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Stressin' and slicin': stress-induced tRNA fragmentation codon-adapts translation to repress cell growth

N[i](https://orcid.org/0000-0002-3898-8064)cola Guzzi **D** & Cristian Bellodi^{[*](https://orcid.org/0000-0001-8047-5329)} D

Transfer RNAs (tRNAs) are central adaptors that decode genetic information during translation and have been long considered static cellular components. However, whether dynamic changes in tRNAs and tRNA-derived fragments actively contribute to gene regulation remains debated. In this issue, Huh et al (2020) highlight tyrosine tRNA $_{\text{GUA}}$ fragmentation at the nexus of an evolutionarily conserved adaptive codonbased stress response that fine-tunes translation to restrain growth in human cells.

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rotein synthesis is a fundamental process that relies on tRNAs for decoding the genetic information in mRNAs. Their central role as universal adaptor molecules in translation contributed to the prevailing notion that tRNAs are static cellular components with limited or no regulatory function. This view is changing due to increasing evidence that tRNA abundance and modifications are rewired to direct cellular translatomes during physiological and stress conditions in different types of normal and malignant cells (Gingold et al, 2014; Goodarzi et al, 2016; Chou et al, 2017). A renewed interest in tRNA biology was further sparked by the discovery of evolutionarily conserved stable tRNA-derived fragments (tRFs) with pleiotropic roles in gene expression control across different cell types and upon stress in organisms from all kingdoms of life (Oberbauer & Schaefer, 2018). Findings that only a small portion of tRNA is cleaved suggested that the functional effects of tRFs on gene expression are mostly uncoupled from the translational impairments caused by the depletion of precursor or mature tRNAs (Thompson et al, 2008). However, an outstanding question is to what extent tRNA fragmentation coordinates the tRNA and tRF cellular pools at the codon level in order to molecularly adapt the stress response.

In this issue of The EMBO journal, Huh et al (2020) set out to address this question in human epithelial cells undergoing oxidative stress, which was known to induce robust tRF accumulation in multiple cell systems (Thompson et al, 2008). The authors employed a highly sensitive probe-based sequencing approach to uncover reductions within a limited subset of tRNAs following stress. Depletion of these tRNAs overlapped, at least in part, with the generation of isodecoder-specific fragments, highlighting tyrosine $tRNA_{GUA}$ ($tRNA_{Tyr}_{GUA}$) fragmentation as one of the most striking oxidativestress-induced event. In an attempt to delineate the consequences of $tRNA^{Tyr}_{GUA}$ depletion, authors performed elegant lossof-function studies and revealed specific translation impairments in distinct tyrosine codon-enriched mRNAs encoding metabolic and growth-promoting factors. These defects were consistent with the cytostatic effects observed following oxidative stress and $tRNA^{Tyr}_{GUA}$ downregulation. Findings that ribosome stalling was increased on tyrosine codons upon depletion of $tRNA^{Tyr}_{GUA}$ strongly support that the availability of this cognate tRNA needs to meet the demand imposed by the mRNA codon usage during stress. This is concordant with previous reports illustrating a direct correspondence between tRNA abundance and codon usage that selectively impacts ribosome elongation rates and gene expression in response to stress and various biological cues (Gingold et al, 2014; Goodarzi et al, 2016).

Emerging evidence further suggests that reprogramming of tRNA modifications may provide an additional layer of translation control that impacts codon optimality in cells exposed to different environmental signals (Chan et al, 2012; Chou et al, 2017). Previous work by the Dedon laboratory demonstrated that increased levels of Trm4 methyltransferase-induced 5-methylcytosine (m5C) modification at the wobble position of $tRNA^{Leu}_{CAA}$ promoted selective translation of UUG codonenriched mRNAs involved in survival upon oxidative stress in yeast (Chan et al, 2012). Additional research indicates that the expression of tRNA modifying enzymes may be regulated and influence tRNA fragmentation across different types of normal and malignant cells and following stress (Blanco et al, 2016; Guzzi et al, 2018). These results raise a key question: do epitranscriptomic modifications impact processing and function of $tRNA^{Tyr}_{GUA}$ and other stress-sensitive isoacceptor tRNAs identified by Huh and colleagues (2020)? This may provide further mechanistic explanations for differences in selectivity of tRNA modulation observed between mammalian cells and unicellular organisms such as yeast and bacteria, which display profound rearrangements of their tRNA pools upon exposure to oxidative stress (Torrent et al, 2018).

One of most prominent mechanism of tRF generation relies on angiogenin (ANG),

Division of Molecular Hematology, Department of Laboratory Medicine, Lund Stem Cell Center, Faculty of Medicine, Lund University, Lund, Sweden *Corresponding author. E-mail: cristian.bellodi@med.lu.se

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Figure 1. Stress-induced tyrosine tRNA fragmentation controls cell growth.

Schematic model depicts the adaptive codon-based stress response governed by tyrosine tRNA^{Tyr}_{GUA} fragmentation in human cells. Through an unknown mechanism, oxidative stress induces rapid depletion of pre-tRNA^{Tyr}_{GUA} and accumulation of tRNA fragments (tRF^{Tyr}_{GUA}) in an exoribonuclease DIS3L2-dependent manner. Left, tRFTyr_{GUA} competes for hnRNPA1 binding with specific growth-associated mRNA subsets promoting their destabilization. Right, pre-tRNA Tyr _{GUA} fragmentation subsequently depletes mature $tRNA^{Tyr}_{GUA}$, thereby hampering the translation of growth-related mRNAs (e.g., USP3, EPCAM, and SCD) that are enriched in tyrosine codons. The synergistic effects on the transcriptome and translatome induced by the accumulation of tRF^{Tyr}_{GUA} and depletion of tRNA^{Tyr}_{GUA} drive an evolutionarily conserved program that limits cell growth upon oxidative stress.

a member of the RNase A superfamily, which is activated under stress to cleave multiple mature cytoplasmic tRNAs into halves (Ivanov et al, 2011). Focusing on the accumulation of tRF^{Tyr} _{GUA}, Huh et al (2020) report a new processing pathway that involves DIS3L2, a Wilms tumor-associated exoribonuclease. Interestingly, tRF^{Tryr} _{GUA} induction occurs rapidly following oxidative stress and is concomitant with a drastic reduction of pre-tRNA Tryr _{GUA} levels without changes in the mature tRNA pool. That DIS3L2 depletion does not fully restore pretRNA levels upon oxidative stress points to an intermediate precursor $\text{t}\text{RF}^{\text{Tyr}}_{\text{GUA}}$ form that is processed. Evidence that this DIS3L2 mediated processing event is evolutionarily conserved in C. elegans suggests that it may have widespread physiological implications for organismal growth. Thus, a future challenge is to determine how DIS3L2-mediated tRF processing is regulated to generate mature tRF^{Tyr}_{GUA} and possibly other tRFs.

There is a growing realization that tRFs may provide additional means to direct genetic information upon exposure to stress, during development and in disease (Oberbauer & Schaefer, 2018). Although still little is known about the function of tRFs, exciting studies suggest that specific fragments perturb mRNA stability and repress translation through critical interactions with RNA binding proteins (RBPs) and translation initiation factors (Ivanov et al, 2011; Goodarzi et al, 2015; Guzzi et al, 2018). Huh et al (2020) expand the tRF-protein interactome by uncovering selective binding between tRF^{Tryr} _{GUA} and two RBPs with multiple roles in RNA regulation, namely hnRNPA1 and La/SSB, in response to oxidative stress. Intriguingly, tRF^{Tyr}_{GUA} may directly compete for hnRNPA1 binding with specific growthassociated mRNA subsets, promoting their destabilization under stress conditions. This effect consolidates the translational repression of growth-promoting transcripts imposed by the subsequent depletion of mature $tRNA^{Tyr}_{CHA}$. As such, these findings highlight divergent control programs that converge on pre-tRNA Tryr _{GUA} to adapt the cellular transcriptome and translatome during oxidative stress (Fig 1).

In conclusion, the recent study by Huh et al (2020) supports evidence that multifaceted gene regulatory mechanisms intrinsic to the genetic code are integral to the cellular stress response. While new exciting horizons are opening within this emerging research field, future work is needed to delineate the signaling cascades that coordinate tRNA fragmentation and reprogram protein synthesis during critical cell state transitions in development and tumorigenesis.

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