **9**, 121–137.
- Fernando N, Panozzo J, Tausz M, Norton RM, Fitzgerald GJ, Myers S, Nicolas ME, Seneweera S. 2014. Intra-specific variation of wheat grain quality in response to elevated [CO₂] at two sowing times under rain-fed and irrigation treatments. *Journal of Cereal Science* **59**, 137–144.
- Fisher DB, Macnicol PK. 1986. Amino acid composition along the transport pathway during grain filling in wheat. *Plant Physiology* **82**, 1019–1023.
- Galmés J, Aranjuelo I, Medrano H, Flexas J. 2013. Variation in Rubisco content and activity under variable climatic factors. *Photosynthesis Research* **117**, 73–90.
- Geiger M, Haake V, Ludewig F, Sonnewald U, Stitt M. 1999. The nitrate and ammonium nitrate supply have a major influence on the response of photosynthesis, carbon metabolism, nitrogen metabolism and growth to elevated carbon dioxide in tobacco. *Plant Cell and Environment* **22**, 1177–1199.
- Grant RF, Kimball BA, Brooks TJ, Wall GW, Pinter Jr PJ, Hunsaker DJ, Adamsen FJ, La Morte RL, Leavitt SW, Thompson TL. 2001. CO₂ effects on mass and energy exchange of wheat under different N fertilization: model theory and testing with a free air CO₂ enrichment (FACE) experiment. *Plant Cell and Environment* **93**, 630–649.
- Harley PC, Loreto F, Marco GD, Sharkey TD. 1992. Theoretical considerations when estimating the mesophyll conductance to CO₂ flux by analysis of the response of photosynthesis to CO₂. *Plant Physiology* **98**, 1429–1436.
- Hayashi H, Chino M. 1986. Collection of pure phloem sap from wheat and its chemical composition. *Plant and Cell Physiology* **27**, 1387–1393.
- He Z, Joshi AK, Zhang W. 2013. Climate vulnerabilities and wheat production. In: Pielke RA, ed. *Climate vulnerability*. Oxford: Academic Press, 57–67.
- Hill CB, Taylor JD, Edwards J, Mather D, Langridge P, Bacic A, Roessner U. 2015. Detection of QTL for metabolic and agronomic traits in wheat with adjustments for variation at genetic loci that affect plant phenology. *Plant Science* **233**, 143–154.
- Högy P, Brunnbauer M, Koehler P, Schwadorf K, Breuer J, Franzaring J, Zhunusbayeva D, Fangmeier A. 2013. Grain quality characteristics of spring wheat (*Triticum aestivum*) as affected by free-air CO₂ enrichment. *Environmental and Experimental Botany* **88**, 11–18.
- Högy P, Fangmeier A. 2008. Effects of elevated atmospheric CO₂ on grain quality of wheat. *Journal of Cereal Science* **48**, 580–591.

- Houshmandfar A, Fitzgerald GJ, Macabuhay AA, Armstrong R, Tausz-Posch S, Löw M, Tausz M.** 2016. Trade-offs between water-use related traits, yield components and mineral nutrition of wheat under Free-Air CO₂ Enrichment (FACE). *European Journal of Agronomy* **76**, 66–74.
- Hunsaker J, Kimball A, Pinter J Jr, LaMorte M, Wall W.** 1996. Carbon dioxide enrichment and irrigation effects on wheat evapotranspiration and water use efficiency. *Transactions of the ASAE* **39**, 1345–1355.
- Jauregui I, Aroca R, Garnica M, Zamarreño ÁM, García-Mina JM, Serret MD, Parry M, Irigoyen JJ, Aranjuelo I.** 2015. Nitrogen assimilation and transpiration: key processes conditioning responsiveness of wheat to elevated [CO₂] and temperature. *Physiologia Plantarum* **155**, 338–354.
- Kimball BA.** 2010. Lessons from FACE: CO₂ effects and interactions with water, nitrogen and temperature. In: Hillel D, Rosenzweig C, eds. *Handbook of climate change and agroecosystems: impacts, adaptation, and mitigation*. London: Imperial College Press, 87–107.
- Kimball BA.** 2016. Crop responses to elevated CO₂ and interactions with H₂O, N, and temperature. *Current Opinion in Plant Biology* **31**, 36–43.
- Kimball BA, Morris CF, Pinter PJ Jr, et al.** 2001a. Elevated CO₂, drought and soil nitrogen effects on wheat grain quality. *New Phytologist* **150**, 295–303.
- Kimball BA, Morris CF, Pinter PJ Jr, Wall GW, Hunsaker DJ, Adamsen FJ, Lamorte RL, Leavitt SW, Thompson TL, Matthias AD.** 2001b. Wheat grain quality as affected by elevated CO₂, drought, and soil nitrogen. *Plant Pathology* **150**, 295–303.
- Kimball BA, Pinter PJ Jr, Garcia RL, LaMorte RL, Wall GW, Hunsaker DJ, Wechsung G, Wechsung F, Kartschall T.** 1995. Productivity and water use of wheat under free-air CO₂ enrichment. *Global Change Biology* **1**, 429–442.
- Kolderup F.** 1978. Interconversions of amino acids in maturing wheat grains. In: *Seed protein improvement in cereals and grain legumes*, Vol. 1. Vienna: International Atomic Energy Agency, 187–204.
- Li A-G, Hou Y-S, Wall GW, Trent A, Kimball BA, Pinter PJ.** 2000. Free-air CO₂ enrichment and drought stress effects on grain filling rate and duration in spring wheat. *Crop Science* **40**, 1263–1270.
- Loladze I.** 2014. Hidden shift of the ionome of plants exposed to elevated CO₂ depletes minerals at the base of human nutrition. *eLife* **3**, e02245.
- López-Ferrer D, Martínez-Bartolomé S, Villar M, Campillos M, Martín-Maroto F, Vázquez J.** 2004. Statistical model for large-scale peptide identification in databases from tandem mass spectra using SEQUEST. *Analytical Chemistry* **76**, 6853–6860.
- MacLean B, Tomazela DM, Shulman N, Chambers M, Finney GL, Frewen B, Kern R, Tabb DL, Liebler DC, MacCoss MJ.** 2010. Skyline: an open source document editor for creating and analyzing targeted proteomics experiments. *Bioinformatics* **26**, 966–968.
- Manderscheid R, Dier M, Erbs M, Sickora J, Weigel H-J.** 2018. Nitrogen supply – a determinant in water use efficiency of winter wheat grown under free air CO₂ enrichment. *Agricultural Water Management* **210**, 70–77.
- Marti J, Slafer GA.** 2014. Bread and durum wheat yields under a wide range of environmental conditions. *Field Crops Research* **156**, 258–271.
- Martín Del Molino IM, Rojo B, Martínez-Carrasco R, Pérez P.** 1988. Amino acid composition of wheat grain. 1: Changes during development. *Journal of the Science of Food and Agriculture* **42**, 29–37.
- Meehl G, Stocker T, Collins W, et al.** 2007. Global climate projections. In: Solomon S, Qin D, Manning M, Chen Z, Marquis M, Averyt K, Tignor M, Miller H, eds. *Climate change 2007: the physical science basis. Contribution of working group I to the fourth assessment report of the Intergovernmental Panel on Climate Change (IPCC)*. New York: Cambridge University Press, 747–845.
- Miglietta F, Giuntoli A, Bindi M.** 1996. The effect of free air carbon dioxide enrichment (FACE) and soil nitrogen availability on the photosynthetic capacity of wheat. *Photosynthesis Research* **47**, 281–290.
- Miller MP, Singer MJ, Nielsen DR.** 1988. Spatial variability of wheat yield and soil properties on complex hills. *Soil Science Society of America Journal* **52**, 1133–1141.
- Mollah M, Norton R, Huzzey J.** 2009. Australian grains free-air carbon dioxide enrichment (AGFACE) facility: design and performance. *Crop and Pasture Science* **60**, 697–707.
- Moulin AP, Anderson DW, Mellinger M.** 1994. Spatial variability of wheat yield, soil properties and erosion in hummocky terrain. *Canadian Journal of Soil Science* **74**, 219–228.
- Myers SS, Zanobetti A, Kloog I, et al.** 2014. Increasing CO₂ threatens human nutrition. *Nature* **510**, 139–142.
- Nie G, Hendrix DL, Webber AN, Kimball BA, Long SP.** 1995. Increased accumulation of carbohydrates and decreased photosynthetic gene transcript levels in wheat grown at an elevated CO₂ concentration in the field. *Plant Physiology* **108**, 975–983.
- O’Leary GJ, Christy B, Nuttall J, et al.** 2015. Response of wheat growth, grain yield and water use to elevated CO₂ under a Free-Air CO₂ Enrichment (FACE) experiment and modelling in a semi-arid environment. *Global Change Biology* **21**, 2670–2686.
- Oliveira HR, Campana MG, Jones H, Hunt HV, Leigh F, Redhouse DI, Lister DL, Jones MK.** 2012. Tetraploid wheat landraces in the Mediterranean basin: taxonomy, evolution and genetic diversity. *PLoS One* **7**, e37063.
- Oury F-X, Leenhardt F, Remesy C, Chanliaud E, Duperrier B, Balfourier F, Charmet G.** 2006. Genetic variability and stability of grain magnesium, zinc and iron concentrations in bread wheat. *European Journal of Agronomy* **25**, 177–185.
- Pacholski A, Manderscheid R, Weigel H-J.** 2015. Effects of free air CO₂ enrichment on root growth of barley, sugar beet and wheat grown in a rotation under different nitrogen supply. *European Journal of Agronomy* **63**, 36–46.
- Pandey V, Sharma M, Deeba F, Maurya VK, Gupta SK, Singh SP, Mishra A, Nautiyal CS.** 2017. Impact of elevated CO₂ on wheat growth and yield under free air CO₂ enrichment. *American Journal of Climate Change* **6**, 573–596.
- Pendall E, Leavitt SW, Brooks T, et al.** 2001. Elevated CO₂ stimulates soil respiration in a FACE wheat field. *Basic and Applied Ecology* **2**, 193–201.
- Peñuelas J, Estiarte M, Kimball BA.** 2000. Flavonoid responses in wheat grown at elevated CO₂: green versus senescent leaves. *Photosynthetica* **37**, 615–619.
- Prioul J, Manicacci D, Damerval C, Méchin V.** 2012. Proteomics in identifying new regulatory mechanisms involved in seed development and ultimately seed quality. In: Agrawal G, Rakwal R. eds. *Seed development: OMICS technologies toward improvement of seed quality and crop yield*. Dordrecht: Springer, 247–264.
- Ray DK, Ramankutty N, Mueller ND, West PC, Foley JA.** 2012. Recent patterns of crop yield growth and stagnation. *Nature Communications* **3**, 1293.
- Reynolds M, Foulkes J, Furbank R, Griffiths S, King J, Murchie E, Parry M, Slafer G.** 2012. Achieving yield gains in wheat. *Plant, Cell & Environment* **35**, 1799–1823.
- Rogers A, Humphries SW.** 2000. A mechanistic evaluation of photosynthetic acclimation at elevated CO₂. *Global Change Biology* **6**, 1005–1011.
- Sanchez de la Puente L, Pérez P, Martínez-Carrasco R, Morcuende R, Martín Del Molino IM.** 2000. Action of elevated CO₂ and high temperatures on the mineral composition of two varieties of wheat. *Agrochimica* **44**, 221–230.
- Shewry PR, Mitchell RAC, Tosi P, Wan Y, Underwood C, Lovegrove A, Freeman J, Toole GA, Mills ENC, Ward JL.** 2012. An integrated study of grain development of wheat (cv. Hereward). *Journal of Cereal Science* **56**, 21–30.
- Sicher RC, Bunce JA.** 1998. Evidence that premature senescence affects photosynthetic decline of wheat flag leaves during growth in elevated carbon dioxide. *International Journal of Plant Sciences* **159**, 798–804.
- Slafer G, Calderini D, Miralles D.** 1996. Yield components and compensation in wheat: opportunities for further increasing yield potential. In: Reynolds MP, Rajaram S, McNab A, eds. *Increasing yield potential in wheat: breaking the barriers*. Mexico DF: CIMMYT, 101–133.
- Soba D, Ben Mariem S, Fuertes-Mendizábal T, Méndez-Espinoza AM, Gilard F, González-Murua C, Irigoyen JJ, Tcherkez G, Aranjuelo I.** 2019. Metabolic effects of elevated CO₂ on wheat grain development and composition. *Journal of Agricultural and Food Chemistry* **67**, 8441–8451.
- Tasleem-Tahir A, Nadaud I, Chambon C, Branlard G.** 2012. Expression profiling of starchy endosperm metabolic proteins at 21 stages of wheat grain development. *Journal of Proteome Research* **11**, 2754–2773.
- Tausz-Posch S, Borowiak K, Dempsey RW, Norton RM, Seneweera S, Fitzgerald GJ, Tausz M.** 2013a. The effect of elevated CO₂ on photochemistry and antioxidative defence capacity in wheat depends on environmental

growing conditions – a FACE study. *Environmental and Experimental Botany* **88**, 81–92.

Tausz-Posch S, Norton RM, Seneweera S, Fitzgerald GJ, Tausz M. 2013b. Will intra-specific differences in transpiration efficiency in wheat be maintained in a high CO₂ world? A FACE study. *Physiologia Plantarum* **148**, 232–245.

Thilakarathne CL, Tausz-Posch S, Cane K, Norton RM, Fitzgerald GJ, Tausz M, Seneweera S. 2015. Intraspecific variation in leaf growth of wheat (*Triticum aestivum*) under Australian Grain Free Air CO₂ Enrichment (AGFACE): is it regulated through carbon and/or nitrogen supply? *Functional Plant Biology* **42**, 299–308.

Thilakarathne CL, Tausz-Posch S, Cane K, Norton RM, Tausz M, Seneweera S. 2013. Intraspecific variation in growth and yield response to elevated CO₂ in wheat depends on the differences of leaf mass per unit area. *Functional Plant Biology* **40**, 185–194.

Tiryakioglu M, Yildirim M, Karanlik S. 2014. Macronutrient concentration and remobilization in spring wheat organs during grain filling. *Turkish Journal of Agriculture and Forestry* **38**, 488–494.

Torres N, Goicoechea N, Zamarreño AM, Antolín MC. 2018. Mycorrhizal symbiosis affects ABA metabolism during berry ripening in *Vitis vinifera* L. cv. Tempranillo grown under climate change scenarios. *Plant Science* **274**, 383–393.

Tubiello FN, Rosenzweig C, Kimball BA, Pinter PJ Jr, Wall GW, Hunsaker DJ, LaMorte RL, Garcia RL. 1999. Testing CERES–Wheat with Free-Air Carbon Dioxide Enrichment (FACE) experiment data: CO₂ and water interactions. *Agronomy Journal* **91**, 247–255.

Ugalde T, Jenner C. 1990. Substrate gradients and regional patterns of dry matter deposition within developing wheat endosperm. II. Amino acids and protein. *Functional Plant Biology* **17**, 395–406.

Uprety DC, Dwivedi N, Raj A, Jaiswal S, Paswan G, Jain V, Maini HK. 2009. Study on the response of diploid, tetraploid and hexaploid species of wheat to the elevated CO₂. *Physiology and Molecular Biology of Plants* **15**, 161–168.

Vensel WH, Tanaka CK, Cai N, Wong JH, Buchanan BB, Hurkman WJ. 2005. Developmental changes in the metabolic protein profiles of wheat endosperm. *Proteomics* **5**, 1594–1611.

Veres S, Malik AI, Rengel Z. 2017. Differential nitrogen supply causes large variability in photosynthetic traits in wheat germplasm. *Crop and Pasture Science* **68**, 703–712.

Verrillo F, Badeck F-W, Terzi V, et al. 2017. Elevated field atmospheric CO₂ concentrations affect the characteristics of winter wheat (cv. Bologna) grains. *Crop and Pasture Science* **68**, 713–725.

Vicente R, Pérez P, Martínez-Carrasco R, Morcuende R. 2017. Improved responses to elevated CO₂ in durum wheat at a low nitrate supply

associated with the upregulation of photosynthetic genes and the activation of nitrate assimilation. *Plant Science* **260**, 119–128.

von Caemmerer S, Farquhar GD. 1981. Some relationships between the biochemistry of photosynthesis and the gas exchange of leaves. *Planta* **153**, 376–387.

Walker C, Armstrong R, Panozzo J, Partington D, Fitzgerald G. 2017. Can nitrogen fertiliser maintain wheat (*Triticum aestivum*) grain protein concentration in an elevated CO₂ environment? *Soil Research* **55**, 518–523.

Wechsung F, Garcia RL, Wall GW, et al. 2000. Photosynthesis and conductance of spring wheat ears: field response to free-air CO₂ enrichment and limitations in water and nitrogen supply. *Plant Cell and Environment* **23**, 917–929.

Wechsung G, Wechsung F, Wall GW, Adamsen FJ, Kimball BA, Garcia RL, Pinter PJ, Kartschall T. 1995. Biomass and growth rate of a spring wheat root system grown in free-air CO₂ enrichment (FACE) and ample soil moisture. *Journal of Biogeography* **22**, 623–634.

Wieser H, Manderscheid R, Erbs M, Weigel HJ. 2008. Effects of elevated atmospheric CO₂ concentrations on the quantitative protein composition of wheat grain. *Journal of Agricultural and Food Chemistry* **56**, 6531–6535.

Wolf J, van Oijen M, Kempenaar C. 2002. Analysis of the experimental variability in wheat responses to elevated CO₂ and temperature. *Agriculture, Ecosystems and Environment* **93**, 227–247.

Wroblewitz S, Hüther L, Manderscheid R, Weigel HJ, Wätzig H, Dänicke S. 2013. The effect of free air carbon dioxide enrichment and nitrogen fertilisation on the chemical composition and nutritional value of wheat and barley grain. *Archives of Animal Nutrition* **67**, 263–278.

Yang L-X, Li S-F, Wang Y-L, Huang J-Y, Yang H-J, Dong G-C, Zhu J-G, Liu G. 2007a. Effects of free-air CO₂ enrichment (FACE) on yield formation of wheat. *Journal of Applied Ecology* **18**, 75–80.

Yang L-X, Wang Y-L, Li S-F, Huang J-Y, Dong G-C, Zhu J-G, Liu G, Han Y. 2007b. Effects of free-air CO₂ enrichment (FACE) on dry matter production and allocation in wheat. *Journal of Applied Ecology* **18**, 339–346.

Zhang D-Y, Chen G-Y, Chen J, Yong Z-H, Zhu J-G, Xu D-Q. 2009. Photosynthetic acclimation to CO₂ enrichment related to ribulose-1,5-bisphosphate carboxylation limitation in wheat. *Photosynthetica* **47**, 152–154.

Zhang Z, Chen J, Lin S, Li Z, Cheng R, Fang C, Chen H, Lin W. 2012. Proteomic and phosphoproteomic determination of ABA's effects on grain-filling of *Oryza sativa* L. inferior spikelets. *Plant Science* **185–186**, 259–273.

Zhu C, Zhu J, Zeng Q, Liu G, Xie Z, Tang H, Cao J, Zhao X. 2009. Elevated CO₂ accelerates flag leaf senescence in wheat due to ear photosynthesis which causes greater ear nitrogen sink capacity and ear carbon sink limitation. *Functional Plant Biology* **36**, 291–299.