CPAP minus during CPAP withdrawal) in extracellular water and weight was observed (R = 0.590; P = 0.004).

Discussion

This study provides strong evidence that the weight gain associated with the treatment of OSA with CPAP is due to fluid accumulation. Extracellular fluid volume during CPAP was not statistically higher than during CPAP withdrawal. However, weight gain occurred after 1 week of CPAP and could not be explained by the level of physical activity, calorie intake, water intake, or BMR, all of which remained stable. The magnitude of weight change in this acute study was remarkably similar to a meta-analysis of previous longer-term studies (0.37 vs. 0.42 kg, respectively) (1). Positive energy balance and increase in fat (4) and lean body mass (3, 4) have been suggested as a mechanism of weight gain during CPAP. If positive energy balance were a prevailing explanation, one would expect progressive weight gain over time. However, the recent evidence of lack of weight gain after an average of 3.8 years of CPAP (9) suggests that weight gain during CPAP is not cumulative. Moreover, fluid accumulation may explain the previously reported increase in lean body mass (3), as the vast majority of lean body mass (70%) is composed of water (10). Similar to mechanical ventilation, the treatment of OSA with CPAP reduces preload and urinary volume and may lead to fluid accumulation (5). In line with other studies, we did not find reduction in B-type natriuretic peptide levels associated with CPAP, indicating that other mechanisms such as antidiuretic hormone release may be involved. Twenty-four-hour urinary volume and sodium excretion between CPAP and CPAP withdrawal groups were similar, suggesting that patients had already reached a steady state after 1 week. Future studies are necessary to understand fluid balance within the first week of CPAP. Finally, we found a positive correlation between changes in weight and extracellular fluid volume. In conclusion, extracellular fluid volume did not increase during CPAP use. However, weight gain occurred within the first week of CPAP use and could be explained by fluid accumulation caused by reversal of nocturia. Our findings provide evidence against a negative metabolic impact promoted by CPAP during OSA treatment.

Author disclosures are available with the text of this letter at www.atsjournals.org.

Sara Herculano, P.T. Gustavo F. Grad, M.D. Luciano F. Drager, M.D. André L. P. de Albuquerque, M.D. *Universidade de Sao Paulo Sao Paulo, Brazil*

Camila M. Melo, C.N. Federal University of Lavras Lavras, Brazil

Geraldo Lorenzi-Filho, M.D. Pedro R. Genta, M.D.* *Universidade de Sao Paulo Sao Paulo, Brazil*

ORCID ID: 0000-0002-6764-165X (P.R.G.).

*Corresponding author (e-mail: prgenta@usp.br).

References

- Drager LF, Brunoni AR, Jenner R, Lorenzi-Filho G, Benseñor IM, Lotufo PA. Effects of CPAP on body weight in patients with obstructive sleep apnoea: a meta-analysis of randomised trials. *Thorax* 2015;70: 258–264.
- Quan SF, Budhiraja R, Clarke DP, Goodwin JL, Gottlieb DJ, Nichols DA, et al. Impact of treatment with continuous positive airway pressure (CPAP) on weight in obstructive sleep apnea. J Clin Sleep Med 2013; 9:989–993.
- Hoyos CM, Killick R, Yee BJ, Phillips CL, Grunstein RR, Liu PY. Cardiometabolic changes after continuous positive airway pressure for obstructive sleep apnoea: a randomised sham-controlled study. *Thorax* 2012;67:1081–1089.
- Tachikawa R, Ikeda K, Minami T, Matsumoto T, Hamada S, Murase K, et al. Changes in energy metabolism after continuous positive airway pressure for obstructive sleep apnea. Am J Respir Crit Care Med 2016;194:729–738.
- Darmon M, Legrand M, Terzi N. Understanding the kidney during acute respiratory failure. *Intensive Care Med* 2017;43:1144– 1147.
- Miyazato M, Tohyama K, Touyama M, Nakamura H, Oshiro T, Ueda S, et al. Effect of continuous positive airway pressure on nocturnal urine production in patients with obstructive sleep apnea syndrome. *Neurourol Urodyn* 2017;36:376–379.
- Kohler M, Stoewhas AC, Ayers L, Senn O, Bloch KE, Russi EW, et al. Effects of continuous positive airway pressure therapy withdrawal in patients with obstructive sleep apnea: a randomized controlled trial. *Am J Respir Crit Care Med* 2011;184:1192–1199.
- Herculano S, Grad GF, Drager LF, Albuquerque ALP, Melo CM, Lorenzi-Filho G, et al. The role of fluid accumulation in acute weight gain during CPAP treatment in patients with obstructive sleep apnea [abstract]. Am J Respir Crit Care Med 2020;201:A4519.
- Ou Q, Chen B, Loffler KA, Luo Y, Zhang X, Chen R, et al.; SAVE investigators. The effects of long-term CPAP on weight change in patients with comorbid OSA and cardiovascular disease: data from the SAVE trial. Chest 2019;155:720–729.
- Sheng HP, Huggins RA. A review of body composition studies with emphasis on total body water and fat. Am J Clin Nutr 1979;32:630–647.

Copyright © 2021 by the American Thoracic Society

Check for updates

BAL Is Safe and Well Tolerated in Individuals with Idiopathic Pulmonary Fibrosis: An Analysis of the PROFILE Study

To the Editor:

Diagnosing interstitial lung disease (ILD) is frequently challenging. Improvements in the recognition of disease-specific radiological patterns have resulted in many cases of ILD being diagnosed noninvasively. Nonetheless, there remain many circumstances

P.L.M. is an Action for Pulmonary Fibrosis Research Fellow. R.G.J. is supported by a National Institute for Health Research Research Professorship (RP-2017-08-ST2-014). T.M.M. is supported by a National Institute for Health Research Clinician Scientist Fellowship (CS-2013-13-017) and a British Lung Foundation Chair in Respiratory Research (C17-3). The PROFILE study was funded by the Medical Research Council (G0901226) and GlaxoSmithKline R&D (CRT114316) and was sponsored by Nottingham University and Royal Brompton and Harefield National Health Service Foundation Trust.

Originally Published in Press as DOI: 10.1164/rccm.202004-1138LE on August 28, 2020

Table 1. Demographics

	No Bronchoscopy (<i>n</i> = 391)	Bronchoscopy (n = 223)	P Value
Age, yr, mean (SD)	71.8 (8.3)	67.8 (8.1)	<0.001
Sex, M, n (%)	302 (77.2)	170 (76.2)	0.776
Smoking history, ever/current, n (%)	276 (70.6)	141 (63.2)	0.06
FVC, %, mean (SD)	76.3 (18.1)	77.8 (18.2)	0.591
DL _{CO} , %, mean (SD)	48.5 (17.6)	47.6 (14.1)	0.79
FEV ₁ /FVC, mean (SD)	79.8 (7.8)	79.6 (7.4)	0.71

in which further investigations beyond imaging are required to establish a definitive ILD diagnosis. This is reflected in recent diagnostic guidelines for hypersensitivity pneumonitis that suggest bronchoscopy and cellular analysis of BAL in patients with newly identified ILD for whom the differential diagnosis includes fibrotic hypersensitivity pneumonitis (1). The current idiopathic pulmonary fibrosis (IPF) guidelines also provide a conditional recommendation to perform BAL in cases of newly detected ILD of apparently unknown cause, in which the computed tomographic pattern is not one of definite usual interstitial pneumonia (2). However, not all centers perform diagnostic bronchoscopy in patients with ILD; in part, this reflects concerns about safety. A small number of retrospective and anecdotal observations have been used to suggest that bronchoscopy in individuals with IPF could be associated with an increased risk of acute exacerbations or acute respiratory deterioration (3, 4). We therefore aimed to clarify the safety of BAL in patients with IPF using the PROFILE (Prospective Observation of Fibrosis in Lung Clinical Endpoints) study cohort (5).

Incident cases of multidisciplinary diagnosed IPF were recruited prospectively as part of the PROFILE study through the following two coordinating centers in the United Kingdom: Nottingham University Hospitals in Nottingham (NCT01134822) and Royal Brompton Hospital in London (NCT01110694). Patients were assessed at baseline, 1 month, 3 months, and 6 months, and annually for 3 years. Fiberoptic bronchoscopy with BAL was undertaken in a subset of the Brompton cohort at baseline. BAL was performed by instillation of 240 ml of warm saline (in four 60-ml aliquots) into a segment of the right middle lobe with gentle aspiration by hand (6). No other bronchoscopic procedures were performed. Immediate, 30-day, and 90-day adverse events as well as overall survival were evaluated and compared between subjects undergoing BAL and those who did not. Continuous variables are presented as means (SD), and categorical variables are presented as proportions. Differences between subject groups were evaluated with the Mann-Whitney test for continuous variables and the Fisher's exact test for categorical variables. Time-to-event curves were calculated using the Kaplan-Meier method and compared using the log-rank test. Associations between continuous explanatory variables and overall survival were explored with a Cox proportional hazards model.

Of 614 subjects who were prospectively recruited into the PROFILE study, 223 underwent bronchoscopy (36%). The 391 individuals with IPF who did not undergo BAL were older (71.8 vs. 67.8 yr; P < 0.001) than subjects in the bronchoscopy cohort but otherwise were well matched (Table 1). All subjects in the bronchoscopy cohort tolerated the procedure well, a cell differential



Figure 1. No significant difference in overall mortality in patients with idiopathic pulmonary fibrosis undergoing bronchoscopy. Shown are Kaplan-Meier curves comparing survival between individuals in the PROFILE (Prospective Observation of Fibrosis in Lung Clinical Endpoints) study undergoing bronchoscopy and those not undergoing bronchoscopy. Log-rank *P* test value is reported.

was available for all, and there were no immediate (<72 h) complications.

In the first 30 days after BAL, six patients (2.7%) reported complications. Two subjects described transient viral-type symptoms after the procedure, one subject described odynophagia (again transient), and three subjects were treated with antibiotics for presumed lower respiratory tract infection, with one case (0.4%) requiring an emergency room attendance but not admission. There was no difference in 30-, 60-, or 90-day all-cause mortality in those undergoing bronchoscopy compared with the no-bronchoscopy cohort. All-cause mortality at 90 days was 1.4% in the bronchoscopy cohort and 3.6% in the nonprocedure cohort.

There was no significant difference (P = 0.45) in overall mortality between patients who underwent bronchoscopy and those who did not (Figure 1). The median survival for patients undergoing bronchoscopy was 3.7 years. There remained no difference in survival after adjustment for age, sex, baseline % predicted FVC, baseline % predicted DL_{CO}, smoking status, and recruitment site in a multivariable Cox proportional hazards model (hazard ratio, 0.84; 95% confidence interval, 0.59–1.22; P = 0.364).

There are a number of limitations to our work. First, subjects were not randomly assigned to either the procedure or no-procedure arm, as the bronchoscopy component of the study was optional. Second, some of the most severe cases of IPF were not included, as only subjects able to safely undergo bronchoscopy were enrolled; allcause mortality was used as an endpoint rather than respiratoryrelated mortality because the necessary level of detail regarding cause of death was not available for the whole cohort. Finally, the nobronchoscopy cohort was on average older than the intervention cohort, a known risk factor for mortality in IPF. However, there remained no difference in overall survival when incorporating this into a Cox model of survival, implying the age difference between groups had no meaningful impact. Although we demonstrate no negative safety signal, we do not address cost effectiveness; something which may impact local decisions to perform BAL in the diagnostic assessment of ILD.

In summary, this prospectively recruited longitudinal cohort study demonstrates that bronchoscopy is a safe and well-tolerated procedure in individuals with IPF. Although the assessment of BAL in the diagnosis of fibrotic lung disease may have recently been overshadowed by the emergence of cryobiopsy (7) and other novel molecular techniques (8), it remains important in distinguishing specific forms of ILD from IPF (something that is highlighted in recently published diagnostic guidelines for hypersensitivity pneumonitis). Furthermore, BAL has an important role in proof-of-concept clinical trials (9) and as a research tool for understanding disease pathogenesis and discovering novel biomarkers (10, 11).

Author disclosures are available with the text of this letter at www.atsjournals.org.

Philip L. Molyneaux, M.B. B.S., Ph.D.* Royal Brompton Hospital London, United Kingdom and National Heart and Lung Institute, Imperial College London London, United Kingdom Jonathan J. Smith National Heart and Lung Institute, Imperial College London London, United Kingdom

Peter Saunders, M.B. B.S. Royal Brompton Hospital London, United Kingdom and

National Heart and Lung Institute, Imperial College London London, United Kingdom

Felix Chua, M.B. B.S., Ph.D. Royal Brompton Hospital London, United Kingdom

Athol U. Wells, M.D. Elisabetta A. Renzoni, M.D., Ph.D. Andrew G. Nicholson, M.D. *Royal Brompton Hospital London, United Kingdom* and *National Heart and Lung Institute, Imperial College London London, United Kingdom*

William A. Fahy, M.B. B.S. GlaxoSmithKline R&D Stevenage, United Kingdom

R. Gisli Jenkins, B.M., Ph.D. University of Nottingham Nottingham, United Kingdom

Toby M. Maher, B.M., Ph.D.[‡] Royal Brompton Hospital London, United Kingdom National Heart and Lung Institute, Imperial College London London, United Kingdom and University of Southern California Los Angeles, California

ORCID IDs: 0000-0002-7929-2119 (R.G.J.); 0000-0001-7192-9149 (T.M.M.).

*Corresponding author (e-mail: p.molyneaux@imperial.ac.uk). [‡]T.M.M. is Associate Editor of *AJRCCM*. His participation complies with American Thoracic Society requirements for recusal from review and decisions for authored works.

References

- Raghu G, Remy-Jardin M, Ryerson CJ, Myers JL, Kreuter M, Vasakova M, et al. Diagnosis of hypersensitivity pneumonitis in adults: an official ATS/JRS/ALAT clinical practice guideline. Am J Respir Crit Care Med 2020;202:e36–e69.
- Raghu G, Remy-Jardin M, Myers JL, Richeldi L, Ryerson CJ, Lederer DJ, et al.; American Thoracic Society, European Respiratory Society, Japanese Respiratory Society, and Latin American Thoracic Society. Diagnosis of idiopathic pulmonary fibrosis: an official ATS/ERS/JRS/ALAT clinical practice guideline. Am J Respir Crit Care Med 2018;198:e44–e68.
- Hiwatari N, Shimura S, Takishima T, Shirato K. Bronchoalveolar lavage as a possible cause of acute exacerbation in idiopathic pulmonary fibrosis patients. *Tohoku J Exp Med* 1994;174: 379–386.
- Sakamoto K, Taniguchi H, Kondoh Y, Wakai K, Kimura T, Kataoka K, et al. Acute exacerbation of IPF following diagnostic bronchoalveolar lavage procedures. *Respir Med* 2012;106:436–442.
- 5. Maher TM, Oballa E, Simpson JK, Porte J, Habgood A, Fahy WA, et al. An epithelial biomarker signature for idiopathic pulmonary fibrosis: an

analysis from the multicentre PROFILE cohort study. *Lancet Respir Med* 2017;5:946–955.

- Molyneaux PL, Cox MJ, Willis-Owen SA, Mallia P, Russell KE, Russell AM, et al. The role of bacteria in the pathogenesis and progression of idiopathic pulmonary fibrosis. Am J Respir Crit Care Med 2014;190: 906–913.
- Troy LK, Grainge C, Corte TJ, Williamson JP, Vallely MP, Cooper WA, et al.; Cryobiopsy versus Open Lung biopsy in the Diagnosis of Interstitial lung disease alliance (COLDICE) Investigators. Diagnostic accuracy of transbronchial lung cryobiopsy for interstitial lung disease diagnosis (COLDICE): a prospective, comparative study. *Lancet Respir Med* 2020;8:171–181.
- Raghu G, Flaherty KR, Lederer DJ, Lynch DA, Colby TV, Myers JL, *et al.* Use of a molecular classifier to identify usual interstitial pneumonia in conventional transbronchial lung biopsy samples: a prospective validation study. *Lancet Respir Med* 2019;7:487–496.
- Lukey PT, Harrison SA, Yang S, Man Y, Holman BF, Rashidnasab A, *et al*. A randomised, placebo-controlled study of omipalisib (PI3K/mTOR) in idiopathic pulmonary fibrosis. *Eur Respir J* 2019;53:1801992.
- Allden SJ, Ogger PP, Ghai P, McErlean P, Hewitt R, Toshner R, et al. The transferrin receptor CD71 delineates functionally distinct airway macrophage subsets during idiopathic pulmonary fibrosis. Am J Respir Crit Care Med 2019;200:209–219.
- Molyneaux PL, Willis-Owen SAG, Cox MJ, James P, Cowman S, Loebinger M, et al. Host-microbial interactions in idiopathic pulmonary fibrosis. Am J Respir Crit Care Med 2017;195:1640–1650.

Copyright © 2021 by the American Thoracic Society

Check for updates

∂ Is IL-6 the Right Target in COVID-19 Severe Pneumonia?

To the Editor:

We read with great interest the article by McElvaney and colleagues, "Characterization of the Inflammatory Response to Severe COVID-19 Illness," published in the *Journal* (1). Systemic inflammation that characterizes acute respiratory distress syndrome (ARDS) in coronavirus disease (COVID-19) is associated with a high mortality rate (2). The term "cytokine storm" has emerged to explain the immunopathogenesis of most severe forms of COVID-19, because the release of many inflammatory cytokines (e.g., IL-6) was correlated with the disease severity (3). However, immune pathogenesis is still unsettled and comparisons with community-acquired pneumonia (CAP) from other origins are scarce. Interestingly, McElvaney and colleagues showed that COVID-19 cytokinemia is distinct, with circulating levels of IL-6 significantly higher in patients with COVID-19 than in those with non–COVID-19 severe CAP (1). These results are challenged by an editorial by Sinha and colleagues showing that plasma concentrations of IL-6 in patients with severe COVID-19 were lower than those observed in patients with ARDS from other origins (4). However, such comparisons are only possible if the respiratory severity is comparable between both groups.

We performed a prospective study that is part of the LYMPHONIE project (clinicaltrials.gov NCT03505281). We included non-immune-compromised patients with severe pneumonia (at least two criteria of the quick Sequential Organ Failure Assessment score and/or need for mechanical ventilation or vasopressors). Patients with COVID-19 all tested positive for severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) by RT-PCR. Plasma was collected within 48 hours of hospital admission and the concentration of IL-6 quantified using a Luminex assay (R&D Systems). Oral consent was obtained from the patient or their legal representatives. Approval was obtained from the ethics committee (Comité de Protection des Personnes Sud-MEDITERRANEE V) (2017-A03404-49).

Thirty-six patients without COVID-19 were enrolled between November 2018 and February 2020 (before the COVID-19 pandemic started in Burgundy, France) and 27 patients with COVID-19 in March and April 2020. Median age (interquartile range) was 67.5 (63–76.5) and 64 (57–71) in the non–COVID-19 and COVID-19 groups, respectively (P = 0.0559). ICU admission was needed for 32 (89%) and 27 (100%) patients of the non–COVID-19 and COVID-19 groups, respectively (P = 0.07). Pa_{O2}:FI_{O2} ratios were not different between groups (115.9 [74.6–157.9] vs. 145 [86–175.7] mm Hg, respectively; P = 0.3073). The IL-6 plasma concentrations were higher in the patients without COVID-19 than in those with COVID-19 (460.4 [138.2–4434.7] vs. 121 [75.7–236.6] pg/ml; P = 0.0003) (Figure 1).

Here, we showed that despite similar respiratory severity, plasma concentrations of IL-6 were dramatically lower in severe forms of COVID-19 than in CAP from other origins. Thus, although immune modulation is a promising therapeutic avenue that is likely to improve outcomes for COVID-19, the most relevant target and strategy remain to be found.



Figure 1. Box plot showing the plasma concentration of IL-6 within 48 hours of hospitalization in 36 patients with non–COVID-19 and 27 patients with COVID-19 severe pneumonia (LYMPHONIE study, 2018–2020). COVID-19 = coronavirus disease.

³This article is open access and distributed under the terms of the Creative Commons Attribution Non-Commercial No Derivatives License 4.0 (http://creativecommons.org/licenses/by-nc-nd/4.0/). For commercial usage and reprints, please contact Diane Gern (dgern@thoracic.org).

Supported by grants from the AOIc2020 (to M.B.), from the INSERM UMR 1231, ANR-11 LABX-0021-01, Labex Lipstic, the FEDER, the Regional Council of Bourgogne Franche-Comte, and by crowdfunding (https://thellie.org/covid-19).

Author Contributions: Concept and design: M.B., C.B., and L.P. Acquisition, analysis, or interpretation of data: M.B., A.B., C.B., and L.P. Drafting of the manuscript: M.B. Critical revision: C.B. and L.P. Supervision: C.B. and L.P.

Originally Published in Press as DOI: 10.1164/rccm.202007-2924LE on September 21, 2020