

Review article:

IMMUNOLOGY OF IL-12: AN UPDATE ON FUNCTIONAL ACTIVITIES AND IMPLICATIONS FOR DISEASE

Karen A.-M. Ullrich^{#1}, Lisa Lou Schulze^{#1}, Eva-Maria Paap¹, Tanja M. Müller¹,
Markus F. Neurath¹, Sebastian Zundler^{1,*}

Shared first authorship

¹ Department of Medicine and
Deutsches Zentrum Immuntherapie, University Hospital Erlangen, Friedrich-Alexander-
University Erlangen-Nuremberg, Germany

* **Corresponding author:** Dr. med. Sebastian Zundler, Department of Medicine and
University Hospital Erlangen, Ulmenweg 18, D-91054 Erlangen, Germany, Phone:
09131/85-35000, E-mail: sebastian.zundler@uk-erlangen.de

<http://dx.doi.org/10.17179/excli2020-3104>

This is an Open Access article distributed under the terms of the Creative Commons Attribution License
(<http://creativecommons.org/licenses/by/4.0/>).

ABSTRACT

As its first identified member, Interleukin-12 (IL-12) named a whole family of cytokines. In response to pathogens, the heterodimeric protein, consisting of the two subunits p35 and p40, is secreted by phagocytic cells. Binding of IL-12 to the IL-12 receptor (IL-12R) on T and natural killer (NK) cells leads to signaling via signal transducer and activator of transcription 4 (STAT4) and subsequent interferon *gamma* (IFN- γ) production and secretion. Signaling downstream of IFN- γ includes activation of T-box transcription factor TBX21 (Tbet) and induces pro-inflammatory functions of T helper 1 (T_H1) cells, thereby linking innate and adaptive immune responses. Initial views on the role of IL-12 and clinical efforts to translate them into therapeutic approaches had to be re-interpreted following the discovery of other members of the IL-12 family, such as IL-23, sharing a subunit with IL-12. However, the importance of IL-12 with regard to immune processes in the context of infection and (auto-) inflammation is still beyond doubt. In this review, we will provide an update on functional activities of IL-12 and their implications for disease. We will begin with a summary on structure and function of the cytokine itself as well as its receptor and outline the signal transduction and the transcriptional regulation of IL-12 secretion. In the second part of the review, we will depict the involvement of IL-12 in immune-mediated diseases and relevant experimental disease models, while also providing an outlook on potential translational approaches.

Keywords: IL-12, T_H1 cells, STAT4, immunology, cytokines, ustekinumab

INTRODUCTION

Dozens of cytokines have meanwhile been described and the knowledge on the stimuli that induce their release, the signaling they trigger and the cellular responses they

cause has substantially increased our understanding of immune processes in the organism. Cytokines are regarded as crucial paracrine or autocrine mediators of inter-cellular communication regulating functions like proliferation, differentiation and maturation (Neurath, 2014).

One important group of cytokines are interleukins, in which the twelfth to be described in 1989 was Interleukin-12 (IL-12). In more than three decades, numerous papers have reported effects and functions of IL-12. Here, we will briefly point out the biochemistry of IL-12 before reviewing its involvement in immune-mediated pathologies.

INTERLEUKIN-12

Two groups independently described IL-12: Kobayashi et al. reported the identification of the natural killer cell stimulating factor (NKSF) in 1989 (Kobayashi et al., 1989) and Stern et al. discovered the cytotoxic lymphocyte maturation factor (CLMF) in 1990 (Stern et al., 1990). Soon thereafter, CLMF and NKSF were found to be identical (Gubler et al., 1991) and the name IL-12 was proposed.

IL-12 consists of two subunits, which are connected by disulphide-bonds (Kobayashi et al., 1989; Stern et al., 1990). The smaller p35 monomer (35 kDa α -chain) is encoded on chromosome 3, while the gene for the larger p40 monomer (40 kDa β -chain) is located on chromosome 5 (Sieburth et al., 1992). Co-expression results in the formation of the biologically active p70 heterodimer (Gubler et al., 1991).

Within the ‘IL-12 family’ of cytokines, monomers combine with different partners to create various cytokines. While p35 may also pair with Epstein-Barr virus induced gene 3 (EBI3) to yield IL-35 (Niedbala et al., 2007), the p40 subunit in combination with the p19 monomer leads to the formation of IL-23 (Lupardus and Garcia, 2008). The fourth member of the family is IL-27, which is composed of EBI3 and the p28 subunit (Vignali and Kuchroo, 2012) (Figure 1).

Due to the lower expression of the IL-12 α -chain compared to the β -chain, only free β -chains, p40 homodimers or the heterodimer are secreted (D'Andrea et al., 1992). Structurally, the p40 subunit shares some features with the IL-6 receptor, whereas the p35 subunit is similar to the granulocyte colony-stimulating factor (G-CSF) and IL-6 (Gubler et al., 1991). It has been demonstrated in mice that p40 homodimers regulate the activity of IL-12 by counteracting IL-12-induced signaling via competition with IL-12p70 for binding to the receptor (Gately et al., 1996). Further functions of p40 homodimers have been described, e.g. roles in the migration of dendritic cells (DCs), allograft rejection or chemotactic activity with regard to macrophages (Cooper and Khader, 2007; Ha et al., 1999).

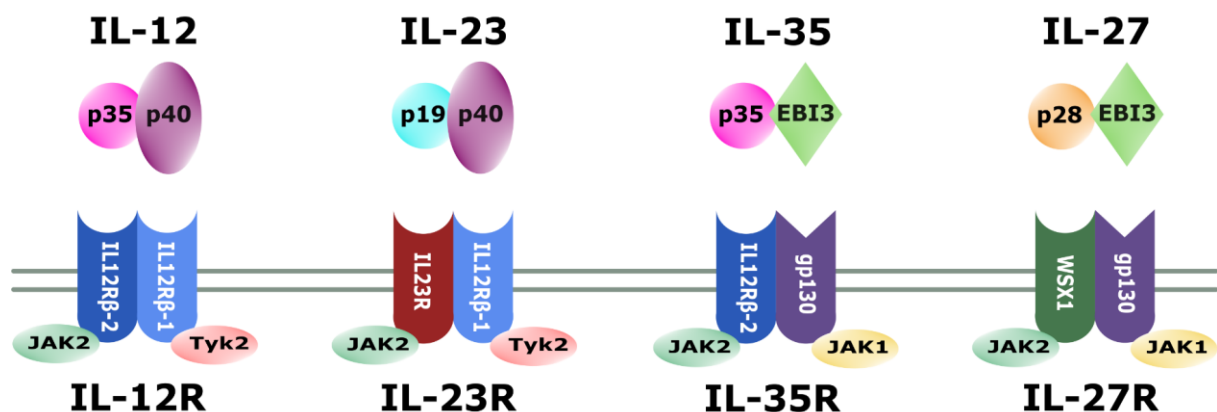


Figure 1: The IL-12 family. Schematic representation of IL-12 family members, their associated receptors and corresponding subunits

THE INTERLEUKIN-12 RECEPTOR

Reflecting the heterodimeric structure of the cytokines of the IL-12 family, the corresponding receptors also consist of two subunits. IL12-receptor $\beta 1$ (IL-12R $\beta 1$) is encoded on chromosome 19 and has a molecular weight of 100 kDa. It is a transmembrane protein with the extracellular domain consisting of 516 amino acids that is responsible for the interaction with IL-12p40 (Chua et al., 1994; Presky et al., 1996). Consistently, it is also part of the receptor for IL-23, where it pairs with IL-23R (Parham et al., 2002). The gene for IL-12R $\beta 2$ is located on chromosome 1 and is translated to a 130 kDa transmembrane protein, with 595 amino acids forming the extracellular domain. Signal transduction into the cell derives from IL-12R $\beta 2$, which interacts with IL-12p35 (Presky et al., 1996; Zou et al., 1997) and is, thus, in combination with glycoprotein 130 (gp130), also part of the IL-35 receptor (Collison et al., 2012). The IL-27 receptor as the fourth receptor of the family is composed of gp130 together with the interleukin 27 receptor subunit alpha (WSX1) (Pflanz et al., 2004) (Figure 1).

Since NK cells and T cells are the main targets of IL-12, the expression of IL-12R is predominantly confined to these cell types (Desai et al., 1992). In particular, antigen contact of naïve T cells induces upregulation of IL-12R $\beta 2$, which is subsequently maintained by interferon *gamma* (IFN- γ) signaling, but may be counteracted by IL-4 (Szabo et al., 1997). This hints at a vital role for the commitment of T cells to different effector T (T_{eff}) cell lineages such as cells with a T helper type 1 (T_{H1}), but not a T_{H2} phenotype and, consistently, only the former cells express IL-12R $\beta 2$.

REGULATION OF IL-12 SECRETION

IL-12 is primarily produced by professional antigen-presenting cells (APCs) such as B cells and DCs as well as phagocytes including monocytes, macrophages and granulocytes (Hsieh et al., 1993; Heufler et al., 1996; Macatonia et al., 1993).

While the production of IL-12p35 is predominantly regulated at the translational level, transcriptional regulation is responsible for the amount of IL-12p40 expressed. The initial signal triggering IL-12 expression is the exposure of the above mentioned cells to bacteria, viruses, fungi or parasites. Pathogen associated molecular patterns (PAMPs) such as lipopolysaccharide (LPS) or CpG DNA expressed or contained in such commensals or pathogens are recognized by pattern recognition receptors (PRRs) of the toll like receptor (TLR) family. This leads to the activation of several transcription factors regulating IL-12 production, most importantly NF- κ B and interferon regulatory elements (IRFs) (Goriely et al., 2008).

Due to the different chromosomal locations (Liu et al., 2003) and as mentioned above, there are important differences in the regulation of p40 and p35 production. The synthesis of the p40 chain greatly exceeds the production of the p35 chain, suggesting that the synthesis of the p35 chain is the rate-limiting step of IL-12 secretion (Snijders et al., 1996). Moreover, most of the TLRs are linked to the expression of IL-12p40, while expression of p35 is induced by only a limited subset of these receptors, including TLR3, 4 and 8. In addition to direct regulation of IL-12 production, activation of TLRs also leads to the secretion of IFN- β and IFN- γ , whose signaling, in turn, induces activation of IRF-1, IRF-7 and IRF-8 (Goriely et al., 2008; Najjar et al., 2017; Gautier et al., 2005). All three IRFs induce p35 and IRF-7 and IRF-8 also induce p40 (Ma et al., 2015; Zhao et al., 2017).

The IL12A gene is transcribed throughout, however, the mRNA contains an inhibitory ATG codon blocking its translation. Upon TLR signaling as described above, transcription is initiated from different genomic positions. Since ATG codons are missing in the resulting mRNA, translation into IL-12p35 proceeds (Wang et al., 2000).

The IL12B gene is subject to transcriptional regulation by a number of transcription factors, most importantly NF- κ B and Ets. Moreover, Spi-1, AP-1, IRF-1, erythroid

Krüppel-like factor 1 (KLF-1), nuclear factor in activated T cells (NFAT) and interferon consensus sequence-binding protein (ICSBP) have been described to be involved (Ma et al., 2015; Zhao et al., 2017). Furthermore, some groups showed an inhibitory effect of Nuclear Receptor Subfamily 4 Group A Member 1 (NR4A1) on the expression of IL-12p40 (Murphy and Crean, 2015; Ipseiz et al., 2014).

In addition, IL-12 expression is regulated via interaction of APCs with T-cells through CD40 and its ligand CD40L. CD40 signaling through H-Ras and K-Ras enhances p38 mitogen-activated protein kinase (MAPK)-mediated pro-inflammatory IL-12 production (Snijders et al., 1996; Nair et al., 2020).

An important positive feedback loop increasing IL-12 secretion is so-called IFN- γ priming (Liu et al., 2003; Ma et al., 2015). IFN- γ release downstream of IL-12 further boosts IL-12 production via induction of IL-12p35 by IRF-1 and of p40 by ICSBP (Wang et al., 2000; Grumont et al., 2001).

Taken together, the high degree of regulatory mechanisms involved in IL-12 secretion illustrates the complexity of the processes, in which this cytokine is involved and underscores that dysregulation might be a switchpoint for disease development.

IL-12 SIGNAL TRANSDUCTION

Binding of the two IL-12 subunits to the two chains of the IL-12 receptor, IL-12R β 1 and IL-12R β 2, activates the Janus kinase (JAK)-signal transducer and activator of transcription (STAT) pathway of signal transduction. Specifically, IL-12R β -1 subsequently recruits the JAK family member tyrosine kinase 2 (TYK2), whereas IL-12R β 2 associates with JAK2, resulting in phosphorylation of JAK2 (Bacon et al., 1995; Zou et al., 1997). This activates the kinase activity of JAK2, which now, *vice versa*, phosphorylates a tyrosine residue of the associated receptor subunit. STAT molecules contain SRC homology domains (SH2), which, in a next step, bind to phospho-IL-12R β 2 exposing the STATs to

JAK and leading to their phosphorylation. Association of these activated transcription factors to homo- or heterodimers enables subsequent nuclear translocation. By binding to specific DNA sequences, they promote or repress gene transcription (Naeger et al., 1999; Zhong et al., 1994; Xu et al., 1996; Jacobson et al., 1995; Lamb et al., 1996) (Figure 2). STAT4 is the most important downstream target of IL-12, while effects on STAT1, STAT3 and STAT5 molecules play minor roles (Trinchieri, 2003).

Moreover, IL-12R signaling activates mitogen-activated protein kinase kinase 3/6 (MKK) and p38 MAPK, which support the secretion of IFN- γ in activated T cells and T_H1 cells. Importantly, this pathway is mediated by a STAT4-independent mechanism and correlates with increased STAT2 (Zhang and Kaplan, 2000).

CELLULAR FUNCTIONS OF IL-12

As already mentioned, a main effect of IL-12 is the induction of IFN- γ production, by which the cytokine is importantly implicated in adaptive as well as innate immune processes (Figure 3) (Trinchieri, 2003; Lyakh et al., 2008).

While having no proliferative effect on resting peripheral T cells or NK cells, IL-12 directly induces proliferation of these cells in case of pre-activation. Moreover, by inducing the transcription of genes of cytotoxic granule-associated molecules, such as perforin and granzymes, and by upregulation of the expression of adhesion molecules, IL-12 enhances the generation and cytotoxic activity of cytotoxic T lymphocytes (CTLs), lymphokine-activated killer (LAK) cells and NK cells, which also secrete IFN- γ (Trinchieri, 1998, 2003; Tait Wojno et al., 2019). Even low concentrations of IL-12 promote IFN- γ production in a highly efficient way. Furthermore, IL-12 is also very important as IFN- γ -inducer in synergy with other activating stimuli. E.g., for T cells and NK cells, IL-12 acts synergistically with IL-2 to rapidly upregulate

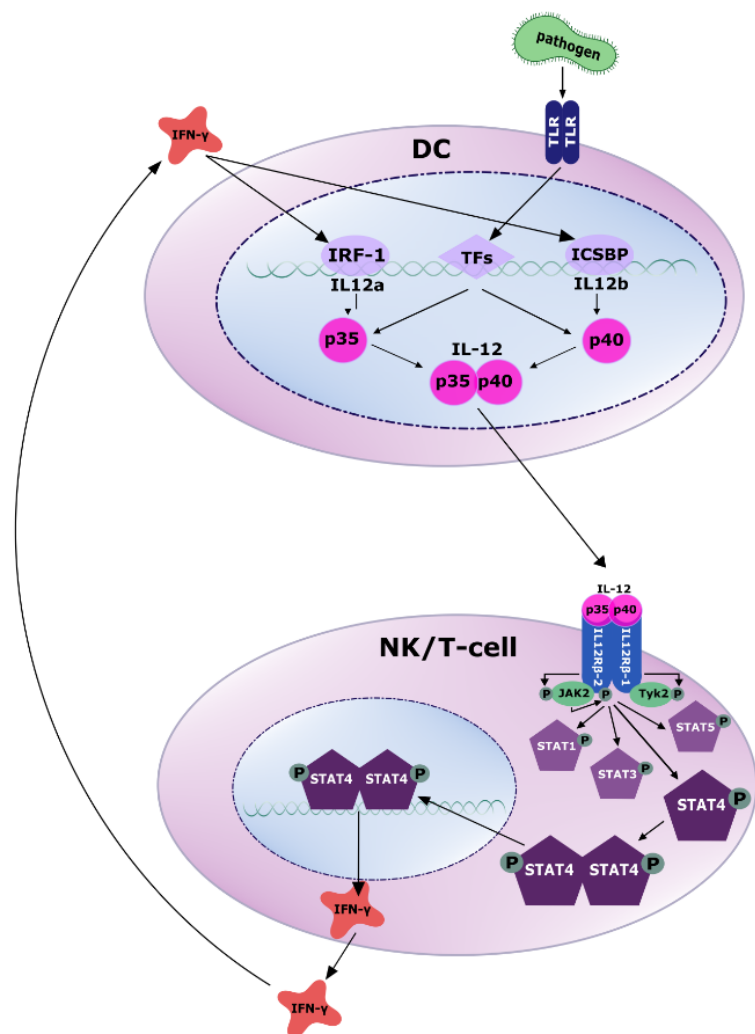


Figure 2: Secretion and signaling of IL-12. Antigen-presenting cells (APCs) like dendritic cells sense PAMPs (pathogen-associated molecular patterns) through toll like receptors (TLRs). Subsequently, several transcription factors are activated to induce the transcription of IL-12p35 and IL-12p40 (for more details confer text). The secreted IL-12 heterodimer binds to its receptor on NK and T cells, recruits the tyrosine kinases JAK2 and TYK2 and activates JAK2 by tyrosine phosphorylation. Activated JAK2 phosphorylates the IL12R β 2 subunit, which in turn activates STAT4 via phosphorylation. Subsequently, phosphorylated STAT4 homo- or heterodimerizes, enabling translocation to the nucleus, where it regulates gene transcription by binding to target DNA. A main target gene is IFN- γ , which in turn induces transcriptional activation of IL-12 production via IRF-1 and ICSBP.

IFN- γ (Trinchieri, 2003; Chan et al., 1992). While the cytokine IL-18 alone is ineffective in regulating IFN- γ , in synergy with IL-12, it induces gene transcription via the transcription factors STAT4 and AP-1. This is particularly important for the induction of IFN- γ by cell types such as macrophages, DCs or B cells that are no conventional sources of this cytokine (Trinchieri, 2003; Walker et al., 1999; Barbulescu et al., 1998). Interestingly, STAT4 is required for the production of IFN- γ downstream of IL-12 in both CD4⁺ and CD8⁺ T cells, but only CD4⁺ T cells require IL-12 and STAT4 for the production of IFN- γ following antigen recognition and T cell receptor (TCR) signaling suggesting alternative regulatory pathways in different lymphocyte subsets (Trinchieri, 2003; Carter and Murphy,

1999). It was also shown that IL-12 pre-treatment of CD4⁺ and CD8⁺ T cells enhances TCR-induced IFN- γ , tumor necrosis factor *alpha* (TNF- α), IL-13, IL-4 and IL-10 production and intensifies oxidative metabolism (Vacaflores et al., 2016, 2017).

Due to and related to its predominant effect on IFN- γ transcription, IL-12 is a potent inducer of T_H1 cell development. However, it is not sufficient to guide this process, since previous signaling downstream of IL-27 released by DCs is required in naïve T cells to make the cells “IL-12-sensitive” by expression of IL12R β 2 (Pot et al., 2010). After release of IL-12 and induction of IFN- γ , IFN- γ signaling activates STAT1, which together with IFN- α , IFN- β and IL-12 induces the T-box transcription factor 21 (TBX21/Tbet), the

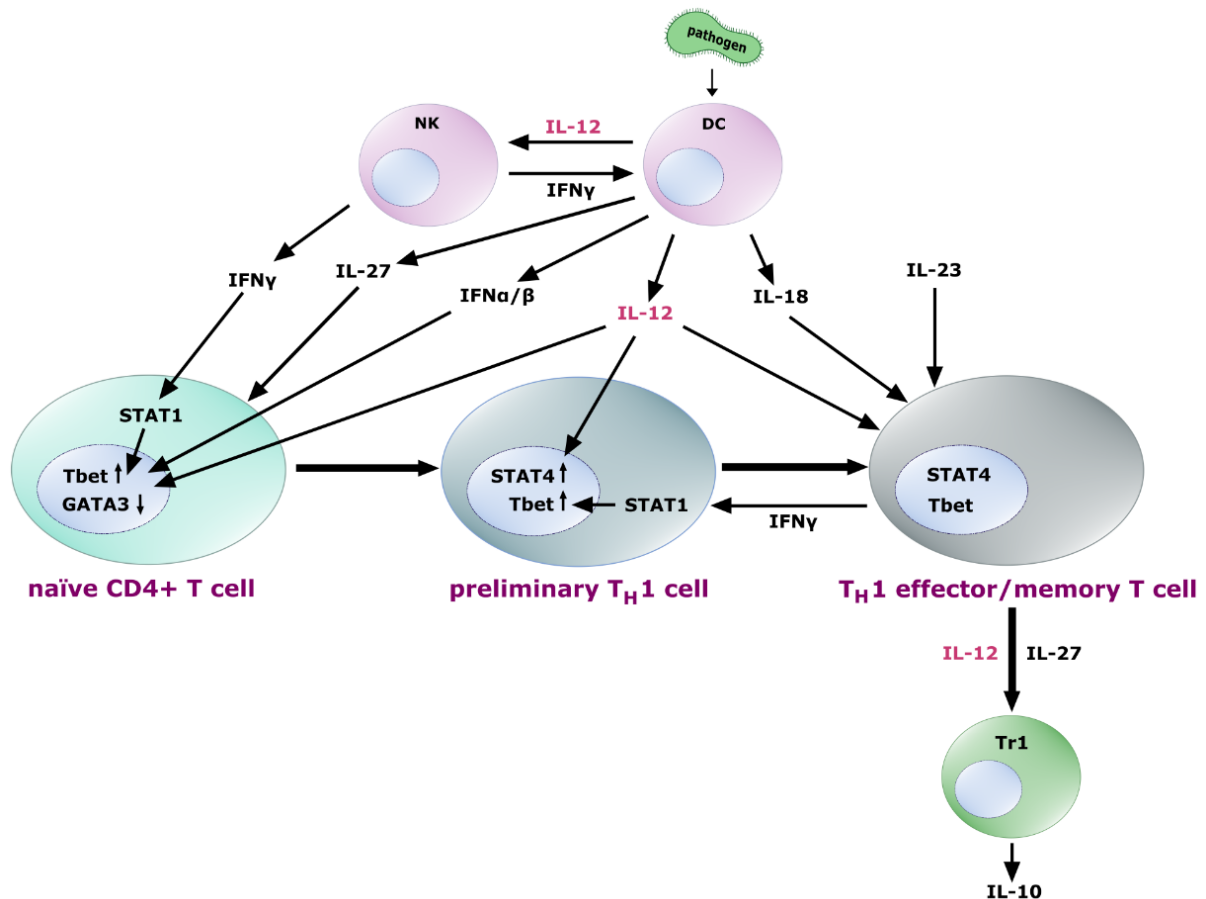


Figure 3: Role of IL-12 in TH1 differentiation. Naïve T cells exposed to IL-27 express the IL-12 receptor heterodimer, sensitizing the cells for the influence of IL-12, which – together with IFN-γ from NK cells and feedback loops, IFN-α and IFN-β induces upregulation of the transcription factor Tbet and down-regulation of GATA3 leading to a preliminary TH1 commitment. Further exposure to IL-12 leads to the upregulation of STAT4 in these early TH1 cells, followed by their differentiation into TH1 effector and memory T cells. IL-18 and IL-23 contribute to the fixation, amplification and maintenance of the TH1 cell effector functions. Differentiated TH1 cells produce IFN-γ to enhance Tbet expression via STAT1, thus closing a positive feedback loop. Further, they are able to promote IL-10 secreting Tr1 cells, a process probably mediated by IL-12 and IL-27 signaling.

key transcription factor for T cell commitment to preliminary TH1 cells (Trinchieri, 2003). For this early phase of TH1 polarization, some exceptions seem to exist, in which IL-12 might not be an absolute requirement and coordinated action of other pathways might compensate, if IL-12 is missing. IL-12, however, is essential for the subsequent maturation phase (Noble et al., 2001) and, together with IL-18 and IL-23 fixes, amplifies and maintains a TH1 phenotype during clonal expansion to effector and memory TH1 cells (Trinchieri, 2003). Furthermore, it has been shown that the expression of genes promoting

the induction of IFN-γ- and IL-21-secreting TH1-biased T follicular helper (TFH1)-like cells are also dependent on IL-12 signals, especially on the expression of Bcl-6 and inducible T-cell costimulator (ICOS) (Powell et al., 2019).

Additionally, it has been shown that IL-12 also primed CD4+ and CD8+ T cells to produce IL-10, when present early during clonal expansion (Gerosa et al., 1996). This might result in the development of IL-10 secreting Type 1 regulatory (Tr1) cells in response to IL-12 and IL-27 (Tait Wojno et al., 2019; Pot

et al., 2010; Wang et al., 2011), which is consistent with the observation of IL-12-dependent Tr1 cell development in visceral leishmaniasis patients (Montes de Oca et al., 2016). T_H2 differentiation, to the contrary, is counteracted by IL-12, since GATA binding protein 3 (GATA3), which is indispensable for T_H2 polarization, is repressed in CD4⁺ and CD8⁺ T cell populations upon treatment with IL-12 or *in vivo* expansion in the presence of IL-12-producing DCs (Billerbeck et al., 2014).

Taken together, IL-12 is a potent cytokine to regulate the immune response in different ways. Thus, its implication in immune-mediated diseases is obvious and will be addressed in the following part.

RELEVANCE OF IL-12 IN DIFFERENT DISEASES AND DISEASE MODELS

Consistent with its central role in orchestrating immune responses, various studies in animal models and humans confirmed that IL-12 contributes to the pathogenesis of several immune-mediated inflammatory diseases. In the following paragraphs, we outline the current knowledge on the impact of IL-12 in the context of inflammatory bowel diseases, psoriasis, diabetes mellitus, multiple sclerosis, rheumatoid arthritis, cancer, lupus erythematosus, primary biliary cholangitis and Sjögren's syndrome.

Inflammatory Bowel Diseases

The inflammatory bowel diseases (IBDs) Crohn's disease (CD) and ulcerative colitis (UC) have a multifactorial pathogenesis and are marked by a misdirected and dysregulated immune response, which arises due to factors such as genetic predisposition, intestinal dysbiosis and a disruption of the intestinal epithelial barrier. Experimental colitis can be observed in genetic knockout models leading to spontaneous development of colitis as well as following the administration of chemicals like oxazolone, trinitrobenzene sulfonic acid (TNBS) or dextran sodium sulfate (DSS). Further, a so called T cell transfer colitis

model can be induced by transfer of colitogenic T cells to immunodeficient mice (Neurath, 2014).

T_H1-like colonic lymphocytes have been identified as predominant in TNBS-induced colitis and anti-IL-12p40 administration was able to eliminate inflammation (Hurtubise et al., 2019) as well as to re-establish tolerance towards the intestinal microbiota (Duchmann et al., 1996). Fuss et al. suggested that Fas pathway activation inducing apoptosis of T_H1 cells might be a leading mechanism (Fuss et al., 1999). Corroborating these results on transcription factor level, mice adoptively receiving CD4⁺ T cells from STAT4-transgenic mice developed pronounced colitis and STAT4-transgenic mice themselves showed higher STAT4 expression in their lamina propria lymphocytes and were more susceptible to colitis (Wirtz et al., 1999). Consistently, spontaneous colitis observed in IL-10^{-/-} mice was rescued by additional knockout of IL-12R and IL-23R (Hurtubise et al., 2019) as well as by administration of a neutralizing anti-IL-12 antibody (Davidson et al., 1998). Blocking IL-12 was also able to avert T cell transfer-induced colitis in immunodeficient mice which received CD4⁺ T cells from IL-10^{-/-} donors, however this was not the case for the blockade of IFN- γ (Davidson et al., 1998) indicating that IFN- γ -independent pathways are involved. In conclusion, T_H1-dependent colitis models seem to be clearly dependent on IL-12, but IFN- γ as a major cytokine of T_H1 cells seemed to be non-essential. An explanation for this redundancy might be that many of these studies focused on IL12p40 at a time, when IL-23 and its effects on IL-17-producing T_H17 cells were not yet described (Moschen et al., 2019).

Following the identification of IL-23, intensive research tried to dissect the mechanisms exerted by IL-12 and IL-23 on their own, although some aspects cannot finally be separated. In murine models, IL-23 was, like IL-12, able to induce STAT4 activation and overexpression was followed by colitis. Furthermore, p40 and IL-23 induction occurred subsequent to bacteria intake in DCs in the

terminal ileum (Becker et al., 2003). In studies focusing on the differences between IL-12 and IL-23, depletion of p19, but not p35 in IL-10^{-/-} mice led to protection from colitis development (Yen et al., 2006). Consistently, while treatment with anti-p40 antibodies protected from systemic inflammatory disease and experimental colitis, treatment with anti-p19 alleviated intestinal but did not affect systemic pathology, suggesting a contribution of IL-23 to local inflammation and an IL-12-regulated systemic inflammation (Uhlir et al., 2006). In TNBS colitis, to the contrary, p19 knockout was not protective due to upregulation of IL-12 suggesting that both cytokines might interact in the pathogenesis of colitis (Becker et al., 2006). A recent study building on a genetically induced intestinal barrier impairment phenotype added further complexity by showing that inflammation in this model was mediated by IL-12 and IL-23 in a temporally distinct, biphasic manner. While inflammation in the early stages was driven by IL-12, the inflammatory response shifted towards IL-23 in older mice (Eftychi et al., 2019). This is consistent with the assumption of some experts that the early stages of CD pathogenesis are driven by T_{H1} activation mediated by IL-12 and the following IFN- γ signaling cascade, whereas the continuation of disease is more likely driven by IL-23 (Becker et al., 2005). What is further complicating a clear separation of IL-12- and IL-23-induced functions is T cell plasticity. It has been shown that IL-12 exposure shifted the phenotype of T_{H17} effector memory cells from the mesenteric lymph nodes of CD as well as UC patients towards a T_{H1}-like profile suggesting that T_{H17} plasticity is taking place at inductive sites before T cell homing to gut tissues (Bsat et al., 2019).

Taken together, undoubtedly, IL-12 as well as IL-23 are implicated in the pathogenesis of IBD. Hence, they are promising targets for IBD therapy, which has further been supported by observations in IBD patients. IL-12 transcription is increased in both subtypes of IBD (Nemeth et al., 2017). Additionally, CD is marked by high levels of IFN- γ produced by lamina propria lymphocytes (Fuss et al.,

1996) substantiating the view that CD seems to be a partly T_{H1}-driven disease. Moreover, it was shown that the expression patterns of the IL-12A and IL-12B genes differed between flare-ups and remission phases and might therefore be suitable biomarkers of different disease phases (Norouzinia et al., 2018).

All these observations led to efforts to translate anti-IL-12/IL-23 strategies into clinical practice. Ustekinumab is a neutralizing p40 antibody. While early clinical studies suggested effects in CD patients with previous anti-TNF treatment, phase III trials completed in recent years have demonstrated broad efficacy and safety in inducing and maintaining remission in CD and recently also in UC (Sands et al., 2019; Feagan et al., 2016; Sandborn et al., 2012; Adedokun et al., 2018; Hanauer et al., 2020). Moreover, a number of anti-p19 antibodies blocking IL-23 (e.g., risankizumab, guselkumab) are currently in phase III trials following promising data in phase II (Sandborn et al., 2020; Sands et al., 2017). Once finished, they will further help to understand the specific contribution of IL-12 and IL-23 to disease pathogenesis.

Multiple sclerosis

Multiple sclerosis (MS) is considered as a T_{H1}-, T_{H17}-, B- and innate immune cell-mediated disorder of the central nervous system (CNS), in which immune cell infiltration to the CNS leads to inflammatory lesions, axonal demyelination and enhanced production of pro-inflammatory cytokines (von Essen et al., 2019; Segal et al., 1998). Early studies focussing on p40 postulated IL-12 as a critical pro-inflammatory cytokine for MS (Adorini, 1999; Karp et al., 2000). Consistently, during the acute paralytic phase of experimental autoimmune encephalomyelitis (EAE), a common animal model for MS, mice showed increased expression of IL-12p40 mRNA in brain and spinal cord as well as in spleen, lymph node and liver. Furthermore, *in vivo* administration of recombinant IL-12 led to higher IFN- γ production and inflammation, which was decreased by treatment with anti-

IL-12 antibodies (Bright et al., 1998; Vandenbroeck et al., 2004). However, later studies showed that p35^{-/-} animals were not protected from EAE, whereas p40^{-/-} mice were resistant to disease development (Becher et al., 2002; Gran et al., 2002). Thus, these effects could not be attributed to IL-12p70, but later be explained by IL-23 and protection of p19^{-/-} mice in the EAE model (Gran et al., 2004).

However, also here, the picture is not completely clear, since it was shown with an adoptive transfer strategy that IL-12-modulated T_H1 cells may induce EAE via an IL-23-independent pathway, while IL-23-modulated T_H17 cells may induce EAE through an IL-12-independent way. This indicates, that clinically similar forms of EAE may be mediated by distinct autoreactive T cell subsets pointing at synergistic or alternative functions of these parallel inflammatory pathways (Grifka-Walk et al., 2015; Kroenke et al., 2008).

Recently, a huge amount of single-nucleotide polymorphisms (SNPs) with suggestive evidence of association with MS were identified by meta-analyses. These include, among others, loci which are related to IL-12 and the IL-12 family, e.g. IL-12A (rs4680534), or their downstream signaling, e.g. JAK1 (rs72922276), STAT4 (rs6738544) and TYK2 (rs34536443) (von Essen et al., 2019; Ban et al., 2009; IMSGC, 2010). Moreover, significantly elevated levels of IL-12 were found in sera of MS patients and in cultured peripheral blood mononuclear cells (PBMCs), which were even higher in chronic progressive compared to relapsing-remitting MS (Musabak et al., 2011; Balashov et al., 1997). Moreover, an indirect effect of IL-12 on the pathogenesis of MS was postulated: IL-12p70 and p35 subunit, but not p40, led to increased expression of IL-7 in the CNS, which is associated with MS and EAE (Jana et al., 2014).

Also, considerably enhanced expression of adhesion molecules involved in leukocyte recruitment to the CNS like C-C-motif-chem-

okine-receptor 5 (CCR5) and P-selectin glycoprotein ligand were induced by IL-12 (Bagaeva et al., 2003; Rabinowich et al., 1993; Deshpande et al., 2006). The pathogenetic relevance of such immune cell homing has been demonstrated by the therapeutic anti-CD49d antibody natalizumab (Tysabri®), a recombinant humanized IgG4 monoclonal antibody, which blocks adhesion and homing via $\alpha 4\beta 1$ and $\alpha 4\beta 7$ integrin and is in successful clinical use for the treatment of MS (Nelson et al., 2018; Benkert et al., 2012).

Surprisingly, clinical trials with the neutralizing anti-IL-12/23 p40 subunit antibody ustekinumab could not show the expected therapeutic benefits in MS treatment in phase II studies (Segal et al., 2008). Moreover, another monoclonal anti-IL-12/23 antibody, ABT-874, demonstrated efficacy in reducing inflammatory lesions and prevention relapses, but the effect size was low compared to other agents and, hence, it was not further developed (Vollmer et al., 2011). Although anti-IL23 monotherapy ameliorated EAE (Chen et al., 2006), anti-p19 antibodies have so far not been tested in MS, probably due to the disappointing results observed with anti-p40 antibodies.

Cancer

Due to its ability to activate cytotoxic cells both from the innate (NK cells) and adaptive (cytotoxic T lymphocytes) immune system, IL-12 appears to be a promising mediator of anti-tumoral immunotherapy, which has been the subject of many reviews (Tait Wojno et al., 2019; Golab and Zagozdzon, 1999; Lasek et al., 2014; Tugues et al., 2015; Yan et al., 2018). Taken together, the main anti-tumoral mechanisms of IL-12 are thought to be the increase of IFN- γ production, which has anti-proliferative and pro-apoptotic effects (Castro et al., 2018); the activation of proliferation and cytotoxicity of NK cells and CD4⁺ and CD8⁺ T cells; the enhancement of antibody-dependent cellular cytotoxicity (ADCC); the induction of anti-angiogenic cytokines and chemokines such as

IFN- γ ; the remodeling of the peritumoral extracellular matrix resulting in collapse of the tumor stroma; and the modulation of anti-tumoral immune response by influencing the major histocompatibility complex (MHC) class I molecules as well as reprogramming myeloid-derived suppressor cells (Lasek et al., 2014; Campbell et al., 2015). Moreover, it was found that IL-12 inhibits metastasis development in experimental tumor models in a dose-dependent way without showing signs of toxicity (Brunda et al., 1993; Yue et al., 2016). Interestingly, induction of IL-12 seems to mimic the natural anti-tumoral immune response and induces a very specific anti-tumor reaction (Smyth et al., 2000). Consistently, neoplastic immune evasion strategies from IL-12-induced anti-tumor immunity exist, since B cells in various chronic lymphoproliferative disorders may silence the gene for IL-12R β 2 (IL12rb2) by methylation (Airoldi et al., 2004), which was associated with enhanced tumor-cell survival and proliferation *in vivo* (Airoldi et al., 2005).

Additionally, it has been described that the IL-12 p40 monomer is released in higher amounts than p40 homodimer or IL-12p70 in mouse and human cancer cells. Accordingly, its serum level in patients with prostate cancer is higher compared to healthy controls. Since p40 monomer helps cancer cells to escape from cell death via internalization of IL-12R β 1, p40 neutralization stimulated apoptosis of different cancer cells *in vitro* and *in vivo* (Kundu et al., 2017).

Possible links between IL-12 polymorphisms and various cancers have been explored in many epidemiological studies. One of the cancer-risk loci frequently identified was the SNP rs3212227 in IL12B, which shows significant association to overall cancer risk, especially among Asians, and, particularly, to hepatocellular and nasopharyngeal cancer. Additionally, the rs568408 polymorphism increases overall cancer risk among Caucasians and the risk for cervical cancer, while rs2243115 enhances the risk for brain tumors (Zheng et al., 2017).

It has also been postulated that the effect of immunotherapy with monoclonal antibodies against the checkpoint receptor programmed cell death protein 1 (PD1) requires intratumoral DCs producing IL-12. In this context, the anti-PD1 antibody indirectly activates these DCs through IFN- γ released from drug-activated T cells (Garris et al., 2018; Yin et al., 2016).

When using IL-12 as systemic therapy, it shows toxic inflammatory responses and even lethal side effects in some cases (Ansell et al., 2002; Lenzi et al., 2007; Portielje et al., 1999). Several studies could show that this is due to IFN- γ (Car et al., 1995), but also TNF- α induction (Barrios et al., 2014). To circumvent the adverse effects, different working groups established targeted transport strategies to limit the toxicity associated with systemic application. One example is a vector, in which IL-12 expression is under control of a composite promoter-containing binding motif for nuclear factor of activated T cells (NFAT), which was very effective in a murine melanoma model (Zhang et al., 2011). Furthermore, gene electrotransfer, which was also used for anti-tumor *in situ* vaccination with TNF- α and IL-12 plasmid DNA in a murine melanoma model, and focused ultrasound therapy can be helpful in a more targeted treatment (Shirley et al., 2015; Chen et al., 2015; Kamensek et al., 2018). Another approach is the delivery of IL-12 messenger RNA via lipid nanoparticles, which suppressed tumorigenesis in MYC oncogene-driven hepatocellular carcinoma (Lai et al., 2018). Besides, redesigning of the IL-12 molecule with deletion of the N-terminal signal peptide keeps the anti-tumor efficiency, but reduces the toxicity of the cytokine (Wang et al., 2017). Moreover, the efficacy of tumor therapy with oncolytic viruses (OVs), which preferentially replicate in cancer cells and kill them while sparing healthy cells, was enhanced through viral expression of IL-12 (Nguyen et al., 2020). One of these modified OVs, genetically engineered Herpes Simplex Virus-1, is used in a phase I clinical trial for

the treatment of recurrent malignant glioma (NCT02062827).

Additionally, specific tumor receptors seem to be a good target for genetically modified immunotherapeutics. One example is MUC16^{ecto}, which is highly expressed on most epithelial ovarian carcinomas but at low levels on normal tissues. This observation has been translated into a chimeric antigen receptor specific T cell strategy with autologous IL-12 secretion, which is investigated for the treatment of ovarian cancer (phase I trial: NCT02498912) (Koneru et al., 2015).

IL-12 also seems to have great potential as adjuvant for tumor therapy especially in combination with other substances like doxorubicin (DOX), decorin (DOC) or oncolytic adenovirus expressing suicide genes, resulting in better anti-tumor immune response as shown in murine colorectal cancer, prostate cancer or 4T1 orthotopic breast cancer models (Oh et al., 2017; Hu et al., 2014; Ahn et al., 2016; Freytag et al., 2013). An IL-12-DOX-combination now in clinical evaluation is GEN-1 (EGEN-001, phase I/II trial: NCT03393884), a novel immunotherapeutic agent comprising a human IL-12-expressing plasmid encompassed within a synthetic polyethyleneglycol-polyethyleneimine-cholesterol (PPC) DNA delivery system to facilitate plasmid delivery *in vivo* and administered with pegylated liposomal doxorubicin. It showed encouraging clinical benefit and biological activity in recurrent or persistent epithelial ovarian, fallopian tube or primary peritoneal cancers (Thaker et al., 2017).

Psoriasis

Psoriasis is a chronic inflammatory skin disease with a multifactorial etiology including genetic predisposition, environmental triggers and dysfunctions of TNF- α , dendritic cells and T cells (Griffiths and Barker, 2007). Characteristic traits are infiltration of leukocytes into the skin, hyperplastic blood vessels and hyperproliferation of keratinocytes. Psoriasis vulgaris with chronic plaque formation is the most common manifestation of psoriasis (Cai et al., 2012), which has long been

considered to be a TH1-like disease. Consistently, in biopsies from psoriatic lesions high levels of IFN- γ mRNA could be detected and, furthermore, epidermal T cells produced IFN- γ (Schlaak et al., 1994). Also, peripheral blood T cells from psoriasis patients were capable of producing significantly more IFN- γ compared to cells from healthy controls (Austin et al., 1999). Anti-CD3/CD28 activated PBMCs, which were subsequently stimulated with IL-12 showed 233 significantly dysregulated genes after 4 h of stimulation, among these 28 IL-12-responsive genes. IL-12 stimulation was also found to significantly increase IFN- γ gene expression in anti-CD3/CD28-stimulated PBMCs and therefore might be of functional relevance for systemic inflammation in psoriasis (Enerbäck et al., 2018). Furthermore, IL-12 and IL-23 levels were enhanced in lesional psoriatic skin compared to healthy and non-lesional skin (Yawalkar et al., 2009). Further evidence suggests that susceptibility to psoriasis is associated with IL-12, although inter-ethnic differences exist. In an Egyptian cohort, the interaction of the single nucleotide polymorphisms rs610604 (IL-12B) and rs11209026 (IL-23R) showed a significant association with psoriasis. Moreover, the association of IL-12B with psoriasis was highly significant, whereas no association between rs20541 (IL-13) and psoriasis could be observed (Haase et al., 2015). Consistently, a case-control analysis of psoriatic patients and controls from a Polish population revealed an association of IL12B rs3212227 and IL23R rs11209026 minor allele carrier status with reduced odds for psoriasis, therefore having a protective effect (Bojko et al., 2018). Interestingly, it has been postulated that IL-12 together with IL-17, IL-2 and adiponectin plasma levels predicted psoriasis with a 100 % sensitivity and specificity (Cataldi et al., 2019).

In experimental disease models, mice with imiquimod-induced psoriasis-like dermatitis, which were treated with an anti-IL-12/IL-23p40 monoclonal antibody (p40 mAb) showed reduced epidermal thickness and increased transepidermal water loss, as

well as suppression of IL-23p19, IL-17A, IL-22 and keratin 16 gene expression, suggesting that p40 mAb not only improves dermatitis symptoms, but is also effective against skin barrier dysfunction in those mice (Takahashi et al., 2018).

However, later data showed that IL-12 receptor signaling in keratinocytes initiates a protective transcriptional program that limits skin inflammation, therefore the use of anti-p40 monoclonal antibodies and thereby collateral targeting of IL-12 might be counterproductive for psoriasis therapy (Kulig et al., 2016). Consistently, the picture of psoriasis as a T_H1-biased disease has been redrawn in the meantime, since T_H17 cells were predominantly found in the dermis of psoriatic skin lesions (Yawalkar et al., 2009; Lowes et al., 2008), and IL-22 expression was also increased (Nogales et al., 2009), suggesting that besides T_H1 cells, T_H17 and T_H22 cells have a major influence on disease development. Thus, different T cell subsets probably contribute to plaque formation caused by immune dysregulation (Cai et al., 2012).

This matches the translational experiences made with ustekinumab. This anti-p40 antibody targeting IL-12 and IL-23 turned out to be efficient and safe in the treatment of psoriasis (Papp et al., 2008; Jeon et al., 2017). In accordance with the above mentioned data on genetic predisposition, IL-12-associated gene loci also seem to affect therapeutic success, since patients heterozygous (CT) for the IL12B variant (rs3213094) showed a better response to ustekinumab than the homozygous reference group (CC) (van den Reek et al., 2017). Additionally, briankinumab as an additional anti-IL-12/23 antibody has been approved for the treatment of psoriasis (Gordon et al., 2012). Moreover, ustekinumab has also been demonstrated to be effective in psoriatic arthritis often associated with psoriatic skin lesions. The phase III clinical trial PSUMMIT-1 (NCT01009086) resulted in more ustekinumab-treated than placebo-treated patients achieving the primary endpoint of 20 % or greater improvement in

American College of Rheumatology response (ACR20) at week 24 (McInnes et al., 2013).

Further therapeutic developments in the last years have questioned the pathogenetic role of T_H1 cells, as anti-IL-17 antibodies like secukinumab and ixekizumab demonstrated comparably efficacy to anti-IL-12/23 antibodies (Langley et al., 2014). Moreover, a phase II trial revealed clinical activity of the monoclonal anti-IL23p19 antibody risankizumab that was superior to ustekinumab (Papp et al., 2017). Thus, taken together, it seems that - from a clinical standpoint - T_H17 cells seem to be more important in the pathogenesis of psoriasis than IL-12-driven T_H1 cells.

Diabetes mellitus

There are two types of diabetes mellitus, type 1 (T1D) and type 2 diabetes mellitus (T2D). Characteristic for T1D is the autoimmune destruction of pancreatic islet cells. Without intervention, this leads to hyperglycemia as a result of insulin deficiency. T1D is considered to develop as a result of a reduction in immunosuppressive regulatory T cells (T_{regs}), promoting the expansion of autoreactive CD4⁺ and CD8⁺ T cells (Marwaha et al., 2014). On the other hand, insulin resistance, decreased insulin secretion relative to hyperglycemia, pancreatic β -cell dysfunction, disturbed renal glucose transport and incretin effects, induced by genetic and environmental risk factors are causative for T2D (Aghaei Meybodi et al., 2017). Several reports provide evidence for a contribution of IL-12 to the pathogenesis of both types of diabetes mellitus.

A frequently used murine model for T1D are nonobese diabetic (NOD) mice. Adoptive transfer of diabetogenic T_H1 but not T_H2 cells, as well as IL-12 administration to NOD mice actively promoted diabetes (Katz et al., 1995; Trembleau et al., 1995). Nevertheless, the literature shows that the pathogenesis of diabetes mellitus is more complex and an interplay of IL-12 with other molecules is involved and still not completely understood. For instance, IFN- γ production upon IL-12 stimulation is

not sufficient to drive disease development, since IFN- γ knockdown induces alternative IL-12-mediated pathways. Surprisingly, IFN- γ also seems to prevent the infiltration of pancreatic β cells and the ability of APCs to activate T cells in T1D, suggesting a rather protective function of IFN- γ (Trembleau et al., 2003). To the contrary, it has been shown that β -catenin accumulates in DCs of NOD mice due to hyperphosphorylation at serine 552, which is followed by activation of protein kinase Akt, driving IL-12 production and subsequent development of pathogenic IFN- γ -producing T cells (Zirnheld et al., 2019). Hence, it seems that IL-12 has pathogenic as well as beneficial effects at different stages of T1D progression (Fujihira et al., 2000; Nicoletti et al., 1999).

Additionally, T_H17 cells were shown to be diabetogenic after conversion into T_H1 cells (Mensah-Brown et al., 2006; Emamaullee et al., 2009; Martin-Orozco et al., 2009; Bending et al., 2009), leading to the assumption that T1D pathogenesis is driven by collaborative immune responses of T_H1 cells (IL-12, IFN- γ) and T_H17 cells (IL-23, IL-17) (Marwaha et al., 2014). A study in NOD mice, which were protected from experimental autoimmune diabetes by double deficiency of the IL-17 and IFN- γ receptors, further supports this assumption (Kuriya et al., 2013).

Several studies confirmed the importance of IL-12 for human T1D. For example, neonatal levels of IL-12 were positively associated with the risk of developing T1D in childhood (Thorsen et al., 2017) and children with T1D had increased IL-12 levels. Moreover, pediatric T1D patients had decreased levels of circulating T_{regs}, which was negatively correlated with the abundance of IL-12 (Ryba-Stanisławowska et al., 2014). Consistently, IFN- γ as well as IL-12 were found to be elevated in T1D patients without microvascular complications (MVC), the latter significantly (Shruthi et al., 2016). Furthermore, a model of T1D pathogenesis has been proposed, wherein IL-12 and IL-18 synergistically enhance cytotoxic T lymphocyte and NK cell

cytotoxic activity and disrupt immune regulation by T_{regs} (Dean et al., 2020).

As mentioned above, IL-12 does also play a role in the pathogenesis of T2D. A number of cytokines, including IL-6, IL-8, IL-10, IL-12 and secreted frizzled related protein 4 (SFRP4), as well as some microRNAs were described to be deregulated and associated with measures of pancreatic islet β cell function and glycemic control (Nunez Lopez et al., 2016). Moreover, it has been found that the repressive histone methylation mark, H3K27me₃, is decreased at the IL-12 promoter of bone marrow progenitors and passed down to wound macrophages in diet-induced obese glucose-intolerant mice. Under diabetic conditions, IL-12 production in macrophages is driven by the H3K27 demethylase Jmjd3 and can be modulated by its inhibition (Gallagher et al., 2015). Furthermore, in a high-fat-diet murine model for T2D it has been shown that the disruption of IL-12 promotes angiogenesis and increases blood flow recovery (Ali et al., 2017). A murine model for the treatment of T2D showed that inhibiting accumulation of group 1 innate lymphoid cells (ILC1) in the adipose tissue via IL-12-neutralizing antibodies alleviates adipose tissue fibrosis and is able to improve glycemic tolerance (Wang et al., 2019). Another study revealed, that supplementation of vitamin D3 may be beneficial for T1D and T2D patients with additional autoimmune thyroiditis. Following vitamin D3 supplementation, concentrations of inflammatory T_H1 cytokines (IFN- γ , TNF- α , IL-2, IL-6 and IL-12) decreased, whereas levels of anti-inflammatory T_H2-profile cytokines (IL-4, IL-5), IL-10 and IL-17 increased (Komisarenko and Bobryk, 2018).

To revolutionize the treatment options for T1D, new approaches have focussed on targeting IL-12 to at least partially replace or complement the exogenous administration of insulin. As part of this, ustekinumab is investigated in new-onset T1D and following a pilot trial demonstrating safety of ustekinumab in T1D, a phase II/III trial is under way (NCT03941132).

Systemic lupus erythematosus

Systemic lupus erythematosus (SLE) is an autoimmune disease characterized by antibodies to double-stranded DNA that can affect various parts of the body including for instance skin, joints, kidneys or nervous system (Wang and Xia, 2019).

Several studies demonstrated that IL-12 levels are increased in SLE patients (Uzrail et al., 2019; Zhou et al., 2019; Guimarães et al., 2017) and that risk loci for IL12RB (You et al., 2015) and genetic variants of IL12B (Paradowska-Gorycka et al., 2016) are associated with SLE. On the other hand, however, it has been shown that there is a T_{H17}/T_{reg} imbalance in patients with SLE (Talaat et al., 2015) as well as a shift in the T_{H1}/T_{H2} balance towards T_{H2} cytokines (Uzrail et al., 2019), suggesting that IL-12 is one cytokine among others in disease pathogenesis.

Recently, ustekinumab has been evaluated in patients with SLE in addition to standard-of-care treatment. It resulted in better efficacy in clinical and laboratory endpoints than placebo, with a safety profile comparable with ustekinumab therapy in other diseases and might therefore be an option to improve SLE therapy (van Vollenhoven et al., 2018).

Primary biliary cholangitis

Primary biliary cholangitis (PBC) is an autoimmune disease which specifically affects small bile ducts of the liver, previously known as primary biliary cirrhosis. Intrahepatic small bile ducts are destroyed by lymphocyte and plasma cell infiltration and anti-mitochondrial autoantibody (AMA) as well as high serum levels of IgM are characteristic for the disease (Tsuneyama et al., 2017).

As outlined for the diseases described above, IL-12 also plays an important role for the pathogenesis of PBC. In an experimental model of PBC, using dnTGF β R2 mice which have a dominant-negative transforming growth factor β receptor restricted to T cells, deletion of IL-12p35 resulted in reduced inflammation, whereas the deletion of IL-12p40 resulted in a complete protection against liver inflammation and bile duct damage (Tsuda et

al., 2013). Consistently, another study in mice could show that depletion of p40, leading to a decrease of the IL-12/ T_{H1} as well as the IL-23/ T_{H1} pathway, completely prevented the development of portal inflammatory cell infiltrates and biliary epithelial cell damage, suggesting that T_{H1} and T_{H17} effector responses affect the autoimmunity to biliary epithelial cells (Kawata et al., 2013).

Furthermore, several investigations showed that specific genetic predispositions are associated with PBC. Importantly, SNPs in the three IL-12-related genes IL12A, IL12RB2 and STAT4 are associated with PBC (Lleo et al., 2012; Hirschfield et al., 2009; Wasik et al., 2017). Moreover, PBC T_{reg} cells were shown to be more sensitive to IL-12-induced IFN- γ expression, fostering the notion that the IL-12-IL-12R β 2-STAT4 pathway in T_{regs} is important for disease pathogenesis and potentially treatment (Liaskou et al., 2018).

As for other IL-12-related diseases, ustekinumab was tested for the therapy of PBC patients with inadequate response to the standard therapy with ursodeoxycholic acid. Although no patient met the primary endpoint of a 40 % decline in alkaline phosphatase (ALP), the reduction of IL-17A, IFN- γ and IFN- α 2 levels in patients' serum demonstrated a pharmacodynamic effect of IL-12p40 inhibition (Hirschfield et al., 2016).

Sjögren's syndrome

Sjögren's syndrome (SS) is an autoimmune disorder resulting in dryness of mouth, eyes and other exocrine gland-connected surfaces, caused by mononuclear cell infiltrations. Further, it is associated with the production of specific autoantibodies (Jonsson et al., 2018). It is classified as primary Sjögren's syndrome (pSS), when symptoms appear without associated condition, whereas secondary Sjögren's syndrome (sSS) occurs together with another underlying autoimmune disorder, like rheumatoid arthritis, lupus erythematosus or scleroderma (Ramos-Casals et al., 2012).

Several mice models have been developed to investigate the role of IL-12 in the pathogenesis of SS. For example, McGrath-Morrow *et al.* generated a transgenic mouse model that overexpresses IL-12 in the lungs, which resulted in bronchial and alveolar abnormalities similar to those observed in Sjögren patients (McGrath-Morrow *et al.*, 2006). Moreover, in IL-12-transgenic SJL mice pilocarpine-stimulated salivary flow was significantly reduced and the number and size of lymphocytic foci was increased. IL-12 overexpression in CBA mice led to mononuclear infiltration of salivary and lacrimal glands, expansion of bronchial lymphoid tissue and decreased mucociliary clearance reminiscent of SS (Vosters *et al.*, 2009). Besides, IL-12 mRNA was predominantly expressed in the proinflammatory stage of autoimmune sialadenitis in MRL/lpr mice with experimental SS (Yanagi *et al.*, 1996) and plasma IL-12 was significantly increased in SS-like NOD mice, while anti-IL-12 alleviated the SS-like symptoms (Qi *et al.*, 2019).

Furthermore, it has been shown that the IL12A rs485497 polymorphism is associated with pSS and that IL-12p70 serum levels in patients with active disease are higher than in control subjects, whereas serum IL-35 levels were associated with low disease activity, indicating an involvement of the IL-12/IL-35 balance in the pathogenesis of pSS (Fogel *et al.*, 2018). Another study revealed increased serum concentrations of IL-10 and IL-12 in pSS patients, which were both significantly correlated with pro-inflammatory IL-6. Additionally, the T_H1/T_H2 ratio was significantly decreased in those patients (Girón-González *et al.*, 2009). Moreover, it has been shown that IL-12p40 and IL-15 levels were significantly decreased, while IL-1 β and TNF- α were significantly elevated in the plasma of SS patients. In addition, significant differences in IL-12p40 were described between patients with or without extra-glandular manifestations (Szodoray *et al.*, 2004) and high expression of IL-12 by mononuclear cell infiltrates in minor salivary glands has been observed (Manoussakis *et al.*, 2007). Interestingly,

mesenchymal stem cell transplantation (MSCT), an experimental therapeutic strategy proposed for SS, downregulated T_H17 and T_{FH} cells, but upregulated T_{regs} and reduced IL-12 production in SS patients as well as in mice, indicating that MSCs improve SS by suppressing the production of IL-12 in DCs (Shi *et al.*, 2018). Taken together, IL-12 might be a potential therapeutic target for SS that deserves further research.

Rheumatoid arthritis

Rheumatoid arthritis (RA) is caused by genetic as well as environmental factors. Characteristics include synovial inflammation and swelling possibly leading to skeletal deformation caused by destruction of cartilage and bones (McInnes and Schett, 2011). As for the other conditions discussed, RA manifestation is influenced by T_H1 and T_H17 cells, associated with IL-12 and IL-23 (Cornelissen *et al.*, 2009).

Collagen-induced arthritis (CIA) is a common mouse model for RA. The incidence and severity of CIA is significantly reduced in IL-12p40-deficient mice (McIntyre *et al.*, 1996). Similarly, treatment of type II collagen immunized DBA/1 mice with a neutralizing anti-IL-12 (p40) antibody mitigated the clinical and histopathological disease severity extensively (Malfait *et al.*, 1998). However, later studies dissecting the role of IL-12 and IL-23 rather provided evidence for a protective function of IL-12 by showing that mice specifically lacking IL-23 (p19^{-/-}) were protected from developing clinical signs of disease, whereas a specific lack of IL-12 (p35^{-/-}) resulted in aggravated collagen-induced arthritis (Murphy *et al.*, 2003). Consistently, IL-12 administration at the time of arthritis onset had a stimulatory effect on disease activity, while it seems to be suppressive in established CIA (Joosten *et al.*, 1997). Nevertheless, IL-12 does not remain uninvolved. Several gene polymorphism studies showed that IL-12B gene polymorphisms have an impact on RA pathogenesis, since, for instance, investigations in the Polish population showed that frequencies of the rs3212227 CC

of the IL12B gene were statistically higher in RA patients compared to controls and that the IL12B 1188A/C allele as well as IL-12p70 protein levels are likely to be associated with RA (Paradowska-Gorycka et al., 2017). Further, in a Bulgarian population, the IL12B rs17860508 polymorphism was associated with RA and RA patients with rs3212227 AA genotype of IL12B showed increased serum levels of IL-12p40 and IL-23 (Manolova et al., 2020). By contrast, a meta-analysis of 17 case-control studies demonstrated that IL-12B rs3212227 and rs6887695 polymorphisms do not confer susceptibility to RA (Yang et al., 2017). However, compared with healthy controls higher IL-12 levels can be found in the synovia (Morita et al., 1998) and serum (Paradowska-Gorycka et al., 2017; Cordero et al., 2001) of RA patients. Quite a number of drugs are used for the treatment of RA and some of their mechanisms have been linked with IL-12. For example, it has been suggested that methotrexate induces a down-regulation of IL-12 (Hobl et al., 2011) and modulates the T_H1/T_H2 balance towards a T_H2 profile by induction of IL-10 secretion and reduction of IL-12R and C-X-C motif chemokine receptor 3 (CXCR3) (Herman et al., 2008). Moreover, the phosphodiesterase 4 (PDE4) inhibitor Apremilast (Otezla[®]) strongly inhibited IL-12/IL-23p40 in cultured synovial fluid mononuclear cells from patients with active RA, psoriatic arthritis or peripheral spondyloarthritis (Kragstrup et al., 2019). Another PDE4 inhibitor, Ibudilast, reduced the expression and/or secretion of TNF and IL-12/IL-23p40 in activated human leukocytes and RA synovial fibroblasts and further inhibited T_H17 cell responses *in vivo* (Clanchy and Williams, 2019). Sinomenine, an alkaloid extracted from the Chinese medicinal plant *Sinomenium acutum*, which is approved in China, suppressed RA progression by regulating the secretion of several inflammatory cytokines, such as IL-12p40, IL-6, TNF- α (Liu et al., 2018). However, therapeutic strategies specifically targeting IL-12 have not been considered so far, since the role of IL-12 in RA does not seem to be essential.

CONCLUDING REMARKS

The history of IL-12 is marked by ups and downs. While it was associated with lots of pathological conditions shortly after its discovery based on methods and strategies detecting or targeting the p40 subunit, the diversification of the IL-12 family led to re-attribution of many of the functions initially assigned to IL-12 and other members like IL-23 gained more attention.

However, it must not be forgotten that targeting IL-12 together with IL-23 has become an established therapeutic strategy in IBD, psoriasis and peripheral spondyloarthritis, while experiences with anti-p19 antibodies are still limited. Thus, IL-12 is a key cytokine to consider in these diseases. Moreover, there are areas, where it seems to be “rediscovered”, particularly in the cancer field, where several promising approaches related to IL-12 are currently investigated.

Taken together, this cytokine positioned at an important nexus between the innate and adaptive response displays crucial functions in health and disease that are translationally relevant. Despite being known for more than 30 years now, we are still far away from completely understanding its involvement in physiological and pathological processes.

Competing interests

None.

Funding

The research of MFN and SZ was supported by the Interdisciplinary Center for Clinical Research (IZKF) and the ELAN program of the University Erlangen-Nuremberg, the Else Kröner-Fresenius-Stiftung, the Fritz Bender-Stiftung, the Dr Robert Pfleger Stiftung, the Litwin IBD Pioneers Initiative of the Crohn’s and Colitis Foundation of America (CCFA), the Kenneth Rainin Foundation, the Ernst Jung-Stiftung for Science and Research, the German Crohn’s and Colitis Foundation (DCCV) and the German Research Foundation (DFG) through individual grants (ZU 377/4-1) and the Collaborative Research Centers TRR241, 643, 796 and 1181.

REFERENCES

- Adedokun OJ, Xu Z, Gasink C, Jacobstein D, Szapary P, Johanns J, et al. Pharmacokinetics and exposure response relationships of ustekinumab in patients with Crohn's disease. *Gastroenterology*. 2018;154:1660-71.
- Adorini L. Interleukin-12, a key cytokine in Th1-mediated autoimmune diseases. *Cell Mol Life Sci*. 1999;55:1610-25.
- Aghaei Meybodi HR, Hasanzad M, Larijani B. Path to personalized medicine for type 2 diabetes mellitus: Reality and hope. *Acta Med Iran*. 2017;55:166-74.
- Ahn HM, Hong J, Yun CO. Oncolytic adenovirus co-expressing interleukin-12 and shVEGF restores anti-tumor immune function and enhances antitumor efficacy. *Oncotarget*. 2016;7:84965-80.
- Airoidi I, Di Carlo E, Banelli B, Moserle L, Cocco C, Pezzolo A, et al. The IL-12Rbeta2 gene functions as a tumor suppressor in human B cell malignancies. *J Clin Invest*. 2004;113:1651-9.
- Airoidi I, Di Carlo E, Cocco C, Sorrentino C, Fais F, Cilli M, et al. Lack of Il12rb2 signaling predisposes to spontaneous autoimmunity and malignancy. *Blood*. 2005;106:3846-53.
- Ali M, Mali V, Haddox S, AbdelGhany SM, El-Deek SEM, Abulfadl A, et al. Essential role of IL-12 in angiogenesis in type 2 diabetes. *Am J Pathol*. 2017;187:2590-601.
- Ansell SM, Witzig TE, Kurtin PJ, Sloan JA, Jelinek DF, Howell KG, et al. Phase 1 study of interleukin-12 in combination with rituximab in patients with B-cell non-Hodgkin lymphoma. *Blood*. 2002;99:67-74.
- Austin LM, Ozawa M, Kikuchi T, Walters IB, Krueger JG. The majority of epidermal T cells in Psoriasis vulgaris lesions can produce type 1 cytokines, interferon-gamma, interleukin-2, and tumor necrosis factor-alpha, defining TC1 (cytotoxic T lymphocyte) and TH1 effector populations: A type 1 differentiation bias is also measured in circulating blood T cells in psoriatic patients. *J Invest Dermatol*. 1999;113:752-9.
- Bacon CM, McVicar DW, Ortaldo JR, Rees RC, O'Shea JJ, Johnston JA. Interleukin 12 (IL-12) induces tyrosine phosphorylation of JAK2 and TYK2: differential use of Janus family tyrosine kinases by IL-2 and IL-12. *J Exp Med*. 1995;181:399-404.
- Bagaeva LV, Williams LP, Segal BM. IL-12 dependent/IFN γ independent expression of CCR5 by myelin-reactive T cells correlates with encephalitogenicity. *J Neuroimmunol*. 2003;137:109-16.
- Balashov KE, Smith DR, Khoury SJ, Hafler DA, Weiner HL. Increased interleukin 12 production in progressive multiple sclerosis: induction by activated CD4+ T cells via CD40 ligand. *Proc Natl Acad Sci U S A*. 1997;94:599-603.
- Ban M, Goris A, Lorentzen AR, Baker A, Mihalova T, Ingram G, et al. Replication analysis identifies TYK2 as a multiple sclerosis susceptibility factor. *Eur J Hum Genet*. 2009;17:1309-13.
- Barbulescu K, Becker C, Schlaak JF, Schmitt E, Meyer zum Büschenfelde KH, Neurath MF. IL-12 and IL-18 differentially regulate the transcriptional activity of the human IFN-gamma promoter in primary CD4+ T lymphocytes. *J Immunol*. 1998;160:3642-7.
- Barrios B, Baez NS, Reynolds D, Iribarren P, Cejas H, Young HA, et al. Abrogation of TNF α production during cancer immunotherapy is crucial for suppressing side effects due to the systemic expression of IL-12. *PLoS One*. 2014;9(2):e90116.
- Becher B, Durell BG, Noelle RJ. Experimental autoimmune encephalitis and inflammation in the absence of interleukin-12. *J Clin Invest*. 2002;110:493-7.
- Becker C, Wirtz S, Blessing M, Pirhonen J, Strand D, Bechthold O, et al. Constitutive p40 promoter activation and IL-23 production in the terminal ileum mediated by dendritic cells. *J Clin Invest*. 2003;112:693-706.
- Becker C, Wirtz S, Neurath MF. Stepwise regulation of TH1 responses in autoimmunity: IL-12-related cytokines and their receptors. *Inflamm Bowel Dis*. 2005;11:755-64.
- Becker C, Dornhoff H, Neufert C, Fantini MC, Wirtz S, Huebner S, et al. Cutting edge: IL-23 cross-regulates IL-12 production in T cell-dependent experimental colitis. *J Immunol*. 2006;177:2760-4.
- Bending D, De la Peña H, Veldhoen M, Phillips JM, Uyttenhove C, Stockinger B, et al. Highly purified Th17 cells from BDC2.5NOD mice convert into Th1-like cells in NOD/SCID recipient mice. *J Clin Invest*. 2009;119:565-72.
- Benkert TF, Dietz L, Hartmann EM, Leich E, Rosenwald A, Serfling E, et al. Natalizumab exerts direct signaling capacity and supports a pro-inflammatory phenotype in some patients with multiple sclerosis. *PLoS One*. 2012;7(12):e52208.
- Billerbeck E, Labitt RN, Vega K, Frias-Staheli N, Dornier M, Xiao JW, et al. Insufficient interleukin-12 signalling favours differentiation of human CD4(+) and CD8(+) T cells into GATA-3(+) and GATA-3(+) Tbet(+) subsets in humanized mice. *Immunology*. 2014;143:202-18.

- Bojko A, Ostasz R, Białecka M, Klimowicz A, Malinowski D, Budawski R, et al. IL12B, IL23A, IL23R and HLA-C*06 genetic variants in psoriasis susceptibility and response to treatment. *Hum Immunol.* 2018; 79:213-7.
- Bright JJ, Musuro BF, Du C, Sriram S. Expression of IL-12 in CNS and lymphoid organs of mice with experimental allergic encephalitis. *J Neuroimmunol.* 1998;82:22-30.
- Brunda MJ, Luistro L, Warriar RR, Wright RB, Hubbard BR, Murphy M, et al. Antitumor and antimetastatic activity of interleukin 12 against murine tumors. *J Exp Med.* 1993;178:1223-30.
- Bsat M, Chapuy L, Rubio M, Wassef R, Richard C, Schwenter F, et al. Differential pathogenic Th17 profile in mesenteric lymph nodes of Crohn's disease and ulcerative colitis patients. *Front Immunol.* 2019;10:1177.
- Cai Y, Fleming C, Yan J. New insights of T cells in the pathogenesis of psoriasis. *Cell Mol Immunol.* 2012;9:302-9.
- Campbell AR, Regan K, Bhave N, Pattanayak A, Parihar R, Stiff AR, et al. Gene expression profiling of the human natural killer cell response to Fc receptor activation: unique enhancement in the presence of interleukin-12. *BMC Med Genomics.* 2015;8:66.
- Car BD, Eng VM, Schnyder B, LeHir M, Shakhov AN, Woerly G, et al. Role of interferon-gamma in interleukin 12-induced pathology in mice. *Am J Pathol.* 1995; 147:1693-707.
- Carter LL, Murphy KM. Lineage-specific requirement for signal transducer and activator of transcription (Stat)4 in interferon γ production from CD4⁺ versus CD8⁺ T cells. *J Exp Med.* 1999;189:1355-60.
- Castro F, Cardoso AP, Gonçalves RM, Serre K, Oliveira MJ. Interferon-gamma at the crossroads of tumor immune surveillance or evasion. *Front Immunol.* 2018;9:847.
- Cataldi C, Mari NL, Lozovoy MAB, Martins LMM, Reiche EMV, Maes M, et al. Proinflammatory and anti-inflammatory cytokine profiles in psoriasis: use as laboratory biomarkers and disease predictors. *Inflamm Res.* 2019;68:557-67.
- Chan SH, Kobayashi M, Santoli D, Perussia B, Trinchieri G. Mechanisms of IFN-gamma induction by natural killer cell stimulatory factor (NKSF/IL-12). Role of transcription and mRNA stability in the synergistic interaction between NKSF and IL-2. *J Immunol.* 1992;148:92-8.
- Chen PY, Hsieh HY, Huang CY, Lin CY, Wei KC, Liu HL. Focused ultrasound-induced blood-brain barrier opening to enhance interleukin-12 delivery for brain tumor immunotherapy: a preclinical feasibility study. *J Transl Med.* 2015;13:93.
- Chen Y, Langrish CL, McKenzie B, Joyce-Shaikh B, Stumhofer JS, McClanahan T, et al. Anti-IL-23 therapy inhibits multiple inflammatory pathways and ameliorates autoimmune encephalomyelitis. *J Clin Invest.* 2006;116:1317-26.
- Chua AO, Chizzonite R, Desai BB, Truitt TP, Nunes P, Minetti LJ, et al. Expression cloning of a human IL-12 receptor component. A new member of the cytokine receptor superfamily with strong homology to gp130. *J Immunol.* 1994;153:128-36.
- Clanchy FIL, Williams RO. Ibudilast inhibits chemokine expression in rheumatoid arthritis synovial fibroblasts and exhibits immunomodulatory activity in experimental arthritis. *Arthritis Rheumatol.* 2019;71:703-11.
- Collison LW, Delgoffe GM, Guy CS, Vignali KM, Chaturvedi V, Fairweather D, et al. The composition and signaling of the IL-35 receptor are unconventional. *Nat Immunol.* 2012;13:290-9.
- Cooper AM, Khader SA. IL-12p40: An inherently agonistic cytokine. *Trends Immunol.* 2007;28:33-8.
- Cordero OJ, Salgado FJ, Mera-Varela A, Nogueira M. Serum interleukin-12, interleukin-15, soluble CD26, and adenosine deaminase in patients with rheumatoid arthritis. *Rheumatol Int.* 2001;21(2):69-74.
- Cornelissen F, van Hamburg JP, Lubberts E. The IL-12/IL-23 axis and its role in Th17 cell development, pathology and plasticity in arthritis. *Curr Opin Investig Drugs.* 2009;10:452-62.
- D'Andrea A, Rengaraju M, Valiante NM, Chehimi J, Kubin M, Aste M, et al. Production of natural killer cell stimulatory factor (interleukin 12) by peripheral blood mononuclear cells. *J Exp Med.* 1992;176:1387-98.
- Davidson NJ, Hudak SA, Lesley RE, Menon S, Leach MW, Rennick DM. IL-12, but not IFN-gamma, plays a major role in sustaining the chronic phase of colitis in IL-10-deficient mice. *J Immunol.* 1998;161:3143-9.
- Dean JW, Peters LD, Fuhrman CA, Seay HR, Posgai AL, Stimpson SE, et al. Innate inflammation drives NK cell activation to impair Treg activity. *J Autoimmun.* 2020;108:102417.
- Desai BB, Quinn PM, Wolitzky AG, Mongini PK, Chizzonite R, Gately MK. IL-12 receptor. II. Distribution and regulation of receptor expression. *J Immunol.* 1992;148:3125-32.

- Deshpande P, King IL, Segal BM. IL-12 driven upregulation of P-selectin ligand on myelin-specific T cells is a critical step in an animal model of autoimmune demyelination. *J Neuroimmunol.* 2006;173:35-44.
- Duchmann R, Schmitt E, Knolle P, Meyer zum Büschenfelde KH, Neurath M. Tolerance towards resident intestinal flora in mice is abrogated in experimental colitis and restored by treatment with interleukin-10 or antibodies to interleukin-12. *Eur J Immunol.* 1996;26:934-8.
- Eftychi C, Schwarzer R, Vlantis K, Wachsmuth L, Basic M, Wagle P, et al. Temporally distinct functions of the cytokines IL-12 and IL-23 drive chronic colon inflammation in response to intestinal barrier impairment. *Immunity.* 2019;51:367-80.e4.
- Emamaullee JA, Davis J, Merani S, Toso C, Elliott JF, Thiesen A, et al. Inhibition of Th17 cells regulates autoimmune diabetes in NOD mice. *Diabetes.* 2009;58:1302-11.
- Enerbäck C, Sandin C, Lambert S, Zawistowski M, Stuart PE, Verma D, et al. The psoriasis-protective TYK2 I684S variant impairs IL-12 stimulated pSTAT4 response in skin-homing CD4+ and CD8+ memory T-cells. *Sci Rep.* 2018;8(1):7043.
- Feagan BG, Sandborn WJ, Gasink C, Jacobstein D, Lang Y, Friedman JR, et al. Ustekinumab as induction and maintenance therapy for Crohn's disease. *N Engl J Med.* 2016;375:1946-60.
- Fogel O, Rivière E, Seror R, Nocturne G, Boudaoud S, Ly B, et al. Role of the IL-12/IL-35 balance in patients with Sjögren syndrome. *J Allergy Clin Immunol.* 2018;142:258-68.e5.
- Freytag SO, Barton KN, Zhang Y. Efficacy of oncolytic adenovirus expressing suicide genes and interleukin-12 in preclinical model of prostate cancer. *Gene Ther.* 2013;20:1131-9.
- Fujihira K, Nagata M, Moriyama H, Yasuda H, Arisawa K, Nakayama M, et al. Suppression and acceleration of autoimmune diabetes by neutralization of endogenous interleukin-12 in NOD mice. *Diabetes.* 2000;49:1998-2006.
- Fuss IJ, Neurath M, Boirivant M, Klein JS, de la Motte C, Strong SA, et al. Disparate CD4+ lamina propria (LP) lymphokine secretion profiles in inflammatory bowel disease. Crohn's disease LP cells manifest increased secretion of IFN-gamma, whereas ulcerative colitis LP cells manifest increased secretion of IL-5. *J Immunol.* 1996;157:1261-70.
- Fuss IJ, Marth T, Neurath MF, Pearlstein GR, Jain A, Strober W. Anti-interleukin 12 treatment regulates apoptosis of Th1 T cells in experimental colitis in mice. *Gastroenterology.* 1999;117:1078-88.
- Gallagher KA, Joshi A, Carson WF, Schaller M, Allen R, Mukerjee S, et al. Epigenetic changes in bone marrow progenitor cells influence the inflammatory phenotype and alter wound healing in type 2 diabetes. *Diabetes.* 2015;64:1420-30.
- Garris CS, Arlauckas SP, Kohler RH, Trefny MP, Garen S, Piot C, et al. Successful anti-PD-1 cancer immunotherapy requires T cell-dendritic cell crosstalk involving the cytokines IFN- γ and IL-12. *Immunity.* 2018;49:1148-61.e7.
- Gately MK, Carvajal DM, Connaughton SE, Gillessen S, Warriar RR, Kolinsky KD, et al. Interleukin-12 antagonist activity of mouse interleukin-12 p40 homodimer in vitro and in vivo. *Ann N Y Acad Sci.* 1996;795:1-12.
- Gautier G, Humbert M, Deauvieux F, Scullier M, Hiscott J, Bates EEM, et al. A type I interferon auto-crine-paracrine loop is involved in Toll-like receptor-induced interleukin-12p70 secretion by dendritic cells. *J Exp Med.* 2005;201:1435-46.
- Gerosa F, Paganin C, Peritt D, Paiola F, Scupoli MT, Aste-Amezaga M, et al. Interleukin-12 primes human CD4 and CD8 T cell clones for high production of both interferon-gamma and interleukin-10. *J Exp Med.* 1996;183:2559-69.
- Girón-González JA, Baturone R, Soto MJ, Márquez M, Macías I, Montes de Oca M, et al. Implications of immunomodulatory interleukins for the hyperimmunoglobulinemia of Sjögren's syndrome. *Cell Immunol.* 2009;259:56-60.
- Golab J, Zagodzón R. Antitumor effects of interleukin-12 in pre-clinical and early clinical studies (Review). *Int J Mol Med.* 1999;3:537-44.
- Gordon KB, Langley RG, Gottlieb AB, Papp KA, Krueger GG, Strober BE, et al. A phase III, randomized, controlled trial of the fully human IL-12/23 mAb briakinumab in moderate-to-severe psoriasis. *J Invest Dermatol.* 2012;132:304-14.
- Goriely S, Neurath MF, Goldman M. How microorganisms tip the balance between interleukin-12 family members. *Nat Rev Immunol.* 2008;8:81-6.
- Gran B, Zhang G-X, Yu S, Li J, Chen X-H, Ventura ES, et al. IL-12p35-deficient mice are susceptible to experimental autoimmune encephalomyelitis: Evidence for redundancy in the IL-12 system in the induction of central nervous system autoimmune demyelination. *J Immunol.* 2002;169:7104-10.

- Gran B, Zhang G-X, Rostami A. Role of the IL-12/IL-23 system in the regulation of T-cell responses in central nervous system inflammatory demyelination. *Crit Rev Immunol.* 2004;24:111-28.
- Griffiths CE, Barker JN. Pathogenesis and clinical features of psoriasis. *Lancet.* 2007;370(9583):263-71.
- Grifka-Walk HM, Giles DA, Segal BM. IL-12-polarized Th1 cells produce GM-CSF and induce EAE independent of IL-23. *Eur J Immunol.* 2015;45:2780-6.
- Grumont R, Hochrein H, O'Keeffe M, Gugasyan R, White C, Caminschi I, et al. c-Rel regulates interleukin 12 p70 expression in CD8(+) dendritic cells by specifically inducing p35 gene transcription. *J Exp Med.* 2001;194:1021-32.
- Gubler U, Chua AO, Schoenhaut DS, Dwyer CM, McComas W, Motyka R, et al. Coexpression of two distinct genes is required to generate secreted bioactive cytotoxic lymphocyte maturation factor. *Proc Natl Acad Sci U S A.* 1991;88:4143-7.
- Guimarães PM, Scavuzzi BM, Stadtlober NP, Franchi Santos L, Lozovoy MAB, Iriyoda TMV, et al. Cytokines in systemic lupus erythematosus: Far beyond Th1/Th2 dualism lupus: cytokine profiles. *Immunol Cell Biol.* 2017;95:824-31.
- Ha SJ, Lee CH, Lee SB, Kim CM, Jang KL, Shin HS, et al. A novel function of IL-12p40 as a chemotactic molecule for macrophages. *J Immunol.* 1999;163:2902-8.
- Haase O, Mosaad H, Eldarouti MA, Elramly AZ, Samir N, Abdelhady MM, et al. TNFAIP3 and IL12B gene polymorphisms associated with psoriasis vulgaris in an Egyptian cohort. *J Eur Acad Dermatol Venerol.* 2015;29:1297-301.
- Hanauer SB, Sandborn WJ, Feagan BG, Gasink C, Jacobstein D, Zou B, et al. IM-UNITI: Three-year efficacy, safety, and immunogenicity of ustekinumab treatment of Crohn's disease. *J Crohns Colitis.* 2020;14:23-32.
- Herman S, Zurgil N, Langevitz P, Ehrenfeld M, Deutsch M. Methotrexate selectively modulates TH1/TH2 balance in active rheumatoid arthritis patients. *Clin Exp Rheumatol.* 2008;26:317-23.
- Heufler C, Koch F, Stanzl U, Topar G, Wysocka M, Trinchieri G, et al. Interleukin-12 is produced by dendritic cells and mediates T helper 1 development as well as interferon-gamma production by T helper 1 cells. *Eur J Immunol.* 1996;26:659-68.
- Hirschfield GM, Liu X, Xu C, Lu Y, Xie G, Lu Y, et al. Primary biliary cirrhosis associated with HLA, IL12A, and IL12RB2 variants. *N Engl J Med.* 2009;360:2544-55.
- Hirschfield GM, Gershwin ME, Strauss R, Mayo MJ, Levy C, Zou B, et al. Ustekinumab for patients with primary biliary cholangitis who have an inadequate response to ursodeoxycholic acid: A proof-of-concept study. *Hepatology.* 2016;64:189-99.
- Hobl EL, Mader RM, Erlacher L, Duhm B, Mustak M, Bröll H, et al. The influence of methotrexate on the gene expression of the pro-inflammatory cytokine IL-12A in the therapy of rheumatoid arthritis. *Clin Exp Rheumatol.* 2011;29:963-9.
- Hsieh CS, Macatonia SE, Tripp CS, Wolf SF, O'Garra A, Murphy KM. Development of TH1 CD4+ T cells through IL-12 produced by Listeria-induced macrophages. *Science.* 1993;260(5107):547-9.
- Hu J, Zhu S, Xia X, Zhang L, Kleinerman ES, Li S. CD8+T cell-specific induction of NKG2D receptor by doxorubicin plus interleukin-12 and its contribution to CD8+T cell accumulation in tumors. *Mol Cancer.* 2014;13:34.
- Hurtubise R, Audiger C, Dominguez-Punaro MC, Chabot-Roy G, Chognard G, Raymond-Marchand L, et al. Induced and spontaneous colitis mouse models reveal complex interactions between IL-10 and IL-12/IL-23 pathways. *Cytokine.* 2019;121:154738.
- IMSGC, International Multiple Sclerosis Genetics Consortium. IL12A, MPHOSPH9/CDK2AP1 and RGS1 are novel multiple sclerosis susceptibility loci. *Genes Immun.* 2010;11:397-405.
- Ipseiz N, Uderhardt S, Scholtyssek C, Steffen M, Schabbauer G, Bozec A, et al. The nuclear receptor Nr4a1 mediates anti-inflammatory effects of apoptotic cells. *J Immunol.* 2014;192:4852-8.
- Jacobson NG, Szabo SJ, Weber-Nordt RM, Zhong Z, Schreiber RD, Darnell JE Jr, et al. Interleukin 12 signaling in T helper type 1 (Th1) cells involves tyrosine phosphorylation of signal transducer and activator of transcription (Stat)3 and Stat4. *J Exp Med.* 1995;181:1755-62.
- Jana M, Mondal S, Jana A, Pahan K. Interleukin-12 (IL-12), but not IL-23, induces the expression of IL-7 in microglia and macrophages: implications for multiple sclerosis. *Immunology.* 2014;141:549-63.
- Jeon C, Sekhon S, Yan D, Afifi L, Nakamura M, Bhutani T. Monoclonal antibodies inhibiting IL-12, -23, and -17 for the treatment of psoriasis. *Hum Vaccin Immunother.* 2017;13:2247-59.

- Jonsson R, Brokstad KA, Jonsson MV, Delaleu N, Skarstein K. Current concepts on Sjögren's syndrome - classification criteria and biomarkers. *Eur J Oral Sci.* 2018;126(Suppl 1):37-48.
- Joosten LA, Lubberts E, Helsen MM, van den Berg WB. Dual role of IL-12 in early and late stages of murine collagen type II arthritis. *J Immunol.* 1997;159:4094-102.
- Kamensek U, Cemazar M, Lamprecht Tratar U, Ursic K, Sersa G. Antitumor in situ vaccination effect of TNF α and IL-12 plasmid DNA electrotransfer in a murine melanoma model. *Cancer Immunol Immunother.* 2018;67:785-95.
- Karp CL, Biron CA, Irani DN. Interferon beta in multiple sclerosis: Is IL-12 suppression the key? *Immunol Today.* 2000;21(1):24-8.
- Katz JD, Benoist C, Mathis D. T helper cell subsets in insulin-dependent diabetes. *Science.* 1995;268(5214):1185-8.
- Kawata K, Tsuda M, Yang GX, Zhang W, Tanaka H, Tsuneyama K, et al. Identification of potential cytokine pathways for therapeutic intervention in murine primary biliary cirrhosis. *PLoS One.* 2013;8(9):e74225.
- Kobayashi M, Fitz L, Ryan M, Hewick RM, Clark SC, Chan S, et al. Identification and purification of natural killer cell stimulatory factor (NKSF), a cytokine with multiple biologic effects on human lymphocytes. *J Exp Med.* 1989;170:827-45.
- Komisarenko YI, Bobryk MI. Vitamin D deficiency and immune disorders in combined endocrine pathology. *Front Endocrinol (Lausanne).* 2018;9:600.
- Koneru M, O'Cearbhaill R, Pendharkar S, Spriggs DR, Brentjens RJ. A phase I clinical trial of adoptive T cell therapy using IL-12 secreting MUC-16(ecto) directed chimeric antigen receptors for recurrent ovarian cancer. *J Transl Med.* 2015;13:102.
- Kragstrup TW, Adams M, Lomholt S, Nielsen MA, Heftdal LD, Schafer P, et al. IL-12/IL-23p40 identified as a downstream target of apremilast in ex vivo models of arthritis. *Ther Adv Musculoskelet Dis.* 2019;11:1759720x19828669.
- Kroenke MA, Carlson TJ, Andjelkovic AV, Segal BM. IL-12- and IL-23-modulated T cells induce distinct types of EAE based on histology, CNS chemokine profile, and response to cytokine inhibition. *J Exp Med.* 2008;205:1535-41.
- Kulig P, Musiol S, Freiburger SN, Schreiner B, Gyulveszi G, Russo G, et al. IL-12 protects from psoriasisiform skin inflammation. *Nat Commun.* 2016;7:13466.
- Kundu M, Roy A, Pahan K. Selective neutralization of IL-12 p40 monomer induces death in prostate cancer cells via IL-12-IFN- γ . *Proc Natl Acad Sci U S A.* 2017;114:11482-7.
- Kuriya G, Uchida T, Akazawa S, Kobayashi M, Nakamura K, Satoh T, et al. Double deficiency in IL-17 and IFN- γ signalling significantly suppresses the development of diabetes in the NOD mouse. *Diabetologia.* 2013;56:1773-80.
- Lai I, Swaminathan S, Baylot V, Mosley A, Dhana-sekaran R, Gabay M, et al. Lipid nanoparticles that deliver IL-12 messenger RNA suppress tumorigenesis in MYC oncogene-driven hepatocellular carcinoma. *J Immunother Cancer.* 2018;6(1):125.
- Lamb P, Seidel HM, Stein RB, Rosen J. The role of JAKs and STATs in transcriptional regulation by cytokines. *Annu Rep Med Chem.* 1996;31:269-78.
- Langley RG, Elewski BE, Lebwohl M, Reich K, Griffiths CE, Papp K, et al. Secukinumab in plaque psoriasis - results of two phase 3 trials. *N Engl J Med.* 2014;371:326-38.
- Lasek W, Zagożdżon R, Jakobisiak M. Interleukin 12: Still a promising candidate for tumor immunotherapy? *Cancer Immunol Immunother.* 2014;63:419-35.
- Lenzi R, Edwards R, June C, Seiden MV, Garcia ME, Rosenblum M, et al. Phase II study of intraperitoneal recombinant interleukin-12 (rhIL-12) in patients with peritoneal carcinomatosis (residual disease < 1 cm) associated with ovarian cancer or primary peritoneal carcinoma. *J Transl Med.* 2007;5:66.
- Liaskou E, Patel SR, Webb G, Bagkou Dimakou D, Akiror S, Krishna M, et al. Increased sensitivity of Treg cells from patients with PBC to low dose IL-12 drives their differentiation into IFN- γ secreting cells. *J Autoimmun.* 2018;94:143-55.
- Liu J, Cao S, Herman LM, Ma X. Differential regulation of interleukin (IL)-12 p35 and p40 gene expression and interferon (IFN)-gamma-primed IL-12 production by IFN regulatory factor 1. *J Exp Med.* 2003;198:1265-76.
- Liu W, Zhang Y, Zhu W, Ma C, Ruan J, Long H, et al. Sinomenine inhibits the progression of rheumatoid arthritis by regulating the secretion of inflammatory cytokines and monocyte/macrophage subsets. *Front Immunol.* 2018;9:2228.

- Lleo A, Gershwin ME, Mantovani A, Invernizzi P. Towards common denominators in primary biliary cirrhosis: the role of IL-12. *J Hepatol.* 2012;56:731-3.
- Lowes MA, Kikuchi T, Fuentes-Duculan J, Cardinale I, Zaba LC, Haider AS, et al. Psoriasis vulgaris lesions contain discrete populations of Th1 and Th17 T cells. *J Invest Dermatol.* 2008;128:1207-11.
- Lupardus PJ, Garcia KC. The structure of interleukin-23 reveals the molecular basis of p40 subunit sharing with interleukin-12. *J Mol Biol.* 2008;382:931-41.
- Lyakh L, Trinchieri G, Provezza L, Carra G, Gerosa F. Regulation of interleukin-12/interleukin-23 production and the T-helper 17 response in humans. *Immunol Rev.* 2008;226:112-31.
- Ma X, Yan W, Zheng H, Du Q, Zhang L, Ban Y, et al. Regulation of IL-10 and IL-12 production and function in macrophages and dendritic cells. *F1000Res.* 2015;4:F1000 Faculty Rev-1465.
- Macatonia SE, Hsieh CS, Murphy KM, O'Garra A. Dendritic cells and macrophages are required for Th1 development of CD4+ T cells from alpha beta TCR transgenic mice: IL-12 substitution for macrophages to stimulate IFN-gamma production is IFN-gamma-dependent. *Int Immunol.* 1993;5:1119-28.
- Malfait AM, Butler DM, Presky DH, Maini RN, Brennan FM, Feldmann M. Blockade of IL-12 during the induction of collagen-induced arthritis (CIA) markedly attenuates the severity of the arthritis. *Clin Exp Immunol.* 1998;111:377-83.
- Manolova I, Ivanova M, Vasilev G, Stoilov R, Miteva L, Stanilova S. Impact of IL12B polymorphisms on genetic susceptibility and IL-12p40 and IL-23 serum levels in rheumatoid arthritis. *Immunol Invest.* 2020;49(1-2):1-14.
- Manoussakis MN, Boiu S, Korkolopoulou P, Kapsogeorgou EK, Kavantzias N, Ziakas P, et al. Rates of infiltration by macrophages and dendritic cells and expression of interleukin-18 and interleukin-12 in the chronic inflammatory lesions of Sjögren's syndrome: Correlation with certain features of immune hyperactivity and factors associated with high risk of lymphoma development. *Arthritis Rheum.* 2007;56:3977-88.
- Martin-Orozco N, Chung Y, Chang SH, Wang YH, Dong C. Th17 cells promote pancreatic inflammation but only induce diabetes efficiently in lymphopenic hosts after conversion into Th1 cells. *Eur J Immunol.* 2009;39:216-24.
- Marwaha AK, Tan S, Dutz JP. Targeting the IL-17/IFN- γ axis as a potential new clinical therapy for type 1 diabetes. *Clin Immunol.* 2014;154:84-9.
- McGrath-Morrow S, Laube B, Tzou SC, Cho C, Cleary J, Kimura H, et al. IL-12 overexpression in mice as a model for Sjögren lung disease. *Am J Physiol Lung Cell Mol Physiol.* 2006;291:L837-46.
- McInnes IB, Schett G. The pathogenesis of rheumatoid arthritis. *N Engl J Med.* 2011;365:2205-19.
- McInnes IB, Kavanaugh A, Gottlieb AB, Puig L, Rahman P, Ritchlin C, et al. Efficacy and safety of ustekinumab in patients with active psoriatic arthritis: 1 year results of the phase 3, multicentre, double-blind, placebo-controlled PSUMMIT 1 trial. *Lancet.* 2013;382(9894):780-9.
- McIntyre KW, Shuster DJ, Gillooly KM, Warriar RR, Connaughton SE, Hall LB, et al. Reduced incidence and severity of collagen-induced arthritis in interleukin-12-deficient mice. *Eur J Immunol.* 1996;26:2933-8.
- Mensah-Brown EP, Shahin A, Al-Shamisi M, Wei X, Lukic ML. IL-23 leads to diabetes induction after subdiabetogenic treatment with multiple low doses of streptozotocin. *Eur J Immunol.* 2006;36:216-23.
- Montes de Oca M, Kumar R, de Labastida Rivera F, Amante FH, Sheel M, Faleiro RJ, et al. Blimp-1-dependent IL-10 production by Tr1 cells regulates TNF-mediated tissue pathology. *PLoS Pathog.* 2016;12(1):e1005398.
- Morita Y, Yamamura M, Nishida K, Harada S, Okamoto H, Inoue H, et al. Expression of interleukin-12 in synovial tissue from patients with rheumatoid arthritis. *Arthritis Rheum.* 1998;41:306-14.
- Moschen AR, Tilg H, Raine T. IL-12, IL-23 and IL-17 in IBD: Immunobiology and therapeutic targeting. *Nat Rev Gastroenterol Hepatol.* 2019;16:185-96.
- Murphy CA, Langrish CL, Chen Y, Blumenschein W, McClanahan T, Kastelein RA, et al. Divergent pro- and antiinflammatory roles for IL-23 and IL-12 in joint autoimmune inflammation. *J Exp Med.* 2003;198:1951-7.
- Murphy EP, Crean D. Molecular interactions between NR4A orphan nuclear receptors and NF- κ B are required for appropriate inflammatory responses and immune cell homeostasis. *Biomolecules.* 2015;5:1302-18.
- Musabak U, Demirkaya S, Genç G, Ilikei RS, Odabasi Z. Serum adiponectin, TNF- α , IL-12p70, and IL-13 levels in multiple sclerosis and the effects of different therapy regimens. *Neuroimmunomodulation.* 2011;18(1):57-66.

- Naeger LK, McKinney J, Salvekar A, Hoey T. Identification of a STAT4 binding site in the interleukin-12 receptor required for signaling. *J Biol Chem.* 1999;274:1875-8.
- Nair A, Chakraborty S, Banerji LA, Srivastava A, Navare C, Saha B. Ras isoforms: Signaling specificities in CD40 pathway. *Cell Commun Signal.* 2020;18(1):3.
- Najar M, Krayem M, Meuleman N, Bron D, Lagneaux L. Mesenchymal stromal cells and toll-like receptor priming: A critical review. *Immune Netw.* 2017;17(2):89-102.
- Nelson SM, Nguyen TM, McDonald JW, MacDonald JK. Natalizumab for induction of remission in Crohn's disease. *Cochrane Database Syst Rev.* 2018;8(8):Cd006097.
- Nemeth ZH, Bogdanovski DA, Barratt-Stopper P, Paglinco SR, Antonioli L, Rolandelli RH. Crohn's disease and ulcerative colitis show unique cytokine profiles. *Cureus.* 2017;9(4):e1177.
- Neurath MF. Cytokines in inflammatory bowel disease. *Nat Rev Immunol.* 2014;14:329-42.
- Nguyen HM, Guz-Montgomery K, Saha D. Oncolytic virus encoding a master pro-inflammatory cytokine interleukin 12 in cancer immunotherapy. *Cells.* 2020;9(2):400.
- Nicoletti F, Di Marco R, Zacccone P, Magro G, Di Mauro M, Grasso S, et al. Endogenous interleukin-12 only plays a key pathogenetic role in non-obese diabetic mouse diabetes during the very early stages of the disease. *Immunology.* 1999;97:367-70.
- Niedbala W, Wei XQ, Cai B, Hueber AJ, Leung BP, McInnes IB, et al. IL-35 is a novel cytokine with therapeutic effects against collagen-induced arthritis through the expansion of regulatory T cells and suppression of Th17 cells. *Eur J Immunol.* 2007;37:3021-9.
- Noble A, Thomas MJ, Kemeny DM. Early Th1/Th2 cell polarization in the absence of IL-4 and IL-12: T cell receptor signaling regulates the response to cytokines in CD4 and CD8 T cells. *Eur J Immunol.* 2001;31:2227-35.
- Nograla KE, Zaba LC, Shemer A, Fuentes-Duculan J, Cardinale I, Kikuchi T, et al. IL-22-producing "T22" T cells account for upregulated IL-22 in atopic dermatitis despite reduced IL-17-producing TH17 T cells. *J Allergy Clin Immunol.* 2009;123:1244-52.e2.
- Norouzinia M, Chaleshi V, Alinaghi S, Beheshti Shirazi SS, Keramatinia A, Nourian M. Evaluation of IL-12A, IL-12B, IL-23A and IL-27 mRNA expression level genes in peripheral mononuclear cells of inflammatory bowel disease patients in an Iranian population. *Gastroenterol Hepatol Bed Bench.* 2018;11(Suppl 1):S45-52.
- Nunez Lopez YO, Garufi G, Seyhan AA. Altered levels of circulating cytokines and microRNAs in lean and obese individuals with prediabetes and type 2 diabetes. *Mol Biosyst.* 2016;13(1):106-21.
- Oh E, Choi IK, Hong J, Yun CO. Oncolytic adenovirus coexpressing interleukin-12 and decorin overcomes Treg-mediated immunosuppression inducing potent antitumor effects in a weakly immunogenic tumor model. *Oncotarget.* 2017;8:4730-46.
- Papp KA, Langley RG, Lebwohl M, Krueger GG, Szapary P, Yeilding N, et al. Efficacy and safety of ustekinumab, a human interleukin-12/23 monoclonal antibody, in patients with psoriasis: 52-week results from a randomised, double-blind, placebo-controlled trial (PHOENIX 2). *Lancet.* 2008;371(9625):1675-84.
- Papp KA, Blauvelt A, Bukhalo M, Gooderham M, Krueger JG, Lacour JP, et al. Risankizumab versus ustekinumab for moderate-to-severe plaque psoriasis. *N Engl J Med.* 2017;376:1551-60.
- Paradowska-Gorycka A, Sowinska A, Stypinska B, Grobelna MK, Walczyk M, Olesinska M, et al. Genetic variants in IL-12B and IL-27 in the Polish patients with systemic lupus erythematosus. *Scand J Immunol.* 2016;84:49-60.
- Paradowska-Gorycka A, Sowinska A, Stypińska B, Haladyj E, Pawlik A, Romanowska-Próchnicka K, et al. IL-12B gene polymorphisms and IL-12 p70 serum levels among patients with rheumatoid arthritis. *Scand J Immunol.* 2017;85:147-54.
- Parham C, Chirica M, Timans J, Vaisberg E, Travis M, Cheung J, et al. A receptor for the heterodimeric cytokine IL-23 is composed of IL-12Rbeta1 and a novel cytokine receptor subunit, IL-23R. *J Immunol.* 2002;168:5699-708.
- Pflanz S, Hibbert L, Mattson J, Rosales R, Vaisberg E, Bazan JF, et al. WSX-1 and glycoprotein 130 constitute a signal-transducing receptor for IL-27. *J Immunol.* 2004;172:2225-31.
- Portielje JE, Kruit WH, Schuler M, Beck J, Lamers CH, Stoter G, et al. Phase I study of subcutaneously administered recombinant human interleukin 12 in patients with advanced renal cell cancer. *Clin Cancer Res.* 1999;5:3983-9.

- Pot C, Apetoh L, Awasthi A, Kuchroo VK. Molecular pathways in the induction of interleukin-27-driven regulatory type 1 cells. *J Interferon Cytokine Res.* 2010; 30:381-8.
- Powell MD, Read KA, Sreekumar BK, Jones DM, Oestreich KJ. IL-12 signaling drives the differentiation and function of a TH1-derived TFH1-like cell population. *Sci Rep.* 2019;9(1):13991.
- Presky DH, Yang H, Minetti LJ, Chua AO, Nabavi N, Wu CY, et al. A functional interleukin 12 receptor complex is composed of two beta-type cytokine receptor subunits. *Proc Natl Acad Sci U S A.* 1996;93: 14002-7.
- Qi J, Li D, Shi G, Zhang X, Pan Y, Dou H, et al. Interleukin-12 exacerbates Sjögren's syndrome through induction of myeloid-derived suppressor cells. *Mol Med Rep.* 2019;20:1131-8.
- Rabinowich H, Herberman RB, Whiteside TL. Differential effects of IL12 and IL2 on expression and function of cellular adhesion molecules on purified human natural killer cells. *Cell Immunol.* 1993;152:481-98.
- Ramos-Casals M, Brito-Zerón P, Sisó-Almirall A, Bosch X. Primary Sjogren syndrome. *BMJ.* 2012;344: e3821.
- Ryba-Stanisławowska M, Rybarczyk-Kapturska K, Myśliwiec M, Myśliwska J. Elevated levels of serum IL-12 and IL-18 are associated with lower frequencies of CD4(+)CD25 (high)FOXP3 (+) regulatory Tt cells in young patients with type 1 diabetes. *Inflammation.* 2014;37:1513-20.
- Sandborn WJ, Gasink C, Gao LL, Blank MA, Johans J, Guzzo C, et al. Ustekinumab induction and maintenance therapy in refractory Crohn's disease. *N Engl J Med.* 2012;367:1519-28.
- Sandborn WJ, Ferrante M, Bhandari BR, Berliba E, Feagan BG, Hibi T, et al. Efficacy and safety of mirikizumab in a randomized phase 2 study of patients with ulcerative colitis. *Gastroenterology.* 2020;158: 537-49.e10.
- Sands BE, Chen J, Feagan BG, Penney M, Rees WA, Danese S, et al. Efficacy and safety of MEDI2070, an antibody against interleukin 23, in patients with moderate to severe Crohn's disease: A phase 2a study. *Gastroenterology.* 2017;153:77-86.e6.
- Sands BE, Sandborn WJ, Panaccione R, O'Brien CD, Zhang H, Johans J, et al. Ustekinumab as induction and maintenance therapy for ulcerative colitis. *N Engl J Med.* 2019;381:1201-14.
- Schlaak JF, Buslau M, Jochum W, Hermann E, Girndt M, Gallati H, et al. T cells involved in psoriasis vulgaris belong to the Th1 subset. *J Invest Dermatol.* 1994; 102:145-9.
- Segal BM, Dwyer BK, Shevach EM. An interleukin (IL)-10/IL-12 immunoregulatory circuit controls susceptibility to autoimmune disease. *J Exp Med.* 1998; 187:537-46.
- Segal BM, Constantinescu CS, Raychaudhuri A, Kim L, Fidelus-Gort R, Kasper LH. Repeated subcutaneous injections of IL12/23 p40 neutralising antibody, ustekinumab, in patients with relapsing-remitting multiple sclerosis: a phase II, double-blind, placebo-controlled, randomised, dose-ranging study. *Lancet Neurol.* 2008;7(9):796-804.
- Shi B, Qi J, Yao G, Feng R, Zhang Z, Wang D, et al. Mesenchymal stem cell transplantation ameliorates Sjögren's syndrome via suppressing IL-12 production by dendritic cells. *Stem Cell Res Ther.* 2018;9(1):308.
- Shirley SA, Lundberg CG, Li F, Burcus N, Heller R. Controlled gene delivery can enhance therapeutic outcome for cancer immune therapy for melanoma. *Curr Gene Ther.* 2015;15(1):32-43.
- Shruthi S, Mohan V, Amutha A, Aravindhan V. Increased serum levels of novel T cell cytokines IL-33, IL-9 and IL-17 in subjects with type-1 diabetes. *Cytokine.* 2016;86:6-9.
- Sieburth D, Jabs EW, Warrington JA, Li X, Lasota J, LaForgia S, et al. Assignment of genes encoding a unique cytokine (IL12) composed of two unrelated subunits to chromosomes 3 and 5. *Genomics.* 1992;14: 59-62.
- Smyth MJ, Taniguchi M, Street SEA. The anti-tumor activity of IL-12: Mechanisms of innate immunity that are model and dose dependent. *J Immunol.* 2000;165: 2665-70.
- Snijders A, Hilkens CM, van der Pouw Kraan TC, Engel M, Aarden LA, Kapsenberg ML. Regulation of bioactive IL-12 production in lipopolysaccharide-stimulated human monocytes is determined by the expression of the p35 subunit. *J Immunol.* 1996;156(3):1207-12.
- Stern AS, Podlaski FJ, Hulmes JD, Pan YC, Quinn PM, Wolitzky AG, et al. Purification to homogeneity and partial characterization of cytotoxic lymphocyte maturation factor from human B-lymphoblastoid cells. *Proc Natl Acad Sci U S A.* 1990;87(17):6808-12.
- Szabo SJ, Dighe AS, Gubler U, Murphy KM. Regulation of the interleukin (IL)-12R beta 2 subunit expression in developing T helper 1 (Th1) and Th2 cells. *J Exp Med.* 1997;185:817-24.

- Szodoray P, Alex P, Brun JG, Centola M, Jonsson R. Circulating cytokines in primary Sjögren's syndrome determined by a multiplex cytokine array system. *Scand J Immunol.* 2004;59:592-9.
- Tait Wojno ED, Hunter CA, Stumhofer JS. The immunobiology of the interleukin-12 family: Room for discovery. *Immunity.* 2019;50:851-70.
- Takahashi T, Koga Y, Kainoh M. Anti-IL-12/IL-23p40 antibody ameliorates dermatitis and skin barrier dysfunction in mice with imiquimod-induced psoriasis-like dermatitis. *Eur J Pharmacol.* 2018;828:26-30.
- Talaat RM, Mohamed SF, Bassyouni IH, Raouf AA. Th1/Th2/Th17/Treg cytokine imbalance in systemic lupus erythematosus (SLE) patients: Correlation with disease activity. *Cytokine.* 2015;72:146-53.
- Thaker PH, Brady WE, Lankes HA, Odunsi K, Bradley WH, Moore KN, et al. A phase I trial of intraperitoneal GEN-1, an IL-12 plasmid formulated with PEG-PEI-cholesterol lipopolymer, administered with pegylated liposomal doxorubicin in patients with recurrent or persistent epithelial ovarian, fallopian tube or primary peritoneal cancers: An NRG Oncology/Gynecologic Oncology Group study. *Gynecol Oncol.* 2017;147:283-90.
- Thorsen SU, Pipper CB, Eising S, Skogstrand K, Hougaard DM, Svensson J, et al. Neonatal levels of adiponectin, interleukin-10 and interleukin-12 are associated with the risk of developing type 1 diabetes in childhood and adolescence: A nationwide Danish case-control study. *Clin Immunol.* 2017;174:18-23.
- Trembleau S, Penna G, Bosi E, Mortara A, Gately MK, Adorini L. Interleukin 12 administration induces T helper type 1 cells and accelerates autoimmune diabetes in NOD mice. *J Exp Med.* 1995;181:817-21.
- Trembleau S, Penna G, Gregori S, Giarratana N, Adorini L. IL-12 administration accelerates autoimmune diabetes in both wild-type and IFN-gamma-deficient nonobese diabetic mice, revealing pathogenic and protective effects of IL-12-induced IFN-gamma. *J Immunol.* 2003;170:5491-501.
- Trinchieri G. Interleukin-12: A cytokine at the interface of inflammation and immunity. *Adv Immunol.* 1998;70:83-243.
- Trinchieri G. Interleukin-12 and the regulation of innate resistance and adaptive immunity. *Nat Rev Immunol.* 2003;3:133-46.
- Tsuda M, Zhang W, Yang GX, Tsuneyama K, Ando Y, Kawata K, et al. Deletion of interleukin (IL)-12p35 induces liver fibrosis in dominant-negative TGFβ receptor type II mice. *Hepatology.* 2013;57:806-16.
- Tsuneyama K, Baba H, Morimoto Y, Tsunematsu T, Ogawa H. Primary biliary cholangitis: its pathological characteristics and immunopathological mechanisms. *J Med Invest.* 2017;64(1.2):7-13.
- Tugues S, Burkhard SH, Ohs I, Vrohings M, Nussbaum K, Vom Berg J, et al. New insights into IL-12-mediated tumor suppression. *Cell Death Differ.* 2015;22:237-46.
- Uhlig HH, McKenzie BS, Hue S, Thompson C, Joyce-Shaikh B, Stepankova R, et al. Differential activity of IL-12 and IL-23 in mucosal and systemic innate immune pathology. *Immunity.* 2006;25:309-18.
- Uzrail AH, Assaf AM, Abdalla SS. Correlations of expression levels of a panel of genes (IRF5, STAT4, TNFSF4, MECP2, and TLR7) and cytokine levels (IL-2, IL-6, IL-10, IL-12, IFN-γ, and TNF-α) with systemic Lupus erythematosus outcomes in Jordanian patients. *Biomed Res Int.* 2019;2019:1703842.
- Vacaflares A, Chapman NM, Harty JT, Richer MJ, Houtman JC. Exposure of human CD4 T cells to IL-12 results in enhanced TCR-induced cytokine production, altered TCR signaling, and increased oxidative metabolism. *PLoS One.* 2016;11(6):e0157175.
- Vacaflares A, Freedman SN, Chapman NM, Houtman JC. Pretreatment of activated human CD8 T cells with IL-12 leads to enhanced TCR-induced signaling and cytokine production. *Mol Immunol.* 2017;81:1-15.
- van den Reek J, Coenen MJH, van de L'Isle Arias M, Zweegers J, Rodijk-Olthuis D, Schalkwijk J, et al. Polymorphisms in CD84, IL12B and TNFAIP3 are associated with response to biologics in patients with psoriasis. *Br J Dermatol.* 2017;176:1288-96.
- van Vollenhoven RF, Hahn BH, Tsokos GC, Wagner CL, Lipsky P, Touma Z, et al. Efficacy and safety of ustekinumab, an IL-12 and IL-23 inhibitor, in patients with active systemic lupus erythematosus: Results of a multicentre, double-blind, phase 2, randomised, controlled study. *Lancet.* 2018;392:1330-9.
- Vandenbroeck K, Alloza I, Gadina M, Matthys P. Inhibiting cytokines of the interleukin-12 family: Recent advances and novel challenges. *J Pharm Pharmacol.* 2004;56:145-60.
- Vignali DA, Kuchroo VK. IL-12 family cytokines: Immunological playmakers. *Nat Immunol.* 2012;13:722-8.
- Vollmer TL, Wynn DR, Alam MS, Valdes J. A phase 2, 24-week, randomized, placebo-controlled, double-blind study examining the efficacy and safety of an anti-interleukin-12 and -23 monoclonal antibody in patients with relapsing–remitting or secondary progressive multiple sclerosis. *Mult Scler.* 2011;17:181-91.

- von Essen MR, Søndergaard HB, Petersen ERS, Selberg F. IL-6, IL-12, and IL-23 STAT-pathway genetic risk and responsiveness of lymphocytes in patients with multiple sclerosis. *Cells*. 2019;8(3):285.
- Vosters JL, Landek-Salgado MA, Yin H, Swaim WD, Kimura H, Tak PP, et al. Interleukin-12 induces salivary gland dysfunction in transgenic mice, providing a new model of Sjögren's syndrome. *Arthritis Rheum*. 2009;60:3633-41.
- Walker W, Aste-Amezaga M, Kastelein RA, Trinchieri G, Hunter CA. IL-18 and CD28 use distinct molecular mechanisms to enhance NK cell production of IL-12-induced IFN-gamma. *J Immunol*. 1999;162:5894-901.
- Wang H, Meng R, Li Z, Yang B, Liu Y, Huang F, et al. IL-27 induces the differentiation of Tr1-like cells from human naive CD4⁺ T cells via the phosphorylation of STAT1 and STAT3. *Immunol Lett*. 2011;136:21-8.
- Wang H, Shen L, Sun X, Liu F, Feng W, Jiang C, et al. Adipose group 1 innate lymphoid cells promote adipose tissue fibrosis and diabetes in obesity. *Nat Commun*. 2019;10(1):3254.
- Wang IM, Contursi C, Masumi A, Ma X, Trinchieri G, Ozato K. An IFN-gamma-inducible transcription factor, IFN consensus sequence binding protein (ICSBP), stimulates IL-12 p40 expression in macrophages. *J Immunol*. 2000;165:271-9.
- Wang P, Li X, Wang J, Gao D, Li Y, Li H, et al. Redesigning interleukin-12 to enhance its safety and potential as an anti-tumor immunotherapeutic agent. *Nat Commun*. 2017;8(1):1395.
- Wang X, Xia Y. Anti-double stranded DNA antibodies: Origin, pathogenicity, and targeted therapies. *Front Immunol*. 2019;10:1667.
- Wasik U, Wunsch E, Norman GL, Rigopoulou EI, Bogdanos DP, Milkiewicz P, et al. Polymorphisms of IL12RB2 May affect the natural history of primary biliary cholangitis: A single centre study. *J Immunol Res*. 2017;2017:2185083.
- Wirtz S, Finotto S, Kanzler S, Lohse AW, Blessing M, Lehr HA, et al. Cutting edge: chronic intestinal inflammation in STAT-4 transgenic mice: characterization of disease and adoptive transfer by TNF- plus IFN-gamma-producing CD4⁺ T cells that respond to bacterial antigens. *J Immunol*. 1999;162:1884-8.
- Xu X, Sun Y-L, Hoey T. Cooperative DNA binding and sequence-selective recognition conferred by the STAT amino-terminal domain. *Science*. 1996;273(5276):794-7.
- Yan J, Smyth MJ, Teng MWL. Interleukin (IL)-12 and IL-23 and their conflicting roles in cancer. *Cold Spring Harb Perspect Biol*. 2018;10(7):a028530.
- Yanagi K, Haneji N, Hamano H, Takahashi M, Higashiyama H, Hayashi Y. In vivo role of IL-10 and IL-12 during development of Sjögren's syndrome in MRL/lpr mice. *Cell Immunol*. 1996;168:243-50.
- Yang X, Xiao F, Luo D, Wang G, Liang S. Interleukin 12B gene polymorphisms and susceptibility to rheumatoid arthritis: a data synthesis. *Clin Rheumatol*. 2017;36:299-307.
- Yawalkar N, Tscherner GG, Hunger RE, Hassan AS. Increased expression of IL-12p70 and IL-23 by multiple dendritic cell and macrophage subsets in plaque psoriasis. *J Dermatol Sci*. 2009;54:99-105.
- Yen D, Cheung J, Scheerens H, Poulet F, McClanahan T, McKenzie B, et al. IL-23 is essential for T cell-mediated colitis and promotes inflammation via IL-17 and IL-6. *J Clin Invest*. 2006;116:1310-6.
- Yin P, Liu X, Mansfield AS, Harrington SM, Li Y, Yan Y, et al. CpG-induced antitumor immunity requires IL-12 in expansion of effector cells and down-regulation of PD-1. *Oncotarget*. 2016;7:70223-31.
- You Y, Zhai ZF, Chen FR, Chen W, Hao F. Autoimmune risk loci of IL12RB2, IKZF1, XKR6, TMEM39A and CSK in Chinese patients with systemic lupus erythematosus. *Tissue Antigens*. 2015;85:200-3.
- Yue T, Zheng X, Dou Y, Zheng X, Sun R, Tian Z, et al. Interleukin 12 shows a better curative effect on lung cancer than paclitaxel and cisplatin doublet chemotherapy. *BMC Cancer*. 2016;16(1):665.
- Zhang L, Kerkar SP, Yu Z, Zheng Z, Yang S, Restifo NP, et al. Improving adoptive T cell therapy by targeting and controlling IL-12 expression to the tumor environment. *Mol Ther*. 2011;19:751-9.
- Zhang S, Kaplan MH. The p38 mitogen-activated protein kinase is required for IL-12-induced IFN-gamma expression. *J Immunol*. 2000;165:1374-80.
- Zhao Q, Du Q, Wei F, Xie J, Ma X. An infectious disease-associated Il12b polymorphism regulates IL-12/23 p40 transcription involving poly(ADP-ribose) polymerase 1. *J Immunol*. 2017;198:2935-42.
- Zheng Y, Wang M, Tian T, Liu K, Liu X, Zhai Y, et al. Role of interleukin-12 gene polymorphisms in the onset risk of cancer: a meta-analysis. *Oncotarget*. 2017;8:29795-807.

Zhong Z, Wen Z, Darnell J. Stat3: a STAT family member activated by tyrosine phosphorylation in response to epidermal growth factor and interleukin-6. *Science*. 1994;264(5155):95-8.

Zhou H, Li B, Li J, Wu T, Jin X, Yuan R, et al. Dysregulated T cell activation and aberrant cytokine expression profile in systemic lupus erythematosus. *Mediators Inflamm*. 2019;2019:8450947.

Zirnheld AL, Villard M, Harrison AM, Kosiewicz MM, Alard P. β -Catenin stabilization in NOD dendritic cells increases IL-12 production and subsequent induction of IFN- γ -producing T cells. *J Leukoc Biol*. 2019; 106:1349-58.

Zou J, Presky DH, Wu CY, Gubler U. Differential associations between the cytoplasmic regions of the interleukin-12 receptor subunits beta1 and beta2 and JAK kinases. *J Biol Chem*. 1997;272:6073-7.