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Nanoparticle-Mediated Immunogenic Cell Death Enables and Potentiates Cancer Immunotherapy

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Abstract

Cancer immunotherapies that train or stimulate the inherent immunological systems to recognize, attack, and eradicate tumor cells with minimal damage to healthy cells have demonstrated promising clinical responses in recent years. However, most of these immunotherapeutic strategies only benefit a small subset of patients and cause systemic autoimmune side effects in some patients. Immunogenic cell death (ICD)-inducing modalities not only directly kill cancer cells but also induce antitumor immune responses against a broad spectrum of solid tumors. Such strategies for generating vaccine-like functions could be used to stimulate a "cold" tumor microenvironment to become an immunogenic, "hot" tumor microenvironment, working in synergy with immunotherapies to increase patient response rates and lead to successful treatment outcomes. This Minireview will focus on nanoparticle-based treatment modalities that can induce and enhance ICD to potentiate cancer immunotherapy.

Graphical Abstract

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Keywords

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1. Introduction

Cancer immunotherapies have enjoyed rapid clinical progress over the past few years, particularly in the areas of chimeric antigen receptor (CAR)-modified T cells and immune modulation by blocking suppressive immune checkpoints.^[1] The generation of an anticancer immune response consists of several key steps (Figure 1):^[2] Antigens released from cancer cells are captured by antigen-presenting cells (APCs). Then, danger associated molecular patterns (DAMPs), such as pro-inflammatory cytokines and factors, released by the dying tumor cells signal APC maturation. Activated APCs travel to the lymph nodes to present the tumor antigens on major histocompatibility complex (MHC) I and MHC II molecules to T cells, resulting in the priming and activation of effector T cell responses against cancer-specific antigens. Finally, activated effector T cells traffic to and infiltrate the tumor bed where they specifically recognize cancer cells through interactions between T cell receptor (TCR) and MHC I-bound cognate antigen and kill cancer cells. Killing the cancer cell releases additional tumor-associated antigens (TAAs) to increase the breadth and depth of the immune response in subsequent revolutions of the cycle.

However, cancer cells are able to evade the host immune system by blocking one or more of these steps.^[3] This can occur by down-regulating surface antigen and MHC I expression to inhibit recognition and attack by the immune system, inducing expression of immunosuppressive molecules, and infiltrating inhibitory immune cells into the tumor microenvironment (TME) to inhibit effector T cell homing and activity.^[4] Current cancer immunotherapies usually focus on one of two strategies: 1) To stimulate key players of the immune system, such as cancer vaccines,^[5] cytokine therapy,^[6] and adoptive T-cell transfer^[7] and 2) to eliminate or inhibit immunosuppressive factors, such as immune checkpoint-blockade (ICB) therapy.^[8] Although cancer immunotherapies have demonstrated some exciting clinical responses, they are limited by high costs, efficacy restricted to certain subsets of patients, resistance to treatment, and dose-limiting autoimmune effects like cytokine release syndrome.^[9]

It is now established that ablative cancer treatments, such as radiotherapy (RT), photodynamic therapy (PDT), hyperthermia (HT) and photothermal therapy (PTT), and certain chemotherapeutics can cause tumor cell death in an immunogenic way.^[10] This immunogenic cell death (ICD) is characterized by release of TAAs, DAMPs, and proinflammatory cytokines, which facilitates the presentation of TAAs to adaptive immune cells, eliciting an antigen-specific immune response against a broad spectrum of solid tumors.^[11] Ultimately, ICD can enhance immune stimulatory or subvert immune suppressive effects for the activation, proliferation, and tumor infiltration of T cells to synergize with current immunotherapies. Such in situ "tumor vaccines" provide a new way to broaden and enhance immunotherapy by combining it with ICD-inducing modalities, such as radio-, photo-, and chemotherapy. In this Minireview, we will first discuss the characteristics and design parameters of nanoparticles (NPs) that allow for maximal immunotherapeutic efficacy, then provide an overview of nanoparticle platforms used to combine ICD-inducing modalities and immunotherapy in the emerging and rapidly growing field of NP-mediated cancer immunotherapy.

2. Immunogenic Cell Death (In Situ Cancer Vaccination)

During ICD, DAMPs and TAAs are released, captured by dendritic cells (DCs) and macrophages, then processed and presented to adaptive immune cells, leading to an antigen-specific immune response. Many immunogenic factors throughout apoptotic cell death have been identified as DAMPs, such as pre-apoptotic calreticulin (CRT) exposure on the cell surface,^[12] ATP release during the blebbing phase of apoptosis,^[13] and post-apoptotic exodus of high mobility group box 1 (HMGB-1)^[14] and heat shock proteins (HSPs).^[15] ATP serves as a chemoattractant to recruit APCs; CRTacts as an "eat-me" signal to facilitate the engulfment of dying tumor cells and their debris by APCs.^[16] Finally, HMGB-1 and HSPs stimulate optimal antigen presentation to T cells (Figure 2).^[14a,17] ICD may also induce a strong inflammatory response by releasing pro-inflammatory cytokines such as TNF- α , IL-6, and IL-1 β .^[11a,18] Such strategies to generate vaccine-like effects in situ do not require sophisticated procedures, have no ethical concerns, and are able to efficiently elicit selective and strong protections by local in situ cell death and systemic cell-mediated immune responses. Leveraging the ICD effects of traditional treatment modalities can increase infiltration of immune effector cells to convert an immunosuppressive TME to an

immunogenic TME, and ultimately increase patient response rates to immune adjuvants and checkpoint blockade immunotherapy.

3. Design of Nanoparticles for ICD

For in situ cancer vaccines, the ICD inducer needs to specifically target tumors. Compared to conventional approaches, nanotechnology offers an opportunity for the efficient delivery of an optimal dose of ICD inducers to specific tissues or cell types, enhancing their potency while reducing their side effects.^[20] NPs can co-load multi-components for simultaneous delivery, protect the payloads from degradation and premature release, and passively or actively target tumors by the enhanced permeability and retention (EPR) effect or surface modification with ligands, respectively.^[21] Typically, polymeric NPs are designed with selective shapes and sizes, for intracellular release of loaded small molecules, and surface functionalization for delivery beyond biological barriers.^[22] Furthermore, inorganic NPs can be used as a localized source of ICD-inducing treatment or accentuate treatment by external energy fields to minimize damage to healthy tissues. Metals in inorganic NPs are chosen for biocompatibility and intrinsic behavior such as magnetic susceptibility and colloidal stability.^[23]

NPs with imaging functions can monitor the localization and treatment effect in real-time, while NPs with an immunomodulatory effect can act as adjuvants or immune potentiators. Furthermore, NPs can be tailor-made in terms of size, shape, structure, payload, and surface properties for transporting themselves and their cargoes through biological barriers and enhancing accumulation in the solid tumor.^[24]

4. Application of Nanoparticles in Enhancing Cancer Immunotherapy

4.1. Hyperthermia

Hyperthermia (HT), the heating of tissue to $39-45^{\circ}$ C, was shown to activate an antitumor immune response by inducing a cell stress known as the unfolded protein response, which leads to up-regulation of HSPs on the cell membrane and TAAs within the tumor cells. The released TAAs and TAA-HSP complexes facilitate the uptake by APCs and aid in the presentation of antigens through MHC class I, activating antigen-specific killer T cells that undergo clonal expansion and traffic to all tumors (primary and metastatic) to kill tumor cells directly.^[25] In addition, the fever-like temperature created by HT has been shown to enhance antigen presentation, improve tumor blood flow, and accelerate leukocyte trafficking to the tumor.^[26] Both treatment efficacy and elicited immune response were found to be dependent on the thermal dose, which requires careful optimization to balance treatment of the primary tumors and the induction of immunological effects against distant tumors. For example, coagulative ablation strategies, in which high temperatures "melt" the tumor tissue and cut off intratumoral (i.t.) blood flow, showed the most destructive effects on the primary tumor but likely also formed a barrier for immune cell infiltration and immune induction.^[27] Another important consideration is that normal tissues may also be severely damaged under conventional hyperthermia treatment, thus, specifically heating the tumor region to the desired temperature without damaging surrounding normal tissues is a major

technical barrier. Fortunately, NP-assisted hyperthermia offers a means for controlling the thermal dose and localizing the treatment.^[28]

4.1.1. Magnetic Fluid Hyperthermia—Magnetic fluid hyperthermia (MFH) involves the combination of magnetic NPs with an alternating magnetic field (AMF), which induces local hyperthermia in tumors in a controlled and uniform manner without the limitation of penetration depth. Toraya-Brown et al. stimulated 100 nm Fe₃O₄ BNF-Starch magnetic NPs with an external magnetic field to heat tumors to 43°C for 30 min,^[29] while the Kobayashi group used cationic liposomes containing magnetic NPs (MNHT) to heat the local tumor tissue to above 43°C without damaging surrounding normal tissues in the presence of AMF. ^[30] In both instances, MFH triggered an antitumor immune response mediated by both CD8⁺ and CD4⁺ T cells, leading to elimination of both heat-treated primary tumors and unheated distant tumors and subsequent rechallenge rejection.^[29,31] Interestingly, the differences in temperature did not show obvious differences in immune response. To enhance MNHT, magnetic NPs were conjugated with a melanogenesis substrate, Npropionyl cysteaminylphenol (NPrCAP), to be specifically taken up by melanoma cells and react with tyrosinase to produce highly cytotoxic free radicals, resulting in chemotherapeutic and immunotherapeutic cell death.^[32] In parallel, they extended their localized hyperthermia approach in combination with immunostimulatory factors; i.t. injection of IL-2 or GM-CSF 24 h after intracellular HT resulted in complete tumor regression in 75% and 40% of mice, respectively.^[33] Recombinant mouse HSP70 gene^[34] and protein^[35] were also able to enhance the therapeutic efficacy of MNHT.

4.1.2. Photothermal Therapy—Light-induced hyperthermia, or photothermal therapy (PTT), involves irradiation of light-absorbing agents accumulated in the tumor with NIR light to convert optical energy into heat for thermal ablation of cancer cells. Theoretically, photothermal agents are non-toxic in the dark and the light is locally applied on the tumor area, leading to high treatment efficacy with few side effects.^[36] Recently, PTT has shown the ability to generate antitumor immunological effects by producing TSAs from ablated tumor cell residues.

A number of inorganic nano-agents (e.g. AuNPs and nanoshells,^[37] CuS NPs,^[38] GO,^[39] MoS₂ nanosheets,^[40] or carbon nanotubes^[41]) with intrinsic capacity for NIR light absorption have been developed to deliver thermal energy and immunoadjuvants. Yata et al. modified AuNPs with CpG oligodeoxynuleotides and then mixed them with a hexapod-like structured DNA containing CpG sequences, obtaining an immunostimulatory Au–DNA hydrogel. Intratumoral injection of the Au–DNA hydrogel followed by laser irradiation increased the local temperature and HSP70 mRNA levels in the tumor, improved TAA-specific IgG levels in the serum, and induced TAA-specific IFN-γ production in splenocytes, significantly retarding tumor growth and extending the survival time of tumorbearing mice.^[37] Guo et al. designed chitosan-coated hollow CuS NPs containing CpG, which could break down after laser excitation, reassemble, and transform into polymer complexes, improving CpG tumor retention, effectively eliminating the primary tumor and simultaneously inhibiting the growth of distant untreated tumors.^[38]

Small molecule photothermal agents can also be co-loaded with immunostimulators into liposomes or polymeric NPs for effective combination of PTT and immunotherapy. Li et al. coated a hyaluronic acid (HA)-CpG conjugate onto fluorophore (IR-7)-loaded liposomes to activate and increase tumor infiltration of DCs and CD8⁺ T cells, thereby eradicating tumors in mice and inhibiting tumor metastasis.^[42] Owing to its immunostimulatory effect, glycated chitosan (GC) can be used as both an immunoadjuvant and a carrier to enhance the uptake and presentation of tumor antigens, increase tumor immunogenicity, and synergize with PTT. Kumar and Srivastava developed biocompatible and biodegradable monodisperse polycaprolactone/GC/Poloxamer blend NPs encapsulating photothermal agent IR 820 for imaging and photo-immunotherapy. Though the NPs enhanced toxicity on MCF-7 cells upon laser treatment, the immune response and efficacy in vivo were not investigated.^[43]

As a potent stimulator of the immune system, PTT has also shown synergy with ICB to reverse immunosuppression. Wang et al. demonstrated the first combination of anti-CTLA-4 therapy with PTT using single-wall carbon nanotubes. This stimulated DC maturation in tumor-draining lymph nodes (TDLNs), induced infiltration of effector T cells, and greatly abrogated Treg cells in non-heated distant tumors, thereby reducing tumor burden in both subcutaneous and lung metastasis models and leading to prolonged survival.^[44] Similar long-term survival was later observed using Prussian blue NP-based PTT complemented with anti-CTLA-4 in a preclinical neuroblastoma model.^[45] In addition, all of the mice that survived past 100 days also rejected a tumor rechallenge, illustrating immune memory. Chen et al. also demonstrated the long-term survival by combining anti-CTLA-4 with a PLGA NP co-loaded with a photothermal agent indocyanine green (ICG) and a TLR-7 agonist imiquimod (R837, Figure 3a). PLGA-ICG-R837 plus anti-CTLA-4 therapy additionally demonstrated long-term survival (70% at 70 days) in a 4T1 lung metastasis rechallenge model compared to no survival for the anti-CTLA-4 only treated mice (Figure 3b).^[46] More recently, Liu et al. demonstrated that plasmonic gold nanostar-mediated PTT in combination with anti-PD-L1 immunotherapy dramatically enhanced the efficacy of immunotherapy, achieving complete eradication of primary treated tumors and distant untreated tumors in some mice implanted with MB49 bladder cancer cells. Furthermore, the treatment induced effective long-lasting immunity, rejecting the formation of challenged tumor in a period of 60 days.^[47]

4.2. Photodynamic Therapy

Photodynamic therapy (PDT) is a clinically used, minimally invasive therapeutic procedure with a two-step modus operandi involving the administration of a photosensitizer (PS) followed by irradiation with a specific wavelength light. Upon irradiation, PS in the presence of oxygen generates highly cytotoxic ROS to cause oxidative stress-based cell death and disrupts tumor vasculature. In addition to local tumor ablation, PDT can increase tumor immunogenicity by inducing CRT exposure and release of tumor cell debris, which improves tumor antigen presentation and activation of T lymphocytes, resulting in the destruction of residual tumor cells and reduction in the risk of distant metastasis.^[48]

Typically PSs are hydrophobic and aggregate in aqueous media, which deleteriously affects their photophysical (decreased ${}^{1}O_{2}$ formation), chemical (decreased solubility), and

biological (insufficient tumor localization) properties.^[49] NPs can overcome these limitations and selectively deliver PSs to tumors, minimize damage to normal tissues, and reduce systemic toxicity when accompanied by spatially controlled light irradiation.^[50] Yu et al. functionalized graphene oxide (GO) with an integrin $\alpha\nu\beta6$ -specific HK peptide and loaded the PS 2-[1-hexyloxyethyl]-2-devinyl pyropheophorbidea (HPPH) onto the surface of GO through hydrophobic interactions and $\pi-\pi$ stacking. The nanophotosensitizer (nPS) exhibited high tumor uptake after intravenous (i.v.) injection and increased infiltration of cytotoxic CD8⁺ T lymphocytes within tumors, suppressing tumor growth upon irradiation in both subcutaneous and lung metastatic mouse models.^[51]

Though PDT alone can activate the immune system, combination with immune adjuvants can further potentiate the immune response. Marrach et al. encapsulated a long-wavelength-absorbing PS, zinc phthalocyanine (ZnPc), within a polymeric NP core composed of PLGA-b-PEG, and then coated the polymeric core with CpG-modified gold NPs. Such NP-treated 4T1 cell lysate primed murine bone marrow-derived dendritic cells (BMDCs) to recognize and phagocytose PDT-killed tumor cells. This phagocytosis led to pro-inflammatory cytokines (IL-2, IL-6, IL-12, and TNF- α) release, evidence of DC maturation and activation. [52]

Combination with ICB further enhanced the effects of PDT. Duan et al. synthesized a nontoxic core–shell NP (ZnP@pyro) with a coordination polymer of Zn²⁺ and pyrophosphate in the core and the PS pyrolipid in the shell. In vivo, the NPs improved tumor immunogenicity upon light irradiation, particularly upon combination with PD-L1. ZnP@pyro with anti-PD-L1 eradicated the irradiated tumors, suppressed non-irradiated tumors, and prevented lung meta-stases in a 4T1 metastatic triple-negative breast cancer murine model (Figure 4). In addition, the combination therapy produced an efficient abscopal effect on two bilateral syngeneic mouse models, leading to complete inhibition of pre-existing non-irradiated tumors by generating a systemic tumor-specific cytotoxic T cell response.^[53]

Nanoparticles are ideal to overcome tumor conditions that limit PDT, such as the requirement for oxygen to generate ROS. To overcome hypoxia in tumor, Lan et al. reported a novel nanoscale metal–organic framework (nMOF), Fe-TBP (TBP = 5,10,15,20-tetra(*p*-benzoato)porphyrin), consisting of Fe₃O clusters and TBP ligands. When irradiated under hypoxic conditions, Fe₃O clusters decomposed intracellular H₂O₂ to produce O₂ through a Fenton-like reaction, and the generated O₂ was then converted to cytotoxic ¹O₂ by photo-excited porphyrins. Fe-TBP mediated PDT induced significant tumor infiltration of cytotoxic T cells, sensitizing anti-PD-L1 treatment and eliciting the abscopal effect in a mouse model of colorectal cancer with greater than 90% regression of tumors (Figure 5).^[54]

Wang et al. combined PDT and ICB onto a single particle with a versatile micelleplex, integrating an acid-activatable cationic micelle, a PS pheophorbide A (PPa) and a siRNA that targeted PD-L1 (Figure 6a). The micelleplex was inert at physiological pH conditions and activated only upon internalization in the acidic endocytic vesicles of tumor cells for fluorescence imaging and PDT (Figure 6b). Compared to PDT alone, the combination of PDT and PD-L1 knockdown showed significantly enhanced efficacy for inhibiting tumor

growth and distant metastasis in a B16F10 melanoma xenograft tumor model (Figure 6c–e). [55]

Indoleamine 2,3-dioxygenase (IDO) inhibition has also shown significant synergy with PDT. Recently, Lu et al. showed that an IDO inhibitor (IDOi) could be loaded into the channels of a chlorin-based nMOF, TBC-Hf (TBC is 5,10,15,20-tetra(*p*-benzoato)chlorin), for anticancer efficacy and immune activation. In addition to increased T cell infiltration and consistent abscopal responses in mouse models of colorectal cancers, the i.t. infiltration of neutrophils and B cells was also observed, which may play compensatory roles in presenting TAAs to T cells.^[56]

Song et al. recently synthesized a chimeric peptide, PpIX-1MT, by integrating the PS PpIX with a small-molecule IDOi 1MT (1-methyltryptophan). Light irradiation of PpIX-1MT NPs induced apoptosis and facilitated the expression of caspase-3 and the production of TAAs, triggering an intense immune response. Upon caspase-3 cleavage, the subsequently released 1MT inhibited the IDO pathway to help activate CD8⁺ T cells and synergistically inhibiting both primary and lung metastasis (Figure 7).^[57]

Subsequently, Xu et al. used upconversion NPs (UCNPs) to simultaneously deliver a PS chlorin e6 (Ce6) and R837. Upon irradiation of i.t. injected UCNP-Ce6-R837, the effective photodynamic destruction of tumors generated a pool of TAAs, which were able to trigger DC maturation, promote cytokines secretion, and resulted in strong antitumor immune responses. PDT with UCNP-Ce6-R837 in combination with anti-CTLA-4 not only showed excellent efficacy in eliminating tumors exposed to the NIR laser but also resulted in strong in situ vaccination, inhibiting the growth of distant tumors and protecting treated mice from tumor rechallenge.^[58] Though the combination of PDT immunostimulatory and/or ICB agents consistently led to impressive anticancer efficacy and immune cell infiltration, PDT is reliant on external light stimulation. While the technology is promising, there is limited clinical potential owing to the shallow penetration depth of light.

4.3. Radiotherapy

Radiotherapy (RT) is a powerful therapeutic modality for cancer, commonly used for its ability to kill cancer cells by causing DNA double strand breaks, which is not limited by tissue penetration. Surface exposure of CRT^[16] and release of HSP70^[59] and HMGB-1^[60] suggest that RT induces ICD in situ,^[61] stimulates DC maturation, and induces IFN γ -producing T cells in vitro and in vivo.^[62] Therefore, RT combined with i.t. injection of CpG adjuvant has shown preclinical success and has been tested in humans in whom it induced rejection of the irradiated tumor as well as tumors outside the radiation field (abscopal effect).^[63] RT has also been reported to synergistically promote antitumor immunity with ICB.^[64] However, not all RT-induced modifications of the tumor and its microenvironment favor immune activation. There has been new evidence of pro-tumorigenic M2 macrophages accumulating in hypoxic areas of irradiated tumors^[65] and increased immunosuppressive Treg cells post-RT.^[66] Intriguingly, the dose and fractionation of RT may play a role in modulating the expansion of effector versus Treg cells.^[67]

Min et al. engineered antigen-capturing NPs (AC-NPs) with different surface chemistry to capture TAAs released after radiation and transport them to APCs to promote anticancer immunity (Figure 8). Mechanistic studies revealed that AC-NPs induced an expansion of CD8⁺ cytotoxic T cells and increased both CD4⁺ T/Treg and CD8⁺ T/Treg ratios, thus significantly improving the efficacy of anti-PD-1 treatment on the B16F10 melanoma model with up to a 20% cure rate compared to 0% without AC-NPs.^[68]

Nanotechnology also allows for nanovectorized ionizing radiation that could boost the quality and magnitude of an immune response in a predictable and designable fashion.^[69] Hindré's group used lipid nanocapsules loaded with a lipophilic complex of Rhenium-188 (LNC¹⁸⁸Re-SSS) for fractionated internal radiation in glioblastoma and hepatocellular carcinoma models.^[35] Intratumoral infusion of LNCs by convection-enhanced delivery led to their complete distribution throughout the tumor and peritumoral space without leaking into the contralateral hemisphere except when large volumes were used. 70% of the ¹⁸⁸Re-SSS activity was present in the tumor region 24 h after injection, and no toxicity was observed in the healthy brain. Double fractionated internal radiotherapy with LNC¹⁸⁸Re-SSS resulted in a cure rate of 50% in a human glioblastoma model on T-cell deficient nude mice.^[70a,b] whereas a higher cure rate of 83% was observed in the immunocompetent 9L rat glioma model.^[70c] Increased cytokine (IL-2 and IFN- γ) production in peripheral blood after internal radiation, recruitment of immune and inflammatory cells (DCs, CD4, CD8, NK, macrophages, micro-glia) into the tumor site, and increased expression of MHC class I and class II were all observed, indicating the immune system plays a role in the treatment efficacy.^[71]

4.4. Chemotherapeutic Drugs

Though apoptosis has historically been regarded as a non-inflammatory, immunologically silent or even tolerogenic, recent evidence has shown that a subset of chemotherapeutics can be pro-inflammatory and induce ICD.^[72] As a systemic agent, chemotherapy has the potential to initiate an immune response in multiple sites. Furthermore, NP-mediated chemotherapy has been reported to enhance ICD and consequently improve antitumor effects of the free ICD inducer. For example, Zhao et al. found that immunogenic oxaliplatin encapsulated in PLGA-mPEG NPs released more DAMPs and induced more dendritic cell and T lymphocyte activation and infiltration than free oxaliplatin, improving anticancer efficacy in immunocompetent mice.^[73]

Doxorubicin (DOX) is a bonafide ICD inducer that has already been widely used in NP formulations. Zheng et al. developed a pH- and GSH-dual-sensitive delivery system for systemic treatment of highly metastatic triple-negative breast cancer by loading DOX into highly integrated mesoporous silica NPs, which induced DC maturation and antitumor cytokine release.^[74] The antitumor efficacy and immunity induced by DOX can be enhanced by combination with immunotherapy. Su et al. found that pre-treatment with TNF-a pDNA polyplexes 48 h prior to liposomal DOX (Doxil) promoted its accumulation in tumors, likely owing to TNF-a-mediated opening of the tumor endothelial tight junctions. Three treatment cycles with TNF-a gene vectors and Doxil significantly delayed tumor growth in subcutaneous murine Neuro2A neuroblastoma, and prevented liver metastasis in systemic

Neuro2A metastasis or human LS174T colon carcinoma metastasis models.^[75] Strong intercalation between DOX and DNA has been exploited for the development of DOX/CpG DNA hydrogels, which elevated the levels of cytokines (IL-12, IL-6, and IFN- γ) in serum as well as in tumor tissue, thus inhibiting tumor growth in various tumor models.^[76] More recently, Liu et al. designed a dual pH-responsive multifunctional NP system by coating TLR-7/8 agonist (R848)-loaded poly(L-histidine) (PHIS) nanocore with acid-cleavable HA-DOX conjugates to treat breast cancer. The components separated in the TME, leading to internalization of HA-DOX through CD44-mediated endocytosis by tumor cells. Intracellular HA–DOX released DOX by hydrolysis of the hydrazone bond at pH \approx 5.5 to induce ICD. Extracellular PHIS/R848 released R848 by ionization of PHIS at pH \approx 6.5 to potentiate the immune response. The combined chemoimmunotherapy led to remarkable tumor growth inhibition.^[77] Kuai et al. reported that high-density lipoprotein-mimicking nanodiscs loaded with DOX induced antitumor T cell responses and enhanced therapeutic efficacy of anti-PD-1, leading to eradication of CT26 and MC38 tumors in 80 to 88% of mice and long-term immunity against rechallenge.^[78]

Another chemotherapeutic with a clinically approved nanoformulation, paclitaxel (PTX), is also known to induce ICD. Roy et al. combined chemo- and immunotherapy using PLGA NPs loaded with PTX and a TLR-4 agonist (SP-LPS). The NPs have both direct cytotoxicity and immunostimulatory activity in vitro, but the in vivo antitumor activity was not significantly different from PTX alone because of suboptimal encapsulation of SP-LPS.^[79] Instead, they conjugated PTX with SP-LPS, which subsequently self-assembled into a NP. These NPs showed higher in vivo antitumor activity and a higher percentage of activated immune cells in the TME than the Taxol-treated group.^[80] They also increased the loading of TLR-4 agonist from 20% to 65% by replacing SP-LPS with the similar P-LPS. The higher P-LPS loading showed improved synergy with PTX when co-encapsulated into PLGA NPs, leading to significant reduction in tumor growth compared with either standalone modality. Flow cytometric analysis of tumor-infiltrating immune cells indicated high infiltration and activation of APCs and T cells (CD4⁺ and CD8⁺), correlating to the enhanced survival of mice.^[81] Similarly, Seth et al. demonstrated the synergy between PTX and a TLR-7 agonist for treatment of B16F10 melanoma by using $poly(\gamma$ -glutamic acid) to co-deliver PTX and imiquimod. The co-delivery system enhanced the proliferation (250%) of DCs and secretion of pro-inflammatory and Th1 cytokines, exemplifying drastic inhibition of tumor growth after i.t. injection and leading to 70% survival as compared to individual components with 0% survival at day 41. The antitumor response generated was also found to have systemic memory response since the vaccinated mice significantly delayed secondary tumor development at a distant site six weeks after treatment.^[82] Heo et al. also demonstrated that primary injection with the HA/PTX complex generated TSAs and enhanced their uptake by tumor-recruited BMDCs. With a secondary injection of separate CpG- or IL-10 siRNA-loaded PLGA NPs, the BMDCs became activated and migrated to TDLNs. As a result, the combination not only efficiently inhibited tumor growth but also increased the animal survival rate.^[83]

4.5. Multipronged Approaches

Immunogenic chemotherapy has also been combined with other ablative therapies, such as PTT and PDT. Tao et al. designed a multifunctional platform for combining chemo-, photothermal-, and immunotherapy, constituted of DOX intercalated into CpG sequences, which in turn were conjugated to gold nanorods. The nanosystem localized gold rods into the tumor site for PTT, whereas the CpG induced immune response thereby enhancing the cytotoxic effects of DOX. The combination resulted in significant antitumor efficacy and also led to a long-term tumor-specific immunity.^[84]

He et al. first demonstrated the synergistic effect of chemotherapy with PDT to elicit antitumor immunity, which could be used to augment the antitumor efficacy of ICB. NCPs carrying oxaliplatin (OxPt) in the core and the PS pyrolipid in the shell were prepared for effective chemotherapy and PDT, which provoked a systemic tumor-specific T-cell response. When combined with anti-PD-L1, the NCP led to not only the regression of the irradiated primary tumors but also regression of the non-irradiated tumors by inducing CD8⁺ and CD4⁺ T cell infiltration in bilateral mouse models of syngeneic colorectal cancer (Figure 9). ^[85] Subsequently, Yang et al. co-loaded a PDT agent Ce6 and DOX into hollow H-MnO₂ nanoshells modified with PEG. The obtained H-MnO₂-PEG/C&D dissociated under reduced pH within the TME to release the payloads, while simultaneously decomposing tumor H₂O₂ to relieve tumor hypoxia. As a result, a remarkable in vivo synergistic therapeutic effect was achieved through the combined chemo-photodynamic therapy and the subsequent antitumor immune response. Further combination with ICB led to inhibition of tumors at distant sites, showing promise for the treatment of tumor metastases.^[86]

5. Future perspectives

Cancer immunotherapy has shown exciting clinical responses owing to its unique advantages, such as the induction of specific antitumor immunity and long term immunological memory response, but the low response rate and the potential side effects are still significant hurdles for the broad application of cancer immunotherapy in the clinic. As shown in this review, a number of known and novel inorganic and polymeric nanoparticles such as AuNPs, MOFs, and micelles have been evaluated as ICD-inducing modalities, which can synergize with immunotherapies to improve the treatment outcomes while limiting systemic toxicities. However, the potency of different nanoparticle platforms or applications cannot be easily compared as each experiment was independently performed. Furthermore, more work is needed to monitor the dynamic immune response and understand the specific impact of each combinatorial approach on the TME, with the goal of providing rationales or guidance for selecting the best combinatorial approach for an individual patient. Specifically, it is important to elucidate whether a NP-mediated treatment is more suited towards combination with an immune adjuvant or ICB. In addition, the timing of the ICD-inducing modalities and immunotherapies might also have an impact on the therapeutic efficacy and should be investigated in more detail. So far, only a few stimuli have been shown to induce bonafide ICD; the identification of more compounds or modalities that render cell death immunogenic is clinically urgent. In order to preferentially deliver ICD-inducing agents to tumors, especially metastatic tumors, nanoparticles need to be injected systemically.

However, most of the studies to date have utilized i.t. delivery, possibly owing to low stability, inefficient tumor accumulation or high toxicity; thus, the development of new nanoparticle platforms that can enhance tissue localization and response after systemic administration is critically needed.

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Biography



Xiaopin Duan obtained a B.S. degree in pharmacy from Hebei University (China) in 2008 and received a Ph.D. degree in Pharmaceutics jointly at Shenyang Pharmaceutical University and Shanghai Institute of Materia Medica, Chinese Academy of Sciences in 2013. She is currently a postdoctoral fellow with Prof. Wenbin Lin at the University of Chicago. Her research focuses on combination therapy and immunotherapy of metastatic cancers using nanomedicine.



Christina Chan received a B.A. degree in chemistry from the University of Chicago in 2013. She is a PhD candidate in the NIH Chemistry-Biology Interface Predoctoral Training Program at the University of Chicago. Her research addresses the combination and delivery of active chemical and biological molecules for the treatment of solid cancers.



Wenbin Lin studied chemical physics at University of Science and Technology of China, received a Ph.D. in chemistry at University of Illinois at Urbana-Champaign, and carried out NSF postdoctoral research with Prof. Tobin J. Marks at Northwestern University. He is currently the James Franck Professor of Chemistry, Radiation and Cellular Oncology, and Ludwig Center for Metastasis Research at the University of Chicago. His group has pioneered the application of metal–organic frameworks in cancer therapy, bioimaging, earth-

abundant metal catalysis, artificial photosynthesis, asymmetric catalysis, and second-order nonlinear optics.

References

- [1]. Ribas A, Wolchok JD, Science 2018, 359, 1350 1355. [PubMed: 29567705]
- [2]. a) Chen DS, Mellman I, Immunity 2013, 39, 1 10; [PubMed: 23890059] b) Chen DS, Mellman I, Nature 2017, 541, 321 – 330. [PubMed: 28102259]
- [3]. a) Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD, Nat. Immunol 2002, 3, 991 998;
 [PubMed: 12407406] b) Schreiber RD, Old LJ, Smyth MJ, Science 2011, 331, 1565 1570.
 [PubMed: 21436444]
- [4]. a) Dewitte H, Verbeke R, Breckpot K, De Smedt SC, Lentacker I, Nano Today 2014, 9, 743 758;b) Sau S, Alsaab HO, Bhise K, Alzhrani R, Nabil G, Iyer AK, J. Controlled Release 2018, 274, 24 – 34.
- [5]. a) Carreno BM, Magrini V, Becker-Hapak M, Kaabinejadian S, Hundal J, Petti AA, Ly A, Lie W-R, Hildebrand WH, Mardis ER, Science 2015, 348, 803 – 808; [PubMed: 25837513] b) Palucka K, Banchereau J, Nat. Rev. Cancer 2012, 12, 265 – 277. [PubMed: 22437871]
- [6]. Dranoff G, Nat. Rev. Cancer 2004, 4, 11 22. [PubMed: 14708024]
- [7]. a) Restifo NP, Dudley ME, Rosenberg SA, Nat. Rev. Immunol 2012, 12, 269 281; [PubMed: 22437939] b) Amrolia PJ, Pule M, Lancet 2015, 385, 488 490. [PubMed: 25319502]
- [8]. a) Gubin MM, Zhang X, Schuster H, Caron E, Ward JP, Noguchi T, Ivanova Y, Hundal J, Arthur CD, Krebber W-J, Nature 2014, 515, 577 581; [PubMed: 25428507] b) Pardoll DM, Nat. Rev. Cancer 2012, 12, 252 264. [PubMed: 22437870]
- [9]. Ledford H, Nature 2015, 519, 17 18. [PubMed: 25739610]
- [10]. a) Zitvogel L, Kepp O, Senovilla L, Menger L, Chaput N, Kroemer G, Clin. Cancer Res 2010, 16, 3100 – 3104; [PubMed: 20421432] b) Dudek AM, Garg AD, Krysko DV, Ruysscher D. De, Agostinis P, Cytokine Growth Factor Rev. 2013, 24, 319 – 333. [PubMed: 23391812]
- [11]. a) Krysko DV, Garg AD, Kaczmarek A, Krysko O, Agostinis P, Vandenabeele P, Nat. Rev. Cancer 2012, 12, 860 – 875; [PubMed: 23151605] b) Kroemer G, Galluzzi L, Kepp O, Zitvogel L, Annu. Rev. Immunol 2013, 31, 51 – 72. [PubMed: 23157435]
- [12]. a) Obeid M, Tesniere A, Ghiringhelli F, Fimia GM, Apetoh L, Perfettini J-L, Castedo M, Mignot G, Panaretakis T, Casares N, Nat. Med 2007, 13, 54 61; [PubMed: 17187072] b) Panaretakis T, Joza N, Modjtahedi N, Tesniere A, Vitale I, Durchschlag M, Fimia G, Kepp O, Piacentini M, Froehlich K, Cell Death Differ. 2008, 15, 1499 1509. [PubMed: 18464797]
- [13]. a) Garg AD, Krysko DV, Verfaillie T, Kaczmarek A, Ferreira GB, Marysael T, Rubio N, Firczuk M, Mathieu C, Roebroek AJ, EMBO J. 2012, 31, 1062 1079; [PubMed: 22252128] b) Martins I, Wang Y, Michaud M, Ma Y, Sukkurwala A, Shen S, Kepp O, Metivier D, Galluzzi L, Perfettini J, Cell Death Differ. 2014, 21, 79 91. [PubMed: 23852373]
- [14]. a) Apetoh L, Ghiringhelli F, Tesniere A, Criollo A, Ortiz C, Lidereau R, Mariette C, Chaput N, Mira JP, Delaloge S, Andre F, Tursz T, Kroemer G, Zitvogel L, Immunol. Rev 2007, 220, 47 59; [PubMed: 17979839] b) Scaffidi P, Misteli T, Bianchi ME, Nature 2002, 418, 191 195. [PubMed: 12110890]
- [15]. a) Binder RJ, J. Immunol 2014, 193, 5765 5771; [PubMed: 25480955] b) Tesniere A, Panaretakis T, Kepp O, Apetoh L, Ghiringhelli F, Zitvogel L, Kroemer G, Cell Death Differ. 2008, 15, 3 – 12. [PubMed: 18007663]
- [16]. Obeid M, Panaretakis T, Joza N, Tufi R, Tesniere A, Van Endert P, Zitvogel L, Kroemer G, Cell Death Differ. 2007, 14, 1848 – 1850. [PubMed: 17657249]
- [17]. Apetoh L, Ghiringhelli F, Tesniere A, Obeid M, Ortiz C, Criollo A, Mignot G, Maiuri MC, Ullrich E, Saulnier P, Yang H, Amigorena S, Ryffel B, Barrat FJ, Saftig P, Levi F, Lidereau R, Nogues C, Mira JP, Chompret A, Joulin V, Clavel-Chapelon F, Bourhis J, Andre F, Delaloge S, Tursz T, Kroemer G, Zitvogel L, Nat. Med 2007, 13, 1050 – 1059. [PubMed: 17704786]
- [18]. Kono H, Rock KL, Nat. Rev. Immunol 2008, 8, 279 289. [PubMed: 18340345]

- [19]. Green DR, Ferguson T, Zitvogel L, Kroemer G, Nat. Rev. Immunol 2009, 9, 353 363.
 [PubMed: 19365408]
- [20]. a) Irvine DJ, Hanson MC, Rakhra K, Tokatlian T, Chem. Rev 2015, 115, 11109 11146;
 [PubMed: 26154342] b) Jo SD, Nam G-H, Kwak G, Yang Y, Kwon IC, Nano Today 2017, 17, 23 37;c) Ng KK, Lovell JF, Zheng G, Acc. Chem. Res 2011, 44, 1105 1113; [PubMed: 21557543] d) Zhang Z, Cao W, Jin H, Lovell JF, Yang M, Ding L, Chen J, Corbin I, Luo Q, Zheng G, Angew. Chem. Int. Ed 2009, 48, 9171 9175; Angew. Chem. 2009, 121, 9335 9339.
- [21]. a) Zheng G, Chen J, Li H, Glickson JD, Proc. Natl. Acad. Sci. USA 2005, 102, 17757 17762;
 [PubMed: 16306263] b) Shao S, Geng J, Yi HA, Gogia S, Neelamegham S, Jacobs A, Lovell JF, Nat. Chem 2015, 7, 438; [PubMed: 25901823] c) Zhang Y, Jeon M, Rich LJ, Hong H, Geng J, Zhang Y, Shi S, Barnhart TE, Alexandridis P, Huizinga JD, Nat. Nanotechnol 2014, 9, 631 638. [PubMed: 24997526]
- [22]. Elsabahy M, Wooley KL, Chem. Soc. Rev 2012, 41, 2545 2561. [PubMed: 22334259]
- [23]. a) Reddy LH, Arias JL, Nicolas J, Couvreur P, Chem. Rev 2012, 112, 5818 5878; [PubMed: 23043508] b) Dykman L, Khlebtsov N, Chem. Soc. Rev 2012, 41, 2256 2282. [PubMed: 22130549]
- [24]. a) Duan X, Li Y, Small 2013, 9, 1521 1532; [PubMed: 23019091] b) Hong E, Dobrovolskaia MA, Adv. Drug Deliv. Rev 2018, 10.1016/j.addr.2018.01.005;c) Sykes EA, Chen J, Zheng G, Chan WC, ACS Nano 2014, 8, 5696 5706; [PubMed: 24821383] d) Sun Y, Xia Y, Science 2002, 298, 2176 2179; [PubMed: 12481134] e) Lu Y, Yin Y, Mayers BT, Xia Y, Nano Lett. 2002, 2, 183 186.
- [25]. a) Homma T, Fujii J, Exp. Cell Res 2016, 349, 128 138; [PubMed: 27743894] b) Chen T, Guo J, Han C, Yang M, Cao X, J. Immunol 2009, 182, 1449 1459; [PubMed: 19155492] c) Slovak R, Ludwig JM, Gettinger SN, Herbst RS, Kim HS, J. Immunother. Cancer 2017, 5, 78. [PubMed: 29037259]
- [26]. Evans SS, Repasky EA, Fisher DT, Nat. Rev. Immunol 2015, 15, 335 349. [PubMed: 25976513]
- [27]. Nguyen HT, Tran KK, Sun B, Shen H, Biomaterials 2012, 33, 2197 2205. [PubMed: 22177288]
- [28]. Moy AJ, Tunnell JW, Adv. Drug Delivery Rev 2017, 114, 175 183.
- [29]. Toraya-Brown S, Sheen MR, Zhang P, Chen L, Baird JR, Demidenko E, Turk MJ, Hoopes PJ, Conejo-Garcia JR, Fiering S, Nanomed. Nanotechnol. Biol. Med 2014, 10, 1273 – 1285.
- [30]. a) Kobayashi T, Biotechnol. J 2011, 6, 1342 1347; [PubMed: 22069094] b) Kobayashi T, Kakimi K, Nakayama E, Jimbow K, Nanomedicine 2014, 9, 1715 – 1726. [PubMed: 25321171]
- [31]. a) Yanase M, Shinkai M, Honda H, Wakabayashi T, Yoshida J, Kobayashi T, Cancer Sci. 1998, 89, 775 782;b) Kawai N, Ito A, Nakahara Y, Futakuchi M, Shirai T, Honda H, Kobayashi T, Kohri K, Prostate 2005, 64, 373 381. [PubMed: 15754344]
- [32]. a) Jimbow K, Tamura Y, Yoneta A, Kamiya T, Ono I, Yamashita T, Ito A, Honda H, Wakamatsu K, Ito S, J. Biomater. Nanobiotechnol 2012, 3, 140 153;b) Jimbow K, Ishii-Osai Y, Ito S, Tamura Y, Ito A, Yoneta A, Kamiya T, Yamashita T, Honda H, Wakamatsu K, J. Skin Cancer 2013, 2013;c) Ito A, Yamaguchi M, Okamoto N, Sanematsu Y, Kawabe Y, Wakamatsu K, Ito S, Honda H, Kobayashi T, Nakayama E, Nanomedicine 2013, 8, 891 902. [PubMed: 23066648]
- [33]. Ito A, Tanaka K, Kondo K, Shinkai M, Honda H, Matsumoto K, Saida T, Kobayashi T, Cancer Sci. 2003, 94, 308 – 313. [PubMed: 12824927]
- [34]. Ito A, Matsuoka F, Honda H, Kobayashi T, Cancer Gene Ther. 2003, 10, 918 925. [PubMed: 14712318]
- [35]. Ito A, Matsuoka F, Honda H, Kobayashi T, Cancer Immunol. Immunother 2004, 53, 26 32.[PubMed: 14551746]
- [36]. Zou L, Wang H, He B, Zeng L, Tan T, Cao H, He X, Zhang Z, Guo S, Li Y, Theranostics 2016, 6, 762 – 772. [PubMed: 27162548]
- [37]. Yata T, Takahashi Y, Tan M, Nakatsuji H, Ohtsuki S, Murakami T, Imahori H, Umeki Y, Shiomi T, Takakura Y, Biomaterials 2017, 146, 136 145. [PubMed: 28918263]
- [38]. Guo L, Yan DD, Yang D, Li Y, Wang X, Zalewski O, Yan B, Lu W, ACS Nano 2014, 8, 5670 5681. [PubMed: 24801008]
- [39]. Tao Y, Ju E, Ren J, Qu X, Biomaterials 2014, 35, 9963 9971. [PubMed: 25224368]

- [40]. Han Q, Wang X, Jia X, Cai S, Liang W, Qin Y, Yang R, Wang C, Nanoscale 2017, 9, 5927 5934. [PubMed: 28436514]
- [41]. Zhou F, Wu S, Song S, Chen WR, Resasco DE, Xing D, Biomaterials 2012, 33, 3235 3242. [PubMed: 22296829]
- [42]. Li L, Yang S, Song L, Zeng Y, He T, Wang N, Yu C, Yin T, Liu L, Wei X, Theranostics 2018, 8, 860 – 873. [PubMed: 29344312]
- [43]. Kumar P, Srivastava R, Mater. Sci. Eng. C 2015, 57, 321 327.
- [44]. Wang C, Xu L, Liang C, Xiang J, Peng R, Liu Z, Adv. Mater 2014, 26, 8154 8162. [PubMed: 25331930]
- [45]. Cano-Mejia J, Burga RA, Sweeney EE, Fisher JP, Bollard CM, Sandler AD, Cruz CRY, Fernandes R, Nanomed. Nanotechnol. Biol. Med 2017, 13, 771 – 781.
- [46]. Chen Q, Xu L, Liang C, Wang C, Peng R, Liu Z, Nat. Commun 2016, 7, 13193. [PubMed: 27767031]
- [47]. Liu Y, Maccarini P, Palmer GM, Etienne W, Zhao Y, Lee C-T, Ma X, Inman BA, Vo-Dinh T, Sci. Rep 2017, 7, 8606. [PubMed: 28819209]
- [48]. a) Agostinis P, Berg K, Cengel KA, Foster TH, Girotti AW, Gollnick SO, Hahn SM, Hamblin MR, Juzeniene A, Kessel D, CA: Cancer J. Clin 2011, 61, 250 281; [PubMed: 21617154] b) Thong PS-P, Ong K-W, Goh NS-G, Kho K-W, Manivasager V, Bhuvaneswari R, Olivo M, Soo K-C, Lancet Oncol. 2007, 8, 950 952; [PubMed: 17913664] c) Castano AP, Mroz P, Hamblin MR, Nat. Rev. Cancer 2006, 6, 535 545. [PubMed: 16794636]
- [49]. Celli JP, Spring BQ, Rizvi I, Evans CL, Samkoe KS, Verma S, Pogue BW, Hasan T, Chem. Rev 2010, 110, 2795 – 2838. [PubMed: 20353192]
- [50]. a) Chatterjee DK, Fong LS, Zhang Y, Adv. Drug Delivery Rev 2008, 60, 1627 1637;b) Lucky SS, Soo KC, Zhang Y, Chem. Rev 2015, 115, 1990 2042. [PubMed: 25602130]
- [51]. Yu X, Gao D, Gao L, Lai J, Zhang C, Zhao Y, Zhong L, Jia B, Wang F, Chen X, ACS Nano 2017, 11, 10147 – 10158. [PubMed: 28901740]
- [52]. Marrache S, Choi JH, Tundup S, Zaver D, Harn DA, Dhar S, Integr. Biol 2013, 5, 215 223.
- [53]. Duan X, Chan C, Guo N, Han W, Weichselbaum RR, Lin W, J. Am. Chem. Soc 2016, 138, 16686
 16695. [PubMed: 27976881]
- [54]. Lan G, Ni K, Xu Z, Veroneau SS, Song Y, Lin W, J. Am. Chem. Soc 2018, 140, 5670 5673.
 [PubMed: 29665677]
- [55]. Wang D, Wang T, Liu J, Yu H, Jiao S, Feng B, Zhou F, Fu Y, Yin Q, Zhang P, Nano Lett. 2016, 16, 5503 – 5513. [PubMed: 27525587]
- [56]. Lu K, He C, Guo N, Chan C, Ni K, Weichselbaum RR, Lin W, J. Am. Chem. Soc 2016, 138, 12502 – 12510. [PubMed: 27575718]
- [57]. Song W, Kuang J, Li C-X, Zhang M, Zheng D, Zeng X, Liu C, Zhang X-Z, ACS Nano 2018, 12, 1978 – 1989. [PubMed: 29420012]
- [58]. Xu J, Xu L, Wang C, Yang R, Zhuang Q, Han X, Dong Z, Zhu W, Peng R, Liu Z, ACS Nano 2017, 11, 4463 – 4474. [PubMed: 28362496]
- [59]. Stangl S, Themelis G, Friedrich L, Ntziachristos V, Sarantopoulos A, Molls M, Skerra A, Multhoff G, Radiother. Oncol 2011, 99, 313 – 316. [PubMed: 21704400]
- [60]. Suzuki Y, Mimura K, Yoshimoto Y, Watanabe M, Ohkubo Y, Izawa S, Murata K, Fujii H, Nakano T, Kono K, Cancer Res. 2012, 72, 3967 3976. [PubMed: 22700877]
- [61]. a) Golden E, Pellicciotta I, Demaria S, Barcellos-Hoff MH, Formenti SC, Front. Oncol 2012, 2, 88; [PubMed: 22891162] b) Rubner Y, Wunderlich R, Rühle P-F, Kulzer L, Werthmçller N, Frey B, Weiss E-M, Keilholz L, Fietkau R, U. S. Gaipl, Front. Oncol 2012, 2, 75. [PubMed: 22848871]
- [62]. a) Prasad SJ, Farrand KJ, Matthews SA, Chang JH, McHugh RS, Ronchese F, J. Immunol 2005, 174, 90 98; [PubMed: 15611231] b) Schaue D, Ratikan JA, Iwamoto KS, McBride WH, Int. J. Radiat. Oncol. Biol. Phys 2012, 83, 1306 1310. [PubMed: 22208977]
- [63]. Mason KA, Hunter NR, Front. Oncol 2012, 2, 101. [PubMed: 22912936]
- [64]. a) Binder DC, Fu Y-X, Weichselbaum RR, Trends Mol. Med 2015, 21, 463 465; [PubMed: 26091823] b) Twyman-Saint Victor C, Rech AJ, Maity A, Rengan R, Pauken KE, Stelekati E,

Benci JL, Xu B, Dada H, Odorizzi PM, Nature 2015, 520, 373 – 377; [PubMed: 25754329] c) Deng L, Liang H, Burnette B, Beckett M, Darga T, Weichselbaum RR, Fu Y-X, J. Clin. Invest 2014, 124, 687 – 695. [PubMed: 24382348]

- [65]. Chiang C-S, Fu S-Y, Wang S-C, Yu C-F, Chen F-H, Lin C-M, Hong J-H, Front. Oncol 2012, 2, 89. [PubMed: 22888475]
- [66]. Schaue D, Xie MW, Ratikan JA, McBride WH, Front. Oncol 2012, 2, 90. [PubMed: 22912933]
- [67]. Demaria S, Formenti SC, Front. Oncol 2012, 2, 153. [PubMed: 23112958]
- [68]. Min Y, Roche KC, Tian S, Eblan MJ, McKinnon KP, Caster JM, Chai S, Herring LE, Zhang L, Zhang T, Nat. Nanotechnol 2017, 12, 877 – 882. [PubMed: 28650437]
- [69]. Vanpouille-Box C, Hindré F, Front. Oncol 2012, 2, 136. [PubMed: 23087900]
- [70]. a) Cikankowitz A, Clavreul A, Tétaud C, Lemaire L, Rousseau A, Lepareur N, Dabli D, Bouchet F, Garcion E, Menei P, J. Neuro-Oncol 2017, 131, 49 58;b) Jestin E, Mougin-Degraef M, Faivre-Chauvet A, Remaud-Le Saec P, Q. J. Nucl. Med. Mol. Imaging 2007, 51, 51 60;
 [PubMed: 17372573] c) Vanpouille-Box C, Lacoeuille F, Roux J, Aubé C, Garcion E, Lepareur N, Oberti F, Bouchet F, Noiret N, Garin E, PLoS One 2011, 6, e16926. [PubMed: 21408224]
- [71]. Vanpouille-Box C, Lacoeuille F, Belloche C, Lepareur N, Lemaire L, LeJeune J-J, Benoit J-P, Menei P, Couturier OF, Garcion E, Biomaterials 2011, 32, 6781 – 6790. [PubMed: 21705077]
- [72]. a) Albert ML, Sauter B, Bhardwaj N, Nature 1998, 392, 86 89; [PubMed: 9510252] b) Inoue H, Tani K, Cell Death Differ. 2014, 21, 39 – 49. [PubMed: 23832118]
- [73]. Zhao X, Yang K, Zhao R, Ji T, Wang X, Yang X, Zhang Y, Cheng K, Liu S, Hao J, Biomaterials 2016, 102, 187 – 197. [PubMed: 27343466]
- [74]. Zheng D-W, Chen J-L, Zhu J-Y, Rong L, Li B, Lei Q, Fan J-X, Zou M-Z, Li C, Cheng S-X, Nano Lett. 2016, 16, 4341 – 4347. [PubMed: 27327876]
- [75]. Su B, Cengizeroglu A, Farkasova K, Viola JR, Anton M, Ellwart JW, Haase R, Wagner E, Ogris M, Mol. Ther 2013, 21, 300 – 308. [PubMed: 23299796]
- [76]. a) Bagalkot V, Lee I-H, Yu MK, Lee E, Park S, Lee J-H, Jon S, Mol. Pharm 2009, 6, 1019 1028; [PubMed: 19338265] b) Mizuno Y, Naoi T, Nishikawa M, Rattanakiat S, Hamaguchi N, Hashida M, Takakura Y, J. Controlled Release 2010, 141, 252 259;c) Nishikawa M, Mizuno Y, Mohri K, Matsuoka N, Rattanakiat S, Takahashi Y, Funabashi H, Luo D, Takakura Y, Biomaterials 2011, 32, 488 494. [PubMed: 20932569]
- [77]. Liu Y, Qiao L, Zhang S, Wan G, Chen B, Zhou P, Zhang N, Wang Y, Acta Biomater. 2018, 66, 310 – 324. [PubMed: 29129789]
- [78]. Kuai R, Yuan W, Son S, Nam J, Xu Y, Fan Y, Schwendeman A, Moon JJ, Sci. Adv 2018, 4, eaao1736.
- [79]. Roy A, Singh MS, Upadhyay P, Bhaskar S, Mol. Pharm 2010, 7, 1778 1788. [PubMed: 20822093]
- [80]. Roy A, Chandra S, Mamilapally S, Upadhyay P, Bhaskar S, Pharm. Res 2012, 29, 2294 2309. [PubMed: 22547032]
- [81]. Roy A, Singh MS, Upadhyay P, Bhaskar S, Int. J. Pharm 2013, 445, 171 180. [PubMed: 23376226]
- [82]. Seth A, Heo MB, Lim YT, Biomaterials 2014, 35, 7992 8001. [PubMed: 24954733]
- [83]. Heo MB, Kim S-Y, Yun WS, Lim YT, Int. J. Nanomed 2015, 10, 5981.
- [84]. Tao Y, Ju E, Liu Z, Dong K, Ren J, Qu X, Biomaterials 2014, 35, 6646 6656. [PubMed: 24818880]
- [85]. He C, Duan X, Guo N, Chan C, Poon C, Weichselbaum RR, Lin W, Nat. Commun 2016, 7, 12499. [PubMed: 27530650]
- [86]. Yang G, Xu L, Chao Y, Xu J, Sun X, Wu Y, Peng R, Liu Z, Nat. Commun 2017, 8, 902. [PubMed: 29026068]



Figure 1.

The cancer-immunity cycle and therapies that might affect the cycle.



Figure 2.

In immunogenic death, dying tumor cells expose CRT, secrete ATP, and release HMGB-1, HSPs, TSAs, and HSP-TSA complexes, all of which favor the engulfment of cell corpses and debris by antigen-presenting cells (mainly DCs) and promote DC maturation. Activated DCs can then prime CD4⁺ and CD8⁺ T cells and thereby trigger immunogenic T helper 1 (Th1) cell and cytotoxic T lymphocyte (CTL) responses, respectively. Adapted from Ref. [19] with permission. Copyright 2009, Nature Publishing Group.

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Figure 3.

Formulation of PLGA-ICG-R837 and their immune-stimulation abilities. a) Structure of PLGA-ICG-R837 and the mechanism of antitumor immune responses induced by PTT in combination with anti-CTLA-4. b) Treatment schedule to induce a tumour vaccine and reject secondary and rechallenge tumors. Adapted from Ref. [46] with permission. Copyright 2016, Nature Publishing Group.

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Figure 4.

ZnP@pyro PDT enhances PD-L1 blockade immunotherapy. a) The core–shell structure of ZnP@pyro. b,c) The treatment with PDT plus anti-PD-L1 eradicated primary 4T1 cancer and reduced the metastases in the lung. Adapted from Ref. [53] with permission. Copyright 2016, American Chemical Society.



Figure 5.

Fe-TBP decomposes H_2O_2 to O_2 through a Fenton-like reaction, then converts O_2 to cytotoxic 1O_2 upon light irradiation (top). Fe-TBP overcomes tumor hypoxia for PDT-primed cancer immunotherapy. The Fe-TBP catalyzed Fenton-like reaction leads to significant antitumor response and sensitizing anti-PD-L1 treatment to induce abscopal effect (bottom). Reprinted from Ref. [54] with permission. Copyright 2018, American Chemical Society.



Figure 6.

Acid-activatable micelleplexes for the combination of PDT with PD-L1 knockdown. a) Chemical structure of the acid-activatable POP micelleplexes co-loaded with PPa and siRNA. The micelleplexes dissociate in an acidic microenvironment owing to the protonation of the tertiary amino groups of PDPA. b) POP-PD-L1 micelleplex induction of ROS and release of RNAi upon cell uptake. c,d,e) Metastatic inhibition in metastatic tumorbearing mice. Reprinted from Ref. [55] with permission. Copyright 2016, American Chemical Society.



Figure 7.

a) Structure of the chimeric peptide PpIX-1MT. The PpIX-1MT NPs accumulated in the tumor area through the EPR effect, activated the CD8⁺ T cells by a series of cascade activations, and inhibited both the primary tumor and lung metastasis effectively. b) In situ PDT in the primary tumor caused apoptosis of tumor cells, production of caspase-3, and release of 1MT from PpIX-1MT NPs. Reprinted from Ref. [57] with permission. Copyright 2018, American Chemical Society.



Figure 8.

a) Schematic of utilizing AC-NPs to improve cancer immunotherapy. Radiation of the primary tumor induces the release of antigens, which were captured by AC-NPs and presented to DCs. The improved immune activation combined with anti-PD-1 treatment eradicates the unirradiated secondary tumor. b,c) Binding of unique TAAs to AC-NPs with different surface chemistry. Reprinted from Ref. [68] with permission. Copyright (2017), Nature Publishing Group.

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Figure 9.

Chemotherapy and PDT of NCP@pyrolipid potentiate PD-L1 blockade to induce systemic antitumor immunity. a) Chemotherapy and PDT of NCP@pyrolipid induce ICD and an inflammatory environment, leading to the release of TAAs, which are processed and presented by infiltrated APCs, to elicit the proliferation of tumor-specific effector T cells in lymphoid organs. b) Combined with PD-L1 blockade, the NCP@pyrolipid chemotherapy/PDT significantly resulted in tumor eradication in the primary sites and also a systemic anti-tumor immune response to reject distant tumors. Adapted from Ref. [84] with permission. Copyright 2016, Nature Publishing Group.