

Cotrimoxazole Prophylaxis Selects for Antimicrobial Resistance in Human Immunodeficiency Virus–Exposed, Uninfected Infants

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(See the Major Article by D'Souza et al on pages 2858-68.)

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Rates of mother-to-child transmission of human immunodeficiency virus (HIV) are falling globally, with an estimated 200 000 child infections averted in 2018 due to the implementation of prevention of mother-to-child-transmission (PMTCT) interventions [1]. As a result, fewer children are living with HIV and a growing number of children born to mothers living with HIV remain uninfected. However, children who are HIV-exposed but uninfected (HEU) continue to have higher all-cause mortality than HIV-unexposed infants [2], primarily due to greater risk and severity of common childhood infections [3]. As an expanding, clinically distinct group of children, a greater understanding of why HEU children fail to survive and thrive to the extent of HIV-unexposed children is warranted [3-5]. Optimal interventions, particularly those to reduce infectious morbidity and mortality, may well deviate from those developed for children

living with HIV or for those who are HIV unexposed [5].

Antimicrobial prophylaxis guidelines for HIV-exposed infants were developed prior to the wide availability of antiretroviral therapy (ART), when mother-tochild transmission rates, and subsequent infant infectious mortality, were high [6, 7]. In this context, treating HEU infants in the same way as HIV-positive infants was justified because ongoing exposure to HIV via breastfeeding and the challenges of infant HIV testing posed a risk that initially uninfected infants who subsequently acquired HIV would miss out on life-saving treatment [7]. The World Health Organization (WHO) recommends that all HIV-exposed infants receive prophylactic cotrimoxazole (CTX) from 4-6 weeks of age [7]; CTX is continued long-term for HIV-positive children living in regions with high prevalence of severe bacterial infections and malaria, and only discontinued for HEU children upon cessation of breastfeeding and conclusive determination of their HIV-negative status [7]. CTX is a broad-spectrum antibiotic made up of 2 folate synthesis inhibitors (trimethoprim and sulfamethoxazole) with activity against Pneumocystis jirovecii and a range of other bacterial, fungal, and Plasmodium species [9]. There is strong evidence that CTX significantly reduces

infectious morbidity and mortality [9], and our group and others [10, 11] have recently shown that long-term continuation of CTX reduces systemic and intestinal levels of inflammatory mediators associated with poor clinical outcomes for children and adults living with HIV. The impact of CTX on the health of HEU infants is far less well supported. Although morbidity and mortality reductions have been observed among HEU infants living in malaria-endemic regions [12], 2 recent randomized controlled trials in nonmalarial regions show that CTX did not improve 18-month survival [13], 12-month survival [8], or incidence of severe pneumonia or diarrhea [8] among HEU infants.

Without strong evidence for a clinical benefit, recommendations to provide CTX prophylaxis to all HEU infants have been called into question [14, 15], particularly in light of substantial improvements in PMTCT coverage and early infant diagnosis of HIV since treatment guidelines were developed. There are rising concerns that CTX may select for antimicrobial resistance (AMR) [13, 16–18], which is already very high among CTX-targeted pathogens in many lowand middle-income countries (LMICs) [9]. Furthermore, there is a growing understanding of how resident microbial populations (the microbiome) contribute

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to child development and long-term health [19], which may be disrupted by early exposure to antibiotics. Exposure to maternal HIV is associated with a distinct gut microbiome among HEU infants relative to HIV-unexposed infants [20, 21], but little is known about how this contributes to their clinical phenotype. Contrary to expectations that CTX might reduce diversity of the gut microbiome through its broad antimicrobial activity, recent studies have not identified global difference in the gut microbiome after long-term CTX treatment of HIVpositive, ART-treated children [10] or after short-term treatment of a mixed population of HIV-positive and HIVnegative children [22]. At a species level, our group has shown that CTX specifically suppressed gut-resident streptococci associated with intestinal inflammation among HIV-positive children who continued treatment for 84 weeks, but this was not evident for global taxa [10]. The impact of CTX on HEU microbiomes has not been previously assessed.

In this issue of Clinical Infectious Diseases, D'Souza and colleagues provide the first evaluation of how CTX affects the gut microbiomes and AMR gene carriage of HEU infants during their first year of life. The substudy is nested within a recently completed randomized controlled trial among HEU children in South Africa, which showed that no CTX (n = 609) was not inferior to CTX prophylaxis administered according to WHO guidelines (n = 611)for the prevention of severe childhood illnesses or death in an urban setting without endemic malaria transmission, low incidence of pneumonia and diarrhea, and good coverage of PMTCT and infant HIV testing [8]. Using stool samples from a small subset of trial participants, this substudy identifies a convincing signature of AMR selection over time among infants who received CTX (n = 34) relative to untreated controls (n = 29), informing on a pertinent concern around the use of prophylactic antibiotics in LMICs. Despite global stability

in the variety of genes associated with microbial taxa and their functions within individuals (a-diversity) in both CTXtreated and untreated groups of HEU infants, the abundance and α-diversity of trimethoprim and sulfonamide resistance genes increased with time only in the CTX-treated group. In parallel, variation between individuals in genes associated with microbial taxa, function, and AMR (β -diversity) decreased among infants receiving CTX, suggesting that prophylaxis exerted an ongoing selection pressure on the microbiome with continued use. These more detailed assessments are consistent with observations from the trial of CTX among HEU children in Botswana, which reported a higher proportion of CTX-resistant Escherichia coli isolates cultured from stool in CTX-treated vs untreated HEU children postrandomization [13].

A strength of the study is its randomized design and longitudinal follow-up (stool was sampled at 6 weeks, 4 months, and 6 months), which allowed for a causal role of CTX to be disaggregated from potential confounding factors known to shape microbiome diversity. Few studies have been able to assess the early effects of CTX on the microbiome of HEU infants or infants living with HIV as CTX is standard of care for both groups [7] and microbial diversity changes with age, particularly in young infants in whom colonization is ongoing [19]. An additional advantage of this study over previous investigations of the gut microbiome of HEU children [20, 21] is the use of shotgun metagenomics rather than targeted 16S sequencing or culture-based AMR testing. This metagenomics approach sequences all microbial genomes present in a sample rather than a subset of preselected species or marker genes, allowing for a less biased and more accurate assessment of microbial taxa, function, and AMR gene carriage [23]. While sequencing read-depth in this study did not allow for confident resolution of potential CTX-driven changes at a species or strain level, the lack of global differences in α -diversity identified at a taxa level goes some way to allay concerns that CTX drives dysbiosis among HEU infants.

This substudy excluded infants who did not provide stool samples at all 3 timepoints, and 15% of infants were lost to follow-up in the wider trial [8], meaning that it likely focuses on children with more favorable clinical outcomes. The study is also insufficiently powered to draw conclusions on the clinical importance of AMR selection by CTX. Thus, selection for AMR genes is insufficient evidence in isolation to change current guidelines on CTX use. Indeed, CTX is already known to select for AMR in HIV-positive cohorts [9] and, due to its broad-spectrum activity, appears to do so more rapidly than more restricted-spectrum antibiotics [24]. However, the beneficial effects of CTX prophylaxis on morbidity, mortality, and inflammation persist with long-term use among children and adults living with HIV despite high resistance rates [6, 10, 25, 26]. Concerns around AMR become more critical in the context of the wider trial in which this substudy was nested, which found similar clinical outcomes among those randomized to take vs not take CTX prophylaxis [8]. Therefore, the risk of driving AMR carriage is not being offset by a clinical benefit for HEU in this context. However, calls to change treatment guidelines for HEU on the basis of the evidence that CTX selects for AMR are dependent on 2 further, as yet untested assumptions: (1) AMR genes will persist long-term after discontinuation of CTX, and (2) AMR gene carriage will compromise clinical management of subsequent infections. Both concerns require validation in future studies to provide proof of harm. Another important consideration is that maternal CTX use may have shaped infant microbiomes prior to randomization via vertical transmission of AMR genes and/or direct exposure of infants to CTX in breast milk. D'Souza and colleagues do not report on maternal CTX treatment, and it remains plausible that the most profound effects of CTX on the infant microbiome and resistome occur in response to initial CTX exposure from their mothers; were this to be the case, changing treatment guidelines for HEU infants would not avert AMR risk.

Current guidelines recommending CTX prophylaxis for HIV-exposed infants are almost exclusively due to the clinical benefits for HIV-positive children, with limited evidence for a benefit for children who remain uninfected in settings where malaria is nonendemic [7]. Minimized vertical transmission and early and ongoing infant HIV testing reflect an ideal scenario for the clinical care of HIV-exposed infants, but cannot be considered the norm in many LMICs [7]. Therefore, a context-sensitive review of the relative risks and benefits of existing treatment guidelines for HEU children is timely. D'Souza and colleagues provide the first direct evidence for AMR selection by CTX treatment of HEU infants using unbiased metagenomics, adding empirical evidence for why global antibiotic prophylaxis should be avoided where more targeted interventions are possible. Population-level AMR is a particularly critical consideration in the 6 countries where >10% of all children are HEU (Eswatini [32.4%], Botswana [27.4%], South Africa [21.6%], Lesotho [21.1%], Namibia [16.4%], and Zimbabwe [13.6%]) [27]. Earlier and ongoing testing for HIV during infancy is already recommended [7]; wider implementation of such guidelines would enable CTX prophylaxis to be provided in a more targeted way, maintaining the essential protection against infections necessary for children living with HIV while avoiding unnecessarily selecting for AMR among HEU infants.

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References

- Joint United Nations Programme on HIV/AIDS. AIDSInfo: elimination of mother-to-child transmission. Available at: http://aidsinfo.unaids.org/. Accessed 20 November 2019.
- Brennan AT, Bonawitz R, Gill CJ, et al. A meta-analysis assessing all-cause mortality in HIV-exposed uninfected compared with HIVunexposed uninfected infants and children. AIDS 2016; 30:2351–60.
- Evans C, Jones CE, Prendergast AJ. HIV-exposed, uninfected infants: new global challenges in the era of paediatric HIV elimination. Lancet Infect Dis 2016; 16:e92–107.
- Wedderburn CJ, Evans C, Yeung S, Gibb DM, Donald KA, Prendergast AJ. Growth and neurodevelopment of HIV-exposed uninfected children: a conceptual framework. Current HIV/AIDS Rep 2019. doi:10.1007/ s11904-019-00459-0.
- Slogrove AL, Becquet R, Chadwick EG, et al. Surviving and thriving-shifting the public health response to HIV-exposed uninfected children: report of the 3rd HIV-exposed uninfected child workshop. Front Pediatr 2018; 6:157.
- Chintu C, Bhat GJ, Walker AS, et al; CHAP Trial Team. Co-trimoxazole as prophylaxis against opportunistic infections in HIV-infected Zambian children (CHAP): a double-blind randomised placebo-controlled trial. Lancet 2004; 364:1865–71.
- World Health Organization. Guidelines on postexposure prophylaxis for HIV and the use of co-trimoxazole prophylaxis for HIV-related infections among adults, adolescents and children: recommendations for a public health approach— December 2014 supplement to the 2013 consolidated ARV guidelines. Geneva, Switzerland: WHO Press, 2014.
- Daniels B, Coutsoudis A, Moodley-Govender E, et al. Effect of co-trimoxazole prophylaxis on morbidity and mortality of HIV-exposed, HIVuninfected infants in South Africa: a randomised controlled, non-inferiority trial. Lancet Glob Health 2019; 7:e1717–27.
- Church JA, Fitzgerald F, Walker AS, Gibb DM, Prendergast AJ. The expanding role of co-trimoxazole in developing countries. Lancet Infect Dis 2015; 15:327–39.
- Bourke CD, Gough EK, Pimundu G, et al. Cotrimoxazole reduces systemic inflammation in HIV infection by altering the gut microbiome and immune activation. Sci Trans Med 2019; 11. doi:10.1126/scitranslmed.aav053.
- 11. Kyosiimire-Lugemwa J, Anywaine Z, Abaasa A, et al. Effect of stopping cotrimoxazole preventive therapy on microbial translocation and inflammatory markers among human immunodeficiency virus-infected Ugandan adults on antiretroviral therapy: the COSTOP Trial Immunology Substudy [manuscript published online ahead of print 23 October 2019]. J Infect Dis 2019. doi:10.1093/ infdis/jiz494.

- Ewing AC, King CC, Wiener JB, et al. Effects of concurrent exposure to antiretrovirals and cotrimoxazole prophylaxis among HIV-exposed, uninfected infants. AIDS 2017; 31:2455–63.
- Lockman S, Hughes M, Powis K, et al. Effect of co-trimoxazole on mortality in HIV-exposed but uninfected children in Botswana (the Mpepu Study): a double-blind, randomised, placebocontrolled trial. Lancet Glob Health 2017; 5:e491–500.
- Gill CJ, Sabin LL, Tham J, Hamer DH. Reconsidering empirical cotrimoxazole prophylaxis for infants exposed to HIV infection. Bull World Health Organ 2004; 82:290–7.
- Graham SM. Cotrimoxazole prophylaxis for infants exposed to HIV infection. Bull World Health Organ 2004; 82:297–8.
- Juma DW, Muiruri P, Yuhas K, et al. The prevalence and antifolate drug resistance profiles of *Plasmodium falciparum* in study participants randomized to discontinue or continue cotrimoxazole prophylaxis. PLoS Negl Trop Dis 2019; 13:e0007223.
- Powis KM, Souda S, Lockman S, et al. Cotrimoxazole prophylaxis was associated with enteric commensal bacterial resistance among HIVexposed infants in a randomized controlled trial, Botswana. J Int AIDS Soc 2017; 20:e25021.
- van der Veen EL, Schilder AG, Timmers TK, et al. Effect of long-term trimethoprim/sulfamethoxazole treatment on resistance and integron prevalence in the intestinal flora: a randomized, double-blind, placebo-controlled trial in children. J Antimicrob Chemother 2009; 63:1011–6.
- Robertson RC, Manges AR, Finlay BB, Prendergast AJ. The human microbiome and child growth—first 1000 days and beyond. Trends Microbiol 2019; 27:131–47.
- Bender JM, Li F, Martelly S, et al. Maternal HIV infection influences the microbiome of HIVuninfected infants. Sci Trans Med 2016; 8:349ra100.
- Claassen-Weitz S, Gardner-Lubbe S, Nicol P, et al. HIV-exposure, early life feeding practices and delivery mode impacts on faecal bacterial profiles in a South African birth cohort. Sci Rep 2018; 8:5078.
- Oldenburg CE, Sié A, Coulibaly B, et al. Effect of commonly used pediatric antibiotics on gut microbial diversity in preschool children in Burkina Faso: a randomized clinical trial. Open Forum Infect Dis 2018; 5:ofy289.
- Quince C, Walker AW, Simpson JT, Loman NJ, Segata N. Shotgun metagenomics, from sampling to analysis. Nat Biotechnol 2017; 35:833–44.
- Willmann M, Vehreschild MJGT, Biehl LM, et al. Distinct impact of antibiotics on the gut microbiome and resistome: a longitudinal multicenter cohort study. BMC Biol 2019; 17:76.
- Bwakura-Dangarembizi M, Kendall L, Bakeera-Kitaka S, et al. A randomized trial of prolonged co-trimoxazole in HIV-infected children in Africa. N Engl J Med 2014; 370:41–53.
- Mulenga V, Ford D, Walker AS, et al; CHAP Trial Team. Effect of cotrimoxazole on causes of death, hospital admissions and antibiotic use in HIVinfected children. AIDS 2007; 21:77–84.
- Slogrove AL, Powis KM, Johnson LF, Stover J, Mahy M. Estimates of the global population of children who are HIV-exposed and uninfected, 2000–18: a modelling study. Lancet Glob Health 2020; 8:e67–75.