

Stress and Pathogenesis of Infectious Disease

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Despite inherent difficulties in defining and measuring stress, a scientific framework has been provided in recent years for understanding how disruptive life experiences might be translated into altered susceptibility to infectious diseases. Studies of the effects of stress on pathogenesis of infectious disease are highly relevant to assessment of the biological importance of the immune impairments that have been associated with stress. With a few notable exceptions, investigations of viral infections in humans and in animal models support the hypothesis that stress promotes the pathogenesis of such infections. Similar conclusions can be drawn from studies of bacterial infections in humans and animals and from a small number of studies of parasitic infections in rodent models. While many of these studies have substantial limitations, the data nonetheless suggest that stress is a potential cofactor in the pathogenesis of infectious disease. Given recent unprecedented advances in the neurosciences, in immunology, and in the field of microbial pathogenesis, the relationship between stress and infection should be a fruitful topic for interdisciplinary research.

"The toxin of fatigue has been demonstrated; but the poisons generated by evil temper and emotional excess over non-essentials have not yet been determined, although without a doubt they exist."

Elie Metchnikoff (1845-1916)

The origin of the concept of stress can be traced to Empedocles in the fifth century B.C.; however, Hans Selye, in the late 1930s, was the first person to use the term *stress* more or less as it is used today [1, 2]. While it is inherently difficult to define the term *stress*—in fact, many experts refuse to do so—the definition provided in *Webster's Medical Dictionary* ("a state of bodily or mental tension resulting from factors that tend to alter an existent equilibrium" [3]) is compatible with current usage by researchers in this field [2]. The factors that alter equilibrium, or the normal regulatory rhythms (homeostasis), are termed stressors and include a variety of psychological, environmental, and physiologic stimuli. The outcome of stress, i.e., regained homeostasis or distress (disease), is influenced by multiple variables, as depicted in figure 1.

The purpose of this review is to examine the evidence bearing on the question, Can stress operate as a cofactor in the pathogenesis of infectious diseases? Given the above defini-

tions, infectious disease agents themselves represent common environmental stressors, and the host response that they provoke can be regarded as stress. Thus the issue to be addressed in this review will be whether additional stressors can influence the outcome of infectious disease.

The topic of this review is timely since in the past decade major advances have been made in characterizing the bidirectional interaction between the brain and the immune system. During this period no fewer than eight books [4-11] and two new journals [12, 13] that are devoted to various aspects of the "brain-immune axis" have appeared. From this body of work, it has been firmly established that the CNS and the immune system are connected. A large number of studies have demonstrated that a variety of psychological stressors, e.g., bereavement, academic pressure, and loss of self-esteem, can result in laboratory evidence of immunologic impairment; the anatomic structures within the nervous system and the neuroendocrine pathways involved in the mediation of these effects have been elucidated (reviewed in [14-19]). A number of stress-responsive neuropeptides and neurotransmitters have been shown to interact with immune cells *in vitro*, and these molecules have been proposed as mediators of stress-induced immunosuppression (table 1). Some of these peptides as well as other neurohormones such as melatonin [46], also have been shown to have immunoenhancing or antistress effects. Communication of the immune system with the brain, on the other hand, has been shown to be mediated via a growing list of "immunotransmitters" (e.g., interleukin-1, interleukin-6, tumor necrosis factor- α , and interferons [14]) and may also involve several neuroendocrine hormones produced by the immune cells themselves [17].

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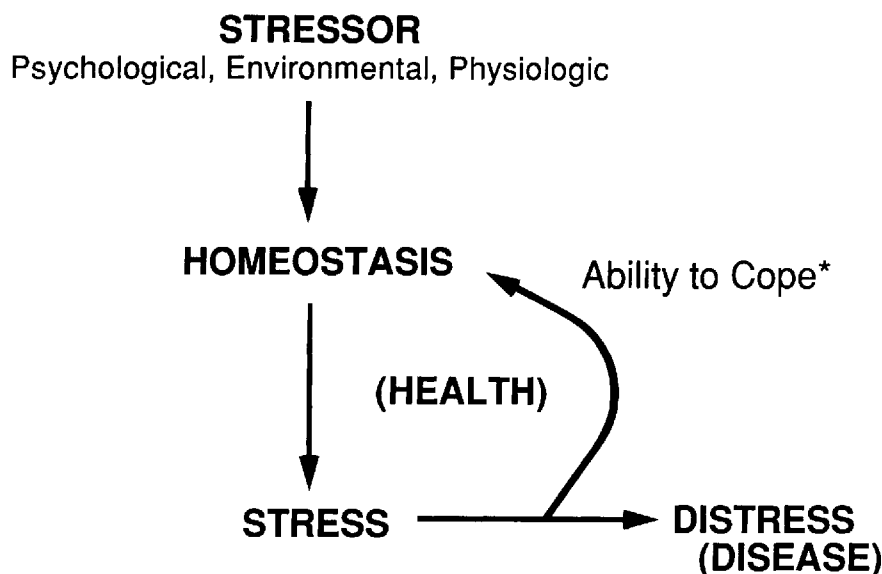


Figure 1. Stress is a state of altered homeostasis provoked by a psychological, environmental, or physiologic stressor.

* Influenced by intensity and duration of stress, age, state of health, previous experience with the stressor, etc.

The biologic significance of the findings in this area of research (psychoneuroimmunology) is less clear, however. The clinical importance of stress-related immunologic disturbance has been investigated in the fields of oncology, rheumatology, and allergy; however, investigations in the field of infectious diseases should yield especially relevant data given its large fund of knowledge regarding the impact of altered immune defenses on microbial pathogenesis. It is disappointing that there is a relative paucity of such data; in fact, most studies of stress and its pertinence to infectious diseases were performed before 1980. The scope of this review will be limited to the studies that have been carried out in mammals. While the use of animal models is distinctly advantageous in that it allows control of a number of important variables, as will be pointed out in this review, natural infections and naturally occurring stressors rarely have been selected for study in animals. In addition, the assessment of stress in animal models

has been limited and in most cases has been largely inferential. Although natural infections and naturally occurring stressors have been examined in humans, there has been a lack of uniformity in the instruments used to measure stress, and often infection has not been microbiologically verified.

Viral Infections

Animal Studies

The results of studies of the effect of stress on the pathogenesis of viral infection in animals are summarized in table 2. As shown, although a variety of viral pathogens have been studied, research in this area has been restricted primarily to rodent models. A number of stressors have been used: forced exercise (swimming or running in a motorized drum), avoidance learning (usually electric shock preceded by a warning stimulus), high-intensity sound, restraint, transportation, differential housing (isolation or crowding), and exposure to cold temperatures. Both the timing and the duration of exposure to the stressor relative to the initiation of infection have varied widely. The most commonly measured pathogenic effect has been altered survival of virus-infected animals.

As shown in table 2, mortality due to viral infection was found to be increased with almost all stressors studied. In a few instances, increased viral replication was noted, and in the cases of neurotropic and cardiotropic viruses, increased evidence of disease involving these organ systems was found. In only one study, of poliomyelitis virus in monkeys that were subjected to avoidance learning, did stress reduce mortality [49]. The fact that the same stressor increased the mortality

Table 1. Proposed mediators of stress-induced immunomodulation.

Mediator	[References]
Corticotropin-releasing factor, adrenocorticotropin, glucocorticoids	[20–23]
β-endorphin	[24–27]
Prolactin	[28–29]
Somatotropin	[30, 31]
Arginine vasopressin	[31, 32]
Norepinephrine, epinephrine	[33–37]
Enkephalin	[38, 39]
Substance P	[40–42]
Vasoactive intestinal peptide	[43–45]

Table 2. Animal models of stress and viral infection.

Virus	Animal	Stressor	Finding	[Reference]
Poliomyelitis	Mouse	Forced exercise	↑ Mortality, paralysis	[47]
	Monkey	Forced exercise	↑ Paralysis	[48]
	Monkey	Avoidance learning	↓ Mortality	[49]
Coxsackievirus B	Mouse	Avoidance learning	↑ Mortality, paralysis	[50]
	Mouse	Avoidance learning	↑ Weight loss, viral dissemination	[51]
	Mouse	Avoidance learning	↑ Mortality, weight loss	[52]
	Mouse	Forced exercise	↑ Mortality, carditis	[53–55]
Vesicular stomatitis	Mouse	Sound	↑ Mortality	[56]
	Mouse	Avoidance learning	↑ Virus in muscle	[57]
	Mouse	Sound	↑ Mortality, encephalitis	[58]
Herpes simplex	Mouse	Avoidance learning	↑ Mortality	[59]
	Mouse	Restraint	↑ Mortality	[59]
Bovine herpesvirus 1	Calves	Transportation	↑ Mortality, pneumonia	[60]
Influenza A	Mouse	Forced exercise	↑ Mortality	[61]
Encephalomyocarditis	Mouse	Isolation	↑ Mortality	[62]
Rabies	Guinea pig	Crowding	Viral reactivation	[63]
Coronavirus	Swine	Cold	↑ Gastroenteritis	[64]

NOTE. ↑ = increased; ↓ = decreased.

of mice infected with poliomyelitis virus [50] suggests that the effect of stress may be animal species-dependent. Friedman et al. [62] observed that the increased mortality due to encephalomyocarditis virus in CD-1 mice subjected to isolation-induced stress was less obvious in BALB/c mice, a finding that suggests strain-dependent or genetic factors may also be important. The potential contribution of animal age to mortality has not been carefully examined. In all of the studies with rodents, the animals used were 4–8 weeks of age, and those used in studies of primates were adults. The sex of the animals did not appear to influence the results in the few studies in which male and female mice were included [59, 62].

The influence of stress on the pathogenesis of herpes simplex virus has been investigated in murine and bovine models. Stress induced by avoidance learning and restraint increased mortality of mice infected with a herpes simplex virus isolate [59]. In a study reported by Filion et al. [60], stress arising from transportation and handling of 6–8-month-old calves fostered the development of pneumonia and death after challenge with bovine herpesvirus 1. Transported animals had elevated levels of plasma cortisol and a diminished blastogenic response to phytohemagglutinin *in vitro*. This study supports the long-held belief that stress from transportation plays a role in the development of bovine pneumonic pasteurellosis (shipping or transit fever). In the same study, the transported calves were found to have no greater susceptibility to *Pasteurella haemolytica*, which is capable of causing the disease as a primary pathogen [65]. However, infection with bovine herpesvirus 1 commonly precedes and may increase susceptibility to secondary bacterial infection.

Fluctuating ambient temperature has been evaluated as a naturally occurring stressor in a swine model of viral gastroenteritis [64]. Pigs maintained at an ambient temperature

of 30°C remained disease-free after challenge with the virulent coronavirus, a transmissible gastroenteritis-causing virus. However, a sudden drop in temperature to 4°C induced severe diarrhea in virus-challenged pigs. Also, pigs raised at temperatures that fluctuated between 20°C and 4°C developed profuse diarrhea when inoculated with this viral agent.

Forced exercise, which is a complex stressor involving both psychological and physiologic elements, has been used commonly in animal studies of infectious disease outcomes. Forced swimming [53, 54] and exhausting exercise of mice on a treadmill [55] have been associated with increased myocarditis after challenge with coxsackie B3 virus. The timing of initiation of exercise relative to the inoculation of virus appears to be crucial [55]. Although increased mortality of mice infected with the coxsackievirus B3 has been associated with swimming exercise [53], this was not the case in a more recent study in which exercise on a treadmill was used [55]. It is interesting that physical conditioning of mice prior to their inoculation with influenza A abrogated the deleterious effect of swimming exercise that was observed in mice that were unconditioned [61]. In a study of forced exercise, chilling, and mechanical trauma during the incubation period of infection due to poliomyelitis virus in monkeys, Levinson et al. [48] found that the incidence and severity of paralysis was greater in monkeys subjected to exhausting exercise.

Human Studies

Studies of the impact of stress on viral infections in humans have dealt with only a limited number of pathogens (table 3). The effect of physical exertion on the pathogenesis of viral infection in humans was most carefully studied in developed countries during the years of epidemic poliomyelitis. Analy-

Table 3. Stress and viral infections in humans.

Type of infectious disease	Pathogen	Subjects	Stressor	Finding	[Reference]
Poliomyelitis	Poliomyelitis virus	Patients	Physical activity	↑ Paralysis	[66]
Hepatitis	Hepatitis A	Patients	Physical activity	No effect	[67, 68]
Upper respiratory tract infection	Rhinovirus	Prisoners	Cold	No effect	[69]
	Rhinovirus	Volunteers	Life events*	↑ Frequency, severity	[70]
	NAI	Marine recruits	Delayed promotion	↑ Frequency	[71]
	NAI	College students	Life events	↑ Reports of symptoms	[72, 73]
	NAI†	Children	Life events	↑ Duration, severity	[74]
	NAI	Prisoners	Life events	↑ Reports of symptoms	[75]
	NAI†	Adults	Life events	↑ Frequency, duration	[76]
	NAI	Medical students	Exams	↑ Reports of symptoms	[77]
	NAI	Adults	Life events	↑ Reports of symptoms	[78]
Herpes labialis (recurrent)	HSV	Nursing students	Unhappiness	↑ Recurrences	[79, 80]
		Nursing students	Unhappiness	No effect	[81]
Herpes genitalis (recurrent)	HSV	Adults	Life events	No effect	[82]
Infectious mononucleosis	EBV	Cadets	Pressure to achieve academically	↑ Clinical illness	[83]

NOTE. ↑ = increased; NAI = naturally acquired infection; HSV = herpes simplex virus; EBV = Epstein-Barr virus.

* Disruptive or stressful life changes as measured by a variety of standardized questionnaires.

† Cultures were performed for viruses and other potential pathogens.

sis of the data from 411 cases of polio that occurred during three epidemics in the United States revealed that the incidence and severity of paralysis were increased only when physical activity was performed after the second or major phase of illness [66]. Moderate physical exertion during the period of recovery from acute viral hepatitis, on the other hand, has been shown to be nondetrimental [67, 68]. Although there are anecdotal reports of an increased incidence of upper respiratory tract infections in conditioned athletes during periods of exhaustive training [84], no controlled studies have been carried out in this group of subjects. Similarly, no data are available regarding the perceived beneficial effects of moderate exercise regimens on the pathogenesis of the common cold.

In contrast to the paucity of studies on the effects of exercise on the pathogenesis of upper respiratory tract infections, there have been numerous studies of the influence of psychological stressors (disruptive life events) on the course of such infectious disease. A variety of psychometric instruments (e.g., the Life Events Inventory, the Daily Hassles Scale, the Life Changes Inventory, and the Schedule of Recent Experience) have been used to assess stress resulting from disruptive life events, most commonly arising from situations of personal failure, loss of or separation from significant others, or a change in social status. In most studies [71–74] naturally acquired upper respiratory tract infections were evaluated, and data regarding frequency of symptomatic infections were usually obtained from subjects' self-reported information. Virologic confirmation of infection rarely has been attempted in such studies. In addition, although most authors have been aware of potential confounding variables such as sleep deprivation, altered nutrition, use of medications, and alcohol consumption, these factors nonetheless are often difficult to control and to measure accurately.

Despite important shortcomings in most of these studies, the results suggest that stressful life events promote the pathogenesis of upper respiratory tract infections. Supportive data were reported in a study in which volunteers were challenged with rhinoviruses at the MRC Common Cold Research Unit in Salisbury, United Kingdom [70]. Recent life stress significantly increased the magnitude of symptomatic infectious disease in these subjects, although personality traits seemed to be a strong determinant of increased susceptibility in that introverts developed worse symptoms and infections than did extroverts. Results of these viral challenge studies are consistent with earlier observations by Jackson et al. [85], who noted that students who were worried or concerned showed an increased susceptibility to upper respiratory tract infection after challenge with nasal secretions or viruses.

Psychological stressors have been regarded as common triggers of recurrent herpes labialis and herpes genitalis, although this belief has not been documented conclusively. In a series of studies carried out in student nurses, unhappiness (a factor that is measured in the Clyde Mood Scale) was found to be predictive of recurrent herpes labialis in some studies [79, 80] but not in others [81]. In a review of studies of recurrent genital herpes, VanderPlate and Aral [86] concluded that most studies lack proper controls and virologic confirmation of infections. In a recent report, Kemeny et al. [82] found no association between recurrent herpes genitalis and stressful life experiences, although depressive mood and alcohol consumption appeared to increase the risk of recurrences.

In a 4-year prospective study of cadets at the U.S. Military Academy at West Point, N.Y., who were seronegative for Epstein-Barr virus on admission to the study, Kasl et al. [83] found that seroconverters who developed symptomatic infectious mononucleosis had a high level of motivation (desire

to stay in the military), had achieved poor grades for academic performance in the year prior to developing mononucleosis, and were more likely to have fathers who were overachievers. Likewise, Glaser et al. [87] demonstrated that first-year medical students had increased levels of antibody to Epstein-Barr virus, herpes simplex virus, and cytomegalovirus during periods of examination stress. Furthermore, students who had high scores for loneliness on the UCLA Loneliness Scale had the highest titers of antibody to Epstein-Barr virus. Although those investigators did not provide any virologic or clinical correlations to the serologic findings, the results suggest that stress fosters reactivation of herpes viruses [88].

Although these data support the postulate that stress can be a cofactor in the pathogenesis of certain viral infections in humans, a number of limitations can be found in these studies. As has been mentioned, many studies have relied on patients' reports of illness, and since stressed individuals are more likely to seek treatment, the results may have been biased in a positive direction. Furthermore, studies that have yielded negative results may be less likely to be reported. It should also be pointed out that the outcome of infection may be influenced by personality traits and primary emotional disturbances, and these characteristics may be difficult to separate from stress produced by psychological stimuli. For example, obsessional symptoms [89] and a high need for power [90] have been associated with increased frequency or severity of upper respiratory tract infections, and delayed recovery from acute respiratory tract infections and influenza have been associated with various personality disturbances [91–93]. Similarly, personality-related effects on infections caused by

herpesviruses have been noted [94–96]. As is the case for studies with animals, the influence of age on the pathogenesis of infection in humans has rarely been evaluated (most studies have been carried out on young to middle-aged individuals), and the potential influence of sex has not been adequately explored. In one large survey, for example, the impact of divorce or separation on acute respiratory tract conditions was observed primarily in women [97].

Bacterial Infections

Animal Studies

It is not surprising that the earliest studies of the effects of stress on the pathogenesis of infectious diseases involved bacterial pathogens. The results of these seminal investigations and of subsequent studies are summarized in table 4. As early as 1890, forced exercise was shown to increase mortality in rats infected with *Bacillus anthracis* [98]. Subsequently, forced exercise in a revolving drum was found to increase mortality due to *Staphylococcus aureus* in rabbits, but this effect was observed only when exercise was initiated at the time of infection [102]. Although exercise before infection with *S. aureus* had no impact on mortality, rabbits were susceptible to *Streptococcus pyogenes* when they were exercised before being inoculated with this organism [102]. In studies with type I *Streptococcus pneumoniae*, forced exercise actually decreased mortality of rabbits when the exercise regimen was begun before bacterial challenge; however, an increased mortality was observed if exercise commenced at the time of inoculation with *S. pneumoniae* [101]. Similarly, mortality was reduced

Table 4. Animal models of stress and bacterial infection.

Bacterial pathogen	Animal	Stressor	Finding	[Reference]
<i>Bacillus anthracis</i>	Rat	Forced exercise	↑ Mortality	[98]
<i>Streptococcus pneumoniae</i>	Rat	Forced exercise	↓ Mortality	[99]
	Guinea pig	Forced exercise	↓ Mortality	[100]
<i>Streptococcus pyogenes</i>	Rabbit	Forced exercise	↓ or ↑ Mortality	[101]
	Rabbit	Forced exercise	↑ Mortality	[102]
<i>Staphylococcus aureus</i>	Rabbit	Forced exercise	↑ Mortality	[102]
	Mouse	Cold	↑ Mortality	[103]
<i>Salmonella typhimurium</i>	Mouse	Cold	↑ Mortality	[103, 104]
	Mouse	Crowding	↑ Mortality	[105]
	Pony	Transportation	↑ Gastroenteritis	[106]
<i>Francisella tularensis</i>	Mouse	Forced exercise	No effect	[61]
	Rat	Forced exercise	No effect	[107]
Lipopolysaccharide of <i>Escherichia coli</i>	Swine	Surgery, anesthesia	↑ Mortality	[108]
	Swine	Weaning, transportation	↓ Mortality	[108]
Viridans streptococci, gram-negative bacilli	Opossum	Captivity	Endocarditis, ↑ Mortality	[109]
<i>Mycobacterium tuberculosis</i>	Mouse	Crowding	↓ or ↑ Mortality	[110]
	Mouse, rat	Forced exercise	↑ Mortality	[111]

NOTE. ↑ = increased; ↓ = decreased.

in rats [99] and guinea pigs [100] when exercise preceded *S. pneumoniae* infection. In more recent studies, forced swimming exercise was found to have no effect on the pathogenesis of *Francisella tularensis* infection in mouse [61] and rat [107] models, although this same stressor significantly increased mortality in mice with influenza A [61]. These studies underscore the potential importance of the timing of the stressor and of the virulence factors of specific microorganisms in the determination of the influence of stress on the outcome of infectious disease.

The importance of bacterial virulence in determination of the influence of a stressor on pathogenesis was also observed in studies of mice that were exposed to cold temperatures and challenged with *S. aureus* [103] or *Salmonella typhimurium* [103, 105]. When mice were infected with highly virulent strains of *S. aureus* or *S. typhimurium*, the temperature at which the infected mice were maintained made no difference in the response to infection since all control animals died. However, when relatively avirulent strains were used, increased mortality was noted in mice that were exposed to environmental temperatures of 5°C and 15°C immediately after infection with these strains [103]. In addition, the stress of cold (5°C) markedly increased the susceptibility of mice to the lethal effect of lipopolysaccharide isolated from *Serratia marcescens* [103].

In other studies with *S. typhimurium*, Edwards and Dean [105] found that the stress of crowding (30–60 versus 2–10 animals per cage) increased the mortality of male and female mice inoculated intraperitoneally with this organism. Transportation-induced stress increased the susceptibility of ponies to development of gastroenteritis due to *S. typhimurium* [106], but it is interesting that this stressor appeared to protect piglets from endotoxic shock following intravenous administration of lipopolysaccharide from *Escherichia coli* [108]. Short hauling (12 hours' duration) but not long hauling (24 hours' duration) increased the morbidity and mortality of calves due to respiratory tract infection in cases in which *P. haemolytica* was a common isolate [112], a fact that suggests the calves were able to adapt to this stressor.

Unlike virtually all other animal species, opossums are susceptible to development of bacterial endocarditis without experimental alteration of the heart valve. Opossums taken into zoos or brought into the research laboratory are at risk of spontaneous death due to this infection. Sherwood et al. [109] have provided evidence that suggests stress arising from captivity plays an important role in the pathogenesis of this infection: in an undefined way, stress elicits valvular lesions that are a substrate for infective endocarditis.

Sex-specific influences of stress on outcome of infectious disease were observed in studies by Tobach and Bloch [110], who found that crowding of female mice before or at the time of infection with *Mycobacterium tuberculosis* reduced mortality. In contrast, crowding of male mice resulted in an increased death rate. In another study of experimental stress,

forced exercise in a metal wheel or in water (swimming) was associated with increased mortality of rodents infected with *M. tuberculosis* [111].

Human Studies

The possibility that “psychic trauma” might adversely influence the clinical course of human tuberculosis has been suspected clinically for many years [113]. In 1919, a researcher in Japan concluded: “Overtaxation of the mind of our youths by our unsatisfactory educational system seems to be the cause of the high mortality of young consumptives in our country” [114]. Death rates due to tuberculosis and other pulmonary infections in divorced persons have been reported to be greater than those in married individuals [115]. In other surveys, bereavement [116] and “chronic life stress” [117] have been regarded as possible predisposing factors in the development of serious respiratory tract infections such as pneumonia. Stress has also been considered as playing a role in the development of dental caries [118–120], although carefully controlled studies of this postulate are lacking.

In the only longitudinal study of the impact of stress on the pathogenesis of bacterial infection in humans, Meyer and Haggerty [121] found increased acquisition of and illness due to group A streptococci in members of families who experienced a variety of psychological stressors such as loss of a family member, other illness in the family, and divorce. While that study provides suggestive evidence that stress promotes the development of streptococcal pharyngitis, research in the field of bacterial pathogenesis in humans generally has not been as carefully controlled for potentially confounding variables as have studies of viral infections.

Parasitic Infections

Animal Studies

A limited number of studies that deal with the interaction of stress and parasitic infection have been reported (table 5). In a series of experiments using a murine model of malaria, Friedman et al. [122–125] demonstrated that a number of variables modulated the influence of stress on outcome of infectious disease. When housing in groups of five versus housing alone was selected as a stressor, the mice housed in groups had a significantly higher mortality after challenge with *Plasmodium berghei* [122–124]. Although sex of the mice had no effect on outcome, the findings were clearly dependent on mouse strain. While increased mortality was consistently observed in mice of the CD-1 strain, no effect was seen in mice of five other strains [124]. Stress induced by avoidance learning had the opposite effect of that induced by type of housing; i.e., mortality was reduced by subjecting mice to electric shock preceded by a warning light. However, avoidance learning promoted the lethality of coxsackie virus B-2 in the same model [123].

Table 5. Animal models of stress and parasitic infection.

Parasite	Animal	Stressor	Finding	[Reference]
<i>Plasmodium berghei</i>	Mouse	Housing in groups	↑ Mortality	[122–124]
	Mouse	Avoidance learning	↓ Mortality	[123, 125]
<i>Hymenolepis nana</i>	Mouse	Social	↑ Reinfection	[126]
	Mouse	Cat	↑ Reinfection	[127]
<i>Entamoeba</i> species	Squirrel	Cold	↑ Cecal organisms	[128]
<i>Trypanosoma cruzi</i>	Mouse	Forced exercise	↑ Mortality, myocarditis	[129]
<i>Toxoplasma gondii</i>	Rat	Cold, heat	↑ Lung disease	[130]
	Mouse	Restraint	↑ Mortality	[131]

NOTE. ↑ = increased; ↓ = decreased.

Two studies of murine gastrointestinal infection due to the dwarf tapeworm *Hymenolepis nana* have been reported, both of which used more naturally occurring stressors. Social stress (association with strange mice and severe fighting) of animals with acquired immunity to this parasite resulted in reinfection [126]. This effect was limited to mice with a low social status and was not observed in dominant males. An increased reinfection rate was also observed when predator (cat)-induced stress was used; in this study, adrenal hypertrophy and increased serum corticosteroid levels were used as markers of stress [127]. In a similar study, a cold environment was used as a natural stressor in ground squirrels. An increased number of naturally acquired *Entamoeba* organisms was found in the gastrointestinal tracts of cold-exposed animals [128]. Extremes of ambient temperature (10°C or 35°C) have been shown to promote pulmonary pathology in rats during acute infection with the RH strain of *Toxoplasma gondii* [130].

In a study in which forced swimming exercise was used as a stressor, Elson and Abelmann [129] demonstrated more-severe myocarditis and increased mortality in mice infected with *Trypanosoma cruzi*. Chao et al [131] recently reported that restraint-induced stress in BALB/c mice promoted the lethality of infection due to *T. gondii* C56, a virulent strain of this protozoan. However, those investigators were unable to detect any adverse effect of this stressor in mice inoculated with *T. gondii* Me49, a relatively avirulent strain. Thus, the strain of both the host and the pathogen appear to affect pathogenesis of infection.

Conclusions

A large body of evidence that has emerged in the past decade indicates that exposure to a variety of psychological, environmental, and physiologic stressors can result in alterations of immune cell functions [14–19]. Most studies have demonstrated that stress is associated with suppressed host-defense mechanisms, although in some instances enhanced immune reactivity has been observed. Similarly, the results of a majority of studies related to pathogenesis of infectious disease suggested that stress interferes with host defense. However, in a distinct minority of cases, stress appeared to enhance

host survival. Few studies have identified the mechanism(s) responsible for stress-induced alterations of microbial pathogenesis, although stress-responsive neuropeptides and catecholamines [132–133] have been implicated.

Although the accumulated data support the hypothesis that stress alters the pathogenesis of infectious disease, substantial limitations have existed in many of the studies with humans and in the animal models. To date, only a small number of viral and bacterial infections have been investigated in both human subjects and animal models. Studies of parasitic infections have been restricted to murine species, and no meaningful reports relating effects of stress on the outcome of fungal disease could be found. Of the variables that are known to influence the outcome of a host-parasite interaction, many have been shown to alter the impact of stress on the pathogenesis of infection; these factors include the microbial species and strain as well as the sex and immune status of the host and the type, timing, and duration of the stressor, relative to the onset of infection (figure 2). The potential importance of some variables, e.g., extremes of age, poor nutritional status, and the presence of concomitant diseases, has not yet been investigated. Given the well-known contribution of these factors to microbial pathogenesis, their influence should be explored. In addition, more studies need to be designed to analyze the potential interaction between the external stressor and personality, since the latter may determine the somatic response to the stressor.

The major advances witnessed in research of microbial pathogenesis coupled with the progress seen in the field of psychoneuroimmunology in recent years should permit greater insight into the cellular and molecular basis of stress-induced influences on the outcome of infectious diseases. Now that it has become abundantly clear that the brain and immune system communicate, it is time for more active interchange between those most familiar with the complexities of the brain—neuroscientists, psychiatrists, psychologists—and investigators of infectious diseases. Because human studies are fraught with variables that are difficult to control, e.g., nutritional factors and the use of medications or substances that might alter host defenses, animal models remain essential. Mounting concerns regarding the ethics of animal experimen-

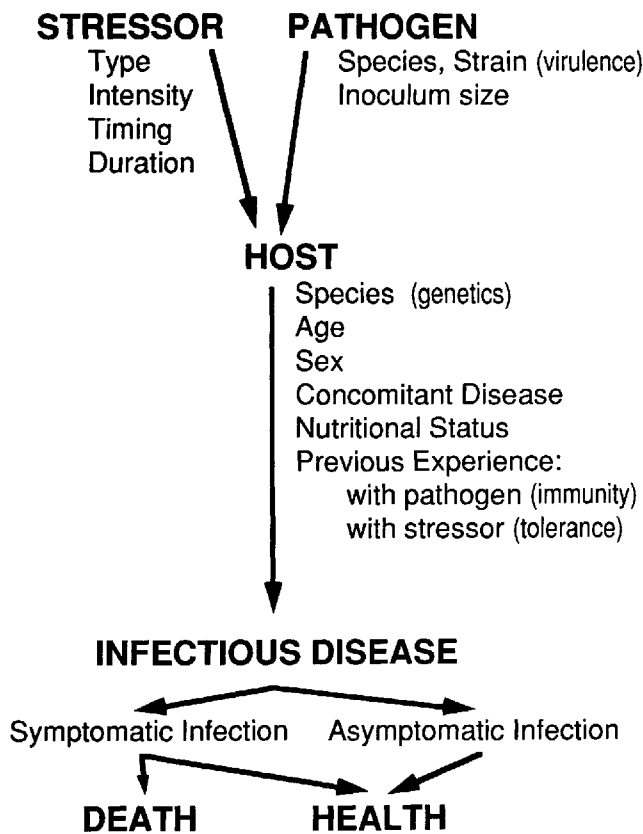


Figure 2. Various factors can modify the impact of a stressor on the pathogenesis of an infectious disease.

tation, however, support the need for greater use of naturally occurring stressors in such research, as well as the use of pathogenic endpoints other than mortality. Since stress appears to have an adverse impact on livestock by promoting the development of infections, these animals represent a particularly relevant model for investigation. Stress-associated immune suppression has been considered as a potential cofactor in the pathogenesis of human immunodeficiency virus infection [134]. Given increased research funding for all aspects of the acquired immunodeficiency syndrome, this is an area of human medicine that is likely to receive increased attention. The question regarding whether stress plays a role in the pathogenesis of postsurgical infections or other types of nosocomial infections might also prove worthy of investigation; certainly, the hospital is a stressful environment [135].

The fact that the human condition is characterized by stress—yet illness due to infectious disease is a relatively rare occurrence in healthy individuals—suggests that successful coping mechanisms are the rule. As pointed out by Hans Selye over 30 years ago, new approaches to prevention and therapy of infectious disease may come from increased understanding of these mechanisms: “If I may venture a prediction, I would like to reiterate my opinion that research on stress will

be most fruitful if it is guided by the principle that we must learn to imitate—and if necessary to correct and complement—the body’s own autopharmacologic efforts to combat the stress factor in disease” [136].

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References

- Mason JW. A historical view of the stress field. *J Human Stress* 1975;1:6–12
- Chrousos GP, Loriaux DL, Gold PW. The concept of stress and its historical development. *Adv Exp Med Biol* 1988;245:3–7
- Webster’s medical desk dictionary. Springfield, MA: Merriam-Webster, 1986
- Ader R, ed. *Psychoneuroimmunology*. New York: Academic Press, 1981
- Cooper EL, ed. *Stress, immunity, and aging*. New York: Marcel Dekker, 1984
- Corson SA, Corson EO, Dartau R, Epp J, Mutschler LA, eds. *Neurohumoral maintenance of immune homeostasis*. Chicago: University of Chicago Press, 1985
- Neruzzi D, Goodwin FK, Costa E, eds. *Hypothalamic dysfunction in neuropsychiatric disorders*. *Adv Biochem Psychopharmacol* 1986;43
- Cotman CW, Brinton RE, Galaburda A, McEwen B, eds. *The neuro-immune endocrine connection*. New York: Raven Press, 1987
- Neuroimmune interactions: proceedings of the second international workshop on neuroimmunomodulation. *Ann NY Acad Sci* 1987;496: 1–756
- Raine CS, ed. *Advances in neuroimmunology*. *Ann N Y Acad Sci* 1988;540
- Goetzl EJ, Spector NH, eds. *Neuroimmune networks: physiology and diseases*. New York: Alan R. Liss, 1989
- Ader R, Cohen H, Felten D, eds. *Brain, behavior and immunity* [journal]. San Diego: Academic Press
- MacLeod RM, Blalock JE, Martin JB, Scapagnini U, eds. *Progress in neuroendocrinimmunology* [journal]. Washington, DC: Fidia Information Network
- Dantzer R, Kelley KW. Stress and immunity: an integrated view of relationships between the brain and the immune system. *Life Sci* 1989;44:1995–2008
- Griffin JFT. Stress and immunity: a unifying concept. *Vet Immunol Immunopathol* 1989;20:263–312
- Rabin BS, Cohen S, Ganguli R, Lysle DT, Cunnick JE. Bidirectional interaction between the central nervous system and the immune system. *CRC Crit Rev Immunol* 1989;9:279–312
- Brown SL, Blalock JE. Neuroendocrine immune interactions. In: Oppenheim JJ, Shevach EM, eds. *Immunophysiology: the role of cells and cytokines in immunity and inflammation*. New York: Oxford University Press, 1990:306–19
- Bonneau RH, Kiecolt-Glaser JK, Glaser R. Stress-induced modulation of the immune response. *Ann N Y Acad Sci* 1990;594:253–69
- Khansari DN, Murgu AJ, Faith RE. Effects of stress on the immune system. *Immunol Today* 1990;11:170–5
- Irwin MR, Vale W, Britton KT. Central corticotropin-releasing factor suppresses natural killer cytotoxicity. *Brain Behav Immun* 1987;1:81–7
- Johnson HM, Torres BA, Smith EM, Dion LD, Blalock JE. Regulation of lymphokine (γ -interferon) production by corticotropin. *J Immunol* 1984;132:246–50
- Smith EM, Brosnan P, Meyer WJ, Blalock JE. An ACTH receptor

- on human mononuclear leukocytes: relation to adrenal ACTH-receptor activity. *N Engl J Med* **1987**;317:1266-9
23. Cupps TR, Fauci AS. Corticosteroid-mediated immunoregulation in man. *Immunol Rev* **1982**;65:133-55
 24. Kay N, Allen J, Morley JE. Endorphins stimulate normal human peripheral blood lymphocyte natural killer activity. *Life Sci* **1984**;35:53-9
 25. Heijnen CJ, Croiset G, Zijlstra J, Ballieux RE. Modulation of lymphocyte function by endorphins. *Ann N Y Acad Sci* **1987**;496:161-5
 26. Peterson PK, Sharp B, Gekker G, Brummitt C, Keane WF. Opioid-mediated suppression of interferon- γ production by cultured peripheral blood mononuclear cells. *J Clin Invest* **1987**;80:824-31
 27. Peterson PK, Sharp B, Gekker G, Brummitt C, Keane WF. Opioid-mediated suppression of cultured peripheral blood mononuclear cell respiratory burst activity. *J Immunol* **1987**;138:3907-12
 28. Russell DH, Kibler R, Matrisian L, Larson DF, Poulos B, Magun BE. Prolactin receptors on human T and B lymphocytes: antagonism of prolactin binding by cyclosporine. *J Immunol* **1985**;134:3027-31
 29. Bernton EW, Meltzer MS, Holaday JW. Suppression of macrophage activation and T-lymphocyte function in hypoprolactinemic mice. *Science* **1988**;239:401-4
 30. Edwards CK 3rd, Schepper JM, Yungler LM, Kelley KW. Somatotropin and prolactin enhance respiratory burst activity of macrophages. *Ann N Y Acad Sci* **1988**;540:698-9
 31. Edwards CK, Ghiasuddin SM, Schepper JM, Yungler LM, Kelley KW. A newly defined property of somatotropin: priming of macrophages for production of superoxide anion. *Science* **1988**;239:769-71
 32. Johnson HM, Torres BA. Regulation of lymphokine production by arginine vasopressin and oxytocin: modulation of lymphocyte function by neurohypophyseal hormones. *J Immunol* **1985**;135(Suppl):773-5
 33. Torres BA, Johnson HM. Arginine vasopressin (AVP) replacement of helper cell requirement in IFN- γ production: evidence for a novel AVP receptor on mouse lymphocytes. *J Immunol* **1988**;140:2179-83
 34. Sanders VM, Munson AE. Norepinephrine and the antibody response. *Pharmacol Rev* **1985**;37:229-48
 35. Koff WC, Dunegan MA. Neuroendocrine hormones suppress macrophage-mediated lysis of herpes simplex virus-infected cells. *J Immunol* **1986**;136:705-9
 36. Felten DL, Felten SY, Bellinger DL, Carlson SL, Ackerman KD, Madden KS, Olschowki JA, Livnat S. Noradrenergic sympathetic neural interactions with the immune system: structure and function. *Immunol Rev* **1987**;100:225-60
 37. Felten DL, Felten SY. Sympathetic noradrenergic innervation of immune organs. *Brain Behav Immun* **1988**;2:293-300
 38. Wybran J. Enkephalins and endorphins as modifiers of the immune system: present and future. *Fed Proc* **1985**;44:92-4
 39. Jankovic BD, Maric D. Enkephalins modulate *in vivo* immune reactions through delta- and mu-opioid receptors. *Ann NY Acad Sci* **1988**;540:691-3
 40. Payan DG, Goetzl EJ. Modulation of lymphocyte function by sensory neuropeptides. *J Immunol* **1985**;135(Suppl 2):783-6
 41. Serra MC, Bazzoni F, Bianca VD, Greskowiak M, Rossi F. Activation of human neutrophils by substance P: effect on oxidative metabolism, exocytosis, cytosolic Ca²⁺ concentration and inositol phosphate formation. *J Immunol* **1988**;141:2118-24
 42. Lotz M, Vaughan JH, Carson DA. Effect of neuropeptides on production of inflammatory cytokines by human monocytes. *Science* **1988**;241:1218-21
 43. O'Dorisio MS, Wood CL, O'Dorisio TM. Vasoactive intestinal peptide and neuropeptide modulation of the immune response. *J Immunol* **1985**;135(Suppl 2):792-6
 44. O'Dorisio MS. Neuropeptides and gastrointestinal immunity. *Am J Med* **1986**;81(Suppl 6B):74-82
 45. O'Dorisio MS, Shannon BT, Fleshman DJ, Campolito LB. Identification of high affinity receptors for vasoactive intestinal peptide on human lymphocytes of B cell lineage. *J Immunol* **1989**;142:3533-6
 46. Maestroni GJM, Conti A. Beta-endorphin and dynorphin mimic the circadian immunoenhancing and anti-stress effects of melatonin. *Int J Immunopharmacol* **1989**;11:333-40
 47. Rosenbaum HE, Harford CG. Effect of fatigue on susceptibility of mice to poliomyelitis. *Proc Soc Exp Biol Med* **1953**;83:678-81
 48. Levinson SO, Milzer A, Lewin P. Effect of fatigue, chilling and mechanical trauma on resistance to experimental poliomyelitis. *American Journal of Hygiene* **1945**;42:204-13
 49. Marsh JT, Lavender JF, Chang S-S, Rasmussen AF. Poliomyelitis in monkeys: decreased susceptibility after avoidance stress. *Science* **1963**;140:1414-5
 50. Johnsson T, Rasmussen AF Jr. Emotional stress and susceptibility to poliomyelitis virus infection in mice. *Arch Gesamte Virusforsch* **1965**;17:392-7
 51. Johnsson T, Lavender JF, Hultin E, Rasmussen AF Jr. The influence of avoidance-learning stress on resistance to Coxsackie B virus in mice. *J Immunol* **1963**;91:569-75
 52. Friedman SB, Ader R, Glasgow LA. Effects of psychological stress in adult mice inoculated with Coxsackie B viruses. *Psychosom Med* **1965**;27:361-8
 53. Gatmaitan BG, Chason JL, Lerner AM. Augmentation of the virulence of murine Coxsackievirus B-3 myocardopathy by exercise. *J Exp Med* **1970**;131:1121-36
 54. Reyes MP, Lerner AM. Interferon and neutralizing antibody in sera of exercised mice with coxsackievirus B-3 myocarditis. *Proc Soc Exp Biol Med* **1976**;151:333-8
 55. Ilbäck N-G, Fohlman J, Friman G. Exercise in coxsackie B3 myocarditis: effects on heart lymphocyte subpopulations and the inflammatory reaction. *Am Heart J* **1989**;177:1298-1302
 56. Jensen MM, Rasmussen AF. Stress and susceptibility to viral infections. II. Sound stress and susceptibility to vesicular stomatitis virus. *J Immunol* **1963**;90:21-3
 57. Yamada A, Jensen MM, Rasmussen AF Jr. Stress and susceptibility to viral infections. III. Antibody response and viral retention during avoidance learning stress. *Proc Soc Exp Biol Med* **1986**;116:677-80
 58. Chang S-S, Rasmussen AF Jr. Stress-induced suppression of interferon production in virus-infected mice. *Nature* **1965**;205:623-4
 59. Rasmussen AF Jr, Marsh JT, Brill NQ. Increased susceptibility to herpes simplex in mice subjected to avoidance-learning stress or restraint. *Proc Soc Exp Biol Med* **1957**;96:183-9
 60. Fillion LG, Willson PJ, Bielefeldt-Ohmann H, Babiuk LA, Thomson RG. The possible role of stress in the induction of pneumonic pasteurellosis. *Can J Comp Med* **1984**;48:268-74
 61. Ilbäck N-G, Friman G, Beisel WR, Johnson AJ, Berendt RF. Modifying effects of exercise on clinical course and biochemical response of the myocardium in influenza and tularemia in mice. *Infect Immun* **1984**;45:498-504
 62. Friedman SB, Glasgow LA, Ader R. Differential susceptibility to a viral agent in mice housed alone or in groups. *Psychosom Med* **1970**;32:285-99
 63. Soave OA. Reactivation of rabies virus in a guinea pig due to the stress of crowding. *Am J Vet Res* **1964**;25:268-9
 64. Shimizu M, Shimizu Y, Kodama Y. Effects of ambient temperatures on induction of transmissible gastroenteritis in feeder pigs. *Infect Immun* **1978**;21:747-52
 65. Shoo MK. Experimental bovine pneumonic pasteurellosis: a review. *Vet Rec* **1989**;124:141-4
 66. Horstmann DM. Acute poliomyelitis: relation of physical activity at the time of onset to the course of the disease. *JAMA* **1950**;142:236-41
 67. Chalmers TC, Eckhardt RD, Reynolds WE, Cigarroa JG, Deane N,

- Reifenstein RW, Smith CW, Davidson CS. The treatment of acute infectious hepatitis. Controlled studies of the effects of diet, rest, and physical reconditioning on the acute course of the disease and on the incidence of relapses and residual abnormalities. *J Clin Invest* 1955;34:1163–235
68. Edlund A. The effect of defined physical exercise in the early convalescence of viral hepatitis. *Scand J Infect Dis* 1971;3:189–96
69. Douglas RG Jr, Lindgren KM, Couch RB. Exposure to cold environment and rhinovirus common cold: failure to demonstrate effect. *N Engl J Med* 1968;279:742–7
70. Totman R, Kiff J, Reed SE, Craig JW. Predicting experimental colds in volunteers from different measures of recent life stress. *J Psychosom Res* 1980;24:155–63
71. Voors AW, Stewart GT, Gutekunst RR, Moldow CF, Jenkins CD. Respiratory infection in marine recruits: influence of personal characteristics. *Am Rev Respir Dis* 1968;98:801–9
72. Jacobs MA, Spilken A, Norman M. Relationship of life change, maladaptive aggression, and upper respiratory infection in male college students. *Psychosom Med* 1969;31:31–44
73. Jacobs MA, Spilken AZ, Norman MM, Anderson LS. Life stress and respiratory illness. *Psychosom Med* 1970;32:233–42
74. Boyce WT, Jensen EW, Cassel JC, Collier AM, Smith AH, Ramey CT. Influence of life events and family routines on childhood respiratory tract illness. *Pediatrics* 1977;60:609–15
75. McClelland DC, Alexander C, Marks E. The need for power, stress, immune function, and illness among male prisoners. *J Abnorm Psychol* 1982;91:61–70
76. Graham NMH, Douglas RM, Ryan P. Stress and acute respiratory infection. *Am J Epidemiol* 1986;124:389–401
77. Glaser R, Rice J, Sheridan J, Fertel R, Stout J, Speicher C, Pinsky D, Kotur M, Post A, Beck M, Keicolt-Glaser J. Stress-related immune suppression: health implications. *Brain Behav Immun* 1987;1:7–20
78. Levy SM, Fernstrom J, Herberman RB, Whiteside T, Lee J, Ward M, Massoudi M. Persistently low natural killer cell activity and circulating levels of plasma beta endorphin: risk factors for infectious disease. *Life Sci* 1991;48:107–16
79. Katcher AH, Brightman V, Luborsky L, Ship I. Prediction of the incidence of recurrent herpes labialis and systemic illness from psychological measurements. *J Dent Res* 1973;52:49–58
80. Friedmann E, Katcher AH, Brightman VI. Incidence of recurrent herpes labialis and upper respiratory infection: a prospective study of the influence of biologic, social, and psychologic predictors. *Oral Medicine* 1977;43:873–8
81. Luborsky L, Mintz J, Brightman VJ, Katcher AH. Herpes simplex virus and moods: a longitudinal study. *J Psychosom Res* 1976;20:543–8
82. Kemeny ME, Cohen F, Zegans LS, Conant MA. Psychological and immunological predictors of genital herpes recurrence. *Psychosom Med* 1989;51:195–208
83. Kasl SV, Evans AS, Niederman JC. Psychosocial risk factors in the development of infectious mononucleosis. *Psychosom Med* 1979;41:445–66
84. Douglas DJ, Hanson PG. Upper respiratory infections in the conditioned athlete [abstract]. *Med Sci Sports* 1978;10:55
85. Jackson GG, Dowling HF, Anderson TO, Riff L, Saporta J, Turck M. Susceptibility and immunity to common upper respiratory viral infection—the common cold. *Ann Intern Med* 1960;53:719–38
86. VanderPlate C, Aral SO. Psychosocial aspects of genital herpes virus infection. *Health Psychol* 1987;6:57–72
87. Glaser R, Kieicolt-Glaser JK, Speicher CE, Holliday JE. Stress, loneliness, and changes in herpesvirus latency. *J Behav Med* 1985;8:249–60
88. Kieicolt-Glaser JK, Glaser R. Psychosocial influences on herpesvirus latency. In: Kurstak E, Lipowski ZJ, Morozov PV, eds. *Viruses, immunity, and mental disorders*. New York: Plenum Medical Book, 1987:403–11
89. Broadbent DE, Broadbent MHP, Phillipotts RJ, Wallace J. Some further studies on the prediction of experimental colds in volunteers by psychological factors. *J Psychosom Res* 1984;28:511–23
90. McClelland DC, Floor E, Davidson RJ, Saron C. Stressed power motivation, sympathetic activation, immune function, and illness. *J Human Stress* 1980;6:11–9
91. Brodman K, Mittelman B, Wechsler D, Weider A, Wolff HG. The relation of personality disturbances to duration of convalescence from acute respiratory infections. *Psychosom Med* 1947;9:37–44
92. Imboden JB, Canter A, Cluff LE. Convalescence from influenza: a study of the psychological and clinical determinants. *Arch Intern Med* 1961;108:393–9
93. Cluff LE, Canter A, Imboden JB. Asian influenza: infection, disease, and psychological factors. *Arch Intern Med* 1966;117:159–63
94. Goldmeier D, Johnson A. Does psychiatric illness affect the recurrence rate of genital herpes? *Br J Vener Dis* 1982;58:40–3
95. Silver PS, Auerbach SM, Vishniavsky N, Kaplowitz LG. Psychological factors in recurrent genital herpes infection: stress, coping style, social support, emotional dysfunction, and symptom recurrence. *J Psychosom Res* 1986;30:163–71
96. Greenfield NS, Roessler R, Crosley AP Jr. Ego strength and length of recovery from infectious mononucleosis. *J Nerv Ment Dis* 1959;125–8
97. Verbrugge LM. Marital status and health. *Journal of Marriage and the Family* 1979;41:267–85
98. Charrin, R. Contribution a l'etude experimentale du surmenage; son influence sur l'infection. *Archives de Physiologie Normale et Pathologique* 1890;2:273–83
99. Oppenheimer EH, Spaeth RA. The relation between fatigue and the susceptibility of rats towards a toxin and an infection. *American Journal of Hygiene* 1922;2:51–66
100. Nicholls EE, Spaeth RA. The relation between fatigue and the susceptibility of guinea pigs to infections of type I pneumococcus. *American Journal of Hygiene* 1922;2:527–35
101. Bailey GH. The effect of fatigue upon the susceptibility of rabbits to intratracheal injections of type I pneumococcus. *American Journal of Hygiene* 1925;5:175–85
102. Abbott AC, Gildersleeve N. The influence of muscular fatigue and of alcohol upon certain of the normal defenses. *University of Pennsylvania Medical Bulletin* 1910;23:169–81
103. Previte JJ, Berry LJ. The effect of environmental temperature on the host-parasite relationship in mice. *J Infect Dis* 1962;110:201–9
104. Miraglia GJ, Berry LJ. Enhancement of salmonellosis and emergence of secondary infection in mice exposed to cold. *J Bacteriol* 1962;84:1173–80
105. Edwards EA, Dean LM. Effects of crowding of mice on humoral antibody formation and protection to lethal antigenic challenge. *Psychosom Med* 1977;39:19–24
106. Owen RA, Fullerton J, Barnum DA. Effects of transportation, surgery, and antibiotic therapy in ponies infected with *Salmonella*. *Am J Vet Res* 1983;44:46–50
107. Friman G, Ilbäck N-G, Beisel WR, Crawford DJ. The effects of strenuous exercise on infection with *Francisella tularensis* in rats. *J Infect Dis* 1982;145:706–14
108. Osborne JC, Meredith JH. The influence of environmental and surgical stressors on susceptibility to bacterial endotoxin. *Exp Med Surg* 1970;28:39–44
109. Sherwood BF, Rowlands DT Jr, Hackel DB, LeMay JC. Bacterial endocarditis, glomerulonephritis, and amyloidosis in the opossum (*Didelphis virginiana*). *Am J Pathol* 1968;53:115–26
110. Tobach E, Bloch H. Effect of stress by crowding prior to and following tuberculous infection. *Am J Physiol* 1956;187:399–402

111. Kreis B, Hirsch A. Effort et tuberculose (étude expérimentale chez la souris et le rat). *Rev Tuberc (Paris)* 1965;29:421–30
112. Cole NA, Camp TH, Rowe LD Jr, Stevens DG, Hutcheson DP. Effect of transport on feeder calves. *Am J Vet Res* 1988;49:178–83
113. Wittkower ED. Psychological aspects of pulmonary tuberculosis. In: Sparer PJ, ed. *Personality, stress and tuberculosis*. New York: International Universities Press, 1956;153:174
114. Ishigami T. The influence of psychic acts on the progress of pulmonary tuberculosis. *American Review of Tuberculosis* 1919;2:470–84
115. Somers AR. Marital status, health, and use of health services: an old relationship revisited. *JAMA* 1979;241:1818–22
116. Parkes CM, Brown RJ. Health and bereavement: a controlled study of young Boston widows and widowers. *Psychosom Med* 1972;34:449–61
117. Hinkle LE Jr, Plummer N. Life stress and industrial absenteeism: the concentration of illness and absenteeism in one segment of a working population. *The Journal of Medicine in Industry* 1952;21:363–75
118. Manhold JH, Doyle JL, Weisinger EH. Effects of social stress on oral and other bodily tissue. II. Results offering substance to a hypothesis for the mechanism of formation of periodontal pathology. *J Periodontol* 1971;42:109–11
119. Manhold JH. Stress, oral disease, and general illness. *Psychosomatics* 1979;20:83–7
120. MacGregor AJ. *The impacted lower wisdom tooth*. New York: Oxford University Press, 1985;22–3
121. Meyer RJ, Haggerty RJ. Streptococcal infections in families: factors altering susceptibility. *Pediatrics* 1962;29:539–49
122. Plaut SM, Ader R, Friedman SB, Ritterson AL. Social factors and resistance to malaria in the mouse: effects of group vs. individual housing on resistance to *Plasmodium berghei* infection. *Psychosom Med* 1969;31:536–52
123. Friedman SB, Glasgow LA, Ader R. Psychosocial factors modifying host resistance to experimental infections. *Ann N Y Acad Sci* 1969;164:381–93
124. Friedman SB, Glasgow LA. Interaction of mouse strain and differential housing upon resistance to *Plasmodium berghei*. *J Parasitol* 1973;59:851–4
125. Friedman SB, Ader R, Grota LJ. Protective effect of noxious stimulation in mice infected with rodent malaria. *Psychosom Med* 1973;35:535–7
126. Weinmann CJ, Rothman AH. Effects of stress upon acquired immunity to the dwarf tapeworm, *Hymenolepis nana*. *Exp Parasitol* 1967;21:61–7
127. Hamilton DR. Immunosuppressive effects of predator induced stress in mice with acquired immunity to *Hymenolepis nana*. *J Psychosom Res* 1974;18:143–53
128. Noble GA. Stress and parasitism. IV. Cold stress and *Entamoeba*. *Exp Parasitol* 1966;19:264–8
129. Elson SH, Abelmann WH. Effects of muscular activity upon the acute myocarditis of C3H mice infected with *Trypanosoma cruzi*. *Am Heart J* 1965;69:629–36
130. El-Fakahany AF, El-Ridi AM, Marii NE. Experimental toxoplasmosis: effect of ambient temperature on lungs. *J Egypt Soc Parasitol* 1988;18:193–6
131. Chao CC, Peterson PK, Filice GA, Pomeroy C, Sharp BM. Effects of immobilization stress on the pathogenesis of acute murine toxoplasmosis. *Brain Behav Immun* 1990;4:162–9
132. Gruchow HW. Catecholamine activity and infectious disease episodes. *J Human Stress* 1979;5:11–7
133. Mason JW, Buescher EL, Belfer ML, Artenstein MS, Mougey EH. A prospective study of corticosteroid and catecholamine levels in relation to viral respiratory illness. *J Human Stress* 1979;5:18–28
134. Glaser R, Kiecolt-Glaser J. Stress-associated immune suppression and acquired immune deficiency syndrome (AIDS). *Adv Biochem Psychopharmacol* 1988;44:203–15
135. Bedell SE, Cleary PD, Delbanco TL. The kindly stress of hospitalization. *Am J Med* 1984;77:592–6
136. Selye H. Stress and disease. *Science* 1955;122:625–31