

Lung Pathology in COVID-19: A Systematic Review

Abstract

Sparse literature is available regarding autopsy findings of coronavirus disease 2019 (COVID-19) despite high mortality due to its highly contagious nature and lack of robust infrastructure for appropriate handling of the infected cases. Based on clinical findings and various diagnostic tests, it is evident that it holds the potential to affect multiple organ systems of the body preferably lungs and immune and coagulation systems. Cytokine storm-induced thrombotic complication such as disseminated intravascular coagulation is a significant feature in severe cases of COVID-19. This review captures the current information on lung histopathology in COVID-19 infection and severe respiratory failure. In COVID-19, lungs are affected bilaterally, become edematous and red/tan mottled to maroon in color with firm consistency. Distinct parenchymal changes, firm thrombi in the peripheral pulmonary vessels along with diffuse alveolar damage, have been the most consistent feature of COVID-19-related lung pathology. Electron microscopy has also been used to demonstrate viral particles.

Keywords: Acute respiratory distress syndrome, alveolar damage, COVID-19, lung pathology, pneumonia

Introduction

It was at the end of 2019 when a new pandemic caused by a novel coronavirus-severe acute respiratory syndrome-coronavirus-2 (SARS-CoV-2) subsequently hit the world after originating from Wuhan, Hubei province, in China, and the disease thus caused in human was named coronavirus disease 2019 (COVID-19) by the World Health Organization (WHO) and on January 30, 2020, it was declared as a global health emergency.^[1] The 21st century, since its beginning, has witnessed three coronavirus pandemics so far namely SARS-CoV which also had originated in China in 2002 and led to 8098 cases with 774 deaths worldwide, followed by Middle East respiratory syndrome-coronavirus (MERS-CoV) in 2012 that began in Saudi Arabia and resulted in 2458 cases with 848 deaths^[2,3] and latest in the row is the most pathogenic and widely spread COVID-19 pandemic, of which 4,993,470 cases have already occurred worldwide with 327,738 deaths as on May 22, 2020, as declared by the WHO.^[4] Compared to SARS-CoV and MERS-CoV, SARS-CoV-2 is far more

infective and transmissible but with lower mortality rate^[5] which is 2%–3% in hospitalized patients.^[6] The reported case fatality rate for SARS is 9.6%^[7] and that for MERS is 34%.^[8] Coronaviruses are known to cause diseases in both humans and animals, affecting pulmonary, intestinal, hepatic, renal, and neurologic systems.^[9] SARS-CoV-2 is the seventh member of the family of coronaviruses that infects humans. Zoonotic coronaviruses after crossing the species barrier resulted in these pandemics associated with severe morbidity and mortality in humans.

Study design

For the present review, a systematic literature search was undertaken over online databases such as PubMed, MEDLINE, medRxiv, bioRxiv, ChemRxiv, Google Scholar, and CNKI using relevant keywords: “SARS-CoV-2,” “2019-nCoV epidemiology,” “COVID-19 lung pathology,” “coronavirus,” “COVID-19 autopsy findings,” SARS, and MERS. Articles written in English or Chinese language and those that were accepted, in preprint stage, in press, and published between August 2016 and May 5, 2020, were included. Only publications covering lung pathology in COVID-19 were chosen. This review is entirely based on the

How to cite this article: Pandey P, Agarwal S, Rajkumar. Lung pathology in COVID-19: A systematic review. *Int J App Basic Med Res* 2020;10:226-33.

**Pinki Pandey,
Savita Agarwal,
Rajkumar¹**

*Departments of Pathology and
¹Neurosurgery, UP University
of Medical Sciences, Etawah,
Uttar Pradesh, India*

Submitted: 19-Jun-2020

Revised: 30-Aug-2020

Accepted: 18-Sep-2020

Published Online: 07-Oct-2020

Address for correspondence:

*Dr. Pinki Pandey,
Type V, B 101, UP University
of Medical Sciences,
Saifai, Etawah - 206 130,
Uttar Pradesh, India.
E-mail: pnkdxt@yahoo.co.in*

Access this article online

Website:
www.ijabmr.org

DOI:
10.4103/ijabmr.IJABMR_381_20

Quick Response Code:



This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: WKHLRPMedknow_reprints@wolterskluwer.com

information published in the literatures mentioned under reference section.

Etiopathogenesis

Coronaviruses are enveloped, positive-sense, single-stranded, RNA viruses which are genotypically and serologically divided into four genera, namely, α , β , γ , and δ infecting mammals and birds.^[2,10] SARS-CoV-2 is β -CoV just like SARS-CoV and MERS-CoV, which are known to cause deadly respiratory tract infection in humans.^[11] Structurally, SARS-CoV-2 genome is of size 29.9 kb.^[12] Infection with SARS-CoV-2 is acquired through respiratory droplets, by contact through mucosal surfaces and potentially by feco-oral route (though it is yet to be studied).^[7,13] SARS-CoV-2, just like SARS-CoV, utilizes host angiotensin-converting enzyme 2 (ACE2) as a receptor for its binding.^[14,15] ACE2 is expressed in many tissues including alveolar cells, bronchial cells, and vascular endothelium in the lung and is involved in the pathogenesis of acute lung injury and pulmonary edema.^[16-18] Viral interaction with its receptor causes proteolytic cleavage involving the viral spike protein, leading to membrane fusion and infection,^[19] releasing the viral RNA genome into the cytoplasm to be translated into structural and polyproteins, resulting in viral replication within endoplasmic reticulum and Golgi apparatus followed by fusion of virus-containing vesicles with plasma membrane and release of virus.^[20,21] Rapid viral replication may lead to epithelial and endothelial cell damage, causing vascular leakage, and virus-mediated ACE2 downregulation and shedding and antibody-dependent enhancement (ADE) altogether cause prominent systemic inflammatory response where various immunological cells including CD4 T cells and CD8 T cells produce and regulate the release of different pro-inflammatory cytokines and chemokines that include interleukin (IL)-8, IL-18, IL-1, tumor necrosis factor- α , IL-6, and IL-10.^[22-24] ADE, observed in many viral infections, further enhances the infection of target cells by assisting in cellular uptake of infectious virus-antibody complexes.^[25]

Two interrelated and mutually reinforcing phenomena, that is, thrombosis and inflammation, are together believed to result in disseminated intravascular coagulation (DIC) and consumption coagulopathy as a terminal event in COVID-19,^[26,27] which is far more likely in nonsurvivors (71.4%) compared to survivors (0.6%).^[28] In a recent report from Italy, hospitalized COVID-19 patients had arterial and venous thromboembolic complications, occurring at a cumulative rate of 21%, and DIC as a secondary outcome was present in 2.2% cases with associated elevated D-dimer levels indicating inflammatory and procoagulant state in these patients, particularly in those patients who succumbed to their illness.^[29] Latest literature showed that COVID-19 acute respiratory distress syndrome (ARDS) patients experienced much

higher (11.7%) rate of thromboembolic complications compared to non-COVID-19 ARDS patients.^[30] A recent series on 12 COVID-19 autopsy cases reported deep-vein thrombosis in seven cases, in which venous thromboembolism was not suspected antemortem.^[31] Apart from hemostatic functions, both coagulation factors and platelets also act as an immunomodulator through their pro-inflammatory properties.^[32]

Clinical Features

The mean incubation period is 5.2 days (95% confidence interval, 4.1–7.0).^[33] The most frequent clinical manifestations in COVID-19-infected patients include fever (83%–98%), dry cough (76%–82%), fatigue, and dyspnea. Patients also present with symptoms pertaining to multiple systems such as headache, confusion, hemoptysis, sputum production, rhinorrhea, sore throat, dyspnea, diarrhea, chest pain, nausea, vomiting, myalgia, and conjunctival injection.^[13,34] Poor prognosis and higher mortality were observed in elderly patients and those with underlying cardiovascular disease, chronic respiratory disease, diabetes, cancer, and hypertension where the reported case fatality rate was 10.5%, 6.3%, 7.3%, 5.6%, and 6.0%, respectively.^[5,35,36] Published reports have documented that in severe cases, patients also develop acute kidney injury, arrhythmia due to cardiac dysfunction, shock, hepatic dysfunction, and hematological abnormalities such as lymphocytopenia.^[37,38]

Diagnostics

Real-time-polymerase chain reaction for detection of SARS-CoV-2 done on nasopharyngeal swab, tracheal aspirate, or bronchoalveolar lavage carries very high specificity. SARS-CoV-2 RNA has also been detected in blood and stool samples. Frequently found hematological abnormalities are leukopenia; leukocytosis; lymphopenia; thrombocytopenia; and elevated levels of alanine aminotransferase, aspartate aminotransferase, and lactate dehydrogenase.^[6,39] There is elevation in D-dimer, ferritin, and C-reactive protein levels, whereas fall in procalcitonin levels has been reported. Radiological findings such as ground-glass opacity, consolidations, and the crazy-paving pattern are frequently observed mainly in the peripheral portions of lower lobes of lung on computed tomography. These are nonspecific signs and are probably due to alveolar septal edema and interstitial hyperplasia.^[40,41]

Pathology

Sparse literature is available regarding autopsy findings of this novel virus despite the high mortality probably because of its highly contagious nature and limited information on its prevention and lack of robust infrastructure for appropriate handling of infected cases at several centers across the globe. However, autopsy remains the gold standard for ascertaining the exact cause (s) of death

and provides necessary information for optimizing clinical management as it permits adequate sampling and study of multiple organs for diagnostic and research purposes.^[42] Significance of autopsies is further emphasized by the finding that in up to 30% cases, antemortem diagnostic errors or unrecognized diagnoses were disclosed on autopsies.^[43,44] The Royal College of Pathologists has presented latest guidelines for pathologists and mortuary technicians on autopsies in confirmed or suspected cases of COVID-19.^[45,46] Pathology-related information obtained through autopsies is essential for understanding its pathogenesis and critical morphological findings. Moreover, postmortem tissue sampling will also be needed for enhanced understanding of current novel virus through *in situ* and molecular studies.^[47]

There is marked resemblance in the histopathological findings of SARS-CoV, MERS-CoV, and SARS-CoV-2. The focus of the current review is to capture the currently available information on lung histopathology in COVID-19 infection and severe respiratory failure. Details of published literature discussing pulmonary involvement from pathologists' perspective are tabulated in Table 1.

Lungs

For pathological study of lung, tissue from patients who succumbed after acquiring SARS-CoV-2 infection at various centers across the globe, was obtained after complete postmortem examinations in seven studies,^[31,42,48-52] whole-lung biopsy in one study,^[18] postmortem autopsy lung tissues in one study,^[53] and thoroscopic resection of lung lobe in one study performed for two cases of carcinoma lung;^[54] in other five studies, postmortem needle core biopsies, ultrasound-based minimally invasive autopsies, and minimally invasive autopsies were performed.^[26,55-58] Of all these studies, the largest series was based on 38 cases^[53] from Northern Italy.

A gross description of COVID-19-related lung pathology was available in 9^[18,31,42,48-53] out of 15 studies included in the present review. In a study by Tian *et al.*,^[54] two patients underwent thoroscopic resection of lung lobe for carcinoma lung, however on gross description, only tumor-related findings were described and lung changes due to COVID-19 were not mentioned probably because lobectomy was done very early during the disease course, much before the gross changes could have become apparent.

Gross Features

In COVID-19, lungs being the primary organ to be affected are bilaterally involved and are heavy ranging on the left side – 680–1269 g and on the right side – 800–1183 g.^[31,42,48-50] There is diffuse or spotty involvement of lung parenchyma which is edematous and red/tan mottled to maroon in color with firm consistency similar

to ARDS. Distinct parenchymal changes also include congestion, punctuate hemorrhages, and hemorrhagic necrosis, particularly at the peripheral edges of the pulmonary lobe, especially right lower lobes. Further, Luo *et al.*^[18] mentioned the presence of hemorrhagic necrosis preferentially in the outer edge of the lower lobe of the right lung, suggesting it as one of the initial sites of origin of main lesions in COVID-19 and could be the result of CD4 and CD8 T cell-induced cytokine storm, which further progresses to severe and at time fatal respiratory dysfunction in critical patients. Buja *et al.*^[50] also described empyema and atelectasis in one of the cases in their study.

Cut surfaces after fixation have shown alternating consolidated tan-gray area and patchy hemorrhagic areas. Notable gross examination finding as reported by several authors^[31,48-52] is the presence of small or large, firm pulmonary thrombo-emboli in the peripheral or central parenchyma and segmental or sub-segmental regions. The upper and lower airways may remain patent with glistening white mucosa, or the bronchi may show mucinous and hemorrhagic exudation or pink froth in the airways.^[18,42,48,53] Menter *et al.*^[51] also observed severe mucous tracheitis/tracheobronchitis in one-third of the patients and in addition to consolidation also described extensive suppurative bronchopneumonic infiltrates. Pericardial and pleural effusions with mild-to-moderate serosanguinous fluid has also been reported.^[48,49] In autopsy cases of SARS-CoV infection, the gross features are described as firm, edematous, heavy lungs with congestion and hemorrhages along with hilar and abdominal lymphadenopathy with small spleen size.^[59,60]

Microscopic Features

Studies have found remarkable similarities in the microscopic features of lung lesions in all the three coronavirus pandemic cases of the 21st century. Diffuse alveolar damage (DAD) has been the consistent feature of COVID-19-related lung pathology. In the largest autopsy series comprising 38 cases,^[55] lung histopathological features relating to cellular and interstitial damage were extensively studied and semi-quantitatively graded. Histopathological features conforming to exudative, proliferative, and fibrotic phases with other associated findings such as alveolar multinucleated giant cells and interstitial and alveolar inflammation were assessed. Bilateral DAD in the exudative and proliferative phases is the most consistent finding.^[31,48-51,53,56]

Features of exudative phase included capillary congestion, pneumocyte injury with sloughing and scattered syncytial giant cell formation, dilated alveolar ducts, interstitial edema, and thickening of alveolar capillaries along with intra-alveolar hyaline membranes which are composed of serum proteins and condensed or organized fibrin; there may be massive fibrinous exudate with intense mononuclear inflammatory cells and multinucleated giant

Table 1: Salient features of the papers and patient characteristics as mentioned in the studies

Study	Type of study, method of sampling, and number of cases	Age range in years, gender	Duration of illness and clinical presentation	Risk factors	Cause of death	DAD on lung pathology
Fox et al. ^[48]	Series of autopsies, 4 cases	44-76, male and female	Three days of mild cough and fever Acute respiratory distress syndrome	Obesity, hypertension, insulin-dependent diabetes, chronic kidney disease	Thrombotic microangiopathy	Present
Barton et al. ^[42]	Original article, complete postmortem examinations, 2 cases	Case (1) 77-year-old male Case (2) 42-year-old male	Case (1) six days of fever and chills Case (2) abdominal pain followed by fever, shortness of breath, and cough	Case (1) obesity, hypertension Case (2) obesity, myotonic muscular dystrophy	Case (1) COVID-19, with coronary artery disease Case (2) Complications of hepatic cirrhosis, with muscular dystrophy, aspiration pneumonia Respiratory failure	Case (1) present Case (2) acute Broncho-pneumonia
Luo et al. ^[18]	Case report	60-year-old male	High fever and cough	Hypertension	Respiratory failure	Interstitial fibrosis, and hyaline degeneration
Zhang et al. ^[55]	Whole-lung biopsy Brief research report postmortem	72-year-old male	Fever and cough	Diabetes and hypertension	Respiratory failure	Present
Tian et al. ^[56]	Trans thoracic needle biopsy Article	59-81 years, three males and one female	Fever	Chronic lymphocytic leukemia, renal transplantation, cirrhosis, hypertension, and diabetes	Not mentioned	Present
Xu et al. ^[57]	Postmortem needle-core biopsies, 4 cases Case report Postmortem needle-core biopsy	50-year-old male	Fever, chills, cough, fatigue, and shortness of breath.	Travel history to Wuhan	Coronavirus pneumonia	Present
Tian et al. ^[54]	Case report Thoracoscopic lung resection Case (1) right middle lobe Case (2) right lower lobe	Case (1) 84-year-old female Case (2) 73-year-old male	Case (1) difficulty in breathing, chest tightness, wheezing, and dry cough Case (2) fever on postoperative day 9 with dry cough, chest tightness, and muscle pain	Case (1) lung adenocarcinoma, hypertension for 30 years, Type 2 diabetes Case (2) lung adenocarcinoma, hypertension	Case (1) Coma Case (2) Recovered alive	Case (1) adenocarcinoma, with alveolar damage Case (2) adenocarcinoma with DAD
Dolhnikoff et al. ^[26]	Letter to editor, ultrasound-based minimally invasive autopsies for COVID-19, ten cases	33-83 years, five males and five females	Not mentioned	Hypertension, diabetes mellitus, ischemic heart disease, and chronic obstructive pulmonary disorders	Not mentioned	Present
Carsana et al. ^[53]	Original article, Postmortem autopsy lung tissues 38 cases	32-86 years, 33 males and 5 females	Not mentioned	Diabetes, hypertension, cardiovascular disorders, chronic obstructive pulmonary disorders	Not mentioned	Present

Contd...

Table 1: Contd...

Study	Type of study, method of sampling, and number of cases	Age range in years, gender	Duration of illness and clinical presentation	Risk factors	Cause of death	DAD on lung pathology
Yao et al. ^[58]	Minimally invasive autopsies from multiple organs	Three cases	Not mentioned	Not mentioned	Coronavirus pneumonia	Not mentioned
Bradley et al. ^[49]	Original article, Autopsy 12 cases	42-84 years, five males and seven females	Respiratory distress (83.3%), fever (58.3%), cough (50%), altered mental status, and gastrointestinal distress	Hypertension, chronic kidney disease, diabetes, and obesity	Coronavirus pneumonia	Present
Buja et al. ^[50]	Original article, Autopsy, three cases	Case (1) 62-year-old male Case (2) 34-year-old male Case (3) 48-year-old male	Case (1) found dead in his car with few-day history of respiratory illness Case (2) headache, shortness of breath, hemoptysis for 4 days, and fever for 1 day Case (3) found dead at his residence	Case (1) obesity Case (2) hypertension, heart failure, Type 2 diabetes mellitus, and microcytic anemia Case (3) obesity	Case (2) respiratory failure	Case (1) present Case (2) acute thromboemboli with pulmonary hemorrhage and infarction Case (3) present
Wichmann et al. ^[31]	Original article, Autopsy, 12 cases	52-87 years, nine males and three females	Not mentioned	Obesity, coronary heart disease, asthma or chronic obstructive pulmonary disease, and diabetes mellitus type 2	Pulmonary embolism in four cases, sudden cardiac death, respiratory failure	Present
Menter et al. ^[51]	Original article, Autopsy, 21 cases	53-96 years, 17 males and 4 females	Dry cough and fever, dyspnea	Hypertension, obesity, cardiovascular diseases, diabetes mellitus, 65% had blood group A	Respiratory failure	Present
Grimes et al. ^[52]	Case report, Autopsy, two cases	Case (1, 2) both middle-aged males	Case (1) fever, chills, myalgia, dry cough, and dyspnea Case (2) fever, chills, productive cough, and dyspnea	Case (1) hypertension Case (2) asthma, hypertension and HIV	Case (1, 2) pulmonary thromboembolism	Pulmonary thromboembolism

DAD: Diffuse alveolar damage

cells in air spaces.^[18,26,42,48-51,53-58] Many inspissated spherical secretions or globules are also described.^[54]

Findings conforming to proliferative phase of DAD include Type-2 pneumocyte hyperplasia of varying degrees with or without reactive atypia, and interstitial myofibroblastic proliferation together causing thickening of alveolar walls, alveolar granulation tissue, obliterating fibrosis, alveolar macrophages, interstitial thickening, and plugs of proliferating fibroblasts or “fibroblast balls” in the interstitium. Mild-to-moderate lymphocytic infiltrate with foci of hemorrhages can also be seen.^[26,48-51,54,56,58]

Bronchi and bronchioles may show chronic inflammation with marked thickening of bronchial mucosa due to edema.^[18,42,49] Other features such as necrotizing bronchiolitis and alveolitis with atrophy, proliferation, and squamous metaplasia in alveolar epithelium have also been reported in SARS-CoV-2.^[31,50,59,60]

In another case reported by Barton *et al.*, the patient had features of acute bronchopneumonia with rare aspirated foreign material of vegetable matter seen within the airways with infiltration of peribronchiolar airspaces by neutrophils and histiocytes and DAD was not present, hence the cause of death in this case was aspiration pneumonia and not COVID-19 although he tested positive for it.^[42]

Features corresponding to fibrotic phase such as pleural involvement, mural fibrosis, scars, microcystic honeycombing, and arterial hypermuscularization were uncommon probably due to the short duration of disease which ranged from 5 to 31 days between the onset of symptoms and death in the largest series of the present review.^[53]

The most remarkable and clinically important finding mentioned in most of the reports is diffusely or focally present platelet-fibrin thrombi involving the peripheral or central pulmonary arterial vessels and capillaries in affected and preserved lung parenchyma and entrapped in these thrombi, many inflammatory cells including neutrophils can be seen.^[18,26,31,42,48,50-53]

Few studies also mentioned about the presence of endothelial tumefaction and platelet-producing pulmonary megakaryocytes displaying nuclear hyperchromasia and atypia within the alveolar capillaries as an indicator of coagulopathy.^[26,48,50]

Patients developing severe disease frequently show deranged coagulation profile with elevated levels of D-dimer in blood, for which targeted therapy in the form of anticoagulants has to be instituted. This hypercoagulable status is further supported by the frequent presence of pulmonary microthrombosis, which leads to hypoxemia and respiratory failure in these cases.^[18,26]

Another finding in favor of viral etiology is the presence of definite or suspected viral cytopathic effects in

pneumocytes characterized by cytomegaly, nuclear enlargement, prominent eosinophilic nucleoli, and eccentrically placed intracytoplasmic inclusions in the intra-alveolar spaces.^[18,26,48,49,54,57]

Blood vessels may show fibrinoid necrosis, wall thickening, luminal stenosis, or occlusion, leading to the development of pulmonary hypertension in later stages in some critical patients.^[18,56] Secondary infection is uncommon and may occur in immunocompromised patients manifesting as superimposed bacterial bronchopneumonia, as described in literature.^[26,48,49,51,56]

Neutrophil extracellular traps showing partially degenerated neutrophils entrapped in fibers lying in close association with CD4+ mononuclear cells aggregates are mentioned in a report by Fox *et al.*^[48] Rarely, interstitial fibrosis, usually of mild-to-moderate degree, and inflammation, mainly of lymphocytic, may be identified.^[18,26,42,48,50,51,54-58]

Ancillary Testing

Few studies have used special stains to highlight the findings seen on hematoxylin and eosin stain or immunohistochemistry (IHC) for typing and distribution of lymphocytic infiltrate and electron microscopy for virus particles. Masson trichrome stains are used for the demonstration of fibrin deposition, pulmonary interstitial fibrosis, and thickening of the vessel wall.^[18,48] IHC markers employed in different studies were CD3, CD4, CD8, CD20, CD79a, CD5, CD38, CD68, CD61, CD45, CD68, CD61, TTF1, p40, Ki67, SARS-CoV-2 antigen, and Rp3 NP protein of SARS-CoV-2. Results of IHC showed the presence of both T and B cells with a mixture of CD4+ and CD8+ lymphocytes in the peribronchiolar region and interstitium, whereas the perivascular area mainly had CD4+ lymphocytes.^[18,48] CD68 and CD61 positivity was seen in macrophages and megakaryocytes, respectively.^[42,48,50] SARS-CoV-2 antigen can be identified in alveolar epithelial cells and macrophages.^[55,58] Electron microscopy has demonstrated viral particles ranging between 60 and 120 nm with viral projections of about 13 nm in length which were localized along the plasmalemmal membranes, in the cytoplasmic vacuoles of pneumocytes, and bronchial mucosal cells.^[49,52,53,58]

Conclusion

Various clinical, biochemical, and radiological parameters suggest multiorgan pathology in COVID-19. Lungs being the primary site for COVID-19 pathology are currently under extensive study, and recent literature have shown DAD as the most consistent finding along with platelet fibrin thrombo-emboli in pulmonary vessels. Increased D-dimer levels indicate the ongoing inflammatory and hypercoagulable state in these patients and represent poor prognostic factor associated with high mortality rate. Being a novel virus, not much is known about its pathogenesis,

treatment, and prevention, hence fast-paced research activities are warranted for optimal management of these cases.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

References

- Mahase E. China coronavirus: WHO declares international emergency as death toll exceeds 200. *BMJ* 2020;368:m408.
- Guo YR, Cao QD, Hong ZS, Tan YY, Chen SD, Jin HJ, et al. The origin, transmission and clinical therapies on coronavirus disease 2019 (COVID-19) outbreak-an update on the status. *Mil Med Res* 2020;7:11.
- Bernheim A, Mei X, Huang M, Yang Y, Fayad ZA, Zhang N, et al. Chest CT findings in coronavirus disease-19 (COVID-19): Relationship to duration of infection. *Radiology* 2020;295:685-91.
- Available from: <https://www.who.int/emergencies/diseases/novel-coronavirus-2019>. [Last accessed on 2020 May 30].
- Wang L, Wang Y, Ye D, Liu Q. Review of the 2019 novel coronavirus (SARS-CoV-2) based on current evidence. *Int J Antimicrob Agents*. 2020; 55:105948.
- Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 2020;395:497-506.
- Jin Y, Yang H, Ji W, Wu W, Chen S, Zhang W, et al. Virology, Epidemiology, Pathogenesis, and Control of COVID-19. *Viruses*. 2020;12: 372.
- World Health Organization. Middle East Respiratory Syndrome Coronavirus (MERS-CoV). Available from: <https://www.who.int/emergencies/mers-cov/en/>. [Last accessed on 2020 Apr 30].
- Liu J, Zheng X, Tong Q, Li W, Wang B, Sutter K, et al. Overlapping and discrete aspects of the pathology and pathogenesis of the emerging human pathogenic coronaviruses SARS-CoV, MERS-CoV, and 2019-nCoV. *J Med Virol* 2020;92:491-4.
- Wan Y, Shang J, Graham R, Baric RS, Li F. Receptor recognition by the novel coronavirus from Wuhan: an analysis based on decade-long structural studies of SARS coronavirus. *J Virol* 2020; 94:e00127-20.
- Yin Y, Wunderink RG. MERS, SARS and other coronaviruses as causes of pneumonia. *Respirology* 2018;23:130-7.
- Wu F, Zhao S, Yu B, Chen YM, Wang W, Song ZG, et al. A new coronavirus associated with human respiratory disease in China. *Nature* 2020;579:265-9.
- Wu YC, Chen CS, Chan YJ. The outbreak of COVID-19: An overview. *J Chin Med Assoc* 2020;83:217-20.
- Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, Herrler T, Erichsen S, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell* 2020;181:271-80.e8.
- Zhou P, Yang XL, Wang XG, Hu B, Zhang L, Zhang W, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature* 2020;579:270-3.
- Donoghue M, Hsieh F, Baronas E, Godbout K, Gosselin M, Stagliano N, et al. A novel angiotensin-converting enzyme-related carboxypeptidase (ACE2) converts angiotensin I to angiotensin 1-9. *Circ Res* 2000;87:E1-9.
- Zou X, Chen K, Zou J, Han P, Hao J, Han Z. Single-cell RNA-seq data analysis on the receptor ACE2 expression reveals the potential risk of different human organs vulnerable to 2019-nCoV infection. *Front Med* 2020;14:185-92.
- Luo W, Yu H, Gou J, Li X, Sun Y, Li J, et al. Clinical pathology of critical patient with novel coronavirus pneumonia (COVID-19). Preprints. 2020, 2020020407. [Available from: <https://www.preprints.org/manuscript/202002.0407/v1>]. [Last accessed 2020 May 30].
- Belouzard S, Chu VC, Whittaker GR. Activation of the SARS coronavirus spike protein via sequential proteolytic cleavage at two distinct sites. *Proc Natl Acad Sci U S A* 2009;106:5871-6.
- Perlman S, Netland J. Coronaviruses post-SARS: Update on replication and pathogenesis. *Nat Rev Microbiol* 2009;7:439-50.
- de Wit E, van Doremalen N, Falzarano D, Munster VJ. SARS and MERS: Recent insights into emerging coronaviruses. *Nat Rev Microbiol* 2016;14:523-34.
- Fu Y, Cheng Y, Wu Y. Understanding SARS-CoV-2-mediated inflammatory responses: From mechanisms to potential therapeutic tools. *Viol Sin* 2020;35:266-71.
- Yang M. Cell Pyroptosis, a Potential Pathogenic Mechanism of 2019-nCoV infection. Available from: <https://ssrn.com/abstract=3527420>. [Last accessed on 2020 Jan 29].
- Savarin C, Bergmann CC. Fine tuning the cytokine storm by IFN and IL-10 following neurotropic coronavirus encephalomyelitis. *Front Immunol* 2018;9:3022.
- Takada A, Kawaoka Y. Antibody-dependent enhancement of viral infection: Molecular mechanisms and *in vivo* implications. *Rev Med Virol* 2003;13:387-98.
- Dolnikoff M, Duarte-Neto AN, de Almeida Monteiro RA, Ferraz da Silva LF, Pierre de Oliveira E, et al. Pathological evidence of pulmonary thrombotic phenomena in severe COVID-19. *J Thromb Haemost* 2020;18:1517-9.
- McGonagle D, O'Donnell JS, Sharif K, Emery P, Bridgewood C. Immune mechanisms of pulmonary intravascular coagulopathy in COVID-19 pneumonia. *Lancet Rheumatol* 2020;2:e437-e445.
- Tang N, Li D, Wang X, Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. *J Thromb Haemost* 2020;18:844-7.
- Lodigiani C, Iapichino G, Carenzo L, Cecconi M, Ferrazzi P, Sebastian T, et al. Venous and arterial thromboembolic complications in COVID-19 patients admitted to an academic hospital in Milan, Italy. *Thromb Res* 2020;191:9-14.
- Helms J, Tacquard C, Severac F, Leonard-Lorant I, Ohana M, Delabranche X, et al. High risk of thrombosis in patients with severe SARS-CoV-2 infection: A multicenter prospective cohort study. *Intensive Care Med* 2020;46:1089-98.
- Wichmann D, Sperhake JP, Lütgehetmann M, Steure S, Edler C, Heineman A, et al. Autopsy findings and venous thromboembolism in patients with COVID-19. *Ann Intern Med* 2020; 173: 268-277.
- Marietta M, Ageno W, Artoni A, De Candia E, Gresele P, Marchetti M, et al. COVID-19 and haemostasis: A position paper from Italian Society on Thrombosis and Haemostasis (SISET). *Blood Transfus* 2020;18:167-9.
- Li Q, Guan X, Wu P, Wang X, Zhou L, Tong Y, et al. Early transmission dynamics in Wuhan, China, of novel coronavirus-infected pneumonia. *N Engl J Med* 2020;382:1199-207.
- Rothan HA, Byrareddy SN. The epidemiology and pathogenesis of coronavirus disease (COVID-19) outbreak. *J Autoimmun* 2020;109:102433.
- The Novel Coronavirus Pneumonia Emergency Response

- Epidemiology Team, Chinese Center of Disease Control and Prevention. The epidemiological characteristics of an outbreak of 2019 novel coronavirus diseases (COVID-19) in China. *Zhonghua Liu Xing Bing Xue Za Zhi* 2020;41:145-51.
36. Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, *et al.* Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: A retrospective cohort study. *Lancet* 2020;395:1054-62.
 37. Yang X, Yu Y, Xu J, Shu H, Xia J, Liu H, *et al.* Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: A single-centered, retrospective, observational study. *Lancet Respir Med* 2020;8:475-81.
 38. Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, *et al.* Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China. *JAMA* 2020;323:1061-9.
 39. Pascarella G, Strumia A, Piliago C, Bruno F, Del Buono R, Costa F, *et al.* COVID-19 diagnosis and management: a comprehensive review. *J Intern Med.* 2020;288:192-206.
 40. Boraschi P. COVID-19 Pulmonary Involvement: Is Really an Interstitial Pneumonia? *Acad Radiol* 2020;27:900.
 41. Guan CS, Lv ZB, Yan S, Du YN, Chen H, Wei LG, *et al.* Imaging features of coronavirus disease 2019 (COVID-19): Evaluation on thin-section CT. *Acad Radiol* 2020;27:609-13.
 42. Barton LM, Duval EJ, Stroberg E, Ghosh S, Mukhopadhyay S. COVID-19 autopsies, Oklahoma, USA. *Am J Clin Pathol* 2020;153:725-33.
 43. Roosen J, Frans E, Wilmer A, Knockaert DC, Bobbaers H. Comparison of premortem clinical diagnoses in critically ill patients and subsequent autopsy findings. *Mayo Clin Proc* 2000;75:562-7.
 44. Tejerina EE, Padilla R, Abril E, Frutos-Vivar F, Ballen A, Rodríguez-Barbero JM, *et al.* Autopsy-detected diagnostic errors over time in the intensive care unit. *Hum Pathol.* 2018;76:85-90.
 45. Osborn M, Lucas S, Stewart R, Ben Swift B, Youd E. Royal College of Pathologist. Briefing on COVID-19. Autopsy Practice Relating to Possible Cases of COVID-19 (2019-nCov, novel coronavirus from China 2019/2020); 2020. Available from: <https://www.rcpath.org/uploads/assets/d5e28baf-5789-4b0f-acecfe370eee6223/fe8fa85a-f004-4a0c-81ee4b2b9cd12cbf/Briefing-on-COVID-19-autopsy-Feb-2020>. [Last accessed 2020 May 30].
 46. Hanley B, Lucas SB, Youd E, Swift B, Osborn M. Autopsy in suspected COVID-19 cases. *J Clin Pathol* 2020;73:239-42.
 47. Monteiro RAA, Duarte-Neto AN, Silva LFF, Oliveira EP, Filho JT, Santos GAB, *et al.* Ultrasound-guided minimally invasive autopsies: A protocol for the study of pulmonary and systemic involvement of COVID-19. *Clinics.* 2020;75:e1972.
 48. Fox SE, Akmatbekov A, Harbert JL, Li G, Brown JQ, Vander Heide RS. Pulmonary and Cardiac Pathology in Covid-19: The First Autopsy Series from New Orleans. medRxiv; 2020. DOI: 10.1101/2020.04.06.20050575. [Last accessed on 2020 Apr 15].
 49. Bradley BT, Maioli H, Johnson R, Chaudhry I, Fink SL, Xu H, *et al.* Histopathology and ultrastructural findings of fatal COVID-19 infections in Washington State: a case series. *Lancet* 2020;396:320-32.
 50. Buja LM, Wolf D, Zhao B, Akkanti B, McDonald M, Lelenwa L, *et al.* Emerging spectrum of cardiopulmonary pathology of the coronavirus disease 2019 (covid-19): Report of three autopsies from Houston, Texas and review of autopsy findings from other United States cities. *Cardiovasc Pathol* 2020;48:107233.
 51. Menter T, Haslbauer JD, Nienhold R, Savic S, Hopfer H, Deigendesch N, *et al.* Postmortem examination of COVID-19 patients reveals diffuse alveolar damage with severe capillary congestion and variegated findings in lungs and other organs suggesting vascular dysfunction. *Histopathology* 2020;77:198-209.
 52. Grimes Z, Bryce C, Sordillo EM, Gordon RE, Reidy J, Paniz Mondolfi AE, *et al.* Fatal pulmonary thromboembolism in SARS-CoV-2-infection. *Cardiovasc Pathol* 2020;48:107227.
 53. Carsana L, Sonzogni A, Nasr A, Rossi R, Pellegrinelli A, Zerbi P, *et al.* Pulmonary post-mortem findings in a series of COVID-19 cases from northern Italy: a two-centre descriptive study. *Lancet Infect Dis.* 2020; 20:1135-1140.
 54. Tian S, Hu W, Niu L, Liu H, Xu H, Xiao SY. Pulmonary pathology of early-phase 2019 novel coronavirus (COVID-19) pneumonia in two patients with lung cancer. *J Thorac Oncol* 2020;15:700-4.
 55. Zhang H, Zhou P, Wei Y, Yue H, Wang Y, Hu M, *et al.* Histopathologic changes and SARS-CoV-2 immunostaining in the lung of a patient with COVID-19. *Ann Intern Med* 2020;172:629-32.
 56. Tian S, Xiong Y, Liu H, Niu L, Guo J, Liao M, *et al.* Pathological study of the 2019 novel coronavirus disease (COVID-19) through postmortem core biopsies. *Mod Pathol* 2020;33:1007-14.
 57. Xu Z, Shi L, Wang Y, Zhang J, Huang L, Zhang C, *et al.* Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *Lancet Respir Med* 2020;8:420-2.
 58. Yao XH, Li TY, He ZC, Ping YF, Liu HW, Yu SC, *et al.* A pathological report of three COVID-19 cases by minimal invasive autopsies. *Zhonghua Bing Li Xue Za Zhi* 2020;49:411-7.
 59. Nicholls JM, Poon LL, Lee KC, Ng WF, Lai ST, Leung CY, *et al.* Lung pathology of fatal severe acute respiratory syndrome. *Lancet* 2003;361:1773-8.
 60. Ding Y, Wang H, Shen H, Li Z, Geng J, Han H, *et al.* The clinical pathology of severe acute respiratory syndrome (SARS): A report from China. *J Pathol* 2003;200:282-9.