ORIGINAL ARTICLE

Characteristics of intestinal microbiota and enzyme activities in mice fed with lily bulb

Yi Wu1 · Chenyang Zhang1 · Haoqing Shao¹ · Huaihao Luo1 · Zhoujin Tan[1](http://orcid.org/0000-0003-3193-073X)

Received: 28 September 2020 / Accepted: 15 December 2020 / Published online: 2 January 2021 © King Abdulaziz City for Science and Technology 2021

Abstract

The aim was to investigate the efect of lily bulbs on the microecological characteristics of intestinal microbiota and enzyme activities in normal mice. Thirty SPF Kunming mice were randomly divided into the control group, *Lilium lancifolium* (LL) group and *Lilium davidi*i var. unicolor (LDU) group. Mice of the latter two groups were given 0.15 g·mL−1 lily bulb solution, respectively, by gavage twice a day, while the control group was given the same volume of sterilized water. After 49 days, intestinal contents and mucosa of all mice were collected and the characteristics of intestinal microbiota and enzyme activities were analyzed. Results showed that the number of *Lactobacillus* spp. and *Bifdobacteria* spp. in the LL group was significantly higher than that in the control group $(t=2.68\times10^7, P=0.000; t=5.96\times10^7; P=0.000)$ and the LDU group $(t=6.12\times10^7, P=0.000; t=2.71\times10^7, P=0.000)$, while the number of total bacteria was significantly lower (*P* = 0.040). Microbial activity in intestinal contents and mucosa of the LDU group $(t=0.43, P=0.001; t=0.69, P=0.000)$ decreased, and microbial activity in intestinal mucosa of the LL group decreased significantly $(t=0.89, P=0.000)$ but increased significantly in intestinal contents of the LL group $(t=0.81, P=0.000)$. The activities of amylase $(t=455.73, P=0.000; t=206.56,$ *P*=0.000) and protease (*t*=52.32, *P*=0.000) increased but the activities of lactase (*t*=443.51, *P*=0.000; *t*=15.71, *P*=0.000) and sucrase $(t=5.82, P=0.000; t=366.82, P=0.000)$ decreased significantly in contents from the LL group and LDU group. Except for the sucrase activity, enzyme activities in mucosa of the LL group were completely opposite to those in contents $(t=44.15, P=0.000; t=1.25, P=0.007; t=14.64, P=0.011)$. In conclusion, dietary lily bulbs increased intestinal contents amylase activities and mucosa lactase activity signifcantly. Lily bulbs, especially *Lilium lancifolium*, can promote the growth of *Lactobacillus* spp. and *Bifdobacteria* spp., and inhibit the growth of total bacteria in the intestines of normal mice. *Lilium lancifolium* bulbs have the potential to be a functional food.

Keywords Lily bulb · Intestinal microbiota · Enzyme activity · Functional food

Introduction

The intestinal tract is an important digestive organ and immune organ of the human body, in which intestinal digestive enzymes and intestinal microbiota play a vital role. Intestinal microbiota, distributed in contents and intestinal mucosa, has a total of 10^{14} colony-forming units (CFU), weighing about 1.0–1.5 kg, and participates in the physiological, pathological and toxicological process (Sekirov

 \boxtimes Huaihao Luo 980195904@qq.com

 \boxtimes Zhoujin Tan tanzhjin@sohu.com et al. [2010\)](#page-6-0). Under physiological conditions, the microecosystem is in dynamic balance (Gilbert et al. [2018](#page-6-1)). For example, in bacterial membrane barriers, *Bifdobacteria* spp. can inhibit colonization and growth of pathogenic bacteria by occupying and competing nutrients, thus maintaining human health. *Lactobacillus* can inhibit *Enterohemorrhagic*, *Escherichia coli*, *Salmonella, Shigella* and *Vibrio cholerae* by secreting rhodobactin (Spinler et al. [2008\)](#page-6-2) . However, once this balance is destroyed, more diseases will occur, especially chronic diseases (Kalliomäki and Walker [2005](#page-6-3)) . Some studies have shown that the occurrence of basic gastrointestinal diseases and nervous system diseases is related to the imbalance of intestinal microecosystem (Han et al. [2009](#page-6-4); Kim et al. [2019](#page-6-5)).

Intestinal digestive enzymes refect the digestive function of monogastric animals to a certain extent, and play an

¹ Hunan University of Chinese Medicine, Xueshi Road 300, Yuelu District, Changsha 410208, Hunan Province, China

important role in nutrient absorption and immunity. According to the sources, intestinal digestive enzymes can be divided into endogenous digestive enzymes and exogenous digestive enzymes. The former is secreted by the small intestinal mucosa, such as amylase and lipase. Substances, such as plant polysaccharides, disaccharides, oligosaccharides and lignin, cannot be metabolized by the host, but are digested by digestive enzymes released by the intestinal microbiota. After being decomposed by enzymes released by intestinal bacteria, these substances are metabolized into butyric acid and methane, amino acids, etc., which are also called intestinal microbial metabolites and further participate in the physiological and pathological processes. However, the contribution of intestinal bacteria and intestinal mucosal bacteria to digestive enzyme activity and immunity may be diferent. Bacteria in intestinal mucosa are closer to the host epithelium than contents. Long et al. found that the bacterial composition of the intestinal mucosa may be diferent from the intestinal contents (Long et al. [2018](#page-6-6)). Zeng et al. discovered mucosa microbiota may play a more important role in the occurrence and development of gastrointestinal diseases (Zeng et al. [2019](#page-7-0)). Therefore, our study focuses not only on changes in enzyme activity in intestinal contents but also on the changes of enzyme activity in the intestinal mucosa.

Lily bulb is the earliest food that can be used as both food and medicine, with a potent ability to enhance immunity and antioxidation, anti-infammatory, anti-depressant (Chang et al. [2007](#page-6-7); Pan et al. [2017;](#page-6-8) Hui et al. [2020\)](#page-6-9), and is very popular in East Asia. It was reported that it contains starch, protein, amino acids and trace elements, and also contains active ingredients such as polyphenols, favonoids, alkaloids and saponins (Liu et al. [2019](#page-6-10)). The compounds in the lily bulbs may vary from region to region. We used two common varieties of lily in China—*Lilium lancifolium* and *Lilium davidii* var. *Unicol* (Fig. [1\)](#page-1-0) in the study. There are few studies on the intestinal microbiota of lily bulb, but the conclusions are signifcant. Okazaki et al. found dextran sulfate sodium (DSS)-induced colitis in rats showed a lower fecal abundance of *bifdobacteria*, but it was recovered by lily bulbs intake (Okazaki et al. [2016a](#page-6-11), [b\)](#page-6-12). In another of his articles, eating lily can reduce the fecal *Firmicutes*/*Bacteroidetes* ratio, and thus resist obesity (Okazaki et al. [2016a](#page-6-11), [b\)](#page-6-12).

In our research, we use two species of lily bulbs and investigate the efect of lily bulbs on the microecological of normal mice from three indicators: main intestinal bacteria communities, enzyme activity, and microbial activity.

Fig. 1 Main components of *Lilium lancifolium* (**a**) and *Lilium davidii* var. unicolor (**b**)

Materials and methods

Animals and procedures

Thirty 4-week-old-specifc pathogen-free (SPF) Kunming mice (All mice are male), weighting 20 ± 2 g, were purchased from Hunan Slaccas Jingda Laboratory Animal Co., Ltd (SCXK (Xiang) 2019-0004). All procedures involving animals were reviewed and approved by the Animal Ethics and Welfare Committee of Hunan University of Chinese Medicine, Animal license SCXK (Xiang) 2019-0009. Mice were reared in a stable environment (temperature 23–25 °C, relative humidity 50–70%, 12 h light/dark cycles, free access to diet and water). According to *Pharmacopoeia of the People's Republic of China (2015)*, mice in *Lilium lancifolium* (LL) group and *Lilium davidii* var. unicolor (LDU) group were given 0.15 g·mL⁻¹ lily bulb solution at a dose of 3 g⋅kg⁻¹⋅day⁻¹, respectively, by gavage, twice a day for 49 days, while the control group was given equal sterile water by gavage.

Reagents

Four point fve grams ultrafne powders of LL and LDU (offered by Hunan Academy of TCM) was dissolved and

in boiling water, respectively, to make the fnal concentration was 0.15 g·mL−1. Foline-phenol was purchased from Hefei Bomei Biotechnology Co., Ltd (Anhui China). DNS reagent, o-Nitrophenyl β-D-galactopyranoside (ONPG) reagent, and four substrate solutions were prepared in the lab. Acetone was purchased from Hunan Huihong reagent Co., Ltd (Hunan China). Fluorescein diacetate (FDA) was purchased from Shanghai Yuanye Biotechnology Co., Ltd (Shanghai China), and sodium phosphate buffer was prepared in the lab.

Extraction of intestinal contents

All mice were sacrifced by cervical dislocation after being gavaged for 49 days. The intestinal contents (from the jejunum to the rectum) of all mice in each group were collected, respectively, in a sterile environment and cooled to 4 °C refrigerator (Guo et al. [2015\)](#page-6-13).

Intestinal mucosa extraction

Intestinal mucosa in each group was collected according to the collection method of sterile intestinal mucosa established by our research group (He et al. [2017\)](#page-6-14) . Under sterile conditions, the contents on the wall of the small intestine (the jejunum to the rectum) were washed with sterile saline, and then the intestinal mucosa was scraped off using coverslips and cooled to -80 °C liquid nitrogen tank.

Determination of microbiota in intestinal contents

A certain amount of intestinal contents from each group were weighed aseptically and transferred into the corresponding group's triangle bottle flled with sterile water and glass beads. Triangle bottles were shaken on a bed temperature incubator at 120 rpm for 30 min to release microorganisms from intestinal contents completely. The number of microorganisms was counted by dilution plate culture counting. The experiment was performed in triplicate. Total numbers of bacteria were detected after being cultured at 37 °C for 24 h, the numbers of *Lactobacillus* spp. and *Bifdobacteria* spp. were determined after being anaerobic cultured at 37 °C for 48 h.

Analysis of enzyme activities of intestinal in mice

The intestinal mucosa and contents were transferred into the corresponding centrifuge tubes flled with glass beads and added an appropriate amount of sterile water. Then, the intestinal mucosa and contents were shaken on a vortex mixer for 10 min to release enzymes completely and centrifuged at 3000 rpm for 8 min. Enzyme activities were analyzed by UV spectrophotometer. Protease activity was determined by the Foline-phenol method, the activities of amylase and sucrase were determined by the DNS colorimeter method and lactase by the ONPG method. The unit of enzyme activity is defned according to the reference (Tan et al. [2012](#page-6-15)).

Determination of microbial activity (Schnürer and Rosswall [1982](#page-6-16); Tang et al. [2020](#page-7-1))

FDA was dissolved in acetone and stored as a stock solution (2 mg/mL) at $-$ 20 °C, the sample (intestinal mucosa and contents) was simultaneously added with FDA (fnal concentration, $10 \mu g/mL$) to sterile sodium phosphate buffer, pH 7.6, and the mixture was incubated at 24 °C on a rotary shaker. Mixture was incubated at 24 °C on a rotary shaker. After 90 min, 2 mL of acetone was added to stop the reaction, and the value of A_{490} was determined. A group of blank control group was established, the sample (intestinal mucosa and contents) was simultaneously added with acetone, FDA (fnal concentration, 10 μg/mL) to sterile sodium phosphate buffer. Three repeated tests were performed on each sample.

Statistical analysis

Measurement data of each group were expressed by the form of mean \pm standard deviation and analyzed using a single factor of variance (one-way ANOVA) in SPSS 21.0 software. LSD-t method was performed for multiple comparisons. Diferences among groups were considered signifcant when *P* values less than 0.05.

Results

Efects of diferent lily bulbs on mice body weight

The weight of the mice can be seen from Fig. [2](#page-3-0). Generally speaking, the weight of each group of mice increased with an increase in feeding time. Specifcally, compared with the control group, the weight of mice in the lily bulb group gradually decreased with time, but there was no signifcant difference between the control group $(P > 0.05)$. The weight of the mice in the LDU group was gradually lower than that in the LL group, but there was no signifcant diference between the two lily bulb groups $(P > 0.05)$.

Efects of lily bulbs on microbiota in mice contents

Intestinal microbiota plays an important role in the human body. It can be seen from Fig. [3](#page-3-1), the total numbers of bacteria in the LL group was signifcantly lower than that in the control group $(t = 2.21 \times 10^6, P = 0, 0.040)$, while the number of *Lactobacillus* spp. and *Bifidobacteria* spp.

Fig. 2 Body weight changes of control group (\bullet) , LL group (\square) , LDU group (Δ) . Each value is the mean \pm SD of five replicates $(n=5)$

Fig. 3 They represented the number of total bacteria, *Bifdobacteria spp* and *Lactobacillus* spp. colony between diferent groups. The colony numbers of the three strains in the LL group and other groups (the control group and the LDU group) were signifcantly diferent. Compared with the control group, $^{A}P < 0.05$, $^{a}P < 0.01$; Compared with the *Lilium lancifolium* group, ${}^{b}P$ < 0.01

was signifcantly higher than those in the control group $(t=2.68\times10^7, P=0.000; t=5.96\times10^7; P=0.000)$. Different from the LL group, the changes of the number of three culturable microbiota in the LDU group was not signifcant difference from that in the control group $(P > 0.05)$. Similar to the control group, the number of bacteria in the LDU group was signifcantly higher than that in the LL group

 $(t=4.0\times10^6, P=0.003)$, while the number of *Lactobacillus* spp. and *Bifdobacteria* spp. was signifcantly lower than that in the LL group $(t=6.12 \times 10^7, P=0.000; t=2.71 \times 10^7,$ $P=0.000$).

Efects of lily bulbs on enzyme activities in mice contents

Table [1](#page-3-2) demonstrates the efects of two kinds of lilies on the enzyme activity in contents. The activities of four digestive enzymes in the LL group were signifcantly diferent from those in the control group. The activities of amylase and protease were signifcantly higher than those in the control group (*t*=455.73, *P*=0.000; *t*=52.32, *P*=0.000), while the activities of sucrase and lactase were significantly lower than those of the control group ($t = 5.82$, $P = 0.000$; $t = 443.51$, $P = 0.000$. The changes in the four digestive enzyme activities of the LDU group were similar to those of the LL group. Except for protease, the other three enzyme activities were signifcantly diferent from the control group (*t* = 206.56, *P* = 0.000; *t* = 366.82, *P* = 0.000; *t* = 15.71, $P=0.000$. The activities of amylase, protease and sucrase in the LDU group were signifcantly lower than those of the LL group (*t*=249.18, *P*=0.000; *t*=52.15, *P*=0.000; $t=9.89$, $P=0.000$), while lactase activity was significantly higher than that in the LL group $(t=76.69, P=0.000)$.

Efects of lily bulbs on mucosa enzyme activity in mice

As shown in Table [2](#page-4-0), contrary to the changes of enzyme activity in mice contents, the activities of amylase and protease in the intestinal mucosa of the LL group (*t*=44.15, *P*=0.000; *t*=1.25, *P*=0.007) and the LDU group (*t*=15.41, *P*=0.012; *t*=0.867, *P*=0.031) were obviously lower than that of the control group. In addition, the amylase and protease activities in the LL group were lower than those in the LDU group, and the amylase activity was signifcantly different $(t=28.74, P=0.001)$. There was no significant difference between the LL group and the control group in the activity of sucrase in the intestinal mucosa, but the activity of sucrase in the LDU group was signifcantly higher than that of the control group $(t=3.49, P=0.015)$. Compared with the control group and the LL group, lactase activity in

Table 1 Efects of diferent lilies on enzyme activity in feces

Each value is the mean \pm standard deviation, $N=5$. Compared with the control group, ${}^{3}P$ < 0.01; compared with the *Lilium lancifolium* group, ${}^{b}P$ < 0.01

Table 2 Efects of diferent lilies on enzyme activity in mucosa

Each value is the mean \pm standard deviation, $N=5$. Compared with the control group, AP < 0.05, ap \geq 0.01; compared with the *Lilium lancifolium* group, ${}^{b}P$ < 0.01

Fig. 4 They represented microbial activity in feces of control $group (①), LL group (②), LDU$ group (Δ) . The microbial activity of LL group was the strongest, followed by the normal group, and fnally the LL group. Compared with the control group, a *P*<0.01; Compared with the *Lilium lancifolium* group, $\frac{b}{P}$ < 0.01

A₄₉₀ vaules

5

intestinal mucosa of the LDU group increased signifcantly $(t = 34.45, P = 0.000; t = 19.81, P = 0.003)$. The lactase activity of the intestinal mucosa in the LL group was higher than that in the control group, with a signifcant diference $(t=14.64, P=0.011).$

Efects of lily bulbs on microbial activity in mice contents

Figure [4](#page-4-1) demonstrates the effects of two lilies on the microbial activity in mice contents. Microbial activity of the LL group was signifcantly higher than that of the control group $(t=0.81, P=0 0.000)$, while that of the LDU group was significantly lower than that of the control group $(t=0.43,$ $P=0.001$). Compared with the LL group, the microbial activity of the LDU group decreased significantly $(t=1.23)$, $P=0.000$).

Efects of lily bulbs on microbial activity in mice mucosa

The results of the microbial activity in mucosa are shown in Fig. [5.](#page-4-2) Compared with the control group, the mucosa microbial activity of the LL group and the LDU group was significantly decreased $(t=0.89, P=0.000; t=0.69, P=0.000)$. There was no signifcant diference in the mucosa microbial activity between the LL group and the LDU group $(P>0.05)$.

65 Control group 60 Lilium lancifolium group 55 Lilium davidii var. unicolor group 50 45 40 $35₅$ 30 25 20 \mathbf{a} 15 $10[°]$

Fig. 5 They represented microbial activity in mucosa of control group (\bullet), LL group (\Box), LDU group (Δ). The microbial activity in the normal group was the strongest, and there was no diference between the LL group and the LDU group. Compared with the control group, a *P*<0.01

Discussion and conclusion

Lily bulbs can regulate the digestive enzyme activity and microbial activity of the intestinal contents in normal mice

The composition of lily bulbs is complex, and intestinal digestive enzymes are easily afected by various active substances in lily. The regulatory mechanism may include the regulation of the intestinal mucosa, intestinal

microbiota and intestinal microenvironment (Long et al. [2017](#page-6-17)). The digestive enzymes in the intestinal contents are mainly endogenous digestive enzymes, which are determined by the enzyme-producing microbial community. The larger number of enzyme-producing microbial communities, the higher digestive enzyme activity, the more complete absorption and utilization of foods, and the more intestinal microbial metabolites. Fluorescein diacetate, also known as FDA, can be hydrolyzed by esterases, proteases, lipases, etc. (Schnürer and Rosswall [1982\)](#page-6-16). The changes in intestinal microbial enzyme activity can be refected using the method of FDA. Combined with changes in intestinal microbiota and digestive enzyme activity, it can help us to better understand changes in the intestinal microenvironment (Tang et al. [2020\)](#page-7-1).

The study has shown that lily bulbs, especially *Lilium lancifolium*, increase the activity of microbial in intestinal contents. At the same time, amylase and protease activities in intestinal contents are increased signifcantly by lily. Amylase and protease are secreted by the intestinal tract and intestinal microorganisms, which are closely related to the digestion and absorption function of the small intestine. Our research shows that eating lily bulbs can improve the activity of amylase and protease in the small intestine, and enhance the digestion and absorption of food. Amylase is the main enzyme for digesting carbohydrates, studies showed that when weaned pigs consume more carbohydrates, the amylase activity will increase (Makkink et al. [1994\)](#page-6-18). Probiotics can stimulate the activity of digestive enzymes. Studies have shown that adding probiotics to aquatic feed can increase the digestion and absorption of fsh, and increase the activity of protease and amylase (Zhang et al. [2020](#page-7-2)). Long-term consumption of lily bulbs in mice and bacteria will stimulate the production of amylase. At the same time, the growth of *Lactobacillus* spp. and *Bifdobacteria* spp. in the intestinal contents of normal mice may also stimulate amylase activity to some extent.

Lily bulbs can regulate the digestive enzyme activity and microbial activity in the intestinal mucosa of normal mice

Maintain the normal morphology and structure of the intestinal mucosa so that intestinal digestive enzymes can digest and absorb nutrients. Normal intestinal mucosa can secrete digestive enzymes to digest and absorb nutrients. Once the structure of intestinal mucosa is destroyed, it will reduce the activity of digestive enzymes. According to reports, the chemical components in traditional Chinese medicine can thicken and lengthen intestinal villi, making it more compact and orderly, which has a positive impact on the production and secretion of digestive enzymes (Long et al. [2017](#page-6-17)). In other words, changes in enzyme activity of intestinal

mucosa can refect changes in the intestinal mucosa structure. Jiang et al. found that the addition of 1.0% glutamine to the feed increased the activities of sucrase, maltase, and lactase signifcantly in the jejunum of 14-day-old pigs (Jiang et al. [2010\)](#page-6-19). Investigations demonstrated that probiotics can increase the activity of diabylase (referring to lactase, sucrase, and maltase) and intestinal lipase in the small intestinal mucosa, and regulate the intestinal microbiota (Yang et al. [2005](#page-7-3)).

Eating lily enhanced the activity of lactase in the intestinal mucosa, which was mainly produced by small intestinal epithelial cells and decomposed lactose into galactose. Galactose is an oligosaccharide that provides nutrients for probiotics. In the studies, the microbial activity of the LDU group was lower than that of the normal group, so were the activity of mucosa amylase and protease. However, lily bulbs increased the activity of lactase in the mucosa, which indicated that lily bulb mainly promote the secretion of lactase in the intestine. Galactose is the product of lactase decomposition, which can promote the growth of probiotics, and probiotics can stimulate the activity of amylase and sucrose (Zhang et al. [2020](#page-7-2)). All the above can promote the digestion and absorption of the body and enhance the intestinal immune function.

Lily bulbs can promote the growth of *Lactobacillus spp.* **and** *Bifdobacteria spp.*

Our experimental results show that eating lily bulbs promote the growth of *Lactobacillus* spp. and *Bifdobacteria* spp. The experiment of Gong et al. confrmed that lily bulbs extract has a positive infuence on the growth of *Bifdobacteria* spp. and even higher than that of Isomalto-oligosaccharide. Gong thinks that several oligosaccharides of lily bulbs promote the growth of *Bifdobacteria* spp. and put forward that lily bulbs have the potential as a bifdus factor or functional food (Gong et al. [2009](#page-6-20)). Zhao et al. found that purifed lily bulb polysaccharide can restore the microbiota of mice with intestinal microbiota disorders (Zhao et al. [2020\)](#page-7-4) . The research by Sim et al. revealed that Korean *L. lancifolium* bulbs are more anti-infammatory and anti-aging than Japanese Lily (Sim et al. [2020\)](#page-6-21). In our study, we also found that *L*. *lancifolium* bulbs increase the number of *Lactobacillus* spp. and *Bifdobacteria* spp. more than *Lilium davidii* var. *Unicol* bulbs.

Our experiments have proven that lily bulbs increased amylase activities in intestine contents signifcantly and improved digestion and absorption of foods. In addition, lily bulbs stimulate intestinal epithelial cells to secrete lactase, increase mucosa lactase activity, and maintain intestinal mucosa immunity. Lily bulbs promote the growth of *Lactobacillus* spp. and *Bifdobacteria* spp. and inhibit the growth of total bacteria in the intestines of normal mice, indicating that lily may maintain the intestinal mucosa barrier. This may be related to the oligosaccharides (such as glucomannan) of lily. Above all, the study might contribute to improving the current understanding the interaction between lily bulbs and intestinal digestive enzymes and microbiota, and is worthy of further study. Lily bulbs, especially *L. lancifolium* bulb has the potential to be functional foods, and it is necessary to fgure out which ingredients of lily bulbs can improve intestinal microecology.

Acknowledgements Thanks for the ethical certifcation of Hunan University of Chinese Medicine.

Author contribution ZT designed the study; HS and CZ performed the experiments; YW analyzed the data; YW wrote the paper; ZT and HL checked the paper. The decision to submit the manuscript for publication was made by all the authors.

Funding No funded projects for this experiment.

Data availability Material and raw data are available.

Compliance with ethical standards

Conflict of interest The authors declare that there is no confict of interests regarding the publication of this paper.

Institutional animal care and use committee statement The study was approved by the Animal Ethics and Welfare Committee of Hunan University of Chinese Medicine.

References

- Chang YZ, Zhong SM, Cao YC, Wang YX (2007) Study on the function of lily dietary fber on the moderating hyperlipaemia and weight lose in the SD. Food Sci Technol 09:245–247. [https://doi.](https://doi.org/10.3969/j.issn.1005-9989.2007.09.080) [org/10.3969/j.issn.1005-9989.2007.09.080](https://doi.org/10.3969/j.issn.1005-9989.2007.09.080)
- Gilbert JA, Blaser MJ, Caporaso JG, Jansson JK, Lynch SV, Knight R (2018) Current understanding of the human microbiome. Nat Med 24:392–400.<https://doi.org/10.1038/nm.4517>
- Gong FH, He S, Zhang DC, Guo (2009) Preliminary study on growth promotion of lily bulbs extract on bifdobacterium. Food Ferment Ind 35(11):64–67. https://doi.org/10.1007/978-3-540-85168-4_52
- Guo KX, Tan ZJ, Xie MZ, She Y, Wang YH (2015) The synergic efect of ultra-micro powder Qiweibaizhusan combined with yeast on dysbacteriotic diarrhea mice. Chin J Appl Environ Biol 21(01):61–67.<https://doi.org/10.3724/SP.J.1145.2013.10002>
- Han XY, Deng H, Cai Y, Fu JH, Yu Z, Ren X, Yin XL, Xie C (2009) Intestinal microbes and chronic diseases. Chin J Micro Ecol 21(11):1039–1042, 1046. https://doi.org/ CNKI:SUN:ZGWS.0.2009-11-023
- He L, Long C, Liu Y, Guo Y, Xiao N, Tan Z (2017) Effects of Debaryomyces hansenii treatment on intestinal microorganisms in mice with antibiotics-induced diarrhea. 3 Biotech 7:347. [https://doi.](https://doi.org/10.1007/s13205-017-0953-9) [org/10.1007/s13205-017-0953-9](https://doi.org/10.1007/s13205-017-0953-9)
- Hui H, Xin A, Cui H, Jin H, Yang X, Liu H, Qin B (2020) Anti-aging efects on Caenorhabditis elegans of a polysaccharide, O-acetyl glucomannan, from roots of Lilium davidii var. unicolor Cotton.

Int J Biol Macromol 155:846–852. [https://doi.org/10.1016/j.ijbio](https://doi.org/10.1016/j.ijbiomac.2020.03.206) [mac.2020.03.206](https://doi.org/10.1016/j.ijbiomac.2020.03.206)

- Jiang ZY, Zheng WC, Lin YC, Zheng CT, Huang L, Chen F (2010) Efects of glutamine on growth performance, serum hormone levels and intestinal mucosal enzyme activity in newborn piglets. Chin J Anim Nutr 22(1):125–131. [https://doi.org/10.3969/j.](https://doi.org/10.3969/j.issn.1006-267x.2010.01.019) [issn.1006-267x.2010.01.019](https://doi.org/10.3969/j.issn.1006-267x.2010.01.019)
- Kalliomäki MA, Walker WA (2005) Physiologic and pathologic interactions of bacteria with gastrointestinal epithelium. Gastroenterol Clin North Am 34(383–399):vii. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.gtc.2005.05.007) [gtc.2005.05.007](https://doi.org/10.1016/j.gtc.2005.05.007)
- Kim S, Kwon SH, Kam TI, Panicker N, Karuppagounder SS, Lee S, Lee JH, Kim WR, Kook M, Foss CA, Shen C, Lee H, Kulkarni S, Pasricha PJ, Lee G, Pomper MG, Dawson VL, Dawson TM, Ko HS (2019) Transneuronal propagation of pathologic α-synuclein from the gut to the brain models Parkinson's disease. Neuron 103:627-641.e7. <https://doi.org/10.1016/j.neuron.2019.05.035>
- Liu P, Lin ZJ, Zhang B (2019) Research progress on chemical constituents and pharmacological efect of *Lilii Bulbus*. Chin J Exp Trad Med Formulae 23(23):201–211. [https://doi.org/10.13422/j.cnki.](https://doi.org/10.13422/j.cnki.syfjx.2017230201) [syfx.2017230201](https://doi.org/10.13422/j.cnki.syfjx.2017230201)
- Long CX, Guo YF, Liu YW, Peng XX, Tan ZJ (2017) Immunoprotective effect of traditional Chinese medicine on intestinal mucosa. World Chin J Digest 25(35):3115–3122. [https://doi.org/10.11569](https://doi.org/10.11569/wcjd.v25.i35.3115) [/wcjd.v25.i35.3115](https://doi.org/10.11569/wcjd.v25.i35.3115)
- Long C, Liu Y, He L, Yu R, Li D, Tan Z, Hui H (2018) Bacterial lactase genes diversity in intestinal mucosa of dysbacterial diarrhea mice treated with Qiweibaizhu powder. 3 Biotech 8:423. [https://doi.](https://doi.org/10.1007/s13205-018-1460-3) [org/10.1007/s13205-018-1460-3](https://doi.org/10.1007/s13205-018-1460-3)
- Makkink CA, Berntsen PJ, den Kamp op BM, Kemp B, Verstegen MW (1994) Gastric protein breakdown and pancreatic enzyme activities in response to two diferent dietary protein sources in newly weaned pigs. J Anim Sci 72:2843–2850. [https://doi.](https://doi.org/10.2527/1994.72112843x) [org/10.2527/1994.72112843x](https://doi.org/10.2527/1994.72112843x)
- Okazaki Y, Chiji H, Kato N (2016a) Protective efect of dietary lily bulb on dextran sulfate sodium-induced colitis in rats fed a highfat diet. J Nutr Sci Vitaminol (Tokyo) 62:206–212. [https://doi.](https://doi.org/10.3177/jnsv.62.206) [org/10.3177/jnsv.62.206](https://doi.org/10.3177/jnsv.62.206)
- Okazaki Y, Sekita A, Chiji H, Kato N (2016b) Consumption of lily bulb modulates fecal ratios of frmicutes and bacteroidetes phyla in rats fed a high-fat diet. Food Sci Biotechnol 25:153–156. [https](https://doi.org/10.1007/s10068-016-0112-9) [://doi.org/10.1007/s10068-016-0112-9](https://doi.org/10.1007/s10068-016-0112-9)
- Pan G, Xie Z, Huang S, Tai Y, Cai Q, Jiang W, Sun J, Yuan Y (2017) Immune-enhancing efects of polysaccharides extracted from Lilium lancifolium Thunb. Int Immunopharmacol 52:119–126. <https://doi.org/10.1016/j.intimp.2017.08.030>
- Schnürer J, Rosswall T (1982) Fluorescein diacetate hydrolysis as a measure of total microbial activity in soil and litter. Appl Environ Microbiol 43:1256–1261. [https://doi.org/10.1128/](https://doi.org/10.1128/AEM.43.6.1256-1261.1982) [AEM.43.6.1256-1261.1982](https://doi.org/10.1128/AEM.43.6.1256-1261.1982)
- Sekirov I, Russell SL, Antunes LC, Finlay BB (2010) Gut microbiota in health and disease. Physiol Rev 90:859–904. [https://doi.](https://doi.org/10.1152/physrev.00045.2009) [org/10.1152/physrev.00045.2009](https://doi.org/10.1152/physrev.00045.2009)
- Sim WS, Choi SI, Jung TD, Cho BY, Choi SH, Park SM, Lee OH (2020) Antioxidant and anti-infammatory efects of Lilium lancifolium bulbs extract. J Food Biochem 44:e13176. [https://doi.](https://doi.org/10.1111/jfbc.13176) [org/10.1111/jfbc.13176](https://doi.org/10.1111/jfbc.13176)
- Spinler JK, Taweechotipatr M, Rognerud CL, Ou CN, Tumwasorn S, Versalovic J (2008) Human-derived probiotic Lactobacillus reuteri demonstrate antimicrobial activities targeting diverse enteric bacterial pathogens. Anaerobe 14:166–171. [https://doi.](https://doi.org/10.1016/j.anaerobe.2008.02.001) [org/10.1016/j.anaerobe.2008.02.001](https://doi.org/10.1016/j.anaerobe.2008.02.001)
- Tan ZJ, Wu H, Liu FL, Cai Y, Cai GX, Zeng A (2012) Efect of ultramicro powder qiweibaizhusan on the intestinal microbiota and enzyme activities in mice. Acta Ecol Sin 32(21):6856–6863. [https](https://doi.org/10.5846/stxb201109271422) [://doi.org/10.5846/stxb201109271422](https://doi.org/10.5846/stxb201109271422)

- Tang Y, Wu Y, Hui HY, Tan ZJ (2020) Effet of Tongxieyaofang prescription on intestinal microbia lactivity in mice with Ganqichenpi diarrhea. Chin J Micro Ecol 32(01):17–20. [https://doi.](https://doi.org/10.13381/j.cnki.cjm.202001004) [org/10.13381/j.cnki.cjm.202001004](https://doi.org/10.13381/j.cnki.cjm.202001004)
- Yang SC, Chen JY, Shang HF, Cheng TY, Tsou SC, Chen JR (2005) Efect of synbiotics on intestinal microfora and digestive enzyme activities in rats. World J Gastroenterol 11:7413–7417. [https://doi.](https://doi.org/10.3748/wjg.v11.i47.7413) [org/10.3748/wjg.v11.i47.7413](https://doi.org/10.3748/wjg.v11.i47.7413)
- Zeng A, Peng M, Liu H, Guo Z, Xu J, Wang S, He L, Tan Z (2019) Effects of Debaryomyces hansenii treatment on intestinal mucosa microecology in mice with antibiotic-associated diarrhea. PLoS ONE 14(11):e0224730. [https://doi.org/10.1371/journ](https://doi.org/10.1371/journal.pone.0224730) [al.pone.0224730](https://doi.org/10.1371/journal.pone.0224730)
- Zhang JJ, Yang HL, Yan YY, Zhang CX, Ye JD, Sun YZ (2020) Efects of fsh origin probiotics on growth performance, immune response and intestinal health of shrimp (Litopenaeus vannamei) fed diets with fsh meal partially replaced by soybean meal. Aquacult Nutr 26:1255–1265.<https://doi.org/10.1111/anu.13081>
- Zhao ZM, Zhao H, Wang YL, Shen Y, Wang CX, Meng FL, Zhang M, ZhangY (2020) Purifcation of lily polysaccharide and its regulatory effect on intestinal flora dysregulation mice. Science and Technology of Food Industry 41(08):295–300, 306. [http://kns.](http://kns.cnki.net/kcms/detail/11.1759.TS.20191104.1456.022.html) [cnki.net/kcms/detail/11.1759.TS.20191104.1456.022.html](http://kns.cnki.net/kcms/detail/11.1759.TS.20191104.1456.022.html)

