

Article

A Comparison of Gene Expression Changes in the Blood of Individuals Consuming Diets Supplemented with Olives, Nuts or Long-Chain Omega-3 Fatty Acids

Virginie Bottero and Judith A. Potashkin *

Center for Neurodegenerative Disease and Therapeutics, The Chicago Medical School, Rosalind Franklin University of Medicine and Science, North Chicago, IL 60064, USA; virginie.bottero@rosalindfranklin.edu

* Correspondence: judy.potashkin@rosalindfranklin.edu; Tel.: +1-847-578-8677; Fax: +1-847-578-8515

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Abstract: Background: The Mediterranean diet, which is rich in olive oil, nuts, and fish, is considered healthy and may reduce the risk of chronic diseases. Methods: Here, we compared the transcriptome from the blood of subjects with diets supplemented with olives, nuts, or long-chain omega-3 fatty acids and identified the genes differentially expressed. The dietary genes obtained were subjected to network analysis to determine the main pathways, as well as the transcription factors and microRNA interaction networks to elucidate their regulation. Finally, a gene-associated disease interaction network was performed. Results: We identified several genes whose expression is altered after the intake of components of the Mediterranean diets compared to controls. These genes were associated with infection and inflammation. Transcription factors and miRNAs were identified as potential regulators of the dietary genes. Interestingly, caspase 1 and sialophorin are differentially expressed in the opposite direction after the intake of supplements compared to Alzheimer's disease patients. In addition, ten transcription factors were identified that regulated gene expression in supplemented diets, mild cognitive impairment, and Alzheimer's disease. Conclusions: We identified genes whose expression is altered after the intake of the supplements as well as the transcription factors and miRNAs involved in their regulation. These genes are associated with schizophrenia, neoplasms, and rheumatic arthritis, suggesting that the Mediterranean diet may be beneficial in reducing these diseases. In addition, the results suggest that the Mediterranean diet may also be beneficial in reducing the risk of dementia.

Keywords: nutrigenomic; olive; nut; fish; Mediterranean diet; nuclear factor interleukin 3 regulated; *NFIL3*; dementia; Alzheimer's disease

1. Introduction

Dietary patterns are associated with different disease risks. Whereas the Western diet increases the risk of cardiovascular disease and some types of cancers, other diets have some beneficial effects on health. The Mediterranean diet is rich in the consumption of fruits, vegetables, olive oil, fish, and nuts. It is characterized by the intake of food rich in polyphenols, monounsaturated fatty acids, and polyunsaturated fatty acids. It has been proposed that some components of the Mediterranean diet are beneficial for the individual's health. Based on observation as well as randomized controlled studies, the Mediterranean diet is proposed to reduce the risk of developing several diseases. A beneficial effect of the diet on cardiovascular risks has been observed (reviewed in [1]), along with a decrease in blood pressure [2–4]. In addition, the Mediterranean diet is proposed to be beneficial for patients with diabetes mellitus by improving glycogenic regulation in patients [1,5,6]. Furthermore, several observational studies found that the Mediterranean diet reduced the risk of developing several cancers, such as colorectal, breast, stomach,

liver, and head and neck cancer [1,7]. Moreover, Mediterranean diets may improve an individual's resilience to Parkinson's disease, depression, and dementia [1].

The beneficial effects of some of the components of the Mediterranean diet have been investigated. The main source of fat in the diet is olive oil. Olive oil contains high levels of monounsaturated fatty acids, as well as other biologically active components such as polyphenol [8]. Many studies have shown the benefits of olive oil on cardiovascular risk factors [9,10] and the benefits for patients with type 2 diabetes [5,11]. Olive oil also helps control the levels of plasma lipids and glucose in patients with metabolic syndrome [12]. Additionally, olive oil may be protective for some forms of cancer [13–15]. It has been suggested that the health benefits of olive oil are related to its anti-hypertensive, anti-inflammatory, and antioxidant effects [11,16–18].

Nuts, another component of the Mediterranean diet, are good sources of monosaturated fatty acids, polyunsaturated fatty acids, and other nutrients such as fibers, vitamin E, and L-arginine. Walnuts, for example, are rich in long-chain omega-3 fatty acids (omega-3). Numerous epidemiologic studies have illuminated the beneficial impact of nut consumption on health outcomes (reviewed in [19]). Indeed, the consumption of nuts has been shown to have a positive effect on patients suffering from obesity, hypertension, diabetes mellitus, and cardiovascular diseases [20–24]. Nuts can reduce oxidative stress, inflammation, and blood pressure while helping with glycemic control [25–28].

Finally, fish consumption, characteristically high in a Mediterranean diet, has been proposed to have potential health benefits. Omega-3 are essential fatty acids that are found in fish oils. These n-3 fatty acids are composed of two crucial components: polyunsaturated fatty acids eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA). Studies have highlighted the beneficial effect of fish consumption and omega-3 on cardiovascular diseases, stroke, atherosclerosis as well as insulin resistance [29,30]. One of the possible mechanisms proposed for the health benefits was a decrease in plasma bioactive lipid components involved in insulin resistance and inflammation [31,32].

Microarray and high-throughput technologies for gene expression are essential tools for identifying differential patterns of gene expression that are characteristic of environmental determinants of health. The use of peripheral blood mononuclear cells (PBMC) to determine the effect of diet has shown that food intake can modify the blood's proteome, metabolome, and gene expression profile [33–35]. Because PBMC can be easily and repeatedly collected, compared to liver, muscles, and adipocyte tissue, they are used frequently to study the impact of diet [35], including that of olive oil, olive leaves, nuts, and omega-3 consumption [36–40].

In this study, we took an integrative network-based approach to identify the genes that may be responsible for the beneficial effects of Mediterranean diets. Previously, we used a similar approach to reveal some of the mechanistic pathways involved in the development of Parkinson's diseases, Alzheimer's disease, and other dementias [41–44]. Among the dietary genes identified, we observed that the Nuclear Factor Interleukin 3 Regulated (*NFIL3*) was downregulated after the intake of all three supplements. In addition, Interleukin 8 (*IL8*), the Serine/Threonine Kinase 17b (*STK17B*), and Serpin Family B Member 2 (*SERPINB2*), and the Regulator of G Protein Signaling 1 (*RGS1*) were downregulated after two supplements were included in the diet. Pathway analysis determined that these genes were associated with infection and inflammation, suggesting potential health benefits of the Mediterranean diet in modulating the immune system. Gene-transcription factor and gene-miRNA network analyses identified important factors involved in the expression of dietary genes. Interestingly, we observed some shared transcription factors involved in the regulation of supplemented diets and dementia, indicating that the supplements may also be beneficial in reducing the risk of Alzheimer's disease.

2. Materials and Methods

2.1. Analysis of PBMC Transcriptomic Studies

We used the curated database BaseSpace Correlation Engine (BSCE, Illumina, Inc., San Diego, CA, USA) to search for gene expression studies in different human diets [45]. Using the search

terms “Mediterranean diet”, “olive diet”, “nut diet”, “fish oil”, “EPA”, “DHA”. “blood”, “human”, “RNA”, and “microarray”, we identified several studies with PMBC from people under different diets. Only human microarray studies with 5 samples or more for cases and controls and curated in BSCE were considered for analysis. In addition, analysis from PMBC collected from obese patients was not included in this analysis. Six microarrays met our inclusion criteria as of 1 October 2019. A description of microarray datasets included in this study is provided in Section 3.1. The arrays from olive oil and olive leaves diets were defined as “olive diet”, whereas the arrays with EPA/DHA were defined as “omega-3 diet”.

The differentially expressed genes were curated by BSCE. Statistical analyses were performed on log scale data. In the parametric test, variances were not assumed equal (Welch *t*-test). A *p*-value cutoff of 0.05 and a fold-change of 1.2 was applied to generate the final list of genes. Genes whose mean normalized test and control intensities are both less than the 20th percentile of the combined normalized signal intensities were removed. Final gene expression data from microarray studies were downloaded from BSCE (Table S1). A Venn diagram analysis was performed with the genes up or downregulated in the three olive diet arrays and the four fish oil arrays independently. The transcription factors Venn diagram was created using the website <http://bioinformatics.psb.ugent.be/webtools/Venn/>. Only genes that were differentially expressed in at least two olive diet arrays and at least two fish oil studies were included for further analysis. The list of genes can be found in Table S2.

2.2. Pathway Enrichment Analysis

Official gene symbols for the genes identified in the supplemented diets gene expression comparison were imported into NetworkAnalyst for pathway analyses (<https://www.networkanalyst.ca/NetworkAnalyst/faces/home.xhtml>) for pathway analyses using the Kyoto Encyclopedia of Genes and Genome (KEGG) pathway database [46]. NetworkAnalyst uses an enrichment network OverRepresentation Analyses (ORA). ORA is a statistical technique to identify gene sets or pathways that have a significant overlap with the selected genes of interest. In NetworkAnalyst, hypergeometric tests are used to compute the *p*-values [47,48].

2.3. Gene-Transcription Factors Interaction Analysis

Gene-transcription factors interactome was performed in NetworkAnalyst. Transcription factor and gene target data were derived from the Encyclopedia of DNA Elements (ENCODE) ChIP-seq data, ChIP Enrichment Analysis (ChEA), or JASPAR database [49–51]. ENCODE uses the BETA Minus Algorithm in which only a peak intensity signal <500 and the predicted regulatory potential score <1 is used. ChEA transcription factor targets the database inferred from integrating literature curated Chip-X data. JASPAR is an open-access database of curated, non-redundant transcription factor (TF)-binding profiles. A Venn diagram analysis was performed with the transcription factors identified with each database. Transcription factors were ranked according to network topology measurements including degree and betweenness centrality.

2.4. Gene-miRNA Interaction Analysis

The gene-miRNA interactome was performed in NetworkAnalyst. The Gene-miRNA Interactome was carried out from comprehensive experimentally validated miRNA-gene interaction data collected from TarBase and miRTarBase [52–54].

2.5. Gene-Disease Association Analysis

Gene-disease association analysis was performed in NetworkAnalyst. The literature curated gene-disease association information was collected from the DisGeNET database, a publicly available collection of genes and variants associated with human diseases [55].

2.6. Diet and Dementia Analysis

Our previous gene expression analysis identified 91 mild cognitive impairment (MCI) genes dysregulated in in the 2 arrays analyzed and 387 Alzheimer's disease (AD) genes dysregulated in at least 2 out of the 4 arrays analyzed [42]. A Venn diagram analysis was carried out between the dementia genes previously identified and the genes identified in the present diet study using the InteractiVenn website (<http://www.interactivenn.net/>).

3. Results

3.1. Gene Expression Comparison of the Diet Supplements

We first identified common genes differentially expressed in several diet studies. The datasets, platforms, and test samples for each study are listed in Table 1. The overall strategy of the study is presented in Figure 1.

Three PBMC microarray datasets from olive-supplemented diets (olive oil or olive leaves) were identified in the BaseSpace Correlation Engine (BSCE) (GSE28358, GSE75025, and GSE87300). We used Venn diagram analysis to identify genes shared among the datasets from olive-based diets (Figure 2a,b, Table S1). A total of 38 genes were differentially expressed in at least two out of the three olive-supplemented diet arrays (Table S2). Similarly, four PBMC microarrays from omega-3-supplemented diet datasets (fish oil or DHA + EPA supplements) were obtained from BSCE and analyzed by (GSE48368, GSE48368, GSE12375, and E-MTAB-48) (Figure 2c,d, Table S1). A total of 36 differentially expressed genes were shared in at least two out of the four datasets analyzed (Table S2). Only one microarray dataset from a diet rich in nuts was available (GSE28358) and 365 genes were identified as differentially regulated (Table S2).

Table 1. Gene expression datasets used in this study. The peripheral blood mononuclear cells (PBMC) transcriptomic studies that were selected for analysis are presented in the table. Age is indicated in years as mean \pm SD, or mean alone, or mean (range) depending of the information available. EPA+DHA: eicosapentaenoic acid and docosahexaenoic acid.

Diets	Datasets	Platform	Sample #	Age	Sex (%F)	Health	Reference
Mediterranean diet + Olive oil	GSE28358	GPL571 [HG-U133A_2] Affymetrix Human Genome U133A 2.0 Array	12	62 \pm 8	45	High risk of coronary artery disease	[36]
Olive oil	GSE75025	GPL10558 Illumina HumanHT-12 V4.0 expression beadchip	12	29 \pm 2	50	Healthy	[37]
Olive leaf extract	GSE87300	GPL13667 [HG-U219] Affymetrix Human Genome U219 Array	15	32	0	Healthy	[38]
Mediterranean diet + Nuts	GSE28358	GPL571 [HG-U133A_2] Affymetrix Human Genome U133A 2.0 Array	10	63 \pm 6	45	High risk of coronary artery disease	[36]
Fish oil 3 weeks	GSE48368	GPL10558 Illumina HumanHT-12 V4.0 expression beadchip	17	27.2 \pm 6.9	71	Healthy	[39]
Fish oil 7 weeks	GSE48368	GPL10558 Illumina HumanHT-12 V4.0 expression beadchip	17	27.2 \pm 6.9	71	Healthy	[39]
EPA + DHA	GSE12375	GPL7144 NuGO array (human) NuGO_Hs1a520180	23	69.9 (67-76)	35	Healthy	[40]
EPA + DHA	E-MTAB-48	A-AFFY-111—Affymetrix Custom Array	23	69.9 (67-76)	35	Healthy	[40]

Next, we compared the genes differentially expressed in the diets. Interestingly, we found that the Nuclear Factor Interleukin 3 Regulated (*NFIL3*) was downregulated in the three types of diets. In addition, we observed that Interleukin 8 (*IL8*), the Serine/Threonine Kinase 17b (*STK17B*), and Serpin

Family B Member 2 (*SERPINB2*) were downregulated in diets rich in olive and nuts. Additionally, the Regulator of G Protein Signaling 1 (*RGS1*) was downregulated in diets rich in nuts and omega-3. We did not find any upregulated genes shared between the diets.

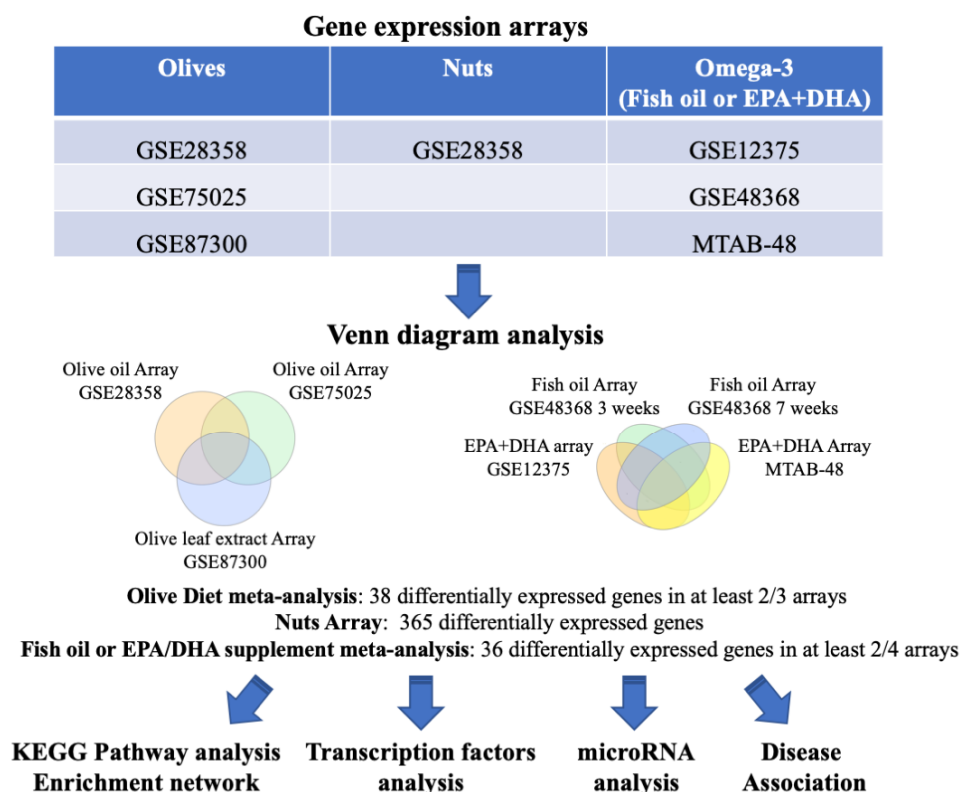


Figure 1. Flowchart of the study. The BaseSpace Correlation Engine (BSCE) was searched to identify microarray data from appropriate diet studies. Venn diagram analysis was used to identify shared differentially regulated genes. The genes shared by olive-, nuts- or omega-3-supplemented diets were analyzed for shared functional pathways, transcription factors, miRNAs regulation, and disease associations.

3.2. Pathway Enrichment Analysis

A pathway analysis using NetworkAnalyst was performed to elucidate the functional and biological role of genes differentially expressed in the different diets. The pathway enrichment network analysis was performed using the Kyoto Encyclopedia of Genes and Genome (KEGG) database (Figure 3, Table S3). The 38 olive supplemented diet genes identified 31 pathways. The top 10 pathways were bladder cancer, Hepatitis B, ErbB signaling, NOD-like receptor signaling, MAPK signaling, Kaposi's sarcoma-associated herpesvirus infection, ubiquitin-mediated proteolysis, epithelial cell signaling in *Helicobacter pylori* infection, pertussis, and salmonella infection. 48 pathways were identified from the genes differentiated regulated during a diet rich in nuts. The top 10 pathways were notch signaling, thyroid hormone signaling, hepatitis B, pathways in cancer, oxytocin signaling, Kaposi's sarcoma-associated herpesvirus infection, endocrine resistance, phosphatidylinositol signaling, and melanogenesis. Finally, nine pathways were identified from the 36 differentially expressed genes in omega-3 rich diets. These pathways are longevity regulating, insulin resistance, 5' AMP-activated protein kinase (AMPK) signaling, forkhead transcription factor family (FoxO) signaling, cyclic guanosine monophosphate-protein kinase G (cGMP-PKG) signaling, herpes simplex infection, primary immunodeficiency, vasopressin-regulated water reabsorption, type II diabetes mellitus, and intestinal immune network for IgA production.

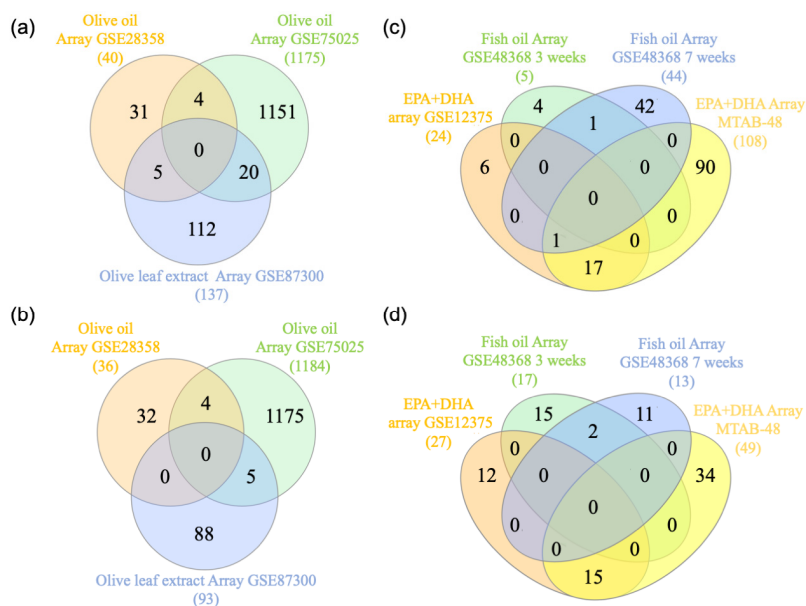


Figure 2. Venn diagram analysis of the genes up and downregulated in olive- and omega-3-supplemented diets. (a) and (b) . Genes in olive-supplemented diets: The genes downregulated (a) and upregulated (b) in the olive oil or leaf extracts arrays were downloaded from BSCE and used to create a Venn diagram using the following website <http://www.interactivenn.net/>. (c) and (d). Genes in Omega-3-supplemented diets: The genes downregulated (c) and upregulated (d) in the fish oil or eicosapentaenoic acid (EPA)/docosahexaenoic acid (DHA) studies were downloaded from BSCE and used to create the Venn diagrams.

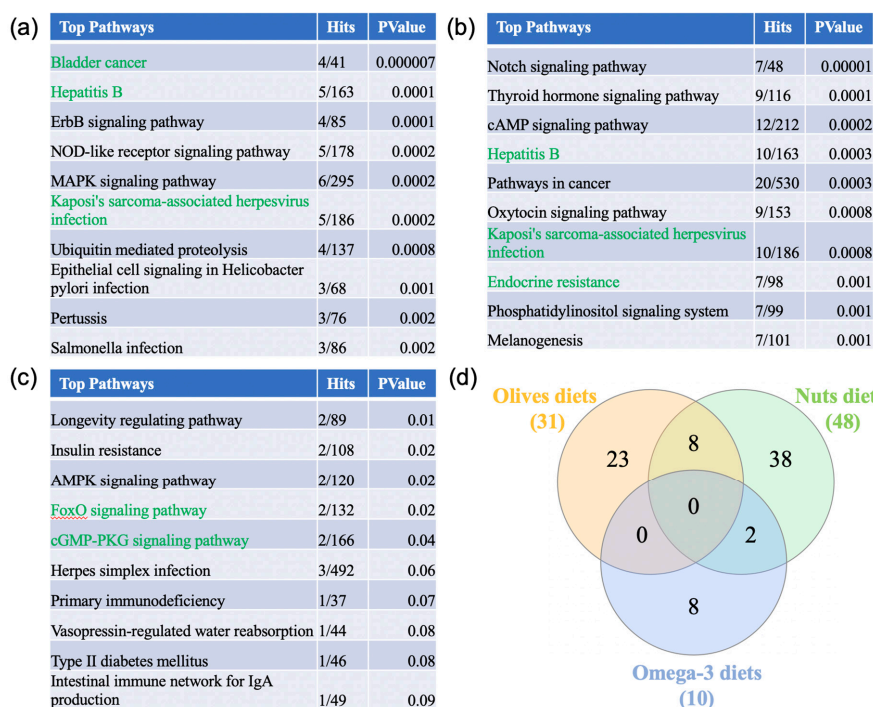


Figure 3. Pathway analysis. (a–c) The genes differentially expressed in 2 out of 3 olive supplement arrays, in the nuts supplement array, in at least 2 out of the 4 arrays from fish or EPA + DHA diets were obtained using Venn analysis. The gene lists were uploaded to <https://www.networkanalyst.ca/NetworkAnalyst/faces/home.xhtml>, where a list enrichment network analysis was performed using the KEGG database. The 10 tops pathways identified from the olive-, nuts-, or omega-3-supplemented diets are listed in (a), (b), and (c) respectively. (d) The pathways shared between the three types of Mediterranean diets were analyzed in a Venn diagram analysis.

Venn analysis showed that no pathway was shared by all three supplements. Moreover, we did not observe any pathways shared between the olive-based and omega-3-based diets. However, eight pathways were shared between the olive-based and nut-rich diets. These pathways were bladder cancer, Hepatitis B, Kaposi’s sarcoma-associated herpesvirus infection, Influenza A, proteoglycans in cancer, HTLV-I infection, endocrine resistance, and Chagas disease. In addition, we observed that cGMP-PKG signaling and FoxO signaling pathways were shared between the nuts and omega-3 diets. Interestingly several of the pathways associated with the different diets are associated with infection and inflammation.

3.3. Gene-Transcription Factors Interaction Analysis

To identify key regulators of the genes differentially expressed in the different types of diets, gene transcription factor interactomes were performed on NetworkAnalyst using three different databases (ENCODE, ChEA, and JASPAR) (Table S4). The transcription factors that were shared by all the databases were identified by Venn analysis (Figure 4a–c). The analysis identified 14, 24, and 15 transcription factors in the olive-, nuts-, and omega-3-supplemented diets, respectively. The list of these transcription factors is presented in Figure 4d. A total of 10 transcription factors shared among the three types of diets included CREB1, EGR1, ELK1, GATA2, GATA3, PPARG, RELA, STAT1, STAT3, and YY1. Eight transcription factors were identified in at least two analyses. HNF4A was shared between olive- and omega-3-supplemented diets. CTCF, IRF1, REST, and SREBF2 were shared between the nuts- and omega-3-supplemented diets. Finally, ARNT, CEBPB, and SREBF1 were shared between the olive- and nuts-supplemented diets.

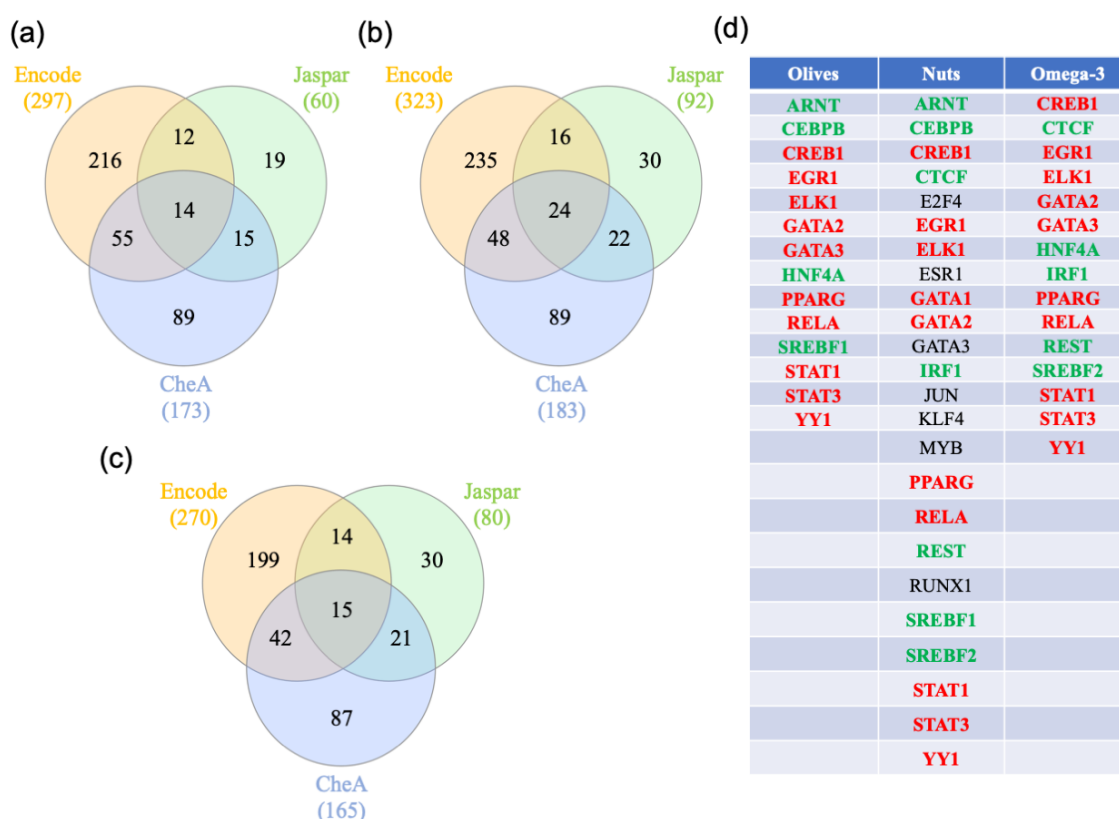


Figure 4. Transcription factor analysis. (a–c) Transcription factor analysis for each diet. The gene lists were uploaded to <https://www.networkanalyst.ca/NetworkAnalyst/faces/home.xhtml>. The gene–transcription factor interaction network was performed with ENCODE, ChEA, and JASPAR, and (a), (b), and (c) represent the results of the Venn analysis performed with olive-, nuts- and omega-3-genes, respectively. The transcription factors interacting with the supplement-regulated genes are listed in (d). Transcription factors in red are shared between the three supplements, the transcription factors in green are shared between two supplements, and the transcription factors in black are unique to a supplement.

3.4. Gene-miRNA Interaction Analysis

To further understand the regulation of the expression of genes differentially expressed during diet supplementation, a gene-miRNA interaction network analysis was performed in NetworkAnalyst. Comprehensive experimentally validated miRNA-gene interaction data were collected from TarBase and miRTarBase. 304, 819, and 166 miRNAs were identified from an olive-rich diet, a nuts-supplemented diet, and an omega-3-supplemented diet, respectively (Figure 5 and Table S5). Interestingly, 100 miRNAs were shared in the three types of supplemented diets (Figure 5). In order to determine the most important miRNA, we performed the Venn analysis after selecting the top miRNAs using a degree cut off of five for the olive-rich and omega-3-supplemented diets and a degree cut off of 25 for the nuts-supplemented diet. This allowed us to identify 17, 19, and 4 miRNAs from an olive-rich diet, a nuts-supplemented diet, and an omega-3-supplemented diet, respectively (Table S5). Interestingly, three miRNAs were shared between the three diets (mir-93-5p, mir-17-5p, and mir-335-5p).

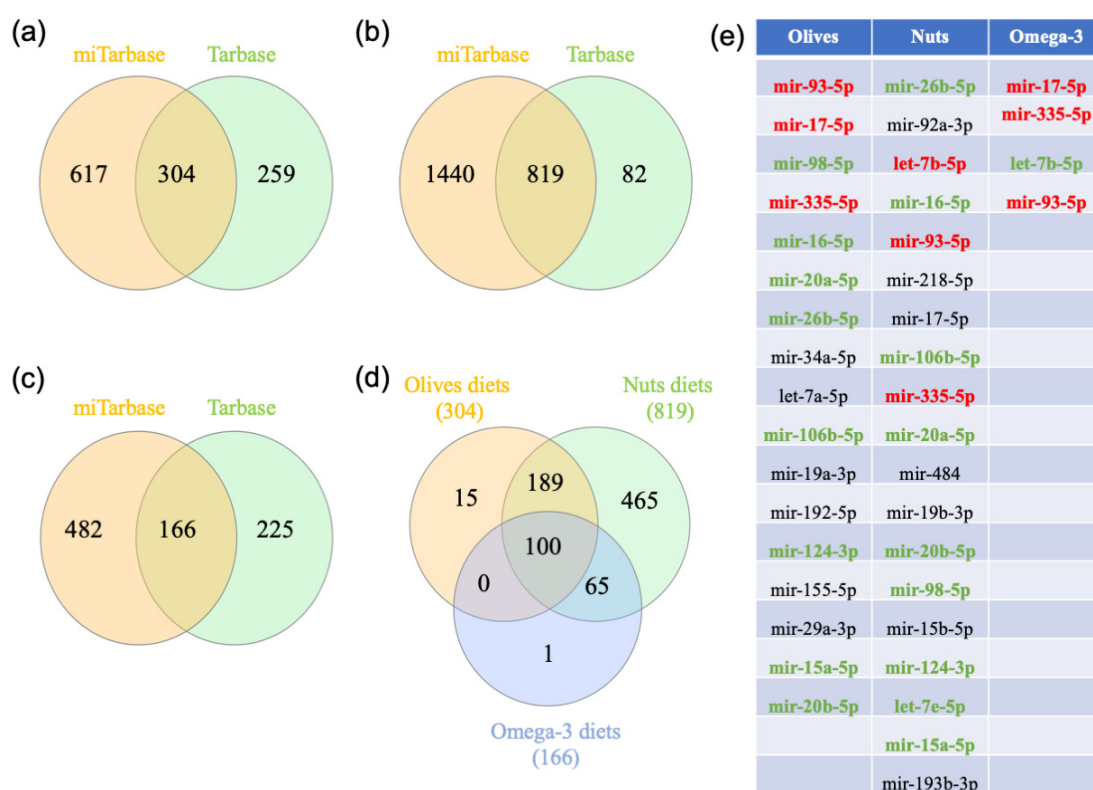


Figure 5. MicroRNA analysis. The diet gene lists were uploaded to <https://www.networkanalyst.ca/NetworkAnalyst/faces/home.xhtml>. A gene-microRNA interactome was performed using the TarBase and miRTarBase databases. (a), (b), and (c) represent the results of the Venn analysis performed with olive-, nuts- and omega-3-genes, respectively. (d) The miRNA shared between the three types of Mediterranean diets were analyzed in a Venn diagram analysis. The most significant miRNAs are listed in (e). MiRNAs in red are shared between the 3 supplements, the miRNAs in green are shared between 2 supplements, and the miRNAs in black are unique to a supplement.

3.5. Gene-Disease Association Analysis

A gene-disease association network analysis was performed in NetworkAnalyst. The differentially expressed genes in the olive-, nuts- or omega-3-rich diets allowed for the identification of 231, 1099, and 9 associated diseases, respectively. These diseases were ranked by decreasing degree followed by decreasing betweenness (Figure 6 and Table S6). Interestingly, six associated diseases were shared between the three Mediterranean diet components: schizophrenia, mammary neoplasms, prostatic neoplasms, neoplasm metastasis, endometrial neoplasms, and rheumatoid arthritis.

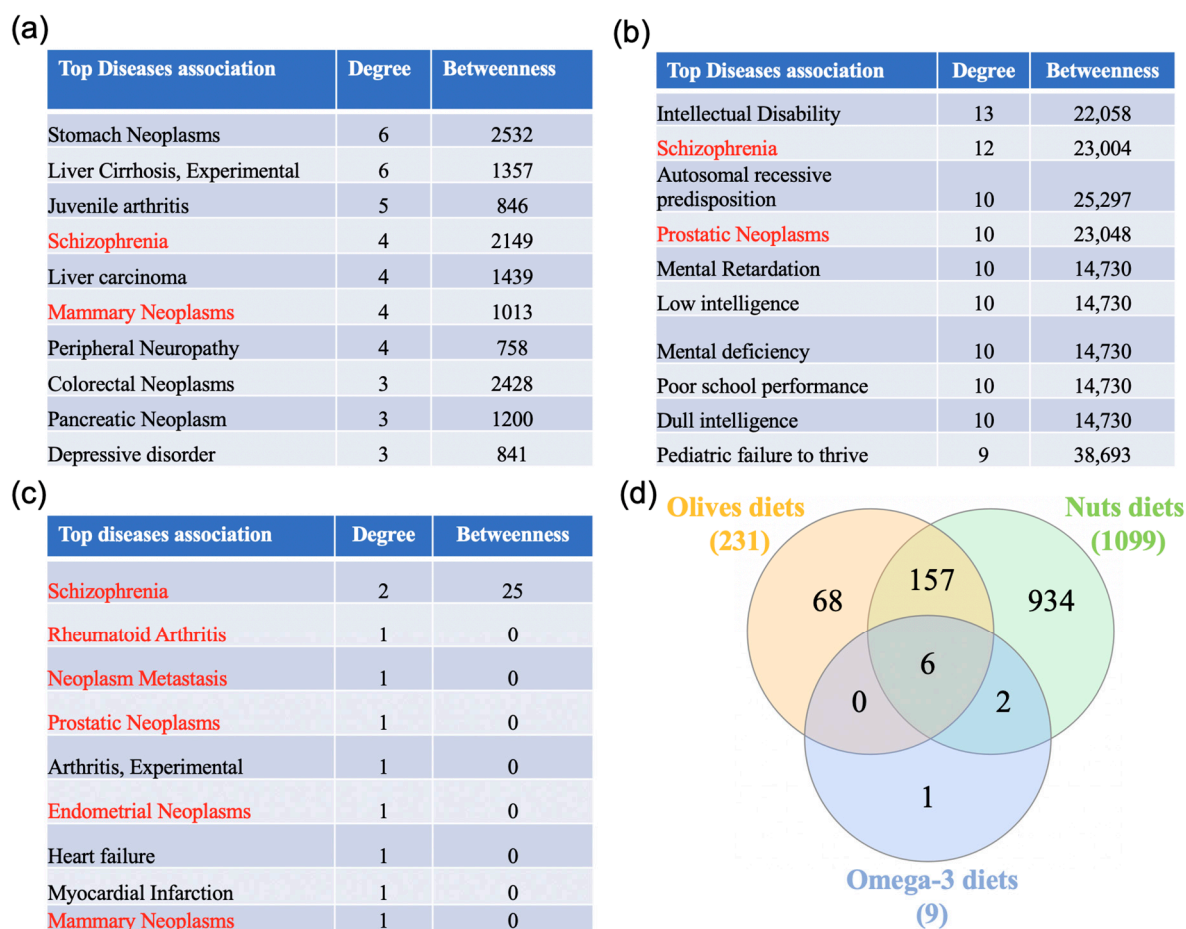


Figure 6. Disease association analysis. (a–c) The diet genes lists were uploaded to <https://www.networkanalyst.ca/NetworkAnalyst/faces/home.xhtml> to perform the disease association analysis. The diseases were ranked by decreasing degree followed by decreasing betweenness, and the top 10 diseases obtained from the olive-, nuts, and omega-2-supplemented diets are presented in (a), (b), and (c), respectively. (d) A Venn analysis was performed to determine the diseases shared between the different types of Mediterranean diets. The associated diseases in red are shared between the three supplements.

3.6. Diets and Dementia Analysis

In our previous study, we performed a gene expression comparison of publicly available arrays from blood samples obtained from mild cognitive impairment (MCI) and Alzheimer’s disease (AD) patients [42]. We obtained 91 MCI genes dysregulated in the two arrays analyzed and 387 AD genes dysregulated in at least two out of the four arrays analyzed. Interestingly, unhealthy eating habits might increase the risk to develop dementia [56] and numerous studies have highlighted the potential beneficial impact of the Mediterranean diet on cognition (reviewed in [57,58]). For example, nuts consumption has been shown to delay cognitive decline in aging populations and improves AD pathology [59,60]. Recently, the impact of a Mediterranean diet and the microbiota on neurodegeneration has been reviewed [61].

We compared the genes dysregulated in MCI and AD patients to the genes identified in this study that are differentially expressed after diet supplementation [42]. Caspase 1 (*CASP1*) was shared between a diet supplemented with olives and AD patients (Figure 7a). In addition, four genes were shared between the diet supplemented with nuts and AD patients, including Actin-Related Protein 2/3 Complex Subunit 3 (*ARPC3*), Sialophorin (*SPN*), Neurobeachin Like 2 (*NBEAL2*), and Mixed-Lineage Leukemia (*MLL*) (Figure 7b). No genes were shared between a diet rich in omega-3 and MCI and AD

patients (Figure 7c). Interestingly, *CASP1* and *SPN* are regulated in dementia and the supplemented diets in the opposite direction, indicating that the supplements may be beneficial in reducing the risk of dementia. We observed that *CASP1* expression was upregulated in the olive-supplemented diet, whereas it was downregulated in the blood of AD patients. On the other hand, the expression of Sialophorin (*SLN*), also known as Leukosialin and CD43, was downregulated in a nut-rich diet, whereas it is upregulated in AD. In the blood, *SLN* is expressed at the surface of T cells and regulates multiple T cell functions. In the brain, *SLN* is expressed on the surface of microglia. Whereas our study indicated an upregulation in blood from AD patients, *SLN* protein expression was downregulated at the surface of microglial cells [62]. We also performed a pathways enrichment analysis between the genes identified in the supplemented diets and the genes identified in our previous dementia analysis. We did not find any shared pathways.

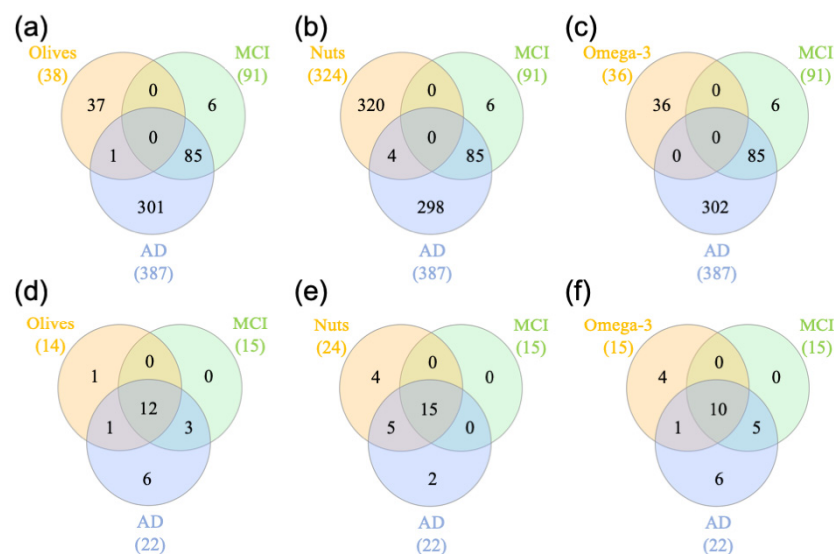


Figure 7. Comparison of gene expression data from supplemented diets to mild cognitive impairment (MCI) and Alzheimer's disease (AD). (a–c) The shared genes between a specific supplemented diet, MCI, and AD were analyzed by Venn (a): olive-supplemented diet, (b): nuts-supplemented diet, (c): omega-3-supplemented diet. (d–f) The shared transcription factors between a specific diet, MCI, and AD were analyzed by Venn (d): Olive-supplemented diet, (e): Nuts-supplemented diet, (f): omega-3-supplemented diet.

Next, we compared the transcription factors obtained in the different diets to the previously identified transcription factors involved in MCI and AD regulation [42]. We observed a significant overlap between the diet and dementia analysis (Figure 7). A total of 12 transcription factors identified in the olive-supplemented diet were also identified in the dementia analysis (PPARG, EGR1, CREB1, ELK1, YY1, GATA2, GATA3, STAT1, CEBPB, RELA, STAT3, and SREBF1) (Figure 7d). A total of 15 transcription factors were shared between a nuts-supplemented diet and dementia (ELK1, EGR1, PPARG, YY1, STAT1, CREB1, GATA2, GATA3, CEBPB, RELA, STAT3, SREBF1, E2F4, JUN, and RUNX1) (Figure 7e). Finally, 10 transcription factors were regulating both the omega-3-supplemented diet and dementia genes (PPARG, YY1, ELK1, CREB1, GATA2, GATA3, EGR1, RELA, STAT1, and STAT3) (Figure 7f). Together, we identified 10 transcription factors that are involved in the regulation of the three different types of supplemented diets, MCI and AD (PPARG, EGR1, CREB1, ELK1, YY1, GATA2, GATA3, STAT1, RELA, and STAT3).

4. Discussion

4.1. Regulation of Gene Expression by Components of a Mediterranean Diet

In this study, we identified several differentially expressed genes that are shared between the various diets. The main finding is that the transcriptional repressor *NFIL3*, also known as E4 binding protein 4 E4BP4, was downregulated in all three supplemented Mediterranean diets. *NFIL3* can regulate the expression of several cytokines and is involved in the development of immune cells [63]. In addition, *NFIL3* is an important regulator of the circadian clock, which acts by repressing the expression of *PER1* and *PER2* [64]. Interestingly, the expression of *NFIL3* is regulated by nutrients. Indeed, the expression of *NFIL3* is activated by insulin and feeding, whereas fasting decreases its expression [65]. *NFIL3* can repress the expression of *FGF21*, a protein with anti-diabetic, and triglyceride-lowering properties [65]. In intestinal epithelial cells, *NFIL3* has been shown to regulate lipid storage and body composition in mice [66]. *NFIL3* also regulates the signaling processes involved in heart functions [67], and, in the brain, *NFIL3* blocks neuronal regeneration by competing with the transcription factor *CREB* [68]. Decreasing expression of *NFIL3* using siRNAs induced neurite outgrowth in a rat neuronal model [68].

IL8 is downregulated in diets supplemented with olive oil and nuts. *IL8* is a key mediator of inflammation. This cytokine functions as a chemoattractant allowing target cells, such as neutrophils, to migrate towards a site of infection. Interestingly, *IL8* is one of the immunological signatures of excess body weight [69,70]. *IL8* might play a role in some obesity-related metabolic complications [69]. Adherence to a Mediterranean diet was shown to reduce the plasma levels of *IL8* and to delay atheroma plaque development [71]. Wine, consumed in moderate quantity, could also decrease the level of *IL8*. Different varieties of grapes, as well as their phenolic compounds, have been shown to reduce *IL8* levels [72].

The serine-threonine kinase *STK17B*, also known as Death-Associated Protein Kinase-Related 2 (*DRAK2*), is downregulated in both diets rich in olive and nuts. Free fatty acid (FFA) increases the expression of *STK17B*, which participates in the apoptosis of islet β -cells [73]. *STK17B* has been proposed as a novel target in the treatment of diabetes. Indeed, knockdown of *STK17B* by siRNA attenuated FFA-induced islet β cells apoptosis [73]. In addition, *STK17B* upregulation was observed by pro-inflammatory cytokines such as *IL-1 β* , and *TNF- α* . The downregulation of *STK17B* was able to prevent the inflammatory-induced cell death of insulinoma cells [74]. The chemical inhibition of *STK17B* has been proposed as a potential diabetes treatment [75]. Together, these studies suggested that the downregulation of *STK17B* could be involved in some of the health benefits of the Mediterranean diet.

SERPINB2, also known as plasminogen Activator Inhibitor 2 (*PAI2*), is downregulated in both diets rich in olive and nuts. The coagulation factor *SERPINB2* inactivates the tissue plasminogen activator and urokinase. The role of *SERPINB2* in diets is largely unknown. It has been shown that, in mice fed a high-fat diet, *SERPINB2* promotes adipose tissue development [76]. Moreover, it has been shown that *SERPINB2* expression is decreased in a methionine-supplemented diet [77].

RGS1, a regulator of the G-protein superfamily, is downregulated in both diets rich in nuts and omega-3. Polymorphic variants in *RGS1* have been linked to chronic inflammatory diseases such as celiac disease, multiple sclerosis, and type I diabetes [78–80]. *RGS1* was upregulated in the epididymal white adipose tissue of high-fat diet-fed mice [81]. In addition, *RGS1* expression was upregulated in both obese adipose tissue and atherosclerotic aortae models [82]. *RGS1* was upregulated in both PBMC and brain samples from Alzheimer's disease patients [83]. Altogether, whereas *RGS1* is upregulated in different diseases, its expression is decreased with Mediterranean diets.

4.2. Gene Expression Regulation by Transcription Factors and miRNA in Mediterranean Diets

Performing a gene-transcription factor interaction network, we identified 10 transcription factors that might regulate the genes differentially expressed in Mediterranean diets. The transcription factor *CREB1* is increased by listroside, a purified olive secoirodoid derivative [84]. It has been proposed that listroside could ameliorate the mitochondrial function in an Alzheimer's disease cell model [84].

In addition, CREB1 is proposed as an early biomarker candidate for obesity-induced pathophysiological changes in the colon [85]. PPARG is considered a key regulator of lipid metabolism [86]. It has been shown that a polymorphism in PPARG was associated with the effect of diet on the health of the individual [87]. Hydroxytyrosol, a component of olive oil, was shown to downregulate STAT3 in K562 cells [88]. Polyphenol extract from olive oil has also been shown to inhibit STAT3 in a model of rheumatoid arthritis [89]. Polymorphism in the *STAT3* gene might interact with higher saturated fat intake and increases the risk of abdominal obesity [90].

MicroRNAs are crucial in the regulation of gene expression and have been implicated in several diseases including metabolic disorders and insulin resistance (reviewed in [91]). We also performed a network analysis to reveal the microRNAs involved in the regulation of the differentially expressed genes by the supplemented diets and observed three shared microRNAs (miR-17-5p, miR-355-5p, and miR-93-5p). MiR-17-5p is involved in angiogenesis, proliferation, apoptosis, and autophagy [92,93], and it plays a role in several cancers, including hepatocellular carcinoma, osteosarcoma, leukemia, lung, gastric, colorectal, prostate, and breast cancer [93]. MiR-17-5p levels increase after high-fat diet consumption and activate adipogenic differentiation [94]. Interestingly, the plant polyphenol curcumin decreases miR-17-5p levels and inhibits adipogenesis [94]. However, another study on retinal inflammation indicated that high-fat diets for 8 weeks induced obesity and insulin resistance, as well as, decreased miR-17-5p [95]. Endoplasmic reticulum stress was proposed to trigger the reduction of miR-17-5p [95]. MiR-17-5p reduces inflammation and lipid accumulation in an atherosclerosis model [96]. Finally, a role for miR-17-5p in aging has been proposed [97]. Surprisingly, whereas several miRNAs are decreased in aging brains, an increase in miR-17-5p levels was observed [98]. However, in neurodegenerative diseases such as Alzheimer's disease, miR-17-5p expression is inhibited and this may be responsible for an increase in APP protein levels [99]. Flavonoids are components of Mediterranean diets that inhibit oxidative stress and neuroinflammation. MiR-355-5p was one of the miRNAs potentially regulated after exposure to flavonoids [100]. In a dietary methionine restriction mouse model, miR-355-5p was elevated in the bone marrow and might be involved in osteoblast differentiation and function [101]. Finally, an increase of miR-355-5p was observed in white adipose tissue of ob/ob mice and mice on a high-fat diet [102]. Altogether, these studies indicate that miR-355-5p might be involved in the network of genes differentially regulated by the Mediterranean diet and, consequently, in the cellular effect of this diet. Little is known about the impact of diets on miR-93-5p; however, its expression increases in coronary artery disease patients, suggesting the potential use of this miRNA as a diagnostic marker [103].

4.3. Mediterranean Diets and Disease Association

Schizophrenia was identified as one of the six diseases associated with olive-, nuts-, and omega-3-rich diets. Schizophrenia patients have a shorter lifespan due to metabolic and cardiovascular disease and often practice unhealthy dietary habits [104–106]. Improving the diet of Schizophrenic patients could be very beneficial [107]. For example, eating a Mediterranean diet improved cardiovascular risks in Schizophrenic patients [108]. In addition, the omega-3 fatty acid supplement was beneficial for Schizophrenic patients by improving their psychopathology, reducing tardive dyskinesia, and attenuating the risk of conversion to psychosis in patients [109–111].

We also observed that rheumatoid arthritis is associated with all three types of supplements used in this study. Compared to a Western diet, eating a Mediterranean diet reduces inflammatory activity, improves physical function, and vitality [112]. In a rat model of rheumatoid arthritis, the gavage of hydroxytyrosol, a typical virgin olive oil phenolic compound, decreased both acute and chronic inflammation [113]. Rheumatoid arthritis patients consume less monounsaturated fatty acids than healthy individuals [114]. Monounsaturated fatty acids are a component of the Mediterranean diet, and, therefore, it may be beneficial for these patients [114]. Although several trials and systemic reviews show that a Mediterranean diet might improve the patient's condition, further clinical studies are necessary to recommend the use of this diet as an adjunct therapy to standard treatment [115–117].

An inverse correlation between the consumption of a Mediterranean diet and cancer risk has been proposed for several neoplasms, including breast and prostate [118–121]. However, recent meta-analyses suggested that further investigations are needed for a better assessment [122,123]. Moreover, it is interesting to note that, in addition to possibly decreasing the risk of cancer development, the Mediterranean diet may improve the quality of life of a cancer survivor [124,125].

4.4. Limitations

This comparison of gene expression using diets with different supplements associated with Mediterranean diets is based on publicly available microarray data, and, therefore, several limitations should be considered. For example, the sample size could influence the power of the analysis. Moreover, the studies were performed at different sites, following different protocols, and these differences might also influence the results. The present study was performed using the statistical criteria described in the methods section with the datasets that were available at the time of the analysis. The gene expression data were curated using the database BSCE. The differential expression of genes at the lower end of the thresholds was applied (1.2 FC, $p < 0.05$) in order to produce a larger set of genes for the initial analysis. These criteria may produce a higher likelihood of false positives. Alternatively, future studies could use more stringent thresholds initially to reduce the number of false positives. It is also important to note that datasets are updated routinely and thus analysis of the data with newer datasets may yield different results. In addition, the results may be influenced by the participant themselves. Their genetics, age, sex and disease risk status, as well as their lifestyle, can influence gene expression. To improve the power of this gene expression comparison, we had to combine data from individuals who had different characteristics (Table 1). Future studies with many more participants will be needed to determine if the Mediterranean diet influences gene regulation differently in young and aging populations, males and females, and in healthy participants and those with a high risk of cardiovascular disease. In addition, it would be interesting to analyze the effect of Mediterranean diet on populations of individuals with different health conditions such as cancer, dementia, or metabolic disorders to determine if there is a benefit. Finally, whereas the individual component of the Mediterranean diet was used for this study, it would be interesting to determine the effect of the full Mediterranean diet and to determine if a synergistic effect of the component would be beneficial.

5. Conclusions

Comparing gene expression profiles, we determined that *NFIL3*, *IL8*, *STK17B*, *SERPINB2*, and *RGS* were differentially expressed by supplementation of different components of the Mediterranean diets. These genes were associated with infection and inflammation. The Mediterranean differentiated genes were regulated by several key transcription factors (HNF4A, IRF1, REST, CTCF, and SREBF2) and miRNA (miR-17-5p, miR-335-5p, miR-93-5p). Finally, we determined several diseases for which the Mediterranean diet could be beneficial (schizophrenia, several neoplasms, rheumatoid arthritis, and dementia).

Supplementary Materials: The following are available online at <http://www.mdpi.com/2072-6643/12/12/3765/s1>, Table S1. Genes down and up-regulated in the selected arrays; Table S2. Genes shared in the different diets; Table S3. Enrichment pathways; Table S4. Gene-Transcription factor analysis; Table S5. miRNA analysis; Table S6. Disease association analysis.

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References

1. Sanchez-Sanchez, M.L.; Garcia-Vigara, A.; Hidalgo-Mora, J.J.; Garcia-Perez, M.A.; Tarin, J.; Cano, A. Mediterranean Diet and Health: A Systematic Review of Epidemiological Studies and Intervention Trials. *Maturitas* **2020**, *136*, 25–37. [[CrossRef](#)] [[PubMed](#)]
2. Nunez-Cordoba, J.M.; Valencia-Serrano, F.; Toledo, E.; Alonso, A.; Martinez-Gonzalez, M.A. The Mediterranean Diet and Incidence of Hypertension: The Seguimiento Universidad de Navarra (SUN) Study. *Am. J. Epidemiol.* **2009**, *169*, 339–346. [[CrossRef](#)]
3. Bendinelli, B.; Masala, G.; Bruno, R.M.; Caini, S.; Saieva, C.; Boninsegni, A.; Ungar, A.; Ghiadoni, L.; Palli, D. A Priori Dietary Patterns and Blood Pressure in the EPIC Florence Cohort: A Cross-Sectional Study. *Eur. J. Nutr.* **2019**, *58*, 455–466. [[CrossRef](#)]
4. De Pergola, G.; D'Alessandro, A. Influence of Mediterranean Diet on Blood Pressure. *Nutrients* **2018**, *10*, 1700. [[CrossRef](#)]
5. Tosatti, J.A.G.; Alves, M.T.; Gomes, K.B. The Role of the Mediterranean Dietary Pattern on Metabolic Control of Patients with Diabetes Mellitus: A Narrative Review. *Adv. Exp. Med. Biol.* **2020**, *1307*, 115–128. [[CrossRef](#)] [[PubMed](#)]
6. Salas-Salvado, J.; Guasch-Ferre, M.; Lee, C.H.; Estruch, R.; Clish, C.B.; Ros, E. Protective Effects of the Mediterranean Diet on Type 2 Diabetes and Metabolic Syndrome. *J. Nutr.* **2015**, *146*, 920S–927S. [[CrossRef](#)] [[PubMed](#)]
7. Schwingshackl, L.; Schwedhelm, C.; Galbete, C.; Hoffmann, G. Adherence to Mediterranean Diet and Risk of Cancer: An Updated Systematic Review and Meta-Analysis. *Nutrients* **2017**, *9*, 1063. [[CrossRef](#)]
8. Sanchez-Rodriguez, E.; Biel-Glesson, S.; Fernandez-Navarro, J.R.; Calleja, M.A.; Espejo-Calvo, J.A.; Gil-Extremuera, B.; de la Torre, R.; Fito, M.; Covas, M.I.; Vilchez, P.; et al. Effects of Virgin Olive Oils Differing in Their Bioactive Compound Contents on Biomarkers of Oxidative Stress and Inflammation in Healthy Adults: A Randomized Double-Blind Controlled Trial. *Nutrients* **2019**, *11*, 561. [[CrossRef](#)]
9. Bendinelli, B.; Masala, G.; Saieva, C.; Salvini, S.; Calonico, C.; Sacerdote, C.; Agnoli, C.; Grioni, S.; Frasca, G.; Mattiello, A.; et al. Fruit, Vegetables, and Olive Oil and Risk of Coronary Heart Disease in Italian Women: The EPICOR Study. *Am. J. Clin. Nutr.* **2011**, *93*, 275–283. [[CrossRef](#)]
10. Romani, A.; Ieri, F.; Urciuoli, S.; Noce, A.; Marrone, G.; Nediani, C.; Bernini, R. Health Effects of Phenolic Compounds Found in Extra-Virgin Olive Oil, by-Products, and Leaf of *Olea europaea* L. *Nutrients* **2019**, *11*, 1776. [[CrossRef](#)]
11. Mazzocchi, A.; Leone, L.; Agostoni, C.; Pali-Scholl, I. The Secrets of the Mediterranean Diet. Does [Only] Olive Oil Matter? *Nutrients* **2019**, *11*, 2941. [[CrossRef](#)] [[PubMed](#)]
12. Esposito, K.; Marfella, R.; Ciotola, M.; Di Palo, C.; Giugliano, F.; Giugliano, G.; D'Armiento, M.; D'Andrea, F.; Giugliano, D. Effect of a Mediterranean-Style Diet on Endothelial Dysfunction and Markers of Vascular Inflammation in the Metabolic Syndrome: A Randomized Trial. *JAMA* **2004**, *292*, 1440–1446. [[CrossRef](#)] [[PubMed](#)]
13. Menendez, J.A.; Lupu, R. Mediterranean Dietary Traditions for the Molecular Treatment of Human Cancer: Anti-Oncogenic Actions of the Main Olive Oil's Monounsaturated Fatty Acid Oleic Acid (18:1n-9). *Curr. Pharm. Biotechnol.* **2006**, *7*, 495–502. [[CrossRef](#)]
14. Psaltopoulou, T.; Kostis, R.I.; Haidopoulos, D.; Dimopoulos, M.; Panagiotakos, D.B. Olive Oil Intake is Inversely Related to Cancer Prevalence: A Systematic Review and a Meta-Analysis of 13,800 Patients and 23,340 Controls in 19 Observational Studies. *Lipids Health Dis.* **2011**, *10*, 127. [[CrossRef](#)] [[PubMed](#)]
15. Fogli, S.; Arena, C.; Carpi, S.; Polini, B.; Bertini, S.; Digiaco, M.; Gado, F.; Saba, A.; Saccomanni, G.; Breschi, M.C.; et al. Cytotoxic Activity of Oleocanthal Isolated from Virgin Olive Oil on Human Melanoma Cells. *Nutr. Cancer* **2016**, *68*, 873–877. [[CrossRef](#)] [[PubMed](#)]
16. Massaro, M.; Scoditti, E.; Carluccio, M.A.; Calabriso, N.; Santarpino, G.; Verri, T.; De Caterina, R. Effects of Olive Oil on Blood Pressure: Epidemiological, Clinical, and Mechanistic Evidence. *Nutrients* **2020**, *12*, 1548. [[CrossRef](#)]
17. Bogani, P.; Galli, C.; Villa, M.; Visioli, F. Postprandial Anti-Inflammatory and Antioxidant Effects of Extra Virgin Olive Oil. *Atherosclerosis* **2007**, *190*, 181–186. [[CrossRef](#)]

18. Mitjavila, M.T.; Fandos, M.; Salas-Salvado, J.; Covas, M.I.; Borrego, S.; Estruch, R.; Lamuela-Raventos, R.; Corella, D.; Martinez-Gonzalez, M.A.; Sanchez, J.M.; et al. The Mediterranean Diet Improves the Systemic Lipid and DNA Oxidative Damage in Metabolic Syndrome Individuals. A Randomized, Controlled, Trial. *Clin. Nutr.* **2013**, *32*, 172–178. [[CrossRef](#)]
19. De Souza, R.G.M.; Schincaglia, R.M.; Pimentel, G.D.; Mota, J.F. Nuts and Human Health Outcomes: A Systematic Review. *Nutrients* **2017**, *9*, 1311. [[CrossRef](#)]
20. Souza, R.G.; Gomes, A.C.; Naves, M.M.; Mota, J.F. Nuts and Legume Seeds for Cardiovascular Risk Reduction: Scientific Evidence and Mechanisms of Action. *Nutr. Rev.* **2015**, *73*, 335–347. [[CrossRef](#)]
21. Jackson, C.L.; Hu, F.B. Long-Term Associations of Nut Consumption with Body Weight and Obesity. *Am. J. Clin. Nutr.* **2014**, *100*, 408S–411S. [[CrossRef](#)] [[PubMed](#)]
22. Mohammadifard, N.; Salehi-Abargouei, A.; Salas-Salvado, J.; Guasch-Ferre, M.; Humphries, K.; Sarrafzadegan, N. The Effect of Tree Nut, Peanut, and Soy Nut Consumption on Blood Pressure: A Systematic Review and Meta-Analysis of Randomized Controlled Clinical Trials. *Am. J. Clin. Nutr.* **2015**, *101*, 966–982. [[CrossRef](#)] [[PubMed](#)]
23. Vigiuliouk, E.; Kendall, C.W.; Blanco Mejia, S.; Cozma, A.I.; Ha, V.; Mirrahimi, A.; Jayalath, V.H.; Augustin, L.S.; Chiavaroli, L.; Leiter, L.A.; et al. Effect of Tree Nuts on Glycemic Control in Diabetes: A Systematic Review and Meta-Analysis of Randomized Controlled Dietary Trials. *PLoS ONE* **2014**, *9*, e103376. [[CrossRef](#)] [[PubMed](#)]
24. Blanco Mejia, S.; Kendall, C.W.; Vigiuliouk, E.; Augustin, L.S.; Ha, V.; Cozma, A.I.; Mirrahimi, A.; Maroleanu, A.; Chiavaroli, L.; Leiter, L.A.; et al. Effect of Tree Nuts on Metabolic Syndrome Criteria: A Systematic Review and Meta-Analysis of Randomised Controlled Trials. *BMJ Open* **2014**, *4*, e004660. [[CrossRef](#)] [[PubMed](#)]
25. Lopez-Uriarte, P.; Nogues, R.; Saez, G.; Bullo, M.; Romeu, M.; Masana, L.; Tormos, C.; Casas-Agustench, P.; Salas-Salvado, J. Effect of Nut Consumption on Oxidative Stress and the Endothelial Function in Metabolic Syndrome. *Clin. Nutr.* **2010**, *29*, 373–380. [[CrossRef](#)]
26. Lorenzon dos Santos, J.; Quadros, A.S.; Weschenfelder, C.; Garofallo, S.B.; Marcadenti, A. Oxidative Stress Biomarkers, Nut-Related Antioxidants, and Cardiovascular Disease. *Nutrients* **2020**, *12*, 682. [[CrossRef](#)]
27. Bitok, E.; Sabate, J. Nuts and Cardiovascular Disease. *Prog. Cardiovasc. Dis.* **2018**, *61*, 33–37. [[CrossRef](#)]
28. Parham, M.; Heidari, S.; Khorramirad, A.; Hozoori, M.; Hosseinzadeh, F.; Bakhtyari, L.; Vafaeimanesh, J. Effects of Pistachio Nut Supplementation on Blood Glucose in Patients with Type 2 Diabetes: A Randomized Crossover Trial. *Rev. Diabet. Stud.* **2014**, *11*, 190–196. [[CrossRef](#)]
29. Sokola-Wysoczanska, E.; Wysoczanski, T.; Wagner, J.; Czyz, K.; Bodkowski, R.; Lochynski, S.; Patkowska-Sokola, B. Polyunsaturated Fatty Acids and Their Potential Therapeutic Role in Cardiovascular System Disorders—A Review. *Nutrients* **2018**, *10*, 1561. [[CrossRef](#)]
30. Kris-Etherton, P.M.; Richter, C.K.; Bowen, K.J.; Skulas-Ray, A.C.; Jackson, K.H.; Petersen, K.S.; Harris, W.S. Recent Clinical Trials Shed New Light on the Cardiovascular Benefits of Omega-3 Fatty Acids. *Methodist Debaquey Cardiovasc. J.* **2019**, *15*, 171–178.
31. Lankinen, M.; Schwab, U.; Erkkila, A.; Seppanen-Laakso, T.; Hannila, M.L.; Mussalo, H.; Lehto, S.; Uusitupa, M.; Gylling, H.; Oresic, M. Fatty Fish Intake Decreases Lipids Related to Inflammation and Insulin Signaling—A Lipidomics Approach. *PLoS ONE* **2009**, *4*, e5258. [[CrossRef](#)]
32. De Mello, V.D.; Erkkila, A.T.; Schwab, U.S.; Pulkkinen, L.; Kolehmainen, M.; Atalay, M.; Mussalo, H.; Lankinen, M.; Oresic, M.; Lehto, S.; et al. The Effect of Fatty or Lean Fish Intake on Inflammatory Gene Expression in Peripheral Blood Mononuclear Cells of Patients with Coronary Heart Disease. *Eur. J. Nutr.* **2009**, *48*, 447–455. [[CrossRef](#)]
33. Kaminski, W.E.; Jendraschak, E.; Kiefl, R.; von Schacky, C. Dietary Omega-3 Fatty Acids Lower Levels of Platelet-Derived Growth Factor mRNA in Human Mononuclear Cells. *Blood* **1993**, *81*, 1871–1879. [[CrossRef](#)]
34. Wittwer, J.; Rubio-Aliaga, I.; Hoefl, B.; Bendik, I.; Weber, P.; Daniel, H. Nutrigenomics in Human Intervention Studies: Current Status, Lessons Learned and Future Perspectives. *Mol. Nutr. Food Res.* **2011**, *55*, 341–358. [[CrossRef](#)]
35. De Mello, V.D.; Kolehmainen, M.; Schwab, U.; Pulkkinen, L.; Uusitupa, M. Gene Expression of Peripheral Blood Mononuclear Cells as a Tool in Dietary Intervention Studies: What Do We Know So Far? *Mol. Nutr. Food Res.* **2012**, *56*, 1160–1172. [[CrossRef](#)]

36. Castaner, O.; Corella, D.; Covas, M.I.; Sorli, J.V.; Subirana, I.; Flores-Mateo, G.; Nonell, L.; Bullo, M.; de la Torre, R.; Portoles, O.; et al. In Vivo Transcriptomic Profile after a Mediterranean Diet in High-Cardiovascular Risk Patients: A Randomized Controlled Trial. *Am. J. Clin. Nutr.* **2013**, *98*, 845–853.
37. D'Amore, S.; Vacca, M.; Cariello, M.; Graziano, G.; D'Orazio, A.; Salvia, R.; Sasso, R.C.; Sabba, C.; Palasciano, G.; Moschetta, A. Genes and miRNA Expression Signatures in Peripheral Blood Mononuclear Cells in Healthy Subjects and Patients with Metabolic Syndrome after Acute Intake of Extra Virgin Olive Oil. *Biochim. Biophys. Acta* **2016**, *1861*, 1671–1680. [[CrossRef](#)]
38. Boss, A.; Kao, C.H.; Murray, P.M.; Marlow, G.; Barnett, M.P.; Ferguson, L.R. Human Intervention Study to Assess the Effects of Supplementation with Olive Leaf Extract on Peripheral Blood Mononuclear Cell Gene Expression. *Int. J. Mol. Sci.* **2016**, *17*, 2019. [[CrossRef](#)]
39. Myhrstad, M.C.; Ulven, S.M.; Gunther, C.C.; Ottestad, I.; Holden, M.; Ryeng, E.; Borge, G.I.; Kohler, A.; Bronner, K.W.; Thoresen, M.; et al. Fish Oil Supplementation Induces Expression of Genes Related to Cell Cycle, Endoplasmic Reticulum Stress and Apoptosis in Peripheral Blood Mononuclear Cells: A Transcriptomic Approach. *J. Intern. Med.* **2014**, *276*, 498–511. [[CrossRef](#)]
40. Bouwens, M.; van de Rest, O.; Dellschaft, N.; Bromhaar, M.G.; de Groot, L.C.; Geleijnse, J.M.; Muller, M.; Afman, L.A. Fish-Oil Supplementation Induces Antiinflammatory Gene Expression Profiles in Human Blood Mononuclear Cells. *Am. J. Clin. Nutr.* **2009**, *90*, 415–424. [[CrossRef](#)]
41. Santiago, J.A.; Bottero, V.; Potashkin, J.A. Dissecting the Molecular Mechanisms of Neurodegenerative Diseases through Network Biology. *Front. Aging Neurosci.* **2017**, *9*, 166. [[CrossRef](#)] [[PubMed](#)]
42. Bottero, V.; Potashkin, J.A. Meta-Analysis of Gene Expression Changes in the Blood of Patients with Mild Cognitive Impairment and Alzheimer's Disease Dementia. *Int. J. Mol. Sci.* **2019**, *20*, 5403. [[CrossRef](#)] [[PubMed](#)]
43. Santiago, J.A.; Bottero, V.; Potashkin, J.A. Transcriptomic and Network Analysis Highlight the Association of Diabetes at Different Stages of Alzheimer's Disease. *Front. Neurosci.* **2019**, *13*, 1273. [[CrossRef](#)] [[PubMed](#)]
44. Santiago, J.A.; Bottero, V.; Potashkin, J.A. Transcriptomic and Network Analysis Identifies Shared and Unique Pathways across Dementia Spectrum Disorders. *Int. J. Mol. Sci.* **2020**, *21*, 2050. [[CrossRef](#)]
45. Kupersmidt, I.; Su, Q.J.; Grewal, A.; Sundaresh, S.; Halperin, I.; Flynn, J.; Shekar, M.; Wang, H.; Park, J.; Cui, W.; et al. Ontology-based meta-analysis of global collections of high-throughput public data. *PLoS ONE* **2010**, *5*, e13066. [[CrossRef](#)]
46. Kanehisa, M.; Goto, S. KEGG: Kyoto Encyclopedia of Genes and Genomes. *Nucleic Acids Res.* **2000**, *28*, 27–30. [[CrossRef](#)]
47. Xia, J.; Gill, E.E.; Hancock, R.E. NetworkAnalyst for Statistical, Visual and Network-Based Meta-Analysis of Gene Expression Data. *Nat. Protoc.* **2015**, *10*, 823–844. [[CrossRef](#)]
48. Zhou, G.; Soufan, O.; Ewald, J.; Hancock, R.E.W.; Basu, N.; Xia, J. NetworkAnalyst 3.0: A Visual Analytics Platform for Comprehensive Gene Expression Profiling and Meta-Analysis. *Nucleic Acids Res.* **2019**, *47*, W234–W241. [[CrossRef](#)]
49. Consortium, E.P. A User's Guide to the Encyclopedia of DNA Elements (ENCODE). *PLoS Biol.* **2011**, *9*, e1001046.
50. Lachmann, A.; Xu, H.; Krishnan, J.; Berger, S.I.; Mazloom, A.R.; Ma'ayan, A. ChEA: Transcription Factor Regulation Inferred from Integrating Genome-Wide ChIP-X Experiments. *Bioinformatics* **2010**, *26*, 2438–2444. [[CrossRef](#)]
51. Khan, A.; Fornes, O.; Stigliani, A.; Gheorghe, M.; Castro-Mondragon, J.A.; van der Lee, R.; Bessy, A.; Cheneby, J.; Kulkarni, S.R.; Tan, G.; et al. JASPAR 2018: Update of the Open-Access Database of Transcription Factor Binding Profiles and Its Web Framework. *Nucleic Acids Res.* **2018**, *46*, D1284. [[CrossRef](#)] [[PubMed](#)]
52. Karagkouni, D.; Paraskevopoulou, M.D.; Chatzopoulos, S.; Vlachos, I.S.; Tastsoglou, S.; Kanellos, I.; Papadimitriou, D.; Kavakiotis, I.; Maniou, S.; Skoufos, G.; et al. DIANA-TarBase v8: A Decade-Long Collection of Experimentally Supported miRNA-Gene Interactions. *Nucleic Acids Res.* **2018**, *46*, D239–D245. [[CrossRef](#)] [[PubMed](#)]
53. Sethupathy, P.; Corda, B.; Hatzigeorgiou, A.G. TarBase: A Comprehensive Database of Experimentally Supported Animal microRNA Targets. *RNA* **2006**, *12*, 192–197. [[CrossRef](#)] [[PubMed](#)]
54. Chou, C.H.; Shrestha, S.; Yang, C.D.; Chang, N.W.; Lin, Y.L.; Liao, K.W.; Huang, W.C.; Sun, T.H.; Tu, S.J.; Lee, W.H.; et al. miRTarBase Update 2018: A Resource for Experimentally Validated microRNA-Target Interactions. *Nucleic Acids Res.* **2018**, *46*, D296–D302. [[CrossRef](#)] [[PubMed](#)]

55. Pinero, J.; Bravo, A.; Queralt-Rosinach, N.; Gutierrez-Sacristan, A.; Deu-Pons, J.; Centeno, E.; Garcia-Garcia, J.; Sanz, F.; Furlong, L.I. DisGeNET: A Comprehensive Platform Integrating Information on Human Disease-Associated Genes and Variants. *Nucleic Acids Res.* **2017**, *45*, D833–D839. [[CrossRef](#)]
56. Samieri, C.; Sonawane, A.R.; Lefevre-Arbogast, S.; Helmer, C.; Grodstein, F.; Glass, K. Using Network Science Tools to Identify Novel Diet Patterns in Prodromal Dementia. *Neurology* **2020**, *94*, e2014–e2025. [[CrossRef](#)]
57. Vinciguerra, F.; Graziano, M.; Hagnas, M.; Frittitta, L.; Tumminia, A. Influence of the Mediterranean and Ketogenic Diets on Cognitive Status and Decline: A Narrative Review. *Nutrients* **2020**, *12*, 1019. [[CrossRef](#)]
58. Roman, G.C.; Jackson, R.E.; Gadhia, R.; Roman, A.N.; Reis, J. Mediterranean Diet: The Role of Long-Chain Omega-3 fatty Acids in Fish; Polyphenols in Fruits, Vegetables, Cereals, Coffee, Tea, Cacao and Wine; Probiotics and Vitamins in Prevention of Stroke, Age-Related Cognitive Decline, and Alzheimer Disease. *Rev. Neurol.* **2019**, *175*, 724–741. [[CrossRef](#)]
59. Sala-Vila, A.; Valls-Pedret, C.; Rajaram, S.; Coll-Adros, N.; Cofan, M.; Serra-Mir, M.; Perez-Heras, A.M.; Roth, L.; Freitas-Simoes, T.M.; Domenech, M.; et al. Effect of a 2-Year Diet Intervention with Walnuts on Cognitive Decline. The Walnuts and Healthy Aging (WAHA) Study: A Randomized Controlled Trial. *Am. J. Clin. Nutr.* **2020**, *111*, 590–600. [[CrossRef](#)]
60. Gorji, N.; Moeini, R.; Memariani, Z. Almond, Hazelnut and Walnut, Three Nuts for Neuroprotection in Alzheimer's Disease: A Neuropharmacological Review of Their Bioactive Constituents. *Pharmacol. Res.* **2018**, *129*, 115–127. [[CrossRef](#)]
61. Gentile, F.; Doneddu, P.E.; Riva, N.; Nobile-Orazio, E.; Quattrini, A. Diet, Microbiota and Brain Health: Unraveling the Network Intersecting Metabolism and Neurodegeneration. *Int. J. Mol. Sci.* **2020**, *21*, 7471. [[CrossRef](#)] [[PubMed](#)]
62. Matsuo, A.; Walker, D.G.; Terai, K.; McGeer, P.L. Expression of CD43 in Human Microglia and Its Downregulation in Alzheimer's Disease. *J. Neuroimmunol.* **1996**, *71*, 81–86. [[CrossRef](#)]
63. Yin, J.; Zhang, J.; Lu, Q. The Role of Basic Leucine Zipper Transcription Factor E4BP4 in the Immune System and Immune-Mediated Diseases. *Clin. Immunol.* **2017**, *180*, 5–10. [[CrossRef](#)] [[PubMed](#)]
64. Mitsui, S.; Yamaguchi, S.; Matsuo, T.; Ishida, Y.; Okamura, H. Antagonistic Role of E4BP4 and PAR Proteins in the Circadian Oscillatory Mechanism. *Genes Dev.* **2001**, *15*, 995–1006. [[CrossRef](#)]
65. Tong, X.; Muchnik, M.; Chen, Z.; Patel, M.; Wu, N.; Joshi, S.; Rui, L.; Lazar, M.A.; Yin, L. Transcriptional Repressor E4-Binding Protein 4 (E4BP4) Regulates Metabolic Hormone Fibroblast Growth Factor 21 (FGF21) during Circadian Cycles and Feeding. *J. Biol. Chem.* **2010**, *285*, 36401–36409. [[CrossRef](#)]
66. Wang, Y.; Kuang, Z.; Yu, X.; Ruhn, K.A.; Kubo, M.; Hooper, L.V. The Intestinal Microbiota Regulates Body Composition through NFIL3 and the Circadian Clock. *Science* **2017**, *357*, 912–916. [[CrossRef](#)]
67. Velmurugan, B.K.; Chang, R.L.; Marthandam Asokan, S.; Chang, C.F.; Day, C.H.; Lin, Y.M.; Lin, Y.C.; Kuo, W.W.; Huang, C.Y. A Minireview of E4BP4/NFIL3 in Heart Failure. *J. Cell Physiol.* **2018**, *233*, 8458–8466. [[CrossRef](#)]
68. MacGillavry, H.D.; Stam, F.J.; Sassen, M.M.; Kegel, L.; Hendriks, W.T.; Verhaagen, J.; Smit, A.B.; van Kesteren, R.E. NFIL3 and cAMP Response Element-Binding Protein Form a Transcriptional Feedforward Loop That Controls Neuronal Regeneration-Associated Gene Expression. *J. Neurosci.* **2009**, *29*, 15542–15550. [[CrossRef](#)]
69. Kim, C.S.; Park, H.S.; Kawada, T.; Kim, J.H.; Lim, D.; Hubbard, N.E.; Kwon, B.S.; Erickson, K.L.; Yu, R. Circulating Levels of MCP-1 and IL-8 Are Elevated in Human Obese Subjects and Associated with Obesity-Related Parameters. *Int. J. Obes.* **2006**, *30*, 1347–1355. [[CrossRef](#)]
70. Sharabiani, M.T.; Vermeulen, R.; Scoccianti, C.; Hosnijeh, F.S.; Minelli, L.; Sacerdote, C.; Palli, D.; Krogh, V.; Tumino, R.; Chiodini, P.; et al. Immunologic Profile of Excessive BODY weight. *Biomarkers* **2011**, *16*, 243–251. [[CrossRef](#)]
71. Casas, R.; Urpi-Sarda, M.; Sacanella, E.; Arranz, S.; Corella, D.; Castaner, O.; Lamuela-Raventos, R.M.; Salas-Salvado, J.; Lapetra, J.; Portillo, M.P.; et al. Anti-Inflammatory Effects of the Mediterranean Diet in the Early and Late Stages of Atheroma Plaque Development. *Mediat. Inflamm.* **2017**, *2017*, 3674390. [[CrossRef](#)] [[PubMed](#)]
72. Colombo, F.; Di Lorenzo, C.; Regazzoni, L.; Fumagalli, M.; Sangiovanni, E.; Peres de Sousa, L.; Bavaresco, L.; Tomasi, D.; Bosso, A.; Aldini, G.; et al. Phenolic Profiles and Anti-Inflammatory Activities of Sixteen Table Grape (*Vitis vinifera* L.) Varieties. *Food Funct.* **2019**, *10*, 1797–1807. [[CrossRef](#)] [[PubMed](#)]

73. Mao, J.; Luo, H.; Wu, J. Drak2 Overexpression Results in Increased Beta-Cell Apoptosis after Free Fatty Acid Stimulation. *J. Cell Biochem.* **2008**, *105*, 1073–1080. [[CrossRef](#)] [[PubMed](#)]
74. Mao, J.; Luo, H.; Han, B.; Bertrand, R.; Wu, J. Drak2 is Upstream of p70S6 Kinase: Its Implication in Cytokine-Induced Islet Apoptosis, Diabetes, and Islet Transplantation. *J. Immunol.* **2009**, *182*, 4762–4770. [[CrossRef](#)]
75. Wang, S.; Xu, L.; Lu, Y.T.; Liu, Y.F.; Han, B.; Liu, T.; Tang, J.; Li, J.; Wu, J.; Li, J.Y.; et al. Discovery of Benzofuran-3(2H)-One Derivatives as Novel DRAK2 Inhibitors That Protect Islet Beta-Cells from Apoptosis. *Eur. J. Med. Chem.* **2017**, *130*, 195–208. [[CrossRef](#)]
76. Lijnen, H.R.; Frederix, L.; Scroyen, I. Deficiency of Plasminogen Activator Inhibitor-2 Impairs Nutritionally Induced Murine Adipose Tissue Development. *J. Thromb. Haemost.* **2007**, *5*, 2259–2265. [[CrossRef](#)]
77. Aissa, A.F.; Amaral, C.L.D.; Venancio, V.P.; Machado, C.D.S.; Hernandez, L.C.; Santos, P.; Curi, R.; Bianchi, M.L.P.; Antunes, L.M.G. Methionine-Supplemented Diet Affects the Expression of Cardiovascular Disease-Related Genes and Increases Inflammatory Cytokines in Mice Heart and Liver. *J. Toxicol Environ. Health A* **2017**, *80*, 1116–1128. [[CrossRef](#)]
78. Smyth, D.J.; Plagnol, V.; Walker, N.M.; Cooper, J.D.; Downes, K.; Yang, J.H.; Howson, J.M.; Stevens, H.; McManus, R.; Wijmenga, C.; et al. Shared and Distinct Genetic Variants in Type 1 Diabetes and Celiac Disease. *N. Engl. J. Med.* **2008**, *359*, 2767–2777. [[CrossRef](#)]
79. Hunt, K.A.; Zhernakova, A.; Turner, G.; Heap, G.A.; Franke, L.; Bruinenberg, M.; Romanos, J.; Dinesen, L.C.; Ryan, A.W.; Panesar, D.; et al. Newly Identified Genetic Risk Variants for Celiac Disease Related to the Immune Response. *Nat. Genet.* **2008**, *40*, 395–402. [[CrossRef](#)]
80. Johnson, B.A.; Wang, J.; Taylor, E.M.; Caillier, S.J.; Herbert, J.; Khan, O.A.; Cross, A.H.; De Jager, P.L.; Gourraud, P.A.; Cree, B.C.; et al. Multiple Sclerosis Susceptibility Alleles in African Americans. *Genes Immun.* **2010**, *11*, 343–350. [[CrossRef](#)]
81. Choi, M.S.; Kim, Y.J.; Kwon, E.Y.; Ryoo, J.Y.; Kim, S.R.; Jung, U.J. High-Fat Diet Decreases Energy Expenditure and Expression of Genes Controlling Lipid Metabolism, Mitochondrial Function and Skeletal System Development in the Adipose Tissue, along with Increased Expression of Extracellular Matrix Remodelling and Inflammation-Related Genes. *Br. J. Nutr.* **2015**, *113*, 867–877. [[PubMed](#)]
82. Moreno-Viedma, V.; Amor, M.; Sarabi, A.; Bilban, M.; Staffler, G.; Zeyda, M.; Stulnig, T.M. Common Dysregulated Pathways in Obese Adipose Tissue and Atherosclerosis. *Cardiovasc. Diabetol.* **2016**, *15*, 120. [[CrossRef](#)] [[PubMed](#)]
83. Leandro, G.S.; Evangelista, A.F.; Lobo, R.R.; Xavier, D.J.; Moriguti, J.C.; Sakamoto-Hojo, E.T. Changes in Expression Profiles Revealed by Transcriptomic Analysis in Peripheral Blood Mononuclear Cells of Alzheimer’s Disease Patients. *J. Alzheimers Dis.* **2018**, *66*, 1483–1495. [[CrossRef](#)] [[PubMed](#)]
84. Grewal, R.; Reutzel, M.; Dilberger, B.; Hein, H.; Zotzel, J.; Marx, S.; Tretzel, J.; Sarafeddin, A.; Fuchs, C.; Eckert, G.P. Purified Oleocanthal and Ligstroside Protect against Mitochondrial Dysfunction in Models of Early Alzheimer’s Disease and Brain Ageing. *Exp. Neurol.* **2020**, *328*, 113248. [[CrossRef](#)]
85. Kim, S.E.; Choo, J.; Yoon, J.; Chu, J.R.; Bae, Y.J.; Lee, S.; Park, T.; Sung, M.K. Genome-Wide Analysis Identifies Colonic Genes Differentially Associated with Serum Leptin and Insulin Concentrations in C57BL/6J Mice Fed a High-Fat Diet. *PLoS ONE* **2017**, *12*, e0171664. [[CrossRef](#)]
86. Anghel, S.I.; Wahli, W. Fat Poetry: A Kingdom for PPAR Gamma. *Cell Res.* **2007**, *17*, 486–511. [[CrossRef](#)]
87. Chmurzynska, A.; Muzsik, A.; Krzyzanowska-Jankowska, P.; Madry, E.; Walkowiak, J.; Bajerska, J. PPARG and FTO Polymorphism Can Modulate the Outcomes of a Central European Diet and a Mediterranean Diet in Centrally Obese Postmenopausal Women. *Nutr. Res.* **2019**, *69*, 94–100. [[CrossRef](#)]
88. Rafehi, H.; Smith, A.J.; Balcerczyk, A.; Ziemann, M.; Ooi, J.; Loveridge, S.J.; Baker, E.K.; El-Osta, A.; Karagiannis, T.C. Investigation into the Biological Properties of the Olive Polyphenol, Hydroxytyrosol: Mechanistic Insights by Genome-Wide mRNA-Seq Analysis. *Genes Nutr.* **2012**, *7*, 343–355. [[CrossRef](#)]
89. Rosillo, M.A.; Alcaraz, M.J.; Sanchez-Hidalgo, M.; Fernandez-Bolanos, J.G.; Alarcon-de-la-Lastra, C.; Ferrandiz, M.L. Anti-Inflammatory and Joint Protective Effects of Extra-Virgin Olive-Oil Polyphenol Extract in Experimental Arthritis. *J. Nutr. Biochem.* **2014**, *25*, 1275–1281. [[CrossRef](#)]
90. Phillips, C.M.; Goumidi, L.; Bertrais, S.; Field, M.R.; Peloso, G.M.; Shen, J.; McManus, R.; Hercberg, S.; Lairon, D.; Planells, R.; et al. Dietary Saturated Fat Modulates the Association between STAT3 Polymorphisms and Abdominal Obesity in Adults. *J. Nutr.* **2009**, *139*, 2011–2017. [[CrossRef](#)]

91. Dongiovanni, P.; Meroni, M.; Longo, M.; Fargion, S.; Fracanzani, A.L. miRNA Signature in NAFLD: A Turning Point for a Non-Invasive Diagnosis. *Int. J. Mol. Sci.* **2018**, *19*, 3966. [[CrossRef](#)] [[PubMed](#)]
92. Otsuka, M.; Zheng, M.; Hayashi, M.; Lee, J.D.; Yoshino, O.; Lin, S.; Han, J. Impaired microRNA Processing Causes Corpus Luteum Insufficiency and Infertility in Mice. *J. Clin. Invest.* **2008**, *118*, 1944–1954. [[CrossRef](#)] [[PubMed](#)]
93. Bobbili, M.R.; Mader, R.M.; Grillari, J.; Dellago, H. OncomiR-17–5p: Alarm Signal in Cancer? *Oncotarget* **2017**, *8*, 71206–71222. [[CrossRef](#)] [[PubMed](#)]
94. Tian, L.; Song, Z.; Shao, W.; Du, W.W.; Zhao, L.R.; Zeng, K.; Yang, B.B.; Jin, T. Curcumin Represses Mouse 3T3-L1 Cell Adipogenic Differentiation via Inhibiting miR-17–5p and Stimulating the Wnt Signalling Pathway Effector Tcf712. *Cell Death Dis.* **2017**, *8*, e2559. [[CrossRef](#)]
95. Coucha, M.; Mohamed, I.N.; Elshaer, S.L.; Mbata, O.; Bartasis, M.L.; El-Remessy, A.B. High Fat Diet Dysregulates microRNA-17–5p and Triggers Retinal Inflammation: Role of Endoplasmic-Reticulum-Stress. *World J. Diabetes* **2017**, *8*, 56–65. [[CrossRef](#)]
96. Tan, L.; Liu, L.; Jiang, Z.; Hao, X. Inhibition of microRNA-17-5p Reduces the Inflammation and Lipid Accumulation, and up-Regulates ATP-Binding Cassette TransporterA1 in Atherosclerosis. *J. Pharmacol. Sci.* **2019**, *139*, 280–288. [[CrossRef](#)]
97. Dellago, H.; Bobbili, M.R.; Grillari, J. MicroRNA-17–5p: At the Crossroads of Cancer and Aging—A Mini-Review. *Gerontology* **2017**, *63*, 20–28. [[CrossRef](#)]
98. Inukai, S.; de Lencastre, A.; Turner, M.; Slack, F. Novel microRNAs Differentially Expressed during Aging in the Mouse Brain. *PLoS ONE* **2012**, *7*, e40028. [[CrossRef](#)]
99. Hebert, S.S.; Horre, K.; Nicolai, L.; Bergmans, B.; Papadopoulou, A.S.; Delacourte, A.; De Strooper, B. MicroRNA Regulation of Alzheimer’s Amyloid Precursor Protein Expression. *Neurobiol. Dis.* **2009**, *33*, 422–428. [[CrossRef](#)]
100. Ruskovska, T.; Maksimova, V.; Milenkovic, D. Polyphenols in Human Nutrition: From the In Vitro Antioxidant Capacity to the Beneficial Effects on Cardiometabolic Health and Related Inter-Individual Variability—An Overview and Perspective. *Br. J. Nutr.* **2020**, *123*, 241–254. [[CrossRef](#)]
101. Plummer, J.; Park, M.; Perodin, F.; Horowitz, M.C.; Hens, J.R. Methionine-Restricted Diet Increases miRNAs That Can Target RUNX2 Expression and Alters Bone Structure in Young Mice. *J. Cell Biochem.* **2017**, *118*, 31–42. [[CrossRef](#)] [[PubMed](#)]
102. Oger, F.; Gheeraert, C.; Mogilenko, D.; Benomar, Y.; Molendi-Coste, O.; Bouchaert, E.; Caron, S.; Dombrowicz, D.; Pattou, F.; Duez, H.; et al. Cell-Specific Dysregulation of microRNA Expression in Obese White Adipose Tissue. *J. Clin. Endocrinol. Metab.* **2014**, *99*, 2821–2833. [[CrossRef](#)] [[PubMed](#)]
103. O’Sullivan, J.F.; Neylon, A.; McGorrian, C.; Blake, G.J. miRNA-93-5p and Other miRNAs as Predictors of Coronary Artery Disease and STEMI. *Int. J. Cardiol.* **2016**, *224*, 310–316. [[CrossRef](#)] [[PubMed](#)]
104. Bushe, C.; Haddad, P.; Peveler, R.; Pendlebury, J. The Role of Lifestyle Interventions and Weight Management in Schizophrenia. *J. Psychopharmacol.* **2005**, *19*, 28–35. [[CrossRef](#)]
105. Strassnig, M.; Brar, J.S.; Ganguli, R. Nutritional Assessment of Patients with Schizophrenia: A Preliminary Study. *Schizophr. Bull.* **2003**, *29*, 393–397. [[CrossRef](#)]
106. McCreadie, R.G. Diet, Smoking and Cardiovascular Risk in People with Schizophrenia: Descriptive Study. *Br. J. Psychiatry* **2003**, *183*, 534–539. [[CrossRef](#)]
107. Joseph, J.; Depp, C.; Shih, P.B.; Cadenhead, K.S.; Schmid-Schonbein, G. Modified Mediterranean Diet for Enrichment of Short Chain Fatty Acids: Potential Adjunctive Therapeutic to Target Immune and Metabolic Dysfunction in Schizophrenia? *Front. Neurosci.* **2017**, *11*, 155. [[CrossRef](#)]
108. Bogomolova, S.; Zarnowiecki, D.; Wilson, A.; Fielder, A.; Procter, N.; Itsiopoulos, C.; O’Dea, K.; Strachan, J.; Ballestrin, M.; Champion, A.; et al. Dietary Intervention for People with Mental Illness in South Australia. *Health Promot. Int.* **2018**, *33*, 71–83. [[CrossRef](#)]
109. Peet, M. The Metabolic Syndrome, Omega-3 Fatty Acids and Inflammatory Processes in Relation to Schizophrenia. *Prostaglandins Leukot. Essent. Fatty Acids* **2006**, *75*, 323–327. [[CrossRef](#)]
110. Emsley, R.; Myburgh, C.; Oosthuizen, P.; van Rensburg, S.J. Randomized, Placebo-Controlled Study of Ethyl-Eicosapentaenoic Acid as Supplemental Treatment in Schizophrenia. *Am. J. Psychiatry* **2002**, *159*, 1596–1598. [[CrossRef](#)]

111. Chen, A.T.; Chibnall, J.T.; Nasrallah, H.A. A Meta-Analysis of Placebo-Controlled Trials of Omega-3 Fatty Acid Augmentation in Schizophrenia: Possible Stage-Specific Effects. *Ann. Clin. Psychiatry* **2015**, *27*, 289–296. [[PubMed](#)]
112. Skoldstam, L.; Hagfors, L.; Johansson, G. An Experimental Study of a Mediterranean Diet Intervention for Patients with Rheumatoid Arthritis. *Ann. Rheum. Dis.* **2003**, *62*, 208–214. [[CrossRef](#)] [[PubMed](#)]
113. Silva, S.; Sepodes, B.; Rocha, J.; Direito, R.; Fernandes, A.; Brites, D.; Freitas, M.; Fernandes, E.; Bronze, M.R.; Figueira, M.E. Protective Effects of Hydroxytyrosol-Supplemented Refined Olive Oil in Animal Models of Acute Inflammation and Rheumatoid Arthritis. *J. Nutr. Biochem.* **2015**, *26*, 360–368. [[CrossRef](#)] [[PubMed](#)]
114. Matsumoto, Y.; Sugioka, Y.; Tada, M.; Okano, T.; Mamoto, K.; Inui, K.; Habu, D.; Koike, T. Monounsaturated Fatty Acids Might Be Key Factors in the Mediterranean Diet that Suppress Rheumatoid Arthritis Disease Activity: The TOMORROW Study. *Clin. Nutr.* **2018**, *37*, 675–680. [[CrossRef](#)] [[PubMed](#)]
115. Forsyth, C.; Kouvari, M.; D’Cunha, N.M.; Georgousopoulou, E.N.; Panagiotakos, D.B.; Mellor, D.D.; Kellett, J.; Naumovski, N. The Effects of the Mediterranean Diet on Rheumatoid Arthritis Prevention and Treatment: A Systematic Review of Human Prospective Studies. *Rheumatol. Int.* **2018**, *38*, 737–747. [[CrossRef](#)]
116. Petersson, S.; Philippou, E.; Rodomar, C.; Nikiphorou, E. The Mediterranean Diet, Fish Oil Supplements and Rheumatoid Arthritis Outcomes: Evidence from Clinical Trials. *Autoimmun. Rev.* **2018**, *17*, 1105–1114. [[CrossRef](#)]
117. Porras, M.; Rada, G.; Duran, J. Effects of Mediterranean diet on the Treatment of Rheumatoid Arthritis. *Medwave* **2019**, *19*, e7640. [[CrossRef](#)]
118. Laudisio, D.; Castellucci, B.; Barrea, L.; Pugliese, G.; Savastano, S.; Colao, A.; Muscogiuri, G. Mediterranean Diet and Breast Cancer Risk: A Narrative Review. *Minerva Endocrinol.* **2020**. [[CrossRef](#)]
119. Dianatinasab, M.; Rezaian, M.; HaghghatNezad, E.; Bagheri-Hosseiniabadi, Z.; Amanat, S.; Rezaeian, S.; Masoudi, A.; Ghiasvand, R. Dietary Patterns and Risk of Invasive Ductal and Lobular Breast Carcinomas: A Systematic Review and Meta-Analysis. *Clin. Breast Cancer* **2020**, *20*, e516–e528. [[CrossRef](#)]
120. Russo, G.I.; Solinas, T.; Urzi, D.; Privitera, S.; Campisi, D.; Cocci, A.; Carini, M.; Madonia, M.; Cimino, S.; Morgia, G. Adherence to Mediterranean Diet and Prostate Cancer Risk in Sicily: Population-Based Case-Control Study. *Int. J. Impot. Res.* **2019**, *31*, 269–275. [[CrossRef](#)]
121. Schneider, L.; Su, L.J.; Arab, L.; Bensen, J.T.; Farnan, L.; Fontham, E.T.H.; Song, L.; Hussey, J.; Merchant, A.T.; Mohler, J.L.; et al. Dietary Patterns Based on the Mediterranean Diet and DASH Diet Are Inversely Associated with High Aggressive Prostate Cancer in PCaP. *Ann. Epidemiol.* **2019**, *29*, 16–22.e1. [[CrossRef](#)] [[PubMed](#)]
122. Sealy, N.; Hankinson, S.E.; Houghton, S.C. Olive Oil and Risk of Breast Cancer: A Systematic Review and Dose-Response Meta-Analysis of Observational Studies. *Br. J. Nutr.* **2020**, 1–9. [[CrossRef](#)] [[PubMed](#)]
123. Urquiza-Salvat, N.; Pascual-Geler, M.; Lopez-Guarnido, O.; Rodrigo, L.; Martinez-Burgos, A.; Cozar, J.M.; Ocana-Peinado, F.M.; Alvarez-Cubero, M.J.; Rivas, A. Adherence to Mediterranean Diet and Risk of Prostate Cancer. *Aging Male* **2019**, *22*, 102–108. [[CrossRef](#)] [[PubMed](#)]
124. Porciello, G.; Montagnese, C.; Crispo, A.; Grimaldi, M.; Libra, M.; Vitale, S.; Palumbo, E.; Pica, R.; Calabrese, I.; Cubisino, S.; et al. Mediterranean Diet and Quality of Life in Women Treated for Breast Cancer: A Baseline Analysis of DEDiCa Multicentre Trial. *PLoS ONE* **2020**, *15*, e0239803. [[CrossRef](#)] [[PubMed](#)]
125. Baguley, B.J.; Skinner, T.L.; Jenkins, D.G.; Wright, O.R.L. Mediterranean-Style Dietary Pattern Improves Cancer-Related Fatigue and Quality of Life in Men with Prostate Cancer Treated with Androgen Deprivation Therapy: A Pilot Randomised Control Trial. *Clin. Nutr.* **2020**. [[CrossRef](#)]

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