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Progress in Neuropsychopharmacology & Biological Psychiatry

journal homepage: www.elsevier.com/locate/pnp



# COVID-19, ferrosenescence and neurodegeneration, a mini-review

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## ABSTRACT

Exacerbation of cognitive, motor and nonmotor symptoms have been described in critically ill COVID-19 patients, indicating that, like prior pandemics, neurodegenerative sequelae may mark the aftermath of this viral infection. Moreover, SARS-CoV-2, the causative agent of COVID-19 disease, was associated with hyperferritinemia and unfavorable prognosis in older individuals, suggesting virus-induced ferrosenescence.

We have previously defined ferrosenescence as an iron-associated disruption of both the human genome and its repair mechanisms, leading to premature cellular senescence and neurodegeneration. As viruses replicate more efficiently in iron-rich senescent cells, they may have developed the ability to induce this phenotype in host tissues, predisposing to both immune dysfunction and neurodegenerative disorders.

In this mini-review, we summarize what is known about the SARS-CoV-2-induced cellular senescence and iron dysmetabolism. We also take a closer look at immunotherapy with natural killer cells, angiotensin II receptor blockers ("sartans"), iron chelators and dipeptidyl peptidase 4 inhibitors ("gliptins") as adjunct treatments for both COVID-19 and its neurodegenerative complications.

## 1. Introduction

COVID-19 disease is caused by the SARS-CoV-2 virus that, after originating in the city of Wuhan, China, spread rapidly throughout the world. Initially, it was believed that this infection affected only the respiratory tract, but the emergence of gastro-intestinal (GI), neurologic and hematologic symptoms attested to its more systemic nature (Pryce-Roberts et al., 2020). The central nervous system (CNS) involvement, found in nearly 40% of critically ill COVID-19 patients, is heralded by strokes, cognitive dysfunction, depression, psychosis and delirium, suggesting that this virus may predispose to neurodegenerative disorders (Banerjee and Viswanath, 2020).

Although Alzheimer's (AD) or Parkinson's disease (PD) have not been described in association with COVID-19, exacerbation of preexisting cognitive, motor and non-motor symptoms was often observed, indicating viral neurotropism (Banerjee and Viswanath, 2020; Brown et al., 2020; Heneka et al., 2020; Mao et al., 2020; Rogers et al., 2020; Jin et al., 2020). In addition, several prior pandemics have been accompanied by neuropsychiatric sequelae, suggesting that these outcomes could be expected in the aftermath of COVID-19 (Troyer et al., 2020; Tipton and Wszolek, 2020).

The association of unfavorable COVID-19 prognosis with advanced

chronological age, hyperferritinemia and lymphopenia suggests that SARS-CoV-2 may disrupt host antiviral immunity by inducing iron dyshomeostasis (Malavolta et al., 2020; Sargiacomo et al., 2020). As SARS-CoV-2, like other viruses, may be iron-dependent, it likely promotes ferrosenescence to acquire this metal and disable host natural killer cells (NKCs) (Drakesmith and Prentice, 2008; Schmidt, 2020a; Cavezzi et al., 2020; Nuñez and Chana-Cuevas, 2018; Colafrancesco et al., 2020).

In a previous article, we defined ferrosenescence as iron-induced disruption of both DNA and its p53-mediated repair, resulting in premature cellular senescence and neurodegeneration (Sfera et al., 2018). As p53 drives the NKC elimination of senescent and virus-infected cells, SARS-CoV-2 may exploit this protein to optimize thriving. Indeed, decreased p53 and NKCs count were documented in severely ill COVID-19 patients, suggesting virus-usurped immunity (Singh and Bharara, 2020a; Market et al., 2020).

In this mini-review, we examine the SARS-CoV-2 hijacking of NKCs and its link to neurodegeneration via iron-induced genomic and mitochondrial damage. We also take a closer look at NKCs immunotherapy, angiotensin II receptor blockers ("sartans"), iron chelators and dipeptidyl peptidase 4 inhibitors ("gliptins") as adjunct therapies for COVID-19 and neurodegeneration.

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https://doi.org/10.1016/j.pnpbp.2020.110230

Received 14 September 2020; Received in revised form 16 December 2020; Accepted 18 December 2020 Available online 26 December 2020 0278-5846/© 2021 Elsevier Inc. All rights reserved.

## 2. SARS-CoV-2 entry portals and iron dyshomeostasis

New research has shown that SARS-CoV-2 accesses host tissues by usurping several cell membrane and mitochondrial receptors associated with cellular senescence and iron metabolism. Viral exploitation of these proteins, including angiotensin converting enzyme 2 (ACE-2), dipeptidyl peptidase 4 (DPP4), furin and cluster of differentiation 147 (CD 147), disrupts the function of NKCs, facilitating the development of both COVID-19 critical illness and neurodegeneration (Schmidt, 2020a; Cavezzi et al., 2020; Nuñez and Chana-Cuevas, 2018; Colafrancesco et al., 2020) (Radzikowska et al., 2020; Xie et al., 2011). Indeed, as viruses replicate more efficiently in senescent cells, they likely hijack the mechanisms controlling their elimination (Earls et al., 2020; Jadidi-Niaragh et al., 2012; Araga et al., 1991; Mihara et al., 2008).

In the following, we focus on the SARS-CoV-2 entry portals and their association with ferrosenescence-induced NKCs dysfunction.

# 3. ACE-2

The attachment of SARS-CoV-2 virus to ACE-2 protein is mediated by the host transmembrane serine protease 2 (TMPRSS2), an enzyme that primes the S antigen, activating the receptor binding site (RBS) (Hoffmann et al., 2020). SARS-CoV-2/ACE-2 binding likely impairs angiotensin II (ANG II) hydrolysis, leading to its unopposed accumulation (Sfera et al., 2020). Novel preclinical studies have linked ANG II to iron dyshomeostasis as this peptide regulates several iron proteins, including hepcidin, divalent metal transporter 1 (DMT1), ferroportin 1 (Fpn1) and transferrin receptor 1 (TfR1) (Chen et al., 2020; Ishizaka et al., 2007a). In addition, human studies have associated angiotensin receptor blockers (ARBs) with hemoglobin, further connecting ANG II with the release of iron (Mohanram et al., 2008; Cheungpasitporn et al., 2015). Moreover, hyperferritinemia and hemolytic anemia, recently associated with COVID-19 critical illness, also link the virus-upregulated ANG II to iron homeostasis (Pagani et al., 2019; Hindilerden et al., 2020; Lazarian et al., 2020).

Several novel studies have reported that ANG II triggers premature cellular senescence, an iron-abounding phenotype, likely facilitating the SARS-CoV-2 infection (Killilea et al., 2004). Indeed, several other viruses, including human immunodeficiency virus (HIV) and hepatitis B and C were demonstrated to induce cellular senescence by upregulating the host iron (Schmidt, 2020a) (Cohen and Torres, 2017; Idrissi et al., 2016; Naggie and Hepatitis, 2017). Conversely, ARBs were found protective against the senescence-mediated pathology, indicating their potential role in averting both COVID-19 complications and neuro-degeneration (Prusty et al., 2017; Blagosklonny, 2017; Song and Kim, 2019; de Cavanagh et al., 2011).

Taken together, this data suggests that the SARS-CoV-2 virus acquires iron and disrupts NKCs by ferrosenescence. In return, this likely promotes neurodegeneration via the accumulation of senescent cells.

### 4. DPP4

The SARS-CoV-2 entry portal, DPP4, expressed on cell membranes and mitochondria, plays a major role in both type 2 diabetes mellitus (T2DM) and antiviral defenses (Sottile et al., 2019; Jiang et al., 2004; Koshiba et al., 2011; Lee et al., 2020). Like ACE-2, DPP4 has been linked to cellular senescence, suggesting that the SARS-CoV-2 virus thrives by promoting this phenotype. Indeed, previous studies have shown that DPP4 is an aging marker that prompts NKCs to eliminate the senescent cells expressing this protein (Kim et al., 2017). Viral attachment to DPP4 likely alters the receptor configuration, contributing to the accumulation of uncleared senescent cells (Klemann et al., 2016). In this regard, earlier studies have associated NKCs dysfunction with neurodegenerative disorders, including PD and AD (Earls et al., 2020) (Maghazachi, 2013a; Martínez-Cué and Rueda, 2020). ferroptosis, an iron-mediated nonapoptotic cell death, associated with both COVID-19 critical illness and neurodegeneration (Cavezzi et al., 2020) (Edeas et al., 2020a; Li et al., 2020). Under normal circumstances, ferroptosis is blocked by the p53 attachment to DPP4, suggesting that SARS-CoV-2 exploitation of this protein likely enables both viral infection and neurodegeneration (Xie et al., 2017; Zhang and Chen, 2019a; Liu et al., 2020a; Shen et al., 2014; Tarangelo and Dixon, 2018)(Fig. 1). Since p53 is instrumental for NKCs antiviral function, the virus may have developed a backup mechanism for the exploitation of this protein, utilizing two avenues, a direct disruption (via the S2 antigen) and an indirect one (via iron upregulation) (Xie et al., 2017) (Shen et al., 2014) (Fig. 2). On the other hand, DPP4 inhibitors (gliptins) are promising treatments for both COVID-19 and neurodegenerative disorders as they inhibit ferroptosis and restore the integrity of p53 (Angelopoulou and Piperi, 2018; Svenningsson et al., 2016; Solerte et al., 2020).

# 5. CD147

Another SARS-CoV-2 entry portal, CD147 or basigin, is expressed on both the host cell surface and mitochondria, an organelle essential for antiviral immunity (Ulrich and Pillat, 2020; Pablo and Ochrietor, 2013; Luo et al., 2014). In addition, CD147 regulates mitochondrial metabolic reprograming and the generation of lactate (Schneiderhan et al., 2009). Excessive lactate levels have been associated with NKCs dysfunction, linking CD147 to COVID-19 critical illness (Calvisi, 2016; Scott and Cleveland, 2016). Interestingly, CD147 and lactate dehydrogenase (LDH) are colocalized with the mitochondrial reticulum, suggesting that SARS-CoV-2/CD147 attachment could augment the release of LDH, a marker of COVID-19 mortality (Henry et al., 2020; Hashimoto et al., 2006). Moreover, CD147 is also the entry portal of Plasmodium falciparum, an iron-dependent pathogen associated with premature cellular senescence, further emphasizing the role of this phenotype in COVID-19 (Asghar et al., 2018; Frimpong et al., 2019; Clark et al., 2014; Huang et al., 2014). Interestingly, antimalarial drugs, chloroquine and hydroxychloroquine may block SARS-CoV-2/CD147 attachment, preventing iron-related pathology (Quiros Roldan et al., 2020).

Taken together, SARS-CoV-2/CD147 binding likely triggers ferrosenescence that, in turn damages the mitochondrion, disrupting NKCsmediated antiviral immunity.

#### 6. Furin

It was recently reported that SARS-CoV-2 exploits another host protein, furin that cleaves the S antigen into S2, a mediator of cell-to-cell fusion and a likely facilitator of viral migration from infected to noninfected cells (Hoffmann et al., 2020) (Xia et al., 2020). Interestingly, furin is a proprotein convertase with numerous physiological functions, including the conversion of pro-hepcidin to active hepcidin, further linking COVID-19 to iron metabolism (Wichaiyo et al., 2015). In addition, as mentioned above, the S2 antigen of SARS-CoV-2 may directly inhibit p53, sabotaging the elimination of virus-infected, senescent and possibly cancer cells (Singh and Bharara, 2020b; Zhang and Chen, 2019b; Ma-Lauer et al., 2016; Aloni-Grinstein et al., 2018; Jyotsana and King, 2020; Dai et al., 2020; Moujaess et al., 2020). Moreover, as p53 participates in both DNA damage repair (DDR) and senescent cells elimination, SARS-CoV-2 exploitation of this protein may predispose to neurodegeneration (Sargiacomo et al., 2020) (Funauchi et al., 2015; Collin et al., 2018; Kuo et al., 2020). Indeed, dysfunctional DDR and genomic damage have been associated with cellular senescence and neurodegenerative disorders (Madabhushi et al., 2014). Furthermore, inefficient DDR may lead to spillage of damaged DNA or mtDNA into the cytosol and the peripheral blood, probably accounting for this recently described COVID-19 marker (Liu, 2020).

Aside from cellular senescence, DPP4 has been directly linked to

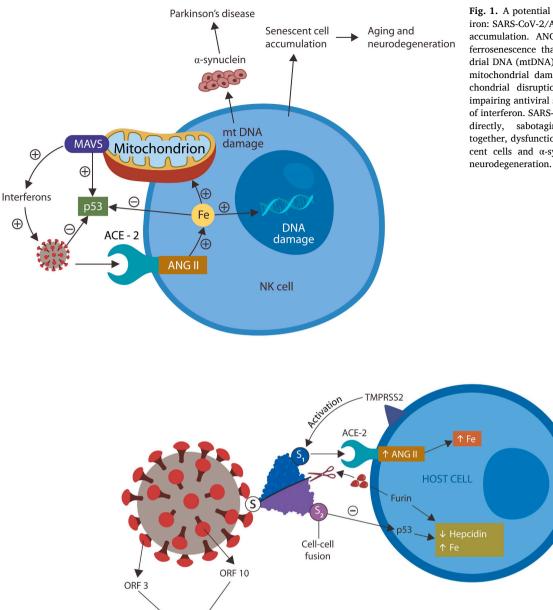


Fig. 1. A potential mechanism of NKCs hijacking by iron: SARS-CoV-2/ACE-2 attachment leads to ANG II accumulation. ANG II upregulates iron, inducing ferrosenescence that damages the DNA, mitochondrial DNA (mtDNA) and p53. ANG II can also inflict mitochondrial damage directly (not shown). Mitochondrial disruption promotes viral infection by impairing antiviral signaling (MAVS) and the release of interferon. SARS-CoV-2 antigen S2 can inhibit p53 directly, sabotaging viral elimination. Taken together, dysfunctional NKCs fail to eliminate senescent cells and a-synuclein, increasing the risk of

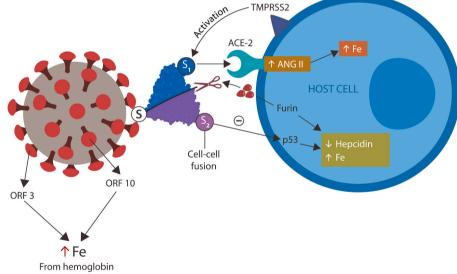


Fig. 2. The SARS-CoV-2-induced ferrosenescence: S antigen usurps host protease TMPRSS2 to prime this protein to S1, enabling ACE-2 attachment. Unopposed ANG II accumulation, augments intracellular iron, inducing DNA and mitochondrial DNA (mtDNA) damage. Furin cleaves, the S antigen into S2, a facilitator of cell-to-cell fusion and a p53 inhibitor. Furin also upregulates the iron protein hepcidin, increasing intracellular iron, further damaging the genome.

#### 7. Viral accessory proteins

Several SARS-CoV-2 accessory viral proteins, including open reading frame (ORF) 3, 10 and possibly 8 were found to facilitate viral infection and iron dyshomeostasis via iron release from hemoglobin (Cavezzi et al., 2020) (Read, 2020). Indeed, novel studies have suggested that ORF proteins may cause the hemolytic anemia documented in severely ill COVID-19 patients (Lazarian et al., 2020) (Lippi and Mattiuzzi, 2020).

# 8. SARS-CoV-2 hijacking of NKCs

Several new studies have found a direct correlation between the COVID-19 prognosis and host hyperferritinemia or lymphopenia,

suggesting that the virus likely exploits iron metabolism to disrupt NKCs antiviral function (Bolondi et al., 2020; Perricone et al., 2020; Edeas et al., 2020b; Vargas-Vargas and Cortés-Rojo, 2020; Gómez-Pastora et al., 2020). Others have reported that hyperferritinemia and premature cellular senescence predispose to both COVID-19 critical illness and neurodegenerative disorders, indicating a role for iron in both conditions (Sottile et al., 2019) (Masaldan et al., 2018; Kale et al., 2020). Conversely, iron deprivation has been shown to lower the viral load and improve prognosis in many viral infections (Liu et al., 2020b; Schmidt, 2020b).

NKCs are vigilant innate immune lymphocytes capable of locating and eliminating senescent, cancer and virus-infected cells without prior sensitization by the major histocompatibility complex (MHC) (Market et al., 2020) (Masselli et al., 2020). Inefficient senescent cells clearance

was demonstrated to dysregulate both neighboring cells and the circulating lymphocytes, increasing the chance of neurodegeneration (Pereira et al., 2019). Indeed, a recent PD preclinical study found that aside from senescent cells, NKCs also clear  $\alpha$ -synuclein, connecting their impairment to neurodegeneration (Earls et al., 2020).

SARS-CoV-2 entry portals have been directly linked to the NKCsdriven elimination of senescent and virus-infected cells, suggesting that this virus triggers ferrosenescence to usurp host immunity (Jurewicz et al., 2007; Abadir et al., 2011). As some of these proteins are expressed on the mitochondrion and are crucial for NKCs antiviral immunity, their exploitation by the SARS-CoV-2, effectively hands control of these lymphocytes to the virus.

Under normal circumstances, the viral-host contact prompts the mitochondrion to release interferons via mitochondrial antiviral signaling (MAVS) protein (Kim et al., 2018). MAVS interaction with p53 activates the NKCs-driven elimination of senescent and virus-infected cells, therefore by exploiting this protein, SARS-CoV-2 hijacks the entire host antiviral defenses (Zhang et al., 2020; Mijit et al., 2020a). Indeed, viral infections and neurodegeneration have been directly corelated with mitochondrial damage (Maghazachi, 2013a) (van Erp et al., 2019; Cowan et al., 2019).

Taken together, SARS-CoV-2-induced ferrosenescence in NKCs, facilitates senescent cells accumulation and viral thriving, supporting the use of exogenous lymphocytes in the treatment of COVID-19 (NCT04365101) (NCT04344548) and neurodegenerative disorders (Earls et al., 2020) (Maghazachi, 2013a) (Song et al., 2020; Machhi et al., 2020).

#### 9. Interventions

Several therapeutic modalities have been developed and are currently in clinical trials for COVID-19, including immunotherapy with NKC, ARBs, iron chelators and DPP4 inhibitors. Aside from protecting against SARS-CoV-2-related cognitive, motor and nonmotor symptoms, these interventions could address the symptoms of neurodegenerative disorders of other etiologies (Fig. 3).

### 10. NKCs immunotherapy

Live, exogenous NKCs derived from the peripheral blood or stem cell precursors are currently in clinical trials for COVID-19 as they likely enhance host antiviral defenses and senescent cells clearance (Market et al., 2020) (Hu et al., 2019) (NCT04344548). NKCs immunotherapy is easily administered as these lymphocytes do not require complex HLA matching and can be augmented by the addition of immune stimulatory molecules (Shimasaki et al., 2020).

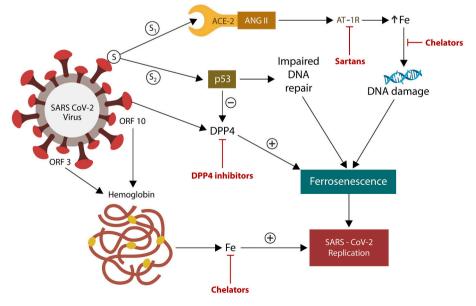
This therapeutic modality has proved effective in amyotrophic lateral sclerosis and multiple sclerosis and is currently being assessed for other neurodegenerative disorders (Maghazachi, 2013b; Garofalo et al., 2020). Indeed, the NKCs immunotherapy could benefit patients with PD as, like influenza H5N1 and H1N1 viruses, SARS-CoV-2 may preferentially target the dopaminergic neurons in substantia nigra (Eldeeb et al., 2020). In addition, as NKCs accelerate the elimination of senescent cells and  $\alpha$ -synuclein, they may comprise a "generic" treatment for multiple neurodegenerative disorders (Earls et al., 2020).

#### 11. Sartans

Preclinical studies have shown that ANG II may interfere with iron absorption, suggesting that early interventions at this level could prevent the development of COVID-19 critical illness (Tajima et al., 2010).

The non-heme iron is absorbed in the duodenum via DMT1 transporter, a protein controlled by the SLC11A1 gene that also regulates NKCs activation (Su et al., 2020; Hedges et al., 2013; Czachorowski et al., 2009; Awomoyi, 2007). For this reason, SARS-CoV-2 exploitation of this gene likely induces iron dyshomeostasis and immune impairment. Conversely, preclinical studies have found that ARBs suppress DMT1 expression, suggesting a protective effect against SARS-CoV-2 at the gut level (Baral et al., 2020; Ishizaka et al., 2007b). In addition, ARBs were demonstrated to possess antiviral properties against other viruses, including dengue and HCV, suggesting that DMT1 inhibition may prevent viral usurpation of iron homeostasis (Baral et al., 2020; Ishizaka et al., 2007b; Eslami et al., 2014; Hernández-Fonseca et al., 2015; Colmenero et al., 2009; Mak et al., 2012; Saavedra, 2020). ARBs are currently in clinical trials against COVID-19 but their early or preventive use could block viral hijacking of SLC11A1 (NCT04335123) (NCT04312009)(NCT04311177). In addition, preclinical studies have found that ARBs may protect against PD, probably by DMT1 and ferrosenescence inhibition (Mertens et al., 2011; Villapol and Saavedra, 2015; Reardon et al., 2000; Lee et al., 2014; Perez-Lloret et al., 2017; Gebre et al., 2018). Furthermore, as ARBs were demonstrated to upregulate p53 and restore DDR, they may slower the development of cellular and immune senescence, lowering the risk of neurodegeneration (Mijit et al., 2020b; Gong et al., 2010; Blagosklonny, 2018).

> **Fig. 3.** The likely action mechanism of "sartans", "gliptins" and iron chelators is schematically illustrated along with viral ferrosenescence mechanisms. The SARS-CoV-2 virus upregulates intracellular iron (via angiotensin II accumulation), triggering DNA damage in host NKCs. In addition, the virus blocks p53, disrupting DNA damage repair (DDR). These two actions trigger ferrosenescence, facilitating viral replication. Viral accessory proteins attack hemoglobin, further increasing iron levels.



# 12. Iron chelators

Iron chelators, including deferoxamine, deferiprone and deferasirox possess antiviral actions against several viruses, including HIV, West Nile and HCV (Duchemin and Paradkar, 2017; Theurl et al., 2004; Georgiou et al., 2000; Williams and Meyer, 2009). As RNA viruses are more sensitive to iron chelators, they may be beneficial for COVID-19 and are currently in clinical trials for this condition (NCT04333550) (Dalamaga et al., 2020). Moreover, a recently developed, potent and natural iron chelator, DIBI, appears a likely SARS-CoV-2 treatment candidate, especially as it is well tolerated by older individuals (Thorburn et al., 2017; Kontoghiorghes and Kontoghiorghe, 2020). Several novel studies have reported that aside from denying iron to viruses, chelators also augment NKCs function, suggesting that they act both as antiviral and immune stimulatory agents (Sherman and Spear, 1993). Furthermore, iron chelators may upregulate p53, augmenting viral and senescent cells elimination, a role that has brought them into the field of neurodegenerative disorders (Zhang and Chen, 2019b; Ma-Lauer et al., 2016) (Muñoz-Fontela et al., 2011; Liang and Richardson, 2003). The SARS-CoV-2 antigen S2 appears to directly block p53, likely triggering ferroptosis, a cell death believed to be involved in both COVID-19 and neurodegeneration (Singh and Bharara, 2020a).

Taken together, iron chelators may inhibit SARS-CoV-2 by withholding iron from the virus, upregulating p53 and the NKCs antiviral function.

## 13. Gliptins

Gliptins, or DPP4 inhibitors are potential COVID-19 treatments currently in clinical trials as they are believed to reduce viral entry (Solerte et al., 2020) (NCT04365517). As gliptins may lower ferroptosis, a phenomenon that plays a major role in AD and PD, these agents may prove beneficial for neurodegenerative disorders (Mousa and Ayoub, 2019; Guiney et al., 2017; Yan and Zhang, 2020). In addition, since DPP4 marks senescent cells, their recognition and elimination by NKCs may avert accumulation (Kim et al., 2017).

Recently DPP4 has been associated with physical exhaustion and chronic fatigue syndrome (CFS), a major complaint of patients with viral infections, including COVID-19, suggesting that DPP4 inhibitors may ameliorate this pathology (Qi et al., 2020; Bühling et al., 1994; Fletcher et al., 2010; Wilson, 2020; Perrin et al., 2020; Silvestre et al., 2019). Several studies have associated CFS with ferroptosis and lipid peroxidation, indicating that iron may play a major role in the pathogenesis of CFS (Brkic et al., 2010; Swinkels et al., 2002). Indeed, cancer studies have connected ferroptosis to the p53-mediated DPP4 inhibition, suggesting that gliptins should be evaluated as CFS treatments (Xie et al., 2017) (Morris and Maes, 2012; Zhou et al., 2020).

Taken together, ferroptosis prevention with DPP4 inhibitors could preempt viral entry, fatigue and the development of neurodegeneration.

# 14. Conclusions

Like other viruses, SARS-CoV-2 may thrive in iron-rich senescent cells, a phenotype it likely promotes. Iron is a major senescence inducer as it disrupts both the DNA and its p53-mediated repair, triggering ferrosenescence. The SARS-CoV-2 virus may have developed the ability to evade host detection by triggering this phenotype in NKCs, disrupting their vigilance.

SARS-CoV-2 exploitation of iron metabolism likely begins at the level of GI absorption by the diversion of SLC11A1 gene, a regulator of both ferroptosis and NKCs activation. Iron metabolism is further usurped at viral entry portals, enabling infection and contributing to neurodegeneration. Because of these reasons, individuals at risk for COVID-19 complications should be started on NKCs immunotherapy and/or antiferrosenescence treatment early after the diagnosis.

#### Authors statement

All authors contributed equally to this manuscript.

#### **Declaration of Competing Interest**

The authors have no conflict of interest and no funding was received for this article.

# References

- Abadir, P.M., Foster, D.B., Crow, M., et al., 2011. Identification and characterization of a functional mitochondrial angiotensin system. Proc. Natl. Acad. Sci. U. S. A. 108 (36), 14849–14854. https://doi.org/10.1073/pnas.1101507108.
- Aloni-Grinstein, R., Charni-Natan, M., Solomon, H., Rotter, V., 2018. p53 and the viral connection: back into the future ‡. Cancers (Basel) 10 (6), 178. Published 2018 Jun 4. https://doi.org/10.3390/cancers10060178.
- Angelopoulou, E., Piperi, C., 2018. DPP-4 inhibitors: a promising therapeutic approach against Alzheimer's disease. Ann. Transl. Med. 6 (12), 255. https://doi.org/ 10.21037/atm.2018.04.41.
- Araga, S., Kagimoto, H., Funamoto, K., Takahashi, K., 1991. Reduced natural killer cell activity in patients with dementia of the Alzheimer type. Acta Neurol. Scand. 84 (3), 259–263. https://doi.org/10.1111/j.1600-0404.1991.tb04948.x.
- Asghar, M., Yman, V., Homann, M.V., et al., 2018. Cellular aging dynamics after acute malaria infection: a 12-month longitudinal study. Aging Cell 17 (1), e12702. https:// doi.org/10.1111/acel.12702.
- Awomoyi, A.A., 2007. The human solute carrier family 11 member 1 protein (SLC11A1): linking infections, autoimmunity and cancer? FEMS Immunol. Med. Microbiol. 49 (3), 324–329. https://doi.org/10.1111/j.1574-695X.2007.00231.x.
- Banerjee, D., Viswanath, B., 2020. Neuropsychiatric manifestations of COVID-19 and possible pathogenic mechanisms: insights from other coronaviruses. Asian J. Psychiatr. 54, 102350. https://doi.org/10.1016/j.ajp.2020.102350.
- Baral, R., White, M., Vassilios, S., Vassiliou, V.V., 2020. Effect of renin-angiotensinaldosterone system inhibitors in patients with COVID-19: a systematic review and meta-analysis of 28,872 patients. Curr. Atheroscler. Rep. 22, 61. https://doi.org/ 10.1007/s11883-020-00880-6.
- Blagosklonny, M.V., 2017. From rapalogs to anti-aging formula. Oncotarget 8 (22), 35492–35507. https://doi.org/10.18632/oncotarget.18033.
- Blagosklonny, M.V., 2018. Disease or not, aging is easily treatable. Aging (Albany NY) 10 (11), 3067–3078. https://doi.org/10.18632/aging.101647.
- Bolondi, G., Russo, E., Gamberini, E., et al., 2020. Iron metabolism and lymphocyte characterisation during Covid-19 infection in ICU patients: an observational cohort study. World J. Emerg. Surg. 15 (1), 41. Published 2020 Jun 30. https://doi. org/10.1186/s13017-020-00323-2.
- Brkic, S., Tomic, S., Maric, D., Novakov Mikic, A., Turkulov, V., 2010. Lipid peroxidation is elevated in female patients with chronic fatigue syndrome. Med. Sci. Monit. 16 (12), CR628–CR632.
- Brown, E.G., Chahine, L.M., Goldman, S.M., Korell, M., Mann, E., Daniel, R., et al., 2020. The Effect of the COVID-19 Pandemic on People with Parkinson's Disease. medRxiv. https://doi.org/10.1101/2020.07.14.20153023, 07.14.20153023.
- Bühling, F., Kunz, D., Reinhold, D., et al., 1994. Expression and functional role of dipeptidyl peptidase IV (CD26) on human natural killer cells. Nat. Immun. 13 (5), 270–279.
- Calvisi, D.F., 2016. CD147/Basigin: a Warburg oncogene in hepatocellular carcinoma? Chin. J. Cancer Res. 28 (3), 377–379. https://doi.org/10.21147/j.issn.1000-9604.2016.03.13.
- Cavezzi, A., Troiani, E., Corrao, S., 2020. COVID-19: hemoglobin, iron, and hypoxia beyond inflammation. A narrative review. Clin. Pract. 10 (2), 1271. https://doi.org/ 10.4081/cp.2020.1271.
- Chen, Y.J., Qian, Z.M., Sheng, Y., Zheng, J., Liu, Y., 2020. Angiotensin II down-regulates transferrin receptor 1 and ferroportin 1 expression in neuro-2a cells via activation of type-1 receptor. Neurosci. Lett. 716, 134684. https://doi.org/10.1016/j. neulet.2019.134684.
- Cheungpasitporn, W., Thongprayoon, C., Chiasakul, T., Korpaisarn, S., Erickson, S.B., 2015. Renin-angiotensin system inhibitors linked to anemia: a systematic review and meta-analysis. QJM: An International Journal of Medicine 108 (11), 879–884. https://doi.org/10.1093/gjmed/hcv049.
- Clark, M.A., Goheen, M.M., Cerami, C., 2014. Influence of host iron status on plasmodium falciparum infection. Front. Pharmacol. 5, 84. https://doi.org/ 10.3389/fphar.2014.00084.
- Cohen, J., Torres, C., 2017. HIV-associated cellular senescence: a contributor to accelerated aging. Ageing Res. Rev. 36, 117–124. https://doi.org/10.1016/j. arr.2016.12.004.
- Colafrancesco, S., Alessandri, C., Conti, F., Priori, R., 2020. COVID-19 gone bad: a new character in the spectrum of the hyperferritinemic syndrome? Autoimmun. Rev. 19 (7), 102573. https://doi.org/10.1016/j.autrev.2020.102573.
- Collin, G., Huna, A., Warnier, M., Flaman, J.M., Bernard, D., 2018. Transcriptional repression of DNA repair genes is a hallmark and a cause of cellular senescence. Cell Death Dis. 9 (3), 259. Published 2018 Feb 15. https://doi.org/10.1038/s41419-0 18-0300-z.
- Colmenero, J., Bataller, R., Sancho-Bru, P., et al., 2009. Effects of losartan on hepatic expression of nonphagocytic NADPH oxidase and fibrogenic genes in patients with

chronic hepatitis C. Am. J. Physiol. Gastrointest. Liver Physiol. 297 (4), G726–G734. https://doi.org/10.1152/ajpgi.00162.2009.

Cowan, K., Anichtchik, O., Luo, S., 2019. Mitochondrial integrity in neurodegeneration. CNS Neurosci. Ther. 25 (7), 825–836. https://doi.org/10.1111/cns.13105.

Czachorowski, M., Lam-Yuk-Tseung, S., Cellier, M., Gros, P., 2009. Transmembrane topology of the mammalian Slc11a2 iron transporter. Biochemistry 48 (35), 8422–8434. https://doi.org/10.1021/bi900606y.

Dai, M., Liu, D., Liu, M., et al., 2020. Patients with cancer appear more vulnerable to SARS-CoV-2: a multicenter study during the COVID-19 outbreak. Cancer Discov. 10 (6), 783–791. https://doi.org/10.1158/2159-8290.CD-20-0422.

Dalamaga, M., Karampela, I., Mantzoros, C.S., 2020. Commentary: could iron chelators prove to be useful as an adjunct to COVID-19 treatment regimens? Metabolism 108, 154260. https://doi.org/10.1016/j.metabol.2020.154260.

de Cavanagh, E.M., Inserra, F., Ferder, L., 2011. Angiotensin II blockade: a strategy to slow ageing by protecting mitochondria? Cardiovasc. Res. 89 (1), 31–40. https://doi. org/10.1093/cvr/cvq285.

Drakesmith, H., Prentice, A., 2008. Viral infection and iron metabolism. Nat. Rev. Microbiol. 6 (7), 541–552. https://doi.org/10.1038/nrmicro1930.

Duchemin, J.B., Paradkar, P.N., 2017. Iron availability affects West Nile virus infection in its mosquito vector. Virol. J. 14 (1), 103. Published 2017 Jun 5. https://doi. org/10.1186/s12985-017-0770-0.

Earls, R.H., Menees, K.B., Chung, J., et al., 2020. NK cells clear α-synuclein and the depletion of NK cells exacerbates synuclein pathology in a mouse model of α-synucleinopathy. Proc. Natl. Acad. Sci. U. S. A. 117 (3), 1762–1771. https://doi. org/10.1073/pnas.1909110117.

Edeas, M., Saleh, J., Peyssonnaux, C., 2020a. Iron: innocent bystander or vicious culprit in COVID-19 pathogenesis? Int. J. Infect. Dis. 97, 303–305. https://doi.org/ 10.1016/j.ijid.2020.05.110.

Edeas, M., Saleh, J., Peyssonnaux, C., 2020b. Iron: innocent bystander or vicious culprit in COVID-19 pathogenesis? Int. J. Infect. Dis. 97, 303–305. https://doi.org/ 10.1016/i.iiid.2020.05.110.

Eldeeb, M.A., Hussain, F.S., Siddiqi, Z.A., 2020. COVID-19 infection may increase the risk of parkinsonism - remember the Spanish flu? [published online ahead of print, 2020 Jun 7]. Cytokine Growth Factor Rev. S1359-6101 (20), 30142–30148. https://doi. org/10.1016/j.cytogfr.2020.06.009.

Eslami, H., Sharifi, A.M., Rahimi, H., Rahati, M., 2014. Protective effect of telmisartan against oxidative damage induced by high glucose in neuronal PC12 cell. Neurosci. Lett. 558, 31–36. https://doi.org/10.1016/j.neulet.2013.10.057.

Fletcher, M.A., Zeng, X.R., Maher, K., Levis, S., Hurwitz, B., Antoni, M., et al., 2010. Biomarkers in chronic fatigue syndrome: evaluation of natural killer cell function and dipeptidyl peptidase IV/CD26. PLoS One 5 (5), e10817. https://doi.org/ 10.1371/journal.pone.0010817.

Frimpong, A., Kusi, K.A., Adu-Gyasi, D., Amponsah, J., Ofori, M.F., Ndifon, W., 2019. Phenotypic evidence of T cell exhaustion and senescence during symptomatic plasmodium falciparum malaria. Front. Immunol. 10, 1345. https://doi.org/ 10.3389/fimmu.2019.01345.

Funauchi, Y., Tanikawa, C., Yi Lo, P.H., et al., 2015. Regulation of iron homeostasis by the p53-ISCU pathway. Sci. Rep. 5, 16497. Published 2015 Nov 12. https://doi.org/ 10.1038/srep16497.

Garofalo, S., Cocozza, G., Porzia, A., et al., 2020. Natural killer cells modulate motor neuron-immune cell cross talk in models of amyotrophic lateral sclerosis. Nat. Commun. 11, 1773. https://doi.org/10.1038/s41467-020-15644-8.

Gebre, A.K., Altaye, B.M., Atey, T.M., Tuem, K.B., Berhe, D.F., 2018. Targeting reninangiotensin system against Alzheimer's disease. Front. Pharmacol. 9, 440. https:// doi.org/10.3389/fphar.2018.00440.

Georgiou, N.A., van der Bruggen, T., Oudshoorn, M., Nottet, H.S., Marx, J.J., van Asbeck, B.S., 2000. Inhibition of human immunodeficiency virus type 1 replication in human mononuclear blood cells by the iron chelators deferoxamine, deferiprone, and bleomycin. J. Infect. Dis. 181 (2), 484–490.

Gómez-Pastora, J., Weigand, M., Kim, J., et al., 2020. Hyperferritinemia in critically ill COVID-19 patients - is ferritin the product of inflammation or a pathogenic mediator? [published online ahead of print, 2020 Jun 21]. Clin. Chim. Acta 509, 249–251. https://doi.org/10.1016/j.cca.2020.06.033.

Gong, Q., Davis, M., Chipitsyna, G., Yeo, C.J., Arafat, H.A., 2010. Blocking angiotensin II Type 1 receptor triggers apoptotic cell death in human pancreatic cancer cells. Pancreas 39 (5), 581–594. https://doi.org/10.1097/MPA.0b013e3181c314cd.

Guiney, S.J., Adlard, P.A., Bush, A.I., Finkelstein, D.I., Ayton, S., 2017. Ferroptosis and cell death mechanisms in Parkinson's disease. Neurochem. Int. 104, 34–48. https:// doi.org/10.1016/j.neuint.2017.01.004.

Hashimoto, T., Hussien, R., Brooks, G.A., 2006. Colocalization of MCT1, CD147, and LDH in mitochondrial inner membrane of L6 muscle cells: evidence of a mitochondrial lactate oxidation complex. Am. J. Physiol. Endocrinol. Metab. 290 (6), E1237–E1244. https://doi.org/10.1152/ajpendo.00594.2005.

Hedges, J.F., Kimmel, E., Snyder, D.T., Jerome, M., Jutila, M.A., 2013. Solute carrier 11A1 is expressed by innate lymphocytes and augments their activation. J. Immunol. 190 (8), 4263–4273. https://doi.org/10.4049/jimmunol.1200732.

Heneka, M.T., Golenbock, D., Latz, E., Morgan, D., Brown, R., 2020. Immediate and longterm consequences of COVID-19 infections for the development of neurological disease. Alzheimers Res. Ther. 12 (1), 69. Published 2020 Jun 4. https://doi. org/10.1186/s13195-020-00640-3.

Henry, B.M., Aggarwal, G., Wong, J., et al., 2020. Lactate dehydrogenase levels predict coronavirus disease 2019 (COVID-19) severity and mortality: a pooled analysis [published online ahead of print, 2020 May 27]. Am. J. Emerg. Med. 38 (9), 1722–1726. https://doi.org/10.1016/j.ajem.2020.05.073.

Hernández-Fonseca, J.P., Durán, A., Valero, N., Mosquera, J., 2015. Losartan and enalapril decrease viral absorption and interleukin 1 beta production by macrophages in an experimental dengue virus infection. Arch. Virol. 160 (11), 2861–2865. https://doi.org/10.1007/s00705-015-2581-1.

Hindilerden, F., Yonal-Hindilerden, I., Akar, E., Yesilbag, Z., Kart-Yasar, K., 2020. Severe autoimmune hemolytic anemia in COVID-19 infection, safely treated with steroids. Mediterr. J. Hematol. Infect. Dis. 12 (1) https://doi.org/10.4084/MJHID.2020.053 e2020053. Published 2020 Jul 1.

Hoffmann, M., Kleine-Weber, H., Schroeder, S., et al., 2020. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. Cell 181 (2), 271–280 e8. https://doi.org/10.1016/j.cell.2020.02.052.

Hu, W., Wang, G., Huang, D., Sui, M., Xu, Y., 2019. Cancer immunotherapy based on natural killer cells: current progress and new opportunities. Front. Immunol. 10, 1205. Published 2019 May 31. https://doi.org/10.3389/fimmu.2019.01205.

Huang, Q., Li, J., Xing, J., et al., 2014. CD147 promotes reprogramming of glucose metabolism and cell proliferation in HCC cells by inhibiting the p53-dependent signaling pathway. J. Hepatol. 61 (4), 859–866. https://doi.org/10.1016/j. jhep.2014.04.035.

Idrissi, M.E., Hachem, H., Koering, C., et al., 2016. HBx triggers either cellular senescence or cell proliferation depending on cellular phenotype. J. Viral Hepat. 23 (2), 130–138. https://doi.org/10.1111/jvh.12450.

Ishizaka, N., Saito, K., Furuta, K., et al., 2007a. Angiotensin II-induced regulation of the expression and localization of iron metabolism-related genes in the rat kidney. Hypertens. Res. 30 (2), 195–202. https://doi.org/10.1291/hypres.30.195.

Ishizaka, N., Saito, K., Furuta, K., et al., 2007b. Angiotensin II-induced regulation of the expression and localization of iron metabolism-related genes in the rat kidney. Hypertens. Res. 30 (2), 195–202. https://doi.org/10.1291/hypres.30.195.

Jadidi-Niaragh, F., Shegarfi, H., Naddafi, F., Mirshafiey, A., 2012. The role of natural killer cells in Alzheimer's disease. Scand. J. Immunol. 76 (5), 451–456. https://doi. org/10.1111/j.1365-3083.2012.02769.x.

Jiang, X., Hsu, K., Denicola, D., Head, J.F., Elliott, R.L., 2004. Iron impairs natural killer cell function. Cancer Res. 64 (7).

Jin, H., Hong, C., Chen, S., Zhou, Y., Wang, Y., Mao, L., et al., 2020. Consensus for prevention and management of coronavirus disease 2019 (COVID-19) for neurologists. Stroke Vasc. Neurol. 2, 146–152. https://doi.org/10.1136/svn-2020 -000382.

Jurewicz, M., McDermott, D.H., Sechler, J.M., et al., 2007. Human T and natural killer cells possess a functional renin-angiotensin system: further mechanisms of angiotensin II-induced inflammation. J. Am. Soc. Nephrol. 18 (4), 1093–1102. https://doi.org/10.1681/ASN.2006070707.

Jyotsana, N., King, M.R., 2020. The impact of COVID-19 on cancer risk and treatment [published online ahead of print, 2020 Jun 29]. Cell. Mol. Bioeng. 1–7. https://doi. org/10.1007/s12195-020-00630-3.

Kale, A., Sharma, A., Stolzing, A., Desprez, P.Y., Campisi, J., 2020. Role of immune cells in the removal of deleterious senescent cells. Immun. Ageing 17, 16. https://doi.org/ 10.1186/s12979-020-00187-9.

Killilea, D.W., Wong, S.L., Cahaya, H.S., Atamna, H., Ames, B.N., 2004. Iron accumulation during cellular senescence. Ann. N. Y. Acad. Sci. 1019, 365–367. https://doi.org/10.1196/annals.1297.063.

Kim, K.M., Noh, J.H., Bodogai, M., et al., 2017. Identification of senescent cell surface targetable protein DPP4. Genes Dev. 31 (15), 1529–1534. https://doi.org/10.1101/ gad.302570.117.

Kim, S.J., Ahn, D.G., Syed, G.H., Siddiqui, A., 2018. The essential role of mitochondrial dynamics in antiviral immunity. Mitochondrion 41, 21–27. https://doi.org/ 10.1016/i.mito.2017.11.007.

Klemann, C., Wagner, L., Stephan, M., von Hörsten, S., 2016. Cut to the chase: a review of CD26/dipeptidyl peptidase-4's (DPP4) entanglement in the immune system. Clin. Exp. Immunol. 185 (1), 1–21. https://doi.org/10.1111/cei.12781.

Kontoghiorghes, G.J., Kontoghiorghe, C.N., 2020. Iron and chelation in biochemistry and medicine: new approaches to controlling iron metabolism and treating related diseases. Cells 9 (6), 1456. Published 2020 Jun 12. https://doi.org/10.3390/cell s9061456.

Koshiba, T., Bashiruddin, N., Kawabata, S., 2011. Mitochondria and antiviral innate immunity. Int J Biochem Mol Biol 2 (3), 257–262.

Kuo, C.L., Pilling, L.C., Atkins, J.C., et al., 2020. COVID-19 Severity is Predicted by Earlier Evidence of Accelerated Aging. Preprint. medRxiv. https://doi.org/10.1101/ 2020.07.10.20147777, 2020.07.10.20147777.

Lazarian, G., Quinquenel, A., Bellal, M., et al., 2020. Autoimmune haemolytic anaemia associated with COVID-19 infection. Br. J. Haematol. 190 (1), 29–31. https://doi. org/10.1111/bjh.16794.

Lee, Y.C., Lin, C.H., Wu, R.M., Lin, J.W., Chang, C.H., Lai, M.S., 2014. Antihypertensive agents and risk of Parkinson's disease: a nationwide cohort study. PLoS One 9 (6), e98961. https://doi.org/10.1371/journal.pone.0098961.

Lee, S., Wu, S., Su, M., Liang, Y., Ku, H., 2020. Dipeptidyl peptidase-4 involved in regulating mitochondria function in cardiomyocytes through Nrf2 and PGC-1α signaling. Res. Square. https://doi.org/10.21203/rs.3.rs-30722/v1.

Li, J., Cao, F., Yin, H., et al., 2020. Ferroptosis: past, present and future. Cell Death Dis. 11, 88. https://doi.org/10.1038/s41419-020-2298-2.

Liang, S.X., Richardson, D.R., 2003. The effect of potent iron chelators on the regulation of p53: examination of the expression, localization and DNA-binding activity of p53 and the transactivation of WAF1. Carcinogenesis 24 (10), 1601–1614. https://doi. org/10.1093/carcin/bgg116.

Lippi, G., Mattiuzzi, C., 2020. Hemoglobin value may be decreased in patients with severe coronavirus disease 2019. Hematol. Transfus. Cell. Ther. 42 (2), 116–117. https://doi.org/10.1016/j.htct.2020.03.001.

Liu, B., 2020. Free DNA, a reason for severe COVID-19 infection? [published online ahead of print, 2020 may 5]. Med. Hypotheses 142, 109812. https://doi.org/ 10.1016/j.mehy.2020.109812.

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Liu, W., Zhang, S., Nekhai, S., Liu, S., 2020a. Depriving iron supply to the virus represents a promising adjuvant therapeutic against viral survival [published online ahead of print, 2020 Apr 20]. Curr. Clin. Microbiol. Rep. 1–7. https://doi.org/ 10.1007/s40588-020-00140-w.

Liu, W., Zhang, S., Nekhai, S., Liu, S., 2020b. Depriving iron supply to the virus represents a promising adjuvant therapeutic against viral survival [published online ahead of print, 2020 Apr 20]. Curr. Clin. Microbiol. Rep. 1–7. https://doi.org/ 10.1007/s40588-020-00140-w.

Luo, Z., Zeng, W., Tang, W., et al., 2014. CD147 interacts with NDUFS6 in regulating mitochondrial complex I activity and the mitochondrial apoptotic pathway in human malignant melanoma cells. Curr. Mol. Med. 14 (10), 1252–1264. https://doi.org/ 10.2174/1566524014666141202144601.

Machhi, J., Kevadiya, B.D., Muhammad, I.K., et al., 2020. Harnessing regulatory T cell neuroprotective activities for treatment of neurodegenerative disorders. Mol. Neurodegener. 15, 32. https://doi.org/10.1186/s13024-020-00375-7.

Madabhushi, R., Pan, L., Tsai, L.H., 2014. DNA damage and its links to neurodegeneration. Neuron 83 (2), 266–282. https://doi.org/10.1016/j. neuron.2014.06.034.

Maghazachi, A.A., 2013a. On the role of natural killer cells in neurodegenerative diseases. Toxins (Basel) 5 (2), 363–375. Published 2013 Feb 19. https://doi.org/10 .3390/toxins5020363.

Maghazachi, A.A., 2013b. On the role of natural killer cells in neurodegenerative diseases. Toxins (Basel) 5 (2), 363–375. Published 2013 Feb 19. https://doi.org/10 .3390/toxins5020363.

Mak, I.T., Landgraf, K.M., Chmielinska, J.J., Weglicki, W.B., 2012. Angiotensin II promotes iron accumulation and depresses PGI<sub>2</sub> and NO synthesis in endothelial cells: effects of losartan and propranolol analogs. Can. J. Physiol. Pharmacol. 90 (10), 1413–1418. https://doi.org/10.1139/y2012-104.

Ma-Lauer, Y., Carbajo-Lozoya, J., Hein, M.Y., et al., 2016. p53 down-regulates SARS coronavirus replication and is targeted by the SARS-unique domain and PLpro via E3 ubiquitin ligase RCHY1. Proc. Natl. Acad. Sci. U. S. A. 113 (35), E5192–E5201. https://doi.org/10.1073/pnas.1603435113.

Malavolta, M., Giacconi, R., Brunetti, D., Provinciali, M., Maggi, F., 2020. Exploring the relevance of senotherapeutics for the current SARS-CoV-2 emergency and similar future global health threats. Cells 9 (4), 909. Published 2020 Apr 8. https://doi. org/10.3390/cells9040909.

- Mao, L., Wang, M., Chen, S., He, Q., Chang, J., Hong, C., 2020. Neurological manifestations of hospitalized patients with COVID-19 in Wuhan. China: a retrospective case series study, medRxiv. JAMA Neurol. 77 (6), 683–690 (02.22.20026500).
- Market, M., Angka, L., Martel, A.B., et al., 2020. Flattening the COVID-19 curve with natural killer cell based immunotherapies. Front. Immunol. 11, 1512. Published 2020 Jun 23. https://doi.org/10.3389/fimmu.2020.01512.
- Martínez-Cué, C., Rueda, N., 2020. Cellular senescence in neurodegenerative diseases. Front. Cell. Neurosci. 14, 16. Published 2020 Feb 11. https://doi.org/10.33 89/fncel.2020.00016.
- Masaldan, S., Clatworthy, S.A.S., Gamell, C., et al., 2018. Iron accumulation in senescent cells is coupled with impaired ferritinophagy and inhibition of ferroptosis. Redox Biol. 14, 100–115. https://doi.org/10.1016/j.redox.2017.08.015.
   Masselli, E., Vaccarezza, M., Carubbi, C., et al., 2020. NK cells: a double edge sword

Masselli, E., Vaccarezza, M., Carubbi, C., et al., 2020. NK cells: a double edge sword against SARS-CoV-2. Adv. Biol. Regul. 77, 100737. https://doi.org/10.1016/j. jbior.2020.100737.

Mertens, B., Varcin, M., Michotte, Y., Sarre, S., 2011. The neuroprotective action of candesartan is related to interference with the early stages of 6-hydroxydopamineinduced dopaminergic cell death. Eur. J. Neurosci. 34 (7), 1141–1148.

Mihara, T., Nakashima, M., Kuroiwa, A., et al., 2008. Natural killer cells of Parkinson's disease patients are set up for activation: a possible role for innate immunity in the pathogenesis of this disease. Parkinsonism Relat. Disord. 14 (1), 46–51. https://doi. org/10.1016/j.parkreldis.2007.05.013.

Mijit, M., Caracciolo, V., Melillo, A., Amicarelli, F., Giordano, A., 2020a. Role of p53 in the regulation of cellular senescence. Biomolecules 10 (3), 420. https://doi.org/ 10.3390/biom10030420.

Mijit, M., Caracciolo, V., Melillo, A., Amicarelli, F., Giordano, A., 2020b. Role of p53 in the regulation of cellular senescence. Biomolecules 10 (3), 420. Published 2020 Mar 8. https://doi.org/10.3390/biom10030420.

Mohanram, A., Zhang, Z., Shahinfar, S., Lyle, P.A., Toto, R.D., 2008. The effect of losartan on hemoglobin concentration and renal outcome in diabetic nephropathy of type 2 diabetes. Kidney Int. 73 (5), 630–636. https://doi.org/10.1038/sj. ki.5002746.

Morris, G., Maes, M., 2012. Increased nuclear factor-kB and loss of p53 are key mechanisms in Myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS). Med. Hypotheses 79 (5), 607–613. https://doi.org/10.1016/j.mehy.2012.07.034.

Moujaess, E., Kourie, H.R., Ghosn, M., 2020. Cancer patients and research during COVID-19 pandemic: a systematic review of current evidence. Crit. Rev. Oncol. Hematol. 150, 102972. https://doi.org/10.1016/j.critrevonc.2020.102972.

Mousa, S.A., Ayoub, B.M., 2019. Repositioning of dipeptidyl peptidase-4 inhibitors and glucagon like peptide-1 agonists as potential neuroprotective agents. Neural Regen. Res. 14 (5), 745–748. https://doi.org/10.4103/1673-5374.249217.

Muñoz-Fontela, C., Pazos, M., Delgado, I., et al., 2011. p53 serves as a host antiviral factor that enhances innate and adaptive immune responses to influenza A virus. J. Immunol. 187 (12), 6428–6436. https://doi.org/10.4049/jimmunol.1101459.
Naggie, S., Hepatitis, C., 2017. Virus, inflammation, and cellular aging: turning back time. Top Antivir Med. 25 (1), 3–6.

Nuñez, M.T., Chana-Cuevas, P., 2018. New perspectives in iron chelation therapy for the treatment of neurodegenerative diseases. Pharmaceuticals (Basel) 11 (4), 109. Published 2018 Oct 19. https://doi.org/10.3390/ph11040109. Pablo, K.A., Ochrietor, J.D., 2013. Deletion of the Basigin gene results in reduced mitochondria in the neural retina. Biochem. Biophys. Res. Commun. 438 (3), 546–550. https://doi.org/10.1016/j.bbrc.2013.07.092.

Pagani, A., Nai, A., Silvestri, L., Camaschella, C., 2019. Hepcidin and anemia: a tight relationship. Front. Physiol. 10, 1294. Published 2019 Oct 9. https://doi.org/10.33 89/fphys.2019.01294.

Pereira, B.I., Devine, O.P., Vukmanovic-Stejic, M., et al., 2019. Senescent cells evade immune clearance via HLA-E-mediated NK and CD8+ T cell inhibition. Nat. Commun. 10, 2387. https://doi.org/10.1038/s41467-019-10335-5.

Perez-Lloret, S., Otero-Losada, M., Toblli, J.E., Capani, F., 2017. Renin-angiotensin system as a potential target for new therapeutic approaches in Parkinson's disease. Expert Opin. Investig. Drugs 26 (10), 1163–1173. https://doi.org/10.1080/ 13543784.2017.1371133.

Perricone, C., Bartoloni, E., Bursi, R., et al., 2020. COVID-19 as part of the hyperferritinemic syndromes: the role of iron depletion therapy. Immunol. Res. 68 (4), 213–224. https://doi.org/10.1007/s12026-020-09145-5.

Perrin, R., Riste, L., Hann, M., Walther, A., Mukherjee, A., Heald, A., 2020. Into the looking glass: post-viral syndrome post COVID-19 [published online ahead of print, 2020 Jun 27]. Med. Hypotheses 144, 110055. https://doi.org/10.1016/j. mehv.2020.110055.

Prusty, S.K., Sahu, P.K., Subudhi, B.B., 2017. Angiotensin mediated oxidative stress and neuroprotective potential of antioxidants and AT1 receptor blockers. Mini-Rev. Med. Chem. 17 (6), 518–528. https://doi.org/10.2174/1389557516666161025094539.

Pryce-Roberts, A., Talaei, M., Robertson, N.P., 2020. Neurological complications of COVID-19: a preliminary review. J. Neurol. 267 (6), 1870–1873. https://doi.org/ 10.1007/s00415-020-09941-x.

Qi, R., Chen, W., Liu, S., et al., 2020. Psychological Morbidities and Fatigue in Patients with Confirmed COVID-19 During Disease Outbreak: Prevalence and Associated Biopsychosocial Risk Factors. Preprint. medRxiv. https://doi.org/10.1101/ 2020.05.08.20031666, 2020.05.08.20031666. Published 2020 May 11.

Quiros Roldan, E., Biasiotto, G., Magro, P., Zanella, I., 2020. The possible mechanisms of action of 4-aminoquinolines (chloroquine/hydroxychloroquine) against Sars-Cov-2 infection (COVID-19): a role for iron homeostasis? Pharmacol. Res. 158, 104904. https://doi.org/10.1016/j.phrs.2020.104904.

Radzikowska, U., Ding, M., Tan, G., et al., 2020. Distribution of ACE2, CD147, CD26 and other SARS-CoV-2 associated molecules in tissues and immune cells in health and in asthma, COPD, obesity, hypertension, and COVID-19 risk factors [published online ahead of print, 2020 Jun 4]. Allergy. https://doi.org/10.1111/all.14429.

Read, R., 2020. Flawed methods in "COVID-19: Attacks the 1-Beta Chain of Hemoglobin and Captures the Porphyrin to Inhibit Human Heme Metabolism". ChemRxiv. https://doi.org/10.26434/chemrxiv.12120912.v1.

Reardon, K.A., Mendelsohn, F.A., Chai, S.Y., Horne, M.K., 2000. The angiotensin converting enzyme (ACE) inhibitor, perindopril, modifies the clinical features of Parkinson's disease. Aust. NZ J. Med. 30 (1), 48–53. https://doi.org/10.1111/ j.1445-5994.2000.tb01054.x.

Rogers, J.P., Chesney, E., Oliver, D., et al., 2020. Psychiatric and neuropsychiatric presentations associated with severe coronavirus infections: a systematic review and meta-analysis with comparison to the COVID-19 pandemic. Lancet Psychiatry 7 (7), 611–627. https://doi.org/10.1016/S2215-0366(20)30203-0.

Saavedra, J.M., 2020. Angiotensin receptor blockers and COVID-19 [published online ahead of print, 2020 Apr 15]. Pharmacol. Res. 156, 104832. https://doi.org/ 10.1016/j.phrs.2020.104832.

Sargiacomo, C., Sotgia, F., Lisanti, M.P., 2020. COVID-19 and chronological aging: senolytics and other anti-aging drugs for the treatment or prevention of corona virus infection? Aging (Albany NY) 12 (8), 6511–6517. https://doi.org/10.18632/ aging.103001.

Schmidt, S.M., 2020a. The role of iron in viral infections. Front. Biosci. (Landmark Ed) 25, 893–911.

Schmidt, S.M., 2020b. The role of iron in viral infections. Front. Biosci. (Landmark Ed) 25, 893–911 (Published 2020 Jan 1).

Schneiderhan, W., Scheler, M., Holzmann, K.H., et al., 2009. CD147 silencing inhibits lactate transport and reduces malignant potential of pancreatic cancer cells in in vivo and in vitro models. Gut 58 (10), 1391–1398. https://doi.org/10.1136/ gut.2009.181412.

Scott, K.E., Cleveland, J.L., 2016. Lactate wreaks havoc on tumor-infiltrating T and NK cells. Cell Metab. 24 (5), 649–650. https://doi.org/10.1016/j.cmet.2016.10.015.

Sfera, A., Bullock, K., Price, A., Inderias, L., Osorio, C., 2018. Ferrosenescence: the iron age of neurodegeneration? Mech. Ageing Dev. 174, 63–75. https://doi.org/10.1016/ j.mad.2017.11.012.

Sfera, A., Osorio, C., Jafri, N., Diaz, E.L., Campo Maldonado, J.E., 2020. Intoxication with endogenous angiotensin II: a covid-19 hypothesis. Front. Immunol. 11, 1472. Published 2020 Jun 19. https://doi.org/10.3389/fimmu.2020.01472.

Shen, J., Sheng, X., Chang, Z., et al., 2014. The heme-p53 interaction: linking iron metabolism to p53 signaling and tumorigenesis. Mol. Cell. Oncol. 3 (1) https://doi. org/10.4161/23723548.2014.965642 e965642. Published 2014 Nov 3.

Sherman, A.R., Spear, A.T., 1993. Iron and immunity. In: Klurfeld, D.M. (Ed.), Nutrition and Immunology. Page 296. Human Nutrition (A Comprehensive Treatise). Springer, Boston, MA. https://doi.org/10.1007/978-1-4615-2900-2\_14.

Shimasaki, N., Jain, A., Campana, D., 2020. NK cells for cancer immunotherapy. Nat. Rev. Drug Discov. 19 (3), 200–218. https://doi.org/10.1038/s41573-019-0052-1.

Silvestre, I.B., Dagda, R.Y., Dagda, R.K., Darley-Usmar, V., 2019. Mitochondrial alterations in NK lymphocytes from ME/CFS patients. J. Immunol. 202 (1 Supplement), 126.39.

Singh, N., Bharara, Singh A., 2020a. S2 subunit of SARS-nCoV-2 interacts with tumor suppressor protein p53 and BRCA: an in silico study [published online ahead of

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print, 2020 Jun 29]. Transl. Oncol. 13 (10), 100814. https://doi.org/10.1016/j. tranon.2020.100814.

Singh, N., Bharara, Singh A., 2020b. S2 subunit of SARS-nCoV-2 interacts with tumor suppressor protein p53 and BRCA: an in silico study [published online ahead of print, 2020 Jun 29]. Transl. Oncol. 13 (10), 100814. https://doi.org/10.1016/j. tranon.2020.100814.

- Solerte, S.B., Di Sabatino, A., Galli, M., Fiorina, P., 2020. Dipeptidyl peptidase-4 (DPP4) inhibition in COVID-19. Acta Diabetol. 57 (7), 779–783. https://doi.org/10.1007/ s00592-020-01539-z.
- Song, P., Kim, J.H., 2019. Benefits of angiotensin receptor blockade: preventing smooth muscle cell senescence and beyond. Korean Circ. J. 49 (7), 627–628. https://doi.org/ 10.4070/kcj.2019.0164.
- Song, P., An, J., Zou, M.H., 2020. Immune clearance of senescent cells to combat ageing and chronic diseases. Cells 9 (3), 671. Published 2020 Mar 10. https://doi.org/1 0.3390/cells9030671.
- Sottile, R., Federico, G., Garofalo, C., et al., 2019. Iron and ferritin modulate MHC class I expression and NK cell recognition. Front. Immunol. 10, 224. Published 2019 Feb 26. https://doi.org/10.3389/fimmu.2019.00224.
- Su, S., Shen, J., Zhu, L., et al., 2020. Involvement of digestive system in COVID-19: manifestations, pathology, management and challenges. Ther. Adv. Gastroenterol. 13 https://doi.org/10.1177/1756284820934626 (1756284820934626).
- Svenningsson, P., Wirdefeldt, K., Yin, L., et al., 2016. Reduced incidence of Parkinson's disease after dipeptidyl peptidase-4 inhibitors-a nationwide case-control study. Mov. Disord. 31 (9), 1422–1423. https://doi.org/10.1002/mds.26734.
- Swinkels, D.W., Aalbers, N., Elving, L.D., Bleijenberg, G., Swanink, C.M., van der Meer, J. W., 2002. Primary haemochromatosis: a missed cause of chronic fatigue syndrome? Neth. J. Med. 60 (11), 429–433.
- Tajima, S., Tsuchiya, K., Horinouchi, Y., et al., 2010. Effect of angiotensin II on irontransporting protein expression and subsequent intracellular labile iron concentration in human glomerular endothelial cells. Hypertens. Res. 33, 713–721. https://doi.org/10.1038/hr.2010.63.
- Tarangelo, A., Dixon, S., 2018. The p53-p21 pathway inhibits ferroptosis during metabolic stress. Oncotarget 9 (37), 24572–24573. https://doi.org/10.18632/ oncotarget.25362.
- Theurl, I., Zoller, H., Obrist, P., et al., 2004. Iron regulates hepatitis C virus translation via stimulation of expression of translation initiation factor 3. J. Infect. Dis. 190 (4), 819–825. https://doi.org/10.1086/422261.
- Thorburn, T., Aali, M., Kostek, L., et al., 2017. Anti-inflammatory effects of a novel iron chelator, DIBI, in experimental sepsis. Clin. Hemorheol. Microcirc. 67 (3–4), 241–250. https://doi.org/10.3233/CH-179205.
- Tipton, P.W., Wszolek, Z.K., 2020. What can Parkinson's disease teach us about COVID-19? Neurol. Neurochir. Pol. 54 (2), 204–206. https://doi.org/10.5603/PJNNS. a2020.0039.
- Troyer, E.A., Kohn, J.N., Hong, S., 2020. Are we facing a crashing wave of neuropsychiatric sequelae of COVID-19? Neuropsychiatric symptoms and potential immunologic mechanisms [published online ahead of print, 2020 Apr 13]. Brain

Behav. Immun. https://doi.org/10.1016/j.bbi.2020.04.027. S0889-1591(20)30489-

- Ulrich, H., Pillat, M.M., 2020. CD147 as a target for COVID-19 treatment: suggested effects of azithromycin and stem cell engagement. Stem Cell Rev. Rep. 16 (3), 434–440. https://doi.org/10.1007/s12015-020-09976-7.
- van Erp, E.A., van Kampen, M.R., van Kasteren, P.B., de Wit, J., 2019. Viral infection of human natural killer cells. Viruses 11 (3), 243. Published 2019 Mar 12. https://doi. org/10.3390/v11030243.
- Vargas-Vargas, M., Cortés-Rojo, C., 2020. Ferritin levels and COVID-19. Rev. Panam. Salud Publica 44. https://doi.org/10.26633/RPSP.2020.72 e72.
- Villapol, S., Saavedra, J.M., 2015. Neuroprotective effects of angiotensin receptor blockers. Am. J. Hypertens. 28 (3), 289–299. https://doi.org/10.1093/ajh/hpu197.
- Wichaiyo, S., Yatmark, P., Morales Vargas, R.E., et al., 2015. Effect of iron overload on furin expression in wild-type and β-thalassemic mice. Toxicol. Rep. 2, 415–422. https://doi.org/10.1016/j.toxrep.2015.01.004.
- Williams, A., Meyer, D., 2009. Desferrioxamine as immunomodulatory agent during microorganism infection. Curr. Pharm. Des. 15 (11), 1261–1268. https://doi.org/ 10.2174/138161209787846801.
- Wilson, C., 2020. Concern coronavirus may trigger post-viral fatigue syndromes. New Sci. 246 (3278), 10–11. https://doi.org/10.1016/S0262-4079(20)30746-6.
- Xia, S., Lan, Q., Su, S., et al., 2020. The role of furin cleavage site in SARS-CoV-2 spike protein-mediated membrane fusion in the presence or absence of trypsin. Signal Transduct. Target Ther. 5 (1), 92. https://doi.org/10.1038/s41392-020-0184-0.
- Xie, H., Liu, L., Shi, W., et al., 2011. Down regulation of CD147 boosts the premature senescence in human skin fibroblasts by destroying the redox balance and inhibiting klotho. J. Dermatol. Sci. 64 (3), 243–245. https://doi.org/10.1016/j. idermsci.2011.09.010.
- Xie, Y., Zhu, S., Song, X., et al., 2017. The tumor suppressor p53 limits ferroptosis by blocking DPP4 activity. Cell Rep. 20 (7), 1692–1704. https://doi.org/10.1016/j. celrep.2017.07.055.
- Yan, N., Zhang, J., 2020. Iron metabolism, ferroptosis, and the links with Alzheimer's disease. Front. Neurosci. 13, 1443. Published 2020 Jan 29. https://doi.org/10.33 89/fnins.2019.01443.
- Zhang, J., Chen, X., 2019a. p53 tumor suppressor and iron homeostasis. FEBS J. 286 (4), 620–629. https://doi.org/10.1111/febs.14638.
- Zhang, J., Chen, X., 2019b. p53 tumor suppressor and iron homeostasis. FEBS J. 286 (4), 620–629. https://doi.org/10.1111/febs.14638.
- Zhang, W., Gong, J., Yang, H., et al., 2020. The mitochondrial protein MAVS stabilizes p53 to suppress tumorigenesis. Cell Rep. 30 (3), 725–738 e4. https://doi.org/10.10 16/j.celrep.2019.12.051.
- Zhou, Xiang, Wang, Weiming, Zhang, Baofu, Lin, Zixia, Wang, Yi, Chen, Gang, 2020. DPP4 modulates ACSL4 to promote lipid peroxidation and regulate ferroptosis in pancreatic cancer [abstract]. In: Proceedings of the Annual Meeting of the American Association for Cancer Research(2020) 2020 Apr 27–28 and Jun 22–24. AACR; Cancer Res, Philadelphia (PA) (80(16 Suppl): Abstract nr 6330).