

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



Contents lists available at ScienceDirect

Best Practice & Research Clinical Rheumatology

journal homepage: www.elsevierhealth.com/berh

8

The interplay between neutrophils, complement, and microthrombi in COVID-19



Rheumatology

Yu Zuo ^a, Yogendra Kanthi ^b, Jason S. Knight ^a, Alfred H.J. Kim ^{c, *}

^a Division of Rheumatology, Department of Internal Medicine, University of Michigan, Ann Arbor, MI, USA

^b Division of Intramural Research National Heart, Lung and Blood Institute Bethesda, Maryland, USA

^c Division of Rheumatology, Department of Medicine, Washington University School of Medicine, Saint Louis, Missouri, USA

Keywords: COVID-19 SARS-CoV-2 Neutrophil extracellular traps NETs Complement Innate immunity Thrombotic microangiopathy

ABSTRACT

As of the end of 2020, coronavirus disease 2019 (**COVID-19**) remains a global healthcare challenge with alarming death tolls. In the absence of targeted therapies, supportive care continues to be the mainstay of treatment. The hallmark of severe COVID-19 is a thromboinflammatory storm driven by innate immune responses. This manifests clinically as acute respiratory distress syndrome, and in some patients, widespread thrombotic microangiopathy. Neutrophils and complement are key players in the innate immune system, and their role in perpetuating fatal severe COVID-19 continues to receive increasing attention. Here, we review the interplay between neutrophils, neutrophil extracellular traps, and complement in COVID-19 immunopathology, and highlight potential therapeutic strategies to combat these pathways.

© 2021 Elsevier Ltd. All rights reserved.

Introduction

The coronavirus disease 2019 (**COVID-19**) is caused by the novel severe acute respiratory syndrome coronavirus 2 (**SARS-CoV-2** [1]. SARS-CoV-2 can present as asymptomatic infection or it may cause a wide spectrum of disease, ranging from mild upper respiratory tract infection to life-threatening sepsis

https://doi.org/10.1016/j.berh.2021.101661 1521-6942/© 2021 Elsevier Ltd. All rights reserved.

^{*} Corresponding author. 660 S. Euclid Ave. Campus Box 8045 St. Louis, MO, USA 63110. *E-mail address:* akim@wustl.edu (A.H.J. Kim).

with multiorgan failure [2]. Given its rapid spread, high contagiousness, and lack of specific effective treatment, this pandemic remains a global healthcare challenge [1,3,4].

SARS-CoV-2 displays a complex relationship with the human immune system [5]. It has a unique ability during early infection to inhibit host type I interferon and natural killer cell responses, thereby compromising the body's antiviral defenses and leading to high viral loads in some patients [5,6]. As a result of initial immune evasion, the body then mounts a compensatory hyperinflammatory response to aid in viral clearance. This is characterized by persistent hyperactivation of innate immunity with the overproduction of proinflammatory cytokines; hyperinfiltration of neutrophils, monocytes, and macrophages in lung parenchyma; and extensive epithelial and endothelial damage [5,7,8]. This fulminant inflammatory storm can lead to acute respiratory distress syndrome, and, in some patients, provoke thrombotic widespread microangiopathy, which results in multiorgan failure and death. Neutrophils and complement are key players in our innate immune system, and their roles to potentiate severe COVID-19 continues to receive much attention. Here, we will review potential roles of neutrophils and complement in COVID-19 immunopathology and highlight some potential therapeutic strategies that may target these pathways.

Neutrophil extracellular traps and COVID-19

Neutrophils are the most abundant immune cells in circulation. They act as the first responder against various infections. For decades it was assumed that phagocytosis was the primary mechanism by which neutrophils neutralized pathogens. However, in 2004, Brinkmann and colleagues described another distinct activity of neutrophils, in which neutrophils release sticky extracellular "spider webs" composed of neutrophil-derived chromatin, microbicidal proteins, and mitochondrial remnants [9]. They named this process neutrophil extracellular trap (**NET**) release or NETosis. NETs act to prevent the spread of infections and also use their antimicrobial peptides to kill pathogens [9,10]. By exposing potential autoantigens, such as nucleic acids and modified proteins, NETs may also contribute to autoimmune responses in susceptible individuals [10]. In recent years, it has also been revealed that NETs are prothrombotic [11]. NETs activate platelets and the clotting cascade, and ultimately form an integral part of arterial and venous thrombi [11–15].

Prior knowledge regarding NETs in viral infection

Many viruses can directly trigger NET release [16]. HIV and respiratory syncytial virus (**RSV**) induce NETosis through toll-like receptor-mediated signaling [17,18]. Viruses can also promote NETosis indirectly through triggers such as IL-8, type I interferon, and activated platelets [16]. NETs contribute to antiviral immunity by immobilizing viral particles and inactivating them through the release of antimicrobial molecules such as myeloperoxidase, α -defensin, and cathelicidins [16]. While NETs help contain viral spread, they may also cause damage to the host. For example, NETs promote alveolar-capillary injury in RSV and influenza models [19–21]. High levels of NETs are observed in patients with pneumonia-associated acute respiratory distress syndrome (**ARDS**) [16], where isolated neutrophils have a lower threshold for spontaneous NET release.

NETs in COVID-19

It was recognized early in the pandemic that many patients hospitalized with COVID-19 demonstrate neutrophilia, which predicts critical illness and in-hospital mortality [22–24]. Furthermore, autopsy reports of COVID-19 lung tissues revealed intra-alveolar and capillary neutrophil hyperinfiltration, acute capillaritis with fibrin deposition, and neutrophilic mucositis [25–27]. In April 2020, high levels of NETs in the blood of patients with severe COVID were first reported [28]. This study assessed three markers of NETs [cell-free DNA, myeloperoxidase (**MPO**)-DNA complexes, and citrullinated histone H3 (**Cit-H3**)] in sera of 50 patients hospitalized with COVID-19 as compared to 30 healthy controls [28]. All three markers were significantly elevated in COVID-19 sera, while high levels of NETs were associated with the severest disease [28]. Notably, COVID-19 sera robustly triggers NET release from healthy donor neutrophils [28]. These findings were soon confirmed by independent groups. In a prospective cohort of 33 patients hospitalized with COVID-19, high levels of MPO-DNA complexes were associated with severe respiratory status and worse clinical outcomes [29]. Upon convalescence, levels of MPO-DNA complexes decreased [29]. Others have also confirmed high circulating NETs among COVID-19 patients and their tendency to tracking with disease severity [30,31].

The first microscopic confirmation of NETs was reported in an autopsy series from New Orleans where degenerative neutrophils with strands of extracellular material that stained weakly positive for DNA were present in COVID-19 lungs [32]. Histological detection of NETs has since been made not only in the pulmonary vasculature, but also in the microvasculature of the kidney and heart [30,31,33]. Notably, the transcriptomic analysis of four COVID-19 patients' lung tissue and bronchial alveolar lavage fluid revealed marked enrichment of NET-associated genes [34]. In summary, several lines of data have supported the presence of NETs in the severest forms of COVID-19 (Table 1).

Table 1Studies investigating NETs in COVID-19.

Reference	Publication Date	Study Design	Study Population	Main Findings
Zuo [28]	April 24, 2020	Cohort	N = 50	Markedly elevated NETs (cell-free DNA, MPO- DNA, and Cit-H3) were detected in the sera of patients hospitalized with COVID-19. Markers of NETs associated with severe respiratory status. Sera from individuals with COVID-19 triggered NET release from control neutrophils <i>in vitro</i> .
Fox [35] Middleton [29]	May 27, 2020 June 29, 2020	Autopsy series Prospective Cohort	N = 10 N = 33	First microscopic confirmation of NETs in alveoli. High level of NETs (MPO-DNA) were detected in COVID-19 blood, where they were associated with worse respiratory status and clinical outcomes. Platelet-derived factors known to trigger NETosis were also elevated. COVID-19 plasma potentiated NET release <i>in vitro</i> , which could be attenuated by neonatal NET-inhibitory factor. Autopsies revealed colocalization of PF4 and NETs in pulmonary vessels.
Nicolai [33]	July 22, 2020	Cohort	N = 38	Immunothrombi enriched with neutrophils were seen in COVID-19 lungs. Distinct neutrophil signatures were appreciated in COVID-19, which range from a hypoactive phenotype in patients with intermediate COVID-19 to excessive neutrophil activation in severe COVID-19. Platelets isolated from patients with severe COVID-19 were bound to neutrophils and potentiated NETosis <i>in vitro</i> .
Leppkes [30]	July 31, 2020	Cohort	N = 71	High level of NETs (cell-free DNA, MPO-DNA, Cit- H3, and neutrophil elastase-DNA complexes) were appreciated in COVID-19 blood, where they were associated with severe disease. Significantly increased low-density neutrophils that have high propensity for spontaneous NETosis were observed in COVID-19 patients. Autopsy of COVID-19 patients demonstrated the occlusion of pulmonary vessels by aggregated NETs.
Wang [34]	Aug 18, 2020	Cohort	N=4	Transcriptomic analysis of COVID-19 lung tissue and BAL fluid revealed marked enrichment of NET-associated genes.
Veras [31]	Sep 14, 2020	Cohort	N=32	High levels of NETs (MPO-DNA) were detected in the blood of COVID-19 patients. SARS-CoV-2 directly induced NETosis through a mechanism that was dependent upon PAD4, effective viral replication, and host cell ACE2.

MPO = myeloperoxidase; Cit-H3 = citrullinated histone H3; PF4 = platelet factor 4; NETs = neutrophil extracellular traps; PAD4 = protein arginine deiminase 4; and BAL = bronchial alveolar lavage.

Y. Zuo, Y. Kanthi, J.S. Knight et al.

NET induction in COVID-19

In COVID-19, NETs are likely produced in both direct and indirect fashion. NETs can be induced by incubating SARS-CoV-2 with healthy neutrophils [31]; in this context, the inhibition of either PAD4 or RNA polymerase abrogates NETosis. SARS-CoV-2 utilizes ACE2 receptor and serine protease TMPRSS2 to enter host cells [36], and the blockade of either ACE2 or serine protease activity antagonizes SARS-CoV-2-triggered NETosis [31]. Other studies have suggested that NETs are produced indirectly in COVID-19 through activated platelets. Platelets from patients with severe COVID-19 demonstrate increased adhesion to neutrophils *in vivo*, while triggering NETosis from healthy neutrophils *in vitro* [33]. Furthermore, platelet-derived molecules such as platelet factor 4 (**PF4**) and RANTES, which are known to trigger NETosis, are markedly elevated in COVID-19 patients [37]. Autopsy specimens have revealed the colocalization of PF4 and NETs in the pulmonary vasculature [37]. Considering that severe COVID-19 is also associated with high levels of myriad cytokines and chemokines, their role as pro-NET factors should also be considered.

NETs and SARS-CoV-2 pathology

NETs have direct cytotoxic effects against epithelial and endothelial cells. Indeed, one study demonstrated that SARS-CoV-2-associated NETs induce pulmonary epithelial cell death, which results in alveolar damage and fibrosis [31]. NETs likely also potentiate microvascular thrombosis in COVID-19, as multiple autopsy studies have revealed NET-containing microthrombi and neutrophil-platelet infiltration in the microvasculature of the lung, kidney, and heart [31,37–39]. While not well investigated, disproportionate NET formation may also lead to pathogenic autoantibody production that likely contributes to further local or systemic damage. In summary, clinical, pathological, and molecular evidence support the presence of NETs in COVID-19. SARS-CoV-2 can directly and indirectly induce NET formation, which contributes to COVID-19 pathology.

Complement: helpful early in COVID-19, detrimental late

Data from clinical, pathological, and molecular studies suggest an important role for complement activation in the development of severe COVID-19 manifestations. Yet, it is also clear that complement is critical for protective responses early in COVID-19.

Complement in immunity and disease

The complement system is composed of over 40 serine proteases, inhibitors, and receptors initiated by three pathways (classical, lectin, and alternative). Each pathway is uniquely triggered to drive a proteolytic cascade, which generates a potent, highly regulated, innate immune response [40]. Its activation induces: 1) membrane perturbation mediated by C4b- and C3b-facilitated opsonization and phagocytosis as well as a lytic process through the membrane attack complex (**MAC and C5b-9**), and 2) the generation of a proinflammatory milieu largely mediated through the anaphylatoxins, C3a and C5a. Using these two phenomena, the complement system also assists in the clearance of apoptotic material and cellular debris. Additionally, intracellular complement activation enables cells to modulate metabolic pathways and thereby regulate immune responsiveness [41].

These same pathways though are utilized in several complement-mediated diseases. In systemic lupus erythematosus and related autoimmune diseases, immune complexes generated by autoantibodies drive type II and III hypersensitivity reactions, driving classical pathway activation to initiate destructive inflammatory responses. In age-related macular degeneration (**AMD**) and atypical hemolytic uremic syndrome (**aHUS**), loss-of-function (commonly haploinsufficiency) of complement regulatory proteins or gain-of-function in complement-activating components promote excessive alternative pathway engagement, leading to retinal damage in AMD and microthrombi featuring endothelial injury in aHUS [42,43].

Complement activation is likely required for controlling early COVID-19

Complement has a well-established role in immunity against viruses. Initiated by natural and crossreacting antibodies and lectins, viruses trigger complement action to drive the opsonization of viruses and virus-infected cells, generation of an antiviral inflammatory state, and boost virus-specific immune responses [44].

Indeed, several lines of data support that SARS-CoV-2 can directly activate complement. *In vitro*, SARS-CoV-2 spike proteins 1 and 2 expressed on the human cell line PIGAnull TF1 primarily drives the activation of the alternative pathway through heparan sulfate [45], and in a preprint, the N protein activated the mannan-binding lectin-associated serine protease (**MASP-2**) [46]. Furthermore, virus infection appears to drive the early activation of complement *in vivo*, as in a murine model of SARS-CoV, infected mice generate C3 activation products (C3a, C3b, iC3b, C3c, and C3d) in the lung within 24 h [47]. While numerous reports have observed elevated complement activation product levels in severe COVID-19 patients (as discussed later), only one report assessed complement component levels in early COVID-19 infection. Here, low complement C3 was found in 57% of day 1 samples in COVID-19 patients who eventually developed nonserious pneumonia [48]. This may be an underestimation of complement activation though, as C3 is an acute phase reactant [49]. Thus, normal C3 levels may also represent elevated complement activation status due to the elevated production of C3 in inflammatory states.

To counteract complement activation, viruses have developed evasion tactics. While these have yet to be observed for coronaviruses, other viruses have well-described mechanisms (reviewed in Ref. [50]). For example, poxviruses express a protein with similar structure and function to complement regulatory proteins, while flaviviruses neutralize complement regulators by binding to them.

While our understanding of the role of complement in early COVID-19 remains in its infancy, we do speculate that complement, along with other immune pathways such as interferons and neutrophil activation, serve to drive important antivirus responses. If compromised, this can lead to inadequate control of SARS-CoV-2, which leads to severe manifestations of COVID-19.

Complement contributes to immunopathology observed in severe COVID-19

Initial data that support a prominent role of complement in severe COVID-19 came from clinicopathological studies. Jeffrey Laurence's group identified complement deposition (specifically C5b-9, C4d, and MASP-2), in the skin of deceased COVID-19 patients with retiform and purpuric lesions and the lung of those with septal capillary injury [25]. Notably, they also found the C4d and C5b-9 colocalized with SARS-CoV-2 S protein in these organs [25]. Similarly, mannose-binding lectins (**MBL**), C4, C3, and C3b-9 were observed in alveolar epithelial cells and alveolar exudate in deceased patients [46]. Furthermore, C5b-9 deposition was also observed in the apical brush border of tubular epithelial cells in the kidney [51].

Additional histopathological features with similarities to other complement-mediated diseases indicate that complement activation is a central player in severe COVID-19. Endothelial cell abnormalities, such as cellular swelling with foamy degeneration in the setting of a thrombotic microangiopathy (**TMA**), have been observed in numerous organs [8,52–56], consistent with C5b-9 mediated injury and a hypercoagulable state [57]. In the lungs of those with COVID-19-associated ARDS, septal microangiopathy was observed characterized by endothelial cell injury, mural fibrin deposition, and variable intraluminal thrombus formation [8,58,59]. Consistent with these pathological observations, C3-deficient mice infected with SARS-CoV demonstrated less lung inflammation and injury and weight loss despite having similar viral loads as compared to wild-type mice [47].

Both proteomic [46,60–64] and transcriptomic [62,65] studies of blood and lung samples from severe COVID-19 confirm the notion that complement activation is an immune signature characteristic of severe disease. The earliest data were observed in Chinese COVID-19 patients, as elevated serum C5a was uniquely noted in those with severe respiratory distress or hypoxia on room air as compared to those with mild symptoms or healthy controls [46]. C5a was confirmed as a potential biomarker of severe disease in a longitudinal analysis of a French cohort, and both anti-C5aR1-treated or C5aR1-knockout mice had attenuated lung injury following C5a instillation into the lungs [66]. MBL was

associated with plasma D-dimer concentration in a Swedish cohort of critically ill COVID-19 patients [67]. Using the Albany Medical Center (New York) cohort, plasma target metabolomics identified complement activation and its regulation as two of the top six GO (gene ontogeny) pathways unique to hospitalized COVID-19 patients who require ICU admission as compared to those on the medical floor [64]. Data from the Columbia University Irving Medical Center/New York-Presbyterian Hospital cohort confirmed these observations as hospitalized patients with COVID-19 exhibited elevated serum levels of factor H and I along with C5, as assessed by mass spectroscopy, as compared to healthy controls [60]. Using immunoassays in the Norwegian cohort, elevated plasma sC5b-9, C5a, C3b/iC3b/C3c, alternative pathway C3 convertase (C3bBbP), and C4d were observed in majority of COVID-19 patients in respiratory failure and were higher as compared to those that were not [61]. Similar observations were made in the lungs at the transcriptomic level, as lung biopsy samples from COVID-19 patients showed a pronounced complement signature (including C3 transcripts) following GO pathway analysis as compared to healthy controls [65]. They confirmed this observation using in vitro SARS-CoV-2-infected primary bronchial epithelial cells or a type II human pneumocyte cell line (A549) with or without ACE2 overexpression, which demonstrates a similar complement signature, which was not observed in influenza A- or respiratory syncytial virus-infected cells [65].

Collectively, these data provide substantial evidence that complement activation occurs in COVID-19, with emerging data that this positively correlates with disease severity.

Interplay between NETs and complement

As discussed above, NETs are webs of extracellular chromatin decorated with factors derived from neutrophil cytoplasm, granules, and mitochondria. NETs restrain invading microbes through both immobilization and killing, but when released intravascularly they are an important nidus for thromboinflammation. For example, NETs activate platelets [68], capture red blood cells [68], and potentiate both intrinsic [13] and extrinsic pathways [69] of coagulation.

Prior knowledge regarding the interplay between NETs and complement

The communication between NETs and complement is bidirectional and, if not tightly regulated, has potential to form a self-amplifying loop. C5a recruits and then primes neutrophils for NETosis through the upregulation of various immune receptors, including TLRs and complement receptors such as CR1 and CR3 [70,71]. Pathogens opsonized with C3b and iC3b may then trigger NETosis through the engagement of neutrophils CR1 and CR3, respectively [72]. Furthermore, once NETs have been released into the extracellular space, complement components—and perhaps particularly C1q—stabilize their structure and prevent clearance by DNase I [73].

Emphasizing the bidirectional nature of the relationship, neutrophils express Factor B, C3, and properdin, the combination of which allows for the production and stabilization of the C3-convertase on the neutrophil surface [74] and also on NETs themselves [75]. At the same time, myeloperoxidase and serine proteases such as cathepsin G and proteinase 3 bind to and activate properdin with the potential to further potentiate complement activation on NETs [76]. The end result is NETs as a fertile platform for the generation of anaphylatoxins C3a and C5a [73], which can lead to further immune system activation.

NETs, complement, and thrombotic microangiopathy

Prior to COVID-19, circulating NET remnants were already known to track closely with the activity of various **TMAs**, including thrombotic thrombocytopenic purpura [77,78], hemolytic uremic syndrome [79,80], transplant-associated TMA [78,81], and likely catastrophic antiphospholipid syndrome [82]. At the same time, these TMAs are also clearly complementopathies as evidenced by the utility of inhibitors of C5 cleavage as effective therapies [83]. One interesting study contrasted transplant-associated TMA with another complication of stem-cell transplant, graft versus host disease (**GVHD**) [84]. While markers of endothelial damage/activation such as thrombomodulin were elevated in both situations, only transplant-associated TMA was associated with elevations in soluble C5b-9, NET

remnants, including myeloperoxidase-DNA complexes and markers of coagulation pathways such as thrombin-antithrombin complexes [84].

The potential for cross talk between complement and NETs has recently been evaluated in COVID-19. Specifically, Skendros and colleagues found that C3 inhibitor compstatin Cp40 disrupted tissue factor expression by COVID-19 neutrophils, while C5a receptor blockade attenuated NETosis triggered by COVID-19 platelet-rich plasma [85]. The group also found that COVID-19 NETs are decorated with particularly high levels of tissue factor and through this and likely other pathways are potent activators of cultured endothelial cells [85]. Taken together, it is not difficult to see how these pathways could conspire to occlude the COVID-19 microvasculature *in vivo*.

Potential approaches to treatment

As we await what will hopefully be definitive antiviral solutions to the current pandemic, antineutrophil and anti-complement therapies have potential to help mitigate the severest manifestations of COVID-19.

Medications already used in the treatment of COVID-19

Patients with COVID-19 are increasingly being treated with the combination of heparin-based anticoagulation and dexamethasone. The former may modulate NETs by the neutralization of cyto-toxic histones [86] and by the potentiation of NET clearance through DNase I [30]. At the same time, corticosteroids such as dexamethasone can also reduce NET formation *in vivo* [87], most likely through the modulation of the inflammatory mediators that activate neutrophils. Other agents being trialed in COVID-19 such as JAK-STAT inhibitors, anakinra (IL-1 receptor antagonist), and colchicine also have potential to reduce NET release [88–90].

DNases

Recombinant DNase I cleaves the DNA scaffold of NETs and thereby has the potential to relieve intravascular thrombosis and airway obstruction [30]. Applying this logic, a small single-center case series has suggested that nebulized endotracheal dornase alfa (a biosynthetic form of human DNase I) reduced oxygen requirements in all treated patients [91]. Going forward, COVIDornase (NCT04355364) and COVASE (NCT04359654) are two studies evaluating nebulized dornase alfa in prospective randomized controlled trials. While there are not (to our knowledge) any active studies for intravenous administration, one wonders if some of the products might find their way into circulation through the damaged airways of COVID-19 [92].

Treatment by modulation of purinergic signaling with dipyridamole

The activation of surface adenosine receptors suppresses NETosis through cyclic AMP-dependent signaling [93]. Dipyridamole is an inexpensive, FDA-approved drug with a favorable safety profile. Dipyridamole potentiates adenosine receptor signaling by both the inhibition of ectonucleoside reuptake and stabilization of intracellular cyclic AMP. Dipyridamole tempers NET release *in vitro*, which prevents NET-dependent thrombosis in mice [93]. In a small study, dipyridamole suppressed D-dimer levels in patients with COVID-19 [94]. Larger studies are now underway to evaluate clinical outcomes (NCT04391179, NCT04424901) [95].

Treatment with complement inhibitors

C5 has been an early experimental target for severe COVID-19 given that eculizumab is already FDAapproved for several complementopathies. Severe COVID-19 patients treated with C5 cleavage inhibitors (i.e., eculizumab, ravulizumab, and LFG316) have resulted in success in several small case series and reports [96–101], and a small number of patients already on a C5 cleavage inhibitor for PNH or TMA-associated lupus nephritis failed to progress to severe COVID-19 [102–104]. A C5a inhibitor, vilobelimab (IFX-1), failed to meet the primary endpoint of improved lung function in this small (n = 30) exploratory study, although two secondary outcome measures, pulmonary embolism and renal impairment, were improved in the IFX-1 treatment group [105]. It remains too early to tell whether inhibiting C5a, C5b-9, or both is required to suppress severe manifestations of COVID-19, but several clinical trials inhibiting C5 cleavage (eculizumab: NCT04346797, NCT04355494, ravulizumab: NCT04369469, NCT04390464, and zilucoplan: NCT04382755) and the C5a receptor (avdoralimab: NCT0431367 and IFX-1: NCT04333420) are currently ongoing.

Given its central position, another attractive inhibitory target of complement activation is C3. AMY-101 is a compstatin-based C3 inhibitor that prevents the conformational change of C3 required for subsequent cleavage to C3 and C3b [106], which was described earlier to attenuate neutrophil tissue factor expression [85]. A phase II study is now active examining the utility of AMY-101 (NCT04395456), and a total of four patients with severe COVID-19 have been successfully treated with AMY-101 [101,107]. There may be a theoretical advantage to inhibiting C3 rather than C5, as greater inhibition of NET formation, neutrophil and lymphocyte recovery, and more rapid LDH decline [101].

Additional targets, including the C1 esterase inhibitor (conestat alfa, NCT04414631 and rocunest, NCT04530136) [108] and MASP-2 [109] have also been tried in pilot populations with success.

Summary

In summary, NET-releasing neutrophils and complement appear to play central roles in the immunopathogenesis of COVID-19. This review has presented various points of cross talk between NETs and complement in SARS-CoV-2 infection. These intermingled pathways conspire to promote a thromboinflammatory storm in some individuals with severe COVID-19. Therefore, the development of novel therapeutic strategies that target NET formation and complement activation may help reduce COVID-19 morbidity and mortality.

Practice points

- SARS-CoV-2 displays a complex relationship with the human immune system. The interplay between neutrophils, neutrophil extracellular traps, and complement—important players in innate immunity—contribute to the thromboinflammatory milieu of COVID-19.
- SARS-CoV-2 can directly and indirectly induce NET formation, which contributes to COVID-19 pathology.
- NETs contribute to COVID-19 pathology by:
 - o direct cytotoxic effects against epithelial and endothelial cells
 - microthrombi formation and microvascular damage in multiple organs
 - perpetuating pathogenic autoantibody production
- While providing early viral containment, complement activation heightens immunopathology and contributes to COVID-19 severity.
- The cross-talk between NETs and complement are key drivers of COVID-19 thrombotic microangiopathy.
- Anti-neutrophil and anti-complement therapies have potential to mitigate the severest manifestations of COVID-19.

Research agenda

- Deep dive into the mechanisms that govern the tripartite immunopathogenesis of COVID-19 and their downstream and long-term sequela.
- Identify clinically actionable biomarkers for NETs and complement pathway activation thus enables precision management.
- Evaluate therapeutics to target NETs and complement in COVID-19.

Funding

The authors have no relevant financial conflicts to report.

Declaration of competing interest

The authors have no relevant conflicts to report.

References

- Zhu N, Zhang D, Wang W, et al. A novel coronavirus from patients with pneumonia in China, 2019. N Engl J Med 2020; 382:727-33.
- [2] Wiersinga WJ, Rhodes A, Cheng AC, et al. Pathophysiology, transmission, diagnosis, and treatment of coronavirus disease 2019 (COVID-19): a review. J Am Med Assoc 2020;324:782–93.
- [3] Zhou P, Yang XL, Wang XG, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. Nature 2020;579:270–3.
- [4] Ruan Q, Yang K, Wang W, et al. Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan, China. Intensive Care Med 2020;46:846–8.
- [5] Maggi E, Canonica GW, Moretta L. COVID-19: unanswered questions on immune response and pathogenesis. J Allergy Clin Immunol 2020;146:18–22.
- [6] Ye Q, Wang B, Mao J. The pathogenesis and treatment of the `Cytokine Storm' in COVID-19. J Infect 2020;80:607-13.
- [7] Lo MW, Kemper C, Woodruff TM. COVID-19: complement, coagulation, and collateral damage. J Immunol 2020;205: 1488–95.
- [8] Menter T, Haslbauer JD, Nienhold R, et al. Postmortem examination of COVID-19 patients reveals diffuse alveolar damage with severe capillary congestion and variegated findings in lungs and other organs suggesting vascular dysfunction. Histopathology 2020 Aug;77(2):198–209.
- [9] Brinkmann V, Reichard U, Goosmann C, et al. Neutrophil extracellular traps kill bacteria. Science 2004;303:1532–5.
- [10] Lee KH, Kronbichler A, Park DD, et al. Neutrophil extracellular traps (NETs) in autoimmune diseases: a comprehensive review. Autoimmun Rev 2017;16:1160–73.
- [11] Thalin C, Hisada Y, Lundstrom S, et al. Neutrophil extracellular traps: villains and targets in arterial, venous, and cancerassociated thrombosis. Arterioscler Thromb Vasc Biol 2019;39:1724–38.
- [12] Massberg S, Grahl L, von Bruehl ML, et al. Reciprocal coupling of coagulation and innate immunity via neutrophil serine proteases. Nat Med 2010;16:887–96.
- [13] von Bruhl ML, Stark K, Steinhart A, et al. Monocytes, neutrophils, and platelets cooperate to initiate and propagate venous thrombosis in mice in vivo. J Exp Med 2012;209:819–35.
- [14] Knight JS, Luo W, O'Dell AA, et al. Peptidylarginine deiminase inhibition reduces vascular damage and modulates innate immune responses in murine models of atherosclerosis. Circ Res 2014;114:947–56.
- [15] Borissoff JI, Joosen IA, Versteylen MO, et al. Elevated levels of circulating DNA and chromatin are independently associated with severe coronary atherosclerosis and a prothrombotic state. Arterioscler Thromb Vasc Biol 2013;33:2032–40.
 [16] Schonrich G, Raftery MJ. Neutrophil extracellular traps go viral. Front Immunol 2016;7:366.
- [17] Funchal GA, Jaeger N, Czepielewski RS, et al. Respiratory syncytial virus fusion protein promotes TLR-4-dependent neutrophil extracellular trap formation by human neutrophils. PloS One 2015;10. e0124082.
- [18] Saitoh T, Komano J, Saitoh Y, et al. Neutrophil extracellular traps mediate a host defense response to human immunodeficiency virus-1. Cell Host Microbe 2012;12:109–16.
- [19] Gupta AK, Joshi MB, Philippova M, et al. Activated endothelial cells induce neutrophil extracellular traps and are susceptible to NETosis-mediated cell death. FEBS Lett 2010;584:3193-7.
- [20] Saffarzadeh M, Juenemann C, Queisser MA, et al. Neutrophil extracellular traps directly induce epithelial and endothelial cell death: a predominant role of histones. PloS One 2012;7:e32366.
- [21] Cortjens B, de Boer OJ, de Jong R, et al. Neutrophil extracellular traps cause airway obstruction during respiratory syncytial virus disease. J Pathol 2016;238:401–11.
- [22] Liu J, Liu Y, Xiang P, et al. Neutrophil-to-lymphocyte ratio predicts critical illness patients with 2019 coronavirus disease in the early stage. J Transl Med 2020;18:206.
- [23] Liu S, Su X, Pan P, et al. Neutrophil extracellular traps are indirectly triggered by lipopolysaccharide and contribute to acute lung injury. Sci Rep 2016;6:37252.
- [24] Liu Y, Du X, Chen J, et al. Neutrophil-to-lymphocyte ratio as an independent risk factor for mortality in hospitalized patients with COVID-19. J Infect 2020;81:e6–12.
- [25] Magro C, Mulvey JJ, Berlin D, et al. Complement associated microvascular injury and thrombosis in the pathogenesis of severe COVID-19 infection: a report of five cases. Transl Res 2020;220:1–13.
- [26] Fox SE, Akmatbekov A, Harbert JL, et al. Pulmonary and cardiac pathology in African American patients with COVID-19: an autopsy series from New Orleans. Lancet Respir Med 2020 Jul;8(7):681–6.
- [27] Barnes BJ, Adrover JM, Baxter-Stoltzfus A, et al. Targeting potential drivers of COVID-19: neutrophil extracellular traps. J Exp Med 2020:217.
- [28] Zuo Y, Yalavarthi S, Shi H, et al. Neutrophil extracellular traps in COVID-19. JCI Insight 2020.
- [29] Middleton EA, He XY, Denorme F, et al. Neutrophil extracellular traps contribute to immunothrombosis in COVID-19 acute respiratory distress syndrome. Blood 2020;136:1169–79.
- [30] Leppkes M, Knopf J, Naschberger E, et al. Vascular occlusion by neutrophil extracellular traps in COVID-19. EBioMedicine 2020;58:102925.

- [31] Veras FP, Pontelli MC, Silva CM, et al. SARS-CoV-2-triggered neutrophil extracellular traps mediate COVID-19 pathology. J Exp Med 2020:217.
- [32] Fox SE, Akmatbekov A, Harbert JL, et al. Pulmonary and cardiac pathology in African American patients with COVID-19: an autopsy series from New Orleans. Lancet Respir Med 2020;8:681–6.
- [33] Nicolai L, Leunig A, Brambs S, et al. Immunothrombotic dysregulation in COVID-19 pneumonia is associated with respiratory failure and coagulopathy. Circulation 2020;142:1176–89.
- [34] Wang J, Li Q, Yin Y, et al. Excessive neutrophils and neutrophil extracellular traps in COVID-19. Front Immunol 2020;11: 2063.
- [35] Darnell ME, Taylor DR. Evaluation of inactivation methods for severe acute respiratory syndrome coronavirus in noncellular blood products. Transfusion 2006;46:1770–7.
- [36] Hoffmann M, Kleine-Weber H, Schroeder S, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. Cell 2020;181:271–280 e8.
- [37] Middleton EA, He XY, Denorme F, et al. Neutrophil extracellular traps (NETs) contribute to immunothrombosis in COVID-19 acute respiratory distress syndrome. Blood 2020 Apr 16;181(2):271–280.e8.
- [38] Tian S, Hu W, Niu L, et al. Pulmonary pathology of early-phase 2019 novel coronavirus (COVID-19) pneumonia in two patients with lung cancer. J Thorac Oncol 2020 May; 15(5):700–4.
- [39] Bosmuller H, Traxler S, Bitzer M, et al. The evolution of pulmonary pathology in fatal COVID-19 disease: an autopsy study with clinical correlation. Virchows Arch 2020;477:349–57.
- [40] Holers VM. Complement and its receptors: new insights into human disease. Annu Rev Immunol 2014;32:433–59.
- [41] Liszewski MK, Elvington M, Kulkarni HS, Atkinson JP. Complement's hidden arsenal: new insights and novel functions inside the cell. Mol Immunol 2017;84:2–9.
- [42] Java A, Apicelli AJ, Liszewski MK, et al. The complement system in COVID-19: friend and foe? JCI Insight 2020;5.
- [43] Liszewski MK, Java A, Schramm EC, Atkinson JP. Complement dysregulation and disease: insights from contemporary genetics. Annu Rev Pathol 2017;12:25–52.
- [44] Agrawal P, Nawadkar R, Ojha H, Kumar J, Sahu A. Complement evasion strategies of viruses: an overview. Front Microbiol 2017;8:1117.
- [45] Yu J, Yuan X, Chen H, et al. Direct activation of the alternative complement pathway by SARS-CoV-2 spike proteins is blocked by factor D inhibition. Blood 2020 Oct 29;136(18):2080–9.
- [46] Gao T, Hu M, Zhang X, et al. Highly pathogenic coronavirus N protein aggravates lung injury by MASP-2-mediated complement over-activation. 2020. p. 20041962. 2020.03.29.
- [47] Gralinski LE, Sheahan TP, Morrison TE, et al. Complement activation contributes to severe acute respiratory syndrome coronavirus pathogenesis. mBio 2018;9.
- [48] Wei XS, Wang XR, Zhang JC, et al. A cluster of health care workers with COVID-19 pneumonia caused by SARS-CoV-2. J Microbiol Immunol Infect 2020 Apr 27. S1684-1182(20)30107-9.
- [49] Gabay C, Kushner I. Acute-phase proteins and other systemic responses to inflammation. N Engl J Med 1999;340:448–54.
 [50] Stoermer KA, Morrison TE. Complement and viral pathogenesis. Virology 2011;411:362–73.
- [51] Diao B, Wang C, Wang R, et al. Human kidney is a target for novel severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection. 2020. p. 20031120. 2020.03.04.
- [52] Varga Z, Flammer AJ, Steiger P, et al. Endothelial cell infection and endotheliitis in COVID-19. Lancet 2020;395:1417-8.
- [53] Su H, Yang M, Wan C, et al. Renal histopathological analysis of 26 postmortem findings of patients with COVID-19 in China. Kidney Int 2020;98:219–27.
- [54] Jhaveri KD, Meir LR, Flores Chang BS, et al. Thrombotic microangiopathy in a patient with COVID-19. Kidney Int 2020;98: 509–12.
- [55] Korkeala H, Makela P. Characterization of lactic acid bacteria isolated from vacuum-packed cooked ring sausages. Int J Food Microbiol 1989;9:33–43.
- [56] Bryce C, Grimes Z, Pujadas E, et al. Pathophysiology of SARS-CoV-2: targeting of endothelial cells renders a complex disease with thrombotic microangiopathy and aberrant immune response. In: The Mount Sinai COVID-19 autopsy experience; 2020. p. 20099960. 2020.05.18.
- [57] Timmermans S, Abdul-Hamid MA, Potjewijd J, et al. C5b9 formation on endothelial cells reflects complement defects among patients with renal thrombotic microangiopathy and severe hypertension. J Am Soc Nephrol 2018;29:2234–43.
- [58] Ackermann M, Verleden SE, Kuehnel M, et al. Pulmonary vascular endothelialitis, thrombosis, and angiogenesis in covid-19. N Engl J Med 2020;383:120–8.
- [59] Schaller T, Hirschbuhl K, Burkhardt K, et al. Postmortem examination of patients with COVID-19. J Am Med Assoc 2020; 323:2518–20.
- [60] D'Alessandro A, Thomas T, Dzieciatkowska M, et al. Serum proteomics in COVID-19 patients: altered coagulation and complement status as a function of IL-6 level. J Proteome Res 2020 Nov 6;19(11):4417–27.
- [61] Holter JC, Pischke SE, de Boer E, et al. Systemic complement activation is associated with respiratory failure in COVID-19 hospitalized patients. Proc Natl Acad Sci U S A 2020 Oct 6;117(40):25018–25.
- [62] Overmyer KA, Shishkova E, Miller IJ, et al. Large-scale multi-omic analysis of COVID-19 severity. medRxiv 2020.
- [63] Cugno M, Meroni PL, Gualtierotti R, et al. Complement activation in patients with COVID-19: a novel therapeutic target. J Allergy Clin Immunol 2020;146:215–7.
- [64] Lam LM, Murphy SJ, Kuri-Cervantes L, et al. Erythrocytes reveal complement activation in patients with COVID-19. 2020. p. 20104398. 2020.05.20.
- [65] Yan B, Freiwald T, Chauss D, et al. SARS-CoV2 drives JAK1/2-dependent local and systemic complement hyper-activation. Res Sq 2020 Jun 9. rs.3.rs-33390.
- [66] Carvelli J, Demaria O, Vely F, et al. Association of COVID-19 inflammation with activation of the C5a-C5aR1 axis. Nature 2020 Dec;588(7836):146–50.
- [67] Eriksson O, Hultstrom M, Persson B, et al. Mannose-binding lectin is associated with thrombosis and coagulopathy in critically ill COVID-19 patients. Thromb Haemostasis 2020 Dec;120(12):1720–4.

- [68] Fuchs TA, Brill A, Duerschmied D, et al. Extracellular DNA traps promote thrombosis. Proc Natl Acad Sci U S A 2010;107: 15880–5.
- [69] Kambas K, Mitroulis I, Apostolidou E, et al. Autophagy mediates the delivery of thrombogenic tissue factor to neutrophil extracellular traps in human sepsis. PloS One 2012;7:e45427.
- [70] Martinelli S, Urosevic M, Daryadel A, et al. Induction of genes mediating interferon-dependent extracellular trap formation during neutrophil differentiation. J Biol Chem 2004;279:44123–32.
- [71] Behnen M, Leschczyk C, Moller S, et al. Immobilized immune complexes induce neutrophil extracellular trap release by human neutrophil granulocytes via FcgammaRIIIB and Mac-1. J Immunol 2014;193:1954–65.
- [72] Palmer LJ, Damgaard C, Holmstrup P, Nielsen CH. Influence of complement on neutrophil extracellular trap release induced by bacteria. J Periodontal Res 2016;51:70–6.
- [73] Leffler J, Martin M, Gullstrand B, et al. Neutrophil extracellular traps that are not degraded in systemic lupus erythematosus activate complement exacerbating the disease. J Immunol 2012;188:3522–31.
- [74] Camous L, Roumenina L, Bigot S, et al. Complement alternative pathway acts as a positive feedback amplification of neutrophil activation. Blood 2011;117:1340–9.
- [75] Wang H, Wang C, Zhao MH, Chen M. Neutrophil extracellular traps can activate alternative complement pathways. Clin Exp Immunol 2015;181:518–27.
- [76] O'Flynn J, Dixon KO, Krol MCF, et al. Myeloperoxidase directs properdin-mediated complement activation. Journal of Innate Immunity 2014;6:417–25.
- [77] Jimenez-Alcazar M, Napirei M, Panda R, et al. Impaired DNase1-mediated degradation of neutrophil extracellular traps is associated with acute thrombotic microangiopathies. J Thromb Haemostasis 2015;13:732–42.
- [78] Arai Y, Yamashita K, Mizugishi K, et al. Serum neutrophil extracellular trap levels predict thrombotic microangiopathy after allogeneic stem cell transplantation. Biol Blood Marrow Transplant 2013;19:1683–9.
- [79] Leffler J, Prohaszka Z, Mikes B, et al. Decreased neutrophil extracellular trap degradation in shiga toxin-associated haemolytic uraemic syndrome. J Innate Immun 2017;9:12–21.
- [80] Ramos MV, Mejias MP, Sabbione F, et al. Induction of neutrophil extracellular traps in shiga toxin-associated hemolytic uremic syndrome. J Innate Immun 2016;8:400–11.
- [81] Gloude NJ, Khandelwal P, Luebbering N, et al. Circulating dsDNA, endothelial injury, and complement activation in thrombotic microangiopathy and GVHD. Blood 2017;130:1259–66.
- [82] Sule G, Kelley WJ, Gockman K, et al. Increased adhesive potential of antiphospholipid syndrome neutrophils mediated by beta 2 integrin mac-1. Arthritis Rheum 2020;72:114–24.
- [83] Legendre CM, Licht C, Muus P, et al. Terminal complement inhibitor eculizumab in atypical hemolytic-uremic syndrome. N Engl J Med 2013;368:2169–81.
- [84] Gavriilaki E, Chrysanthopoulou A, Sakellari I, et al. Linking complement activation, coagulation, and neutrophils in transplant-associated thrombotic microangiopathy. Thromb Haemostasis 2019;119:1433–40.
- [85] Skendros P, Mitsios A, Chrysanthopoulou A, et al. Complement and tissue factor-enriched neutrophil extracellular traps are key drivers in COVID-19 immunothrombosis. J Clin Invest 2020 Nov 2;130(11):6151–7.
- [86] Buijsers B, Yanginlar C, Maciej-Hulme ML, et al. Beneficial non-anticoagulant mechanisms underlying heparin treatment of COVID-19 patients. EBioMedicine 2020;59:102969.
- [87] Vargas A, Boivin R, Cano P, et al. Neutrophil extracellular traps are downregulated by glucocorticosteroids in lungs in an equine model of asthma. Respir Res 2017;18:207.
- [88] Furumoto Y, Smith CK, Blanco L, et al. Tofacitinib ameliorates murine lupus and its associated vascular dysfunction. Arthritis Rheum 2017;69:148–60.
- [89] Yadav V, Chi L, Zhao R, et al. Ectonucleotidase tri(di)phosphohydrolase-1 (ENTPD-1) disrupts inflammasome/interleukin 1beta-driven venous thrombosis. J Clin Invest 2019;129:2872–7.
- [90] Apostolidou E, Skendros P, Kambas K, et al. Neutrophil extracellular traps regulate IL-1beta-mediated inflammation in familial Mediterranean fever. Ann Rheum Dis 2016;75:269–77.
- [91] Weber AG, Chau AS, Egeblad M, et al. Nebulized in-line endotracheal dornase alfa and albuterol administered to mechanically ventilated COVID-19 patients: a case series. Mol Med 2020;26:91.
- [92] Aitken ML, Burke W, McDonald G, et al. Recombinant human DNase inhalation in normal subjects and patients with cystic fibrosis. A phase 1 study. J Am Med Assoc 1992;267:1947–51.
- [93] Ali RA, Gandhi AA, Meng H, et al. Adenosine receptor agonism protects against NETosis and thrombosis in antiphospholipid syndrome. Nat Commun 2019;10:1916.
- [94] Liu X, Li Z, Liu S, et al. Potential therapeutic effects of dipyridamole in the severely ill patients with COVID-19. Acta Pharm Sin B 2020;10:1205–15.
- [95] Kanthi Y, Knight JS, Zuo Y, Pinsky DJ. New (re)purpose for an old drug: purinergic modulation may extinguish the COVID-19 thromboinflammatory firestorm. JCI Insight 2020;5.
- [96] Diurno F, Numis FG, Porta G, et al. Eculizumab treatment in patients with COVID-19: preliminary results from real life ASL Napoli 2 Nord experience. Eur Rev Med Pharmacol Sci 2020;24:4040–7.
- [97] Mahajan R, Lipton M, Broglie L, et al. Eculizumab treatment for renal failure in a pediatric patient with COVID-19. J Nephrol 2020 Dec;33(6):1373–6.
- [98] Laurence J, Mulvey JJ, Seshadri M, et al. Anti-complement C5 therapy with eculizumab in three cases of critical COVID-19. Clin Immunol 2020;219:108555.
- [99] Zelek WM, Cole J, Ponsford MJ, et al. Complement inhibition with the C5 blocker LFG316 in severe COVID-19. Am J Respir Crit Care Med 2020 Nov 1;202(9):1304–8.
- [100] Peffault de Latour R, Bergeron A, Lengline E, et al. Complement C5 inhibition in patients with COVID-19 a promising target? Haematologica 2020 Dec 1;105(12):2847–50.
- [101] Mastellos DC, Pires da Silva BGP, Fonseca BAL, et al. Complement C3 vs C5 inhibition in severe COVID-19: early clinical findings reveal differential biological efficacy. Clin Immunol 2020;220:108598.
- [102] Araten DJ, Belmont HM, Schaefer-Cutillo J, et al. Mild clinical course of COVID-19 in 3 patients receiving therapeutic monoclonal antibodies targeting C5 complement for hematologic disorders. Am J Case Rep 2020;21. e927418.

- [103] Kulasekararaj AG, Lazana I, Large J, et al. Terminal complement inhibition dampens the inflammation during COVID-19. Br J Haematol 2020;190:e141–3.
- [104] Pike A, Muus P, Munir T, et al. COVID-19 infection in patients on anti-complement therapy: the Leeds National Paroxysmal Nocturnal Haemoglobinuria service experience. Br J Haematol 2020 Oct;191(1):e1–4.
- [105] Vlaar APJ, de Bruin S, Busch M, et al. Anti-C5a antibody IFX-1 (vilobelimab) treatment versus best supportive care for patients with severe COVID-19 (PANAMO): an exploratory, open-label, phase 2 randomised controlled trial. Lancet Rheumatol 2020 Dec;2(12):e764–73.
- [106] Mastellos DC, Yancopoulou D, Kokkinos P, et al. Compstatin: a C3-targeted complement inhibitor reaching its prime for bedside intervention. Eur J Clin Invest 2015;45:423–40.
- [107] Mastaglio S, Ruggeri A, Risitano AM, et al. The first case of COVID-19 treated with the complement C3 inhibitor AMY-101. Clin Immunol 2020;215:108450.
- [108] Urwyler P, Moser S, Charitos P, et al. Treatment of COVID-19 with conestat alfa, a regulator of the complement, contact activation and kallikrein-kinin system. Front Immunol 2020;11:2072.
- [109] Rambaldi A, Gritti G, Mico MC, et al. Endothelial injury and thrombotic microangiopathy in COVID-19: treatment with the lectin-pathway inhibitor narsoplimab. Immunobiology 2020:152001.