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Author manuscript *Semin Cell Dev Biol.* Author manuscript; available in PMC 2021 April 01.

Published in final edited form as:

Semin Cell Dev Biol. 2020 April; 100: 52-61. doi:10.1016/j.semcdb.2019.10.003.

# Homeobox gene Meis1 modulates cardiovascular regeneration

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# Abstract

Regeneration of cardiomyocytes, endothelial cells and vascular smooth muscle cells (three major lineages of cardiac tissues) following myocardial infarction is the critical step to recover the function of the damaged heart. Myeloid ecotropic viral integration site 1 (Meis1) was first discovered in leukemic mice in 1995 and its biological function has been extensively studied in leukemia, hematopoiesis, the embryonic pattering of body axis, eye development and various genetic diseases, such as restless leg syndrome. It was found that Meis1 is highly associated with Hox genes and their cofactors to exert its regulatory effects on multiple intracellular signaling pathways. Recently with the advent of bioinformatics, biochemical methods and advanced genetic engineering tools, new function of Meis1 has been found to be involved in the cell cycle regulation of cardiomyocytes and endothelial cells. For example, inhibition of Meis1 expression increases the proliferative capacity of neonatal mouse cardiomyocytes, whereas overexpression of Meis1 results in the reduction in the length of cardiomyocyte proliferative window. Interestingly, downregulation of one of the circular RNAs, which acts downstream of Meis1 in the cardiomyocytes, promotes angiogenesis and restores the myocardial blood supply, thus reinforcing better regeneration of the damaged heart. It appears that Meis1 may play double roles in modulating proliferation and regeneration of cardiomyocytes and endothelial cells post-myocardial infarction. In this review, we propose to summarize the major findings of Meis1 in modulating fetal development and adult abnormalities, especially focusing on the recent discoveries of Meis1 in controlling the fate of cardiomyocytes and endothelial cells.

# Keywords

Homeobox gene; Meis1; cardiomyocytes; endothelial cells; regeneration; heart

Conflict of interest

Authors disclose no conflict of interest.

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# 1. Introduction

A successful clinical application of cardiac regeneration would save almost 3.7 million lives per year worldwide caused by myocardial infarction (MI)-induced heart failure (HF) [1]. To this end, stem cell-based therapy for heart regeneration has attracted more attention in the past years. However, current clinical trials using stem cells alone shows certain drawbacks of low cell viability and low retention following transplantation and these issues have not been overcome yet. Combinatorial approaches, such as gene-based therapy together with stem cell, may facilitate the regeneration of damaged heart post-MI [2]. In the past two decades, researchers have highlighted multiple genes, such as *Vegf* and *Angiopoietin-2* [2, 3], which are involved in the development and regeneration of the cardiovascular system, as potential therapeutic targets [4, 5]. The recent discovery of *Myeloid ecotropic viral integration site 1* (*Meis1*) in regulating cardiomyocyte (CM) cell cycle [6] is one of them, which has opened a new window for researchers to unravel its role in the regeneration of the cardiovascular system.

Homeodomain containing transcription factors are the conserved group of proteins known as the Hox proteins, which are organized into chromosomal clusters and have been established as key players in the patterning of body axis [7]. These transcription factors are known to be involved in various aspects of developmental processes during embryogenesis (reviewed in [8]). Apart from embryonic development, Hox proteins are also involved in several processes at the adult stage, such as hematopoiesis [9–13], angiogenesis [9], apoptosis [14], receptor signaling as well as in diseased conditions like oncogenesis [15–21]. Meis1 belongs to the family of highly conserved three amino-acid loop extension (TALE) homeobox class that was first identified as a common viral integration site in myeloid leukemic cells of BXH-2 mice [21].

Ever since its discovery, researchers have found the roles of Meis1 in limb development [22] and several pathological disorders, including restless leg syndrome [23, 24], eye defects [25], cardiac defects [26] *etc.*, which has drawn tremendous amount of attention to explore this protein as a therapeutic target. It was found that deficiency of Meis1 in murine results in lethality at embryonic day 14.5 with reduction of megakaryocytes and definitive hematopoietic stem cells (HSCs) along with malformed capillaries and smaller eye lenses [13, 24, 27]. Furthermore, haploinsufficiency of Meis1 causes micro-ophthalmic traits and visual impairment in adult mice, suggesting its role in varied cellular and developmental processes in a Hox-Pbx independent manner in contrast to the leukemic conditions [25].

Since majority of the previous studies have focused on leukemia, the detailed mechanism regarding *Meis1* in modulating leukemogenesis has been well studied (reviewed in [12]); however, its molecular responsibilities in normal hematopoiesis, mammalian vascular network, and cardiac regeneration remains largely unknown although clear phenotypic changes have been observed in the animal models. In this review, we have provided a comprehensive picture of this gene in regulating organogenesis during the embryonic development and the progression of various diseases, eventually leading the readers in the path of unraveling the role of *Meis1* in modulating cardiovascular system, especially CMs and endothelial cells (ECs) - the two critical cardiac components.

### 2. Meis1 is an established Hox co-factor

Functions of *Hox* genes depend on the interaction with their cofactors. By increasing the DNA binding efficiency and specificity of the Hox proteins, these cofactors regulate various highly complex and ordered cellular processes like hematopoiesis [10]. Among all *Hox* cofactors, *Pbx1* and *Meis1* have established themselves as key regulators of differentiation and maturation of HSCs whose expression follows a similar trend as *Hox* genes during embryonic development and at the later stages [28]. Comprehensive knowledge of the protein structure and domain organization will enable us to gain insight into Meis1's mechanism of action where it binds to the DNA as well as to other proteins in order to form hetero dimeric, trimeric or tetrameric complexes.

Meis1 generally forms dimers with its common partner Pbx1, although it can also interact with other proteins to form higher-order complexes. Through a varied combination of interactions, the transcription of each Hox-regulated gene is controlled at different developmental stages by forming a Hox regulatory protein complex mostly with Meis1, Pbx1 and Prep1 [29]. These three cofactors share a common structural organization containing a DNA-binding homeodomain towards the carboxy-terminus and two proteinprotein interaction domains towards the amino-terminus [30, 31]. The characteristic feature of these proteins is the presence of the 60 residues long helix-loop-helix DNA-binding homeodomain. The loop connecting helix-1 and helix-2 is a highly conserved sequence of three amino acids, which gives this protein family its name -Three Amino acid Loop Extension (TALE) [30]. Two protein-protein interaction domains enable the accommodation of additional proteins which depends on the cell-type and developmental stage. It has been well documented through various studies demonstrating that the combination of different proteins in the Hox regulatory complex gives rise to patterning of organs in the embryo and at the later stage of development in maintaining hematopoiesis or in the progression of leukemia [14–17, 20].

#### 2.1. Embryonic development

Embryonic patterning and development are centrally governed by homeobox-containing transcription factors whose functions are partly regulated by their co-factors, like *Meis1* and *Prep1* [29]. As mentioned earlier, the phenotype of *Meis1* deficient mice, which die at midgestation due to fetal hematopoietic failure, marks its importance as an essential developmental gene [13]. During organogenesis, *Meis1* is expressed and has functional roles in many organs and embryonic structures, such as in the central nervous system and sensory structures of the head, including the ears, eyes, nose [27, 32, 33] as well as in the four cardiac chambers [26], lungs and limb [22, 34]. These observations collectively underscore the molecular mechanisms determined by Hox-Pbx-Meis1 complex in the mammalian embryo, which is briefly addressed for different stages of development in the following sections.

The function of Hox-Pbx-Meis1 protein complex in limb patterning was already verified in an earlier study, which shows Meis1 cooperatively binds with Pbx1 in a unique way independent of the tryptophan residue amino-terminal to the promoter of CYP17 gene containing cAMP-responsive sequence (CRS1). This leads to promotion of growth and

differentiation via modulating c-AMP-dependent transcriptional pathway [35] [36]. Further studies on vertebrate limb growth showed that expressions of Meis1 and Pbx1 are restricted in the proximal regions of the vertebrate limb where *Meis1* controls *Pbx1* activity by promoting nuclear import of the Pbx1 protein. This process is essential for cell fate specification and differentiation patterns along the proximodistal (P-D) axis of limb giving rise to distinct limb domains during development. The mechanism was also observed in Drosophila *melanogaster*, chicken, and murine embryonic development hinting toward its conserved nature [22]. Moreover, expression analysis and functional assays in mouse myoblast cell line demonstrated that Meis1 accompanied by c-AMP responsive element binding protein (Creb) and Nuclear transcription factor Y (Nf-Y) cooperatively bind to the conserved promoter/enhancer site of *Myostatin (MSTN). MSTN* is a key player of vertebrate skeletal muscle growth during early myogenesis, which acts as a strong inhibitor of skeletal muscle growth in vertebrates [37]. Together, these observations indicate that *Meis1* not only associates with Pbx1, but also becomes an ally of other DNA binding proteins, like Creb and Nf-Y, in limb patterning and development.

In a *pbx* dependent manner, *meis1* is shown to be responsible for proper hindbrain segmentation in zebrafish where the gene is expressed in bilaterally symmetric regions of ventral telencephalon [38–40]. In mice, it was identified to act upstream of Paired box protein 6 (Pax6) and positively regulate its expression by binding to its enhancer. This interaction is essential for lens ectoderm development and brings essential specification of the eyes [32, 33]. A study of *Meis1* loss-of-function and conditional functional rescue has further provided insights into its role in series of events throughout eye development, in which Meis1 binds to Hox/Pbx independent sites during optic cup development, eye patterning, retinal proliferation and differentiation [25]. This is the first time when *Meis1* has been reported to be functional without any association of *Pbx-Hox* genes and making it as a part of the *Notch* signaling pathway. It will be interesting to find out the Hox independent action of Meis1 that can establish its multifaceted mechanisms of action.

#### 2.2 Leukemia

Accumulation of multiple genetic alterations results in disruption of the normal growth and differentiation mechanism, which is believed to be the main cause behind many cancers, including leukemia. Studies have shown that Meis1 locus in chromosome 11 serves as a viral integration site in 15% tumors arising in BXH-2 mice, thus resulting in expression of an ecotropic murine leukemia virus that alters expression of cellular proto-oncogenes leading to myeloid leukemia [21]. Concomitant with this discovery, the MEIS1 locus 2p23-p12 in human is also subjected to chromosomal translocations, thereby altering gene expression in human leukemia. Since Meis1 is found to be frequently upregulated in human acute myeloid leukemia (AML) and acute lymphoid leukemia (ALL) [18, 41, 42], the characterization of this protein and its interacting partners have given us more insights into understanding the blood and bone marrow cancers.

It is believed that Meis1 acts as a cofactor of HoxA7 and HoxA9 in induction of AML of mouse model [21, 43, 44], suggesting its role as a Hox-cofactor to facilitate DNA binding specificity and increase the efficiency of Hox proteins. This explanation of Meis1's role

coincides with the observation from differential expression study throughout murine hematopoietic ontogeny, where the expression profiles of Meis1 and Pbx1 (already established as a Hox cofactor) closely resemble with that of Hox genes during different stages of hematopoiesis [28]. Further investigation confirms that along with forming a dimeric DNA-binding complex with HoxA9 [45], Meis1's Pbx interaction domain binds to Pbx2 and Pbx3 in order to collaborate with Hox proteins forming a Meis1-Pbx-Hox trimeric complex in myeloid progenitors [12, 16, 46].

The molecular mechanism underlying oncogenicity of Meis1 has been revealed from its association with HoxA9 and Pbx, which stabilizes the complex in order to promote transcription of *Cyclin dependent kinase 6 (Cdk6), Fms related tyrosine kinase 3 (Flt3), (Myb proto-oncogene (c-Myb)*, resulting in hyper-proliferation of myeloid leukemic cells (Figure 1) [16, 17, 20]. Meanwhile, activation of *B Lymphoma Mo-MLV Insertion Region 1 (Bmi1)* through this complex leads to down-regulation of *Cyclin dependent kinase inhibitor 2a (Cdkn2a)* augmenting the process of cell proliferation (Figure 1) [20, 47].

# 2.3 Hematopoiesis

Hematopoiesis is a fundamental process of formation of blood cellular components during embryonic development and throughout life. It is well established that hematopoiesis is dependent on *Hox* genes, which are found to be preferentially expressed in HSC-enriched subpopulations followed by a downregulation through differentiation and maturation [10]. This pattern was observed at both adult and fetal stages of hematopoiesis. The trend of expression level change in *Hox* genes in HSCs is consistent with that of *Meis1*, whereas *Pbx1* follows a different expression pattern [28]. Knockout studies found that mice lacking *Meis1* show extensive hemorrhaging in the trunk and die at embryonic day 14.5 due to failure in definitive hematopoiesis. The number of HSCs in Meis1<sup>-/-</sup> fetal liver is found to be dramatically reduced, where the cells fail to protect themselves in irradiated mice [13, 27, 48], denoting the importance of *Meis1* mediated *Hox* regulated processes in hematopoiesis.

It is well known that HSCs use glycolytic metabolism to meet their energy demands [14, 49]. Inducible deletion of Meis1 in adult HSCs results in downregulation of Hif-1alpha (Hif-1 $\alpha$ ) and Hif-2alpha (Hif-2 $\alpha$ ), thereby increasing their oxygen consumption and decreasing the glycolytic flux [14]. This results in the production of higher level of Reactive Oxygen Species (ROS) associated with increased expression of proteins p16(INK4) and p19(ARF) (Figure 2). These findings imply that Meis1 acts upstream of a transcriptional network that regulates metabolism and oxidative stress response of HSCs [14].

#### 2.4. Vascular Network

Vasculogenesis, angiogenesis and arteriogenesis are three common processes to form vertebrate blood vessel network. Vasculogenesis is defined by the aggregation of angioblasts for *in situ* differentiation and growth of blood vessels [50–52], while angiogenesis is the process where new blood vessels sprout from the existing ones [53]. Arteriogenesis refers to a process where collateral arteries are promoted by increasing radius of existing arteries to accommodate more supply of blood [54, 55].

Depending on the growth of each organ in the body, the requirement of blood to supply an adequate amount of oxygen for every single cell increases, thus maintaining normal tissue function. Angiogenesis and arteriogenesis play a big role in maintaining the health of each tissue and organ by branching into several vessels and capillaries. Unraveling the mechanism of blood vessel formation can offer us therapeutic options to improve or even cure ischemic disorders that are now one of the leading cause of mortality [56]. For example, to prevent proliferation of the cancerous cells in a solid tumor, impediment of angiogenesis could act as a therapeutic procedure, whereas the condition of patient with coronary artery disease can be ameliorated by stimulating angiogenic factors to increase either arteriogenesis or angiogenesis at the site of infarction. The basic frameworks of vasculature are conserved among vertebrates, which make it possible to assign homologies between distinct blood vessels and to directly compare the formation of these vessels in different vertebrate species [57].

**2.4.1 Meis1 is a key gene regulating vertebrate vasculogenesis**—Zebrafish is an ideal model to study hematopoiesis and vascular development due to their externally fertilized and transparent embryos, which enable *in vivo* visualization of early embryonic processes from birth of HSCs in the mesoderm to migration of blood cells [11, 58–60]. In addition, 70% of genetic similarities between human and zebrafish makes it advantageous for experimental and genetic analysis of vascular development [61].

Kawahara et al. reported, for the first time, that meis1 plays a role in zebrafish vascular network development by regulating ECs [62]. Differentiation of ECs and the formation of vasculature, *i.e.* the *de novo* emergence of vascular network, by endothelial progenitor cells [63] in *meis1* morphants (MO) were found to be affected with abnormal patterning of pan EC markers, such as platelet and endothelial cell adhesion molecule 1 (pecam1, also known as cd31) and friend leukemia integration 1 (fli1). Diminished expression of artery markers, such as eph related receptor tyrosine kinase ligand B2 (ephrinB2), with upregulation of vein markers, such as fms related tyrosine kinase 4 (flt4) and eph receptor B4 (ephB4), in presumptive arterial vessels meis1 MO imply that meis1 is involved in arterial vessel development. At the same time, expression of fetal liver kinase 1(flk1, also known as kdr or *vegfr*) gene was found to be significantly reduced in *meis1* morpholino injected zebrafish embryo, suggesting direct regulation of this vascular endothelial growth factor receptor (vegfr) in vertebrate embryonic development [62]. Epistasis analysis on zebrafish has concomitantly resulted in a similar outcome, where it is found to be involved in hemangioblast formation acting downstream of hoxd4a (Figure 3) [9]. Hemangioblasts are the common precursors of the blood and endothelial lineages, malformation of which directly affects the three fundamental processes in the body namely, vasculogenesis, angiogenesis, and hematopoiesis.

*Meis1* global KO mice developed ocular and angiogenesis defects with failure to produce megakaryocytes and displaying extensive hemorrhaging [27, 64]. Meis1 was found to be expressed in specific and discrete locations throughout the mouse embryo, including the aorta-gonad-mesonephros (AGM) mesenchyme, in the hemogenic embryonic arterial endothelium and in the hematopoietic clusters within the arteries [13]. In all three *Meis1* expressing cell populations, impaired expression of a critical endothelial transcription factor,

*Runt-related transcription factor 1 (Runx1)*, caused disruption of vascular patterning probably through affecting ECs [13]. A comparable phenotype in zebrafish and mice indicate that *Meis1*'s role in vascularization is conserved throughout the vertebral species.

HSC differentiation gives rise to several types of cells, of which hemogenic ECs are the ones involved in aorta formation [65]. A chemical defined hematopoietic differentiation model accompanied by whole genome gene profiling study as well as CRISPR/CAS9 gene editing technology recognized *MEIS1* acting as a regulator in generation of hemogenic endothelial cells, megakaryopoiesis and thrombopoiesis in human pluripotent stem cells through targeting endothelial cell-specific genes, *TAL1* and *FLI1* [66]. Both *TAL1* and *FLI1* are known to control angiogenic response of ECs through stimulating cell morphogenesis and angiogenic migration [67, 68]. These results give us an insight into the regulation of vascular patterning in vertebrates via maturing ECs from its progenitors through *MEIS1*. Further investigation into EC specific *MEIS1*'s role will strengthen our understanding of vasculogenesis, and arteriogenesis.

**2.4.2.** A possible relationship between Meis1 and EC proliferation—The above discussion provides us perception that the regulation of vascular patterning in vertebrates is likely through modulation of either mature ECs of its progenitors by *Meis1*. Consequently, investigation into EC-specific role of *Meis1* will further strengthen our understanding of angiogenesis and/or arteriogenesis through EC proliferation. The reduced expression of EC markers as well as molecular players in *vegf* signaling pathway due to Meis1 downregulation in the above-mentioned studies in zebrafish and murine places this gene, which was previously known to be as a proto-oncogene followed by known to be a cardiogenic transcription factor, in the list of regulators of EC regulation.

Interestingly, Meis1's role in the regulation of hemogenic ECs that are required for aorta formation by acting upstream of endothelial-specific proteins, TAL1 and FLI1, demonstrate that Meis1 is a crucial factor in EC lineage specification and maintenance. Both TAL1 and FLI1 are known to control angiogenic response of ECs by stimulating cell morphogenesis and angiogenic migration [67, 68]. The study on cardiac super-enhancers [69] show injection of Nfix shRNA at the infarcted site of adult heart results in comparatively higher blood vessels through angiogenesis than those in the heart of wild-type (WT) animal. As it is already established by the study that Meis1 positively regulates expression of Nfix by binding to its super-enhancer region, downregulation of this circular RNA leads to better recovery from myocardial infarction [69]. Collectively, these data suggest a inhibitory role of Meis1 in the process of angiogenesis.

#### 2.5 Other genetic diseases

Other than leukemia, Meis1 is known to be associated with restless leg syndrome (RLS), in which several non-synonymous variants in Meis1 have been identified in RLS patients [23, 70]. Meis1 locus is the most important RLS susceptibility gene [71]. Similar to RLS, Meis1 is also involved in locomotor hyperactivity [72]. It was found that Meis1 is strongly expressed in dopaminergic neurons of the substantia nigra, in the spinal cord and, in the red nucleus [73]. As a part of a transcriptional regulatory network governing motor neurons, it is

believed that Meis1 plays a critical link to the pathophysiology of RLS. The involvement of Meis1 in eye development mentioned early suggests its role in microphthalmia and genetic eye defects [25].

# 3. Meis1 in the cardiovascular system

Human cardiac tissue is probably the most difficult part in the body to regenerate following ischemic damage (*i.e.*, MI). The underlying principle that inhibits cardiac regeneration is the fact that adult CMs quickly lose their ability to proliferate after birth and remains an extreme low ability of regeneration in contrast to the hearts of amphibians and zebrafish [74]. However, the most important discovery in the field of cardiac regeneration is that neonatal mouse heart can efficiently regenerate until 7 days through proliferation of CMs, which lays a foundation for researchers to study the genetic and epigenetic markers that can potentially preserve this property of CMs beyond this proliferative window even at the adult stage [75].

#### 3.1 Cardiac development

Meis1 has previously been found to play vital roles in cardiogenesis and heart development in zebrafish and murine [26, 76], suggesting it is a conserved cardiac-specific gene across species; however, its expression level in heart changes dramatically from the embryonic stage to the adult stage with very low level of Meis1 protein in adult heart [77]. The implication of stage-dependent Meis1 expression remains largely unknown in mammalian heart development and maintenance (Table 1).

**3.1.1 Embryonic stage**—Studies found that Meis1 homozygous deletion results in lethality in mice with death occurring between embryonic days 11.5 and 14.5 due to internal hemorrhages and hematopoietic defects [13, 27], suggesting Meis1 as an important molecular player in embryonic development. By knockdown (KD) of *Meis1* by anti-sense mRNA, meis1 morphants exhibit heart edema and malformed heart loop, along with weak heartbeats and abnormal circulation of blood cells in zebrafish [11, 62]. Disruption of Meis1 also leads to malformed cardiac outflow tracts with overriding the aorta and ventricular septal defect in mice [26]. These findings signify that Mei1 exerts important regulatory impacts on CMs, ECs, and/or vascular smooth muscle cells (VSMCs) during embryonic development.

Heart development is a complex process with a range of differentiation steps where cardiogenic mesoderm differentiates into endoderm and myocardium [78]. Generation of myocardium involves three sequential transitions from cardiac precursors to primitive CMs and finally definitive CMs [79]. Although it is recognized that Meis1 is an important cardiac-related gene, based on phenotypic changes in the animal model, its mechanism in modulating cardiogenesis at different stages is not explored to a large extent; however, a few studies that have resulted in bridging the gap of knowledge in this regard draw a clear and interesting picture. Investigation of epigenetic regulation of cardiac lineage development showed MEIS1HOXA9 motif was enriched at a subset of enhancers at the cardiac precursor stage. Enhancer-based gene network analysis showed Meis1 along with Gata4 activate the enhancer of *Myocd*, an essential gene in differentiation and proliferation of VSMCs [80].

This therefore validates the proposal by Stankunas *et al.* that Hox proteins are functionally associated with Meis1 partners during heart development at the molecular level [26].

Series of events during cardiogenesis takes place through controlled expression of several temporal genes that result in chromatin and histone modifications [81]. At the embryonic stage, histone chaperone HIRA modulates gene expression through deposition of histone variant H3.3, which is an euchromatic marker of gene activity [82]. Conditional KO of HIRA in embryonic mouse hearts leads to cardiac septal defects accompanied by differential down-regulation of *Meis1*, *Tbx2* and *Gata6* [83]. These data suggest that HIRA directly binds the enhancers of *Meis1* and *Gata6* and the promoter of *Tbx2* activate their expression (Figure 4). A combination of ChIP-Seq and RNA-Seq studies in cardiac development has found that Meis1 and Nkx2–5 sequential binds to the enhancers of several cardiac regulatory genes, like *Popeye domain-containing protein 2 (Popdc2)* [84]. Together, these studies give us a partial picture of upstream and downstream genes of Meis1, where HIRA binds to the enhancer of Meis1 and activates its expression, which then triggers activation of several down-stream cardiac-specific genes during myocardial differentiation.

**3.1.2** Neonatal and adult stage—As mentioned earlier, *Meis1* regulates cell cycle exit and regenerative capacity of neonatal CMs [6]. *Meis1* deletion in neonatal mouse heart extended CMs proliferative window from 7 to 14 days. Additionally, by deleting *Meis1* in adult CMs, cyclin-dependent kinase inhibitors were down-regulated, which were usually upregulated in the postnatal stage, resulting in cell cycle re-entry and promoting CMs proliferation without cell hypertrophy or deleterious effect on cardiac function[6]. In contrast, overexpression of *Meis1* in neonatal CMs reduced CMs proliferation and inhibited the heart regeneration response[6]. Xiang *et al* [85] found Meis1 was directly bound and repressed by Tbx20 (a cardiac-specific gene), specifically in adult CM, to promote adult CM proliferation and preserve cardiac performance after myocardial infarction (Figure 4).

In addition, Meis1 was also found to inhibit angiotensin II-induced CMs hypertrophy by binding to *poly (rC)-binding protein 2 (PCBP2)* promoter and promoting its expression [86]. Expression of Meis1 is dramatically down-regulated in human failing heart tissues and murine hypertrophic heart tissues. Meanwhile, angiotensin II-induced hypertrophic is significantly inhibited by *Meis1* overexpression or promoted by *Meis1* knockdown [86]. *Meis1* inactivation in the mouse neural crest leads to altered sympatho-vagal regulation of cardiac rhythmicity in adults, which is characterized by chronotropic incompetence and cardiac conduction defects, thus increasing susceptibility to sudden cardiac death [73]. The role of *Meis1* as an anti-cardiomyocyte proliferative gene was again validated by Pandey *et al.* when they found translational inhibition of Meis1 by three micro RNAs (miRNA), miR-548c-3p, miR-509–3p, and miR-23b-3p increases proliferation of cardiomyocytes significantly [87] (Figure 5).

In the past decade, non-coding RNAs have emerged as vital molecular players in maintaining cardiac health by regulating gene network either in cell cycle progression of cardiomyocytes or induction of certain molecular phenomenon in the heart under healthy as well as diseased conditions. A recent study by Huang *et al.* has discovered that one of the crucial non-coding circular RNAs implicated in the cardiovascular system is regulated by

Meis1 via binding to its super-enhancer region [69]. Silencing of this conserved mammalian circular RNA Nfix has a similar phenotype as that of cardiomyocyte-specific *Meis1* KO, while cardiac regeneration of post-MI is greatly enhanced in circRNA Nfix KD adult mice in comparison to that of WT mice, in which enhancing angiogenesis is resulted by ubiquitin-dependent degradation of Ybx1 and rescuing miRNAmiR-214. In contrast, overexpression of this circular RNA inhibited the neonatal cardiac regeneration post-MI [69].

The mammalian heart undergoes a series of coordinated metabolic changes from the embryonic stage to the postnatal stage [88]. Maturation of the fetal heart is accompanied by higher myocardial metabolic activity switching from glycolysis to oxidative phosphorylation in order to produce ATP in an efficient manner [88]. This transition of mitochondrial activity is closely linked with cell proliferation, where higher metabolic activity reinforces the event of cell division [89]. A recent study on fetal, neonatal, and adult sheep CMs by Drake et al. has revealed that Meis1 is a key regulator of metabolic activity switch at birth corroborating with the loss of proliferative capacity of post-natal mammalian CMs. Suppression of Meis1 expression leads to increased oxygen consumption in fetal CMs, presumably mimicking the characteristics of adult stage, but no effect was found in the CMs isolated from neonatal sheep in contrast to the results by Mahmoud *et al.* [6], which is probably due to the difference in the nature of CMs at the post-natal stage between murine and sheep. siRNA mediated Meis1 knockdown resulted in downregulation of important metabolic genes glycolytic genes aldolase (ALDO), enolase (ENO) and post-glycolytic gene lactate dehydrogenase B (LDHB), which may underscore its role as CM metabolic regulator [77]. This observation implicates a normal down-regulation of Meis1 with age to provide a switch in oxidative metabolism [77].

The change in metabolic activity of neonatal and adult CMs explains the sudden change of expression of Meis1 between two stages (neonatal vs adult). It is important to understand what makes Meis1 as a negative regulator of cardiac regeneration post-MI even though it is expressed minimally at the post-natal stage. The previous results indicate an interaction between metabolic activity of CMs and cell cycle, which is regulated through Meis1 likely under different mechanisms, depending on the age of CM.

#### 3.2. Cardiac regeneration through augmented vascularization

As stated above, exogenous stem cell therapy for infarcted heart still faces several challenges in terms of techniques and logistics, such as lack of availability of blood supplies to the newly transplanted cells sitting on the scar tissue, efficiency of the delivery system and low cell viability and low retention. In order to increase the efficiency of cardiac regeneration or survivability, the newly transplanted CMs located in the post-infarct environment must be supplied for a sufficient supply of oxygenated blood to meet the required myocardial oxygen demand. To this end, angiogenic pre-treatment in the scar tissue by stimulating endothelial growth or vascular proliferative genes would initiate angiogenesis or arteriogenesis [90].

Animal study shows that EC angiogenesis play an essential role in CM regeneration post-MI by supplying blood perfusion to the infarcted heart and guiding CM migration [3], thus defining the important sequence of events that has to take place in order to promote myocardial regeneration or repair, *i.e.*, angiogenesis precedes cardiomyocyte migration. Any

stimulus that enhances EC or VSMC proliferation to form collateral blood vessels or drives the existing branches into small capillaries can be used as the first step in cardiac regenerative therapy. Under such pre-vascularization condition, the implanted stem cell or CMs or *in situ* programmed existing cells will have a fair chance to survive in the damaged cardiac tissue.

Importantly, adult mouse heart appeared to be promisingly recovered post-silencing of circRNA Nfix in the periphery of the infarcted region [69]. This improvement in the prognosis is brought about by a combination of restoration of CMs and angiogenesis. Thus, it provides a promise to use Meis1 transcript factor as a potential therapeutic target in modulating ECs and CMs, thereby increasing vascularization or cardiomyogenesis in the infarcted heart as it acts immediately upstream of Nfix by binding and activating its enhancer; however, how Meis1 modulates ECs or CMs as an upstream element of endothelial-specific genes, such as *Tal1, Fli1, Runx1, Scl1, etc*, [9, 66] remain not clear.

#### 3.3. Meis1, inflammation, and cardiac repair

Cardiac tissue repair does not only involve CMs and ECs, but also dependent on the immune cells, like macrophages that are recruited immediately after tissue injury. The inflammatory response after tissue injury is deleterious yet essential for tissue repair, where macrophages play a critical role in the initiation, maintenance and shut down the process [91]. The remarkable capacity of neonatal mouse heart to repair itself after apical resection or myocardial infarction is also attributed to the type of macrophage population residing in the cardiac tissue [92]. Calvo *et al.* showed that neonatal cardiac macrophage composition differs from that of adult heart. The interesting findings from this study underscore the important distinction between the resident macrophage (embryonic origin) and the monocyte derived macrophage with respect to cardiac tissue injury. Neonatal heart selectively expands the number of resident macrophages which is reparative and produces minimal inflation whereas adult heart recruits more monocyte derived macrophages leading to prolonged inflammation [92].

Meis1 is essential for macrophage differentiation of myeloid progenitors in cooperation with Hoxa9 [93]. *Meis1* is downregulated during monocyte-macrophage terminal differentiation along with most of the *Hoxa* genes [16]. This suggests the monocyte-derived macrophages result from downregulation of *Meis1*, but it will be interesting to see the level of Meis1 in the resident macrophages. This link between *Meis1* and the cardiac macrophages will uncover many puzzles associated with inflammation and cardiac regeneration.

# Conclusion

In summary, the discovery of Meis1 in leukemic pathogenesis has been beneficial in understanding of its biological roles in cancer and HSCs, it also gives us an opportunity in grasping some of the vital mechanisms in cardiac development and regeneration. It is commonly agreed that Meis1 plays critical roles in disease progression, including in MI, apart from being an important developmental gene.

Gene therapy is a promising strategy in this era of genomics and transcriptomics. Multiple roles of Meis1 in fetal development and adult disease conditions of the mammalians gives us hope to utilize the known information to ameliorate the impairment caused by this gene through gene therapy. Meanwhile, stem cell-based therapy has caught a lot of attention to serve as an alternative promising strategy to reinstate the damaged heart, which can overcome the fundamental challenge of rare or non-proliferative nature of adult CMs. Yet, this faces another obstruction in which the newly generated CMs located on the ischemic and fibrotic-scarred tissue is unable to obtain sufficient supply of blood to ensure their survival and proliferation. In other words, myocardial regeneration and survival after transplantation largely surely depend on suitable angiogenesis or arteriogenesis to restore blood flow in the ischemic territory in the heart. Based on the literature evidence (Table 1), modulation of Meis1 in CMs, ECs and VSMCs may offer a new approach to improve the outcomes of stem cell-based therapy.

# Acknowledgment

This work was supported by the National Institute of Health (1R15HL140528) and the Seed Grant from the College of Veterinary Medicine at Virginia Tech. The funding sources was not involved in preparing and submitting this manuscript. We would like to thank the online image software Biorender (https://biorender.com/) for making it helpful to create the figures.

# Abbreviations:

AGM	Aorta gonad mesenchyme	
ALDO	Aldolase	
ALL	Acute lymphoid leukemia	
AML	Acute myeloid leukemia	
Bmi1	B lymphoma mo-mlv insertion region 1	
CMs	Cardiomyocytes	
Cdk6	Cyclin Dependent kinase 6	
Cdkn2a	Cyclin Dependent kinase inhibitor 2a	
C-Myb	Myb-like DNA-binding gene	
Creb	Camp response element binding	
ECs	endothelial cells	
ENO	Enolase	
Fli1	Friend leukemia integration 1	
Flk1	Fetal liver kinase 1 (Aliases: Kdr, Vegfr2)	
Flt3	FMS related tyrosine kinase 3 (Aliases: CD135)	

HF	heart failure		
Hif1a or 2a	Hypoxia inducible factor 1a or 2a		
Hox	Homeobox		
HSCs	Hematopoietic stem cells		
KDR	Kinase insert domain receptor (Aliases: Vegfr2)		
LDHB	Lactate dehydrogenase b		
Meis1	Myeloid ecotropic viral integration site 1		
MI	Myocardial infarction		
MSTN	Myostatin		
Myocd	Myocardin		
Nf-Y	Nuclear transcription factor Y		
Nkx2.5	NK2 homeobox 5		
Pbx	Pre-b-cell leukemia homeobox		
PCBP2	Poly (rc)-binding protein 2		
P-D axis	Proximo-Distal axis		
PECAM1	Platelet and Endothelial Cell Adhesion Molecule-1 (Aliases: CD31)		
Popdc2	Popeye domain-containing protein 2		
Prep1	Pbx regulating protein 1		
ROS	Reactive Oxygen Species		
RLS	Restless leg syndrome		
Runx1	Runt related transcription factor 1		
SCL	Stem cell leukemia (Aliases: Tal1)		
shRNA	Short hairpin RNA		
Tal1	Tal bhlh transcription factor 1 (Aliases: SCL)		
TALE	Three amino acid loop extension		
Tbx2	box 2		
Vegfr	Vascular endothelial growth factor receptor		
VSMCs	vascular smooth muscle cells		

- Mehra R, Global public health problem of sudden cardiac death, J Electrocardiol 40 (6 Suppl) (2007) S118–122. 10.1016/j.jelectrocard.2007.06.023. [PubMed: 17993308]
- [2]. Lee SJ, Lee CK, Kang S, Park I, Kim YH, Kim SK, Hong SP, Bae H, et al., Angiopoietin-2 exacerbates cardiac hypoxia and inflammation after myocardial infarction, J Clin Invest 128 (11) (2018) 5018–5033. 10.1172/JCI99659. [PubMed: 30295643]
- [3]. Ingason AB, Goldstone AB, Paulsen MJ, Thakore AD, Truong VN, Edwards BB, Eskandari A, Bollig T, et al., Angiogenesis precedes cardiomyocyte migration in regenerating mammalian hearts, J Thorac Cardiovasc Surg 155 (3) (2018) 1118–1127 e1111. 10.1016/j.jtcvs.2017.08.127. [PubMed: 29452461]
- [4]. Miyagawa S, Sawa Y, Taketani S, Kawaguchi N, Nakamura T, Matsuura N, Matsuda H, [myocardial regeneration therapy for heart failure: Hepatocyte growth factor enhances the effect of cellular cardiomyoplasty], J Cardiol 41 (1) (2003) 36–38. [PubMed: 12564112]
- [5]. Mathison M, Gersch RP, Nasser A, Lilo S, Korman M, Fourman M, Hackett N, Shroyer K, et al., In vivo cardiac cellular reprogramming efficacy is enhanced by angiogenic preconditioning of the infarcted myocardium with vascular endothelial growth factor, J Am Heart Assoc 1 (6) (2012) e005652 10.1161/JAHA.112.005652. [PubMed: 23316332]
- [6]. Mahmoud AI, Kocabas F, Muralidhar SA, Kimura W, Koura AS, Thet S, Porrello ER, Sadek HA, Meis1 regulates postnatal cardiomyocyte cell cycle arrest, Nature 497 (7448) (2013) 249–253. 10.1038/nature12054. [PubMed: 23594737]
- [7]. Mercader N, Selleri L, Criado LM, Pallares P, Parras C, Cleary ML, Torres M, Ectopic meis1 expression in the mouse limb bud alters p-d patterning in a pbx1-independent manner, The International journal of developmental biology 53 (8–10) (2009) 1483–1494. 10.1387/ ijdb.072430nm. [PubMed: 19247936]
- [8]. Garcia-Fernandez J, The genesis and evolution of homeobox gene clusters, Nat Rev Genet 6 (12) (2005) 881–892. 10.1038/nrg1723. [PubMed: 16341069]
- [9]. Amali AA, Sie L, Winkler C, Featherstone M, Zebrafish hoxd4a acts upstream of meis1.1 to direct vasculogenesis, angiogenesis and hematopoiesis, PLoS One 8 (3) (2013) e58857 10.1371/ journal.pone.0058857. [PubMed: 23554940]
- [10]. Alharbi RA, Pettengell R, Pandha HS, Morgan R, The role of hox genes in normal hematopoiesis and acute leukemia, Leukemia 27 (5) (2013) 1000–1008. 10.1038/leu.2012.356. [PubMed: 23212154]
- [11]. Cvejic A, Serbanovic-Canic J, Stemple DL, Ouwehand WH, The role of meis1 in primitive and definitive hematopoiesis during zebrafish development, Haematologica 96 (2) (2011) 190–198. 10.3324/haematol.2010.027698. [PubMed: 21048033]
- [12]. Argiropoulos B, Yung E, Humphries RK, Unraveling the crucial roles of meis1 in leukemogenesis and normal hematopoiesis, Genes & development 21 (22) (2007) 2845–2849.
   10.1101/gad.1619407. [PubMed: 18006680]
- [13]. Azcoitia V, Aracil M, Martinez AC, Torres M, The homeodomain protein meis1 is essential for definitive hematopoiesis and vascular patterning in the mouse embryo, Dev Biol 280 (2) (2005) 307–320. 10.1016/j.ydbio.2005.01.004. [PubMed: 15882575]
- [14]. Kocabas F, Zheng J, Thet S, Copeland NG, Jenkins NA, DeBerardinis RJ, Zhang C, Sadek HA, Meis1 regulates the metabolic phenotype and oxidant defense of hematopoietic stem cells, Blood 120 (25) (2012) 4963–4972. 10.1182/blood-2012-05-432260. [PubMed: 22995899]
- [15]. Schneider E, Staffas A, Rohner L, Malmberg ED, Ashouri A, Krowiorz K, Pochert N, Miller C, et al., Micro-ribonucleic acid-155 is a direct target of meis1, but not a driver in acute myeloid leukemia, Haematologica 103 (2) (2018) 246–255. 10.3324/haematol.2017.177485. [PubMed: 29217774]
- [16]. Wong P, Iwasaki M, Somervaille TC, So CW, Cleary ML, Meis1 is an essential and rate-limiting regulator of mll leukemia stem cell potential, Genes & development 21 (21) (2007) 2762–2774. 10.1101/gad.1602107. [PubMed: 17942707]
- [17]. Pineault N, Buske C, Feuring-Buske M, Abramovich C, Rosten P, Hogge DE, Aplan PD, Humphries RK, Induction of acute myeloid leukemia in mice by the human leukemia-specific

fusion gene nup98-hoxd13 in concert with meis1, Blood 101 (11) (2003) 4529–4538. 10.1182/ blood-2002-08-2484. [PubMed: 12543865]

- [18]. Rozovskaia T, Feinstein E, Mor O, Foa R, Blechman J, Nakamura T, Croce CM, Cimino G, et al., Upregulation of meis1 and hoxa9 in acute lymphocytic leukemias with the t(4 : 11) abnormality, Oncogene 20 (7) (2001) 874–878. 10.1038/sj.onc.1204174. [PubMed: 11314021]
- [19]. Afonja O, Smith JE Jr., Cheng DM, Goldenberg AS, Amorosi E, Shimamoto T, Nakamura S, Ohyashiki K, et al., Meis1 and hoxa7 genes in human acute myeloid leukemia, Leuk Res 24 (10) (2000) 849–855. [PubMed: 10996203]
- [20]. Lawrence HJ, Rozenfeld S, Cruz C, Matsukuma K, Kwong A, Komuves L, Buchberg AM, Largman C, Frequent co-expression of the hoxa9 and meis1 homeobox genes in human myeloid leukemias, Leukemia 13 (12) (1999) 1993–1999. [PubMed: 10602420]
- [21]. Moskow JJ, Bullrich F, Huebner K, Daar IO, Buchberg AM, Meis1, a pbx1-related homeobox gene involved in myeloid leukemia in bxh-2 mice, Mol Cell Biol 15 (10) (1995) 5434–5443.
   [PubMed: 7565694]
- [22]. Mercader N, Leonardo E, Azpiazu N, Serrano A, Morata G, Martinez C, Torres M, Conserved regulation of proximodistal limb axis development by meis1/hth, Nature 402 (6760) (1999) 425– 429. 10.1038/46580. [PubMed: 10586884]
- [23]. Schulte EC, Knauf F, Kemlink D, Schormair B, Lichtner P, Gieger C, Meitinger T, Winkelmann J, Variant screening of the coding regions of meis1 in patients with restless legs syndrome, Neurology 76 (12) (2011) 1106–1108. 10.1212/WNL.0b013e318211c366. [PubMed: 21422461]
- [24]. Allen RP, Donelson NC, Jones BC, Li Y, Manconi M, Rye DB, Sanyal S, Winkelmann J, Animal models of rls phenotypes, Sleep Med 31 (2017) 23–28. 10.1016/j.sleep.2016.08.002. [PubMed: 27839945]
- [25]. Marcos S, Gonzalez-Lazaro M, Beccari L, Carramolino L, Martin-Bermejo MJ, Amarie O, Mateos-San Martin D, Torroja C, et al., Meis1 coordinates a network of genes implicated in eye development and microphthalmia, Development 142 (17) (2015) 3009–3020. 10.1242/ dev.122176. [PubMed: 26253404]
- [26]. Stankunas K, Shang C, Twu KY, Kao SC, Jenkins NA, Copeland NG, Sanyal M, Selleri L, et al., Pbx/meis deficiencies demonstrate multigenetic origins of congenital heart disease, Circ Res 103 (7) (2008) 702–709. 10.1161/CIRCRESAHA.108.175489. [PubMed: 18723445]
- [27]. Hisa T, Spence SE, Rachel RA, Fujita M, Nakamura T, Ward JM, Devor-Henneman DE, Saiki Y, et al., Hematopoietic, angiogenic and eye defects in meis1 mutant animals, EMBO J 23 (2) (2004) 450–459. 10.1038/sj.emboj.7600038. [PubMed: 14713950]
- [28]. Pineault N, Helgason CD, Lawrence HJ, Humphries RK, Differential expression of hox, meis1, and pbx1 genes in primitive cells throughout murine hematopoietic ontogeny, Experimental hematology 30 (1) (2002) 49–57. [PubMed: 11823037]
- [29]. Penkov D, Mateos San Martin D, Fernandez-Diaz LC, Rossello CA, Torroja C, Sanchez-Cabo F, Warnatz HJ, Sultan M, et al., Analysis of the DNA-binding profile and function of tale homeoproteins reveals their specialization and specific interactions with hox genes/proteins, Cell Rep 3 (4) (2013) 1321–1333. 10.1016/j.celrep.2013.03.029. [PubMed: 23602564]
- [30]. Burglin TR, Analysis of tale superclass homeobox genes (meis, pbc, knox, iroquois, tgif) reveals a novel domain conserved between plants and animals, Nucleic Acids Res 25 (21) (1997) 4173– 4180. 10.1093/nar/25.21.4173. [PubMed: 9336443]
- [31]. Mukherjee K, Burglin TR, Comprehensive analysis of animal tale homeobox genes: New conserved motifs and cases of accelerated evolution, J Mol Evol 65 (2) (2007) 137–153. 10.1007/ s00239-006-0023-0. [PubMed: 17665086]
- [32]. Zhang X, Friedman A, Heaney S, Purcell P, Maas RL, Meis homeoproteins directly regulate pax6 during vertebrate lens morphogenesis, Genes & development 16 (16) (2002) 2097–2107. 10.1101/gad.1007602. [PubMed: 12183364]
- [33]. Longobardi E, Penkov D, Mateos D, De Florian G, Torres M, Blasi F, Biochemistry of the tale transcription factors prep, meis, and pbx in vertebrates, Dev Dyn 243 (1) (2014) 59–75. 10.1002/ dvdy.24016. [PubMed: 23873833]

- [34]. Hill TP, Taketo MM, Birchmeier W, Hartmann C, Multiple roles of mesenchymal beta-catenin during murine limb patterning, Development 133 (7) (2006) 1219–1229. 10.1242/dev.02298.
   [PubMed: 16495310]
- [35]. Bischof LJ, Kagawa N, Moskow JJ, Takahashi Y, Iwamatsu A, Buchberg AM, Waterman MR, Members of the meis1 and pbx homeodomain protein families cooperatively bind a campresponsive sequence (crs1) from bovine cyp17, J Biol Chem 273 (14) (1998) 7941–7948. 10.1074/jbc.273.14.7941. [PubMed: 9525891]
- [36]. Lund J, Bakke M, Mellgren G, Morohashi K, Doskeland SO, Transcriptional regulation of the bovine cyp17 gene by camp, Steroids 62 (1) (1997) 43–45. 10.1016/s0039-128x(96)00157-2.
  [PubMed: 9029713]
- [37]. Grade CVC, Mantovani CS, Fontoura MA, Yusuf F, Brand-Saberi B, Alvares LE, Creb, nf-y and meis1 conserved binding sites are essential to balance myostatin promoter/enhancer activity during early myogenesis, Mol Biol Rep 44 (5) (2017) 419–427. 10.1007/s11033-017-4126-z. [PubMed: 28956216]
- [38]. Waskiewicz AJ, Rikhof HA, Hernandez RE, Moens CB, Zebrafish meis functions to stabilize pbx proteins and regulate hindbrain patterning, Development 128 (21) (2001) 4139–4151. [PubMed: 11684652]
- [39]. Dibner C, Elias S, Frank D, Xmeis3 protein activity is required for proper hindbrain patterning in xenopus laevis embryos, Development 128 (18) (2001) 3415–3426. [PubMed: 11566848]
- [40]. Choe SK, Vlachakis N, Sagerstrom CG, Meis family proteins are required for hindbrain development in the zebrafish, Development 129 (3) (2002) 585–595. [PubMed: 11830560]
- [41]. Kawagoe H, Humphries RK, Blair A, Sutherland HJ, Hogge DE, Expression of hox genes, hox cofactors, and mll in phenotypically and functionally defined subpopulations of leukemic and normal human hematopoietic cells, Leukemia 13 (5) (1999) 687–698. [PubMed: 10374871]
- [42]. Imamura T, Morimoto A, Takanashi M, Hibi S, Sugimoto T, Ishii E, Imashuku S, Frequent coexpression of hoxa9 and meis1 genes in infant acute lymphoblastic leukaemia with mll rearrangement, Br J Haematol 119 (1) (2002) 119–121. [PubMed: 12358913]
- [43]. Nakamura T, Jenkins NA, Copeland NG, Identification of a new family of pbx-related homeobox genes, Oncogene 13 (10) (1996) 2235–2242. [PubMed: 8950991]
- [44]. Nakamura T, Largaespada DA, Shaughnessy JD Jr., Jenkins NA, Copeland NG, Cooperative activation of hoxa and pbx1-related genes in murine myeloid leukaemias, Nat Genet 12 (2) (1996) 149–153. 10.1038/ng0296-149. [PubMed: 8563752]
- [45]. Shen WF, Rozenfeld S, Kwong A, Kom ves LG, Lawrence HJ, Largman C, Hoxa9 forms triple complexes with pbx2 and meis1 in myeloid cells, Mol Cell Biol 19 (4) (1999) 3051–3061. 10.1128/mcb.19.4.3051. [PubMed: 10082572]
- [46]. Wang GG, Pasillas MP, Kamps MP, Meis1 programs transcription of flt3 and cancer stem cell character, using a mechanism that requires interaction with pbx and a novel function of the meis1 c-terminus, Blood 106 (1) (2005) 254–264. 10.1182/blood-2004-12-4664. [PubMed: 15755900]
- [47]. Yuan X, Braun T, An unexpected switch: Regulation of cardiomyocyte proliferation by the homeobox gene meis1, Circ Res 113 (3) (2013) 245–248. 10.1161/CIRCRESAHA.113.302023.
  [PubMed: 23868827]
- [48]. Gonzalez-Lazaro M, Rosello-Diez A, Delgado I, Carramolino L, Sanguino MA, Giovinazzo G, Torres M, Two new targeted alleles for the comprehensive analysis of meis1 functions in the mouse, Genesis 52 (12) (2014) 967–975. 10.1002/dvg.22833. [PubMed: 25363539]
- [49]. Simsek T, Kocabas F, Zheng J, Deberardinis RJ, Mahmoud AI, Olson EN, Schneider JW, Zhang CC, et al., The distinct metabolic profile of hematopoietic stem cells reflects their location in a hypoxic niche, Cell Stem Cell 7 (3) (2010) 380–390. 10.1016/j.stem.2010.07.011. [PubMed: 20804973]
- [50]. Risau W, Sariola H, Zerwes HG, Sasse J, Ekblom P, Kemler R, Doetschman T, Vasculogenesis and angiogenesis in embryonic-stem-cell-derived embryoid bodies, Development 102 (3) (1988) 471–478. [PubMed: 2460305]
- [51]. Poole TJ, Coffin JD, Vasculogenesis and angiogenesis: Two distinct morphogenetic mechanisms establish embryonic vascular pattern, The Journal of experimental zoology 251 (2) (1989) 224– 231. 10.1002/jez.1402510210. [PubMed: 2671254]

- [52]. Patan S, Vasculogenesis and angiogenesis, Cancer Treat Res 117 (2004) 3–32. [PubMed: 15015550]
- [53]. Folkman J, Tumor angiogenesis, Advances in cancer research 43 (1985) 175–203. [PubMed: 2581424]
- [54]. Van Royen N, Piek JJ, Schaper W, Bode C, Buschmann I, Arteriogenesis: Mechanisms and modulation of collateral artery development, J Nucl Cardiol 8 (6) (2001) 687–693. 10.1067/ mnc.2001.118924. [PubMed: 11725265]
- [55]. Helisch A, Schaper W, Arteriogenesis: The development and growth of collateral arteries, Microcirculation 10 (1) (2003) 83–97. 10.1038/sj.mn.7800173. [PubMed: 12610665]
- [56]. Carmeliet P, Mechanisms of angiogenesis and arteriogenesis, Nat Med 6 (4) (2000) 389–395. 10.1038/74651. [PubMed: 10742145]
- [57]. Ellertsdottir E, Lenard A, Blum Y, Krudewig A, Herwig L, Affolter M, Belting HG, Vascular morphogenesis in the zebrafish embryo, Dev Biol 341 (1) (2010) 56–65. 10.1016/ j.ydbio.2009.10.035. [PubMed: 19895803]
- [58]. Jing L, Zon LI, Zebrafish as a model for normal and malignant hematopoiesis, Dis Model Mech 4
  (4) (2011) 433–438. 10.1242/dmm.006791. [PubMed: 21708900]
- [59]. Martin CS, Moriyama A, Zon LI, Hematopoietic stem cells, hematopoiesis and disease: Lessons from the zebrafish model, Genome Med 3 (12) (2011) 83 10.1186/gm299. [PubMed: 22206610]
- [60]. Gore AV, Pillay LM, Venero Galanternik M, Weinstein BM, The zebrafish: A fintastic model for hematopoietic development and disease, Wiley Interdiscip Rev Dev Biol 7 (3) (2018) e312 10.1002/wdev.312. [PubMed: 29436122]
- [61]. Howe K, Clark MD, Torroja CF, Torrance J, Berthelot C, Muffato M, Collins JE, Humphray S, et al., The zebrafish reference genome sequence and its relationship to the human genome, Nature 496 (7446) (2013) 498–503. 10.1038/nature12111. [PubMed: 23594743]
- [62]. Minehata K, Kawahara A, Suzuki T, Meis1 regulates the development of endothelial cells in zebrafish, Biochemical and biophysical research communications 374 (4) (2008) 647–652. 10.1016/j.bbrc.2008.07.075. [PubMed: 18656453]
- [63]. Kassmeyer S, Plendl J, Custodis P, Bahramsoltani M, New insights in vascular development: Vasculogenesis and endothelial progenitor cells, Anat Histol Embryol 38 (1) (2009) 1–11. 10.1111/j.1439-0264.2008.00894.x. [PubMed: 18983622]
- [64]. Charboneau A, East L, Mulholland N, Rohde M, Boudreau N, Pbx1 is required for hox d3-mediated angiogenesis, Angiogenesis 8 (4) (2005) 289–296. 10.1007/s10456-005-9016-7.
  [PubMed: 16328158]
- [65]. Richard C, Drevon C, Canto PY, Villain G, Bollerot K, Lempereur A, Teillet MA, Vincent C, et al., Endothelio-mesenchymal interaction controls runx1 expression and modulates the notch pathway to initiate aortic hematopoiesis, Dev Cell 24 (6) (2013) 600–611. 10.1016/ j.devcel.2013.02.011. [PubMed: 23537631]
- [66]. Wang H, Liu C, Liu X, Wang M, Wu D, Gao J, Su P, Nakahata T, et al., Meis1 regulates hemogenic endothelial generation, megakaryopoiesis, and thrombopoiesis in human pluripotent stem cells by targeting tal1 and fli1, Stem Cell Reports 10 (2) (2018) 447–460. 10.1016/ j.stemcr.2017.12.017. [PubMed: 29358086]
- [67]. Lazrak M, Deleuze V, Noel D, Haouzi D, Chalhoub E, Dohet C, Robbins I, Mathieu D, The bhlh tal-1/scl regulates endothelial cell migration and morphogenesis, J Cell Sci 117 (Pt 7) (2004) 1161–1171. 10.1242/jcs.00969. [PubMed: 14970264]
- [68]. Liu F, Walmsley M, Rodaway A, Patient R, Fli1 acts at the top of the transcriptional network driving blood and endothelial development, Curr Biol 18 (16) (2008) 1234–1240. 10.1016/ j.cub.2008.07.048. [PubMed: 18718762]
- [69]. Huang S, Li X, Zheng H, Si X, Li B, Wei G, Li C, Chen Y, et al., Loss of super-enhancerregulated circrna nfix induces cardiac regeneration after myocardial infarction in adult mice, Circulation 139 (25) (2019) 2857–2876. 10.1161/CIRCULATIONAHA.118.038361. [PubMed: 30947518]
- [70]. Smith JG, Magnani JW, Palmer C, Meng YA, Soliman EZ, Musani SK, Kerr KF, Schnabel RB, et al., Genome-wide association studies of the pr interval in african americans, PLoS Genet 7 (2) (2011) e1001304 10.1371/journal.pgen.1001304. [PubMed: 21347284]

- [71]. Schormair B, Plag J, Kaffe M, Gross N, Czamara D, Samtleben W, Lichtner P, Strohle A, et al., Meis1 and btbd9: Genetic association with restless leg syndrome in end stage renal disease, J Med Genet 48 (7) (2011) 462–466. 10.1136/jmg.2010.087858. [PubMed: 21572129]
- [72]. Spieler D, Kaffe M, Knauf F, Bessa J, Tena JJ, Giesert F, Schormair B, Tilch E, et al., Restless legs syndrome-associated intronic common variant in meis1 alters enhancer function in the developing telencephalon, Genome Res 24 (4) (2014) 592–603. 10.1101/gr.166751.113. [PubMed: 24642863]
- [73]. Bouilloux F, Thireau J, Venteo S, Farah C, Karam S, Dauvilliers Y, Valmier J, Copeland NG, et al., Loss of the transcription factor meis1 prevents sympathetic neurons target-field innervation and increases susceptibility to sudden cardiac death, Elife 5 (2016) 10.7554/eLife.11627.
- [74]. Jewhurst K, McLaughlin KA, Beyond the mammalian heart: Fish and amphibians as a model for cardiac repair and regeneration, J Dev Biol 4 (1) (2015) 10.3390/jdb4010001.
- [75]. Porrello ER, Mahmoud AI, Simpson E, Hill JA, Richardson JA, Olson EN, Sadek HA, Transient regenerative potential of the neonatal mouse heart, Science 331 (6020) (2011) 1078–1080. 10.1126/science.1200708. [PubMed: 21350179]
- [76]. Maves L, Tyler A, Moens CB, Tapscott SJ, Pbx acts with hand2 in early myocardial differentiation, Dev Biol 333 (2) (2009) 409–418. 10.1016/j.ydbio.2009.07.004. [PubMed: 19607825]
- [77]. Lindgren IM, Drake RR, Chattergoon NN, Thornburg KL, Down-regulation of meis1 promotes the maturation of oxidative phosphorylation in perinatal cardiomyocytes, FASEB J 33 (6) (2019) 7417–7426. 10.1096/fj.201801330RR. [PubMed: 30884246]
- [78]. Brade T, Pane LS, Moretti A, Chien KR, Laugwitz KL, Embryonic heart progenitors and cardiogenesis, Cold Spring Harb Perspect Med 3 (10) (2013) a013847 10.1101/ cshperspect.a013847. [PubMed: 24086063]
- [79]. Murry CE, Keller G, Differentiation of embryonic stem cells to clinically relevant populations: Lessons from embryonic development, Cell 132 (4) (2008) 661–680. 10.1016/j.cell.2008.02.008.
   [PubMed: 18295582]
- [80]. Wamstad JA, Alexander JM, Truty RM, Shrikumar A, Li F, Eilertson KE, Ding H, Wylie JN, et al., Dynamic and coordinated epigenetic regulation of developmental transitions in the cardiac lineage, Cell 151 (1) (2012) 206–220. 10.1016/j.cell.2012.07.035. [PubMed: 22981692]
- [81]. Martinez SR, Gay MS, Zhang L, Epigenetic mechanisms in heart development and disease, Drug Discov Today 20 (7) (2015) 799–811. 10.1016/j.drudis.2014.12.018. [PubMed: 25572405]
- [82]. Akiyama T, Suzuki O, Matsuda J, Aoki F, Dynamic replacement of histone h3 variants reprograms epigenetic marks in early mouse embryos, PLoS Genet 7 (10) (2011) e1002279 10.1371/journal.pgen.1002279. [PubMed: 21998593]
- [83]. Saleh RNM, Dilg D, Abou Zeid AA, Hashad DI, Scambler PJ, Chapgier ALA, Hira directly targets the enhancers of selected cardiac transcription factors during in vitro differentiation of mouse embryonic stem cells, Mol Biol Rep 45 (5) (2018) 1001–1011. 10.1007/s11033-018-4247z. [PubMed: 30030774]
- [84]. Dupays L, Shang C, Wilson R, Kotecha S, Wood S, Towers N, Mohun T, Sequential binding of meis1 and nkx2–5 on the popdc2 gene: A mechanism for spatiotemporal regulation of enhancers during cardiogenesis, Cell Rep 13 (1) (2015) 183–195. 10.1016/j.celrep.2015.08.065. [PubMed: 26411676]
- [85]. Xiang FL, Guo M, Yutzey KE, Overexpression of tbx20 in adult cardiomyocytes promotes proliferation and improves cardiac function after myocardial infarction, Circulation 133 (11) (2016) 1081–1092. 10.1161/CIRCULATIONAHA.115.019357. [PubMed: 26841808]
- [86]. Zhang Y, Si Y, Ma N, Meis1 promotes poly (rc)-binding protein 2 expression and inhibits angiotensin ii-induced cardiomyocyte hypertrophy, IUBMB Life 68 (1) (2016) 13–22. 10.1002/ iub.1456. [PubMed: 26597775]
- [87]. Pandey R, Yang Y, Jackson L, Ahmed RP, Micrornas regulating meis1 expression and inducing cardiomyocyte proliferation, Cardiovasc Regen Med 3 (2016)
- [88]. Piquereau J, Ventura-Clapier R, Maturation of cardiac energy metabolism during perinatal development, Front Physiol 9 (2018) 959 10.3389/fphys.2018.00959. [PubMed: 30072919]

- [89]. Dedkova EN, Blatter LA, Measuring mitochondrial function in intact cardiac myocytes, J Mol Cell Cardiol 52 (1) (2012) 48–61. 10.1016/j.yjmcc.2011.08.030. [PubMed: 21964191]
- [90]. Mathison M, Rosengart TK, Heart regeneration: The endothelial cell comes first, J Thorac Cardiovasc Surg 155 (3) (2018) 1128–1129. 10.1016/j.jtcvs.2017.09.106. [PubMed: 29452462]
- [91]. Fujiwara N, Kobayashi K, Macrophages in inflammation, Curr Drug Targets Inflamm Allergy 4 (3) (2005) 281–286. [PubMed: 16101534]
- [92]. Lavine KJ, Epelman S, Uchida K, Weber KJ, Nichols CG, Schilling JD, Ornitz DM, Randolph GJ, et al., Distinct macrophage lineages contribute to disparate patterns of cardiac recovery and remodeling in the neonatal and adult heart, Proc Natl Acad Sci U S A 111 (45) (2014) 16029–16034. 10.1073/pnas.1406508111. [PubMed: 25349429]
- [93]. Calvo KR, Knoepfler PS, Sykes DB, Pasillas MP, Kamps MP, Meis1a suppresses differentiation by g-csf and promotes proliferation by scf: Potential mechanisms of cooperativity with hoxa9 in myeloid leukemia, Proc Natl Acad Sci U S A 98 (23) (2001) 13120–13125. 10.1073/ pnas.231115398. [PubMed: 11687616]



# Meis1 modulates hyper-proliferation of myeloid leukemic cells

#### Fig 1. Illustration of Meis1 as a Hox cofactor in myeloid leukemic cells.

Meis1, as a part of Pbx-HoxA9- Meis1 trimeric complex, regulates the expression of cell cycle regulators like Cdk6, c-Myb, Flt3 etc. resulting in hyper-proliferation. Meis1: Myeloid Ecotropic Insertion Site 1; Cdk6: Cyclin Dependent kinase 6; C-Myb: Myb-like DNAbinding gene; Pbx: Pre-B cell regulating protein 1; Flt3: FMS related tyrosine kinase 3.







**Fig 3. Illustration of EC specific role of Meis1.** HoxD4 acts upstream of Meis1 which regulates several EC genes like Tal1, Runx1, Fli1 etc. through various intermediates proteins. Normal proliferation and maintenance of ECs is widely regulated by Meis1 acting upstream of several genes. ECs: endothelial cells; Tal: Tal bhlh transcription factor 1; Runx1: Runt related transcription factor 1; Fli1: Friend leukemic integration 1.



# Meis1 is a key regulator in embryonic cardiomyocyte proliferation

# Fig4. Meis1 is a key regulator of cardiogenesis by modulating embryonic CMs.

In embryonic CM, HIRA positively regulates *Meis1* transcription. Meis1 acts as transcriptional regulators of cardiac genes, *Myocd* and *Popdc2*, by binding to their promoters and enhancers respectively. Normal expression of Meis1 at embryonic stage is required for cardiogenesis and formation of distal outflow tract. CMs, cardiomyocytes; Myocd: Myocardin; Popdc2: Popeye domain-containing protein 2.



### Roles of Meis1 in neonatal and adult cardiomyocyte proliferation

#### Fig 5: Illustration of the role of Meis1 in neonatal/ adult CM.

Over expression of Meis1 in neonatal cardiomyocytes is accompanied by Pcbp-2 overexpression, which inhibits the transcription of *Ang2*. This causes neonatal CMs in premature stage via cell cycle arrest, inhibitions of both neonatal heart regeneration and CM hypertrophy. On the other hand, transcriptional inhibition of Meis1 by overexpressing Tbx20- or miRNA-mediated translational inhibition leads to increased proliferation of CMS at neonatal and adult (?) stage and preserves cardiac function of neonatal heart post-MI. CMs: cardiomyocytes; Pcbp-2: Poly (rc)-binding protein 2; Ang2: angiotensin 2; Tbx20: T-box 20.

# Table 1.

## Summary of Meis1 in Modulating Cardiovascular System (see footnote for abbreviations)

Genetic alteration/technique used	in-vitro or in-vivo model	Functional effect/Observations	Reference
Targeted disruption of Meis1 (KO)	Mouse	Embryonic lethality at mid-gestation day 14.5; abnormal vascular patterning	[13, 27]
Morpholino injected KD	Zebrafish	Malformed heart and disrupted vessels	[11, 57]
Targeted disruption of <i>Meis1</i> (KO)	C57BL/6 mouse	<i>Meis1</i> -null embryos display an overriding aorta and ventricular septal defect (Congenital abnormalities)	[26]
Cardiomyocyte specific Meis1 KO	C57BL/6 mouse	Cardiomyocyte cell cycle arrest	[6]
Chip-Seq analysis of embryonic heart	Mouse	Meis1 is involved in spatio-tempral regulation during cardiogenesis.	[81]
Overexpression of Tbx20	Mouse differentiated cardiomyocytes	Overexpression of Tbx20, an inhibitor Meis1 promotes proliferation and improves cardiac function after Ml.	[82]
Induction of hypertrophy in mice	C57BL/6 mouse, Human heart samples, neonatal rat cardiomyocytes	Meis1 promotes PCBP2 expression in cardiomyocytes and inhibits Angl1 induced cardiac hypertrophy	[83]
miRNA overexpression in cardiomyocytes resulting in functional inhibition of Meis1	Isolated rat cardiomyocytes	Functional inhibition of Meis1 in adult cardiomyocytes, increases proliferation.	[84]
RNA-Seq and ChIP-Seq analysis of WT and histone chaperon <i>Hira</i> null cells	Mouse embryonic stem cells and early cardiomyocyte progenitors	<i>Meis1</i> is directly regulated by Hira iduring early cardiogenesis	[80]
Loss of function assay	Mouse	Circular RNA Nfix acts downstream of Meis1 and loss of which promotes angiogenesis and better heart regeneration post-MI,	[66]
siRNA mediated knock-down	Isolated sheep cardiomyocytes	Meis1 down regulation promotes oxidative phosphorvlation in perinatal cardiomvocytes	[74]

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