

## Coffee Intake and Colorectal Cancer Incidence According to T-Cell Response

Tomotaka Ugai, MD, PhD,<sup>1,2,†</sup> Koichiro Haruki , MD, PhD,<sup>1,†</sup> Juha P. Väyrynen, MD, PhD,<sup>1,3,†</sup> Rong Zhong, PhD,<sup>1,†</sup> Jennifer Borowsky, MBChB,<sup>4</sup> Kenji Fujiyoshi , MD, PhD,<sup>1</sup> Mai Chan Lau, PhD,<sup>1</sup> Melissa Zhao, MD,<sup>1</sup> Naohiko Akimoto , MD, PhD,<sup>1</sup> Tzuu-Wang Chang, PhD,<sup>1</sup> Junko Kishikawa, MD, PhD,<sup>1</sup> Kota Arima, MD, PhD,<sup>1</sup> Shan-Shan Shi, PhD,<sup>1</sup> Simeng Gu, MB,<sup>1</sup> Charles S. Fuchs, MD, MPH,<sup>5,6,7</sup> Edward L. Giovannucci, MD, ScD,<sup>2,8,9</sup> Marios Giannakis, MD, PhD,<sup>3,10,11</sup> Xuehong Zhang, MD, ScD,<sup>8,9</sup> Mingyang Song, MD, ScD,<sup>8,12,13</sup> Jeffrey A. Meyerhardt , MD, MPH,<sup>3,†</sup> Molin Wang, PhD,<sup>2,9,14,‡</sup> Jonathan A. Nowak, MD, PhD,<sup>1,‡</sup> Shuji Ogino , MD, PhD, MS<sup>1,2,10,15,\*,‡</sup>

<sup>1</sup>Department of Pathology, Brigham and Women's Hospital, Program in MPE Molecular Pathological Epidemiology, Harvard Medical School, Boston, MA, USA;

<sup>2</sup>Department of Epidemiology, Harvard T.H. Chan School of Public Health, Boston, MA, USA; <sup>3</sup>Department of Medical Oncology, Dana-Farber Cancer Institute and Harvard Medical School, Boston, MA, USA; <sup>4</sup>Conjoint Gastroenterology Department, QIMR Berghofer Medical Research Institute, Queensland, Brisbane, Australia; <sup>5</sup>Yale Cancer Center, New Haven, CT, USA; <sup>6</sup>Department of Medicine, Yale School of Medicine, New Haven, CT, USA; <sup>7</sup>Smilow Cancer Hospital, New Haven, CT, USA;

<sup>8</sup>Department of Nutrition, Harvard T.H. Chan School of Public Health, Boston, MA, USA; <sup>9</sup>Channing Division of Network Medicine, Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, MA, USA; <sup>10</sup>Broad Institute of MIT and Harvard, Cambridge, MA, USA; <sup>11</sup>Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, MA, USA; <sup>12</sup>Clinical and Translational Epidemiology Unit, Massachusetts General Hospital and Harvard Medical School, Boston, MA, USA; <sup>13</sup>Division of Gastroenterology, Massachusetts General Hospital, Boston, MA, USA; <sup>14</sup>Department of Biostatistics, Harvard T.H. Chan School of Public Health, Boston, MA, USA and <sup>15</sup>Cancer Immunology and Cancer Epidemiology Programs, Dana-Farber Harvard Cancer Center, Boston, MA, USA

<sup>†</sup>Co-first authors.

<sup>‡</sup>Co-senior authors.

\*Correspondence to: Shuji Ogino, MD, PhD, MS, Department of Pathology, Brigham and Women's Hospital, Program in MPE Molecular Pathological Epidemiology, Harvard Medical School, 221 Longwood Ave, EBRC Room 404A, Boston, MA 02115, USA (e-mail: sogino@bwh.harvard.edu).

### Abstract

We hypothesized that the associations between coffee intake and colorectal cancer (CRC) incidence might differ by immune cell densities in CRC tissue. Using the Nurses' Health Study and the Health Professionals Follow-up Study, we examined the association of coffee intake with incidence of CRC classified by intraepithelial or stromal T-cell subset densities by multiplex immunofluorescence assay for CD3, CD4, CD8, CD45RO (PTPRC), and FOXP3. We applied an inverse probability-weighted Cox proportional hazards regression model to control for selection bias and potential confounders. During follow-up of 133 924 participants (3 585 019 person-years), we documented 3161 incident CRC cases, including 908 CRC cases with available data on T-cell densities in tumor tissue. The association between coffee intake and CRC was not statistically significantly different by intraepithelial or stroma T-cell subset ( $P_{\text{heterogeneity}} > .38$ ). Hence, there is no sufficient evidence for differential effect of coffee intake on incidence of CRC subtypes classified by T-cell infiltrates.

Observational studies have found conflicting evidence on the association between coffee consumption and the incidence of colorectal cancer (CRC) (1). Coffee contains complex mixtures of biochemically active compounds, some of which, including polyphenols and caffeine, are suggested to influence adaptive immune responses (2-5). However, the human data linking coffee to immune cells in CRC are lacking. We hypothesized that the associations between coffee intake and CRC incidence might differ by T-cell response to CRC.

We used data from 2 large prospective cohort studies in the United States: the Nurses' Health Study (NHS,  $n = 121\,701$  women aged 30-55 years at enrollment followed since 1976) and the Health Professionals Follow-up Study (HPFS,  $n = 51\,529$  men aged 40-75 years at enrollment followed since 1986) (6,7). Study participants have been sent follow-up questionnaires biennially to update information on demographics, diet, lifestyle factors, and medical history. The follow-up rate has been over 90% for each questionnaire cycle in both cohorts. To assess dietary

Received: 28 April 2020; Revised: 27 July 2020; Accepted: 11 August 2020

© The Author(s) 2020. Published by Oxford University Press.

This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs licence (<http://creativecommons.org/licenses/by-nc-nd/4.0/>), which permits non-commercial reproduction and distribution of the work, in any medium, provided the original work is not altered or transformed in any way, and that the work is properly cited. For commercial re-use, please contact [journals.permissions@oup.com](mailto:journals.permissions@oup.com)

**Table 1.** Age-standardized characteristics according to coffee intake in the NHS (women, 1980-2012) and the HPFS (men, 1986-2012)<sup>a</sup>

Characteristic	Coffee intake							
	Women				Men			
	0 cups/d (n = 19 497)	<1 cups/d (n = 7388)	1-3 cups/d (n = 38 147)	≥3 cups/d (n = 21 666)	0 cups/d (n = 7594)	< 1 cups/d (n = 9416)	1-3 cups/d (n = 21 041)	≥3 cups/d (n = 9175)
Coffee intake, cup/d (median, IQR)	0	0.5 (0.2, 0.8)	2.2 (1.5, 2.5)	4.3 (3.5, 5.0)	0	0.5 (0.2, 0.8)	2.0 (1.4, 2.5)	4.3 (3.5, 4.9)
Age, y	56.2 (11.4)	62.2 (11.4)	61.9 (11.3)	59.7 (10.6)	61.2 (11.1)	65.3 (11.4)	65.1 (11.2)	62.7 (10.4)
Body mass index, kg/m <sup>2</sup>	25.9 (5.2)	25.6 (5.0)	25.3 (4.5)	25.0 (4.3)	25.5 (3.5)	25.5 (3.5)	25.9 (3.4)	26.2 (3.4)
Physical activity, METS-h/wk <sup>b</sup>	15.7 (18.0)	16.5 (16.8)	16.9 (17.5)	16.2 (17.3)	26.6 (24.4)	26.6 (23.8)	25.6 (22.8)	24.8 (21.6)
Pack-years of smoking	8.4 (16.6)	7.7 (14.9)	11.8 (17.6)	20.7 (22.7)	5.2 (12.7)	8.4 (14.9)	13.5 (18.3)	20.3 (22.2)
Family history of CRC, %	11.9	13.3	13.6	13.2	11.5	12.5	12.5	12.5
History of diabetes, %	9.3	9.5	7.2	5.5	7.0	7.7	7.8	7.5
History of previous endoscopy, %	35.6	43.6	39.9	36.6	50.7	55.6	53.9	52.7
Current multivitamin use, %	50.1	55.1	52.4	49.4	41.8	44.8	43.6	44.6
Regular aspirin use, %	29.4	30.9	32.6	33.9	40.8	45.4	48.7	50.3
Regular NSAID use, %	13.6	18.0	18.0	18.8	12.1	13.9	16.4	17.7
Postmenopausal, %	73.9	78.0	78.3	78.7				
Current hormone use, % <sup>c</sup>	22.8	28.4	27.2	26.3				
Dietary intake								
Total calorie, kcal/d	1650 (467)	1668 (437)	1672 (433)	1709 (442)	1954 (551)	1904 (543)	1974 (548)	2082 (573)
Alcohol, g/d (median, IQR)	0.3 (0.0,2.4)	1.1 (0.0,4.6)	2.7 (0.5,9.2)	2.8 (0.6,9.6)	0.7 (0.0,5.5)	4.6 (0.9,11.9)	8.2 (2.2,17.3)	9.2 (2.4,20.1)
Total red meat, servings/wk	6.7 (4.1)	6.1 (3.5)	6.5 (3.5)	7.0 (3.7)	5.8 (4.4)	5.6 (4.0)	6.4 (4.3)	7.5 (4.8)
Calcium, mg/d	879 (387)	958 (371)	925 (345)	932 (337)	999 (412)	962 (395)	908.9 (357)	915.2 (345)
Folate, μg/d	423 (253)	452 (229)	426 (203)	400 (194)	552 (267)	576 (270)	537 (246)	509 (233)
AHEI score	45.1 (10.5)	47.1 (9.9)	46.4 (9.5)	45.9 (9.4)	48.3 (10.9)	49.6 (10.0)	48.1 (9.8)	46.7 (9.8)

<sup>a</sup>Cumulative average values are presented. Mean ( $\pm$ SD) or median (IQR) for continuous variable or percentages for categorical variables are presented. All variables are age-standardized except age. AHEI = Alternative Healthy Eating Index; CRC = colorectal cancer; HPFS = Health Professionals Follow-up Study; IQR = interquartile range; METS = metabolic equivalent task score; NHS = Nurses' Health Study; NSAID = nonsteroidal antiinflammatory drug.

<sup>b</sup>Physical activity is represented by the product sum of the metabolic equivalent score (METS) of each specific recreational activity and hours spent on that activity per week.

<sup>c</sup>Proportion of current menopausal hormone use is calculated among postmenopausal women only.

intake in each cohort, food frequency questionnaires (FFQs) were initially collected in 1980 for the NHS and in 1986 for the HPFS. For the NHS, a 61-item semi-quantitative FFQ was used at baseline (8), which was expanded to approximately 130 foods and beverage items in 1984, 1986, and every 4 years thereafter. For the HPFS cohorts, baseline dietary intake was assessed using a 131-item FFQ that was also used for updates generally every 4 years subsequently (9). For each item, FFQs prompted participants to report their average intake over the preceding year for a specified serving size of each food and beverage from 9 possible responses, ranging from never or almost never to 6 or more times per day. Intakes of various nutrients were calculated by multiplying the frequency of each food or beverage consumed by the nutrient content of the specified portion size and then summing the contributions from all foods and beverages in the FFQ. To capture long-term exposure, we calculated the cumulative average of each factor from baseline up to the most updated cycle. We excluded participants who had a history of cancer (except for nonmelanoma skin cancer), inflammatory bowel disease, implausibly high or low caloric intake, and incomplete data on coffee intake. The follow-up durations for this analysis were 1980 to 2012 for the NHS and 1986 to 2012 for the HPFS. In addition to self-reported incident CRC cases, lethal unreported CRC cases were ascertained through use of the National Death Index, and all CRC cases were confirmed by medical record review.

We simultaneously measured the expression of CD3, CD4, CD8, CD45RO (PTPRC), and FOXP3 in T cells within tumor epithelial and stromal regions in CRC tissue microarrays by a multiplex immunofluorescence assay. For 908 CRC patients, there

were 1694 available tissue microarray cores (mean 1.9 cores per patient), while 192 of 1886 (10%) tumor cores from these patients were missing or unrepresentative, based on pathologist review. Informed consent was obtained from all study participants at enrollment. This study was approved by the institutional review boards of Brigham and Women's Hospital and Harvard T.H. Chan School of Public Health (Boston, MA) and those of the participating registries as required.

All statistical analyses were conducted using SAS software (version 9.4, SAS Institute, Cary, NC). All *P* values were 2-sided, and a *P* less than .005 was considered statistically significant given multiple hypothesis testing. Our primary hypothesis testing was an assessment of heterogeneity in the associations of coffee intake with incidence of CRC classified by each T-cell subset. All other assessments were secondary analyses. Leveraging covariate data of 3161 CRC cases, the inverse probability weighting method was integrated into the Cox proportional hazards model (10) to control for selection bias due to tissue availability and potential confounders. The Cox model was used with stratification by age, sex (ie, cohort), and calendar year of the current questionnaire cycle. We also included the covariates described in the footnote to Table 2. The assumption of proportionality of hazards was verified by including an interaction term between coffee intake and follow-up time. To test whether strength of the coffee-CRC association might differ across the ordinal subtypes, we used the meta-regression method with a subtype-specific random effect term (11).

Table 1 shows age-standardized characteristics of participants according to coffee intake in the NHS and the HPFS. During follow-up of 133 924 participants (3 585 019

**Table 2.** Incidence of CRC, overall and by intraepithelial and stroma T-cell subset density, according to coffee intake in the combined cohorts of the NHS (1980-2012) and HPFS (1986-2012)

Characteristics	Coffee intake, cups/d				P <sub>trend</sub> <sup>a</sup>	P <sub>heterogeneity</sub> <sup>b</sup>
	0	<1	1-3	≥3		
<b>Overall</b>						
Person-years	402 273	604 468	1 712 111	866 168		
Cases (n = 3161), No.	269	579	1621	692		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.08 (0.94 to 1.26)	1.08 (0.94 to 1.23)	0.99 (0.89 to 1.15)	.22	
<b>Intraepithelial</b>						
<b>CD3<sup>+</sup>CD4<sup>+</sup>FOXP3<sup>+</sup> cells</b>						
Low density (n = 625), No.	48	110	328	139		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.23 (0.87 to 1.73)	1.26 (0.93 to 1.72)	1.11 (0.79 to 1.55)	.68	.39
Intermediate density (n = 142), No.	11	18	79	34		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.79 (0.38 to 1.63)	1.29 (0.70 to 2.36)	1.05 (0.54 to 2.04)	.50	
High density (n = 141), No.	17	26	71	27		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.68 (0.37 to 1.26)	0.62 (0.36 to 1.07)	0.53 (0.29 to 0.98)	.099	
<b>CD3<sup>+</sup>CD4<sup>+</sup>CD45RO<sup>+</sup> cells</b>						
Low density (n = 302), No.	28	48	161	65		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.04 (0.65 to 1.67)	1.11 (0.74 to 1.67)	0.92 (0.58 to 1.44)	.94	.71
Intermediate density (n = 305), No.	27	51	159	68		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.92 (0.58 to 1.45)	1.06 (0.71 to 1.60)	0.91 (0.58 to 1.43)	.76	
High density (n = 301), No.	21	55	158	67		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.16 (0.70 to 1.92)	1.17 (0.74 to 1.86)	1.07 (0.65 to 1.76)	.81	
<b>CD3<sup>+</sup>CD4<sup>+</sup>CD45RO<sup>-</sup> cells</b>						
Low density (n = 514), No.	41	94	264	115		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.22 (0.85 to 1.77)	1.17 (0.84 to 1.63)	1.05 (0.73 to 1.52)	.47	.72
Intermediate density (n = 197), No.	16	21	115	45		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.69 (0.36 to 1.30)	1.38 (0.83 to 2.31)	1.06 (0.60 to 1.87)	.31	
High density (n = 197), No.	19	39	99	40		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.92 (0.53 to 1.60)	0.80 (0.49 to 1.32)	0.71 (0.41 to 1.23)	.17	
<b>CD3<sup>+</sup>CD8<sup>+</sup>CD45RO<sup>+</sup> cells</b>						
Low density (n = 423), No.	37	87	206	93		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.26 (0.86 to 1.86)	1.06 (0.74 to 1.51)	0.96 (0.65 to 1.42)	.15	.39
Intermediate density (n = 242), No.	20	25	144	53		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.62 (0.35 to 1.12)	1.26 (0.80 to 2.00)	0.96 (0.57 to 1.59)	.39	
High density (n = 243), No.	19	42	128	54		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.03 (0.59 to 1.77)	1.06 (0.65 to 1.73)	0.97 (0.57 to 1.66)	.77	
<b>CD3<sup>+</sup>CD8<sup>+</sup>CD45RO<sup>-</sup> cells</b>						
Low density (n = 571), No.	43	107	295	126		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.29 (0.90 to 1.84)	1.29 (0.93 to 1.79)	1.11 (0.78 to 1.58)	.55	.51
Intermediate density (n = 168), No.	20	28	88	32		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.71 (0.40 to 1.26)	0.72 (0.44 to 1.16)	0.59 (0.34 to 1.03)	.16	
High density (n = 169), No.	13	19	95	42		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.72 (0.35 to 1.48)	1.15 (0.63 to 2.09)	1.04 (0.55 to 1.99)	.42	
<b>Stroma</b>						
<b>CD3<sup>+</sup>CD4<sup>+</sup>FOXP3<sup>+</sup> cells</b>						
Low density (n = 485), No.	43	81	253	108		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.00 (0.69 to 1.45)	1.07 (0.77 to 1.49)	0.91 (0.63 to 1.30)	.38	.80
Intermediate density (n = 211), No.	14	34	111	52		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.18 (0.63 to 2.19)	1.40 (0.80 to 2.45)	1.39 (0.77 to 2.53)	.29	
High density (n = 211), No.	19	39	113	40		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.99 (0.57 to 1.72)	0.98 (0.60 to 1.60)	0.77 (0.44 to 1.33)	.19	
<b>CD3<sup>+</sup>CD4<sup>+</sup>CD45RO<sup>+</sup> cells</b>						
Low density (n = 302), No.	26	52	148	76		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.10 (0.68 to 1.77)	1.03 (0.68 to 1.57)	1.08 (0.69 to 1.71)	.93	.51
Intermediate density (n = 304), No.	28	47	167	62		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.86 (0.54 to 1.37)	1.10 (0.73 to 1.64)	0.85 (0.54 to 1.34)	.62	
High density (n=301), No.	22	55	162	62		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.17 (0.71 to 1.92)	1.22 (0.77 to 1.91)	0.97 (0.59 to 1.59)	.38	

(continued)

Table 2. (continued)

Characteristics	Coffee intake, cups/d				P <sub>trend</sub> <sup>a</sup>	P <sub>heterogeneity</sub> <sup>b</sup>
	0	<1	1-3	≥3		
CD3 <sup>+</sup> CD4 <sup>+</sup> CD45RO <sup>-</sup> cells						
Low density (n = 365), No.	31	64	190	80		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.04 (0.68 to 1.59)	1.06 (0.73 to 1.55)	0.92 (0.60 to 1.39)	.41	.74
Intermediate density (n = 270), No.	22	37	146	65		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.86 (0.50 to 1.46)	1.21 (0.77 to 1.91)	1.11 (0.68 to 1.83)	.33	
High density (n = 272), No.	23	53	141	55		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.19 (0.73 to 1.95)	1.09 (0.69 to 1.70)	0.88 (0.54 to 1.45)	.18	
CD3 <sup>+</sup> CD8 <sup>+</sup> CD45RO <sup>+</sup> cells						
Low density (n = 383), No.	32	74	185	92		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.30 (0.85 to 1.97)	1.11 (0.76 to 1.63)	1.06 (0.70 to 1.60)	.42	.75
Intermediate density (n = 262), No.	18	40	147	57		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.04 (0.59 to 1.82)	1.45 (0.88 to 2.37)	1.28 (0.75 to 2.18)	.28	
High density (n = 262), No.	26	40	145	51		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.75 (0.46 to 1.21)	0.90 (0.59 to 1.37)	0.67 (0.42 to 1.08)	.19	
CD3 <sup>+</sup> CD8 <sup>+</sup> CD45RO <sup>-</sup> cells						
Low density (n = 503), No.	37	94	254	118		
HR (95% CI) <sup>c</sup>	1 (Referent)	1.35 (0.92 to 1.99)	1.29 (0.91 to 1.83)	1.20 (0.82 to 1.76)	.83	.94
Intermediate density (n = 202), No.	19	32	121	30		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.80 (0.46 to 1.39)	1.05 (0.65 to 1.70)	0.58 (0.33 to 1.03)	.070	
High density (n = 202), No.	20	28	102	52		
HR (95% CI) <sup>c</sup>	1 (Referent)	0.70 (0.40 to 1.24)	0.86 (0.53 to 1.41)	0.89 (0.53 to 1.51)	.69	

<sup>a</sup>Trend test was performed using the median intake of each category. CI = confidence interval; CRC = colorectal cancer; HPFS = Health Professionals Follow-up Study; HR = hazard ratio; METS = metabolic equivalent task score; NHS = Nurses' Health Study.

<sup>b</sup>The meta-regression method with a subtype-specific random effect term was used to evaluate the heterogeneity.

<sup>c</sup>Cox proportional hazards model was used with stratification by age, sex (ie, cohort), and calendar year of current questionnaire cycle. We additionally adjusted for family history of CRC, history of diabetes, history of endoscopy, pack-years of smoking (0, 0 to <5, 5 to <20, 20 to <40, and ≥40), body mass index (quartiles), physical activity (METS, quartiles), multivitamin use, regular use of aspirin, regular use of nonsteroidal antiinflammatory drugs, alcohol consumption (0 to <5, 5 to <15, 15 to <30, ≥30 g/d), total calorie intake (quartiles), total red meat intake (quartiles), folate intake (quartiles), calcium intake (quartiles), and Alternative Healthy Eating Index (quartiles). For women, we further adjusted for menopause status and menopausal hormone therapy (premenopause vs postmenopause with never, past, or current use of menopausal hormone therapy).

person-years), we documented 3161 CRC cases, including 908 CRC cases with available data on T-cell densities in tumor tissue.

Table 2 shows hazard ratios for incidence of CRC overall and by intraepithelial and stromal T-cell subset in relation to coffee intake. We did not observe a statistically significant association between coffee intake and overall CRC incidence. Furthermore, the association of coffee intake with CRC incidence did not statistically significantly differ by intraepithelial or stroma T-cell subset ( $P_{\text{interaction}} > .38$ ). Supplementary Table 1 (available online) shows CRC by intraepithelial and stromal T-cell subset, according to coffee intake.

Evidence indicates that certain modifiable factors may influence the immune system and reduce CRC incidence, thereby suggesting their antitumor effects through immunomodulation (12,13). In fact, epidemiological studies showed that the inverse association of marine omega-3 polyunsaturated fatty acids intake with CRC incidence was stronger for tumors containing a higher density of FOXP3<sup>+</sup> cells (14) and that the association of cigarette smoking with CRC risk was stronger for tumors containing a higher density of CD3<sup>+</sup> cells (7). However, the current study does not support the hypothesis that coffee may modify CRC incidence through its effect on T cells. One of the limitations of this study is modest statistical power. With the sample size of 908 cases and 80% power, the maximum detectable ratio of hazard ratio (ie, a test for heterogeneity) is 0.66. Therefore, it is possible that we were not able to detect smaller effects than this number. Considering the importance of cancer immunology and the paucity of immuno-epidemiological studies

(12,13,15), additional efforts are needed to study the relationships of modifiable factors with incidence CRC classified by various types of immune cells.

## Funding

This work was supported by U.S. National Institutes of Health (NIH) grants (P01 CA87969 to M.J. Stampfer; UM1 CA186107 to M.J. Stampfer; P01 CA55075 to W.C. Willett; UM1 CA167552 to W.C. Willett; U01 CA167552 to W.C. Willett and L.A. Mucci; P50 CA127003 to CSF; R01 CA118553 to CSF; R01 CA169141 to CSF; R01 CA137178 to ATC; K24 DK098311 to ATC; R35 CA197735 to SO; R01 CA151993 to SO; R01 CA248857 to SO, U. Peters, and A.I. Phipps; R00 CA215314 to MS; and K07 CA188126 to XZ); by Nodal Award (2016–02) from the Dana-Farber Harvard Cancer Center (to SO); by the Stand Up to Cancer Colorectal Cancer Dream Team Translational Research Grant (SU2C-AACR-DT22-17 to CSF and MG), administered by the American Association for Cancer Research, a scientific partner of SU2C; by the American Cancer Society Mentored Research Scholar Grant (MRSG-17-220-01-NEC to MS); and by grants from the Project P Fund, The Friends of the Dana-Farber Cancer Institute, Bennett Family Fund, and the Entertainment Industry Foundation through National Colorectal Cancer Research Alliance. TU, KH, and KF were supported by fellowship grants from the Uehara Memorial Foundation. KH was supported by the Mitsukoshi Health and Welfare Foundation.

TU was supported by Yasuda Medical Foundation. JB was supported by a grant from the Australia Awards-Endeavour Scholarships and Fellowships Program. KA and TU were supported by a grant from Overseas Research Fellowship (201860083 to KA; 201960541 to TU) MG was supported by a Conquer Cancer Foundation of ASCO Career Development Award. JAM research is supported by the Douglas Gray Woodruff Chair fund, the Guo Shu Shi Fund, Anonymous Family Fund for Innovations in Colorectal Cancer, Project P fund, and the George Stone Family Foundation.

## Notes

**Role of the funder:** The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Disclosures:** C.S.F. previously served as a consultant for Agios, Bain Capital, Bayer, Celgene, Dicerna, Five Prime Therapeutics, Gilead Sciences, Eli Lilly, Entrinsic Health, Genentech, KEW, Merck, Merrimack Pharmaceuticals, Pfizer Inc, Sanofi, Taiho, and Unum Therapeutics; C.S.F. also serves as a Director for CytomX Therapeutics and owns unexercised stock options for CytomX and Entrinsic Health. M.G. receives research funding from Bristol-Myers Squibb and Merck. J.A.M. received institutional research funding from Boston Biomedical. J.A.M. has also served as an advisor/consultant to Ignyta, Array Pharmaceutical, and Cota. This study was not funded by any of these commercial entities. No other conflicts of interest exist. The other authors declare that they have no conflicts of interest.

**Author contributions:** T.U., K.H., J.A.N., and S.O.: developed the main concept and designed the study. C.S.F., M.G., J.A.N., and S.O.: wrote grant applications. T.U., K.H., J.P.V., R.Z., J.B., K.F., M.C.L., M.Z., N.A., T.C., J.K., K.A., S.S., S.G., C.S.F., J.A.N., and S.O.: were responsible for collection of tumor tissue, and acquisition of epidemiologic, clinical and tumor tissue data, including histopathological, immunohistochemical, and immunofluorescent characteristics. T.U., K.H., and S.O.: performed data analysis and interpretation. T.U., K.H., and S.O.: drafted the manuscript. All authors contributed to editing and critical revision for important intellectual contents.

**Disclaimer:** The content is solely the responsibility of the authors and does not necessarily represent the official views of NIH.

**Use of Standardized Official Symbols:** We use HUGO (Human Genome Organisation)-approved official symbols (or root symbols) for genes and gene products, including CD3, CD4, CD8,

FOXP3, PTPRC; all of which are described at [www.genenames.org](http://www.genenames.org). Gene symbols are italicized, whereas symbols for gene products are not italicized.

## Data Availability

The data underlying this article cannot be shared publicly. Further information including the procedures to obtain and access data from the Nurses' Health Studies and Health Professionals Follow-up Study is described at <https://www.nurseshealthstudy.org/researchers> (contact email: [nhsaccess@channing.harvard.edu](mailto:nhsaccess@channing.harvard.edu)) and <https://sites.sph.harvard.edu/hpfs/for-collaborators/>.

## References

- Vieira AR, Abar L, Chan DSM, et al. Foods and beverages and colorectal cancer risk: a systematic review and meta-analysis of cohort studies, an update of the evidence of the WCRF-AICR Continuous Update Project. *Ann Oncol.* 2017; 28(8):1788–1802.
- Bhattacharyya S, Md Sakib Hossain D, Mohanty S, et al. Curcumin reverses T cell-mediated adaptive immune dysfunctions in tumor-bearing hosts. *Cell Mol Immunol.* 2010;7(4):306–315.
- Horrigan LA, Kelly JP, Connor TJ. Immunomodulatory effects of caffeine: friend or foe? *Pharmacol Ther.* 2006;111(3):877–892.
- Sharif K, Watad A, Bragazzi NL, et al. Coffee and autoimmunity: more than a mere hot beverage! *Autoimmun Rev.* 2017;16(7):712–721.
- Zou T, Yang Y, Xia F, et al. Resveratrol Inhibits CD4+ T cell activation by enhancing the expression and activity of Sirt1. *PLoS One.* 2013;8(9):e75139.
- Michels KB, Willett WC, Fuchs CS, et al. Coffee, tea, and caffeine consumption and incidence of colon and rectal cancer. *J Natl Cancer Inst.* 2005;97(4):282–292.
- Hamada T, Nowak JA, Masugi Y, et al. Smoking and risk of colorectal cancer sub-classified by tumor-infiltrating T cells. *J Natl Cancer Inst.* 2019;111(1):42–51.
- Willett WC, Sampson L, Stampfer MJ, et al. Reproducibility and validity of a semiquantitative food frequency questionnaire. *Am J Epidemiol.* 1985;122(1):51–65.
- Rimm EB, Giovannucci EL, Stampfer MJ, et al. Reproducibility and validity of an expanded self-administered semiquantitative food frequency questionnaire among male health professionals. *Am J Epidemiol.* 1992;135(10):1114–1126. discussion 1127–1136.
- Liu L, Nevo D, Nishihara R, et al. Utility of inverse probability weighting in molecular pathological epidemiology. *Eur J Epidemiol.* 2018;33(4):381–392.
- Wang M, Spiegelman D, Kuchiba A, et al. Statistical methods for studying disease subtype heterogeneity. *Stat Med.* 2016;35(5):782–800.
- Ogino S, Nowak JA, Hamada T, et al. Insights into pathogenic interactions among environment, host, and tumor at the crossroads of molecular pathology and epidemiology. *Annu Rev Pathol Mech Dis.* 2019;14(1):83–103.
- Ogino S, Nowak JA, Hamada T, et al. Integrative analysis of exogenous, endogenous, tumour and immune factors for precision medicine. *Gut.* 2018; 67(6):1168–1180.
- Song M, Nishihara R, Cao Y, et al. Marine omega-3 polyunsaturated fatty acid intake and risk of colorectal cancer characterized by tumor-infiltrating T cells. *JAMA Oncol.* 2016;2(9):1197–1206.
- Berntsson J, Eberhard J, Nodin B, et al. Pre-diagnostic anthropometry, sex, and risk of colorectal cancer according to tumor immune cell composition. *Oncimmunology.* 2019;8(12):e1664275.